



Neutrophils as a Protagonist and Target in Chronic Rhinosinusitis

Hai Wang · Li Pan · Zheng Liu

Department of Otolaryngology-Head and Neck Surgery, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China

Neutrophils have traditionally been acknowledged as the first immune cells that are recruited to inflamed tissues during acute inflammation. By contrast, their importance in the context of chronic inflammation has been studied in less depth. Neutrophils can be recruited and are largely present in the nasal mucosa of patients with chronic rhinosinusitis (CRS) both in Asians and in Caucasians. Increased infiltration of neutrophils in patients with CRS has been linked to poor corticosteroid response and disease prognosis. Meanwhile, tissue neutrophils may possess specific phenotypic features distinguishing them from resting blood counterparts and are endowed with particular functions, such as cytokines and chemokines production, thus may contribute to the pathogenesis of CRS. This review aims to summarize our current understanding of the pathophysiologic mechanisms of CRS, with a focus on the roles of neutrophils. We discuss recruitment, function, and regulation of neutrophils in CRS and outline the potential therapeutic strategies targeting neutrophils.

Keywords. *Chronic Rhinosinusitis; Immune Response; Nasal Polyp; Neutrophil*

INTRODUCTION

Chronic rhinosinusitis (CRS) is a multifactorial airway disease characterized by persistent inflammation of sinonasal mucosa and associates with over 12 weeks of persistent symptoms including congestion, stuffiness, nasal discharge, pain or facial pressure, impairment or loss of sense of smell, cough, and fatigue [1,2]. It affects a considerable percentage of the population worldwide and renders substantial social and humanistic burden, and direct and indirect healthcare costs [2,3]. CRS has been thought to be a dichotomous disease according to the presence or absence of nasal polyps and are divided into two phenotypes: CRS with nasal polyps (CRSwNP) and CRS without nasal polyps (CRSsNP) [1]. Generally, CRSsNP is associated with a skewed type-1 response leading to a predominance of neutro-

philic inflammation, whereas CRSwNP has a type-2 response dominated eosinophilic inflammation in Caucasian patients [4]. Patients with CRSwNP often have a higher grade of disease severity and are more likely to have a recurrent disease compared to patients with CRSsNP [5]. Recently, increasing evidences demonstrate that the inflammation patterns of CRS may vary across different geographic areas and populations with distinct racial backgrounds [4,6,7]. Significant type-2 response and eosinophilic inflammation can only be observed in 20% to 60% of patients with CRSwNP in Asia countries including China, Korea, Japan, and Malaysia [4,8,9]. In contrast, enhanced type-1 and type-17 response are found in both Asian patients with CRSsNP and CRSwNP as compared with their Caucasian counterparts [4,10]. Based on the tissue eosinophils percentage [7] or the tissue eosinophil cationic protein (ECP)/myeloperoxidase (MPO) ratio [4], CRSwNP is classified into eosinophilic and noneosinophilic type. Cao et al. [7] previously proposed a cutoff of percentage of eosinophils to classify the eosinophilic CRSwNP, which is defined when the percentage of eosinophils exceeds 10% of total infiltrating cells. This cutoff is calculated as twice of the standard deviation of the mean eosinophil percentage in control subjects [7]. Based on this classification, the authors [7] found that about half of Chinese CRSwNP patients

• Received May 7, 2019
Revised June 28, 2019
Accepted June 28, 2019

• Corresponding author: **Zheng Liu**
Department of Otolaryngology-Head and Neck Surgery, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, No. 1095 Jiefang Avenue, Wuhan 430030, China
Tel: +86-27-83663529, Fax: +86-27-83663529
E-mail: zhengliuent@hotmail.com

Copyright © 2019 by Korean Society of Otorhinolaryngology-Head and Neck Surgery.

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

presented noneosinophilic endotype with less impairment of olfactory function, lower frequency of asthma comorbidity, and lower ratio of ethmoid sinus/maxillary sinus CT score [11,12].

Eosinophilic and noneosinophilic CRSwNP demonstrate distinct features of epithelial cell activation, innate lymphoid cell and dendritic cell infiltration, CD4⁺ and CD8⁺ T cell subset polarization, and tissue remodeling [11,13-17]. Particularly, eosinophilic CRSwNP is consistently characterized by type-2 response, whereas noneosinophilic CRSwNP has type-1 and type-17 response skewed inflammation [6,7]. Despite these differences, both subtypes of CRSwNP have been found to have upregulated neutrophil infiltration as compared to controls, and there is a higher upregulation in noneosinophilic CRSwNP than in eosinophilic CRSwNP in Chinese [6,11,16,17]. It has also been reported that CRSwNP in Caucasian patients, which is highly eosinophilic, also has significant levels of neutrophils [4,18-20]. In addition, compared to patients with eosinophilic and noneosinophilic CRSwNP, a further increase of neutrophil infiltration has been noted in patients with CRSsNP [16]. Neutrophilia affects the treatment outcome of CRS patients. It may lead to the poor response to corticosteroid therapy [21]. A recent prospective study reported by Liao et al. [22] showed that in addition to eosinophilic inflammation, significant neutrophilic inflammation was also associated with difficult-to-treat CRS in patients receiving guideline-recommended, glucocorticoid and endoscopic surgery centered treatment, underscoring a crucial role for neutrophils in the pathophysiological processes of CRS. However, compared to eosinophils, studies regarding the role of neutrophils in CRS are still in infant stage, urging the need to explore the function and regulation of neutrophilic inflammation in CRS.

This review aims to summarize our current understanding of the pathophysiologic mechanisms of CRS, with a focus on the roles of neutrophils. We discuss recruitment, function, and regulation of neutrophils in CRS and outline the potential therapeutic strategies targeting neutrophils.

HIGHLIGHTS

- The inflammation patterns of chronic rhinosinusitis (CRS) vary across different geographic areas and populations with distinct racial backgrounds with more neutrophilic inflammation in Asian patients.
- Corticosteroids are the mainstay treatment for eosinophilic CRS, especially for eosinophilic CRS with nasal polyps, whereas macrolide antibiotics are possibly more effective for neutrophilic CRS.
- Targeting the activation and infiltration of neutrophils may offer potential therapeutic approaches to improve clinical outcomes in patients with steroid-insensitive CRS characterized by neutrophilic inflammation.

LOCAL RECRUITMENT OF NEUTROPHILS IN CRS

Neutrophils are terminally differentiated granulocytes that develop in the bone marrow under the control of several key transcription factors including PU.1, CCAAT/enhancer binding protein α (C/EBP α), growth factor independence 1, and C/EBP ϵ [23,24]. Following the instruction of chemokines, neutrophils rapidly leave the circulation and arrive at inflammatory sites according to the chemokine gradient by a carefully orchestrated process [25]. This process is initiated by the endothelial expression of adhesion molecules and followed by adhesion, extravasation and migration of neutrophils [23,25]. Chemokine (C-X-C motif) ligand (CXCL) family members, including CXCL1, CXCL2, CXCL5, and CXCL8, comprise the major chemokines that direct neutrophil migration through engaging G-protein-coupled receptor chemokine (C-X-C motif) receptor (CXCR) 1 and CXCR2 on neutrophils [24,26]. Both tissue infiltrating leukocytes and tissue structure cells (e.g. epithelial and endothelial cells) have been reported to produce neutrophil chemokines in response to inflammatory stimuli or foreign invaders [26]. In CRS, nasal epithelial cells are an important source of neutrophil chemokines as they have been documented to secrete CXCL8 in response to diesel exhaust particle, bacteria, or inflammatory stimuli [27-30]. In addition, CXCL1, CXCL2, and CXCL8 can be produced by fibroblasts [31,32], neutrophils [6] and mast cells [33] in nasal tissues under the stimulation with different triggers. Elevated production of CXCL8 has been found in patients with CRSsNP and CRSwNP in comparison with control subjects in China and Western countries [4,34]. Kim et al. [35] found increased expression of CXCL1, CXCL2, and CXCL8 in uncinate process mucosal samples from patients with CRSsNP compared with those from controls in Korea. Cho et al. [36] reported enhanced CXCL8 levels in nasal secretions from patients with CRSwNP compared with those from control subjects in America. Based on the unsupervised cluster analysis of protein levels of a number of cytokines and chemokines in CRS tissue lysates, Liao et al. [22] and Wei et al. [37] independently identified a neutrophil-dominated cluster associated with high levels of CXCL8 in patients in Wuhan and Sichuan city in China, respectively. These studies collectively implicate a critical role of CXCL family members in recruitment of neutrophils in CRS. However, the relevant contribution of different CXCL family members to the recruitment of neutrophils in different types of CRS need to be further clarified.

The interleukin (IL)-17 family of cytokines includes six members that play various roles in mucosal host defense and chronic inflammation [38]. Among them, IL-17A has been implicated in the recruitment and retention of mucosal neutrophils by promoting neutrophil-associated chemokine production, such as CXCL1 and CXCL8 [39,40]. Increased IL-17A levels have been demonstrated in Asian patients with CRSsNP or CRSwNP [4,6].

Wang et al. [6] found that IL-17A promoted IL-36 γ production, which can further induce CXCL1, CXCL2, and CXCL8 production in CRSwNP. In addition, Wang et al. [41] found that IL-17A significantly upregulated production of serum amyloid A, which is able to promote chemotaxis of neutrophils towards epithelium in CRSwNP. However, based on cluster analysis, Liao et al. [22] and Wei et al. [37] did not find any association between IL-17A levels and the number of tissue neutrophils. Saitoh et al. [42] and Makihara et al. [43] even found that the number of IL-17A-positive cells positively correlated with the number of eosinophils, but not neutrophils, in Japanese patients with CRSwNP. These discrepancies may reflect the pleiotropic activities of IL-17A in immune modulation in different inflammatory settings. For example, IL-17A has been reported to induce recruitment of eosinophils into airways in a murine model of asthma [44].

Thus more studies are needed to define the role of IL-17A in neutrophilic inflammation in CRS.

Lipid mediators are also strong inducers of neutrophil chemotaxis. Generated from arachidonic acid by 5 lipoxygenase, leukotriene B4 (LTB4), which is sensed by the LTB4 receptor on neutrophils, induces the migration of neutrophils [24]. Jeanson et al. [45] found that activation of unfolded protein response led to the secretion of LTB4 by nasal epithelial cells from patients with CRSwNP *in vitro*.

Colonized bacteria in nasal cavity in patients with CRS may induce the chemotaxis of neutrophils to local tissues. Rammal et al. [29] found that both gram-positive (*Staphylococcus aureus*) and gram-negative (*Pseudomonas aeruginosa*) bacteria stimulated CCL5 production from diseased nasal mucosal explants from CRS patients. It was also shown that *N*-formyl peptides,

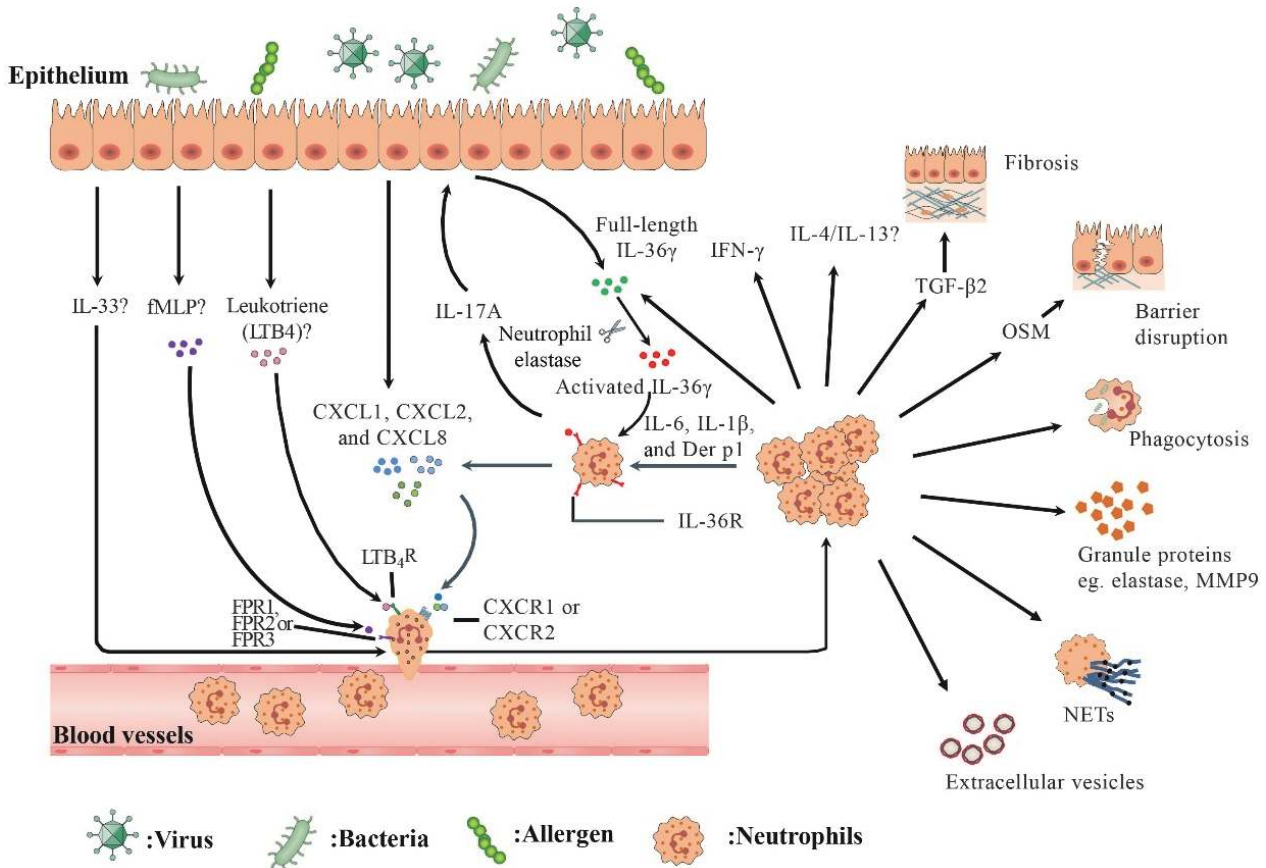


Fig. 1. An overview of recruitment and function of neutrophils in human chronic rhinosinusitis (CRS). In response to environmental stimuli, epithelial cells secrete chemokine (C-X-C motif) ligand (CXCL) family chemokines (eg., CXCL1, CXCL2, and CXCL8) and leukotrienes to recruit neutrophils by engaging chemokine (C-X-C motif) receptor (CXCR) 1/2 and leukotriene B4 receptor (LTB4R) on neutrophils, respectively. fMet-Leu-Phe (fMLP), which can be produced by colonized bacteria or damaged tissue cells, may recruit neutrophils via formyl-peptide receptor (FPR) 1/2/3. Interleukin (IL)-1 cytokine family such as IL-36 γ and IL-33 participate in regulating neutrophil inflammation in CRS. IL-17A upregulates the production of full-length IL-36 γ by epithelial cells. Activated IL-36 γ generated by neutrophil elastase processing acts on neutrophils via IL-36R, which is upregulated by IL-6, IL-1 β and *Dermatophagoides pteronyssinus* group (Der p) 1, and induces CXCL1, CXCL2, and CXCL8 production from neutrophils. Recruited neutrophils can kill bacteria by phagocytosis, degranulation, and formation of neutrophil extracellular traps (NETs). Activated neutrophils release extracellular vesicles. Neutrophils can also produce IL-17A, IL-36 γ , IL-4/13, interferon γ (IFN- γ), oncostatin M (OSM), and transforming growth factor- β 2 (TGF- β 2), which may be involved in the modulation of inflammation and tissue modeling in CRS.

such as fMet-Leu-Phe, derived from bacterial proteins activated human neutrophils and subsequently recruited them to inflammatory sites (Fig. 1) [26,46].

FUNCTION OF NEUTROPHILS IN INNATE IMMUNITY IN CRS

Currently, debate still exists on the role and profile of bacterial infection in the pathogenesis of CRS. Nevertheless, microbiota alteration has been demonstrated in patients with CRS by a number of studies [47,48]. For example, by means of conventional bacteria culture-dependent examination on nasal smear samples, Dobretsov et al. [49] reported increased colonization of *S. aureus* and *Staphylococcus epidermidis* in the nasal cavity of patients with CRSwNP compared with control subjects in western countries. Mahdavinia et al. [50] found that patients with CRS had significantly lower *Corynebacterium* species and *Peptoniphilus* species levels compared with those in control subjects by using high-throughput sequencing analysis of middle meatus swab samples. By metagenome analysis, Mahdavinia et al. [50] further found that lipopolysaccharide (LPS) biosynthesis proteins and bacterial invasion of epithelial cell pathways were significantly enhanced in patients with CRS. Bacteria prefer to exist in the form of biofilms and only 1% of all bacteria exist in the planktonic form [51]. Bacterial biofilms are a complex and organized community of bacteria encased in a self-produced exopolymeric matrix [52]. Bacterial biofilm formation may modify the inflammation process in human diseases [53]. Wang et al. [54] found that there was significantly higher number of neutrophils in biofilms-positive CRS patients compared with biofilms-negative counterparts. Neutrophils are essential for the clearance of bacteria either through phagocytosis or formation of neutrophil extracellular traps (NETs) [46].

Phagocytosis

Neutrophils are professional phagocytes. Following opsonization of the microbe and ligation of the respective opsonic receptor such as Fc γ receptors, C-type lectins, or complement receptors, neutrophils can be rapidly activated and engulf invading pathogens [46,55]. This phagocytosis process is orchestrated by a number of membrane proteins and complicated intracellular signaling cascades together with cytoskeletal rearrangements [46]. Production of oxygen radicals and their reaction products, which are collectively known as reactive oxygen species (ROS), is critical for the killing of the microbes in phagosomes [56]. However, some bacteria may impair the phagocytic capacity of neutrophils. *S. aureus*, which is frequently detected in Caucasian patients with CRSwNP, can inhibit phagocytosis and the subsequent killing of bacteria by neutrophils via multiple mechanisms [57]. Neutrophil phagocytosis can be regulated by toll-like receptor 2 ligand or inflammatory stimuli [58,59]. For ex-

ample, pretreatment with Pam3CSK4 (a toll-like receptor 2 agonist) enhanced the phagocytic function of neutrophils against methicillin-resistant *S. aureus* [58]. After exposing to bronchoalveolar lavage of children with neutrophil-predominant severe asthma, neutrophils exhibited greater phagocytic capability, which is possibly a result of regulation by local inflammatory milieu [59]. However, whether the phagocytic function of neutrophils is altered in CRS remains unexplored.

Neutrophil proteases

Neutrophils have at least three types of granules including azurophilic (primary), specific (secondary), and gelatinase (tertiary) granules [60]. Azurophilic granules are consisted of MPO, neutral protease cathepsin G, elastase and proteinase 3, and defensins. Specific granules contain antimicrobial peptide lactoferrin. Gelatinase granules contain gelatinase proteins such as matrix metalloproteinase 9 [60]. During phagocytosis, azurophilic and specific granules fuse with the phagosome and release antimicrobial contents and cooperatively hydrolyze microorganisms within phagosomes, thus playing an important role in killing pathogens [25]. During degranulation or exocytosis, these preformed mediators are released from the granules into the surrounding environment [60]. Neutrophil elastase activity was found increased in nasal polyp elutes from both eosinophilic and noneosinophilic CRSwNP as compared with that in elutes from control tissues, implying a potential role of neutrophil elastase in host defense in CRS [6].

Evidence also suggests that neutrophil proteases play an important role in the coordination and escalation of inflammatory reactions through the processing of the extracellular matrix, cytokines, chemokines, and receptors [61]. For example, neutrophil elastase is able to process multiple IL-1 family cytokines, including IL-1 α , IL-1 β , IL-33, IL-36 α , IL-36 β , and IL-36 γ [62]. Wang et al. [6] found that nasal polyp neutrophil elastase cleaved full-length IL-36 γ into activated IL-36 γ , highlighting the role of neutrophil proteases in the regulation of inflammation in CRS. Apart from elastase, neutrophil-derived MMP9 and MMP2 has been demonstrated elevated in both eosinophilic and noneosinophilic CRSwNP [6,16], although their function in CRS remains to be elucidated.

NET formation

In addition to phagocytosis and degranulation, neutrophils release their DNA in a netlike configuration to create traps known as NETs [46,63]. NETs are important for the sequestration and killing of bacteria, fungi and virus [64]. The release of nuclear content and the exteriorization of chromatin form a scaffold for the extracellular exposure of histones and antimicrobial proteins including neutrophil elastase, MPO, calprotectin, cathelicidins, and defensins [63]. NETs may kill microorganisms through the action of their antimicrobial components [64]. The formation of NETs can be triggered by microorganisms or endogenous stimu-

li, such as damage-associated molecular patterns and immune complexes, which has been extensively reviewed elsewhere [63]. Different functional attributes may activate different pathways to generate NETs [63]. Many stimuli, such as fungi and bacteria, activate MPO-elastase pathway leading to NETs formation. Under the stimulation, neutrophils generate ROS through the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase pathway [63]. ROS stimulates MPO to trigger the activation and translocation of elastase from azurophilic granules to the nucleus, where elastase proteolytically processes histones and disrupts chromatin packaging. Subsequently, MPO binds chromatin and synergizes with elastase to decondense chromatin, finally leading to the formation of NETs [63]. Nevertheless, some NET stimuli, such as immune complexes, ionomycin, and nicotine, may trigger NET formation independently of NADPH oxidase, relying instead on mitochondrial ROS [63]. Similar to elastase, the enzyme peptidyl arginine deiminase type 4 is critical for NET formation by regulating the initiation of DNA decondensation and nuclear envelope degradation; however, the underlying mechanisms require further investigations [46].

Currently, how neutrophils decide between phagocytosis and NET formation remains an open question. However, several studies have indicated that the pathogen attributes may be critical for this decision. Branzk et al. [65] found that neutrophils sensed the size of microbes and selectively released NETs in response to pathogens that are too large to be phagocytosed. Godridge et al. [66] found that distinct particle-associated cell-wall components, such as β -glucan, could selectively trigger a signaling cascade to initiate phagocytosis. McDonald et al. [67] found that although neutrophils were effective phagocytes by engulfing bacteria in static *in vitro*, upon sensing bacteria under flow condition in blood, they released their NETs into the vasculature to ensnare bacteria and to prevent dissemination.

Nevertheless, persistent NETs release can be detrimental to host. Recent studies have underscored pathogenic roles of NETs in a variety of chronic inflammatory conditions [63]. For example, elevated production of NETs has been demonstrated in the sputum of patients with cystic fibrosis and chronic obstructive pulmonary disease (COPD) and associated with disease severity [68]. Heretofore, there is few studies regarding the formation and function of NETs in CRS. Very recently, Cao et al. [69] found the formation of NETs in diseased nasal mucosa from patients with CRS. Tan et al. [70] found increased levels of autoantibodies against dsDNA, chromatin and histone in nasal polyps. It is interesting to explore whether the production of these autoantibodies is induced by relevant components in NETs in CRS in future.

Neutrophil extracellular vesicles

Extracellular vesicles (EVs) are membrane-derived vesicles that mediate intercellular communications [71]. In response to inflammatory stimuli during inflammatory processes, neutrophils

also produce EVs. On the basis of the generation mechanisms, neutrophil-derived EVs can be categorized into two subtypes [72]. Neutrophil-derived trails (NDTRs) are generated during neutrophil extravasation [72], whereas neutrophil-derived microvesicles (NDMV) are generated when neutrophils arrive at the local inflammatory sites. Although these two subtypes of neutrophil-derived EVs share some common features such as membrane components, receptors, and ligands, there are substantial differences between these two subtypes of neutrophil-derived EVs [72]. NDTRs particularly contain various chemokines including CCL2, CCL5, CXCL1, and CXCL12, which may be slowly released into the surrounding tissues [72]. NDMVs express granule-associated markers and contain granule proteins such as MPO, lactoferrin, elastase, MMP9, and proteinase 3 [72, 73]. NDTRs have been discovered in the airways of influenza-infected mice [74], and NDMVs have been found in the bronchoalveolar fluids of patients with pneumonia [75] and sepsis [76]. Recently, the functions of EVs have garnered increasing attentions. NDTRs have been shown to guide the migration of immune cells into the inflammatory sites as they release various chemokines [74], whereas NDMVs can mediate either proinflammatory or anti-inflammatory responses depending on distinct target cells [72]. For example, NDMVs promote the production of IL-6 and CXCL8 by endothelial cells, although they inhibit the production of IL-6 and IL-1 β and enhance anti-inflammatory cytokine transforming growth factor- β (TGF- β) expression by natural killer (NK) cells and macrophages [72]. A recent study by Genschmer et al. [77] presents provocative new evidence that EVs released by activated neutrophils were able to bypass α 1-antitrypsin, thus contributing to COPD. The authors [77] further showed that EVs were stable since EVs from COPD patients transferred the COPD phenotype to mice. However, more studies are needed to clarify the presence and stability of EVs and whether their degradation limits their pathogenic activity in pathological conditions including CRS (Fig. 1).

FUNCTION OF NEUTROPHILS IN INFLAMMATION AND TISSUE REMODELING IN CRS

Although known as innate immune cells specialized in the elimination of invading pathogens, a more complex role of neutrophils in modulating local inflammatory responses is emerging [25]. Activated neutrophils shape both innate and adaptive immune responses by producing a variety of biological mediators [78]. Peripheral blood neutrophils are known as short-lived leukocytes; however, under the control and regulation of local milieu, neutrophils in inflammatory sites may undergo profound changes with a prolonged lifespan and unique functional phenotype [63]. A recent study by Wang et al. [6] found that IL-36 receptor (IL-36R), which was undetectable on blood neutrophils,

was highly expressed on the tissue neutrophils in both CRSsNP and CRSwNP. IL-36 γ belongs to the recently identified IL-36 cytokine family, which is grouped into the IL-1 family. Binding to a specific IL-36 receptor complex composed of IL-36R and a co-receptor IL-1R accessory protein, IL-36 γ can lead to NF- κ B (nuclear factor kappa B) activation and subsequent IL-6 and CXCL8 production. The authors [6] further found that IL-36R expression on blood neutrophils could be up-regulated by IL-6, IL-1 β , and *Dermatophagoides pteronyssinus* group 1. IL-36 γ promoted the secretion of CXCL1, CXCL2, CXCL8, and IL-17A from tissue neutrophils in CRS [6], suggesting that tissue neutrophils have disease-specific functional and phenotypic characteristics in CRS [6]. These findings argue for the further comprehensive comparison of tissue and peripheral neutrophils to address the regulation of neutrophils in local inflammatory site in CRS.

As one of classic Th1 cytokines, interferon γ (IFN- γ) has a significant role not only in innate immunity against pathogens but also in adaptive immunity in orchestrating leukocyte attraction, maturation, and differentiation [79]. IFN- γ was reported to induce activated but insufficient autophagy in CRS, leading to p62-dependent apoptosis of nasal epithelial cells [80]. Enhanced IFN- γ expression has been shown in both CRSsNP and CRSwNP in Asian patients, and a more predominant elevation was found in CRSsNP compared to CRSwNP [6,7]. IFN- γ can be produced by various cell types in addition to CD4⁺ T helper cells. Very recently, Lee et al. [81] identified that in addition to T cells, NK cells, and B cells, neutrophils also had IFN- γ expression in both CRSsNP and CRSwNP. They found that neutrophils were the predominant immune cell type having IFN- γ expression in CRS, suggesting an important function of neutrophils in inflammatory process in CRS [81].

CRSsNP, and eosinophilic and noneosinophilic CRSwNP present distinct patterns of tissue remodeling [11]. CRSsNP is highly fibrotic and CRSwNP is edematous. Nevertheless, noneosinophilic CRSwNP is less edematous and more fibrotic in comparison with eosinophilic CRSwNP [16]. Shi et al. [16] found that neutrophils were the major source of TGF- β 2 in CRS. They found that the number of TGF- β 2-positive cells positively correlated with the number of myofibroblasts and the expression level of fibronectin in CRS. Therefore, neutrophils may contribute to the tissue fibrosis in CRS by producing TGF- β 2, particularly for CRSsNP and noneosinophilic CRSwNP.

Epithelial barrier dysfunction initiates and promotes the inflammatory and immune response in CRS [2]. Loss of barrier function makes the subepithelial components more susceptible to antigenic exposure and elicits immune response [2]. Pothoven et al. [18] found that neutrophils might disrupt nasal polyp epithelial barrier by producing oncostatin M (OSM). They demonstrated that neutrophils were a major source of OSM in CRSwNP, and granulocyte-macrophage colony stimulating factor (GM-CSF), but not IL-25, IL-33, or thymic stromal lymphopoietin, promot-

ed neutrophil OSM production [17]. Since neutrophil itself can produce GM-CSF, Pothoven et al. [18] implied an autocrine regulatory pathway for OSM secretion in CRSwNP (Fig. 1).

Interestingly, the authors [18] showed that OSM-producing neutrophils expressed the type 2 immune response marker arginase-1, suggesting that OSM-producing neutrophils have a N2 phenotype in CRSwNP. Some studies have shown that neutrophils were able to selectively produce type 2 cytokines including IL-4, IL-5, IL-9, and IL-13 following IL-33 and IL-25 challenge [82,83], indicating a pro-allergic role of neutrophils in certain conditions. Sun et al. [82] found that IL-33-treated neutrophils displayed a distinct expression profile of cytokine and chemokine receptors as compared with resting and LPS-treated neutrophils, such as IL-13R α 1, IL-R2, and CXCR5. Similar to M1/M2 macrophage classification, N1 and N2 neutrophil concept has also been proposed. N2 neutrophils express macrophage mannose receptor, arginase 1, chitinase-like 3, IL-10, and TGF- β and associate with tissue repair, whereas N1 neutrophils have an inflammatory phenotype expressing IL-1 β and IL-12 [84]. Pothoven et al. [18] discovered that GM-CSF and follistatin-like 1 were able to induce a shifting of neutrophils from a N1 phenotype to N2 phenotype. However, the N1/N2 classification has not been widely accepted and the presence of N1 and N2 neutrophils in CRS requires further confirmation.

REGULATION OF NEUTROPHILIC INFLAMMATION BY IL-1 CYTOKINE FAMILY IN CRS

Composed of several *bona fide* proinflammatory cytokines including IL-1 α , IL-1 β , IL-18, IL-33, IL-36 α , IL-36 β , and IL-36 γ , IL-1 family cytokines are instrumental as initiators and amplifiers of inflammation and are frequently among the first cytokines produced in response to infection or injury due to their constitutive expression in barrier tissues [85]. Kim et al. [86] found that the concentration of IL-1 β in tissue homogenates was enhanced in noneosinophilic CRSwNP compared with eosinophilic CRSwNP and controls. Based on the unsupervised cluster analysis of protein levels of cytokines and chemokines in CRS tissue lysates, Liao et al. [22] identified a neutrophil-dominated cluster associated with high levels of IL-1 β in tissue homogenates in patients in Wuhan, implicating its role in regulating neutrophilic inflammation in CRS.

The recently published study by Wang et al. [6] thoroughly explored the expression pattern of IL-36 family and its function in regulating neutrophilic inflammation in CRS. The authors [6] found that all IL-36 family members including IL-36 α , IL-36 β , IL-36 γ , IL-36Ra, and IL-38 as well as their receptor IL-36R were upregulated in all types of CRS, eosinophilic and non-eosinophilic CRSwNP and CRSsNP, in comparison with controls. Importantly, IL-36 γ was the most abundant isoform expressed and mainly

produced by epithelial cells in CRS [6]. The production of IL-36 γ in nasal epithelial cells was induced by double-stranded RNA, IL-1 β , tumor necrosis factor- α (TNF- α), and IL-17A [6]. They further identified that neutrophils were the principal IL-36R-positive cell type in polyps and found that IL-36 γ could induce the production of CXCL1, CXCL2, and CXCL8 from neutrophils in nasal polyps, which may lead to recruitment of neutrophils into nasal mucosa [6]. Interestingly, IL-36 γ could also induce IL-17A production from neutrophils and later IL-17A promoted IL-36 γ production from nasal epithelial cells, suggesting a positive feedback loop of IL-17A/IL-36 γ in CRS [6]. Full-length IL-36 γ was produced by nasal epithelial cells in full-length form and was cleaved into active form with hundreds-fold increased biological capacity by neutrophil-derived elastase in CRS [6]. Taken together, this study implicates a critical role for IL-36 γ in regulating neutrophilic inflammation in CRS.

Although IL-33 is well known for its function in initiation and exaggeration of type 2 inflammation by activating type 2 innate lymphoid cells, eosinophils, and Th2 cells [87], IL-33 may also have a role in recruiting neutrophils possibly by increasing CXCL1 production and preventing CXCR2 downregulation on neutrophils [88,89]. Kim et al. [90] found that IL-33 protein levels were elevated in noneosinophilic polyp tissues compared with eosinophilic polyp tissues. In addition, IL-33 levels were positively correlated with the levels of IL-1 β , TNF- α and IL-17A, and the number of tissue neutrophils [90]. Importantly, the authors [90] discovered that anti-IL-33 treatment reduced infiltration of neutrophils, but not eosinophils, in a murine model of nasal polyps induced by ovalbumin and staphylococcal enterotoxin B, which showed a mixed type-1/2/17 inflammatory response. Kim et al.'s study [90] highlights a previously unrecognized role for IL-33 beyond type 2 response and a novel mechanism underlying neutrophilic inflammation in CRSwNP. However, the expression of IL-33 in CRS remains controversial. Liao et al. [91] found that IL-33 expression was increased in the epithelial cells in both eosinophilic and noneosinophilic CRSwNP. Baba et al. [92] found that there was no difference in IL-33 levels between polyps and controls. Thus the results regrading IL-33 in CRS should be explained with caution (Fig. 1).

AGE AND NEUTROPHILIC INFLAMMATION IN CRS

Age-related differences in clinical presentation and neutrophilic immune signature has recently been noted in CRS; however, conflicting results were reported [93-95]. Kim et al. [93] reported that the number of human neutrophil elastase-positive neutrophils, *myeloperoxidase* gene expression, and neutrophil-associated cytokines including IL-17A and IL-23, and neutrophil chemokine receptor CXCR2 were decreased in patients older than 40 years as compared with young-aged (18-39 years) pa-

tients with noneosinophilic CRSwNP in Korean population. However, no age-related change of neutrophilic inflammation has been observed in control subjects and patients with CRSs-NP or eosinophilic CRSwNP [93]. Furthermore, the authors [93] showed that elderly patients with noneosinophilic CRSwNP have better objective surgical outcomes compared with the non-elderly. In contrast, Morse et al. [94] found that although aged and younger CRS patients had similar frequencies of nasal polyps and asthma, elderly patients with CRS (≥ 60 years) showed elevated expression of neutrophil associated mediators in mucus including IL-1 β , IL-6, CXCL8, IL-17A, and TNF- α compared with the younger counterparts (<60 years) in America. Increase in neutrophil associated mediators was linked with a higher prevalence of bacterial infection/colonization and a less likelihood to response to corticosteroids in aged patients. Cho et al. [95] found less ECP levels in nasal lavage samples in elderly subjects (60-77 years) compared with those in nonelderly subjects (16-59 years); however, there was no change of human neutrophil elastase levels and no correlation between neutrophil elastase levels and ages in patients with CRSwNP in America. These discrepancies may be due to the different techniques used to measure neutrophil-associated markers. Also, the inherited difference between the Caucasian and Asian patients could not be excluded.

THERAPEUTIC IMPLICATIONS

Eosinophilic inflammation in CRS, which is dominated by skewed type 2 response, is a relatively well-established endotype. In addition to corticosteroid therapy, biological therapies targeting IL-5, IL-4R, and immunoglobulin E are emerging for controlling eosinophilic inflammation in CRS [3]. In contrast, the neutrophilic inflammation in CRS remains very poorly defined.

Corticosteroids serve as a mainstay in CRS therapy; however, not all CRS patients respond well to corticosteroid treatment [21]. In Liao et al.'s cluster study [22], the authors identified a cluster of Chinese patients characterized by severe neutrophilic inflammation with high frequency of difficult-to-treat case despite the endoscopic sinus surgery and postoperative corticosteroid treatment, indicating a poor response of neutrophilic inflammation to corticosteroid treatment. Indeed, Wen et al. [21] found that oral prednisone was able to suppress type 2 cytokine expression and eosinophilic inflammation, whereas it could not suppress type 17 cytokine expression and neutrophilic inflammation in Chinese patients with CRSwNP. Increased neutrophilia in CRSwNP has been associated with the poor response to oral corticosteroid therapy in Chinese patients [21]. In contrast to its suppressive effect on eosinophils, pharmacological studies show that corticosteroids prevent apoptosis of neutrophils and promote the neutrophilic inflammation [96].

Beyond the well-known antimicrobial effect, macrolides have

intrinsic anti-inflammation and immunomodulation properties, especially for neutrophilic inflammation [97,98]. They block the activation of transcription factor nuclear factor κ B and inhibit the production of various inflammatory cytokines including CXCL8 and TNF- α [97,98]. Macrolides can also suppress the secretion of airway mucus, induce the apoptosis of neutrophils, and even diminish the formation of bacterial biofilms [97,98]. Luo et al. [99] found that long-term clarithromycin treatment inhibited CXCL8 and MPO levels in Chinese patients with CRSsNP and clarithromycin was more effective for CRSsNP patients with high CXCL8 levels. However, there are conflicting results regarding the efficacy of macrolides in CRS [1,100,101]. Videler et al. [101] reported that there was a lack of efficacy of long-term, low-dose azithromycin in patients with CRS; however, the authors [101] did not performed the subgroup analysis given the insufficient study power. The value of macrolide in CRS treatment, particularly for neutrophilic inflammation, requires to be established by more well-designed studies with larger sample size.

Blocking the trafficking or activation of the neutrophils may serve as a therapeutic strategy for neutrophilic inflammation in CRS, especially for CRSsNP and noneosinophilic CRSwNP. Strategies to inhibit neutrophil recruitment may include blockage of chemokine receptors (CXCR1 and CXCR2 antagonists) and inhibition of IL-36 γ , IL-17A, and LTB4. Several clinical trials assessed the effect of CXCR1 and CXCR2 antagonists and inhibitors on neutrophilic inflammation, and some have reported promising results. For example, clinical trials with a CXCR2 antagonist (MK 7123) in patients with COPD have shown that MK-7123 treatment resulted in decreased neutrophil counts and reduced MMP9 and MPO levels in plasma and sputum, and relieved disease symptoms [102]. Dual CXCR1/CXCR2 antagonist SCH527123 has been shown effective in reducing neutrophil levels in the circulation and airways in mild atopic asthmatic subjects [103]. Derived from omega-3 essential polyunsaturated fatty acids, resolvins inhibit neutrophil transmigration through downregulating neutrophil chemokines including CXCL1 and LTB4, making them a potential therapeutic candidate to suppressing neutrophilic inflammation [104]. Anti-IL-17A biologics (Brodalumab, ixekizumab, and secukinumab) have been approved by U.S. Food and Drug Administration for the treatment of psoriasis, a skin disorder characterized by enhanced neutrophil infiltration, and these agents substantially improve quality of life of patients with psoriasis [105]. However, the clinical efficacy of these agents for the treatment of CRSsNP and noneosinophilic CRSwNP remains to be studied in future.

CONCLUSION

Although increasing evidence highlight the importance of neutrophils in the pathogenesis of CRS, many aspects of neutrophilic

inflammation in CRS remain unknown. These include the association between microbiome and the function of neutrophils; the difference between tissue-specific neutrophils and their blood counterparts; the heterogeneity and plasticity of neutrophils in different types of CRS; and the formation and function of NETs in CRS. Elucidating these questions will improve our understanding of the intricate role of neutrophils during the development of CRS and possibly facilitate the development of innovative therapeutics.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

This work was supported by National Natural Science Foundation of China (NSFC; grants 81630024, 81570899, and 81325006), Natural Science Foundation of Hubei Province (grant 2017CFA 016), and “Ten thousand plan” - National high level talents special support plan of China.

ORCID

Hai Wang <https://orcid.org/0000-0003-0351-2151>
 Li Pan <https://orcid.org/0000-0003-2682-7500>
 Zheng Liu <https://orcid.org/0000-0002-4168-6702>

AUTHOR CONTRIBUTIONS

Conceptualization: ZL. Data curation: HW. Formal analysis: HW, LP. Funding acquisition: ZL. Methodology: HW. Project administration: ZL. Visualization: HW, ZL. Writing - original draft: HW. Writing - review & editing: all authors.

REFERENCES

1. Fokkens WJ, Lund VJ, Mullol J, Bachert C, Alobid I, Baroody F, et al. EPOS 2012: European position paper on rhinosinusitis and nasal polyps 2012: a summary for otorhinolaryngologists. *Rhinology*. 2012 Mar;50(1):1-12.
2. Schleimer RP. Immunopathogenesis of chronic rhinosinusitis and nasal polyposis. *Annu Rev Pathol*. 2017 Jan;12(1):331-57.
3. Bachert C, Zhang L, Gevaert P. Current and future treatment options for adult chronic rhinosinusitis: focus on nasal polyposis. *J Allergy Clin Immunol*. 2015 Dec;136(6):1431-40.
4. Wang X, Zhang N, Bo M, Holtappels G, Zheng M, Lou H, et al. Diversity of TH cytokine profiles in patients with chronic rhinosinus-

- itis: a multicenter study in Europe, Asia, and Oceania. *J Allergy Clin Immunol*. 2016 Nov;138(5):1344-53.
5. Huvenne W, van Bruaene N, Zhang N, van Zele T, Patou J, Gevaert P, et al. Chronic rhinosinusitis with and without nasal polyps: what is the difference? *Curr Allergy Asthma Rep*. 2009 May;9(3):213-20.
 6. Wang H, Li ZY, Jiang WX, Liao B, Zhai GT, Wang N, et al. The activation and function of IL-36 γ in neutrophilic inflammation in chronic rhinosinusitis. *J Allergy Clin Immunol*. 2018 May;141(5):1646-58.
 7. Cao PP, Li HB, Wang BF, Wang SB, You XJ, Cui YH, et al. Distinct immunopathologic characteristics of various types of chronic rhinosinusitis in adult Chinese. *J Allergy Clin Immunol*. 2009 Sep;124(3):478-84.
 8. Zhang Y, Gevaert E, Lou H, Wang X, Zhang L, Bachert C, et al. Chronic rhinosinusitis in Asia. *J Allergy Clin Immunol*. 2017 Nov;140(5):1230-9.
 9. Fan Y, Chen S, Qu X, Zuo K, Li X, Huang J, et al. A lower prevalence of asthma among patients with chronic rhinosinusitis in southern China. *J Allergy Clin Immunol*. 2011 Feb;127(2):520-2.e1-5.
 10. Zhang N, Van Zele T, Perez-Novo C, Van Bruaene N, Holtappels G, DeRuyck N, et al. Different types of T-effector cells orchestrate mucosal inflammation in chronic sinus disease. *J Allergy Clin Immunol*. 2008 Nov;122(5):961-8.
 11. Cao PP, Wang ZC, Schleimer RP, Liu Z. Pathophysiologic mechanisms of chronic rhinosinusitis and their roles in emerging disease endotypes. *Ann Allergy Asthma Immunol*. 2019 Jan;122(1):33-40.
 12. Cao PP, Zhang YN, Liao B, Ma J, Wang BF, Wang H, et al. Increased local IgE production induced by common aeroallergens and phenotypic alteration of mast cells in Chinese eosinophilic, but not non-eosinophilic, chronic rhinosinusitis with nasal polyps. *Clin Exp Allergy*. 2014 Mar;44(5):690-700.
 13. Payne SC, Early SB, Huyett P, Han JK, Borish L, Steinke JW. Evidence for distinct histologic profile of nasal polyps with and without eosinophilia. *Laryngoscope*. 2011 Oct;121(10):2262-7.
 14. Kim DK, Jin HR, Eun KM, Mutusamy S, Cho SH, Oh S, et al. Non-eosinophilic nasal polyps shows increased epithelial proliferation and localized disease pattern in the early stage. *PLoS One*. 2015 Oct;10(10):e0139945.
 15. Shi LL, Song J, Xiong P, Cao PP, Liao B, Ma J, et al. Disease-specific T-helper cell polarizing function of lesional dendritic cells in different types of chronic rhinosinusitis with nasal polyps. *Am J Respir Crit Care Med*. 2014 Sep;190(6):628-38.
 16. Shi LL, Xiong P, Zhang L, Cao PP, Liao B, Lu X, et al. Features of airway remodeling in different types of Chinese chronic rhinosinusitis are associated with inflammation patterns. *Allergy*. 2013 Jan;68(1):101-9.
 17. Ma J, Shi LL, Deng YK, Wang H, Cao PP, Long XB, et al. CD8(+) T cells with distinct cytokine-producing features and low cytotoxic activity in eosinophilic and non-eosinophilic chronic rhinosinusitis with nasal polyps. *Clin Exp Allergy*. 2016 Sep;46(9):1162-75.
 18. Pothoven KL, Norton JE, Suh LA, Carter RG, Harris KE, Biyasheva A, et al. Neutrophils are a major source of the epithelial barrier disrupting cytokine oncostatin M in patients with mucosal airways disease. *J Allergy Clin Immunol*. 2016 Jun;139(6):1966-78.e9.
 19. Bachert C, Zhang N, Holtappels G, De Lobel L, van Cauwenberge P, Liu S, et al. Presence of IL-5 protein and IgE antibodies to staphylococcal enterotoxins in nasal polyps is associated with comorbid asthma. *J Allergy Clin Immunol*. 2010 Nov;126(5):962-8.e1-6.
 20. Tomassen P, Vandeplas G, Van Zele T, Cardell LO, Arebro J, Olze H, et al. Inflammatory endotypes of chronic rhinosinusitis based on cluster analysis of biomarkers. *J Allergy Clin Immunol*. 2016 May;137(5):1449-56.e4.
 21. Wen W, Liu W, Zhang L, Bai J, Fan Y, Xia W, et al. Increased neutrophilia in nasal polyps reduces the response to oral corticosteroid therapy. *J Allergy Clin Immunol*. 2012 Jun;129(6):1522-8.e5.
 22. Liao B, Liu JX, Li ZY, Zhen Z, Cao PP, Yao Y, et al. Multidimensional endotypes of chronic rhinosinusitis and their association with treatment outcomes. *Allergy*. 2018 Jul;73(7):1459-69.
 23. Soehnlein O, Steffens S, Hidalgo A, Weber C. Neutrophils as protagonists and targets in chronic inflammation. *Nat Rev Immunol*. 2017 Apr;17(4):248-61.
 24. Coffelt SB, Wellenstein MD, de Visser KE. Neutrophils in cancer: neutral no more. *Nat Rev Cancer*. 2016 Jul;16(7):431-46.
 25. Ley K, Hoffman HM, Kubes P, Cassatella MA, Zychlinsky A, Hedrick CC, et al. Neutrophils: new insights and open questions. *Sci Immunol*. 2018 Dec;3(30):eaat4579.
 26. de Oliveira S, Rosowski EE, Huttenlocher A. Neutrophil migration in infection and wound repair: going forward in reverse. *Nat Rev Immunol*. 2016 May;16(6):378-91.
 27. Shiozawa A, Miwa M, Ono N, Homma H, Hirotsu M, Ikeda K. Comparative analysis of cytokine release from epithelial cell cultures of the upper airway. *Rhinology*. 2015 Jun;53(2):135-41.
 28. Shimizu S, Kouzaki H, Kato T, Tojima I, Shimizu T. HMGB1-TLR4 signaling contributes to the secretion of interleukin 6 and interleukin 8 by nasal epithelial cells. *Am J Rhinol Allergy*. 2016 May;30(3):167-72.
 29. Rammal A, Tewfik M, Rousseau S. Differences in RANTES and IL-6 levels among chronic rhinosinusitis patients with predominant gram-negative and gram-positive infection. *J Otolaryngol Head Neck Surg*. 2017 Jan;46(1):7.
 30. Ozturk AB, Bayraktar R, Gogebakan B, Mumbuc S, Bayram H. Comparison of inflammatory cytokine release from nasal epithelial cells of non-atopic non-rhinitic, allergic rhinitic and polyp subjects and effects of diesel exhaust particles in vitro. *Allergol Immunopathol (Madr)*. 2017 Sep-Oct;45(5):473-81.
 31. Tsai YJ, Hao SP, Chen CL, Wu WB. Thromboxane A2 regulates CXCL1 and CXCL8 chemokine expression in the nasal mucosa-derived fibroblasts of chronic rhinosinusitis patients. *PLoS One*. 2016 Jun;11(6):e0158438.
 32. Shimizu S, Tojima I, Takezawa K, Matsumoto K, Kouzaki H, Shimizu T. Thrombin and activated coagulation factor X stimulate the release of cytokines and fibronectin from nasal polyp fibroblasts via protease-activated receptors. *Am J Rhinol Allergy*. 2017 Jan;31(1):13-8.
 33. Zhai GT, Wang H, Li JX, Cao PP, Jiang WX, Song J, et al. IgD-activated mast cells induce IgE synthesis in B cells in nasal polyps. *J Allergy Clin Immunol*. 2018 Nov;142(5):1489-99.e23.
 34. Van Zele T, Claeys S, Gevaert P, Van Maele G, Holtappels G, Van Cauwenberge P, et al. Differentiation of chronic sinus diseases by measurement of inflammatory mediators. *Allergy*. 2006 Nov;61(11):1280-9.
 35. Kim DW, Eun KM, Roh EY, Shin S, Kim DK. Chronic rhinosinusitis without nasal polyps in Asian patients shows mixed inflammatory patterns and neutrophil-related disease severity. *Mediators Inflamm*. 2019 Jan;2019:7138643.
 36. Cho DY, Nayak JV, Bravo DT, Le W, Nguyen A, Edward JA, et al. Expression of dual oxidases and secreted cytokines in chronic rhinosinusitis. *Int Forum Allergy Rhinol*. 2013 May;3(5):376-83.
 37. Wei B, Liu F, Zhang J, Liu Y, Du J, Liu S, et al. Multivariate analysis of inflammatory endotypes in recurrent nasal polyposis in a Chinese population. *Rhinology*. 2018 Sep;56(3):216-26.
 38. Weaver CT, Hatton RD, Mangan PR, Harrington LE. IL-17 family cytokines and the expanding diversity of effector T cell lineages. *Annu Rev Immunol*. 2007;25(1):821-52.
 39. Pridgeon C, Bugeon L, Donnelly L, Straschil U, Tudhope SJ, Fenwick P, et al. Regulation of IL-17 in chronic inflammation in the human lung. *Clin Sci (Lond)*. 2011 Jun;120(12):515-24.
 40. Niu YZ, Gong GQ, Chen S, Chen JJ, Kong WJ, Wang YJ. Effects of IL-17 on expression of GRO- α and IL-8 in fibroblasts from nasal

- polyps. *J Huazhong Univ Sci Technolog Med Sci.* 2014 Aug;34(4):591-5.
41. Wang H, Bai J, Ding M, Liu W, Xu R, Zhang J, et al. Interleukin-17A contributes to the expression of serum amyloid A in chronic rhinosinusitis with nasal polyps. *Eur Arch Otorhinolaryngol.* 2013 May; 270(6):1867-72.
 42. Saitoh T, Kusunoki T, Yao T, Kawano K, Kojima Y, Miyahara K, et al. Role of interleukin-17A in the eosinophil accumulation and mucosal remodeling in chronic rhinosinusitis with nasal polyps associated with asthma. *Int Arch Allergy Immunol.* 2010;151(1):8-16.
 43. Makihara S, Okano M, Fujiwara T, Kariya S, Noda Y, Higaki T, et al. Regulation and characterization of IL-17A expression in patients with chronic rhinosinusitis and its relationship with eosinophilic inflammation. *J Allergy Clin Immunol.* 2010 Aug;126(2):397-400. e1-11.
 44. Wakashin H, Hirose K, Maezawa Y, Kagami S, Suto A, Watanabe N, et al. IL-23 and Th17 cells enhance Th2-cell-mediated eosinophilic airway inflammation in mice. *Am J Respir Crit Care Med.* 2008 Nov;178(10):1023-32.
 45. Jeanson L, Kelly M, Coste A, Guerrero IC, Fritsch J, Nguyen-Khoa T, et al. Oxidative stress induces unfolding protein response and inflammation in nasal polyposis. *Allergy.* 2012 Mar;67(3):403-12.
 46. Liew PX, Kubes P. The neutrophil's role during health and disease. *Physiol Rev.* 2019 Apr;99(2):1223-48.
 47. Hamilos DL. Host-microbial interactions in patients with chronic rhinosinusitis. *J Allergy Clin Immunol.* 2014 Mar;133(3):640-53.e4.
 48. Mahdavinia M, Keshavarzian A, Tobin MC, Landay AL, Schleimer RP. A comprehensive review of the nasal microbiome in chronic rhinosinusitis (CRS). *Clin Exp Allergy.* 2016 Jan;46(1):21-41.
 49. Dobretsov K, Negm H, Ralli M, Passali D. The theory of a "staphylococcus superantigen" in chronic rhinosinusitis with nasal polyps: myth or reality? *Eur Rev Med Pharmacol Sci.* 2019 Mar;23(1 Suppl):48-54.
 50. Mahdavinia M, Engen PA, LoSavio PS, Naqib A, Khan RJ, Tobin MC, et al. The nasal microbiome in patients with chronic rhinosinusitis: analyzing the effects of atopy and bacterial functional pathways in 111 patients. *J Allergy Clin Immunol.* 2018 Jul;142(1):287-90.e4.
 51. Sanclement JA, Webster P, Thomas J, Ramadan HH. Bacterial biofilms in surgical specimens of patients with chronic rhinosinusitis. *Laryngoscope.* 2005 Apr;115(4):578-82.
 52. Donlan RM, Costerton JW. Biofilms: survival mechanisms of clinically relevant microorganisms. *Clin Microbiol Rev.* 2002 Apr;15(2):167-93.
 53. Galli J, Calo L, Ardito F, Imperiali M, Bassotti E, Passali GC, et al. Damage to ciliated epithelium in chronic rhinosinusitis: what is the role of bacterial biofilms? *Ann Otol Rhinol Laryngol.* 2008 Dec; 117(12):902-8.
 54. Wang X, Du J, Zhao C. Bacterial biofilms are associated with inflammatory cells infiltration and the innate immunity in chronic rhinosinusitis with or without nasal polyps. *Inflammation.* 2014 Jun; 37(3):871-9.
 55. Dale DC, Boxer L, Liles WC. The phagocytes: neutrophils and monocytes. *Blood.* 2008 Aug;112(4):935-45.
 56. Segal AW. How neutrophils kill microbes. *Annu Rev Immunol.* 2005 Apr;23(1):197-223.
 57. Nasser A, Moradi M, Jazireian P, Safari H, Alizadeh-Sani M, Pourmand MR, et al. *Staphylococcus aureus* versus neutrophil: scrutiny of ancient combat. *Microb Pathog.* 2019 Jun;131:259-69.
 58. Chen Y, Lu S, Zhang Y, Yu J, Deng L, Chen H, et al. TLR2 agonist Pam3CSK4 enhances the antibacterial functions of GM-CSF induced neutrophils to methicillin-resistant *Staphylococcus aureus*. *Microb Pathog.* 2019 May;130:204-12.
 59. Grunwell JR, Stephenson ST, Tirouvanziam R, Brown LA, Brown MR, Fitzpatrick AM. Children with neutrophil-predominant severe asthma have proinflammatory neutrophils with enhanced survival and impaired clearance. *J Allergy Clin Immunol. Pract.* 2019 Feb; 7(2):516-25.e6.
 60. Sheshachalam A, Srivastava N, Mitchell T, Lacy P, Eitzen G. Granule protein processing and regulated secretion in neutrophils. *Front Immunol.* 2014 Sep;5:448.
 61. Korkmaz B, Horwitz MS, Jenne DE, Gauthier F. Neutrophil elastase, proteinase 3, and cathepsin G as therapeutic targets in human diseases. *Pharmacol Rev.* 2010 Dec;62(4):726-59.
 62. Clancy DM, Sullivan GP, Moran HB, Henry CM, Reeves EP, McElvaney NG, et al. Extracellular neutrophil proteases are efficient regulators of IL-1, IL-33, and IL-36 cytokine activity but poor effectors of microbial killing. *Cell Rep.* 2018 Mar;22(11):2937-50.
 63. Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol.* 2018 Feb;18(2):134-47.
 64. Castanheira FV, Kubes P. Neutrophils and NETs in modulating acute and chronic inflammation. *Blood.* 2019 May;133(20):2178-85.
 65. Branzk N, Lubojemska A, Hardison SE, Wang Q, Gutierrez MG, Brown GD, et al. Neutrophils sense microbe size and selectively release neutrophil extracellular traps in response to large pathogens. *Nat Immunol.* 2014 Nov;15(11):1017-25.
 66. Goodridge HS, Reyes CN, Becker CA, Katsumoto TR, Ma J, Wolf AJ, et al. Activation of the innate immune receptor Dectin-1 upon formation of a 'phagocytic synapse'. *Nature.* 2011 Apr;472(7344):471-5.
 67. McDonald B, Urrutia R, Yipp BG, Jenne CN, Kubes P. Intravascular neutrophil extracellular traps capture bacteria from the bloodstream during sepsis. *Cell Host Microbe.* 2012 Sep;12(3):324-33.
 68. Wright TK, Gibson PG, Simpson JL, McDonald VM, Wood LG, Baines KJ. Neutrophil extracellular traps are associated with inflammation in chronic airway disease. *Respirology.* 2016 Apr;21(3):467-75.
 69. Cao Y, Chen F, Sun Y, Hong H, Wen Y, Lai Y, et al. LL-37 promotes neutrophil extracellular trap formation in chronic rhinosinusitis with nasal polyps. *Clin Exp Allergy.* 2019 Jul;49(7):990-9.
 70. Tan BK, Li QZ, Suh L, Kato A, Conley DB, Chandra RK, et al. Evidence for intranasal antinuclear autoantibodies in patients with chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol.* 2011 Dec;128(6):1198-206.e1.
 71. Thery C, Ostrowski M, Segura E. Membrane vesicles as conveyors of immune responses. *Nat Rev Immunol.* 2009 Aug;9(8):581-93.
 72. Hong CW. Extracellular vesicles of neutrophils. *Immune Netw.* 2018 Dec;18(6):e43.
 73. Dalli J, Montero-Melendez T, Norling LV, Yin X, Hinds C, Haskard D, et al. Heterogeneity in neutrophil microparticles reveals distinct proteome and functional properties. *Mol Cell Proteomics.* 2013 Aug;12(8):2205-19.
 74. Lim K, Hyun YM, Lambert-Emo K, Capece T, Bae S, Miller R, et al. Neutrophil trails guide influenza-specific CD8+ T cells in the airways. *Science.* 2015 Sep;349(6252):aaa4352.
 75. Hess C, Sadallah S, Hefti A, Landmann R, Schifferli JA. Ectosomes released by human neutrophils are specialized functional units. *J Immunol.* 1999 Oct;163(8):4564-73.
 76. Prakash PS, Caldwell CC, Lentsch AB, Pritts TA, Robinson BR. Human microparticles generated during sepsis in patients with critical illness are neutrophil-derived and modulate the immune response. *J Trauma Acute Care Surg.* 2012 Aug;73(2):401-6.
 77. Genschmer KR, Russell DW, Lal C, Szul T, Bratcher PE, Noerager BD, et al. Activated PMN exosomes: pathogenic entities causing matrix destruction and disease in the lung. *Cell.* 2019 Jan;176:113-26.e15.
 78. Mantovani A, Cassatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity.

- Nat Rev Immunol. 2011 Jul;11(8):519-31.
79. Plataniias LC. Mechanisms of type-I- and type-II-interferon-mediated signalling. *Nat Rev Immunol.* 2005 May;5(5):375-86.
 80. Wang BF, Cao PP, Wang ZC, Li ZY, Wang ZZ, Ma J, et al. Interferon- γ -induced insufficient autophagy contributes to p62-dependent apoptosis of epithelial cells in chronic rhinosinusitis with nasal polyps. *Allergy.* 2017 Sep;72(9):1384-97.
 81. Lee M, Kim DW, Khalmuratova R, Shin SH, Kim YM, Han DH, et al. The IFN- γ -p38, ERK kinase axis exacerbates neutrophilic chronic rhinosinusitis by inducing the epithelial-to-mesenchymal transition. *Mucosal Immunol.* 2019 May;12(3):601-11.
 82. Sun B, Zhu L, Tao Y, Sun HX, Li Y, Wang P, et al. Characterization and allergic role of IL-33-induced neutrophil polarization. *Cell Mol Immunol.* 2018 Aug;15(8):782-93.
 83. Petersen BC, Budelsky AL, Baptist AP, Schaller MA, Lukacs NW. Interleukin-25 induces type 2 cytokine production in a steroid-resistant interleukin-17RB+ myeloid population that exacerbates asthmatic pathology. *Nat Med.* 2012 May;18(5):751-8.
 84. Ma Y, Yabluchanskiy A, Iyer RP, Cannon PL, Flynn ER, Jung M, et al. Temporal neutrophil polarization following myocardial infarction. *Cardiovasc Res.* 2016 May;110(1):51-61.
 85. Martin SJ. Cell death and inflammation: the case for IL-1 family cytokines as the canonical DAMPs of the immune system. *FEBS J.* 2016 Jul;283(14):2599-615.
 86. Kim DK, Eun KM, Kim MK, Cho D, Han SA, Han SY, et al. Comparison between signature cytokines of nasal tissues in subtypes of chronic rhinosinusitis. *Allergy Asthma Immunol Res.* 2019 Mar;11(2):201-11.
 87. Cayrol C, Girard JP. Interleukin-33 (IL-33): a nuclear cytokine from the IL-1 family. *Immunol Rev.* 2018 Jan;281(1):154-68.
 88. Hueber AJ, Alves-Filho JC, Asquith DL, Michels C, Millar NL, Reilly JH, et al. IL-33 induces skin inflammation with mast cell and neutrophil activation. *Eur J Immunol.* 2011 Aug;41(8):2229-37.
 89. Alves-Filho JC, Sonogo F, Souto FO, Freitas A, Verri WA Jr, Auxiliadora-Martins M, et al. Interleukin-33 attenuates sepsis by enhancing neutrophil influx to the site of infection. *Nat Med.* 2010 Jun;16(6):708-12.
 90. Kim DK, Jin HR, Eun KM, Mo JH, Cho SH, Oh S, et al. The role of interleukin-33 in chronic rhinosinusitis. *Thorax.* 2017 Jul;72(7):635-45.
 91. Liao B, Cao PP, Zeng M, Zhen Z, Wang H, Zhang YN, et al. Interaction of thymic stromal lymphopoietin, IL-33, and their receptors in epithelial cells in eosinophilic chronic rhinosinusitis with nasal polyps. *Allergy.* 2015 Sep;70(9):1169-80.
 92. Baba S, Kondo K, Kanaya K, Suzukawa K, Ushio M, Urata S, et al. Expression of IL-33 and its receptor ST2 in chronic rhinosinusitis with nasal polyps. *Laryngoscope.* 2014 Apr;124(4):E115-22.
 93. Kim DW, Kim DK, Jo A, Jin HR, Eun KM, Mo JH, et al. Age-related decline of neutrophilic inflammation is associated with better post-operative prognosis in non-eosinophilic nasal polyps. *PLoS One.* 2016 Feb;11(2):e0148442.
 94. Morse JC, Li P, Ely KA, Shilts MH, Wannemuehler TJ, Huang LC, et al. Chronic rhinosinusitis in elderly patients is associated with an exaggerated neutrophilic proinflammatory response to pathogenic bacteria. *J Allergy Clin Immunol.* 2019 Mar;143(3):990-1002.e6.
 95. Cho SH, Hong SJ, Han B, Lee SH, Suh L, Norton J, et al. Age-related differences in the pathogenesis of chronic rhinosinusitis. *J Allergy Clin Immunol.* 2012 Mar;129(3):858-60.e2.
 96. Cox G. Glucocorticoid treatment inhibits apoptosis in human neutrophils: separation of survival and activation outcomes. *J Immunol.* 1995 May;154(9):4719-25.
 97. Tamaoki J. The effects of macrolides on inflammatory cells. *Chest.* 2004 Feb;125(2 Suppl):41S-50S.
 98. Tauber SC, Nau R. Immunomodulatory properties of antibiotics. *Curr Mol Pharmacol.* 2008 Jan;1(1):68-79.
 99. Luo Q, Chen F, Liu W, Li Z, Xu R, Fan Y, et al. Evaluation of long-term clarithromycin treatment in adult Chinese patients with chronic rhinosinusitis without nasal polyps. *ORL J Otorhinolaryngol Relat Spec.* 2011;73(4):206-11.
 100. Wallwork B, Coman W, Mackay-Sim A, Greiff L, Cervin A. A double-blind, randomized, placebo-controlled trial of macrolide in the treatment of chronic rhinosinusitis. *Laryngoscope.* 2006 Feb;116(2):189-93.
 101. Videler WJ, Badia L, Harvey RJ, Gane S, Georgalas C, van der Meulen FW, et al. Lack of efficacy of long-term, low-dose azithromycin in chronic rhinosinusitis: a randomized controlled trial. *Allergy.* 2011 Nov;66(11):1457-68.
 102. Rennard SI, Dale DC, Donohue JF, Kannies F, Magnussen H, Sutherland ER, et al. CXCR2 antagonist MK-7123: a phase 2 proof-of-concept trial for chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2015 May;191(9):1001-11.
 103. Todd CM, Salter BM, Murphy DM, Watson RM, Howie KJ, Milot J, et al. The effects of a CXCR1/CXCR2 antagonist on neutrophil migration in mild atopic asthmatic subjects. *Pulm Pharmacol Ther.* 2016 Dec;41:34-9.
 104. Dalli J, Winkler JW, Colas RA, Amardottir H, Cheng CY, Chiang N, et al. Resolvin D3 and aspirin-triggered resolvin D3 are potent immunoresolvents. *Chem Biol.* 2013 Feb;20(2):188-201.
 105. Natsis NE, Gottlieb AB. Bimekizumab for the treatment of psoriatic disease. *Expert Opin Biol Ther.* 2018 Dec;18(12):1193-7.