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New global observations of the terrestrial carbon cycle from GOSAT: Patterns of plant fluorescence with gross primary productivity

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Our ability to close the Earth’s carbon budget and predict feedbacks in a warming climate depends critically on knowing where, when and how carbon dioxide is exchanged between the land and atmosphere. Terrestrial gross primary production (GPP) constitutes the largest flux component in the global carbon budget, however significant uncertainties remain in GPP estimates and its seasonality. Empirically, we show that global spaceborne observations of solar induced chlorophyll fluorescence – occurring during photosynthesis – exhibit a strong linear correlation with GPP. We found that the fluorescence emission even without any additional climatic or model information has the same or better predictive skill in estimating GPP as those derived from traditional remotely-sensed vegetation indices using ancillary data and model assumptions. In boreal summer the generally strong linear relationship between fluorescence and GPP models weakens, attributable to discrepancies in savannas/croplands (18–48% higher fluorescence-based GPP derived by simple linear scaling), and high-latitude needleleaf forests (28–32% lower fluorescence). Our results demonstrate that retrievals of chlorophyll fluorescence provide direct global observational constraints for GPP and open an entirely new viewpoint on the global carbon cycle. Citation: Frankenberg, C., et al. (2011), New global observations of the terrestrial carbon cycle from GOSAT: Patterns of plant fluorescence with gross primary productivity, Geophys. Res. Lett., 38, L17706, doi:10.1029/2011GL048738.

Gross primary production (GPP) through photosynthesis by terrestrial ecosystems constitutes the largest global land carbon flux [Zhao and Running, 2010; Beer et al., 2010]. Currently there are two main spatially explicit approaches to quantify GPP globally: 1) meteorology-driven full land surface carbon cycle models [Friedlingstein et al., 2006; Sitch et al., 2008]; and, 2) remote sensing-driven [Zhao and Running, 2010] and/or flux tower based [Beer et al., 2010; Jung et al., 2011] semi-empirical models focused on GPP or net primary production (NPP). Significant uncertainties related with the first approach are due to differing model sensitivities to meteorological parameters and uncertain global meteorological data sets [Friedlingstein et al., 2006; Sitch et al., 2008]. Uncertainties with the second approach exist because GPP cannot directly be estimated from the remote sensing measurements but is also modeled as a function of leaf area index (LAI) and fraction of absorbed photosynthetically active radiation (fAPAR) or greenness indices such as the normalized difference or enhanced vegetation indices (NDVI, EVI) [Zhao et al., 2005]. These indices are often contaminated by atmospheric interference, and may contribute a misleading signal when vegetation becomes stressed, e.g., green canopies that are not photosynthesizing [Huete et al., 2002].

Remote sensing of solar-induced chlorophyll fluorescence (F_s) [Krause and Weis, 1991], as intended with the FLEX satellite mission, offers a direct physiology-based measure of global photosynthetic activity. Absorbed photosynthetically active radiation (PAR) within 400–700 nm wavelengths drives photosynthesis, but can also be dissipated into heat or re-radiated at longer wavelengths (660–800 nm), which is termed fluorescence. At the laboratory and field scale, chlorophyll fluorescence has been intensively studied [Moya et al., 2004; Corp et al., 2006; Baker, 2008; Campbell et al., 2008; Genty et al., 1989] but spaceborne remote sensing of fluorescence is more difficult [Frankenberg et al., 2011] and accurate data has so far not been available. Concerning solar-induced steady state fluorescence, an implicit direct correlation with GPP exists as both depend on absorbed radiation. Also, field studies [Flexas et al., 2002; Damm et al., 2010; Rascher et al., 2009] as well as theoretical modelling [Van der Tol et al., 2009] show a clear positive correlation of CO2 assimilation and stomatal conductance with F_s, especially because increases in heat dissipation under high light conditions cause a consequent reduction of both fluorescence and photosynthesis yield. These physiological signals provided by fluorescence are not directly achievable with traditional vegetation remote sensing products, which model GPP using a multitude of ancillary data and model assumptions, all of which are prone to errors. Especially light use efficiency (LUE) is difficult to model on a global scale as it
Figure 1. (a) Annual average (June 2009 through May 2010) of retrieved chlorophyll-a fluorescence at 755 nm on a 2° x 2° grid. Only grid-boxes with more than 15 soundings constituting the average are displayed. (b) Latitudinal monthly averages of chlorophyll fluorescence from June 2009 through end of August 2010.
GPP ($r^2 = 0.74$), but significantly worse correlations against the other MODIS vegetation index products ($r^2 = 0.47–0.63$) and the CASA model ($r^2 = 0.52$) (Figure 2). Two biome types caused most of the differences in the comparisons: needleleaf forest for MPI-BGC and MODIS, and evergreen broadleaf forest for CASA. The MODIS greenness indices showed saturation at high values, particularly in high northern latitude needleleaf forests; this may be attributed to problems with using greenness as an indicator for photosynthetic activity. This becomes evident in the correlation of vegetation indices with $F_s$, where the relationship appears curvilinear and needleleaf forests deviate most strongly regarding all indices, especially at low temperatures (Figure 2). Calculation of GPP from vegetation indices thus requires ancillary information, which can add further uncertainties. It is important to note that the chlorophyll

Figure 2. (top) Scatter-plot of 4° × 4° grid cell averages of fluorescence ($F_s$) vs. GPP model estimates (small dots color-coded by latitude, only grid boxes over vegetated areas and with a 1-s precision error in $F_s$ of <0.04 Wm$^{-2}$ µm$^{-1}$ sr$^{-1}$ are shown). The linear regression line in all panels equals a linear fit through the origin on the basis of the MPI-BGC GPP model. (bottom) Normalized $F_s/cos(SZA)$ vs. MODIS LAI, NDVI and fPAR. The large symbols in all plots are biome averages, further separated for northern and southern hemisphere and based on 1x1° biome classification see auxiliary material.

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Table 1. Linear Pearson Correlation Coefficients ($r^2$) With Chlorophyll Fluorescence on 4° × 4° Grid Cells for the Annual Average and Different Seasons

<table>
<thead>
<tr>
<th>Season</th>
<th>MPI-BGC GPP</th>
<th>MODIS GPP</th>
<th>CASA GPP</th>
<th>MODIS LAI</th>
<th>MODIS NDVI</th>
<th>MODIS fPAR</th>
<th>MODIS MPI GPP</th>
<th>CASA MPI GPP</th>
</tr>
</thead>
<tbody>
<tr>
<td>JJA</td>
<td>0.76</td>
<td>0.56</td>
<td>0.57</td>
<td>0.53</td>
<td>0.48</td>
<td>0.44</td>
<td>0.82</td>
<td>0.74</td>
</tr>
<tr>
<td>SON</td>
<td>0.86</td>
<td>0.78</td>
<td>0.64</td>
<td>0.73</td>
<td>0.64</td>
<td>0.63</td>
<td>0.87</td>
<td>0.80</td>
</tr>
<tr>
<td>DJF</td>
<td>0.88</td>
<td>0.76</td>
<td>0.77</td>
<td>0.63</td>
<td>0.63</td>
<td>0.61</td>
<td>0.87</td>
<td>0.80</td>
</tr>
<tr>
<td>MAM</td>
<td>0.81</td>
<td>0.72</td>
<td>0.64</td>
<td>0.63</td>
<td>0.51</td>
<td>0.53</td>
<td>0.86</td>
<td>0.77</td>
</tr>
<tr>
<td>Annual</td>
<td>0.80</td>
<td>0.74</td>
<td>0.52</td>
<td>0.64</td>
<td>0.46</td>
<td>0.46</td>
<td>0.81</td>
<td>0.63</td>
</tr>
<tr>
<td>JJA-DJF</td>
<td>0.89</td>
<td>0.65</td>
<td>0.72</td>
<td>0.70</td>
<td>0.53</td>
<td>0.80</td>
<td>0.78</td>
<td>0.86</td>
</tr>
</tbody>
</table>

*Seasons: June–August 2009 (JJA), September–November 2009 (SON), December–February 2009–2010 (DJF) and March–May 2010 (MAM). Vegetation-free areas are excluded in the analysis. In addition, the correlation of the difference between JJA and DJF is displayed (JJA-DJF, see Figure S12 in Text S1). The two right columns indicate the correlation coefficients of MODIS against MPI-BGC and CASA against MPI-BGC, respectively.*
fluorescence emission is the only dataset not sharing any information with all other datasets used here. In comparison with CASA, evergreen broadleaf forests are consistently low-biased against all other measurements, probably because LUE in CASA is only a function of climatic parameters [Potter et al., 1993].

Goodness of fit with the comparison products is not consistent seasonally. High $r^2$ with MPI-BGC GPP is observed in boreal autumn (SON) and winter (DJF) but is largely reduced in boreal summer (JJA) in all models (Table 1), most notably for MODIS and CASA GPP. Correlation of the raw fluorescence signal with MPI-BGC is as good as MODIS GPP with MPI-BGC, even though no interpretative model has yet been applied to the fluorescence data. For the seasonal amplitude (difference JJA-DJF), the correlation is significantly greater ($r^2 = 0.89$) than for MODIS GPP ($r^2 = 0.78$), which underestimates the seasonal variability especially in the southern hemisphere (see also Figure S12 in Text S1). The seasonal variability in GPP is of prime interest because a) systematic seasonal biases in models or vegetation indices may cancel out in the annual mean [Turner et al., 2006] and b) seasonal variability in GPP largely determines the seasonal cycle of atmospheric CO$_2$ abundances. For all seasons, correlation is best with MPI-BGC GPP, underlining that chlorophyll fluorescence provides direct constraints on the timing and amplitude of GPP.

With the exception of CASA, the latitudinal cross-sections of fluorescence and model GPP, especially with MPI-BGC, agree well in almost all seasons (Figure 3). The fluorescence latitudinal distribution and change in time are mostly within the uncertainty range of MPI-BGC, with two notable exceptions during JJA, causing the correlation deterioration. First, the fluorescence is elevated between 10–40°N. Second, the fluorescence signal in the northernmost latitudes from 55–70°N is much lower, exhibiting a decline further south than the models. The discrepancy at 10–40°N is mostly due to African savannas and croplands in Asia which constitute 38% of total global GPP [Beer et al., 2010]. The fluorescence signal in the northernmost latitudes from 55–70°N is much lower, exhibiting a decline further south than the models. The discrepancy at 10–40°N is mostly due to African savannas and croplands in Asia which constitute 38% of total global GPP [Beer et al., 2010] (fluorescence 18–48% higher than expected, see Figures S11, S14, and S15 in Text S1). High-latitude needleleaf forests (55–70°N), on the other hand, exhibit a 30% lower than expected fluorescence signal. We hypothesize that differences in fluorescence yield and light-use efficiency, potentially caused by water or nutrient limitation may be the reason for the discrepancy (see also auxiliary material, Figure S15 in Text S1). At high latitudes under low light conditions, deviations in the response of fluorescence as a function of GPP may also play a role as fluorescence and photosynthesis can compete under those circumstances [Van der Tol et al., 2009]. However, at 10–40°N in boreal summer, high light conditions prevail and a stricter correlation of GPP with fluorescence is expected (but a deviation from the linear correlation cannot

Figure 3. Latitudinal cross sections of fluorescence ($F_s$) and model GPP estimates for different seasons. The different y-axes are scaled according to the slope of the linear regression line as displayed in Figure 2 (i.e., fluorescence signals are directly comparable to GPP under the assumption of the linear correlation). The green-shaded area represents the ensemble range of the MPI-BGC GPP estimate [Beer et al., 2010; Jung et al., 2011].
yet be excluded with certainty). Savannas are difficult to model from meteorological observations because savanna vegetation imposes relatively more biological control over fluxes, and is less controlled by meteorological variability than are wetter ecosystems [Baldocchi and Xu, 2007]. Croplands are difficult to model on a global scale because of large variability in LUE, as well as uncertain irrigation and fertilization practices. Hence, ancillary data needed to derive GPP from vegetation indices may be biased. For MODIS, GPP summer biases in needleleaf forests (positive) and croplands (negative) have also been observed in site-level evaluations and attributed to errors in LUE and an oversensitive parameterization of vapor pressure deficit dependency [Turner et al., 2005]. Further, the area with highest deviations is almost devoid of flux towers, increasing uncertainties in MPI-BGC. Even though the discrepancies cannot yet be unequivocally resolved, our results point to underestimations of light use efficiency for savannas and croplands in boreal summer (Figure S15 in Text S1).

[10] We acknowledge that the observed linear relationship is empirical and further studies are needed regarding the exact quantitative relationship of steady-state fluorescence [Maxwell and Johnson, 2000] with GPP under various light and temperature conditions and especially considering spatial scales largely exceeding leaf-level and laboratory scales. Eventually, this will unleash the full potential of global space-borne observations of fluorescence. However, we demonstrated the utility and simplicity of using raw fluorescence without any ancillary datasets or model assumptions as a direct linear predictor of GPP at the global scale. Moreover, the GOSAT satellite samples only once per day, does not cover the entire earth and was not even intended to retrieve fluorescence. Nevertheless, it can be anticipated that chlorophyll fluorescence retrievals from GOSAT and OCO-2 (taking about 50 times more data than GOSAT, hence largely reducing the uncertainty) in conjunction with their global atmospheric CO₂ measurements will provide an exceptional combination of a vegetation and atmospheric perspective on the global carbon budget, constraining our model predictions for future atmospheric CO₂ abundances.

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