

Novel Aminonaphthoquinone Mannich Bases Derived from Lawsone and their Copper(II) Complexes: Synthesis, Characterization and Antibacterial Activity

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Uma série de novas Bases de Mannich (**HL1-HL13**) derivadas da 2-hidroxi-1,4-naftoquinona (lawsone), benzaldeídos substituídos [$C_6H_2R^1R^2R^3C(O)H$] e várias aminas primárias (NH_2R^4 , $R^4 = n$ -butil, benzil, alil, 2-furfúril) e seus complexos de Cu^{2+} , $[Cu(L1)_2]$ - $[Cu(L13)_2]$, foram sintetizados e caracterizados por métodos analíticos e espectroscópicos. As estruturas dos complexos **1** ($R^1 = R^2 = R^3 = H$; $R^4 = Bu$), **2** ($R^1 = R^3 = H$; $R^2 = NO_2$; $R^4 = Bu$) e **7** ($R^1 = OH$; $R^2 = R^3 = H$; $R^4 = Bu$) foram determinadas por estudos de difração de raios-X de monocristal. Todos os compostos cristalizam em grupos espaciais centrossimétricos, com um cobre no centro de inversão. Dois L^- coordenam-se através dos átomos de oxigênio do naftalen-2-olato e do nitrogênio da amina secundária, formando anéis quelatos de seis membros ao redor do átomo de cobre em um ambiente $trans-N_2O_2$. A atividade antimicrobiana de todos os compostos foi testada em sete diferentes linhagens de bactérias: *Bacillus cereus*, *Bacillus subtilis*, *Escherichia coli*, *Enterococcus faecalis*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa* e *Staphylococcus aureus*. Em geral, as bases de Mannich foram mais ativas que os complexos, sendo **HL11** ($R^1 = OH$; $R^2 = H$; $R^3 = Me$; $R^4 = Bn$) e **HL13** ($R^1 = OH$; $R^2 = H$; $R^3 = Br$; $R^4 = Bn$) os inibidores mais potentes. O MIC para o composto mais ativo **HL11** contra *S. Coli* foi $20 \mu mol L^{-1}$ ($8 \mu g mL^{-1}$), melhor que o cloranfenicol ($90 \mu mol L^{-1}$) e bem abaixo da maioria dos valores descritos para outras naftoquinonas.

A series of novel Mannich bases (**HL1-HL13**) derived from 2-hydroxy-1,4-naphthoquinone (lawsone), substituted benzaldehydes [$C_6H_2R^1R^2R^3C(O)H$] and various primary amines (NH_2R^4 , $R^4 = n$ -butyl, benzyl, allyl, 2-furfuryl), and their Cu^{2+} complexes, $[Cu(L1)_2]$ - $[Cu(L13)_2]$, have been synthesized and fully characterized by analytical and spectroscopic methods. The structures of complexes **1** ($R^1 = R^2 = R^3 = H$; $R^4 = Bu$), **2** ($R^1 = R^3 = H$; $R^2 = NO_2$; $R^4 = Bu$) and **7** ($R^1 = OH$; $R^2 = R^3 = H$; $R^4 = Bu$) were determined by single crystal X-ray diffraction studies. All complexes crystallize in centrosymmetric space groups, with a copper atom in the inversion centre. Two L^- coordinate through the naphthalen-2-olate oxygen and secondary amine-N atoms, forming six-membered chelate rings around the copper atom in a $trans-N_2O_2$ environment; spectroscopic data confirm that the other complexes exhibit similar molecular arrangement. The antimicrobial activity of all compounds has been tested on seven different strains of bacteria: *Bacillus cereus*, *Bacillus subtilis*, *Escherichia coli*, *Enterococcus faecalis*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. In general, Mannich bases were more active than complexes, **HL11** ($R^1 = OH$; $R^2 = H$; $R^3 = Me$; $R^4 = Bn$) and **HL13** ($R^1 = OH$; $R^2 = H$; $R^3 = Br$; $R^4 = Bn$) being the most potent inhibitors. The MIC for the most active compound **HL11** against *S. Coli* was $20 \mu mol L^{-1}$ ($8 \mu g mL^{-1}$), better than Chloramphenicol ($90 \mu mol L^{-1}$) and well below most values reported for other naphthoquinones.

Keywords: aminonaphthoquinones, copper complexes, Mannich bases, crystal structure determination, antibacterial activity

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Introduction

Natural and synthetic naphthoquinones are known for a wide range of biological activities,¹ amongst which anti-cancer,^{2,3} tripanocidal,⁴ molluscicidal,⁵ antimalarial,⁶ leishmaniscide,⁷ bacteriostatic and bactericidal.^{8,9} The most accepted mechanism for the antimicrobial activity of naphthoquinones is based on the generation of reactive oxygen species by two successive reduction processes to form radical anion and dianion species that are toxic to bacteria.^{10,11}

It has been shown that the incorporation of amino groups or a nitrogen atom into naphthoquinones often results in increased anticancer,¹²⁻¹⁴ molluscicidal¹⁵⁻¹⁷ and antibacterial activities.^{10,18-22} We therefore evaluated the antimicrobial activity of a novel series of 2-hydroxy-3-alkylamine-1,4-naphthoquinones, known as *Mannich bases*. These compounds were first synthesized over sixty years ago,²³ and their antimalarial^{23,24} and molluscicidal²⁵ activities have been described. In spite of the fact that metal complexation of a number naphthoquinones or naphthoquinone derived compounds has resulted in increased cytotoxicity,²⁶ antimalarial^{27,28} and anticancer²⁹ activity, transition metal complexes of Mannich bases derived from 2-hydroxy-1,4-naphthoquinone (lawsone) have not yet been reported. Herein we describe the synthesis of novel Mannich bases **HL1-HL13** from lawsone, a number of primary amines and substituted benzaldehydes and of their copper(II) complexes, [Cu(L)₂] (**1-13**) (Scheme 1), their characterization by analytical and spectroscopic techniques, and the X-ray diffraction studies of three complexes. Furthermore, we report the results of antibacterial activity screening of all compounds and discuss growth inhibition as a function of structural changes and metal complexation.

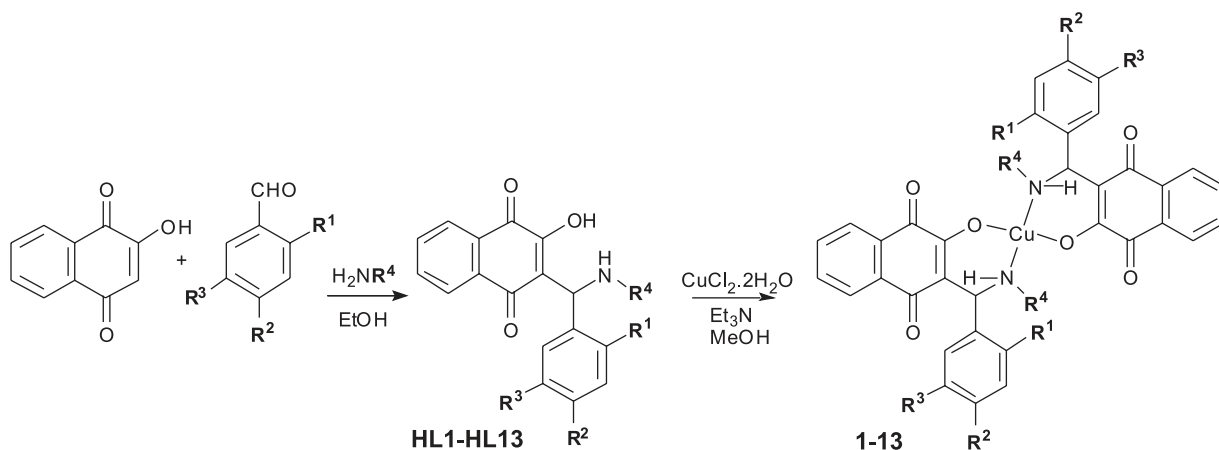
Experimental

Materials and methods

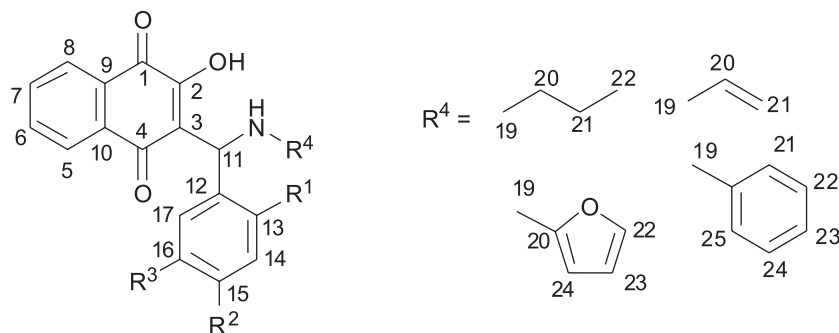
Reagents and solvents were used without further purification. Microanalyses were performed using a Perkin-Elmer CHN 2400 micro analyser at the Central Analítica, Instituto de Química, USP, São Paulo, Brazil. Melting points were obtained with a Mel-Temp II, Laboratory Devices-USA apparatus and are uncorrected. IR spectra (KBr pellets) were recorded on a FT-IR Spectrum One (Perkin Elmer) spectrophotometer. ¹H and ¹³C NMR spectra were recorded with a Varian Unit Plus 300 MHz spectrometer in CDCl₃ or d⁶-DMSO; coupling constants are reported in Hertz (Hz) and chemical shifts in parts per million (ppm) relative to internal standard Me₄Si. The hydrogen signals were attributed thought coupling constant values and ¹H × ¹H-COSY experiments. Electronic spectra were taken on a Diode Array 8452A (Hewlett Packard-HP) spectrophotometer using spectroscopic grade solvents (Tedia Brazil) in 10⁻³ and 10⁻⁴ mol L⁻¹ solutions. Electron paramagnetic resonance (EPR) spectra of the solid samples were obtained at liquid nitrogen temperature (77 K), on a Bruker ESP300E equipment with modulation frequency of 100 kHz, operating at 9.5 GHz (X-band).

Synthesis of the Mannich bases **HL1-HL13**

Compounds **HL1-HL13** (Figure 1) were synthesized according to the general procedure described in the literature^{24,30} with modifications. They were obtained by reacting a suspension of lawsone (5 mmol, 0.870 g), in 15 mL of ethanol, with the respective amine (5.5 mmol). After formation of the lawsonate solution, the aldehyde (6 mmol) is added and the mixture, left stirring at room



Scheme 1. Synthesis of Mannich bases **HL1-HL13** and of complexes **1-13**.



$R^1 = R^2 = R^3 = H; R^4 = Bu$, **HL1**
 $R^1 = R^3 = H; R^2 = NO_2; R^4 = Bu$, **HL2**
 $R^1 = R^3 = H; R^2 = NO_2; R^4 = Bn$, **HL3**
 $R^1 = R^2 = Cl; R^3 = H; R^4 = Bu$, **HL4**
 $R^1 = R^2 = Cl; R^3 = H; R^4 = Bn$, **HL5**
 $R^1 = OH; R^2 = R^3 = H; R^4 = \text{aryl}$, **HL6**
 $R^1 = OH; R^2 = R^3 = H; R^4 = Bu$, **HL7**

$R^1 = OH; R^2 = R^3 = H; R^4 = Bn$, **HL8**
 $R^1 = OH; R^2 = R^3 = H; R^4 = CH_2C_4H_3O$, **HL9**
 $R^1 = OH; R^2 = H; R^3 = Me; R^4 = Bu$, **HL10**
 $R^1 = OH; R^2 = H; R^3 = Me; R^4 = Bn$, **HL11**
 $R^1 = OH; R^2 = H; R^3 = Br; R^4 = Bu$, **HL12**
 $R^1 = OH; R^2 = H; R^3 = Br; R^4 = Bn$, **HL13**

Figure 1. Mannich bases **HL1-HL13**.

temperature for 12 h in the dark. The orange solids were filtered, washed with ethanol, diethyl ether and dried under vacuum.

3-[N-(n-butyl)aminobenzyl]-2-hydroxy-1,4-naphthoquinone (HL1)

From 0.54 mL of butylamine and 0.61 mL of benzaldehyde. Yield: 1.258 g, 74%, mp 143-144 °C. Anal. Calc. for $C_{21}H_{21}NO_3$: C, 75.20; H, 6.31; N, 4.18%. Found: C, 75.14; H, 6.29; N, 4.23%. IR (KBr) ν_{max}/cm^{-1} : 3435 (O-H), 3060 (C-H), 2960 (C-H), 2933 (C-H), 1680 (C=O), 1588 (C=C), 1529 (δ N-H), 1276 (C-O). 1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.02 (ddd, J 7.60, 1.22, 0.45 Hz, 1H, *H5* or *H8*); 7.93 (ddd, J 7.57, 1.41, 0.45 Hz, 1H, *H8* or *H5*); 7.81 (td, J 7.60, 7.60, 1.41 Hz, 1H, *H6* or *H7*); 7.73-7.66 (m, 3H, *H7* or *H6* and *Ph*); 7.49-7.38 (m, 3H, *Ph*); 5.59 (br s, 1H, *H11*); 2.97 (br t, J 7.65, 7.65 Hz, 2H, *H19*); 1.76-1.64 (m, 2H, *H20*); 1.48-1.34 (m, 2H, *H21*); 0.94 (t, J 7.36, 7.36 Hz, 3H, *H22*). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.6, 178.7, 170.8, 138.9, 134.8, 134.0, 131.7, 131.1, 128.6, 128.1, 127.9, 125.6, 125.3, 111.5, 59.0, 45.6, 27.9, 19.5, 13.7. UV-Vis ($CHCl_3$) λ/nm , log ϵ : 267 (4.19), 338 (3.15), 437(3.20).

3-[N-(n-butyl)amino-3-nitrobenzyl]-2-hydroxy-1,4-naphthoquinone (HL2)

From 0.54 mL of butylamine and 0.907 g of *p*-nitrobenzaldehyde. Yield: 1.141 g, 60%, mp 172 °C. Anal. Calc. for $C_{21}H_{20}N_2O_5$: C, 66.31; H, 5.30; N, 7.36%. Found: C, 66.20; H, 5.49; N, 7.51%. IR (KBr) ν_{max}/cm^{-1} : 3437 (O-H), 3086 (C-H), 2962 (C-H), 1683 (C=O), 1588 (C=C), 1520

(δ N-H), 1281 (C-O). 1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.33 (d, J 8.85 Hz, 2H, *H14*; *H16*); 8.03 (ddd, J 7.62, 1.34, 0.49 Hz, 1H, *H5* or *H8*); 7.97 (d, J 8.85 Hz, 2H, *H13*; *H17*); 7.93 (ddd, J 7.62, 1.39, 0.49 Hz, 1H, *H8* or *H5*); 7.82 (td, J 7.62, 7.62, 1.39 Hz, 1H, *H6* or *H7*); 7.70 (td, J 7.62, 7.62, 1.34 Hz, 1H, *H7* or *H6*); 5.78 (s, 1H, *H11*); 3.02 (t, J 7.67, 7.67 Hz, 2H, *H19*); 1.73 (t, J 7.67, 7.67 Hz, 2H, *H20*); 1.40 (sx, J 7.33 Hz, 2H, *H21*); 0.95 (t, J 7.33, 7.33 Hz, 3H, *H22*). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.2, 178.5, 170.7, 146.8, 146.0, 134.6, 133.9, 131.6, 131.1, 128.6, 125.5, 123.5, 110.3, 57.9, 45.6, 27.8, 19.4, 13.6. UV-Vis ($CHCl_3$) λ/nm , log ϵ : 268 (4.36), 336 (3.60), 427(3.26).

3-[N-(benzyl)amino-3-nitrobenzyl]-2-hydroxy-1,4-naphthoquinone (HL3)

From 0.60 mL of benzylamine and 0.907 g of *p*-nitrobenzaldehyde. Yield: (1.513 g, 73%); mp 139-140 °C. Anal. Calc. for $C_{24}H_{18}N_2O_5$: C, 69.56; H, 4.38; N, 6.76%. Found: C, 68.80; H, 4.39; N, 6.67%. IR (KBr) ν_{max}/cm^{-1} : 3445 (O-H), 3066 (C-H), 3032 (C-H), 2969 (C-H), 1677 (C=O), 1592 (C=C), 1523 (δ N-H), 1272 (C-O). 1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.31 (d, J 8.86 Hz, 2H, *H14*; *H16*); 8.04 (ddd, J 7.60, 1.38, 0.40 Hz, 1H, *H5* or *H8*); 7.95 (ddd, J 7.48, 1.41, 0.40 Hz, 1H, *H8* or *H5*); 7.92 (d, J 8.86 Hz, 2H, *H13*; *H17*); 7.83 (td, J 7.60, 7.60, 1.41 Hz, 1H, *H6* or *H7*); 7.70 (td, J 7.48, 7.48, 1.38 Hz, 1H, *H7* or *H6*); 7.52-7.49 (m, 5H, *Ph*); 5.76 (s, 1H, *H11*); 4.28 (s, 2H, *H19*). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.1, 178.6, 170.7, 146.9, 145.7, 134.6, 133.8, 132.1, 131.6, 131.1, 130.2, 128.9, 128.8, 128.6, 125.5, 125.2, 123.5,

110.1, 57.8, 49.3. UV-Vis (CH_2Cl_2) λ_{nm} , log ϵ : 273 (4.47), 337 (3.74), 389 (3.22).

3-[N-(n-butyl)amino-2,4-dichlorobenzyl]-2-hydroxy-1,4-naphthoquinone (HLA)

From 0.60 mL of benzylamine and 1.050 g of 2-4-dichloro-benzaldehyde. Yield: 1.071 g, 53%; mp 142-143 °C. Anal. Calc. for $\text{C}_{21}\text{H}_{19}\text{Cl}_2\text{NO}_3$: C, 62.39; H, 4.74; N, 3.46%. Found: C, 62.00; H, 4.64; N, 3.51%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3437 (O-H), 3064 (C-H), 2958 (C-H), 2870 (C-H), 1678 (C=O), 1616 (C=C), 1588 (C=C), 1531 (δ N-H), 1272 (C-O). ^1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.01 (dd, J 7.46, 1.33 Hz, 1H, H_5 or H_8); 7.97 (dd, J 7.46, 1.37 Hz, 1H, H_8 or H_5); 7.96 (d, J 8.51 Hz, 1H, H_{17}); 7.83 (td, J 7.46, 7.46, 1.37 Hz, 1H, H_6 or H_7); 7.76 (d, J 2.19, 1H, H_{14}); 7.72 (td, J 7.46, 7.46, 1.33 Hz, 1H, H_7 or H_6); 7.55 (dd, J 8.51, 2.19, 1H, H_{16}); 5.99 (s, 1H, H_{11}); 3.05 (br t, J 7.35, 7.35 Hz, 2H, H_{19}); 1.80-1.65 (m, 2H, H_{20}); 1.38-1.34 (m, 2H, H_{21}); 0.96 (t, J 7.28, 7.28 Hz, 3H, H_{22}). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.1, 178.9, 171.3, 134.8, 134.5, 134.4, 133.9, 133.8, 131.9, 131.6, 131.1, 128.8, 127.7, 125.5, 125.2, 109.6, 55.4, 45.9, 27.7, 19.3, 13.5. UV-Vis (CHCl_3) λ_{nm} , log ϵ : 265 (4.29), 337 (3.42), 422 (3.29).

3-[N-(n-benzyl)amino-2,4-dichlorobenzyl]-2-hydroxy-1,4-naphthoquinone (HL5)

From 0.54 mL of butylamine and 1.050 g of 2-4-dichloro-benzaldehyde. Yield: 1.731 g, 79%; mp 145-146 °C. Anal. Calc. for $\text{C}_{24}\text{H}_{17}\text{Cl}_2\text{NO}_3$: C, 65.77; H, 3.91; N, 3.20%. Found: C, 65.67; H, 3.87; N, 3.24%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3453 (O-H), 3065 (C-H), 3033 (C-H), 2969 (C-H), 1677 (C=O), 1592 (C=C), 1523 (δ N-H), 1271 (C-O). ^1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.02 (ddd, J 7.37, 1.50, 0.47 Hz, 1H, H_5 or H_8); 8.00 (dd, J 7.37, 1.50, 0.47 Hz, 1H, H_8 or H_5); 7.86 (d, J 8.63 Hz, 1H, H_{17}); 7.84 (td, J 7.46, 7.46, 1.50 Hz, 1H, H_6 or H_7); 7.74 (td, J 7.46, 7.46, 1.50 Hz, 1H, H_7 or H_6); 7.73 (d, J 2.16 Hz, 1H, H_{14}); 7.55-7.49 (m, 6H, H_{16} ; Ph); 5.78 (s, 1H, H_{11}); 4.35 (d, J 13.00 Hz, 1H, H_{19}); 4.21 (d, J 13.00 Hz, 1H, H_{19}). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 183.9, 179.3, 171.3, 134.6, 134.5, 134.4, 134.0, 133.9, 132.1, 132.0, 131.7, 131.2, 130.3, 128.9, 128.8, 128.6, 127.8, 125.6, 125.2, 109.4, 55.1, 49.7. UV-Vis (CHCl_3) λ_{nm} , log ϵ : 275 (4.14), 334 (3.40), 400 (3.10).

3-[N-(allyl)amino-2-hydroxybenzyl]-2-hydroxy-1,4-naphthoquinone (HL6)

From 0.41 mL of allylamine and 0.63 mL of 2-hydroxy-benzaldehyde. Yield: 1.475 g, 88%; mp 164-165 °C. Anal. Calc. for $\text{C}_{20}\text{H}_{17}\text{NO}_4$: C, 71.63; H, 5.11; N, 4.18%.

Found: C, 71.50; H, 5.05; N, 4.27%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3255 (O-H), 3072 (C-H), 2980 (C-H), 2950 (C-H), 1678 (C=O), 1593 (C=C), 1561 (δ N-H), 1277 (C-O). ^1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.02 (ddd, J 7.61, 1.35, 0.47 Hz, 1H, H_5 or H_8); 7.97 (dd, J 7.53, 1.41, 0.47 Hz, 1H, H_8 or H_5); 7.83 (td, J 7.47, 7.47, 1.41 Hz, 1H, H_6 or H_7); 7.73 (td, J 7.47, 7.47, 1.35 Hz, 1H, H_7 or H_6); 7.42 (dd, J 7.69, 1.65 Hz, 1H, H_{14} or H_{17}); 7.26 (td, J 8.06, 8.06, 1.65 Hz, 1H, H_{16} or H_{15}); 6.96 (dd, J 8.06, 0.99 Hz, 1H, H_{17} or H_{14}); 6.86 (td, J 7.69, 7.69, 0.99 Hz, 1H, H_{15} or H_{16}); 6.10-5.95 (m, 2H, H_{20}); 5.85 (s, H_{11}); 5.50-5.42 (m, 2H, H_{21}); 3.74-3.60 (m, 2H, H_{19}). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.1, 179.6, 171.5, 155.5, 134.6, 133.9, 131.6, 131.1, 129.8, 129.4, 128.6, 125.5, 125.2, 123.8, 122.0, 119.1, 116.1, 110.1, 53.3, 47.9. UV-Vis (DMSO) λ_{nm} , log ϵ : 278 (4.34), 452 (3.33).

3-[N-(n-butyl)amino-2-hydroxybenzyl]-2-hydroxy-1,4-naphthoquinone (HL7)

From 0.54 mL of butylamine and 0.63 mL of 2-hydroxy-benzaldehyde. Yield: 1.405 g, 80%; mp 137-138 °C (with dec.). Anal. Calc. for $\text{C}_{21}\text{H}_{21}\text{NO}_4$: C, 71.78; H, 6.02; N, 3.99%. Found: C, 71.12; H, 6.03; N, 3.92%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3233 (O-H), 3069 (C-H), 2959 (C-H), 2875 (C-H), 1681 (C=O), 1590 (C=C), 1528 (δ N-H), 1275 (C-O). ^1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.02 (ddd, J 7.66, 1.35, 0.50 Hz, 1H, H_5 or H_8); 7.98 (ddd, J 7.47; 1.35; 0.50 Hz, 1H, H_8 or H_5); 7.84 (td, J 7.47, 7.47, 1.35 Hz, 1H, H_6 or H_7); 7.73 (td, J 7.47, 7.47, 1.35 Hz, 1H, H_7 or H_6); 7.44 (dd, J 7.74, 1.56 Hz, 1H, H_{14} or H_{17}); 7.27 (td, J 8.03, 8.03, 1.56 Hz, 1H, H_{16} or H_{15}); 6.99 (dd, J 8.03, 1.05 Hz, 1H, H_{17} or H_{14}); 6.86 (td, J 7.74, 7.74, 1.05 Hz, 1H, H_{15} or H_{16}); 5.86 (s, 1H, H_{11}); 3.03 (t, J 7.50, 7.50 Hz, 2H, H_{19}); 1.79-1.66 (m, 2H, H_{20}); 1.50-1.38 (m, 2H, H_{21}); 0.96 (br t, J 7.33, 7.33 Hz, 3H, H_{22}). ^{13}C NMR (DMSO- d_6 , 75 MHz): δ (ppm) 184.1, 179.6, 171.6, 155.4, 134.5, 133.9, 131.6, 131.2, 129.6, 128.7, 125.5, 125.2, 123.6, 119.1, 115.9, 109.9, 54.1, 45.7, 27.8, 19.4, 13.5. UV-Vis (DMSO) λ_{nm} , log ϵ : 277 (4.33), 450 (3.30).

3-[N-(benzyl)amino-2-hydroxybenzyl]-2-hydroxy-1,4-naphthoquinone (HL8)

From 0.60 mL of benzylamine and 0.63 mL of 2-hydroxy-benzaldehyde. Yield: 1.792 g, 93%; mp 165-166 °C. Anal. Calc. for $\text{C}_{24}\text{H}_{19}\text{NO}_4$: C, 74.79; H, 4.97; N, 3.63%. Found: C, 74.78; H, 4.90; N, 3.72%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3436 (O-H), 3067 (C-H), 2744 (C-H), 1681 (C=O), 1589 (C=C), 1516 (δ N-H), 1276 (C-O). ^1H NMR (DMSO- d_6 , 300 MHz): δ (ppm) 8.04 (ddd, J 7.50, 1.36, 0.48 Hz, 1H, H_5 or H_8); 7.99 (ddd, J 7.50, 1.43, 0.48 Hz, 1H, H_8 or H_5); 7.84 (td, J 7.50, 7.50, 1.43 Hz, 1H, H_6

or *H7*); 7.73 (td, *J* 7.50, 7.50, 1.36 Hz, 1H, *H7* or *H6*); 7.51-7.46 (m, 5H, *Ph*); 7.38 (dd, *J* 7.71, 1.62 Hz, 1H, *H14* or *H17*); 7.24 (td, *J* 8.05; 8.05, 1.62 Hz, 1H, *H16* or *H15*); 6.94 (dd, *J* 8.05, 1.00 Hz, 1H, *H17* or *H14*); 6.84 (td, *J* 7.71, 7.71, 1.00 Hz, 1H, *H15* or *H16*); 5.82 (s, 1H, *H11*); 4.26 (d, *J* 13.09, 1H); 4.17 (d, *J* 13.09, 1H). ¹³C NMR (DMSO-d₆, 75 MHz): δ (ppm) 184.1, 179.7, 171.5, 155.6, 134.6, 133.9, 132.8, 131.7, 131.1, 130.0, 129.4, 128.9, 128.7, 128.5, 125.5, 125.2, 123.6, 119.0, 115.9, 110.0, 53.8, 49.4. UV-Vis (DMSO) λ/nm, log ε: 277 (4.29), 443 (3.25).

3-[N-(furfurylmethyl)amino-2-hydroxybenzyl]-2-hydroxy-1,4-naphthoquinone (HL9)

From furfurylamine (0.49 mL) and 2-hydroxybenzaldehyde (0.63 mL). Yield: 1.653 g, 88%; mp 138 °C. Anal. Calc. for C₂₄H₁₉NO₄·H₂O: C, 67.17; H, 4.87; N, 3.56%. Found: C, 68.94; H, 4.99; N, 3.69%. IR (KBr) ν_{max}/cm⁻¹: 3234 (O-H), 2943 (C-H), 1683 (C=O), 1591 (C=C), 1551 (δ N-H), 1279 (C-O). ¹H NMR (DMSO-d₆, 300 MHz): δ (ppm) 8.02 (br d, *J* 7.61 Hz, 1H, *H5* or *H8*); 7.97 (br d, *J* 7.56 Hz, 1H, *H8* or *H5*); 7.83 (td, *J* 7.61, 7.61, 1.20 Hz, 1H, *H6* or *H7*); 7.72 (td, *J* 7.56, 7.56, 1.20 Hz, 1H, *H7* or *H6*); 7.51-7.46 (m, 5H, *Ph*); 7.32 (br d, *J* 7.89 Hz, 1H, *H14* or *H17*); 7.23 (br td, *J* 7.89; 7.89, 1.62 Hz, 1H, *H16* or *H15*); 6.92 (br d, *J* 7.89 Hz, 1H, *H17* or *H14*); 6.82 (br t, *J* 7.48 Hz, 1H, *H15* or *H16*); 5.78 (s, 1H, *H11*); 4.23 (d, *J* 14.51, 1H); 4.17 (d, *J* 14.51, 1H). ¹³C NMR (DMSO-d₆, 75 MHz): δ (ppm) 184.2, 179.7, 171.4, 155.7, 143.9, 134.6, 133.9, 131.7, 131.2, 129.4, 128.6, 125.5, 125.3, 123.7, 119.2, 119.0, 116.0, 111.6, 111.0, 110.1, 53.9, 41.9. UV-Vis (CHCl₃) λ/nm, log ε: 272 (4.03), 305 (3.93), 366 (3.74).

3-[N-(n-butyl)amino-2-hydroxy-5-methyl-benzyl]-2-hydroxy-1,4-naphthoquinone (HL10)

From butylamine (0.54 mL) and 2-hydroxy-5-methylbenzaldehyde (0.817 g). Yield: 1.279 g, 70%; mp 154-155 °C. Anal. Calc. for C₂₂H₂₃NO₄: C, 72.31; H, 6.34; N, 3.83. Found: C, 71.15; H, 6.28; N, 3.90%. IR (KBr) ν_{max}/cm⁻¹: 3065, 2959, 2870, 1686, 1591, 1551, 1508, 1477, 1432, 1374, 1278. ¹H NMR (DMSO-d₆, 300 MHz): δ (ppm) 8.02 (ddd, *J* 7.67; 1.30; 0.40 Hz, 1H, *H5* or *H8*); 7.98 (ddd, *J* 7.54; 1.37; 0.40 Hz, 1H, *H8* or *H5*); 7.84 (td, *J* 7.54; 7.54; 1.30 Hz, 1H, *H6* or *H7*); 7.73 (td, *J* 7.47; 7.47; 1.37 Hz, 1H, *H7* or *H6*); 7.25 (d, *J* 1.94 Hz, 1H, *H17*); 7.07 (dd, *J* 8.22, 1.94 Hz, 1H, *H15*); 6.87 (d, *J* 8.22 Hz, *H14*); 5.81 (s, 1H, *H11*); 3.00 (t, *J* 7.55, 7.55 Hz, 2H, *H19*); 2.25 (s, 3H, *CH*₃); 1.77-1.65 (m, 2H, *H20*); 1.48-1.34 (m, 2H, *H21*); 0.96 (t, *J* 7.34, 7.34 Hz, 3H, *H22*). ¹³C NMR (DMSO-d₆, 75 MHz): δ (ppm) 184.1, 179.5, 171.4, 153.1, 134.5, 133.9,

131.6, 131.1, 129.8, 128.6, 127.5, 125.5, 125.2, 123.5; 115.9; 110.2; 54.1; 45.7; 27.9; 20.3; 19.4; 13.5. UV-Vis (CHCl₃) λ/nm, log ε: 274 (4.10), 325 (3.63), 373 (3.38), 470 (2.93).

3-[N-(benzyl)amino-2-hydroxy-5-methyl-benzyl]-2-hydroxy-1,4-naphthoquinone (HL11)

From benzylamine (0.60 mL) and 2-hydroxy-5-methylbenzaldehyde (0.817 g). Yield: 1.338 g, 67%; mp 147-148 °C. Anal. Calc. for C₂₅H₂₁NO₄: C, 75.17; H, 5.30; N, 3.51. Found: C, 74.54; H, 5.36; N, 3.51%. IR (KBr) ν_{max}/cm⁻¹: 3266; 3065; 2957; 2862; 1686; 1590; 1539; 1274; 1221. ¹H NMR (DMSO-d₆, 300 MHz): δ (ppm) 8.04 (ddd, *J* 7.61, 1.39, 0.43 Hz, 1H, *H5* or *H8*); 7.99 (ddd, *J* 7.54, 1.41, 0.43 Hz, 1H, *H8* or *H5*); 7.84 (td, *J* 7.43, 7.43, 1.41 Hz, 1H, *H6* or *H7*); 7.73 (td, *J* 7.43, 7.43, 1.39 Hz, 1H, *H7* or *H6*); 7.50-7.47 (m, 5H, *Ph*); 7.20 (d, *J* 2.03 Hz, 1H, *H17*); 7.04 (dd, *J* 8.16, 2.03 Hz, 1H, *H15*); 6.83 (d, *J* 8.16 Hz, 1H, *H14*); 5.77 (s, *H11*); 4.24 (d, *J* 13.13 Hz, 1H, *H19*); 4.16 (d, *J* 13.13 Hz, 1H, *H19'*); 2.23 (s, 3H, *CH*₃). ¹³C NMR (DMSO-d₆, 75 MHz): δ (ppm) 184.1, 179.9, 171.5, 153.2, 134.6, 133.9, 132.8, 131.7, 131.2, 130.1, 129.9, 128.9, 128.7, 128.6, 127.6, 125.6, 125.3, 123.5, 116.0, 110.2, 54.0, 49.5, 20.4. UV-Vis (CHCl₃) λ/nm, log ε: 250 (4.46), 310 (4.15), 382 (3.90).

3-[N-(n-butyl)amino-2-hydroxy-5-bromo-benzyl]-2-hydroxy-1,4-naphthoquinone (HL12)

From butylamine (0.54 mL) and 2-hydroxy-5-bromobenzaldehyde (1.206 g). Yield: 1.316 g, 61%; mp 166-167 °C. Anal. Calc. for C₂₁H₂₀BrNO₄: C, 58.62; H, 4.68; N, 3.26. Found: C, 58.74; H, 4.54; N, 3.42%. IR (KBr) ν_{max}/cm⁻¹: 3202; 2961; 2934; 2867; 1679; 1591; 1524; 1273; 1229. ¹H NMR (DMSO-d₆, 300 MHz): δ (ppm) 8.04 (d, *J* 7.20 Hz, 1H, *H5* or *H8*); 7.98 (d, *J* 7.20 Hz, 1H, *H8* or *H5*); 7.84 (t, *J* 7.20, 7.20 Hz, 1H, *H6* or *H7*); 7.73 (t, *J* 7.20, 7.20 Hz, 1H, *H7* or *H6*); 7.64 (d, *J* 2.15 Hz, 1H, *H17*); 7.42 (dd, *J* 8.58, 2.15 Hz, 1H, *H15*); 6.94 (d, *J* 8.58 Hz, 1H, *H14*); 5.82 (s, 1H, *H11*); 3.05-2.95 (m, 2H, *H19*); 1.78-1.65 (m, 2H, *H20*); 1.49-1.35 (m, 2H, *H21*); 0.96 (t, *J* 7.04, 7.04 Hz, 3H, *H22*). ¹³C NMR (DMSO-d₆, 75 MHz): δ (ppm) 183.9, 179.5, 171.1, 154.8, 134.4, 133.9, 132.0, 131.6, 131.2, 130.9, 126.5, 125.6, 125.2, 118.2, 110.2, 109.8, 53.4, 45.8, 27.8, 19.3, 13.6. UV-Vis (CHCl₃) λ/nm, log ε: 273 (3.12), 354 (2.27), 457 (2.05).

3-[N-(benzyl)amino-2-hydroxy-5-bromo-benzyl]-2-hydroxy-1,4-naphthoquinone (HL13)

From benzylamine (0.60 mL) and 2-hydroxy-5-bromobenzaldehyde (1.206 g). Yield: 1.736 g, 75%; mp 160-161 °C. Anal. Calc. for C₂₄H₁₈BrNO₄: C, 62.08; H, 3.91; N, 3.02. Found: C, 62.09; H, 4.01; N, 3.11%. IR (KBr) ν_{max}/cm⁻¹: 3224 (O-H), 3068 (C-H), 2958 (C-H), 1688 (C=O), 1589

(C=C), 1539 (δ N-H), 1275 (C-O). $^1\text{H NMR}$ (DMSO- d_6 , 300 MHz): δ (ppm) 8.04 (dd, J 7.62, 1.36 Hz, 1H, $H5$ or $H8$); 7.99 (d, J 7.54, 1.41 Hz, 1H, $H8$ or $H5$); 7.85 (td, J 7.44, 7.44, 1.41 Hz, 1H, $H6$ or $H7$); 7.74 (td, J 7.44, 7.44, 1.36 Hz, 1H, $H7$ or $H6$); 7.54 (d, J 2.37 Hz, 1H, $H17$); 7.50-7.46 (m, 5H, Ph); 7.39 (dd, J 8.63, 2.37 Hz, 1H, $H15$); 6.89 (d, J 8.63 Hz, 1H, $H14$); 5.77 (s, 1H, $H11$); 4.24 (d, J 13.15 Hz, 1H, $H19$); 4.15 (d, J 13.15 Hz, 1H, $H19'$). $^{13}\text{C NMR}$ (DMSO- d_6 , 75 MHz) δ (ppm): 184.5, 180.3, 172.1, 155.6, 135.1, 134.5, 133.3, 132.5, 132.3, 131.8, 131.5, 130.7, 129.5, 129.3, 129.1, 127.1, 126.2, 125.9, 118.8, 110.7, 110.4, 53.9, 50.1. UV-Vis (CHCl_3) λ/nm , log ϵ : 271 (3.10), 377 (2.60), 434 (1.94).

Synthesis of complexes [Cu(L) $_2$] 1-13 from HL1-HL13, respectively

To a suspension of 1 mmol of the ligand in 10 mL MeOH, was added a solution of $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ (83 mg, 0.5 mmol) in 2 mL MeOH. After addition of Et_3N (0.14 mL, 1 mmol), the suspension was left under stirring in the dark for 12h at room temperature. The resulting solids were filtered off, washed with methanol, diethyl ether and dried under vacuum (Figure 2).

[Cu(L1) $_2$] (1)

From 335 mg of **HL1**. Yield: 337 mg, 92%; mp 198 °C. Slow evaporation of a CHCl_3 solution yielded brown crystals suitable for X-ray diffraction analysis. Anal. calc. for $\text{C}_{42}\text{H}_{40}\text{N}_2\text{O}_6\text{Cu} \cdot 2\text{H}_2\text{O}$: C, 65.65; H, 5.77; N, 3.65%. Found: C, 64.69; H, 5.76; N, 3.54%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3468 (O-H), 3281 (N-H), 3064 (C-H), 2958 (C-H), 2928 (C-H), 1674 (C=O), 1621 (C=C), 1591 (C=C), 1273 (C-O). UV-Vis (CHCl_3) λ/nm , log ϵ : 315 (4.13), 425 (3.78), 538 (2.25).

[Cu(L2) $_2$] (2)

From 380 mg of **HL2**. Yield: 280 mg, 68%; mp 208 °C. Slow evaporation of the complex solution in a methanol/isopropanol mixture yielded brown crystals suitable for X-ray diffraction analysis. Anal. calc. for $\text{C}_{42}\text{H}_{38}\text{N}_4\text{O}_{10}\text{Cu}$. H_2O : C, 60.03; H, 4.80; N, 6.67%. Found: C, 59.21; H, 4.70; N, 6.82%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3460 (O-H), 3273 (N-H), 3077 (C-H), 2956 (C-H), 2930 (C-H), 1675 (C=O), 1617 (C=C), 1592 (C=C), 1546 (δ N-H), 1272 (C-O). UV-Vis (CHCl_3) λ/nm , log ϵ : 298 (4.64), 324 (4.05), 415 (3.72), 536 (2.17).

[Cu(L3) $_2$] (3)

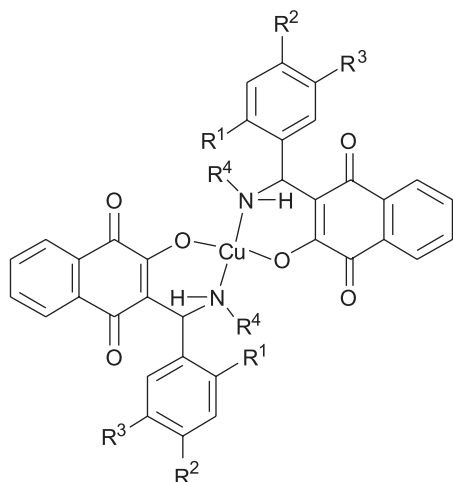
From 414 mg of **HL3**. Yield: 280 mg, 63%; mp 176-177 °C. Anal. Calc. for $\text{C}_{48}\text{H}_{34}\text{N}_4\text{O}_{10}\text{Cu} \cdot 2\text{H}_2\text{O}$: C, 62.23; H, 4.13; N, 6.05%. Found: C, 61.19; H, 4.12; N, 6.16%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3459 (O-H), 3151 (N-H), 2933 (C-H), 1671 (C=O), 1591 (C=C), 1547 (δ N-H), 1274 (C-O). UV-Vis (CHCl_3) λ/nm , log ϵ : 298 (4.51), 319 (4.16), 413 (3.70), 556 (2.17).

[Cu(L4) $_2$] (4)

From 404 mg of **HL4**. Yield: 322 mg, 74%; mp 171 °C. Anal. Calc. for $\text{C}_{42}\text{H}_{36}\text{Cl}_4\text{N}_2\text{O}_6\text{Cu} \cdot 0.5\text{H}_2\text{O}$: C, 57.38; H, 4.24; N, 3.19%. Found: C, 56.49; H, 4.28; N, 3.28%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3446 (O-H); 3273 (N-H), 2959 (C-H), 2931 (C-H); 2871 (C-H), 1678 (C=O), 1625 (C=C), 1591 (C=C), 1548 (δ N-H), 1275 (C-O). UV-Vis (CHCl_3) λ/nm , log ϵ : 289 (4.40), 318 (3.97), 412 (3.70), 530 (2.22).

[Cu(L5) $_2$] (5)

From 438 mg of **HL5**. Yield: 276 mg, 59%; mp 172-173 °C. Anal. Calc. for $\text{C}_{48}\text{H}_{32}\text{Cl}_4\text{N}_2\text{O}_6\text{Cu} \cdot \text{H}_2\text{O}$: C, 60.30; H, 3.58; N, 2.93%. Found: C, 58.87; H, 3.56; N, 3.02%. IR (KBr) $\nu_{\text{max}}/\text{cm}^{-1}$: 3428 (O-H), 3266 (N-H), 3066 (C-H),



- $\text{R}^1 = \text{R}^2 = \text{R}^3 = \text{H}$; $\text{R}^4 = \text{Bu}$, **1**
 $\text{R}^1 = \text{R}^3 = \text{H}$; $\text{R}^2 = \text{NO}_2$; $\text{R}^4 = \text{Bu}$, **2**
 $\text{R}^1 = \text{R}^3 = \text{H}$; $\text{R}^2 = \text{NO}_2$; $\text{R}^4 = \text{Bz}$, **3**
 $\text{R}^1 = \text{R}^2 = \text{Cl}$; $\text{R}^3 = \text{H}$; $\text{R}^4 = \text{Bu}$, **4**
 $\text{R}^1 = \text{R}^2 = \text{Cl}$; $\text{R}^3 = \text{H}$; $\text{R}^4 = \text{Bz}$, **5**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{R}^3 = \text{H}$; $\text{R}^4 = \text{allyl}$, **6**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{R}^3 = \text{H}$; $\text{R}^4 = \text{Bu}$, **7**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{R}^3 = \text{H}$; $\text{R}^4 = \text{Bz}$, **8**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{R}^3 = \text{H}$; $\text{R}^4 = \text{CH}_2\text{C}_4\text{H}_3\text{O}$, **9**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = \text{Me}$; $\text{R}^4 = \text{Bu}$, **10**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = \text{Me}$; $\text{R}^4 = \text{Bz}$, **11**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = \text{Br}$; $\text{R}^4 = \text{Bu}$, **12**
 $\text{R}^1 = \text{OH}$; $\text{R}^2 = \text{H}$; $\text{R}^3 = \text{Br}$; $\text{R}^4 = \text{Bz}$, **13**

Figure 2. Complexes 1-13.

2926 (C-H), 1676 (C=O), 1625 (C=C), 1591 (C=C), 1549 (δ N-H), 1277 (C-O). UV-Vis (CHCl₃) λ /nm, log ϵ : 298 (4.43), 320 (4.03), 411 (3.73), 553 (2.16).

[Cu(L6)₂] (6)

From 335 mg of **HL6**. Yield: 296 mg, 81%; mp > 310 °C. Anal. Calc. for C₄₀H₃₂N₂O₈Cu·0.5H₂O: C, 64.81; H, 4.49; N, 3.78%. Found: C, 63.29; H, 4.60; N, 4.01%. IR (KBr) ν_{\max} /cm⁻¹: 3478 (O-H), 3153 (N-H), 1670 (C=O), 1593 (C=O), 1534 (δ N-H), 1279 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 276 (4.65), 454 (3.63).

[Cu(L7)₂] (7)

From 351 mg of **HL7**. Yield: 306 mg, 80%; mp 201 °C. Slow evaporation of the complex solution in THF/dioxane yielded brown crystals suitable for X-ray diffraction analysis. Anal. Calc. for C₄₂H₄₀N₂O₈Cu·2H₂O: C, 63.03; H, 5.54; N, 3.50%. Found: C, 62.40; H, 5.63; N, 3.41%. IR (KBr) ν_{\max} /cm⁻¹: 3260 (N-H), 2954 (C-H), 2866 (C-H), 1683 (C=O), 1593 (C=C), 1530 (δ N-H), 1276 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 277 (4.58), 455 (3.58).

[Cu(L8)₂] (8)

From 385 mg of **8**. Yield: 287 mg, 69%; mp 187 °C. Anal. Calc. for C₄₈H₃₆N₂O₈Cu·2H₂O: C, 66.39; H, 4.64; N, 3.23%. Found: C, 66.50; H, 4.60; N, 3.31%. IR (KBr) ν_{\max} /cm⁻¹: 3474 (O-H), 3274 (N-H), 3067 (C-H), 2944 (C-H), 1668 (C=O), 1591 (C=C), 1533 (δ N-H), 1279 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 276 (4.63), 449 (3.64).

[Cu(L9)₂] (9)

From 375 mg of **9**. Yield: 260 mg, 64%; mp > 310 °C. Anal. Calc. for C₄₄H₃₂N₂O₁₀Cu·1.5H₂O: C, 62.97; H, 4.20; N, 3.34%. Found: C, 61.93; H, 4.01; N, 3.25%. IR (KBr) ν_{\max} /cm⁻¹: 3478 (O-H), 3245 (N-H), 1669 (C=O), 1591 (C=C), 1534 (δ N-H), 1281 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 276 (4.65), 449 (3.61).

[Cu(L10)₂] (10)

From 365 mg of **10**. Yield: 277 mg, 70%; mp 218 °C. Anal. Calc. for C₄₄H₄₄N₂O₈Cu: C, 66.69; H, 5.60; N, 3.54%. Found: C, 66.27; H, 5.64; N, 3.67%. IR (KBr) ν_{\max} /cm⁻¹: 3254 (N-H), 2954 (C-H), 2868 (C-H), 1693 (C=O), 1590 (C=C), 1500 (δ N-H), 1277 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 277 (4.64), 452 (3.69).

[Cu(L11)₂] (11)

From 399 mg of **11**. Yield: 330 mg, 77%; mp 196 °C. Anal. Calc. for C₅₀H₄₀N₂O₈Cu·H₂O: C, 68.37; H, 4.82; N, 3.19%. Found: C, 67.21; H, 4.68; N, 3.29%. IR (KBr) ν_{\max} /cm⁻¹: 3400 (O-H); 3251 (N-H); 3028 (C-H), 2918 (C-H),

1674 (C=O), 1590 (C=C), 1535 (δ N-H), 1279 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 276 (4.63), 447 (3.63).

[Cu(L12)₂] (12)

From 430 mg of **HL12**. Yield: 152 mg, 33%; mp 186-187 °C. Anal. Calc. for C₄₂H₃₈Br₂N₂O₈Cu·1.5H₂O: C, 53.15; H, 4.35; N, 2.95%. Found: C, 51.75; H, 4.22; N, 3.10%. IR (KBr) ν_{\max} /cm⁻¹: 3422 (O-H), 3253 (N-H), 2959 (C-H), 2933 (C-H), 2869 (C-H), 1675 (C=O), 1589 (C=C), 1532 (δ N-H), 1277 (C-O). UV-Vis (CHCl₃) λ /nm, log ϵ : 276 (4.59), 347 (3.58), 428 (3.58).

[Cu(L13)₂] (13)

From 464 mg of **HL13**. Yield: 294 mg, 60%; mp 202-203 °C. Anal. Calc. for C₄₈H₃₄Br₂N₂O₈Cu·2H₂O: C, 56.18; H, 3.73; N, 2.73%. Found: C, 55.77; H, 3.67; N, 2.80%. IR (KBr) ν_{\max} /cm⁻¹: 3454 (O-H), 3213 (N-H), 2946 (C-H), 1673 (C=O), 1591 (C=C), 1531 (δ N-H), 1279 (C-O). UV-Vis (DMSO) λ /nm, log ϵ : 276 (4.65), 450 (3.65).

X-ray crystallography

The x-ray diffraction data for compounds were collected using a Bruker KAPPA CCD diffractometer,³¹ at 295K and Mo graphite monochromatic radiation. The cell parameters for the molecules were obtained and refined using the PHICHI³² and DIRAX³³ programs, respectively, catching reflections with random orientation in hkl planes. Intensities were corrected by Lorentz polarization and absorption with the SADABS program.³⁴ The structure was solved by Direct Methods using the SHELXS-97 program.³⁵ The anisotropy parameters of non-H atoms were refined with the SHELXL-97 program.³⁶ In **1**, **2** and **7** the aromatic, methyl, methyne and methylene H-atoms were geometrically included in the refinement. Aromatic carbons were refined with U_{iso}(H) = 1.2 Ueq Csp², methylene carbons with U_{iso}(H) = 1.2 Ueq Csp³, methine carbons with U_{iso}(H) = 1.2 Ueq Csp³ and methyl carbons with U_{iso}(H) = 1.5 Ueq Csp³. The hydrogen atom of the water molecules, N-H amine in the three compounds and O-H hydroxyl for **2** were localized experimentally in the Fourier map. For **2** the hydrogen atom coordinates corresponding to a water molecule could not be localized experimentally in the Fourier map. In view of this we opted for using the SQUEEZE³⁷ tool contained in the WinGX³⁸ package, in order to exclude any electronic density contributions relative to the disordered water molecules. This procedure is in accordance with the elemental analysis of the complex, confirming a species free from any crystallization solvate. Consequently we do not comment in this work on the hydrogen bonds for **2**.

The solution and refinement of **1** suggested the presence of disordered C21carbon of the butyl moiety. X-ray data are listed in Table 1 and ORTEP-3³⁹ for Windows was used to draw the Figures.

Antibacterial assays

The antibacterial evaluation was performed with Gram-positive (*Bacillus cereus* ATCC 33019, *Bacillus subtilis* ATCC 6633, *Enterococcus faecalis* ATCC 29212, *Staphylococcus aureus* ATCC 25923) and Gram-negative (*Escherichia coli* ATCC 25922, *Klebsiella pneumoniae* ATCC 700603, *Pseudomonas aeruginosa* ATCC 27853) bacteria as test-microorganisms.

Minimum inhibitory concentration (MIC) was determined by the microdilution broth technique according to the M7-A6 document.⁴⁰ The assays were carried in 96-well tissue culture microplates filled with Mueller Hinton broth (100 μL per well).⁴¹ The inoculum suspension of each strain was prepared in Mueller Hinton broth (108 bacteria cells per mL, corresponding to O. D. = 0.08-0.1 at 625 nm) and diluted to 1:10. All samples were tested in eighth concentrations from 3 to 0.02×10^{-3} mol L⁻¹. The inoculum suspension (5 μL per well) was applied into the microplates which were incubated at 37 °C overnight. An aqueous solution of *p*-iodonitrotetrazolium violet (p-INT) (Sigma) (20 μL) was added⁴² and the microplates were incubated once more for 1-2 hours at 37 °C. The

Table 1. Crystallographic data and refinement parameters for **1**, **2** and **7**

Formula	C ₄₂ H ₄₀ N ₂ O ₆ Cu·2H ₂ O (1)	C ₄₂ H ₃₈ N ₄ O ₁₀ Cu·2C ₃ H ₈ O ₁ (2)	C ₄₂ H ₄₀ N ₂ O ₈ Cu·H ₈ C ₄ O ₂ ·2H ₂ O (7)
Formula weight	768.37	942.49	888.44
T / K	295	295	295
Radiation, λ / Å	0.71073	0.71073	0.71073
Crystal System, space group	Triclinic, <i>P</i> -1	Monoclinic, <i>C</i> 2/c	Monoclinic, <i>P</i> 2 ₁ / <i>n</i>
Unit cell dimensions, <i>a</i> , <i>b</i> , <i>c</i> / Å	<i>a</i> = 9.473(2) <i>b</i> = 10.124(2) <i>c</i> = 11.984(2)	<i>a</i> = 26.187(5) <i>b</i> = 10.382(2) <i>c</i> = 21.158(4)	<i>a</i> = 10.271(2) <i>b</i> = 17.274(4) <i>c</i> = 12.286(3)
α , β , γ / degree	α = 107.30(3) β = 90.81(3) γ = 117.02(3)	β = 115.55(3)	β = 97.97(3)
Volume / Å ³	962.4(3)	5189.6(18)	2158.8(8)
Z, Calculated density / g cm ⁻³	1/1.326	4/1.206	2/1.367
Absorption coefficient / mm ⁻¹	0.622	0.481	0.572
<i>F</i> (000)	403	1980	934
Crystal size / mm ³	0.35 × 0.08 × 0.06	0.30 × 0.24 × 0.17	0.30 × 0.24 × 0.15
Theta range / degree	5.43 to 27.49	2.14 to 25.50	5.13 to 25.50
Index range	-10 ≤ <i>h</i> ≤ 12, -13 ≤ <i>k</i> ≤ 13, -15 ≤ <i>l</i> ≤ 15	-31 ≤ <i>h</i> ≤ 29, -11 ≤ <i>k</i> ≤ 12, -24 ≤ <i>l</i> ≤ 25	-12 ≤ <i>h</i> ≤ 10, -20 ≤ <i>k</i> ≤ 20, -14 ≤ <i>l</i> ≤ 14
Reflections collected	14396	72031	20068
Independent reflections	4361 [<i>R</i> _(int) = 0.0751]	4815 [<i>R</i> _(int) = 0.0763]	3979 [<i>R</i> _(int) = 0.0365]
Completeness to theta max.	98.6%	99.6%	99.0%
Max. and min. transmission	0.8116 and 0.9636	0.9228 and 0.8693	0.9192 and 0.8472
Refinement method	Full-matrix least-squares on <i>F</i> ²	Full-matrix least-squares on <i>F</i> ²	Full-matrix least-squares on <i>F</i> ²
Data / restraints / parameters	4361 / 0 / 258	4815 / 0 / 301	3979 / 0 / 277
Goodness-of-fit on <i>F</i> ²	1.059	1.109	1.060
Final <i>R</i> indices [<i>I</i> > 2 sigma(<i>I</i>)]	<i>R</i> ₁ = 0.0572, <i>wR</i> ₂ = 0.1473	<i>R</i> ₁ = 0.0608, <i>wR</i> ₂ = 0.1469	<i>R</i> ₁ = 0.0465, <i>wR</i> ₂ = 0.1246
<i>R</i> indices (all data)	<i>R</i> ₁ = 0.0851, <i>wR</i> ₂ = 0.1611	<i>R</i> ₁ = 0.0808, <i>wR</i> ₂ = 0.1567	<i>R</i> ₁ = 0.0685, <i>wR</i> ₂ = 0.1429
Extinction coefficient	none	0.0002(2)	None
Largest diff. peak and hole (e ⁻ Å ⁻³)	0.538 and -0.762	0.397 and -0.219	0.900 and -0.722

MIC was defined as the lowest concentration of the extracts that inhibited the antibacterial visible growth as indicated by the p-INT colorimetric reagent. For sterility and growth control the Mueller Hinton broth was used without solvent or compounds. All strains were subcultured twice to verify the cell viability. Tests were performed in triplicate.

Results and Discussion

Syntheses

The Mannich bases **HL1-HL13** (Figure 1) were synthesized from the reactions of 2-hydroxy-1,4-naphthoquinone (lawsone) with an various primary amines and aldehydes in ethanol under stirring at room temperature. The orange products are stable in the solid state, but undergo decomposition when left in solution for a long period of time. Compounds **HL1-HL7** and **HL11-HL13** are obtained in a pure state, but **HL8-HL10** need to be recrystallised from hot ethanol. They were obtained in yields ranging from 53 (**HL4**) to 93% and formulated on the basis of analytical and spectroscopic data (see Experimental).

The ^1H spectra of compounds **HL1-HL13** exhibit peaks due to the four naphthoquinone aromatic hydrogens H5-H8 that appear in the δ 7-8 ppm region as dd or ddd (H5 and H8) and td (H6 and H7) (see Figure 1 for numbering and experimental for data). The other chemical shifts are compatible with the structures proposed for these compounds. In general, the butyl hydrogens appear in the δ 0.9 to 3.1 ppm region as multiplets (H19-H21) or triplets (methyl H22); in the case of the benzyl group; the CH_2 hydrogens H19 appear as a doublet around δ 4.2–4.6 and the phenyl H21-H25, as multiplets. The (substituted) phenyl group hydrogens H13-17 appear as expected, depending on the substitution pattern. Attributions were made on the basis of $^1\text{H} \times ^1\text{H}$ (COSY experiments), J values and multiplicity. All expected resonances were observed in the ^{13}C NMR spectra of compounds **HL1-HL13**. The resonances arising from the carbonyl carbons were found around δ 185 and 179, and those attributed to C2 bound to the hydroxyl group, at about δ 171.

Complexes **1-13** (Figure 2) were obtained by addition of triethylamine to a methanolic suspension of the ligand and $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ (2:2:1), under stirring at room temperature for 12 h in yields varying from 60 to 92%, except for complex **12**, isolated in 30%. Elemental analysis confirmed the proposed formulation. Due to low solubility in methanol, acetonitrile and water, conductivity measurements could not be carried out.

All compounds were also characterized by EPR and UV-Vis spectroscopy, and the structures of **1**, **2** and **7**, determined by X-ray diffraction analyses.

Description of the X-ray structures

Good quality crystals suitable for single crystal X-ray diffraction analyses were obtained for compounds **1**, **2** and **7**. The molecular structures of **1**, **2** and **7** are shown in Figures 3, 4 and 5, respectively and selected bond lengths and angles are given in Table 2.

All complexes crystallize in centrosymmetric space groups, with a copper atom in the inversion centre. Two deprotonated ligands (L^-) coordinate through the naphthalen-2-olate oxygen and secondary amine-N atoms, forming two six-membered chelate rings around the copper atom in a *trans*- N_2O_2 environment. Bond angles $\text{O}(1)\text{-Cu-N}(1)$ (and β parameters:⁴³ 88.43(9), **1**, 91.33(10), **2**, and 90.67(9)°, **7**) and Cu-N and Cu-O distances indicate slightly distorted square-planar coordination of the complexes. The

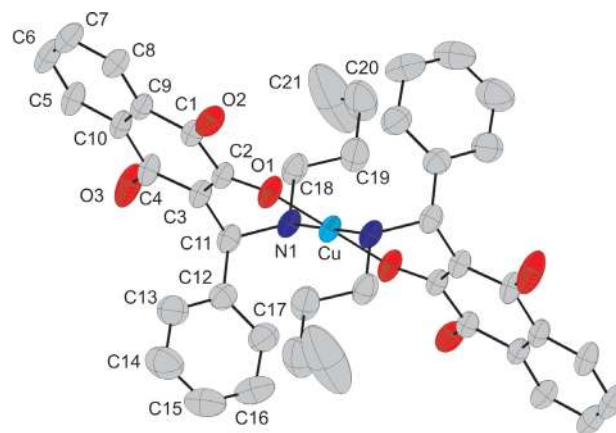


Figure 3. ORTEP view of $[\text{Cu}(\text{L1})_2] \cdot 2\text{H}_2\text{O}$ **1** with labeled atoms and 50% probability ellipsoids; H atoms were omitted for the sake of clarity.

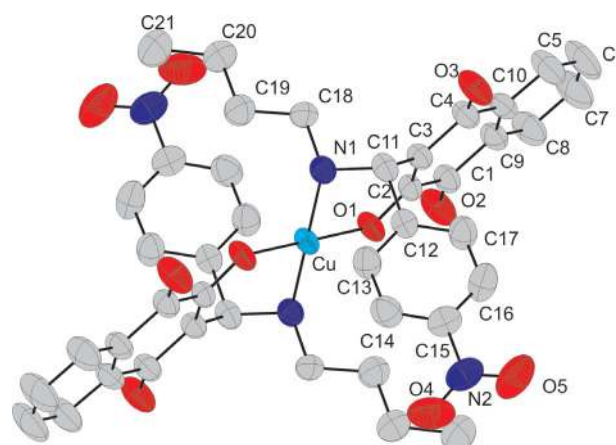


Figure 4. ORTEP view of $[\text{Cu}(\text{L2})_2] \cdot \text{H}_2\text{O}$ **2** with labeled atoms and 50% probability ellipsoids; H atoms were omitted for the sake of clarity.

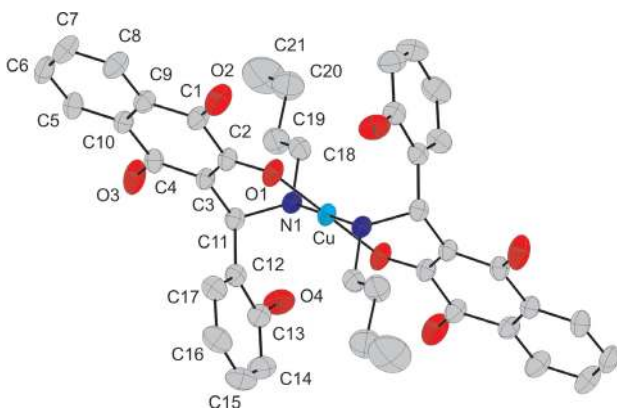


Figure 5. ORTEP view of $[\text{Cu}(\text{L7})_2]\cdot 2\text{H}_2\text{O}$ **7** with labeled atoms and 50% probability ellipsoids; H atoms were omitted for the sake of clarity.

Table 2. Selected bond distances (Å) and angles (°) for complexes **1**, **2** and **7**

Bond distances / Å	1	2	7
Cu-N(1)	2.023(2)	2.023(3)	1.996(2)
Cu-O(1)	1.945(2)	1.928(2)	1.942(2)
N(1)-C(11)	1.509(4)	1.489(4)	1.512(3)
N(1)-C(18)	1.510(4)	1.492(4)	1.495(4)
O(1)-C(2)	1.310(3)	1.300(3)	1.297(3)
O(2)-C(1)	1.230(4)	1.216(4)	1.223(4)
O(3)-C(4)	1.243(4)	1.232(4)	1.239(3)
Bond angles / (°)			
O(1)-Cu-N(1)	88.43(9)	91.33(10)	90.67(9)
Cu-N(1)-C(11)	110.46(17)	109.08(18)	106.52(16)
C(3)-C(11)-N(1)	109.0(3)	110.6(2)	110.0(2)
C(2)-C(3)-C(11)	120.5(2)	120.6(2)	120.6(2)
O(1)-C(2)-C(3)	125.4(2)	125.8(3)	125.6(2)
C(2)-O(1)-Cu	126.86(19)	127.19(18)	126.29(18)
C(11)-N(1)-C(18)	111.0(2)	110.6(2)	114.9(2)

Cu–N_{amine} (2.023(2), **1**, 2.023(3), **2**, and 1.996(2) Å, **7**) and Cu–O_{phenolate} distances (1.945(2), **1**, 1.928(2), **2**, and 1.942(2) Å, **7**) are in the normal range when compared to those observed for other copper(II) complexes containing the same coordination environment.^{44–46} In the same molecule, the two ligands have different absolute configurations at the chiral C11 carbon. In the structures of compounds **1** and **2** the butyl and phenyl groups block the axial positions preventing further coordination to donor molecules (*e.g.* coordinating solvent), normally observed in the structures of analogous complexes.^{47,48}

The packing arrangement of **1** exhibits molecules of **1** and water linked by N–H···O and O–H···O classic hydrogen bonds along the [100] crystallographic direction, resulting in a 1D supramolecular arrangement, Figure 6. The water

molecules are responsible for the 1D self-assembly formed in the solid state, and the network is cemented by bifurcated and linear H-bonds around these molecules. The O4[#]–H4A[#]···O1 and O4[#]–H4A[#]···O2 bifurcated hydrogen bond forms a five membered ring between complex **1** and water, these interactions providing stabilization of the 1D network [symmetry code for (**1**): (#) = $x, 1+y, z$; (##) = $1+x, y, z$]. In addition the crystalline structure is stabilized by C–H···O and C–H···N intramolecular interactions (Figure 7). Table 3 shows all H-bond parameters. All hydrogen bonds were calculated using PLATON³⁷ (Table 3) and agree with the literature.³⁷

The crystalline structure of complex **7** shows a 2D self-arrangement governed by O–H···O and C–H···O classical and non classical hydrogen bonds, respectively. The lattice grows in the [100] direction through O6^{*i*}–H6A^{*i*}···O3, O6^{*i*}–H6B^{*i*}···O2^{*iii*} and O4^{*i*}–H4^{*i*}···O6^{*ii*} interactions and in the [001] direction, through C18–H18B···O5^{*iii*} interactions, as shown in Figure 8 [symmetry code for **7**: (*i*) = $-1-x, -y, 1-z$; (*ii*) = $-x, 1-y, 1-z$; (*iii*) = $-1+x, y, z$, Table 4]. The network in the [100] direction is built around the water molecules and in the [001] direction, *via* the dioxane interactions. In addition, intramolecular interactions *via* N–H···O and C–H···O, specifically the N1–H1···O4 hydrogen bond prevents the copper atom from interacting with the hydroxyl O(4), as illustrated in Figure 9. Similar type of interaction has been observed in an analogous copper compound.⁴⁵ The geometric parameters for these interactions are listed in Table 4. All hydrogen bonds were calculated using PLATON³⁷ and agree with the literature.³⁷

FT-IR spectra

The FT-IR spectra of the complexes show a broad band near 3400 cm⁻¹ assigned to O–H stretching of the water molecules present.⁴⁹ The new bands in the 3200–3300 cm⁻¹ range can be assigned to $\nu_{\text{N-H}}$.^{48–50} These bands were not observed in the spectra of the free ligands, due to the presence of the broad $\nu_{\text{O-H}}$ band centered around 3400 cm⁻¹. This shift in $\nu_{\text{N-H}}$ frequency confirms complexation to the Cu²⁺ center. Several weak bands observed in the 2850–3100 cm⁻¹ range are attributed to aliphatic and aromatic C–H groups.⁴⁹ The strong carbonyl $\nu_{\text{C=O}}$ band around 1680 cm⁻¹ and aromatic ring $\nu_{\text{C=C}}$ bands, around 1590 and 1470 cm⁻¹ were not altered by complexation.⁴⁸ The strong $\nu_{\text{C-O}}$ band around 1280 cm⁻¹ was attributed to the naphthoquinonato group.^{47,51}

EPR Spectra

The EPR spectra of complexes **1–13** were measured in the solid state at liquid nitrogen temperature. The Hamiltonian parameters (Table 5) obtained in

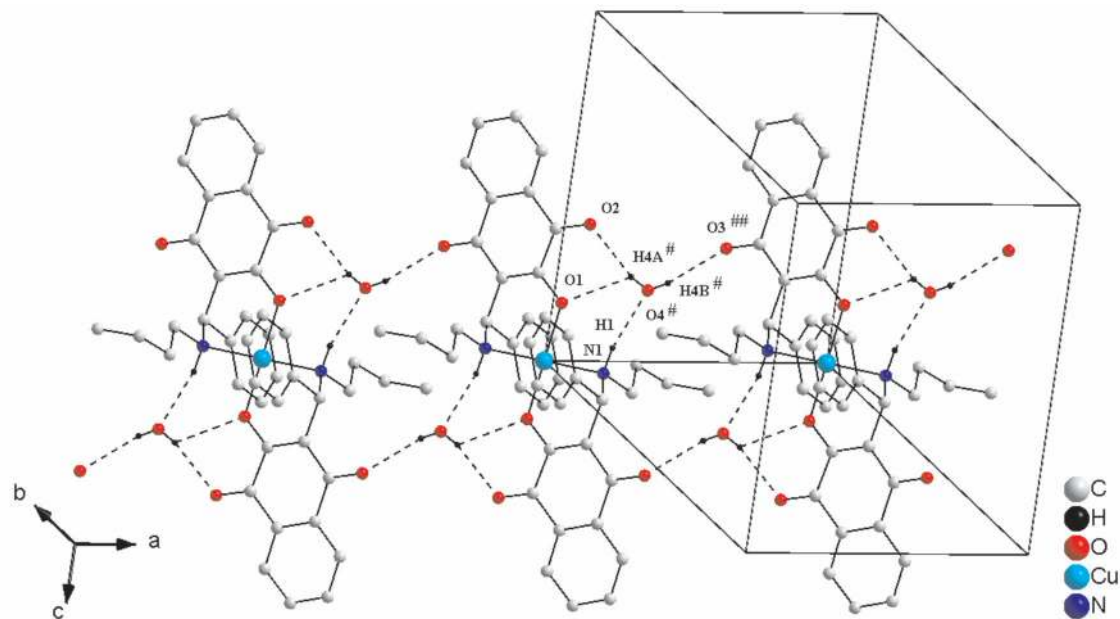


Figure 6. View of the self-assembly 1D by classical hydrogen bonds along [100] crystallography direction. (Symmetry code for **1**: (#) = $x, 1+y, z$; (##) = $1+x, y, z$).

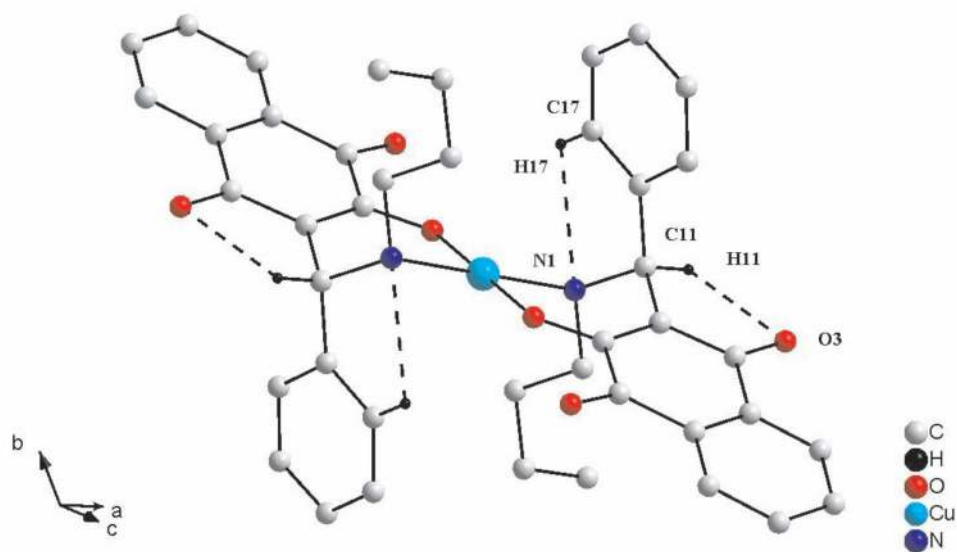


Figure 7. View of intramolecular interactions *via* non classical hydrogen bonds in complex **1**.

Table 3. Geometric parameters for non-classical H bonds for complex **1** (Å, degree)

D-H...A	D-H	H...A	D...A	\angle D-H...A
C(17)-H(17)...N(1) _{intra}	0.93	2.60	2.926(5)	101
C(11)-H(11)...O(3) _{intra}	0.99(4)	2.37(4)	2.856(5)	109(3)
O(4) ^a -H(4A) ^a ...O(1)	0.76(7)	2.56(7)	3.079(5)	126(5)
O(4) ^a -H(4A) ^a ...O(2)	0.76(7)	2.23(6)	2.972(5)	165(6)
O(4) ^a -H(4B) ^a ...O(3) ^b	0.69(7)	2.24(7)	2.913(5)	168(7)
N(1)-H(1)...O(4) ^a	0.88(4)	2.16(4)	3.018(5)	167(4)

Symmetry code for **1**: ^(a) $x, 1+y, z$; ^(b) $1+x, y, z$.

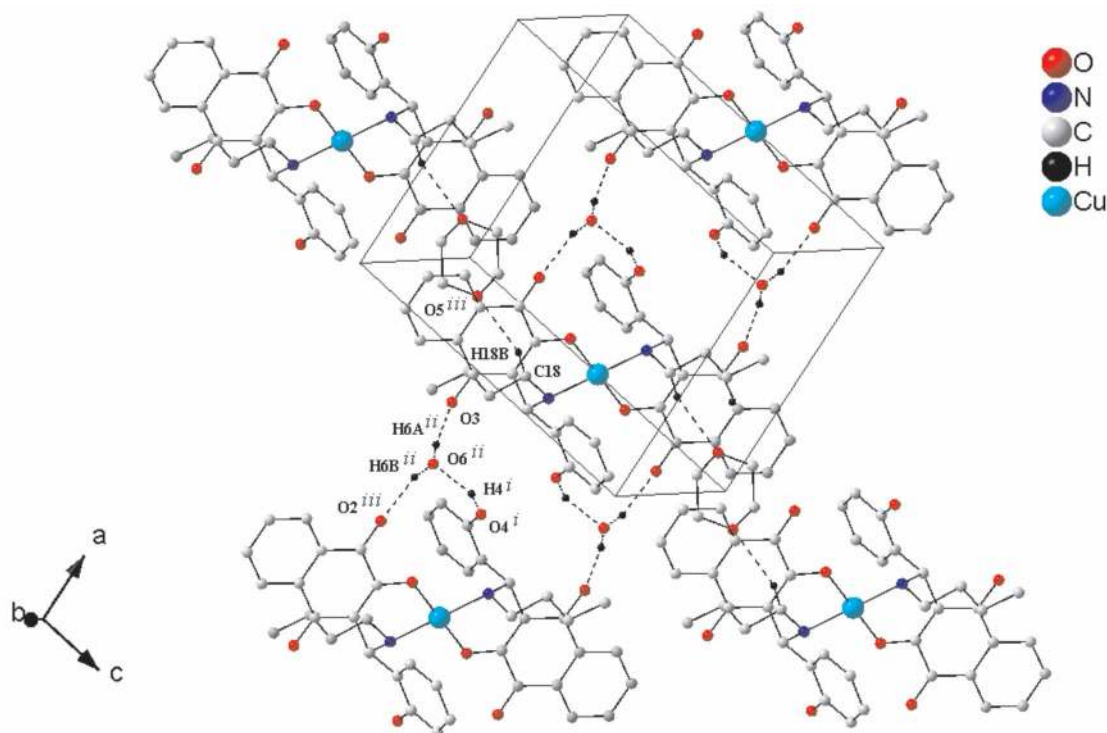


Figure 8. View of the 2D self-arrangement governed by water and dioxane molecules along [100] and [001] crystallography directions, respectively (symmetry code for **7**: (i) = $-1-x, -y, 1-z$; (ii) = $-x, 1-y, 1-z$; (iii) = $-1+x, y, z$).

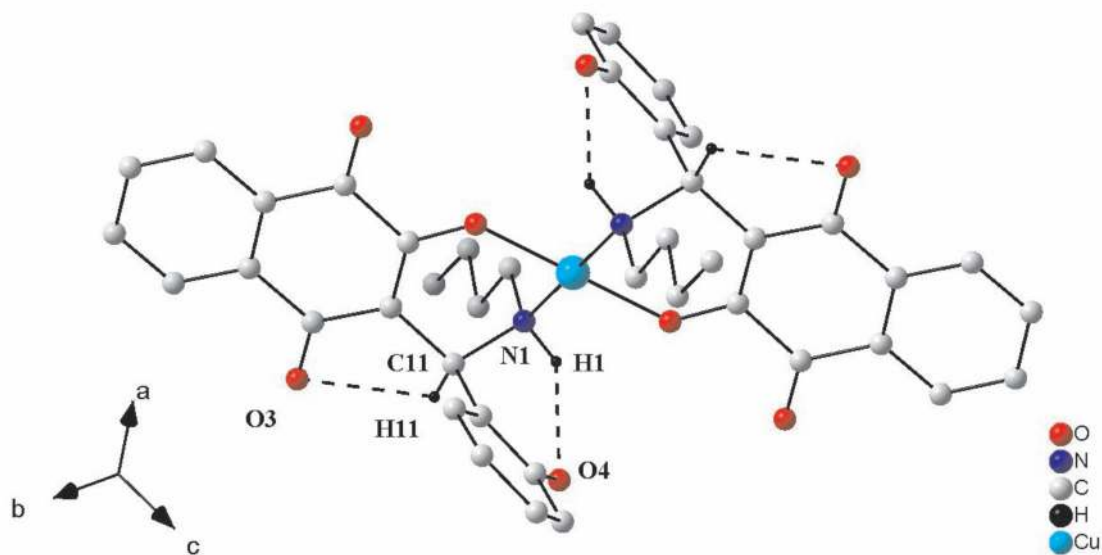


Figure 9. View of the intramolecular interactions in complex **7**.

the simulations of the EPR spectra $g_{\parallel} > g_{\perp} > 2$ and $A_{\parallel} = (188 - 200) \times 10^{-4} \text{ cm}^{-1}$ are typical of elongated octahedral or square planar geometry, suggesting copper(II) sites with axial symmetry.⁵² All complexes, except for **2** and **12**, show values of $g_{\parallel}/A_{\parallel}$ between 110 and 122 thus confirming the square planar environment around the Cu^{2+} centre⁵³ as established for compounds **1**, **2** and **7** by X-ray analysis. The experimental values, $g_{\parallel} > g_{\perp}$, also indicate

that the unpaired electron is predominantly in the dx^2-y^2 orbital, which gives ${}^2B_{1g}$ as the ground state. The very low parallel and perpendicular components of the hyperfine coupling constant for complexes **2** and **12** (not resolved in the spectra) has been explained by considering a mixture of the Cu^{2+} dz^2 and dx^2-y^2 orbitals as the ground state. It is found that a 10% mixture of dx^2-y^2 and dz^2 results in a 20% reduction in dipolar anisotropy.⁵⁴

Table 4. Geometric parameters for non-classical H bonds for complex **7** (Å, degree)

D-H...A	D-H	H...A	D...A	∠D-H...A
N(1)-H(1)···O(4) _{intra}	0.93(4)	2.08(4)	2.818(3)	136(3)
C(11)-H(11)···O(3) _{intra}	0.98	2.40	2.836(4)	107
O(4) ⁱ -H(4) ⁱ ···O(6) ⁱⁱ	0.82	1.95	2.699(3)	151
O(6) ⁱⁱ -H(6A) ⁱⁱ ···O(3)	0.89	1.95	2.833(3)	169
O(6) ⁱⁱ -H(6B) ⁱⁱ ···O(2) ⁱⁱⁱ	0.86	2.03	2.866(3)	163
C(18)-H(18B)···O(5) ⁱⁱⁱ	0.97	2.53	3.461(8)	160

Symmetry code for (**7**): (i) = -1-x, -y, 1-z; (ii) = -x, 1-y, 1-z; (iii) = -1+x, y, z.

Table 5. Spin-Hamiltonian parameters used in the simulated spectra of the copper(II) ion in complexes **1-13** in decreasing order of field strength

Complexes	A _⊥ (×10 ⁻⁴ cm ⁻¹)	A _∥ (×10 ⁻⁴ cm ⁻¹)	g _⊥	g _∥	g _∥ /A _∥
1	15	200	2.0500	2.2070	110
3	25	197	2.0350	2.2100	112
5	27	198	2.0350	2.2100	112
10	25	192	2.0470	2.2055	115
11	25	195	2.0470	2.2350	115
7	20	190	2.0550	2.2070	116
13	25	193	2.0440	2.2380	116
9	25	190	2.0450	2.2380	118
4	25	190	2.0450	2.2380	118
8	25	188	2.0480	2.2380	119
6	25	188	2.0488	2.2400	119
12	----	----	2.0630	2.1880	----
2	----	----	2.0580	2.1880	----

The relatively low g_∥ values of the complexes are consistent with a N₂O₂ environment around the Cu²⁺ ions.⁵² As expected, the ligand field strength depends mainly on the nature of the R⁴ substituent on the nitrogen, the highest ligand field being observed for complexes of ligands containing R⁴ = butyl, independently of the nature of R² and R³. The presence of the hydroxyl group (R¹=OH), however, may lead to a decrease in the ligand field, as observed for complex **7**, compared with complex **1** (Table 5), due to the presence of a O-H...N1 hydrogen bond that reduces the Lewis basicity of N1. This interaction is evidenced in the supramolecular arrangement of the structure of **7**. This interaction may be present in all complexes containing R¹ = OH (**6-13**).

UV-Vis spectra

The electronic spectra of complexes **1-5** and **12**, recorded in CHCl₃ solution, are characterized by two

intense absorptions observed in the 425-315 nm range that are presumably due to a charge-transfer band from the naphthoquinonato moiety to the metal ion, and to ligand based transitions.⁵⁵ The band around 290 nm was not altered by coordination as it corresponds to π-π* transitions of the naphthoquinone ring.⁴⁸⁻⁵⁰ The d-d band appears between 530-550 nm and can be attributed to a ²A_{1g} ← ²B_{1g} transition which supports the square planar geometry for the complexes in solution.⁵⁰

The spectra of complexes **6-11** and **13** were recorded in DMSO solution due to their low solubility in CHCl₃. All exhibit two intense absorption bands: the band observed around 450 nm was assigned to charge-transfer from the naphthoquinonate moiety to the metal ion and that at 277 nm, also present in the spectra of the respective free ligands, to π-π* transitions of the naphthoquinone ring. No d-d transition band was observed, which suggests coordination of DMSO molecules and distorted octahedral coordination environment of the Cu²⁺ ion.

Antibacterial activity

The antibacterial activity of Mannich bases **HL1-HL13** and complexes **1-13** was evaluated against seven strains of bacteria: *Bacillus cereus* (**BC**), *Bacillus subtilis* (**BS**), *Escherichia coli* (**EC**), *Enterococcus faecalis* (**EF**), *Klebsiella pneumoniae* (**KP**), *Pseudomonas aeruginosa* (**PA**) and *Staphylococcus aureus* (**SA**). The results are reported in Table 6 where the MIC values are expressed in $\mu\text{mol L}^{-1}$. Chloramphenicol was used as a positive control in all tests. Compounds 2-hydroxy-1,4-naphthoquinone (**A**), 2-amino-3-hydroxy-1,4-naphthoquinone (**B**) and lapachol (**C**) were also tested for comparison.

Two Mannich bases and four complexes exhibited similar or higher activity than Chloramphenicol against three strains of bacteria (*B. subtilis*, *E. coli* (**EC**) and

S. aureus), whereas the other compounds, including 2-hydroxy-1,4-naphthoquinone (**A**, Entry 14), 2-amino-3-hydroxy-1,4-naphthoquinone (**B**, Entry 15) and lapachol (**C** Entry 16) only inhibited bacterial growth above $200 \mu\text{mol L}^{-1}$; $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ only inhibit bacterial growth above $3000 \mu\text{mol L}^{-1}$.

Mannich bases **HL11** (Entry 20) and **HL13** (Entry 13) strongly inhibited the growth of *E. coli* (at 20 and $40 \mu\text{mol L}^{-1}$, *i.e.* 8 and $22 \mu\text{g mL}^{-1}$, respectively) and *S. aureus* (at $40 \mu\text{mol L}^{-1}$, *i.e.* 19 and $22 \mu\text{g mL}^{-1}$, respectively). As shown in Figure 1, these compounds contain a 2-hydroxyphenyl group ($\text{R}^1 = \text{OH}$) which is substituted at the 5-position with $\text{R}^3 = \text{Me}$ (**HL11**) or Br (**HL13**), respectively, and $\text{R}^4 = \text{Bn}$. Thus, the nature of the R^3 substituent appears to be of lesser importance than that of the lateral chain R^4 , considering that compound **HL10** (Entry 10) with $\text{R}^3 = \text{Me}$ (similar to **HL11**)

Table 6. Antibacterial activity data obtained from the standard dilution techniques of Mannich bases **HL1-HL13**, complexes **1-13**, 2-hydroxy-1,4-naphthoquinone (**A**), 2-amino-3-hydroxy-1,4-naphthoquinone (**B**) and lapachol (**C**), tested against the following strains of bacteria: *Bacillus cereus* (**BC**), *Bacillus subtilis* (**BS**), *Escherichia coli* (**EC**), *Enterococcus faecalis* (**EF**), *Klebsiella pneumoniae* (**KP**), *Pseudomonas aeruginosa* (**PA**) and *Staphylococcus aureus* (**SA**). Positive control: Chloramphenicol

Compound	Entry	BC	BS	EC	EF	KP	PA	SA
HL1	1	> 200	> 200	> 200	> 200	> 200	> 200	> 200
HL2	2	> 200	> 200	> 200	> 200	> 200	> 200	> 200
HL3	3	> 200	> 200	> 200	> 200	> 200	> 200	> 200
HL4	4	> 200	> 200	> 200	> 200	180	> 200	> 200
HL5	5	> 200	> 200	180	> 200	> 200	> 200	> 200
HL6	6	> 200	> 200	180	> 200	> 200	> 200	> 200
HL7	7	> 200	> 200	> 200	> 200	> 200	> 200	> 200
HL8	8	> 200	180	180	> 200	> 200	> 200	180
HL9	9	180	> 200	180	180	> 200	> 200	180
HL10	10	> 200	> 200	> 200	> 200	> 200	> 200	> 200
HL11	11	> 200	90	20	> 200	> 200	> 200	40
HL12	12	180	180	> 200	> 200	> 200	180	180
HL13	13	> 200	180	40	> 200	> 200	> 200	40
A	14	> 200	> 200	> 200	> 200	> 200	> 200	> 200
B	15	> 200	> 200	> 200	> 200	> 200	> 200	> 200
C	16	> 200	> 200	> 200	> 200	> 200	> 200	> 200
Chloramphenicol	17	40-90	20-40	90	90	40-90	40	40-90
1	18	> 200	> 200	> 200	> 200	> 200	> 200	> 200
2	19	> 200	> 200	180	> 200	> 200	> 200	> 200
3	20	> 200	> 200	> 200	> 200	> 200	> 200	> 200
4	21	> 200	> 200	> 200	> 200	> 200	> 200	> 200
5	22	> 200	> 200	180	> 200	> 200	> 200	> 200
6	23	> 200	> 200	90	> 200	> 200	> 200	> 200
7	24	> 200	> 200	> 200	> 200	> 200	> 200	> 200
8	25	> 200	> 200	> 200	> 200	> 200	> 200	> 200
9	26	180	> 200	180	> 200	> 200	> 200	> 200
10	27	180	180	90	> 200	> 200	> 200	180
11	28	180	90	180	180	180	180	90
12	29	180	90	90	> 200	> 200	> 200	90
13	30	> 200	90	40	> 200	> 200	> 200	90
$\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$	31	>3000	>3000	>3000	>3000	>3000	>3000	>3000

Minimum inhibitory concentrations (MICs) are reported in $\mu\text{mol L}^{-1}$.

and $R^4 = \text{Bu}$ is much less active than **HL11** against all strains of bacteria. Solubility differences might be responsible for the changes in activity.

With a few exceptions, the complexes were less active than the respective pro-ligands, which is probably due to their lower solubility. Thus, the activity of **HL11** (Entry 11) decreased upon complexation, from 20 to 90 $\mu\text{mol L}^{-1}$ for **11** (*E. coli*) and from 20 to 90 $\mu\text{mol L}^{-1}$ for **11** (*S. aureus*) (Entry 28), although slight increase in growth inhibition of all the other bacteria strains was observed (from $>200 \mu\text{mol L}^{-1}$ for **HL11** to 180 $\mu\text{mol L}^{-1}$ for **11**).

Improvement of the activity of **HL10** and **H12** that only inhibited the growth of all strains of bacteria above 180-200 $\mu\text{mol L}^{-1}$ (Entries 10 and 12) was observed upon complexation: complex **10** (Entry 27) exhibits slight activity against *B. cereus*, *B. subtilis* and *S. aureus* (180 $\mu\text{mol L}^{-1}$) and growth inhibition against *E. coli* (90 $\mu\text{mol L}^{-1}$), and complex **12**, against *E. coli*, *E. faecalis* and *S. aureus* above 90 $\mu\text{mol L}^{-1}$ (Entry 29). Complexes **10** and **12** were formed from Mannich bases **HL10** and **HL12** that only differed from the very active ones, **HL11** and **HL13** with respect to the R^4 group, butyl, instead of benzyl.

The effect of metal complexation on naphthoquinone antimicrobial agents has been discussed in the literature.⁵⁶ Although metal chelation of the anion of 5-hydroxy-1,4-naphthoquinone (juglone) has resulted in complexes with similar antibacterial effect⁵⁷ or higher antibacterial activity, e.g. against *Bacillus ssp* and *S. aureus*, than juglone, complexation of the anions of a series of 5-amino-8-hydroxy-1,4-naphthoquinones with M^{2+} ($M = \text{Ni, Co, Fe, Cu}$ and Cr) resulted in reduced bacterial activity or lack of inhibition effect.⁵⁵ Considering that redox active metals have been shown to be instrumental in naphthoquinone toxicity,¹¹ the decreased activity observed in our work and by others is probably associated with decreased bioavailability of the aminonaphthoquinones as the result of decreased solubility upon complexation.

Conclusions

In conclusion, of the thirteen novel aminonaphthoquinones **HL1-HL13** synthesized from lawsone, via the Mannich reaction, and their respective copper(II) complexes $[\text{Cu}(\text{L1})_2] - [\text{Cu}(\text{L13})_2]$, those containing a 2-hydroxyphenyl group ($R^1 = \text{OH}$) which is substituted at the 5-position, either with $R^3 = \text{Me}$ or Br , and with $R^4 = \text{benzyl}$ or butyl groups have strongly inhibited the growth especially of *Escherichia coli* and *Staphylococcus aureus*. In general complexes were found to be less active than the respective pro-ligands, probably due to their lower solubility. Further

work is in progress to improve the bioavailability of the complexes of **HL10-HL13**.

Supplementary Information

Supplementary data are available free of charge at <http://jbcs.sbq.org.br>, as PDF file.

Crystallographic data for the structural analysis of the three complexes have been deposited with the Cambridge Crystallographic Data Center, CCDC No. 703509 (**1**), 703510 (**2**) and 703511 (**7**). Copies of this information may be obtained free of charge from The Director, CCDC, 12 Union Road, Cambridge, CB2 1EZ, UK (fax: +44 1233336 033; e-mail: deposit@ccdc.cam.ac.uk).

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References

1. Thompson, R. H.; *Naturally Occurring Quinones IV: Recent Advances*, Chapman & Hall: London, 1997.
2. Sacau, E. P.; Braun, A. E.; Ravelo, A. G.; Ferro, E. A.; Tokuda, H.; Mukainaka, T.; Nishino, H.; *Bioorg. Med. Chem.* **2003**, *11*, 483.
3. Chen, J.; Huang, Y.; Liu, G.; Afrasiabi, Z.; Sinn, E.; Padhye, S.; Ma, Y.; *Toxicol. Appl. Pharmacol.* **2004**, *194*, 40.
4. Ferreira, V. F.; Jorquera, A.; Souza, A. M. T.; Silva, M. N.; de Souza, M. C. B. V.; Gouvêa, R. M.; Rodrigues, C. R.; Pinto, A. V.; Castro, H. C.; Santos, D. O.; Araújo, H. P.; Bourguignon, S. C.; *Bioorg. Med. Chem.* **2006**, *14*, 5459.
5. Santos, A. F.; Ferraz, P. A. L.; Pinto, A. V.; Pinto, M. C. F. R.; Goulart, M. O. F.; Sant'Ana, A. E. G.; *Int. J. Parasitol.* **2000**, *30*, 1199.
6. Santos, E. V. M.; Carneiro, J. W. M.; Ferreira, V. F.; *Bioorg. Med. Chem.* **2004**, *12*, 87.
7. Kayser, O.; Kiderlen, A. F.; Laatsch, H.; Croft, S.; *Acta Trop.* **2000**, *77*, 307.
8. Gafner, S.; Wolfender, J. L.; Nianga, M.; Stoeckli-Evans, H.; Hostettman, K.; *Phytochemistry* **1996**, *42*, 1315.
9. Machado, T. B.; Pinto, A. V.; Pinto, M. C. F. R.; Leal, I. C. R.; Silva, M. G.; Amaral, A. C. F.; Kuster, R. M.; Netto-dosSantos, K. R.; *Int. J. Antimicrob. Agents* **2003**, *21*, 279.
10. Medina, L. F. C.; Hertz, P. F.; Stefani, V.; Henriques, J. A. P.; Zannotto-Filho, A.; Brandelli, A.; *Biochem. Cell Biol.* **2006**, *84*, 720.
11. Zang, R.; Hirsh, O.; Mohsen, M.; Samuni, A.; *Arch. Biochem. Biophys.* **1994**, *312*, 385.

12. Vargas, M. D.; Pinto, A. C.; Echevarria, A.; Esteves-Souza, A.; Camara, C. A.; Cunha, A. C.; Torres, J. C.; Lima, E. L. S.; *J. Braz. Chem. Soc.* **2006**, *17*, 439.
13. Esteves-Souza, A.; Figueiredo, D. V.; Esteves, A.; Câmara, C. A.; Vargas, M. D.; Pinto, A. C.; Echevarria, A.; *Braz. J. Med. Biol. Res.* **2007**, *30*, 1399.
14. Cunha, A. S.; Vargas, M. D.; Gattass, C. R.; Pinto, A. C.; Camara, C. A.; Esteves, A. S.; Lima, E. L. S.; *Oncol. Rep.* **2008**, *20*, 225.
15. Silva, M. S.; Camara, C. A.; Barbosa, T. P.; Soares, A. Z.; Cunha, L. C.; Pinto, A. C.; Vargas, M. D.; *Bioorg. Med. Chem.* **2005**, *13*, 193.
16. Barbosa, T. P.; Camara, C. A.; Silva, T. M. S.; Martins, R. M.; Pinto, A. C.; Vargas, M. D.; *Bioorg. Med. Chem.* **2005**, *13*, 6464.
17. Camara, C. A.; Silva, T. M. S.; Silva, T. G.; Barbosa, T. P.; Martins, R. M.; Vargas, M. D.; Pinto, A. C.; *An. Acad. Bras. Ciênc.* **2008**, *80*, 329.
18. Tandon, V. K.; Yadav, D. B.; Singh, R. V.; Chaturvedi, A. K.; Shukla, P. K.; *Bioorg. Med. Chem. Lett.* **2005**, *15*, 5324.
19. Tandon, V. K.; Yadav, D. B.; Chaturvedi, A. K.; Shukla, P. K.; *Bioorg. Med. Chem. Lett.* **2005**, *15*, 3288.
20. Riffel, A.; Medina, L. F.; Stefani, V.; Santos, R. C.; Bizani, D.; Brandelli, A.; *Braz. J. Med. Biol. Res.* **2002**, *35*, 811.
21. Anaconda, J. R.; Moreno, A.; *Main Group Met. Chem.* **1999**, *22*, 573.
22. Tandon, V. K.; Vaish, M.; Khanna, J. M.; Anand, N.; *Arch. Pharmazie* **1990**, *323*, 383.
23. Leffer, M. T.; Hathaway, R. J.; *J. Am. Chem. Soc.* **1948**, *70*, 3222.
24. Baramée, A.; Coppin, A.; Mortuaire, M.; Pelinski, L.; Tomavo, S.; Brocard, J.; *Bioorg. Med. Chem.* **2006**, *14*, 1294.
25. Santos, A. F.; Ferraz, P. A. L.; Pinto, A. V.; Pinto, M. C. F. R.; Goulart, M. O. F.; Sant'Ana, A. E. G.; *Int. J. Parasitol.* **2000**, *30*, 1199.
26. Zang, O.; Hirsh, M.; Mohsen, A.; Samuni; *Arch. Biochem. Biophys.* **1994**, *312*, 385.
27. Gokhale, N. H.; Shirisha, K.; Padhye, S. B.; Croft, S. L.; Kendrick, H. D.; Mckee, V.; *Bioorg. Med. Chem. Lett.* **2006**, *16*, 430.
28. Gokhale, N. H.; Shirisha, K.; Padhye, S. B.; Croft, S. L.; Kendrick, H. D.; Davies, W.; Anson, C. E.; Powell, A. K.; *J. Inorg. Biochem.* **2003**, *95*, 249.
29. Chen, J.; Huang, Y.; Liu, G.; Afrasiabi, Z.; Sinn, E.; Padhye, S.; Ma, Y.; *Toxicol. Appl. Pharmacol.* **2004**, *197*, 40.
30. Dalgliesh, C.E.; *J. Am. Chem. Soc.* **1949**, *71*, 1697.
31. Nonius; *COLLECT*, Nonius BV: Delft, The Netherlands, 1998.
32. Duisenberg, A. J. M.; Hooft, R. W. W.; Schreurs, A. M. M.; Kroon, J.; *J. Appl. Crystallogr.* **2000**, *33*, 893.
33. Duisenberg, A. J. M.; *J. Appl. Crystallogr.* **1992**, *25*, 92.
34. Sheldrick, G. M.; *SADABS; Program for Empirical Absorption Correction of Area Detector Data*, University of Göttingen: Germany, 1996.
35. Sheldrick, G. M.; *SHELXS97; Program for Crystal Structure Solution*, University of Göttingen: Germany, 1997.
36. Sheldrick, G. M.; *SHELXL97; Program for Crystal Structure Refinement*, University of Göttingen: Germany, 1997.
37. Spek, A. L.; *J. Appl. Crystallogr.* **2003**, *36*, 7.
38. Farrugia, L. J.; *J. Appl. Crystallogr.* **1999**, *32*, 837.
39. Farrugia, L. J.; *J. Appl. Crystallogr.* **1997**, *30*, 565.
40. *National Committee for Clinical Laboratory Standards Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically*, 6th ed.; Approved Standard NCCLS Document M7-A6: Wayne, PA, 2003.
41. Langfield, R. D.; Scarano, F. J.; Heitzman, M. E.; Kondo, M.; Hammond, G. B.; Neto, C. C.; *J. Ethnopharmacol.* **2004**, *94*, 279.
42. Eloff, J. N.; *Planta Med.* **1998**, *65*, 711.
43. Addison, A. W. In *Copper Coordination Chemistry: Biochemical and Inorganic Perspectives*; Karlin, K. D.; Zubieta, J., eds., Academic Press: New York, 1983.
44. Muppidi, V. K.; Das, S.; Raghavaiah, P.; Pal, S.; *Inorg. Chem. Commun.* **2007**, *10*, 234.
45. Xie, Y.; Ni, J.; Liu, X.; Liu, Q.; Xu, X.; *Transition Met. Chem.* **2003**, *28*, 367.
46. Thakuria, H.; Das, G.; *Polyhedron* **2007**, *26*, 149.
47. Xie, Y.; Bu, W.; Chan, A. S. -C.; Xu, X.; Liu, Q.; Zhang, Z.; Yu, J.; Fan, Y.; *Inorg. Chim. Acta* **2000**, *310*, 257.
48. Muppidi, V. K.; Zacharias, P. S.; Pal, S.; *J. Solid State Chem.* **2007**, *180*, 132.
49. Muppidi, V. K.; Das, S.; Raghavaiah, P.; Pal, S.; *Inorg. Chem. Commun.* **2007**, *10*, 234.
50. Thakuria, H.; Das, G.; *Polyhedron* **2007**, *26*, 149.
51. Xie, Y.; Bu, W.; Chan, A. S. -C.; Xu, X.; Liu, Q.; Zhang, Z.; Yu, J.; Fan, Y.; *Inorg. Chim. Acta* **2000**, *310*, 257.
52. Neves, A.; Rossi, L. M.; Bortoluzzi, A. J.; Mangrich, A. S.; Haase, W.; Werner, R.; *J. Braz. Chem. Soc.* **2001**, *12*, 747.
53. Sakaguchi, U.; Addison, A. W.; *J. Chem. Soc., Dalton Trans.* **1979**, 600.
54. Mendes, I. C.; Moreira, J. P.; Speziali, N. L.; Mangrich, A. S.; Takahashi, J. A.; Beraldo, H.; *J. Braz. Chem. Soc.* **2006**, *17*, 1571.
55. Ma, S.; Zhu, W.; Xu, M.; Wang, Y.; Guo, Q.; Liu, Y.; *Polyhedron* **2003**, *22*, 3249.
56. Brandelli, A.; Bizani, D.; Martinelli, M.; Stefani, V.; Gerbase, A. E.; *Braz. J. Pharm. Sci.* **2004**, *40*, 247.
57. Joshi, C. R.; Jagtap, G. S.; Chalgen, S. V.; *Indian J. Pharmaceut. Sci.* **1988**, *50*, 107.
58. Bakolachristianopoulou, M. N.; Ecateriniadou, L. B.; Sarris, K. J.; *Eur. J. Med. Chem.* **1986**, *21*, 385.

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Novel Aminonaphthoquinone Mannich Bases Derived from Lawsone and their Copper (II) Complexes: Synthesis, Characterization and Antibacterial Activity

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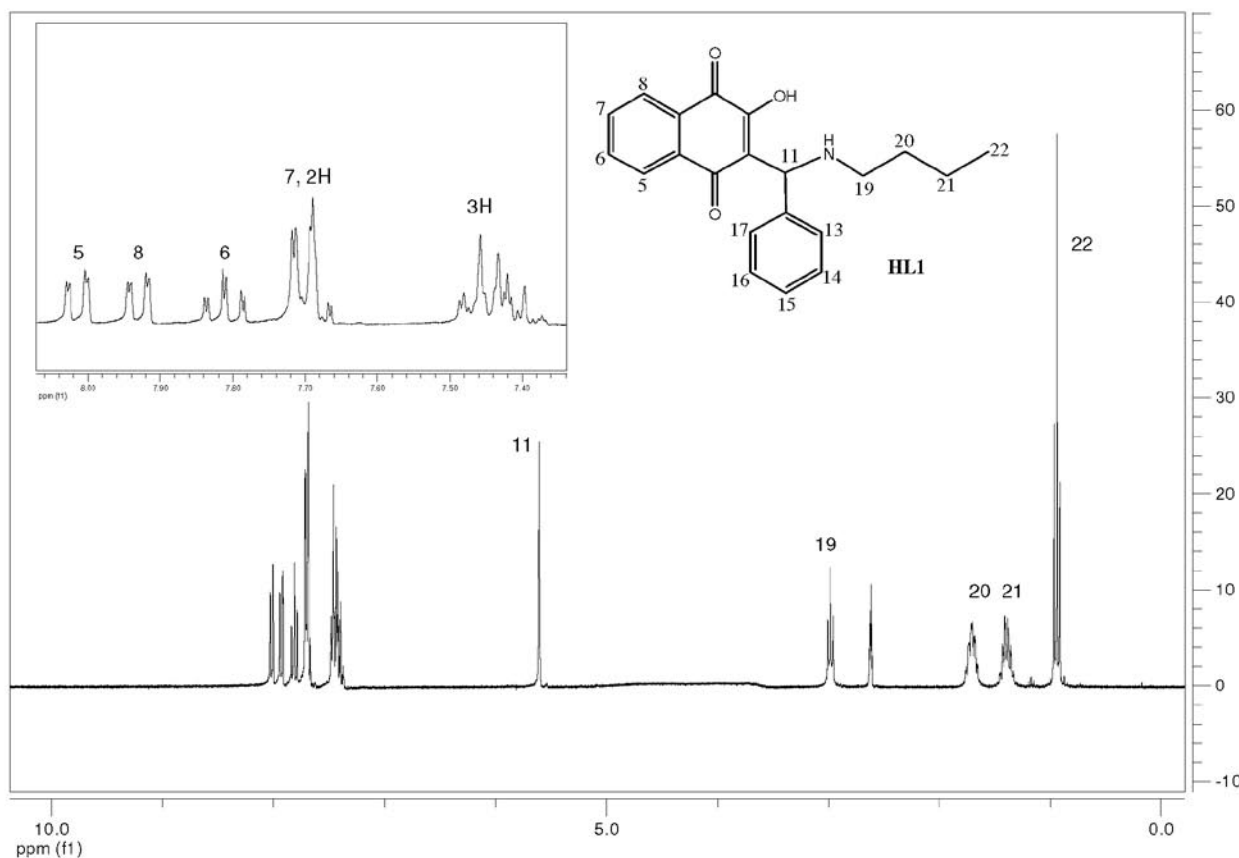


Figure S1. ¹H NMR spectrum of HL1.

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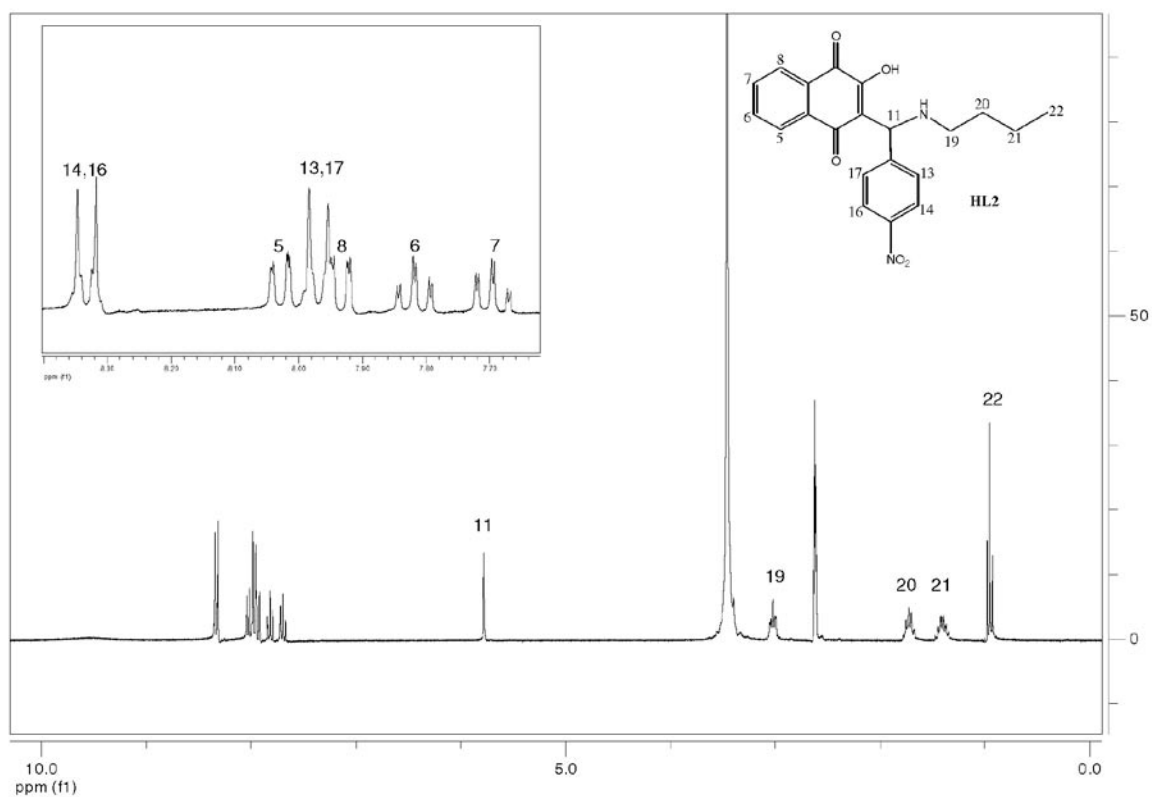


Figure S2. ¹H NMR spectrum of HL2.

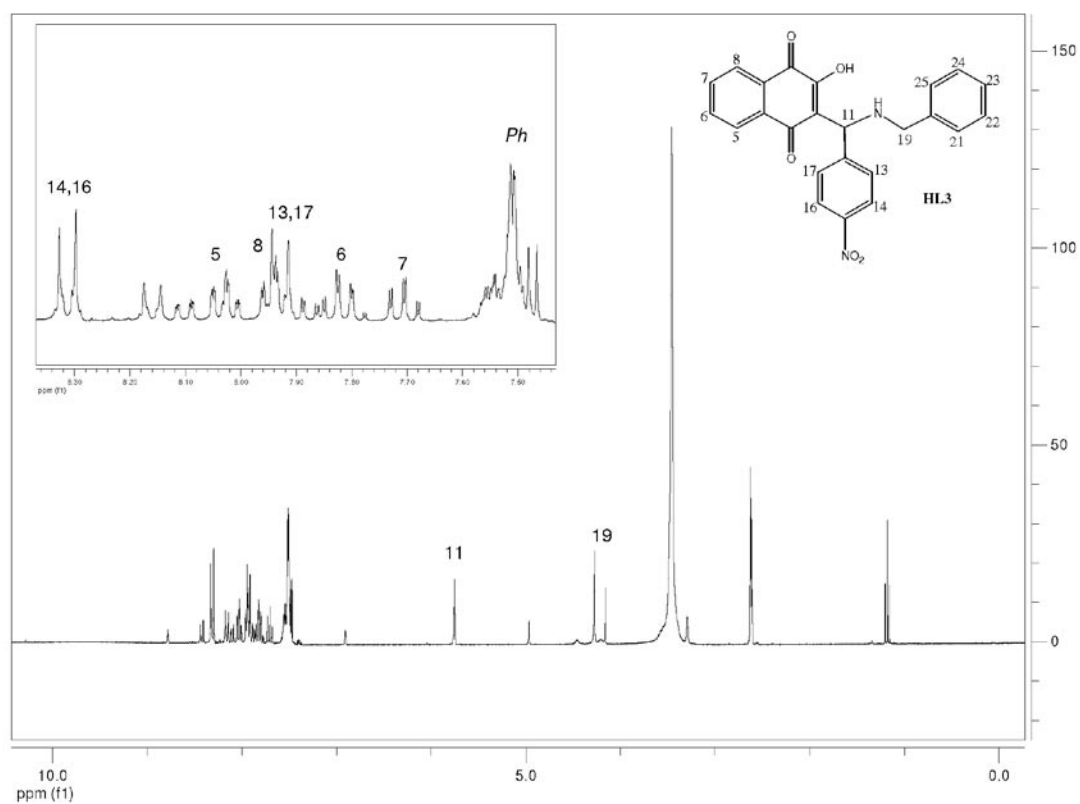
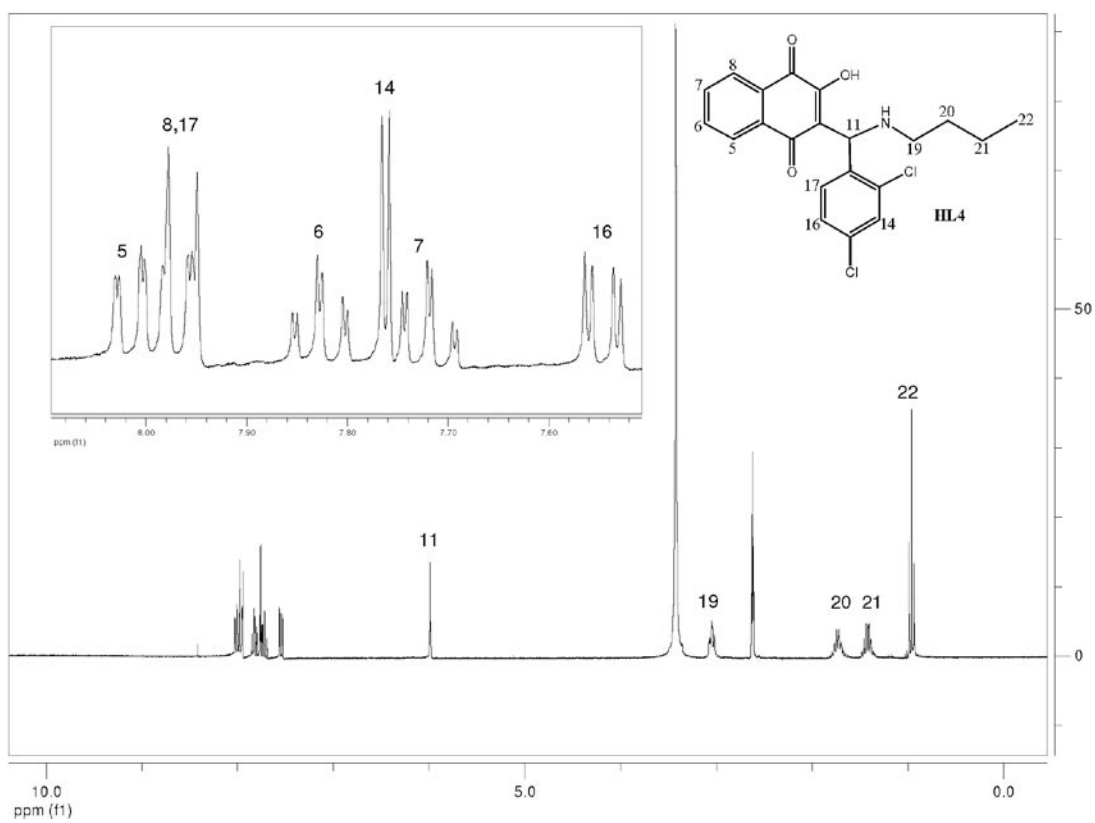
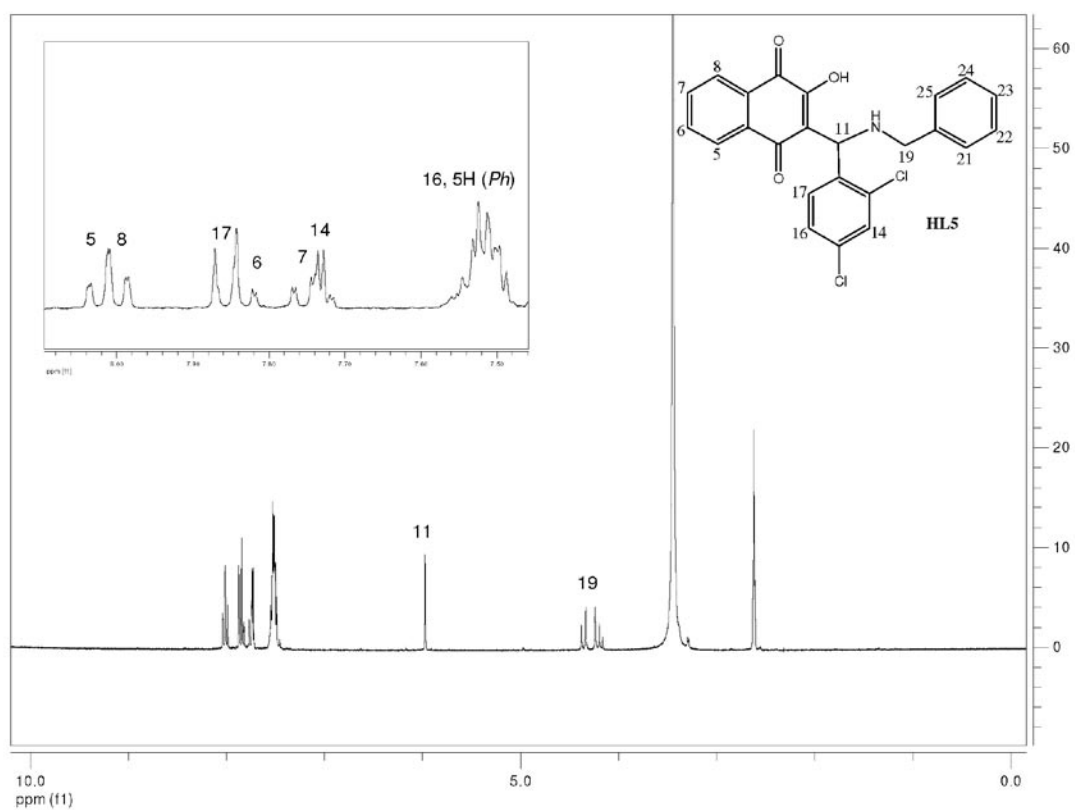


Figure S3. ¹H NMR spectrum of HL3.

Figure S4. ¹H NMR spectrum of HL4.Figure S5. ¹H NMR spectrum of HL5.

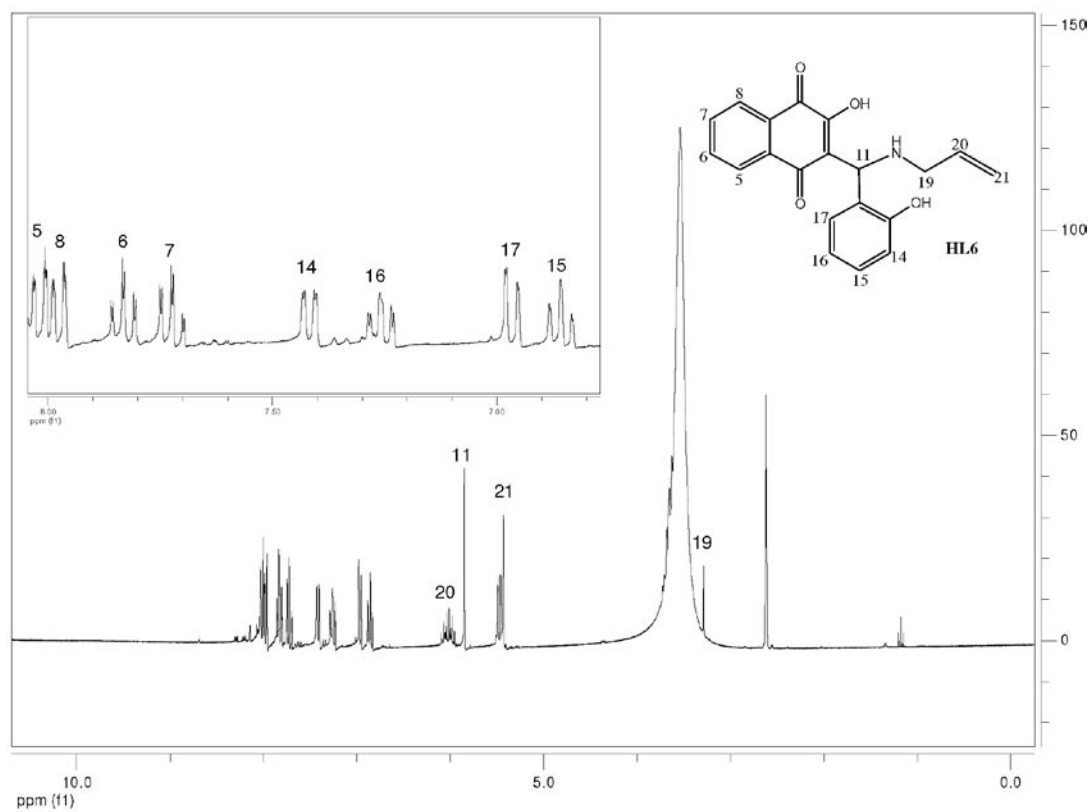


Figure S6. ¹H NMR spectrum of HL6.

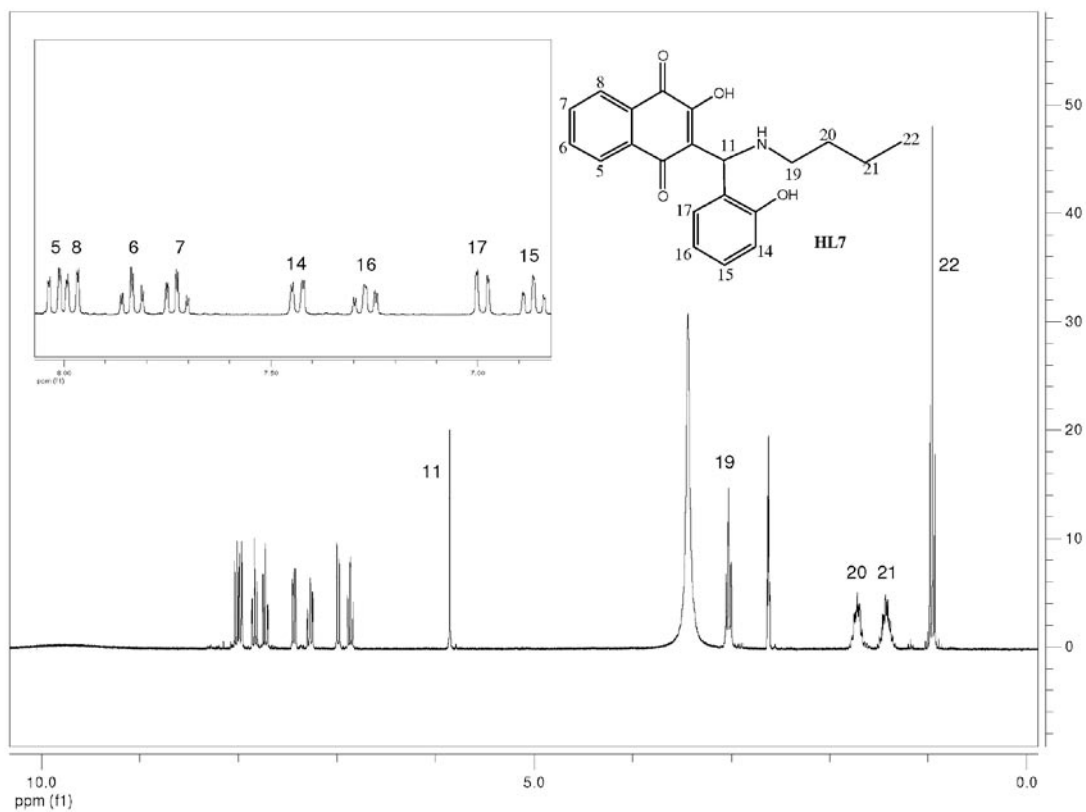
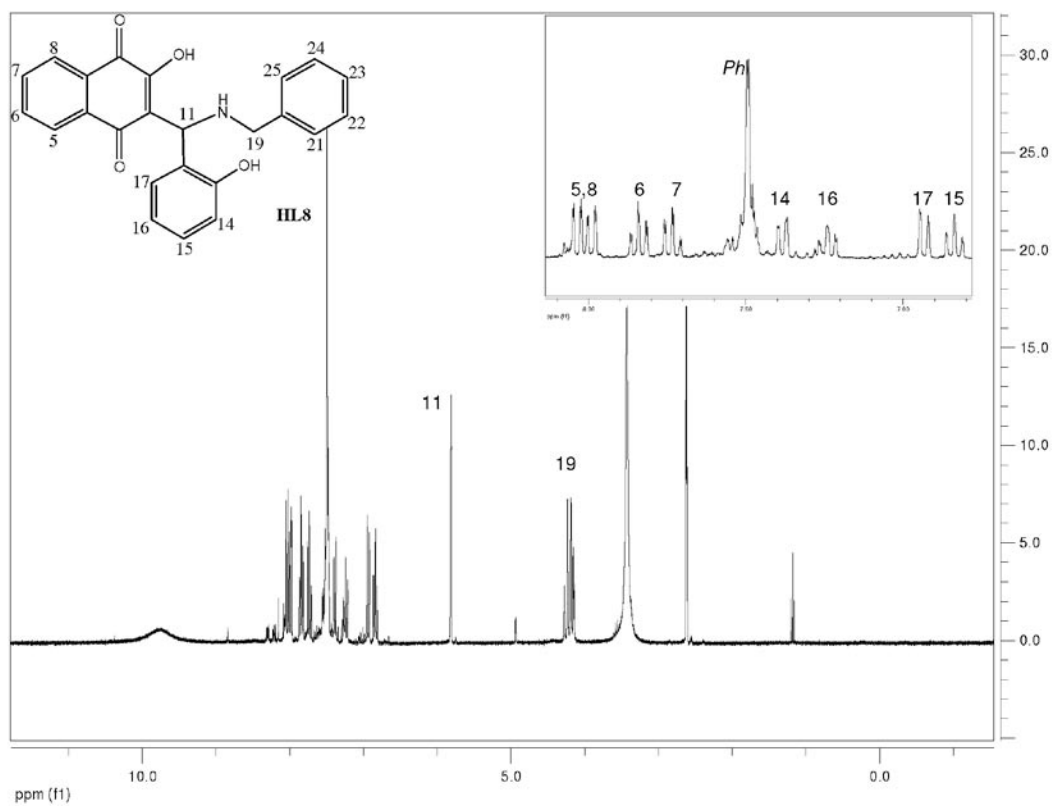
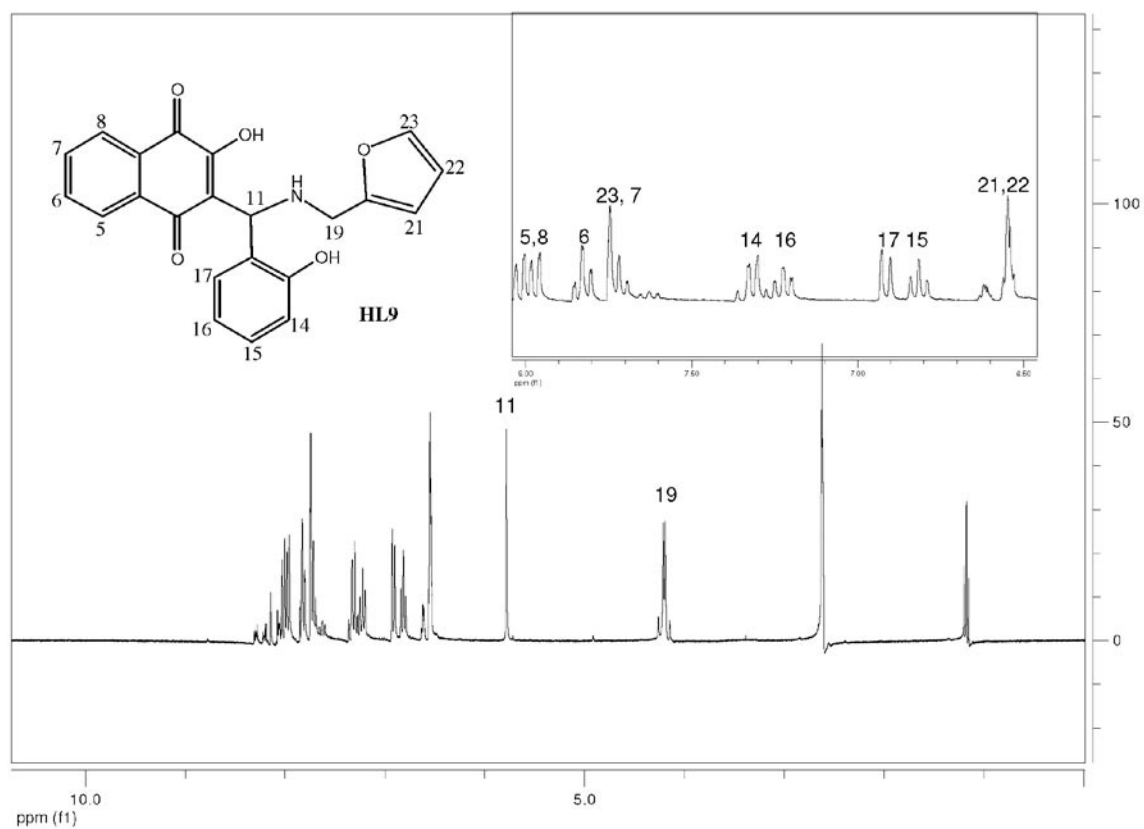
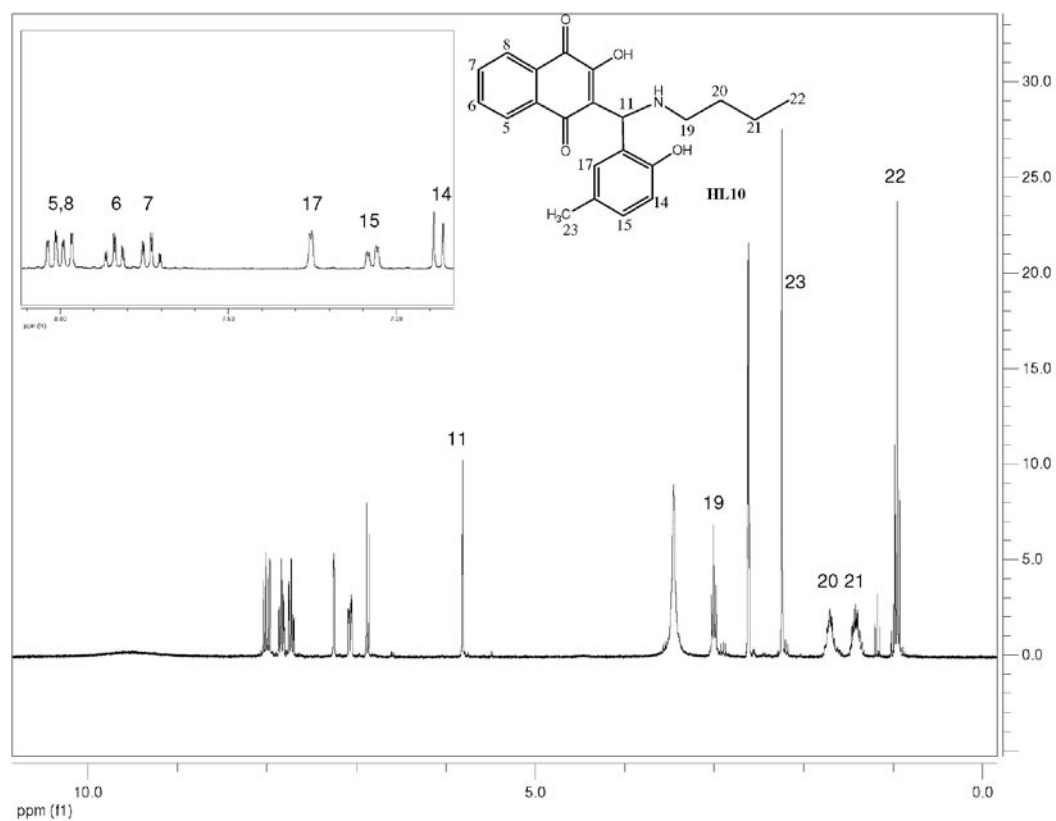
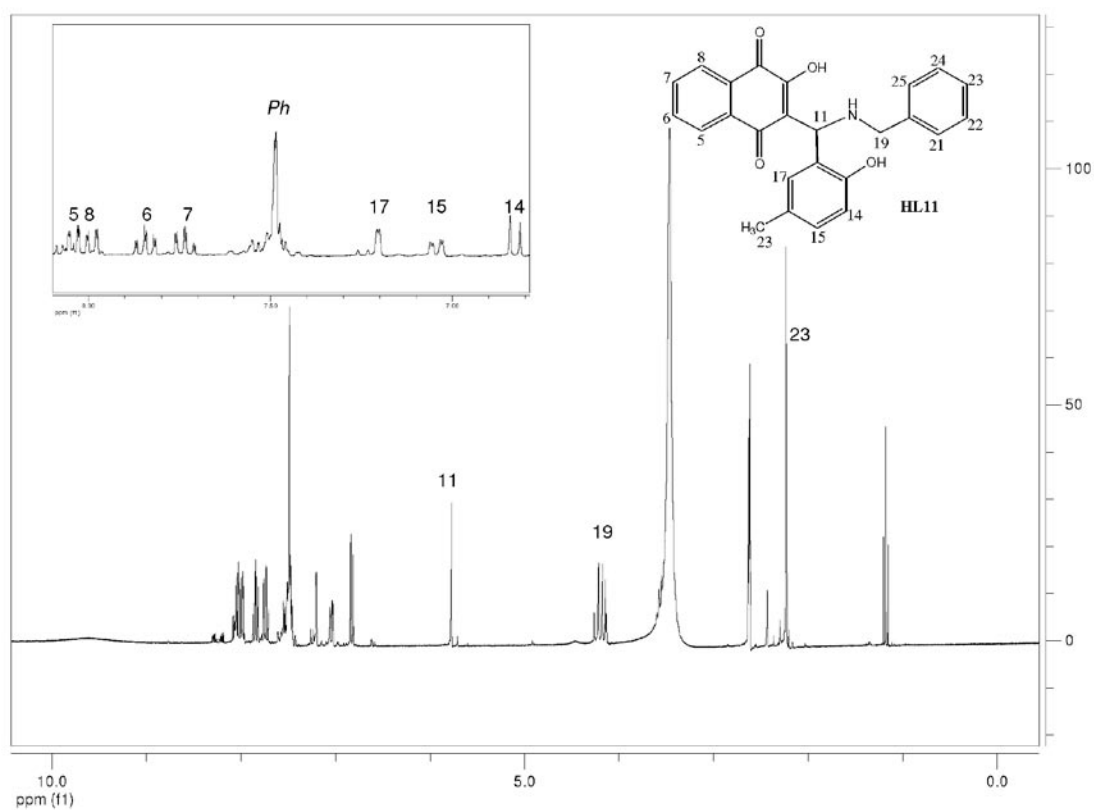
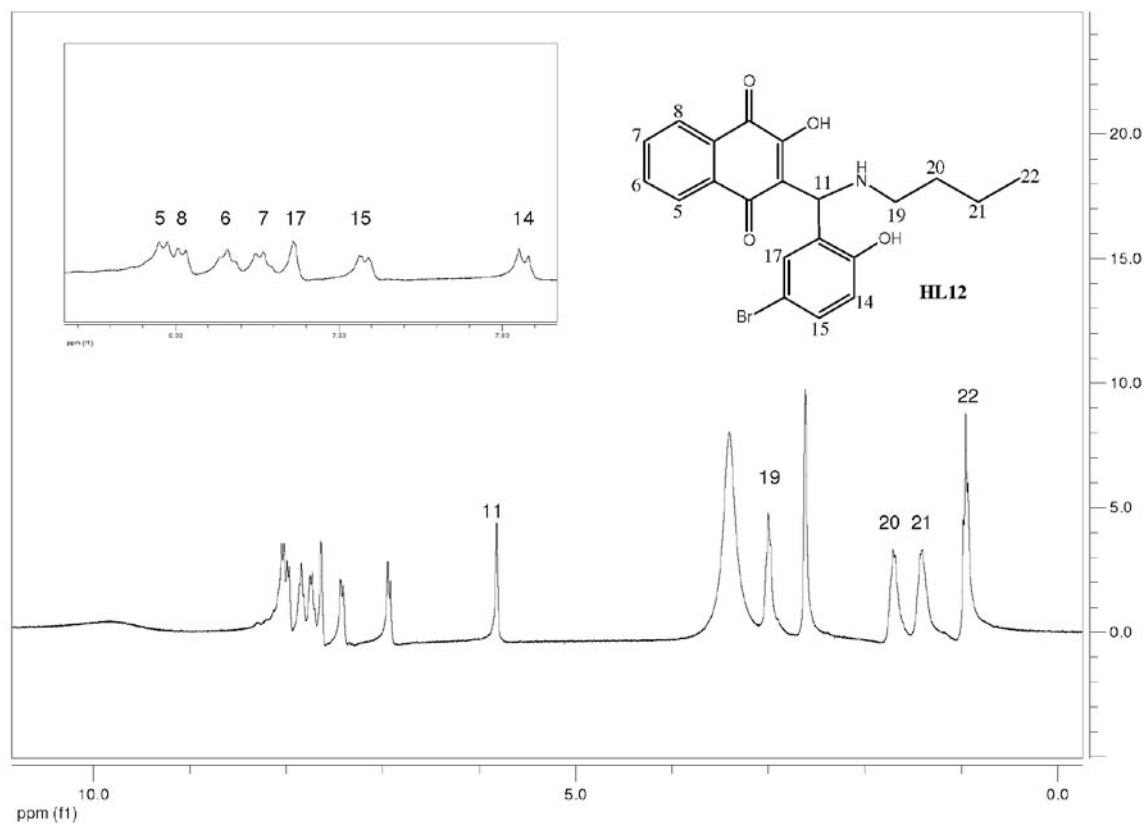
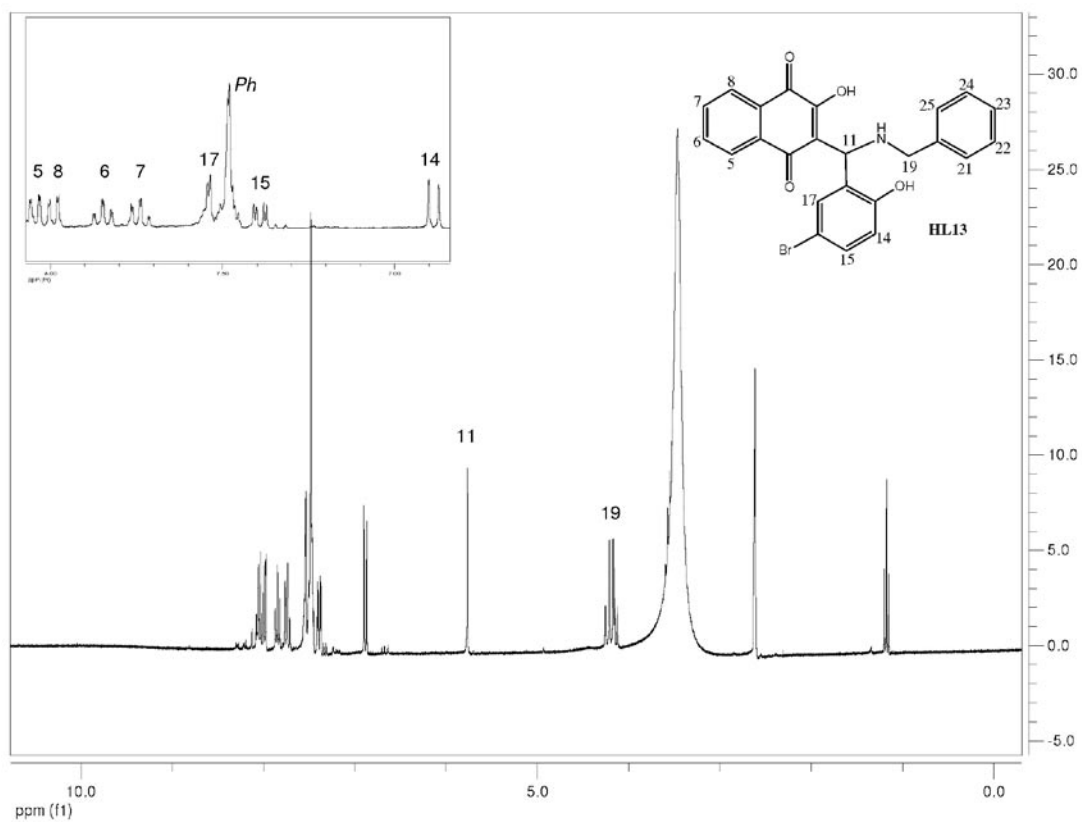
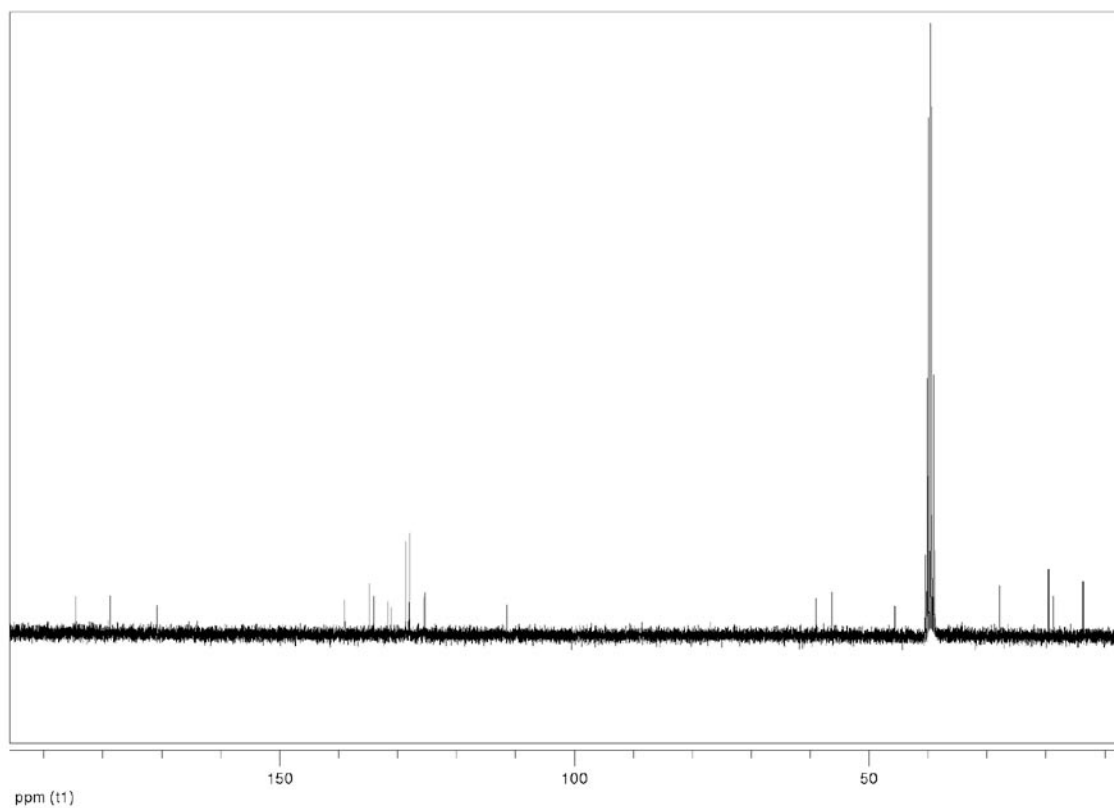
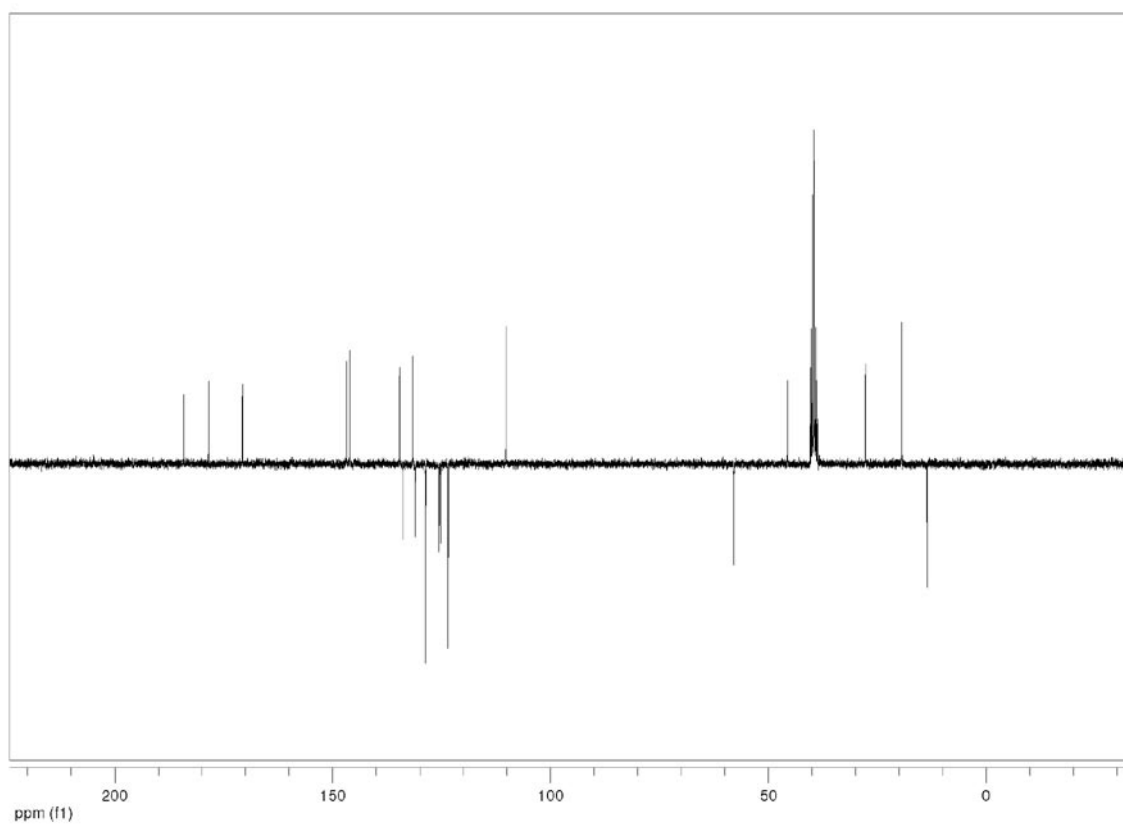


Figure S7. ¹H NMR spectrum of HL7.

**Figure S8.** ¹H NMR spectrum of HL8.**Figure S9.** ¹H NMR spectrum of HL9.

**Figure S10.** ¹H NMR spectrum of HL10.**Figure S11.** ¹H NMR spectrum of HL11.

**Figure S12.** ¹H NMR spectrum of HL12.**Figure S13.** ¹H NMR spectrum of HL13.

**Figure S14.** ¹³C NMR spectrum of HL1.**Figure S15.** ¹³C NMR spectrum of HL2.

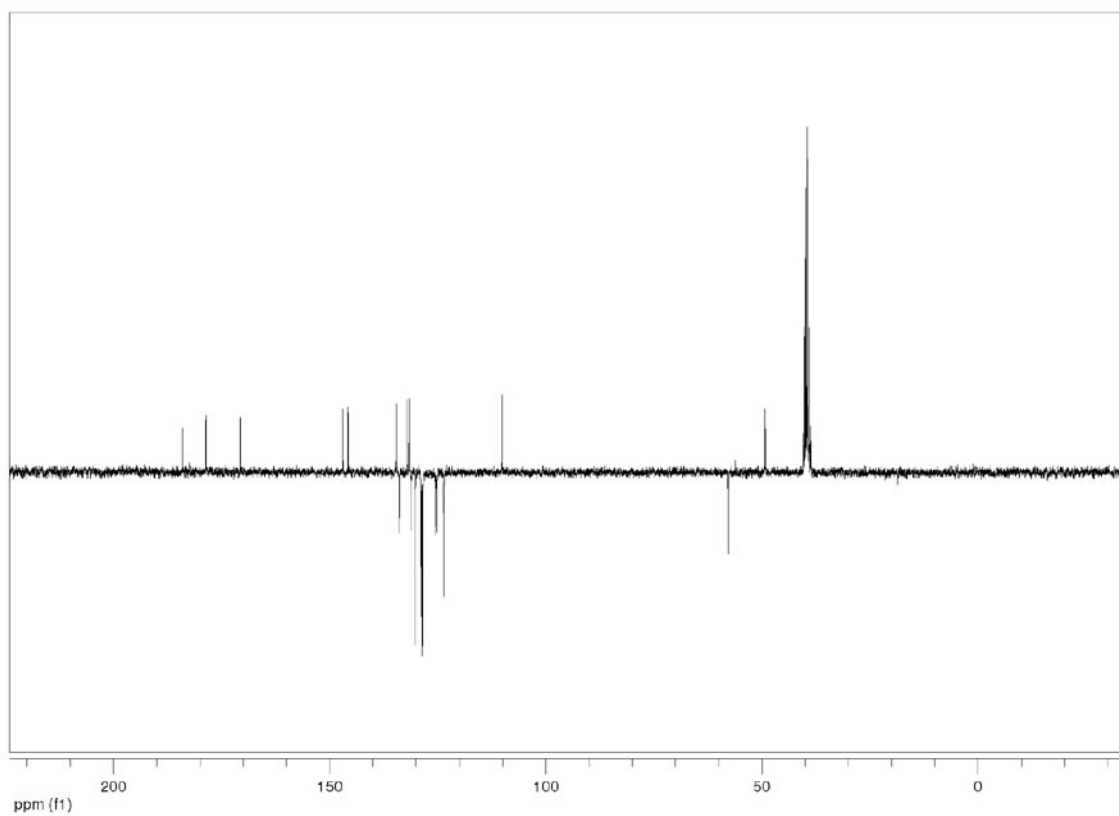


Figure S16. ^{13}C NMR spectrum of HL3.

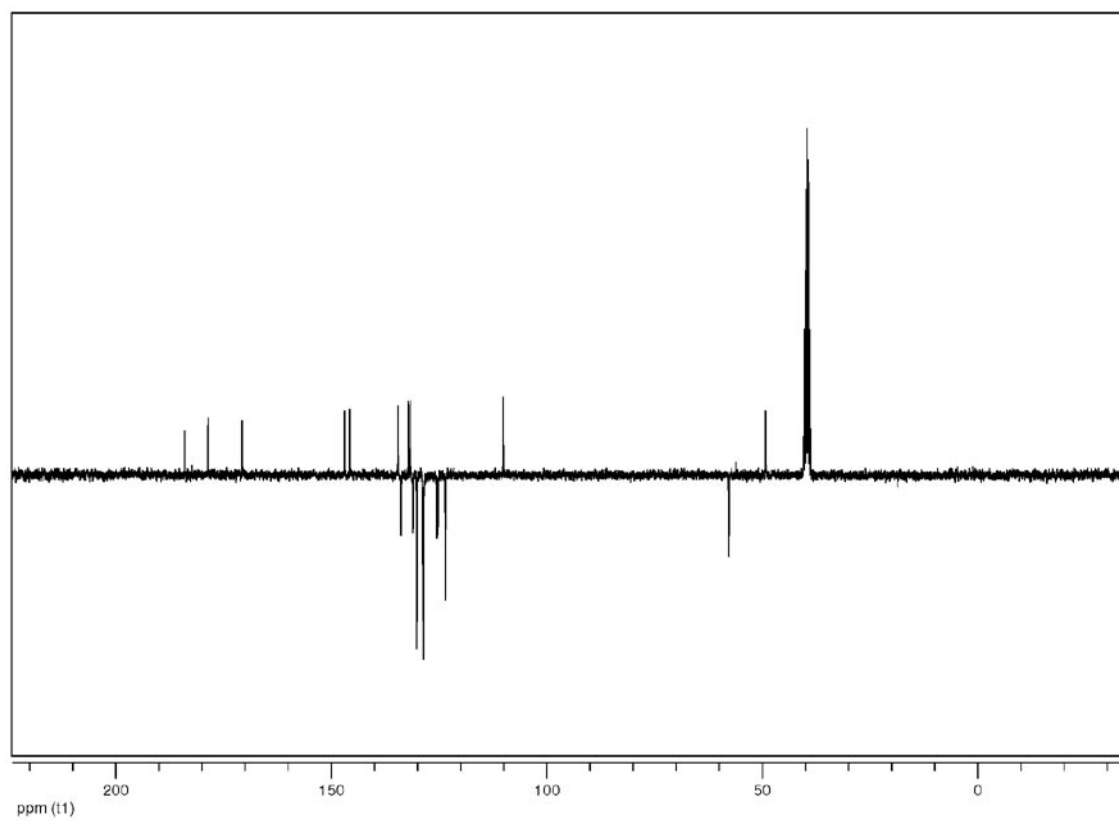
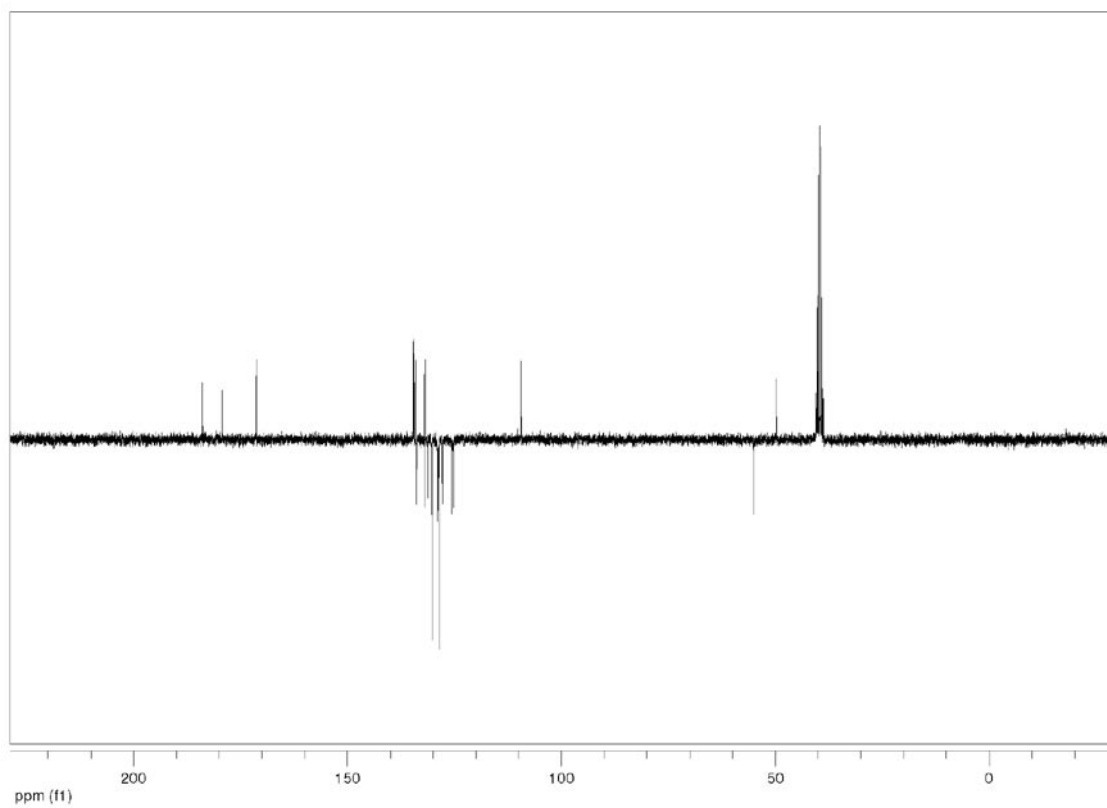
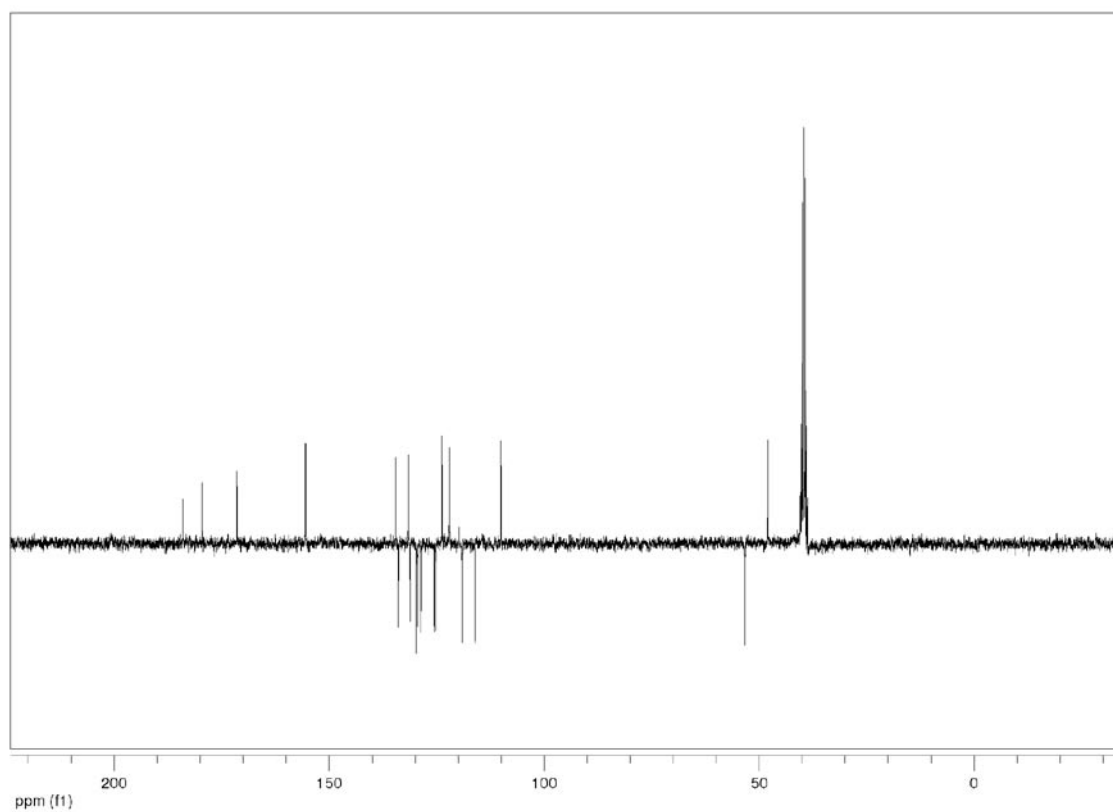


Figure S17. ^{13}C NMR spectrum of HL4.

**Figure S18.** ^{13}C NMR spectrum of HL5.**Figure S19.** ^{13}C NMR spectrum of HL6.

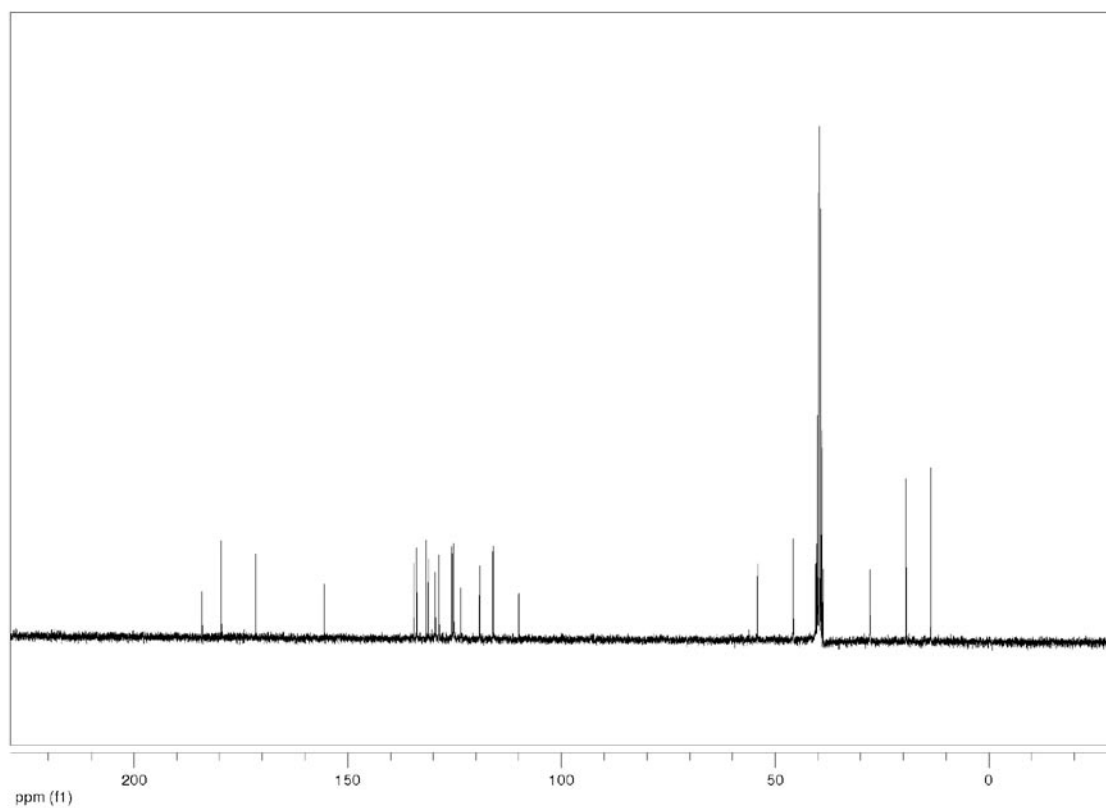


Figure S20. ^{13}C NMR spectrum of HL7.

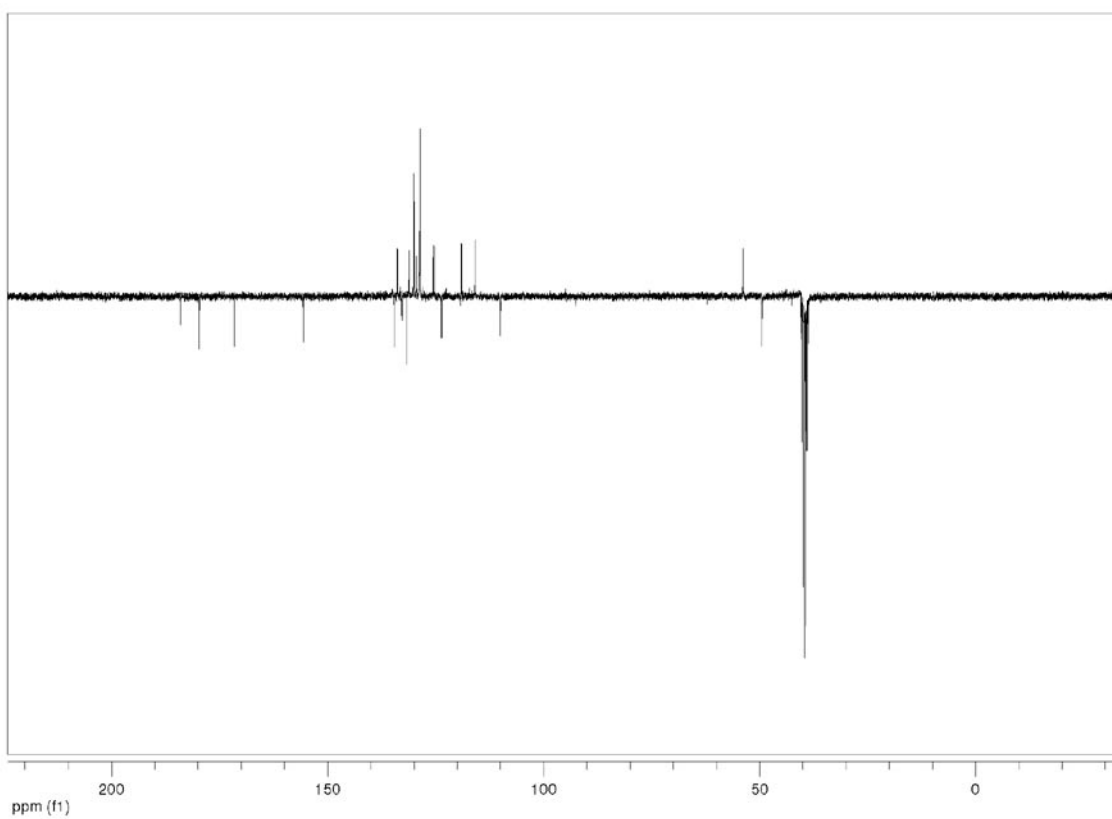
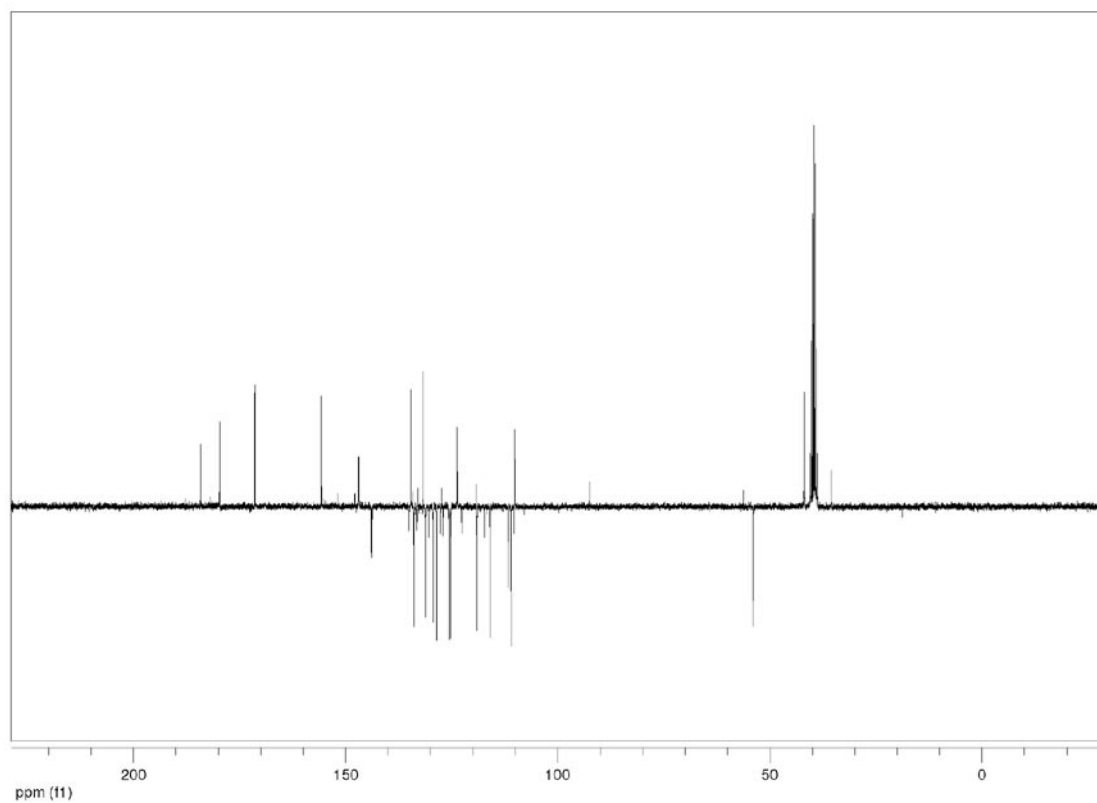
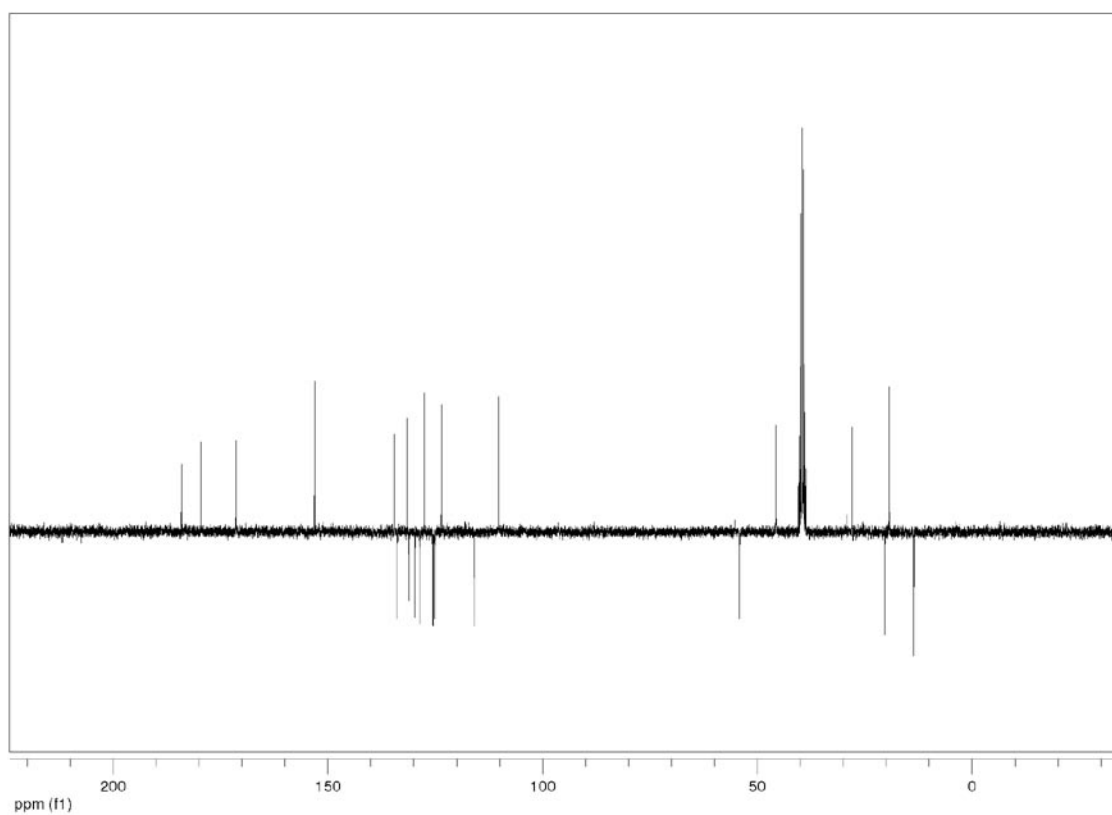


Figure S21. ^{13}C NMR spectrum of HL8.

**Figure S22.** ¹³C NMR spectrum of HL9.**Figure S23.** ¹³C NMR spectrum of HL10.

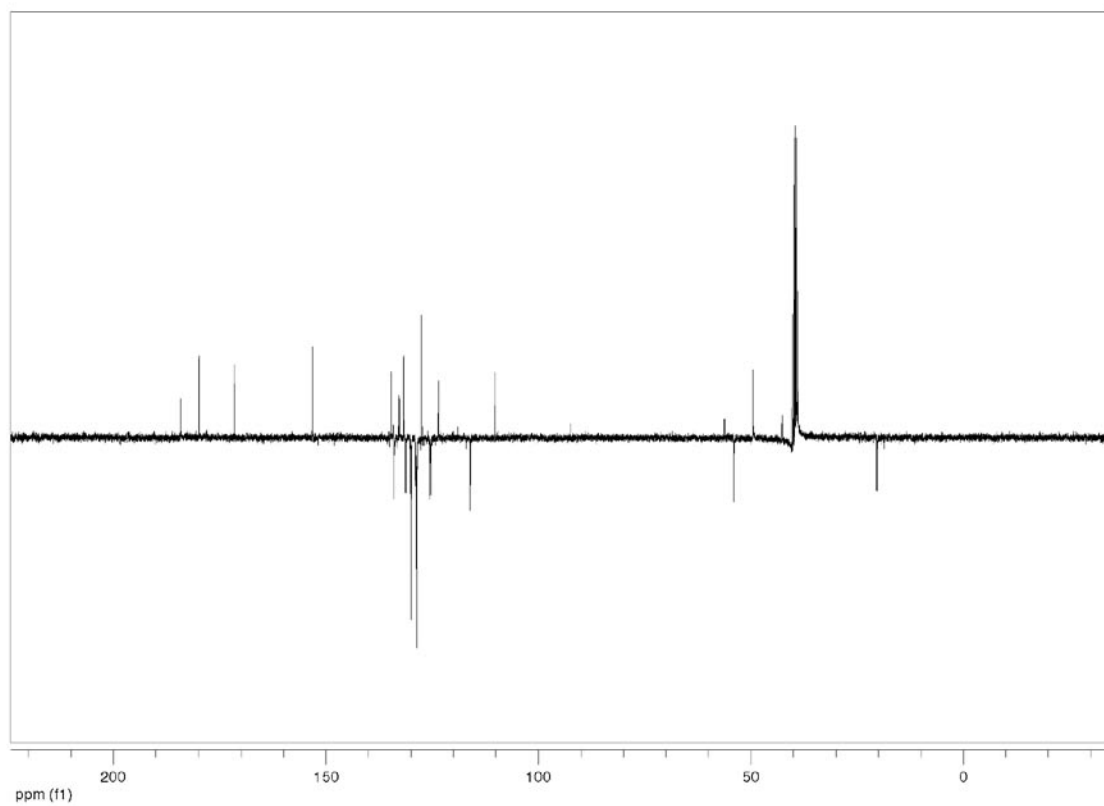


Figure S24. ^{13}C NMR spectrum of HL11.

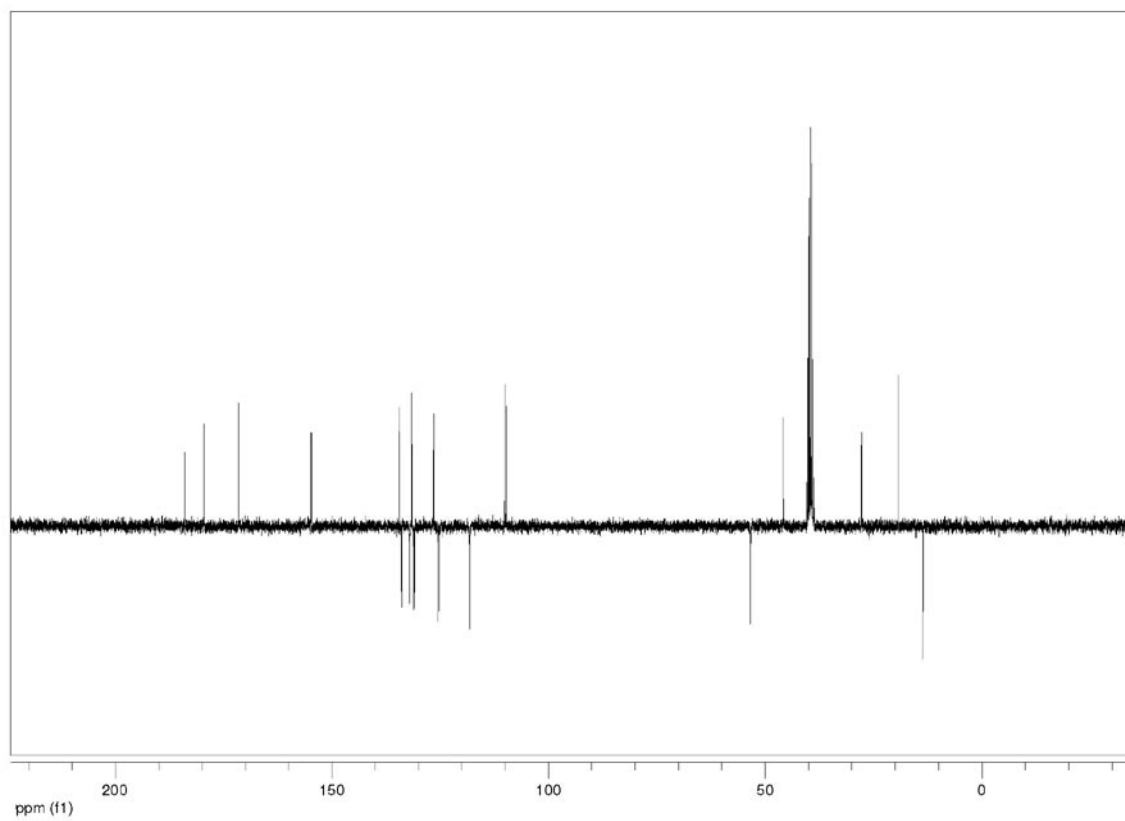


Figure S25. ^{13}C NMR spectrum of HL12.

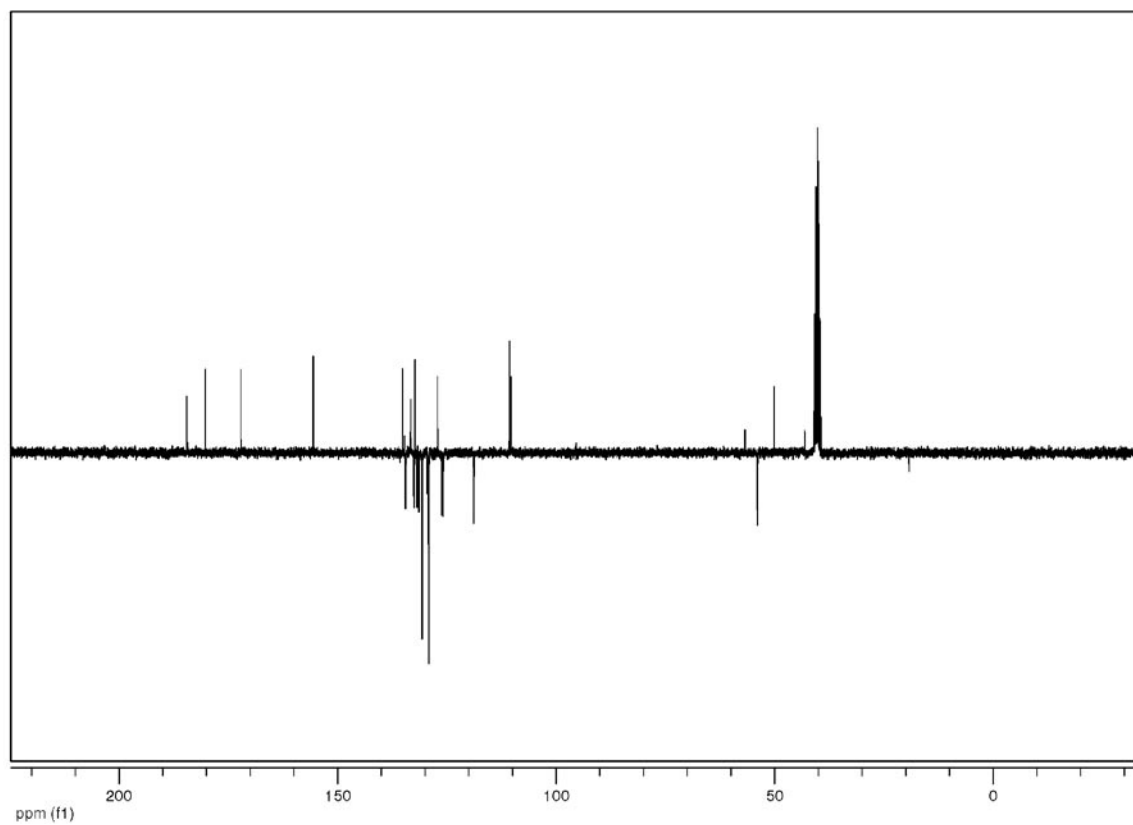


Figure S26. ^{13}C NMR spectrum of HL1.