

## Nutrition, Epigenetics, and Metabolic Syndrome

Junjun Wang,<sup>1</sup> Zhenlong Wu,<sup>1</sup> Defa Li,<sup>1</sup> Ning Li,<sup>2</sup> Scott V. Dindot,<sup>3-5</sup> M. Carey Satterfield,<sup>3,6</sup>  
Fuller W. Bazer,<sup>3,6</sup> and Guoyao Wu<sup>1,3,6</sup>

### Abstract

**Significance:** Epidemiological and animal studies have demonstrated a close link between maternal nutrition and chronic metabolic disease in children and adults. Compelling experimental results also indicate that adverse effects of intrauterine growth restriction on offspring can be carried forward to subsequent generations through covalent modifications of DNA and core histones. **Recent Advances:** DNA methylation is catalyzed by S-adenosylmethionine-dependent DNA methyltransferases. Methylation, demethylation, acetylation, and deacetylation of histone proteins are performed by histone methyltransferase, histone demethylase, histone acetyltransferase, and histone deacetyltransferase, respectively. Histone activities are also influenced by phosphorylation, ubiquitination, ADP-ribosylation, sumoylation, and glycosylation. Metabolism of amino acids (glycine, histidine, methionine, and serine) and vitamins (B6, B12, and folate) plays a key role in provision of methyl donors for DNA and protein methylation. **Critical Issues:** Disruption of epigenetic mechanisms can result in oxidative stress, obesity, insulin resistance, diabetes, and vascular dysfunction in animals and humans. Despite a recognized role for epigenetics in fetal programming of metabolic syndrome, research on therapies is still in its infancy. Possible interventions include: 1) inhibition of DNA methylation, histone deacetylation, and microRNA expression; 2) targeting epigenetically disturbed metabolic pathways; and 3) dietary supplementation with functional amino acids, vitamins, and phytochemicals. **Future Directions:** Much work is needed with animal models to understand the basic mechanisms responsible for the roles of specific nutrients in fetal and neonatal programming. Such new knowledge is crucial to design effective therapeutic strategies for preventing and treating metabolic abnormalities in offspring born to mothers with a previous experience of malnutrition. *Antioxid. Redox Signal.* 17, 282–301.

### Introduction

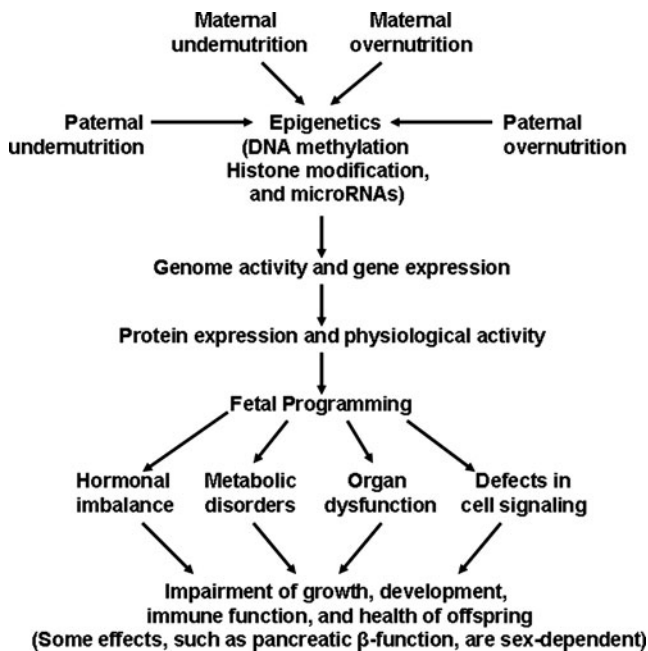
THE UNITED NATIONS Food and Agriculture Organization recently reported that 925 million people worldwide suffer from hunger (190). This population includes not only males and females of reproductive age, but also children who will grow to be adults and produce their own offspring. Currently, deficiencies in protein (amino acids), vitamin A, and iron remain major nutritional problems in developing nations (146), and a deficiency in folic acid can also occur when individuals have a low intake of fresh green vegetables or no consumption of folate-fortified foods (31). In both developed and developing countries, gestating women can experience undernutrition owing to: 1) severe nausea and vomiting known as hyperemesis gravidarum; 2) early or closely-spaced pregnancies; and 3) multi-fetal pregnancies resulting from assisted reproductive technologies (84, 104, 199). These conditions may result in intrauterine growth restriction (IUGR) and impaired health of the offspring (Fig. 1).

The other end of the nutrition spectrum is overnutrition, which can lead to overweight or obesity in human beings (115). One billion adults worldwide are overweight and more than 300 million are obese (126). Many overweight and obese women unknowingly enter pregnancy and continue overeating during gestation (39). These women usually gain more weight during the first pregnancy and accumulate more fat during subsequent pregnancies (128). Maternal obesity before or during gestation may result in adverse pregnancy outcomes, including maternal insulin resistance and hyperglycemia often associated with large-for-gestation age infants, IUGR, and impaired health of the offspring (Fig. 1). Thus, obesity is a global epidemic that negatively affects infants, children, and adults (48).

Extensive epidemiological studies have linked maternal undernutrition and overnutrition with the etiology of many chronic diseases in offspring when they reach adulthood (108). The metabolic syndrome can be defined as a cluster of disorders that include obesity, hyperglycemia (fasting serum

<sup>1</sup>State Key Laboratory of Animal Nutrition and <sup>2</sup>State Key Laboratory of AgroBiotechnology, China Agricultural University, Beijing, China.

<sup>3</sup>Center for Animal Biotechnology and Genomics, Departments of <sup>4</sup>Veterinary Pathobiology, <sup>5</sup>Molecular and Cellular Medicine, and <sup>6</sup>Animal Science, Texas A&M University, College Station, Texas.



**FIG. 1. Impacts of maternal and paternal nutrition on fetal programming.** Either undernutrition or overnutrition of the mother or father affects expression of the fetal genome, which may have lifelong consequences. Thus, alterations in fetal nutrition may result in developmental adaptations that permanently change the structure, physiology, and metabolism of offspring, thereby predisposing individuals to metabolic, endocrine, and cardiovascular diseases in adult life.

glucose ( $>6.1$  mM), hyperinsulinemia, hyperlipidemia, hypertension, and insulin resistance (an impaired response of cells or tissues to physiological concentrations of insulin) (78). These factors, independently or collectively, contribute to a high risk for metabolic disease, a major cause of death in developed nations and developing countries (126). This phenomenon led to the new concept of fetal programming that is defined as an adaptive process whereby nutrition and other environmental factors alter developmental pathways during the critical period of prenatal growth, thereby inducing changes in postnatal metabolism and susceptibility of adults to chronic disease (11). Such findings have prompted animal and human studies to identify the biological mechanisms responsible for the effects of intrauterine nutrition on long-term health consequences of the offspring (156, 200). Results of molecular studies indicate that fetal programming can be explained by epigenetics, which can be defined as stable and heritable alterations of gene expression through covalent modifications of DNA and core histones without changes in the DNA sequence (43). The present article highlights recent advances in this exciting area of biomedical research.

### Impacts of Maternal Nutrition on Fetal Programming

#### *Nutritional requirements for embryonic and fetal development*

The conceptus (embryo/fetus and associated membranes) requires water, amino acids, lipids, carbohydrates, minerals, and vitamins for growth and development (13). In humans, the blastocyst stage of development is reached on about Day 5

after fertilization of the oocyte and then enters the uterus where the embryo differentiates into the inner cell mass (which will develop into the fetus) and the outer layer of cells (trophoblasts which will develop into the placenta and associated membranes). This is followed by implantation (Days 7 to 9 post-fertilization in humans), which is the first stage in sequence of events leading to placentation (the formation and growth of the placenta within the uterus). By Week 4 after fertilization, the basic structure of the mature placenta will have been established. Before placentation, the embryo receives nutrients from uterine secretions and oxygen from its surrounding environment (14). These secretions, including glucose and amino acids (*e.g.*, arginine, leucine, proline, and glutamine), play crucial roles in activating cell signaling and metabolic pathways necessary for protein synthesis and cytoskeletal remodeling in the conceptus (196, 201). A deficiency of nutrients during this period or an inability of the conceptus to respond to the nutrients can result in abnormal development or even death of the conceptus (13).

The placenta transports nutrients, respiratory gases, and the products of their metabolism between the maternal and fetal circulations (186). Rates of uteroplacental blood flows depend on placental vascular growth (a result of angiogenesis) and placental vascularization that are greatly influenced by the availability of nitric oxide and polyamines (143, 198). To support increased uterine and placental blood flows, placental angiogenesis increases markedly from the first to the second third of gestation and continues to increase during late gestation. Uptake of nutrients by the uterus or the fetus is determined by both rate of blood flow and concentrations of nutrients in the arterial and venous blood. To support the rapid rate of placental and fetal growth, uptake of both macro- and micro-nutrients by the uterus is greater in pregnant women relative to that for nonpregnant women (143). Thus, impaired placental blood flow contributes to IUGR in mammalian pregnancies.

Energy is required for a variety of physiological processes in the fetus, including nutrient transport, cell motility, and biosynthetic pathways (64). Dietary macronutrients (glucose, amino acids, and fatty acids) are the ultimate sources of energy substrates during fetal growth (201). In the pregnant mother and her growing fetus, glucose is the major fuel for red blood cells, brain, retinal cells, and medullary cells of the kidney, while the maternal and fetal hearts utilize both glucose and lactate. Additionally, the small intestine of both the mother and the fetus oxidizes glutamate, aspartate, and glutamine to meet most of their energy requirements (78). Through  $\beta$ -oxidation, fatty acids are the major energy substrates for the maternal liver, skeletal muscle, heart, and kidneys (21). The fetal liver also oxidizes long-chain fatty acids (LCFA) to  $\text{CO}_2$  at relatively high rates. However, fetal muscle has a low capacity for oxidation of LCFA due to relatively low concentrations of carnitine and low carnitine palmitoyl-transferase-I activity (78).

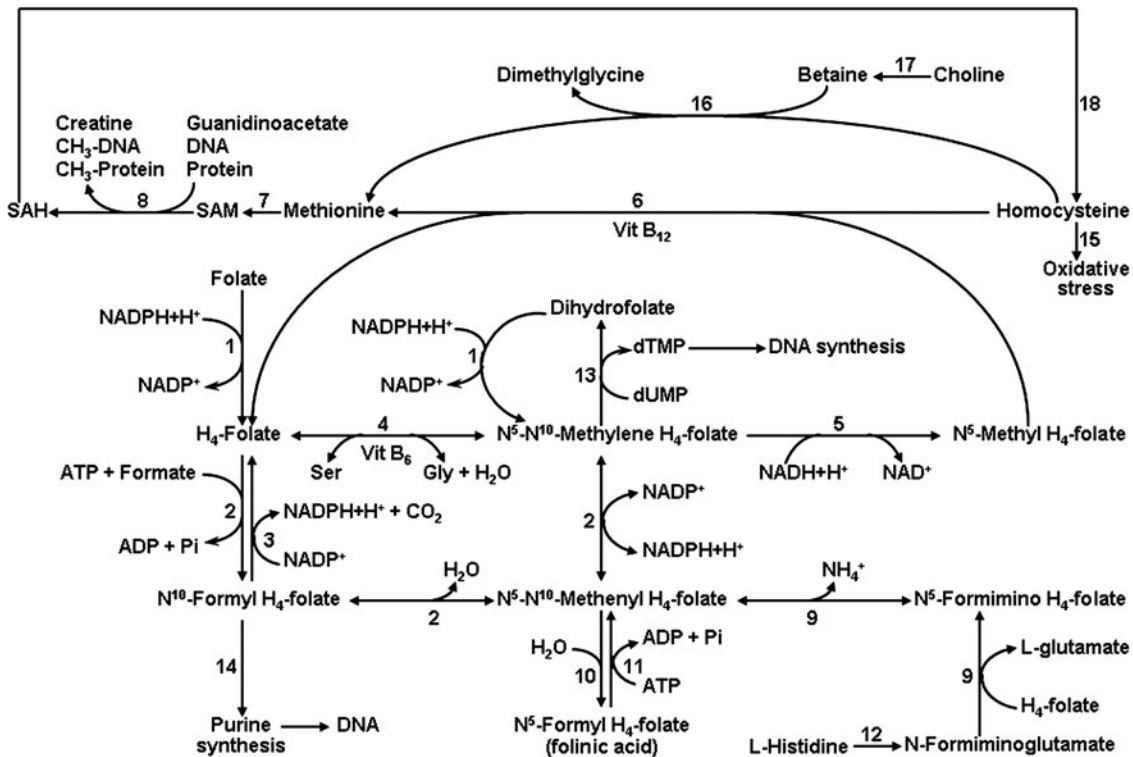
The human placenta is permeable to LCFA and transfers them to the fetus for metabolic utilization. In both maternal and fetal tissues (*e.g.*, the liver), fatty acids are synthesized from acetyl-CoA which is a product of oxidation of glucose and amino acids (200). In most mammals (including humans), the fetus accumulates only a small amount of lipids prior to mid-gestation, but a relatively large amount of lipids accumulate exponentially thereafter (21). Because placental and

fetal tissues have  $\Delta^9$  desaturase, they are able to synthesize the  $\omega 9$  (oleic acid) family of unsaturated fatty acids from palmitate (C16:0; a saturated fatty acid). Humans cannot synthesize linoleic acid or  $\alpha$ -linolenic acid, which are  $\omega 6$  and  $\omega 3$  fatty acids, respectively. Therefore, these two nutritionally essential unsaturated fatty acids must be provided in the diet and used to synthesize the long-chain  $\omega 6$  and  $\omega 3$  polyunsaturated fatty acids. Physiologically and nutritionally important long chain  $\omega 6$  and  $\omega 3$  polyunsaturated fatty acids include arachidonic acid (20:4 $\omega 6$ ), eicosapentaenoic acid (20:5 $\omega 3$ ), and docosahexaenoic acid (22:6 $\omega 3$ ).

Amino acids serve not only as building blocks for proteins, but also as essential precursors for synthesis of a variety of physiologically important molecules, including hormones, small peptides (*e.g.*, glutathione), neurotransmitters, nitric oxide, creatine, carnitine, and polyamines (196). Additionally, through multiple cell signaling pathways, amino acids regulate key metabolic pathways that are vital to human health, growth, development, and reproduction (197). Notably, some amino acids (histidine, methionine, glycine, serine) partici-

pate in one-carbon unit metabolism, which is essential for DNA synthesis, as well as cell growth and development (Fig. 2). Based on nitrogen balance and growth, amino acids have been traditionally classified as either nutritionally essential (EAA) or nonessential (NEAA). The EAA are histidine, isoleucine, leucine, lysine, methionine, phenylalanine, threonine, tryptophan, and valine. The NEAA are alanine, arginine, asparagine, aspartate, cysteine, glutamate, glutamine, glycine, proline, serine, taurine, and tyrosine. However, the basis for classification of an amino acid as a NEAA has conceptual limitations because there is no compelling evidence that certain NEAA (*e.g.*, arginine and glutamine) can be adequately synthesized by the mother or fetus to support optimal survival, growth, and development of the conceptus (203, 204). The synthesis of proteins in the fetus depends on the balanced provision of all of the constituent amino acids, namely both EAA and NEAA.

Nutritionally significant macro- and micro-minerals cannot be synthesized by the body and must, therefore, be provided in the diet (195, 213). The macro-minerals are sodium,



**FIG. 2. One-carbon unit metabolism for provision of methyl donors in cells.** Folate, histidine, methionine, glycine and serine participate in the transfer of one-carbon units for the synthesis of purines and DNA as well as methylation reactions in animals. The enzymes that catalyzed the indicated reactions are: 1) folate reductase; 2)  $N^5$ - $N^{10}$ -methylene  $H_4$ -folate dehydrogenase (a trifunctional enzyme possessing  $N^{10}$ -formyl  $H_4$ -folate synthetase,  $N^5$ - $N^{10}$ -methylene  $H_4$ -folate dehydrogenase, and  $N^5$ - $N^{10}$ -methenyl  $H_4$ -folate cyclohydrolase activities); 3)  $N^{10}$ -formyl  $H_4$ -folate dehydrogenase; 4) serine hydroxymethyl transferase; 5)  $N^5$ - $N^{10}$ -methylene  $H_4$ -folate reductase; 6) methionine synthase; 7) S-adenosylmethionine synthase; 8) S-adenosylmethionine as a major methyl group donor in methyltransferase reactions; 9) formiminotransferase-cyclodeaminase (a bifunctional enzyme possessing glutamate formiminotransferase and formimidoyl  $H_4$ -folate cyclodeaminase activities); 10) serine hydroxymethyl transferase,  $N^5$ - $N^{10}$ -methenyl  $H_4$ -folate cyclohydrolase, and spontaneous reaction at pH 4 to 7.0; 11)  $N^5$ - $N^{10}$ -methenyl  $H_4$ -folate synthetase; 12) a sequential series of enzymes: histidase, urocanase, and imidazolone propionate hydrolase; 13) thymidylate synthase; 14) formyltransferase; 15) an oxidant for cells; 16) betaine:homocysteine methyltransferase; 17) choline dehydrogenase; and 18) S-adenosylhomocysteine hydrolase. dTMP, deoxythymidine 5'-monophosphate; dUMP, deoxyuridine 5'-monophosphate; Gly, glycine;  $H_4$ -folate, tetrahydrofolate; SAH, S-adenosylhomocysteine; SAM, S-adenosylmethionine; Ser, serine; Vit, vitamin.

potassium, chloride, calcium, phosphorus, magnesium, and sulfur (177). The micro-minerals are iodine, iron, zinc, selenium, manganese, copper, cobalt, chromium, molybdenum, silicon, fluoride, vanadium, and nickel (177). The inorganic nutrients are the backbone for development, maintenance, and function of the skeleton system, and they serve as second messengers in cell signaling, and maintain polarity of the plasma membrane. Minerals also regulate extracellular and intracellular osmolality, which is important for maintaining cell and blood volume as well as cell viability and shape (213). Most minerals are either cofactors for enzymes or components of metalloproteins, and also participate in gene expression, electron transport, and redox reactions (21).

Vitamins are organic nutrients that are required in small amounts for biochemical reactions. Water-soluble vitamins include vitamin C and members of the B complex that are absorbed by the small intestine into the portal circulation (127). Members of the B complex vitamins are vitamin B<sub>1</sub> (thiamin), vitamin B<sub>2</sub> (riboflavin), vitamin B<sub>3</sub> (niacin), vitamin B<sub>5</sub> (pantothenic acid), vitamin B<sub>6</sub> (pyridoxal, pyridoxine, and pyridoxamine), vitamin B<sub>12</sub> (cobalamin), biotin, and folate (21). Because of their water solubility, there is limited storage of these vitamins in the body (127). As a result, these micro-nutrients must be provided regularly in the diet. Lipid-soluble vitamins include vitamin A, vitamin D, vitamin E, and vitamin K. Most vitamins, except niacin and vitamin D, cannot be synthesized by humans and must be supplied in the diet (177). Vitamins serve as cofactors for enzymes involved in gene expression, metabolism of nutrients (e.g., fatty acids, glucose, and amino acids), one-carbon-unit transfer (Fig. 2), and ATP production.

#### *Impacts of maternal undernutrition on offspring metabolism and health*

In both humans and animals, birth weight is the predominant indicator of prior nutrient availability *in utero*. Epidemiological studies of the Dutch famine (November 1944 and May 1945) have shown that undernutrition of women during the second or third trimester, but not the first trimester, reduced weight, length, and head circumference of infants at birth (102). The growth-stunting effects were more pronounced for third trimester exposure than second trimester

exposure (132) because most growth of the fetus occurs during late gestation (201). However, this does not necessarily mean that a severe deficiency in specific nutrients (e.g., micronutrients and amino acids) during the first trimester of pregnancy had little impact on maternal health or overall fetal growth and development. Indeed, maternal malnutrition during early gestation has a great effect on adversely influencing metabolism and health of offspring (200). Of note, in response to maternal undernutrition, the fetus develops mechanisms for adapting to the extrauterine environment that may confer an evolutionary advantage for survival of the species (152, 162). However, when environmental cues during prenatal life inappropriately program offspring, there are adverse consequences, including the increased prevalence of adult onset of disease (21, 156).

Maternal undernutrition increases risk for adult-onset of metabolic syndrome (11, 32). This notion is supported by four lines of evidence. First, at 4 years of age, children with low birth weight have elevated levels of glucose and insulin in response to a glucose challenge (206). Second, individuals exposed to the Dutch winter famine of 1944–1945 *in utero* had higher rates of insulin resistance, vascular disease, morbidity, and mortality in adulthood (102). Indeed, exposure of pregnant women to famine during any stage of gestation was associated with insulin resistance in adult offspring (102, 206). Of particular interest, maternal undernutrition during early gestation had greater effects in increasing the incidence of cardiovascular and metabolic disorders in the offspring in later life, compared with maternal undernutrition during mid- or late-gestation (102). Third, maternal undernutrition was correlated with the incidences of coronary heart disease and hypertension in a cohort of 4630 adult men (10, 42) and 3447–4130 women (10, 51) born in Helsinki, Finland between 1924 and 1933 (Table 1). Moreover, among adult men and women, small size at birth (an indicator of IUGR) was associated with increased mortality rates from cardiovascular causes (80). Fourth, a cohort study of 15,000 Swedish men and women born between 1915 and 1929 perhaps provides by far the most convincing evidence for the close association between reduced fetal growth rate and increased risk of death from ischemic heart disease (94). These results highlight the importance of birth weight as a simple measure on predicting and potentially managing potential health risks in adults.

TABLE 1. EFFECTS OF BIRTH WEIGHT ON HAZARD RATIOS FOR CORONARY HEART DISEASE AND THE CUMULATIVE INCIDENCE OF HYPERTENSION IN ADULT MEN AND WOMEN

Birth weight (g)	Adult men		Adult women	
	Hazard ratios for coronary heart disease <sup>a</sup>	Cumulative incidence of hypertension (%) <sup>b</sup>	Hazard ratios for coronary heart disease <sup>c</sup>	Cumulative incidence of hypertension (%) <sup>d</sup>
≤ 2500	3.63	—	1.34	—
2501–3000	1.86	19.0	1.38	21.1
3001–3500	1.99	17.0	1.24	16.3
3501–4000	2.08	14.1	1.17	13.0
> 4000	1.00	12.5	1.00	12.1
<i>P</i> value for trend	0.006	< 0.001	0.007 <sup>e</sup>	< 0.001

<sup>a</sup>Eriksson et al. (42). A cohort of 4630 adult men was studied.

<sup>b</sup>Barker (10). A cohort of 4630 adult men was studied.

<sup>c</sup>Forsen et al. (51). A cohort of 3447 adult women was studied.

<sup>d</sup>Barker (10). A cohort of 4130 adult women was studied.

<sup>e</sup>Adjusted for gestation and placental weight.



Thus, in terms of long-term health, the embryo/fetus is most vulnerable to nutritional insults during early gestation.

Experimental evidence from a variety of species and animal models has provided a wealth of knowledge regarding effects of reduced nutrient availability *in utero* on the development of adult disease (3). For example, global undernutrition increases the risk for dyslipidemia, obesity, and type-2 diabetes mellitus in rats (156). Also, maternal protein restriction induces hypertension and vascular dysfunction in adult female offspring of rats (65, 149). Extensive research with large animals (e.g., sheep and nonhuman primates) provides further evidence for fetal programming of vascular dysfunction and metabolic abnormalities. For example, in sheep, maternal undernutrition induces left ventricular hypertrophy and alters gene expression in the left ventricle of the fetal heart (62, 180). Additionally, maternal nutrient restriction results in increased myocardial lipid and altered gene expression in offspring at 1 year of age (28). Moreover, in sheep, maternal nutrient restriction impairs renal function, increases the development of glomerulosclerosis, and enhances apoptosis in kidneys, while altering the expression of proteins involved in regulating inflammatory processes (155, 192). In both pigs and sheep, maternal undernutrition also results in decreased skeletal muscle fiber number, increased deposition of adipose tissue, and increased connective tissue content (15, 58). Collectively, these alterations in normal development result in: a) reduced postnatal growth, including both whole-body and skeletal muscle growth; b) reduced efficiency of nutrient utilization; and c) suboptimal function of multiple organs.

#### *Impacts of maternal overnutrition on metabolism and health of offspring*

Maternal obesity has been linked to metabolic perturbations, obesity, and cardiovascular disorders in the offspring of a number of mammalian species, including humans, mice, rats, and sheep (6, 83, 85, 156). In nonhuman primates, a maternal high fat diet promotes the development of non-alcoholic fatty liver and atherosclerosis (110) and alteration of seven metabolites in the fetus related to one-carbon unit metabolism (33). Notably, changes in the fetal metabolome result, at least in part, from an altered fetal hepatic chromatin structure that leads to aberrant gene expression in the liver (1) and other insulin-sensitive tissues (162).

Feeding a high-fat diet during pregnancy or the neonatal period stimulates adipogenesis, induces cardiovascular dysfunction characterized by elevated systolic blood pressure, and impairs endothelium-dependent relaxation in rats (3, 61, 83). Importantly, the response to maternal dietary treatment differed between male and female offspring in that female offspring were more susceptible to elevated systolic and diastolic blood pressure when their mothers were fed a high-fat diet during gestation and/or lactation (145). This interesting phenomenon may be explained by imprinting of genes and sex-related differences in hormone secretion. In contrast, endothelium-dependent relaxation of blood vessels in response to acetylcholine was consistently blunted in both males and females exposed to a high-fat diet either *in utero* or during lactation (137). A major underlying mechanism for the high fat-induced vascular dysfunction is likely the reduced release of nitric oxide (a major vasodilator) from endothelial cells (198).

Maternal obesity due to a high-fat diet reduced mitochondrial copy number in the kidney in 1-year-old rats, while altering expression of a number of mitochondrial genes in the rat aorta (166). These findings suggest that mitochondria play a central role in developmental programming of vascular function. Similar results were observed in the offspring of pregnant mice fed a high-fat diet, including increased adiposity, dyslipidemia, hypertension, and insulin resistance (38, 148). In sheep, maternal obesity downregulates expression of genes involved in myogenesis and placental angiogenesis (212) and fetal skeletal muscle (173), while reducing the phosphorylation of AMP-activated kinase in the fetal and neonatal liver (134) as well as energy metabolism (185). Importantly, these changes in gene and protein expression in response to maternal overnutrition result in increased fetal and/or neonatal adiposity (50, 121) and increased expression of the leptin gene in peri-renal and subcutaneous adipose tissue depots (121). The consequences of maternal overnutrition continue to manifest in offspring as evidenced by impaired insulin sensitivity and reduced glucose utilization on postnatal Day 210 despite growing in an identical nutritional environment from birth (150). Similarly, a maternal high-fat diet during pregnancy results in impaired glucose homeostasis in rat offspring characterized by elevated concentrations of insulin in plasma at 1 year of age (166).

#### **Impact of Maternal Nutrition on Gene Expression in Offspring**

##### *Maternal nutrition and gene expression in offspring*

As noted previously, studies of offspring born to women during the Dutch famine in the 1940s suggested a link between the fetal environment (including nutrition) and postnatal health, particularly cardiovascular function or dysfunction, in humans (9, 30, 69, 133). Interestingly, 60 years after birth, offspring with early prenatal experience of the famine exhibited less DNA methylation of the imprinted *IGF2* gene, as compared to the same-sex siblings without exposure to prenatal malnutrition (67). Furthermore, individuals with periconceptual exposure to the famine had lower methylation of the *INSIGF* gene, but higher methylation levels for several other genes (*IL10*, *LEP*, *ABCA1*, *GNASAS*, and *MEG3*) (171). These genes are closely linked with nutrient metabolism, cardiovascular function, and inflammation. Interactions between undernutrition and sex also affect methylation of the *INSIGF*, *LEP*, and *GNASAS* genes (171).

In monkeys, a 30% reduction in maternal nutrient intake did not alter fetal weight at stages 0.5 and 0.9 of gestation (full term being 1.0) (176). However, tissue-specific global methylation status (176) and the availability of methyl group donors (153) were altered in the kidneys of nonhuman primates at both 0.5 and 0.9 of gestation. Furthermore, emerging evidence indicates modifications of hepatic phosphoenolpyruvate carboxykinase (a key enzyme in gluconeogenesis) after exposure of fetal baboons to moderately reduced nutrient availability (124). These results indicate that even a relatively mild nutrient restriction can induce alterations in gene expression and provide a potential mechanism responsible for the increased incidence of endothelial dysfunction, renal dysfunction, and hypertension in offspring with previous experience of malnutrition during the period of fetal growth. Such an effect of fetal programming on gene expression in

offspring has been experimentally tested in a number of species, including nonhuman primates (124), rats (5), mice (38), cattle (23), and sheep (162). Thus, effects of changes in nutrition or endocrine status during the fetal can negatively impact subsequent development of the offspring.

#### *Transgenerational effects of maternal nutrition*

Epidemiological studies have indicated the presence of transgenerational effects of maternal malnutrition in humans (209). For example, starvation in young boys (7–11 years of age) resulted in increased incidence of metabolic syndrome in them as adult men, and in their children and grandchildren (79). Likewise, food deprivation in female fetuses for 7–12 weeks was associated with increased prevalence of metabolic syndrome in them as adult women, and in their children and grandchildren (22). Results of animal studies also indicated that the effect of maternal undernutrition on hormone secretion and metabolism can be passed transgenerationally from mother (F<sub>0</sub>) to daughter (F<sub>1</sub>) and to the F<sub>2</sub> progeny (16). For example, maternal calorie restriction in rats caused: a) decreases in the mass and number of pancreatic  $\beta$ -cells in the first generation of female offspring; and b) impaired responses of F<sub>2</sub> pancreatic  $\beta$ -cells to pregnancy, leading to insulin resistance and gestational hyperglycemia (107, 168). Similar results were obtained for female rats fed a low-protein diet (210) or female mice subjected to caloric undernutrition (76) during gestation. In guinea pigs, even modest maternal undernutrition (70% of normal food intake) resulted in alterations in the cardiac structure (*e.g.*, increases in wall thickness and mass of the left ventricle), hypothalamo-pituitary-adrenal function (*e.g.*, increased basal levels of cortisol), and elevated blood pressure in adult male F<sub>1</sub> and F<sub>2</sub> offspring through the maternal line (4, 16). Interestingly, some of the transgenerational effects of maternal undernutrition-induced IUGR and developmental defects on diabetes and cardiovascular function are sex-dependent (168, 187), which can be explained well by differential genomic imprinting (35). Likewise, chronic high-fat feeding in fathers resulted in hypomethylation of the *II13ra2* gene and  $\beta$ -cell dysfunction in female, but not male, offspring (123).

#### **Fetal Programming**

##### *The mechanism of fetal programming: Epigenetics*

The genetic code established by the DNA sequence exhibits minimal rates of change after the formation of the diploid chromatin state at fertilization (43). Therefore, an extensive search for alternative mechanisms of gene regulation resulted in the now widely accepted notion that changes in gene expression can be manifest by mitotically and/or meiotically heritable alterations in the DNA-protein complex without any change in the DNA sequence (156). This phenomenon was originally termed “epigenetics” by Conrad Waddington in 1940 (182) based on the Greek prefix “epi” meaning over or above. Molecular mechanisms about epigenetics and genomic imprinting have evolved over the past 40 years since the first identification in the 1970s of DNA methylation as a regulatory factor (70). The 1980s (109, 183, 193) and 1990s (12, 18, 63) witnessed pioneering research on the maintenance and regulation of DNA methylation patterns in mammals. In the mid 1990s, histone modifications were discovered as an epigenetic

determinant of chromatin structure and, therefore, genome activity (29, 174). The identification of noncoding microRNAs and their functions early in the 2000s further expanded the field of epigenetic research (4, 95). Excitingly, the first whole epigenome analysis was completed for yeast in 2005 (136). It is now known that epigenetic modifications can be carried forward from one generation of cells to the next (mitotic inheritance) and between generations in a species (meiotic inheritance) (156).

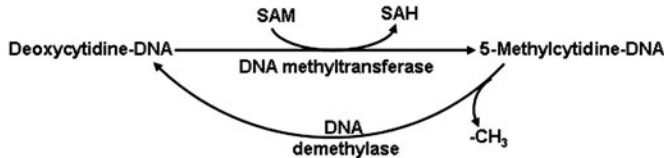
At present, the four mechanisms responsible for mediating epigenetic effects are: a) chromatin modifications, b) DNA methylation (occurring at the 5'-position of cytosine residues within CpG dinucleotides throughout the mammalian genome), c) histone modifications (acetylation, methylation, phosphorylation, ubiquitination, and sumoylation), and d) RNA-based mechanisms such as small noncoding RNAs or inhibitory RNAs (Fig. 3). The enzymes involved in these reactions include specific DNA and protein methyltransferases, DNA demethylases, histone acetylase (lysine acetyltransferase), GCN5-related N-acetyltransferase (a super family of acetyltransferase), and histone deacetyltransferase (18, 26, 30).

##### *DNA methylation*

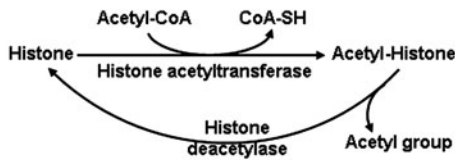
The methylation of DNA is catalyzed by DNA methyltransferases (*DNMT1*, *DNMT3a*, and *DNMT3b*) and is a reversible physiological process in the eukaryotic genome (45). This biochemical event involves the donation of a methyl group from S-adenosylmethionine to the 5'-position of a cytosine nucleotide linked to a guanine nucleotide (CpG dinucleotide) by a phosphodiester bond. Regions with a high CpG dinucleotide content form CpG islands, which are located in the regulatory regions of many genes, including promoters and enhancers (37). Due to their presence in these regulatory regions, changes in methylation status can either facilitate (hypomethylation) or inhibit (hypermethylation) the expression of a gene (106). In normal mammalian cells, over 85% of CpG dinucleotides are methylated, which plays a role in maintaining the integrity of chromatin structure and transcriptional regulation (56). Except for imprinted genes, CpG islands present in the promoter regions of genes are usually unmethylated. Approximately 1% to 2% of the human genome consists of CpG dinucleotide whose methylation is inversely related to transcriptional activity in cells (187).

Epigenetic modifications are erased and re-established in a tissue-specific manner during embryonic development (184). The haploid genomes of the sperm and oocyte possess different patterns of DNA methylation that arise from sex-specific patterns of *de novo* methylation. After fertilization, the paternal genome is rapidly demethylated prior to the first cell division of the zygote. In contrast, the maternal genome is protected from this demethylation event, and, instead, is gradually demethylated during the development of the blastocyst. At the blastocyst stage, most methyl marks are removed, remaining present at elements regulating genomic imprinting and retroviral elements (43, 56). After implantation, *de novo* DNA methylation occurs, giving rise to tissue-specific patterns of DNA methylation (36). The process of demethylation and remethylation during the development of germ cells is referred to as “reprogramming” and this physiological event is regulated, in part, by the phosphatidylinositol 3-kinase pathway (161).

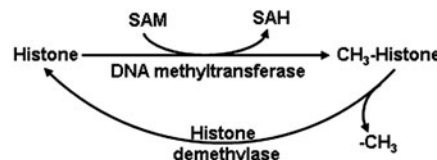
## 1. DNA methylation



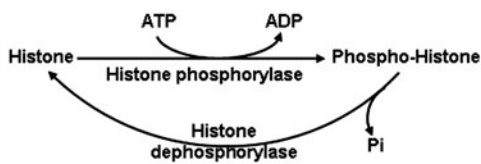
## 2. Histone acetylation



## 3. Histone methylation



## 4. Histone phosphorylation



## 5. Histone ubiquitination

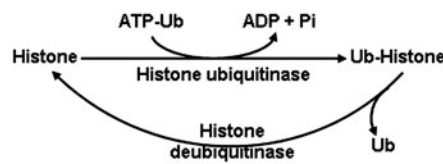


FIG. 3. Biochemical reactions involving DNA methylation and histone modifications. These reactions are localized in specific compartments of the cell. SAH, S-adenosylhomocysteine; SAM, S-adenosylmethionine; Ub, ubiquitin.

Epigenetic information is heritable between cell generations through the action of the methyltransferase, *DNMT1* (184). In somatic cells, this enzyme recognizes and methylates CpG dinucleotides present on the newly synthesized strand. As DNA synthesis occurs, the parental strand remains methylated, whereas the newly synthesized daughter strand is not. *DNMT1* “reads” the parental strand and methylates CpG dinucleotides on the daughter strand that are complementary to methylated CpG dinucleotides on the parental DNA strand (Fig. 4). The importance of DNA methylation for successful pregnancy outcome is evident by the fact that all homozygous *DNMT1* knockout mice die at the morula stage of embryonic development (20).

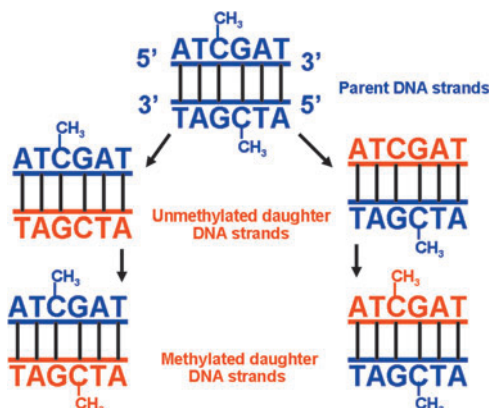


FIG. 4. Passage of epigenetic information during DNA replication in germlines. When double-stranded DNA is divided into daughter DNA, a specific DNA methyltransferase can methylate the cytosine in CpG islands of the daughter DNA at the blastocyst stage of conceptus development. Tissue-specific gene expression results from differences in DNA methylation and promoter activity, leading to differentiated somatic cells.

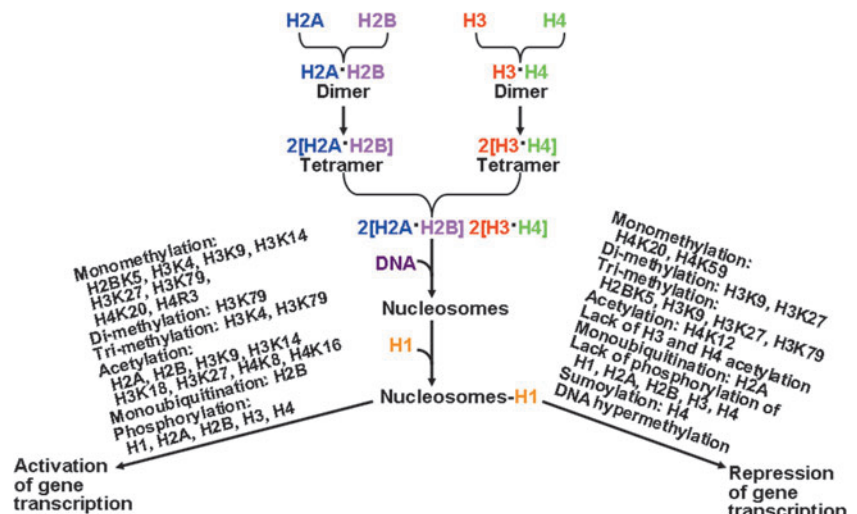
## Histone modifications

Histones are basic proteins that facilitate the packaging of DNA in the nucleus and the regulation of gene expression in cells. The histone proteins are H1, H5, H2A, H2B, H3, and H4. Two copies of the H2A, H2B, H3, and H4 histones assemble to form one octameric histone protein core (Fig. 5). DNA is wrapped around the core twice in a left-handed super-helical turn to make a structural unit called the nucleosome. In the histone protein core, a heterotetramer of histones H3 and H4 is flanked by two heterodimers of histones H2A and H2B (59). Histones H1 and H5 are linker histones (131). Histone H1 binds the nucleosome as well as the entry and exit sites of the DNA. Through changes in phosphorylation and O-glycosylation, linker histones modify chromatin structure (2). Post-translational modifications imparted onto the N-terminal tails of each histone protein alter the properties and structures of chromatin (131). These post-translational modifications include methylation of arginine or lysine, acetylation of lysine, phosphorylation of serine and threonine, ubiquitination of lysine, ADP-ribosylation, sumoylation of lysine, and glycosylation (147). The biochemical changes regulate gene expression by modulating: a) the accessibility of the transcriptional machinery to the DNA binding elements; and b) DNA replication, recombination, and chromosomal organization. At present, more than 100 specific modifications of chromatin are associated with either actively transcribed or silenced genes (45). Therefore, histone proteins play an important role in gene expression (Fig. 5).

Histone methylation is catalyzed by histone methyltransferases (HMT), which transfer one to three methyl groups from S-adenosylmethionine to lysine or arginine residues of histone proteins (7). There are four known HMTs, including SUV1, SUV2, SUV3, and RIZ. Examples of the enzymes are H3K4-, H3K9-, and H3K27-specific methyltransferases, which target lysine residues 4, 9, and 27 of histone H3, respectively. The H3K27-specific methyltransferase



**FIG. 5. Roles of histone modifications in the regulation of gene transcription.** Methylation of histones, acetylation, or ubiquitination can either activate or repress gene expression, depending on specific histone proteins and the sites of modifications. In general, phosphorylation of histones promotes transcription, DNA repair, and apoptosis. H, histone; K, lysine residue; R, arginine residue.



is also known as the enhancer of zeste homologue 2 (EZH2). Other significant histone methylases are H3K79- and H4H20-specific (131). Silencing of differentiation genes is associated with the methylated H3K27 chromatin mark. In a variety of cell types, histone methylation is generally associated with transcriptional repression, but methylation of some lysine (*e.g.*, lysine 4 of histone 3) and arginine residues (*e.g.*, those on H3 and H4) of histones can result in transcriptional activation (7). While histone methylation was previously thought to be a permanent modification, recent studies uncovered histone demethylating enzymes (45). It is now known that demethylation of histone proteins is performed by histone demethylases. Lysine-specific demethylase 1 (a flavin-dependent monoamine oxidase) can demethylate mono- and di-methylated lysines, specifically lysines 4 and 9 of histone 3 (H3K4 and H3K9) (54). This enzyme cannot demethylate trimethylated lysines. However, the Jumonji domain-containing (JmjC) histone demethylases can demethylate mono-, di-, or tri-methylated lysines (187). It is possible that arginine-specific demethylase exists in cells to demethylate mono- and di-methylated arginines in histone proteins (19). Alternatively, methylated arginine residues can be converted into citrulline by peptidyl arginine deiminase 4, which acts on unmodified and mono-methylated arginine residues in proteins (7).

Histone acetylation also influences gene expression (43). Acetylation of lysine residues at the N terminus of histone proteins is catalyzed by histone acetyltransferases (HAT) (59). Superfamilies of HAT include *GNAT*, *CBP/p300*, and *MYST*. Examples of histone acetyltransferases are *HAT1*, *HATp300*, *CDY1*, and *CLOCK*. The source of the acetyl group in histone acetylation is acetyl-CoA. Thus, metabolism of energy substrates or the energy status of the cell can modulate histone acetylation (184). Notably, acetylation of lysine residues neutralizes the positive charge on histones, thereby decreasing the interaction of the N termini of histones with the negatively charged phosphate groups of DNA (187). Such a modification of histone proteins transforms the condensed chromatin into a more relaxed structure and makes RNA polymerase and transcription factors easier to access the promoter region of the DNA, leading to enhanced gene transcription. Thus, histone acetylation is usually associated with activation of gene transcription (59). Histone deacetyla-

tion is catalyzed by histone deacetylases (HDAC; also known as lysine deacetylases). In this reaction, the acetyl group is transferred from the acetylated histone protein to coenzyme A. Based on intracellular localization, three classes of HDAC are I: HDAC 1, 2, 3, and 8 (exclusively in the nucleus); II: HDAC 4, 5, 6, 7, 9, and 10 (predominantly residing in the cytoplasm and shuttling in and out of the nucleus); and III: sirtuin enzymes (SIRT) 1, 2, 3, 4, 5, 6, and 7 (localized in either nucleus, cytoplasm, or mitochondria) (187). The histone deacetylase *CBP/p300* can interact with numerous transcription regulators and likely plays a major role in repressing gene transcription (57).

The biological activity of histones is also regulated by phosphorylation. Histone H3 phosphorylation, which occurs during both mitosis and meiosis, is catalyzed by ATP-dependent protein kinases (26, 207). This event is coordinated in both space and time as a signaling mechanism to modulate nucleosome structure and thus DNA accessibility (73). Phosphorylation of H2A (at serine 1 and threonine T119), H3 (at serine 10 and 28; threonine 3 and 11), and H4 (at serine 1) results in transcriptional activation, as well as mitosis and meiosis (26, 131). Phosphorylation of H2A also enhances DNA repair, while phosphorylation of H4 promotes spermatogenesis. Likewise, histone H1 phosphorylation is associated with decondensation of chromatin and activation of transcription (2), and histone-3 phosphorylation is critical for cytokine-induced gene expression (207). In contrast, phosphorylation of H2B (at serine 14) stimulates apoptosis, which plays an important role in cell metabolism and growth (101). Thus, histone phosphorylation can allow cells to rapidly respond and adapt to changes in maternal nutritional status by regulating numerous processes, including transcription, DNA repair, and apoptosis (Fig. 5).

The monoubiquitination of histones H2A and H2B plays a critical role in regulating DNA repair, gene transcription, and mRNA translation (188). In this process, ubiquitin (a 76-amino acid polypeptide) is conjugated to histones H2A and H2B in a series of reactions involving three separate enzymatic activities: a) ATP-dependent ubiquitin activating enzyme (E1; activation of ubiquitin), b) ubiquitin-conjugating enzyme (E2; conjugation via a thioester bond to a cysteine residue); and c) ubiquitin-protein isopeptide ligase (E3; transfer of ubiquitin

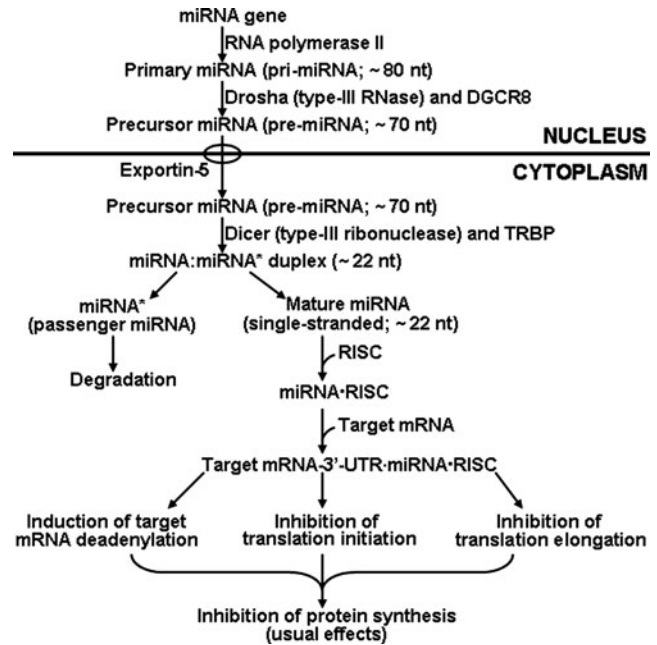


from the E2 enzyme to a target lysine residue in a histone) (135). Monoubiquitination of H2B promotes transcription initiation and elongation through efficient reassembly of nucleosomes (49). There is evidence that H2B monoubiquitination is a prerequisite for Lys-4 H3 and Lys-79 H3 methylation, whereas H2A monoubiquitination is associated with transcriptional silencing (187). Although H2A and H2B ubiquitination appear to perform separate functions, there is cross talk between ubiquitinated H2A and H2B proteins to modulate Lys-4 H3 methylation and RNA polymerase II activity during transcription elongation (49, 169).

#### RNA-based epigenetic mechanisms

Epigenetic regulation of gene expression by RNA-based mechanisms can occur at both the post-transcriptional level and the level of chromatin (187). These mechanisms are mediated by small-interfering RNAs or small non-coding RNAs which act through their respective pathways to induce DNA methylation or histone modifications to silence or enhance gene expression (82, 169). In mammals, the small non-coding RNAs include Piwi RNA and microRNA (miRNA) (74). Piwi RNA is restricted to the germ line (*e.g.*, testes in males and oocytes in females), and regulates gene expression possibly through sequence-specific targeting of heterochromatin formation factors to the mobile elements in the genome and the degradation of mobile element transcripts (172). In contrast, miRNAs are widespread in cells and tissue to regulate gene expression at the post-transcriptional level (44). Since the discovery of *lin-4* as the first miRNA in 1993 (93), approximately 1000 human miRNAs have been identified, annotated and catalogued (103, 113).

The biogenesis of mature (functional) miRNA from its miRNA gene involves both the nucleus and cytoplasm in mammalian cells (Fig. 6). In the nucleus, an miRNA gene is transcribed by RNA polymerase II to produce primary miRNA with ~80 nucleotides (nt) in length (44). The primary miRNA, which is a long capped and polyadenylated transcript, is processed by Droscha (type-III RNase) and its associated protein DGCR8 (Pasha) to generate precursor miRNA (~70 nt). After being transported to the cytoplasm by exportin-5, the precursor miRNA is cleaved by Dicer (type-III ribonuclease) in association with its binding partner TRBP (transacting RNA-binding protein) to generate a small imperfect miRNA:miRNA\* duplex (~22 nt) (74). This duplex is unwound to form a) a passenger strand miRNA (miRNA\*) for degradation; and b) a mature (functional) single-stranded miRNA with approximately 22 nt (72). The mature miRNA is loaded into an RNA-induced silencing complex (RISC) containing several components, including Dicer, TRBP, Argonaute proteins, GW182 protein, and fragile X mental retardation protein (FMRP1) (44). The mature miRNA then pairs with the 3'-untranslated region of the target mRNA, which usually results in: a) induction of target mRNA deadenylation and, thus, degradation; b) inhibition of the assembly of the 80S ribosome complex and, thus, translation initiation; and c) inhibition of translation elongation possibly through premature termination and subsequent ribosome drop-off (44). This leads to decreased synthesis of proteins which are either inhibitors or activators of metabolic pathways depending on cell types, tissues, and developmental stage. Therefore, miRNAs have versatile functions in physiological



**FIG. 6. Biogenesis of microRNA and its role in the regulation of gene expression.** The biogenesis of mature (functional) miRNA from its miRNA gene involves both the nucleus and cytoplasm in mammalian cells. A single-stranded mature (functional) miRNA with approximately 22 nucleotides (nt) binds the 3'-untranslated region of the target mRNA. This usually results in decreased protein synthesis through induction of mRNA deadenylation, reduction of translation initiation, and inhibition of translation elongation. DGCR8, associated protein DGCR8 of Droscha (also known as Pasha); TRBP (transacting RNA-binding protein); miRNA\*, passenger strand miRNA for degradation; RISC, RNA-induced silencing complex (RISC) containing several components, including Argonaute proteins, PW182 protein, and fragile X mental retardation protein.

processes and play crucial roles in growth, development, and health.

#### Aberrant Epigenetic Expression in the Metabolic Syndrome

##### Aberrant DNA methylation

Since the early report that dietary deficiency of methionine caused abnormal cell metabolism through DNA hypomethylation (194), there has been growing evidence indicating that disruption of epigenetic mechanisms results in metabolic disorders related to aging, obesity, and diabetes in animals and humans (46, 101). Aging is associated with decreased expression of *DNMT1* and *DNMT3*, global hypomethylation of DNA, and genomic instability in mammalian tissues, although gene-specific hypermethylation can occur (193). For example, increased DNA methylation in the *NDUFB6* promoter with aging is attributable to decreased expression of the *NDUFB6* gene and whole-body energy metabolism in humans (99). There is evidence that increased DNA methylation of the glucokinase promoter plays a role in decreasing expression of the glucokinase protein in the liver of aged rats (75), thereby increasing the risk for hyperglycemia.

Epigenetic defects (epimutations) can play a role in the development of obesity (3). Of interest, loss-of-function of the histone demethylase, *Jhdm2a*, is associated with reduced expression of peroxisome proliferator-activated receptor- $\alpha$  and medium-chain acyl-CoA dehydrogenase in skeletal muscle, impairment of cold-induced expression of uncoupling protein-1, and obesity in mice (165). Accordingly, hypomethylation increases risk of obesity in humans and animals (71). For example, maternal high-fat diet increased global and gene (dopamine and opioid) specific hypomethylation of DNA in the brains of offspring that resulted in alterations in gene expression and behavior (181). In addition, secretion of leptin by white adipose tissue is controlled by epigenetic mechanisms. Specifically, the leptin promoter that contains a CpG island has a high degree of methylation in preadipocytes (116, 208), and the promoter is demethylated in association with induction of leptin expression in differentiated adipocytes (208). Furthermore, a high-fat diet increases DNA methylation of one CpG site in the leptin promoter of adult rat adipocytes (119). Finally, NAD-dependent sirtuins (class III HDAC), which target both histones and nonhistone proteins, regulate nutrient metabolism, mitogenesis, adipogenesis, and insulin secretion (154). Additionally, defects in orchestration of circadian rhythm by HDAC3 lead to abnormal lipid metabolism in the liver (46).

Diabetes may also have an epigenetic basis (99, 100). First, recent work revealed that *COX7A1* (part of complex 4 of the respiratory chain) is a target of DNA methylation (144). Interestingly, methylation of the DNA for the *COX7A1* promoter is increased in skeletal muscle of diabetic animals, and its gene transcription is reduced in association with reduced glucose uptake by muscle (144). Second, genetic mutation in the *Pdx1*/insulin promoter factor-1, which is a transcription factor regulating development of the pancreas and  $\beta$ -cell differentiation, results in a monogenic form of diabetes (178). Interestingly, IUGR resulting from uteroplacental insufficiency is associated with the silencing of *Pdx1* and the development of type 2 diabetes in adult offspring of rats (130). Third, DNA methylation of the *PPARGC1A* gene is elevated in pancreatic islets from patients with type 2 diabetes and the degree of DNA methylation is inversely correlated with *PPARGC1A* expression (100). Fourth, hypomethylation at chromosome 6q24 is associated with transient neonatal diabetes in humans, and these subjects develop type 2 diabetes as adults (167).

#### *Aberrant histone modifications*

Abnormal modifications of histone proteins contribute to metabolic disorders (101). *GLUT4* is a major glucose transporter in insulin-sensitive tissues; therefore, intensive research on epigenetic regulation of diabetes has focused on this protein. In animal studies, reduced expression of *GLUT4* is associated with elevated HDAC activity and hyperglycemia (112). Normally, *GLUT4* expression is regulated by the transcription factor myocyte enhancer factor 2 (*MEF2*), which interacts with HDAC5 in the nucleus. However, when the histone tails that are colocalized with the *GLUT4* gene are deacetylated by HDAC5 in skeletal muscle, the formation of condense chromatin structures impairs *GLUT4* gene expression (111). In contrast, enhanced HAT activity to acetylate histones is positively correlated with oxygen consumption in human skeletal muscle (129). Similarly, Class II HDAC en-

zymes limit *GLUT4* gene expression during adipocyte differentiation (187). Likewise, histone code modifications by HDAC1 and HDAC4 repress *GLUT4* expression in IUGR offspring (140). Furthermore, phosphorylation of HDAC5 has a stimulatory effect on *GLUT4* transcription mediated by AMP-activated protein kinase in skeletal muscle (111).

A novel finding in epigenetic research is the regulation of insulin secretion by histone modifications (125, 156). In normal pancreatic  $\beta$ -cells, the insulin gene exhibits hyperacetylation of H4 and hypermethylation of H3K4. This pattern is not observed for the insulin gene in other cells (*e.g.*, HeLa cells) that do not produce insulin (27). Recruitment of histone acetyltransferase and the histone methyltransferase SET7/9 to the insulin promoter is necessary to activate the gene for insulin (27). HDAC may inhibit pancreatic development in the fetus (66). Also, reduced levels of histone H3K27 trimethylation and the histone methyltransferase *Ezh2*, as well as the loss of H2A ubiquitination are associated with elevated expression of *Ink4a/Arf* and reduced beta-cell regeneration in aging mice (88). Thus, reduced activity of HAT (*e.g.*, CBP/p300) is associated with a monogenic form of diabetes (8).

Another exciting new development in histone modifications is their crucial roles in cellular antioxidative reactions and cellular signaling. Poor glycemic control increases NF- $\kappa$ B activity in monocytes and expression of genes encoding inflammatory cytokines through an interaction between NF- $\kappa$ B and HAT (*e.g.*, CBP/p300) (117). The H3K4 methyltransferase SET7/9 affects the recruitment of NF- $\kappa$ B p65 to gene promoters and thus expression of NF- $\kappa$ B-dependent inflammatory genes (98). Histone methylase (SET7) and histone demethylase (LSD1) can regulate epigenetic changes in the NF- $\kappa$ B p65 promoter induced by hyperglycemia (19). Vascular smooth muscle cells from diabetic mice display reduced activities of H3K9 methyltransferase and lysine-specific histone demethylase-1, as well as reduced levels of H3K9 trimethylation and elevated levels of H3K4 dimethylation (179). Abnormal H3K9 dimethylation has also been reported for lymphocytes from patients with type-2 diabetes (118).

#### *Aberrant expression of small non-coding RNAs*

Aberrant expression of miRNAs has been implicated in obesity, insulin resistance, diabetes, cardiovascular complications, and placental dysfunction (72, 120). For example, high fat feeding results in upregulation of miR-143 but downregulation of primary miR-27a in mouse mesenteric adipose tissue via mechanisms involving expression of the PPAR-family of genes (113). Also, enhanced levels of miR-335 have been reported for the liver and white adipose tissues of three murine models of obesity (leptin-deficient *ob/ob* mice, leptin receptor-deficient *db/db* mice, and *KKAy* 44 mice) (122). Similarly, type-2 diabetes in humans is associated with down expression of miR-27a and miR-378, which promotes the formation of lipid droplets, adipocyte differentiation, and adipogenesis in white adipocytes (113). Furthermore, miR-17-5P and miR-132 in omental fat and blood are positively correlated with human obesity (68). Interestingly, emerging evidence indicates that miR-17-92, miR-21, miR-103, miR-143, miR-371, and miR-378 promote, but let-7, miR-27, miR-130, miR-138, miR-369-5P, and miR-448 inhibit, oxidative stress and inflammation in animals with obesity and atherosclerosis (72). Finally, mis-expression of miRNAs contributes to

inflammatory processes in placental dysfunction (120), cardiovascular diseases (e.g., coronary artery disease, ischemic injury, hypertension, and atherosclerosis) (158), the metabolic syndrome (103), and aging (158). Besides microRNAs, little is known about effects of nutrition on expression of other small non-coding RNAs in animal tissues.

### Inhibition to Regulate Epigenetics and Ameliorate Metabolic Syndrome

Environmental cues appear to be the primary trigger to induce changes in the epigenome and include such factors as nutrients, stress, environmental pollutants, and toxins (55). The extent to which cells are able to respond to these cues lies in large part with the plasticity of cells. Thus, the developing embryo/fetus is a prime target for epigenetic modifications (43) and interventions during pregnancy may be an important strategy for ameliorating or preventing metabolic disorders (including vascular insulin resistance) in adult life (Fig. 7). Despite a recognized role for epigenetics in fetal programming for metabolic syndrome (175), work in this field remains very limited.

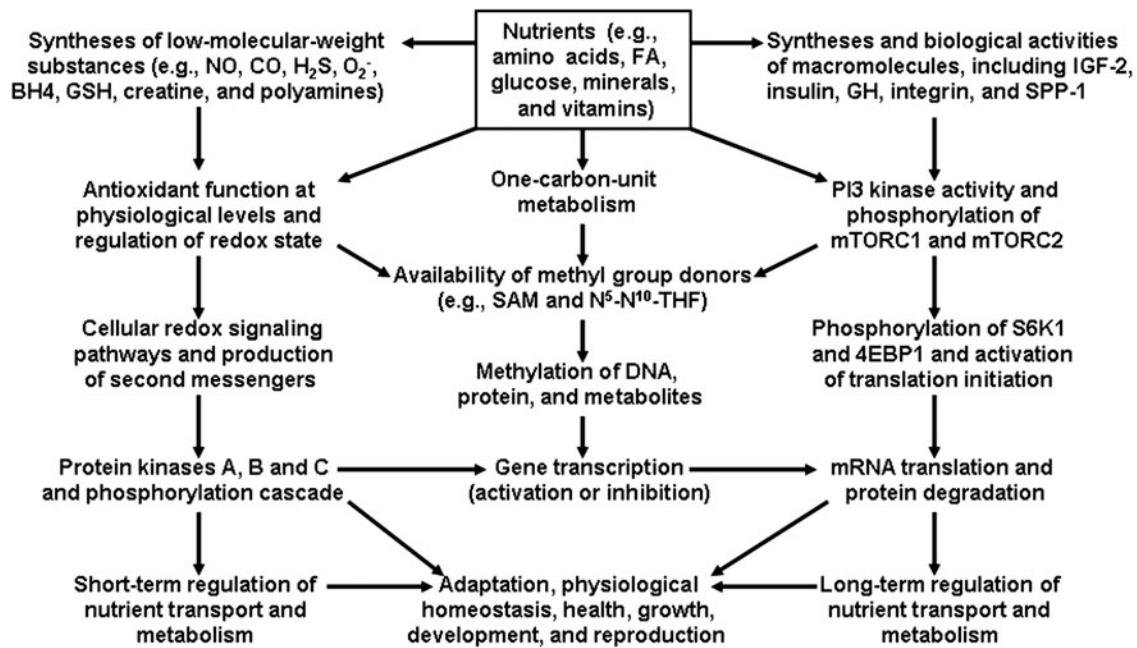
#### Inhibition of DNA methylation and histone deacetylation

Based on epigenetic research, chemically synthesized drugs have been used to target deleterious effects of DNA methyl-

ation and histone deacetylation to reverse metabolic disorders (164). The classic DNMT inhibitor is 5-azacytidine, a derivative of cytidine. Results of both *in vitro* and *in vivo* studies indicate that 5-azacytidine increases expression of the PPAR $\gamma$  gene in white adipocytes in a dose-dependent manner and the metabolic pattern of the cells (53). Also of particular interest is the application of isoform-specific HDAC inhibitors (e.g., short-chain fatty acids, hydroxamic acids, cyclic tetrapeptides, and benzamides) to treat vascular complications associated with diabetes (17, 60). For example, myocardial ischemia-reperfusion injury is associated with increased HDAC activity, but decreased acetylation of histones H3 and H4 in the heart of mice (57). Notably, administration of synthetic HDAC inhibitors (trichostatin A and Scriptaid) to these animals reduced myocardial infarction and cell death (57). Similarly, HDAC inhibitors attenuated load- and agonist-induced cardiac hypertrophy and abolished the associated activation of autophagy in cardiomyocytes (24).

#### Inhibition of miRNA expression and activity

Because abnormally high levels of some miRNAs cause translational repression in tissues (particularly insulin-sensitive tissues) and are associated with the development of chronic diseases (72, 113), modulation of their expression and activity may be a potentially effective therapeutic means for prevention and treatment of the metabolic syndrome. At



**FIG. 7. Roles of macro- and micro-nutrients in epigenetics and physiological responses.** Nutrients, particularly amino acids, regulate cellular redox state, the secretion of hormones (e.g., insulin and insulin-like growth factors), physiological functions, and whole-body homeostasis in humans and animals through three mechanisms: (1) the expression of genes; (2) the production of signaling gases and other metabolites; and (3) MTOR activation. S-Adenosylmethionine is the major methyl group donor in cells and its synthesis is affected by amino acids (e.g., methionine, serine, glycine, and histidine), B vitamins (including folate, vitamin B<sub>12</sub>, and vitamin B<sub>6</sub>), choline, and creatine. Methylation of DNA and protein contributes to epigenetics, which results in transcriptional activation or inhibition of select genes. Changes in intracellular protein turnover (protein synthesis and degradation) and protein kinase cascades can alter physiological responses in the fetus and offspring. CO, carbon monoxide; 4EBP1, eIF4E-binding protein-1; GH, growth hormone; H<sub>2</sub>S, hydrogen sulfide; IGF, insulin-like growth factor; MTOR, mechanistic target of rapamycin; NO, nitric oxide; SAM, S-adenosylmethionine; S6K1, ribosomal protein S6 kinase-1; SPP1, secreted phosphoprotein 1.



present, the expression or activity of a specific miRNA can be inhibited or silenced using: a) a locked nucleic acid oligonucleotide (a Morpholino oligonucleotide); b) an antisense 2'-O-methyl RNA oligonucleotide; c) an antagomir (a small synthetic RNA oligonucleotide that is perfectly complementary to the specific miRNA target), or d) a steric-blocking oligonucleotide to prevent the maturation of miRNA or block the miRNA-binding site of a target mRNA (40, 89). Although research in this area is still limited, results from both *in vitro* and *in vivo* studies have been promising. For example, specific inhibition of miR-375 (a pancreatic islet-specific microRNA) by transfection with an antisense 2'-O-methyl oligonucleotide complementary to miR-375 decreased the expression of the *myotrophin* and *PDK1* genes and increased glucose-stimulated insulin secretion in cultured murine pancreatic  $\beta$ -cells (138). The use of a similar technology led to the discovery that miR-143 plays a key role in the differentiation of cultured human pre-adipocytes by inhibiting expression of the *ERK5* gene (41). Based on the *in vitro* studies, intravenous administration of an antagomir against miR-122 three times within 24 h (80 mg/kg body weight/day) to mice reduced expression of 11 genes involved in cholesterol synthesis (including hydroxy-3-methylglutaryl-CoA reductase) in the liver, thereby decreasing concentrations of cholesterol in plasma (89). Likewise, subcutaneous injection of an anti-miR-33 oligonucleotide to mice weekly for 4 weeks promoted the reversal transport of cholesterol from peripheral tissues to the plasma, liver, and feces, as well as regression of established atherosclerosis, and ameliorated dyslipidemia (as indicated by reduced concentrations of triacylglycerols and cholesterol in plasma) (141). Employing the similar technology, Rayner *et al.* (142) recently reported that inhibition of miR-33a/b increased the expression of: a) the sterol-response-element-binding protein genes *SREBF1* and *SREBF2*; b) the cholesterol transporter gene *ABCA1*; and c) genes involved in fatty acid oxidation (*CROT*, *CPT1A*, *HADHB*, and *PRKAA1*), while reducing the expression of genes involved in fatty acid synthesis (*SREBF1*, *FASN*, *ACLY*, and *ACACA*) in nonhuman primates. This anti-microRNA strategy resulted in increased concentrations of high-density lipoproteins and reduced concentrations of very-low-density lipoproteins in plasma to reduce risk for cardiovascular disease.

#### Targeting epigenetically disturbed metabolic pathways

Maternal malnutrition may affect growth and health of F1 and subsequent offspring (200). It is now known that IUGR is associated with reduced expression of the imprinted gene *IGF2* in animals (52). Interestingly, loss of imprinting of the *IGF2* gene results in enhanced signaling of *IGF2/IGF1* at the receptor level due to a positive feedback mechanism (35) which leads to abnormal metabolism of nutrients (*e.g.*, uncontrolled utilization of glucose via glycolysis) (170). Inhibition of the *IGF2* receptor can reverse the epigenetically disturbed metabolic pathways in animals with loss of imprinting of the *IGF2* gene (81). Additionally, results of recent studies indicate that alterations in DNA methylation and histone modifications in cardiovascular diseases all result in elevated levels of hyperhomocysteinemia due to reduced expression of the enzymes involved in the utilization of homocysteine (25, 47, 87). Accordingly, reducing circulating levels of homocysteine through dietary supplementation with

folic acid can improve function of the cardiovascular system (139, 191).

#### Nutritional interventions with amino acids and vitamins

Besides serving as building blocks of proteins, amino acids are signaling molecules, regulators of gene expression and the protein phosphorylation cascade, and they are key precursors for syntheses of hormones and low-molecular weight nitrogenous substances with enormous biological importance (Fig. 7). Amino acids also modulate cellular redox state and the secretion of hormones from endocrine organs (*e.g.*, insulin, growth hormone, lactogen, and insulin-like factors) (197). Physiological concentrations of metabolites (*e.g.*, nitric oxide, polyamines, glutathione, taurine, thyroid hormones, and serotonin) of amino acids are required for the functions of cells and whole-body homeostasis (197). It is noteworthy that maintenance and regulation of the epigenetic state, which depends on one-carbon unit metabolism, require adequate provision of methionine, serine, glycine, histidine, choline, creatine, and B vitamins (including folate, vitamin B<sub>12</sub>, and vitamin B<sub>6</sub>) (Fig. 2). These nutrients play an important role in regulating the availability of S-adenosylmethionine, a major methyl donor for DNA and protein methylation by specific DNA and protein methyltransferases (200). Thus, restriction of essential B vitamins, folate, and methionine during the periconceptual period in sheep resulted in altered DNA methylation, insulin resistance, and elevated blood pressure observed most notably in adult male offspring (157). Conversely, supplementing the maternal diet with folate, vitamin B12, choline, or betaine increases DNA methylation of the agouti gene in the offspring, thereby leading to low agouti expression and preventing the development of obesity (77). Interestingly, the effect of maternal supplementation with methyl donors can be inherited in the F2 generation through germline epigenetic modifications (34).

S-Adenosylmethionine is synthesized from methionine by S-adenosylmethionine synthase (also known as methionine adenosyltransferase) (197). In addition to its role as a methyl donor, S-adenosylmethionine is required for the synthesis of polyamines, cysteine, taurine, and creatine. Polyamines are required for the proliferation of endothelial cells and the remodeling of the vasculature (96). Importantly, the nutritional and physiological state of the animal will alter the production of these bioactive substances, thus regulating the availability of methyl donors. When cysteine or taurine is deficient in the diet, their synthesis from methionine will be increased *in vivo*, thus decreasing total S-adenosylmethionine availability for DNA or protein methylation. Inadequate synthesis of glycine and serine, coupled with low supplies from the diet, can also impair one-carbon unit metabolism (200). Therefore, an amino acid deficiency can alter the epigenetic code through changes in DNA methylation as well as histone modifications (127).

Both epidemiological and experimental evidence indicate that IUGR contributes to a plethora of metabolic disorders and chronic diseases in adults (Fig. 1). These problems include: 1) hormonal imbalances (*e.g.*, increased plasma levels of glucocorticoids and renin; decreased plasma levels of insulin, growth hormone, and insulin-like growth factor-I); 2) metabolic disorders (*e.g.*, insulin resistance,  $\beta$ -cell dysfunction, dyslipidemia, glucose intolerance, impaired energy homeostasis, obesity, type-II diabetes, oxidative stress,

mitochondrial dysfunction, and aging); 3) organ dysfunction and abnormal development (*e.g.*, testes, ovaries, brain, heart, skeletal muscle, liver, thymus, and small intestine); and 4) cardiovascular disorders (*e.g.*, coronary heart disease, hypertension, stroke, and atherosclerosis) (3, 156, 162, 199). These novel findings led to the design of nutritional means (including supplementation with functional amino acids) to enhance fetal growth and development (200).

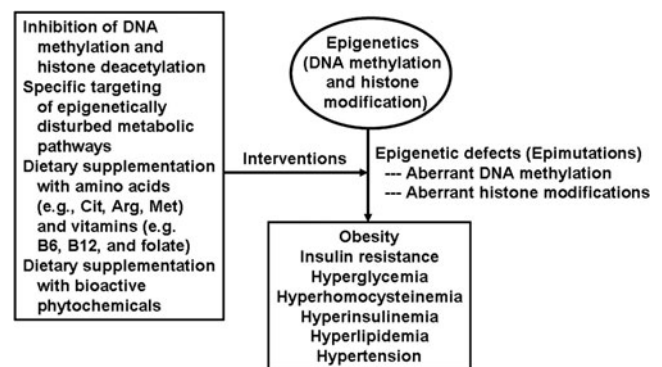
Enteral feeding or intravenous administration of amino acids (*e.g.*, arginine and citrulline) is effective in increasing their circulating concentrations in the mother and her fetus(es) (90–92). Therefore, these nutrients may provide an effective solution to fetal growth restriction in underfed and overfed dams and postnatal metabolic disorders. The importance of amino acids in supporting fetal growth, development, and health is an emerging area of investigation and will undoubtedly shape the future of nutritional management in medicine and animal production (201). The translation of basic research on amino acid nutrition into practice has yielded fruitful outcomes. For example, in pigs, arginine supplementation to pregnant sows during gestation increases placental vascularity (97) and embryonic/fetal survival (105). Interestingly, piglets from gilts that received arginine supplementation during gestation have higher rates of neonatal survival and growth (203). Additionally, supplementing the gestational diet for gilts with 0.4% L-arginine plus 0.6% L-glutamine between Days 30 and 114 of gestation enhanced the efficiency of nutrient utilization, reduced variation in piglet birth weight, and increased litter birth weight (203). In sheep, maternal arginine administration during late gestation increased the development of fetal peri-renal brown adipose tissue (152), which is enriched with endothelial cells. The increased mass of brown adipose tissue may lead to enhancement of blood flow to tissues and the ability of neonates to combat cold exposure at birth. Moreover, intravenous administration of L-arginine-HCl (3 × 27 mg/kg body weight per day) enhanced fetal growth in ovine models of both undernutrition-induced and naturally-occurring IUGR (91). Finally, during late (Week 33) gestation, daily intravenous infusion of L-arginine (20 g/day) for 7 days to women with unknown causes of IUGR increased birth weight at term (Week 39) by 6.4% (205). The beneficial effects of arginine likely result from increases in angiogenesis, the number and size of blood vessels, and ultimately the transfer of nutrients from the mother to the conceptus through utero-placental blood flows (151, 202). Future studies are required to determine cardiovascular function and health of offspring from mothers supplemented with arginine during pregnancy.

Physiological effects of supplemental nutrients depend on many factors, including species, developmental stage, nutritional status, dose, duration, window of receptivity, and the content of dietary components. Thus, too much of a good thing can be detrimental to health and care must be taken to ensure the safety of use of nutrients in epigenetic interventions. For example, a high dose of supplemental arginine can disturb acid-base balance and cause antagonism among basic amino acids to yield undesirable effects (*e.g.*, reduced growth, skin lesions, and organ dysfunction) on animals and humans (196). For this reason, it is important that arginine be administered in divided doses on each day to: a) prevent gastrointestinal tract disorders due to abrupt production of large amounts of NO; b) increase the availability of circulating arginine over a longer

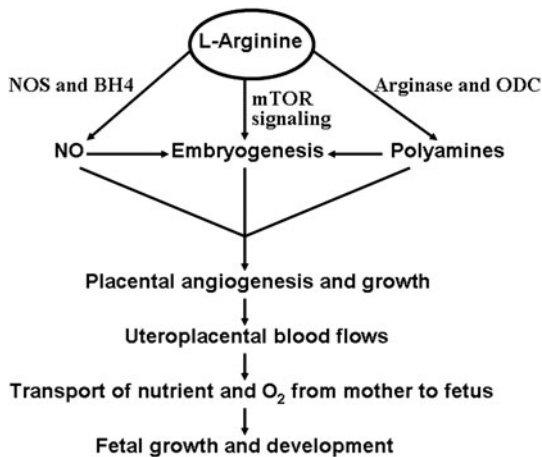
period of time; and c) avoid a potential imbalance among dietary amino acids. Likewise, dietary supplementation with high doses of folate may not benefit all the people and rather may cause harm to some subjects. For example, high intake of dietary folate by animals and humans may reduce the cytotoxicity of natural killer cells, impair the response to antifolate-based drugs, and result in phenotypic changes in subsequent generations (159). Additionally, because folate interferes with the actions of vitamin B12-dependent enzymes, excess folate in diet can worsen vitamin B12 deficiency in subjects whose diets are inadequate in vitamin B12, which may increase risks for: a) cognitive impairment and anemia in the elderly; and b) in pregnant mothers, adverse fetal programming on insulin resistance and obesity in their offspring (159).

#### Nutritional interventions with bioactive phytochemicals

Besides specific nutrients, botanicals have been identified as possible epigenetic modulators to ameliorate metabolic syndrome by regulating expression and activity of DNMT, HMT, HAT, and HDAC (86). For example, PMI 5011 (*A. dracunculoides* L), which has hypoglycemic activity in diabetic animals, reduced *DNMT1* and *DNMT3b* gene expression in NIH 3T3 fibroblasts. Similar results were obtained for ALT-S (*A. tuberosum* L) (86). There are also reports that phytochemicals (including epigallocatechingallate, resveratrol, genistein, curcumin, and isothiocyanates) can interfere with enzymatic activities of DNMT, Class I, II, IV HDAC, HAT, and Class III HDAC sirtuins (SIRT), thereby beneficially modulating inflammatory responses and immunological senescence (163). Plant extracts may also affect interactions among histone proteins or between nucleosomes and binding factors. Thus, dietary ingredients in food can be epigenetic medicine to remedy abnormalities in DNA methylation, histone modifications, chromatin remodeling, and microRNA patterns. Based on current knowledge of epigenetics, we propose pharmacological and nutritional interventions to prevent and treat metabolic disorders caused by epigenetic defects (Fig. 8). However, as noted above for folate supplementation, cautions



**FIG. 8. Pharmacological and nutritional interventions of epigenetic defects.** Epigenetic defects can occur in response to environmental stress (*e.g.*, malnutrition and heat stress), abnormal endocrine status, and abnormal levels of metabolites in the body. Chemically synthesized drugs (*e.g.*, 5'-azacytidine as an inhibitor of DNA methyltransferase), physiological metabolites (*e.g.*, butyrate as an inhibitor of histone deacetylase), specific nutrients (*e.g.*, amino acids and vitamins), or bioactive phytochemicals may be used to prevent and treat epimutations.



**FIG. 9. Role of L-arginine in enhancing fetal growth and development in response to maternal undernutrition or overnutrition.** Through the tetrahydrobiopterin (BH4)-dependent synthesis of nitric oxide (NO) and production of polyamines from arginine-derived ornithine as well as activation of the mechanistic target of rapamycin (MTOR) signaling pathway, L-arginine can promote embryogenesis and placental vascular growth. Antioxidants are essential to maintain sufficient levels of BH4 in cells for NO synthase (NOS). Enhancement of uteroplacental blood flow ensures adequate transport of nutrients and oxygen from mother to fetus to support fetal growth and development under the conditions of maternal malnutrition. ODC, ornithine decarboxylase.

(e.g., timing, dose, frequency, and length of administration) should be taken when using epigenetic compounds to treat human disorders.

### Conclusion

Either undernutrition or overnutrition during pregnancy (particularly the periconception period), remains a significant problem in both medicine and animal agriculture as it can result in epigenetic changes of some genes in both animals and humans. These changes are affected by multiple factors (e.g., sex, gestational period, and the severity of malnutrition) and may persist in offspring throughout postnatal life, and may carry over to the next generation. A unified explanation for impairment of fetal growth and development in response to both maternal undernutrition and overnutrition may be reduced utero-placental blood flow and, therefore, the reduced transfer of nutrients from mother to fetus (Fig. 9). This hypothesis is gaining support from results of studies with animal models, including rats, pigs, and sheep (114, 145, 160, 211). Nutrients, particularly amino acids and B vitamins, are essential for the regulation of epigenetics and vascular function. Compelling evidence indicates that the fetal and early neonatal periods of development are extremely sensitive to environmental cues, which have long-lasting consequences to postnatal growth, health, and likely athletic performance. Much research is needed with both cell and animal models to understand basic mechanisms responsible for effects mediated by specific nutrients in fetal and neonatal programming. Such new knowledge is expected to aid in designing effective therapeutic means for metabolic abnormalities in offspring that experience an adverse intrauterine environment.

### Acknowledgments

This work was supported, in part, by grants from the Thousand-People-Talent program at China Agricultural University, National Natural Science Foundation of China (3081010390, 30972156, 31129006, and 31172217), National Institutes of Health (1R21 HD049449), American Heart Association (10GRNT4480020), National Research Initiative Competitive Grants from the Animal Reproduction Program (2008-35203-19120), and Animal Growth & Nutrient Utilization Program (2008-35206-18764 and 2009-35206-05211) of the USDA National Institute of Food and Agriculture, and Texas AgriLife Research Hatch Project (H-8200).

### Author Disclosure Statement

For all authors, there are no financial or other contractual agreements that might cause conflicts of interest or be perceived as causing conflicts of interest.

### References

1. Aagaard-Tillery KM, Grove K, Bishop J, Ke X, Fu Q, McKnight R, and Lane RH. Developmental origins of disease and determinants of chromatin structure: Maternal diet modifies the primate fetal epigenome. *J Mol Endocrinol* 41: 91–102, 2008.
2. Ahmad W, Shabbiri K, Nazar N, Nazar S, Qaiser S, and Mughal MAS. Human linker histones: Interplay between phosphorylation and O- $\beta$ -GlcNAc to mediate chromatin structural modifications. *Cell Div* 6: 15, 2011.
3. Ainge H, Thompson C, Ozanne SE, and Rooney KB. A systematic review on animal models of maternal high fat feeding and offspring glycaemic control. *Int J Obes* 35: 325–335, 2011.
4. Ambros V. microRNAs: Tiny regulators with great potential. *Cell* 107: 823–826, 2001.
5. Anderson CM, Lopez F, Zimmer A, and Benoit JN. Placental insufficiency leads to developmental hypertension and mesenteric artery dysfunction in two generations of Sprague-Dawley rat offspring. *Biol Reprod* 74: 538–544, 2006.
6. Armitage JA, Lakasing L, Taylor PD, Balachandran AA, Jensen RI, Dekou V, Ashton N, Nyengaard JR, and Poston L. Developmental programming of aortic and renal structure in offspring of rats fed fat-rich diets in pregnancy. *J Physiol* 565: 171–184, 2005.
7. Bannister AJ and Kouzarides T. Reversing histone methylation. *Nature* 436: 1103–1106, 2005.
8. Barbacci E, Chalkiadaki A, Masdeu C, Haumaitre C, Lokmane L, Loirat C, Cloarec S, Talianidis I, Bellanne-Chantelot C, and Cereghini S. HNF1 $\beta$ /TCF2 mutations impair transactivation potential through altered coregulator recruitment. *Hum Mol Genet* 13: 3139–3149, 2004.
9. Barker DJP and Osmond C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. *Lancet* 8489: 1077–1081, 1986.
10. Barker DJP. Growth and living conditions in childhood and hypertension in adult life: A longitudinal study. *J Hypertens* 20: 1951–1956, 2002.
11. Barker DJP. The origins of the developmental origins theory. *J Intern Med* 261: 412–417, 2007.
12. Barlow DP. Gametic imprinting in mammals. *Science* 270: 1610–1613, 1995.
13. Bazer FW, Wu G, Spencer TE, Johnson GA, Burghardt RC, and Bayless K. Novel pathways for implantation and



- establishment and maintenance of pregnancy in mammals. *Mol Human Reprod* 16: 135–152, 2010.
14. Bazer FW, Wu G, Johnson GA, Kim JY, and Song GW. Uterine histotroph and conceptus development: Select nutrients and secreted phosphoprotein 1 affect MTOR cell signaling in ewes. *Biol Reprod* 85: 1094–1107.
  15. Berard J and Bee G. Effects of dietary L-arginine supplementation to gilts during early gestation on foetal survival, growth and myofiber formation. *Animal* 4: 1680–1687, 2010.
  16. Bertram C, Khan O, Ohri S, Phillips DI, Matthews SG, and Hanson MA. Transgenerational effects of prenatal nutrient restriction on cardiovascular and hypothalamic-pituitary-adrenal function. *J Physiol* 586: 2217–2229, 2008.
  17. Bieliauskas AV and Pflum MK. Isoform-selective histone deacetylase inhibitors. *Chem Soc Rev* 37: 1402–1413, 2008.
  18. Bird A. Molecular biology. DNA methylation *de novo*. *Science* 286: 2287–2288, 1999.
  19. Brasacchio D, Okabe J, Tikellis C, Balcerzyk A, George P, Baker EK, Calkin AC, Brownlee M, Cooper ME, and El-Osta A. Hyperglycemia induces a dynamic cooperativity of histone methylase and demethylase enzymes associated with gene-activating epigenetic marks that coexist on the lysine tail. *Diabetes* 58: 1229–1236, 2009.
  20. Brown KD and Robertson KD. DNMT1 knockout delivers a strong blow to genome stability and cell viability. *Nat Genet* 39: 289–290, 2007.
  21. Burdge GC and Lillycrop KA. Nutrition, epigenetics, and developmental plasticity: Implications for understanding human disease. *Annu Rev Nutr* 30: 315–339, 2010.
  22. Bygren LO, Kaati G, and Edvinsson S. Longevity determined by paternal ancestors' nutrition during their slow growth period. *Acta Biotheor* 49: 53–59, 2001.
  23. Cafe LM, Hennessy DW, Hearnshaw H, Morris SG, and Greenwood PL. Consequences of prenatal and preweaning growth for feedlot growth, intake and efficiency of Piedmontese- and Wagyu-sired cattle. *Anim Prod Sci* 49: 461–467, 2009.
  24. Cao DJ, Wang ZV, Battiprolu PK, Jiang N, Morales CR, Kong YL, Rothermel BA, Gillette TG, and Hill JA. Histone deacetylase (HDAC) inhibitors attenuate cardiac hypertrophy by suppressing autophagy. *Proc Natl Acad Sci USA* 108: 4123–4128, 2011.
  25. Castro R, Rivera I, Blom HJ, Jakobs C, and Tavares de Almeida I. Homocysteine metabolism, hyperhomocysteinaemia and vascular disease: An overview. *J Inherit Metab Dis* 29: 3–20, 2006.
  26. Cerutti H and Casas-Mollano JA. Histone H3 phosphorylation: Universal code or lineage specific dialects? *Epigenetics* 4: 71–75, 2009.
  27. Chakrabarti SK, Francis J, Ziesmann SM, Garmey JC, and Mirmira RG. Covalent histone modifications underline the developmental regulation of insulin gene transcription in pancreatic beta cells. *J Biol Chem* 278: 23617–23623, 2003.
  28. Chan LL, Sébert SP, Hyatt MA, Stephenson T, Budge H, Symonds ME, and Gardner DS. Effect of maternal nutrient restriction from early to midgestation on cardiac function and metabolism after adolescent-onset obesity. *Am J Physiol Regul Integr Comp Physiol* 296: R1455–R1463, 2009.
  29. Chen ZJ and Pikaard CS. Epigenetic silencing of RNA polymerase I transcription: A role for DNA methylation in nucleolar dominance. *Genes Dev* 11: 2124–2136, 1997.
  30. Chmurzynska A. Fetal programming: Link between early nutrition, DNA methylation, and complex diseases. *Nutr Rev* 68: 87–98, 2010.
  31. Christian P. Micronutrients, birthweight, and survival. *Annu Rev Nutr* 30: 83–104, 2010.
  32. Cleal JK, Poore KR, Boullin JP, Khan O, Chau R, Hambidge O, Torrens C, Newman JP, Poston L, Noakes DE, Hanson MA, and Green LR. Mismatched pre- and postnatal nutrition leads to cardiovascular dysfunction and altered renal function in adulthood. *Proc Natl Acad Sci USA* 104: 9529–9533, 2007.
  33. Cox J, Williams S, Grove K, Lane RH, and Aagaard-Tillery KM. A maternal high-fat diet is accompanied by alterations in the fetal primate metabolome. *Am J Obstet Gynecol* 201: 281.e281–289, 2009.
  34. Cropley JE, Suter CM, Beckman KB, and Martin DI. Germline epigenetic modification of the murine A vy allele by nutritional supplementation. *Proc Natl Acad Sci USA* 103: 17308–17312, 2006.
  35. Cui H, Cruz-Correa M, Giardiello FM, Hutcheon DF, Kafonek DR, Brandenburg S, Wu Y, He X, Powe NR, and Feinberg AP. Loss of IGF2 imprinting: A potential marker of colorectal cancer risk. *Science* 299: 1753–1755, 2003.
  36. Dindot SV, Kent KC, Evers B, Loskutoff N, Womack J, and Piedrahita JA. Conservation of genomic imprinting at the XIST, IGF2 and GTL2 loci in the bovine. *Mamm Genome* 15: 966–974, 2004.
  37. Dindot SV, Person R, Strivens M, Garcia R, and Beaudet AL. Epigenetic profiling at mouse imprinted gene clusters reveals novel epigenetic and genetic features at differentially methylated regions. *Genome Res* 19: 1374–1383, 2009.
  38. Dunn GA and Bale TL. Maternal high-fat diet promotes body length increases and insulin insensitivity in second-generation mice. *Endocrinology* 150: 4999–5009, 2009.
  39. Edwards LE, Hellerstedt WL, Alton IR, Story M, and Himes JH. Pregnancy complications and birth outcomes in obese and normal-weight women: Effects of gestational weight change. *Obstet Gynecol* 87: 389–394, 1996.
  40. Elmen J, Lindow M, Schutz S, Lawrence M, Petri A, Obad S, Lindholm M, Hedtjarn M, Hansen HF, Berger U, Gullans S, Kearney P, Sarnow P, Straarup EM, and Kauppinen S. LNA-mediated microRNA silencing in nonhuman primates. *Nature* 452: 896–899, 2008.
  41. Esau C, Kang X, Peralta E, Hanson E, Marcusson EG, Ravichandran LV, Sun Y, Koo S, Perera RJ, Jain R, Dean NM, Freier SM, Bennett CF, Lollo B, and Griffey R. MicroRNA-143 regulates adipocyte differentiation. *J Biol Chem* 279: 52361–52365, 2004.
  42. Eriksson JG, Forsen T, Tuomilehto J, Osmond C, and Barker DJP. Early growth and coronary heart disease in later life: Longitudinal study. *Br Med J* 322: 949–953, 2001.
  43. Everitts AG, Zee BM, and Garcia BA. Modern approaches for investigating epigenetic signaling pathways. *J Appl Physiol* 109: 927–933, 2010.
  44. Fabian MR, Sonenberg N, and Filipowicz W. Regulation of mRNA translation and stability by microRNAs. *Annu Rev Biochem* 79: 351–379, 2010.
  45. Feinberg AP. Epigenetics at the epicenter of modern medicine. *JAMA* 299: 1345–1350, 2008.
  46. Feng D, Liu T, Sun Z, Bugge A, Mullican SE, Alenghat T, Liu XS, and Lazar MA. A circadian rhythm orchestrated by histone deacetylase 3 controls hepatic lipid metabolism. *Science* 331: 1315–1319, 2011.
  47. Fernandez AZ, Siebel AL, and El-Osta A. Atherogenic factors and their epigenetic relationships. *Int J Vasc Med* 2010: 437809, 2010.

48. Flegal KM, Carroll MD, Ogden CL, and Curtin LR. Prevalence and trends in obesity among US adults, 1999–2008. *JAMA* 303: 235–241, 2010.
49. Fleming AB, Kao CF, Hillyer C, Pikaart M, Osley MA. H2B ubiquitylation plays a role in nucleosome dynamics during transcription elongation. *Mol Cell* 31: 57–66, 2008.
50. Ford SP, Zhang L, Zhu M, Miller MM, Smith DT, Hess BW, Moss GE, Nathanielsz PW, and Nijland MJ. Maternal obesity accelerates fetal pancreatic beta- cell but not alpha-cell development in sheep: Prenatal consequences. *Am J Physiol Regul Integr Comp Physiol* 297: R835–843, 2009.
51. Forsen T, Eriksson JG, Tuomilehto J, Osmond C, and Barker, DJP. Growth in utero and during childhood among women who develop coronary disease: Longitudinal study. *Br Med J* 319: 1403–1407, 1999.
52. Fowden AL, Ward JW, Wooding FP, Forhead AJ, and Constancia M. Programming placental nutrient transport capacity. *J Physiol* 572: 5–15, 2006.
53. Fujiki K, Kano F, Shiota K, and Murata M. Expression of the peroxisome proliferator activated receptor gamma gene is repressed by DNA methylation in visceral adipose tissue of mouse models of diabetes. *BMC Biol* 7: 38, 2009.
54. Godmann M, Auger V, Ferraroni-Aguiar V, Di Sauro A, Sette C, Behr R, and Kimmins S. Dynamic regulation of histone H3 methylation at lysine 4 in mammalian spermatogenesis. *Biol Reprod* 77: 754–764, 2007.
55. Godfrey KM, Lillycrop KA, Burdge GC, Gluckman PD, and Hanson MA. Epigenetic mechanisms and the mismatch concept of the developmental origins of health and disease. *Pediatr Res* 61: 5R–10R, 2007.
56. Golding MC, Williamson GL, Stroud TK, Westhusin ME, and Long CR. Examination of DNA methyltransferase expression in cloned embryo reveals an essential role for Dnmt1 in bovine development. *Mol Reprod Dev* 78: 306–317, 2011.
57. Granger A, Abdullah I, Huebner F, Stout A, Wang T, Huebner T, Epstein JA, and Gruber PJ. Histone deacetylase inhibition reduces myocardial ischemia-reperfusion injury in mice. *FASEB J* 22: 35–49, 2008.
58. Greenwood PL, Hunt AS, Hermanson JW, and Bell AW. Effects of birth weight and postnatal nutrition on neonatal sheep: II. Skeletal muscle growth and development. *J Anim Sci* 78: 50–61, 2000.
59. Grunstein M. Histone acetylation in chromatin structure and transcription. *Nature* 389: 349–352, 1997.
60. Haberland M, Montgomery RL, and Olsen EN. The many roles of histone deacetylation in development and physiology: Implications for disease and therapy. *Nat Rev Genet* 10: 32–42, 2009.
61. Hales CN and Barker DJ. Type 2 (non-insulin-dependent) diabetes mellitus: The thrifty phenotype hypothesis. *Diabetologia* 35: 595–601, 1992.
62. Han HC, Austin KJ, Nathanielsz PW, Ford SP, Nijland MJ, and Hansen TR. Maternal nutrient restriction alters gene expression in the ovine fetal heart. *J Physiol* 558: 111–121, 2004.
63. Hansen RS, Gartler SM, Scott CR, Chen SH, and Laird CD. Methylation analysis of CGG sites in the CPG island of the human FMR1 gene. *Hum Mol Genet* 1: 571–578, 1992.
64. Hardie DG, Hawley SA, and Scott JW. AMP-activated protein kinase: Development of the energy sensor concept. *J Physiol* 574: 7–15, 2006.
65. Harrison M and Langley-Evans SC. Intergenerational programming of impaired nephrogenesis and hypertension in rats following maternal protein restriction during pregnancy. *Br J Nutr* 101: 1020–1030, 2009.
66. Haumaitre C, Lenoir O, and Scharfmann R. Histone deacetylase inhibitors modify pancreatic cell fate determination and amplify endogenous progenitors. *Mol Cell Biol* 28: 6373–6383, 2008.
67. Heijmans BT, Tobi EW, Stein AD, Putter H, Blauw GJ, Susser ES, Slagboom PE, and Lumey LH. Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proc Natl Acad Sci USA* 105: 17046–17049, 2008.
68. Heneghan FM, Miller N, McAnena OJ, O'Brien T, and Kerin MJ. Differential miRNA expression in omental adipose tissue and in the circulation of obese patients identifies novel metabolic biomarkers. *J Clin Endocrinol Metab* 96: E846–850, 2011.
69. Hill DJ and Duville B. Pancreatic development and adult diabetes. *Pediatr Res* 48: 269–274, 2000.
70. Holliday R and Pugh JE. DNA modification mechanisms and gene activity during development. *Science* 187: 226–232, 1975.
71. Hosoki K, Ogata T, Kagami M, Tanaka T, and Saitoh S. Epimutation (hypomethylation) affecting the chromosome 14q32.2 in a girl with upd914)mat-like phenotype. *Eur J Hum Genet* 16: 1019–1023, 2008.
72. Hulsmans M, De Keyzer D, and Holvoet P. MicroRNAs regulating oxidative stress and inflammation in relation to obesity and atherosclerosis. *FASEB J* 25: 2515–2527, 2011.
73. Hurd PJ, Bannister AJ, Halls K, Dawson MA, Vermeulen M, Olsen JV, Ismail H, Somers J, Mann M, Owen-Hughes T, Gout I, and Kouzarides T. Phosphorylation of histone H3 Thr-45 is linked to apoptosis. *J Biol Chem* 284: 16575–16583, 2009.
74. Ilnytsky Y and Kovalchuk O. Non-targeted radiation effects—An epigenetic connection. *Mutat Res* 714: 113–25, 2011.
75. Jiang MH, Fei J, Lan MS, Lu ZP, Liu M, Fan WW, Gao X, and Lu DR. Hypermethylation of hepatic Gck promoter in ageing rats contributes to diabetogenic potential. *Diabetologia* 51: 1525–1533, 2008.
76. Jimenez-Chillaron JC, Isganaitis E, Charalambos M, Gesta S, Pentinat-Pelegrin T, Faucette RR, Otis JP, Chow A, Diaz R, Ferguson-Smith A, and Patti ME. Intergenerational transmission of glucose intolerance and obesity by in utero undernutrition in mice. *Diabetes* 58: 460–468, 2009.
77. Jirtle RL and Skinner MK. Environmental epigenomics and disease susceptibility. *Nat Rev Genet* 8: 253–262, 2007.
78. Jobgen WS, Fried SK, Fu WJ, Meininger CJ, and Wu G. Regulatory role for the arginine-nitric oxide pathway in metabolism of energy substrates. *J Nutr Biochem* 17: 571–588, 2006.
79. Kaati G, Bygren LO, Pembrey M, and Sjöström M. Trans-generational response to nutrition, early life circumstances and longevity. *Eur J Hum Genet* 15: 784–790, 2007.
80. Kajantie E, Osmond C, Barker DJP, Forsen T, Phillips DIW, and Eriksson JG. Size at birth as a predictor of mortality in adulthood: A follow-up of 350 000 person-years. *Int J Epidemiol* 34: 655–663, 2005.
81. Kaneda A, Wang CJ, Cheong R, Timp W, Onyango P, Wen B, Iacobuzio-Donahue CA, Ohlsson R, Andraos R, Pearson MA, Sharov AA, Longo DL, Ko MS, Levchenko A, and Feinberg AP. Enhanced sensitivity to IGF-II signaling links loss of imprinting of IGF2 to increased cell pro-

- liferation and tumor risk. *Proc Natl Acad Sci USA* 104: 20926–20931, 2007.
82. Kawasaki H, Taira K, and Morris KV. siRNA induced transcriptional gene silencing in mammalian cells. *Cell Cycle* 4: 442–448, 2005.
  83. Khan IY, Dekou V, Douglas G, Jensen R, Hanson MA, Poston L, and Taylor PD. A high-fat diet during rat pregnancy or suckling induces cardiovascular dysfunction in adult offspring. *Am J Physiol Regul Integr Comp Physiol* 288: R127–133, 2005.
  84. Khan KS, Wojdyla D, Say L, Gulmezoglu AM, and Van Look PF. WHO analysis of causes of maternal death: A systematic review. *Lancet* 367: 1066–1074, 2006.
  85. Kiel DW, Dodson EA, Artal R, Boehmer TK, and Leet TL. Gestational weight gain and pregnancy outcomes in obese women. *Obstet Gynecol* 110: 752–758, 2007.
  86. Kirk H, Cefalu WT, Ribnicky D, Liu Z, and Eilertsen KJ. Botanicals as epigenetic modulators for mechanisms contributing to development of metabolic syndrome. *Metabolism* 57: S16–S23, 2008.
  87. Krishna SM, Dear AE, Norman PE, and Golledge J. Genetic and epigenetic mechanisms and their possible role in abdominal aortic aneurysm. *Atherosclerosis* 212: 16–29, 2010.
  88. Krishnamurthy J, Ramsey MR, Ligon KL, Torrice C, Koh A, Bonner-Weir S, and Sharpless NE. p16INK4a induces an age-dependent decline in islet regeneration potential. *Nature* 443: 453–457, 2006.
  89. Krutzfeldt M, Rajewsky N, Braich R, Rajeev KG, Tuschl T, Manoharan M, and Stoffel M. Silencing of miRNAs *in vivo* with ‘antagomirs’. *Nature* 438: 685–689, 2005.
  90. Lassala A, Bazer FW, Cudd TA, Li P, Li XL, Satterfield MC, Spencer TE, and Wu G. Intravenous administration of L-citrulline to pregnant ewes is more effective than L-arginine for increasing arginine availability in the fetus. *J Nutr* 139: 660–665, 2009.
  91. Lassala A, Bazer FW, Cudd TA, Datta S, Keisler DH, Satterfield MC, Spencer TE, and Wu G. Parenteral administration of L-arginine prevents fetal growth restriction in undernourished ewes. *J Nutr* 140: 1242–1248, 2010.
  92. Lassala A, Bazer FW, Cudd TA, Datta S, Keisler DH, Satterfield MC, Spencer TE, and Wu G. Parenteral administration of L-arginine enhances fetal survival and growth in sheep carrying multiple pregnancies. *J Nutr* 141: 849–855, 2011.
  93. Lee RC, Feinbaum RL, and Ambross V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementary to *lin-14*. *Cell* 75: 843–854, 1993.
  94. Leon DA, Lithell HO, Vagero D, Koupilova I, Mohsen R, Berglund L, Lithell UB, and McKeigue PM. Reduced fetal growth rate and increased risk of death from ischaemic heart disease: Cohort study of 15000 Swedish men and women born 1915–29. *Br Med J* 317: 241–245, 1998.
  95. Lewis BP, Shih IH, Jones-Rhoades MW, Bartel DP, and Burge CB. Prediction of mammalian microRNA targets. *Cell* 115: 787–798, 2003.
  96. Li H, Meininger CJ, Hawker JR, Kelly KA, Morris SM, and Wu G. Activities of arginase I and II are limiting for endothelial cell proliferation. *Am J Physiol Regul Integr Comp Physiol* 282: R64–R69, 2002.
  97. Li XL, Bazer FW, Johnson GA, Burghardt RC, Erikson DW, Frank JW, Spencer TE, Shinzato I, and Wu G. Dietary supplementation with 0.8% L-arginine between days 0 and 25 of gestation reduces litter size in gilts. *J Nutr* 140: 1111–1116, 2010.
  98. Li Y, Reddy MA, Miao F, Shanmugam N, Yee JK, Hawkins D, Ren B, and Natarajan R. Role of the histone H3 lysine 4 methyltransferase, SET7/9, in the regulation of NF- $\kappa$ B-dependent inflammatory genes. *J Biol Chem* 283: 26771–26781, 2008.
  99. Ling C, Poulsen P, Simonsson S, Ronn T, Holmkvist J, Almgren P, Hagert P, Nilsson E, Mabey AG, Nilsson P, Vaag A, and Groop L. Genetic and epigenetic factors are associated with expression of respiratory chain component NDUFB6 in human skeletal muscle. *J Clin Invest* 117: 3427–3435, 2007.
  100. Ling C, Del Guerra S, Lupi R, Ronn T, Granhall C, Luthman H, Masiello P, Marchetti P, Groop L, and Del Prato S. Epigenetic regulation of *PPARGC1A* in human type 2 diabetic islets and effect on insulin secretion. *Diabetologia* 51: 615–622, 2008.
  101. Ling C and Groop L. Epigenetics: A molecular link between environmental factors and type 2 diabetes. *Diabetes* 58: 2718–2725, 2009.
  102. Lumey LH. Reproductive outcome in women prenatally exposed to undernutrition: a review of findings from the Dutch famine birth cohort. *Proc Nutr Soc* 57: 129–135, 1998.
  103. Lynn FC. Meta-regulation: MicroRNA regulation of glucose and lipid metabolism. *Trends Endocrinol Metab* 20: 452–459, 2009.
  104. Marsal K. Intrauterine growth restriction. *Curr Opin Obstet Gynecol* 14: 127–135, 2002.
  105. Mateo RD, Wu G, Bazer FW, Park JC, Shinzato I, and Kim SW. Dietary L-arginine supplementation enhances the reproductive performance of gilts. *J Nutr* 137: 652–656, 2007.
  106. Matouk CC and Marsden PA. Epigenetic regulation of vascular endothelial gene expression. *Circ Res* 102: 873–887, 2008.
  107. Martin JF, Johnston CS, Han CT, and Benyshek DC. Nutritional origins of insulin resistance: A rat model for diabetes-prone human populations. *J Nutr* 130: 741–744, 2000.
  108. Maulik N and Maulik G. (Eds). *Nutrition, Epigenetic Mechanisms, and Human Disease*. New York: CRC Press, 2011, pp. 426.
  109. McClelland M and Nelson M. The effect of site-specific DNA methylation on restriction endonucleases and DNA modification methyltransferases. *Gene* 74: 291–304, 1988.
  110. McCurdy CE, Bishop JM, Williams SM, Grayson BE, Smith MS, Friedman JE, and Grove KL. Maternal high-fat diet triggers lipotoxicity in the fetal livers of nonhuman primates. *J Clin Invest* 119: 323–335, 2009.
  111. McGee SL, van Denderen BJ, Howlett KF, Mollica J, Schertzer JD, Kemp BE, and Hargreaves M. AMP-activated protein kinase regulates GLUT4 transcription by phosphorylating histone deacetylase 5. *Diabetes* 57: 860–867, 2008.
  112. McGee SL and Hargreaves M. Histone modifications and exercise adaptations. *J Appl Physiol* 110: 258–263, 2011.
  113. McGregor RA and Choi MS. MicroRNAs in the regulation of adipogenesis and obesity. *Curr Mol Med* 11: 304–316, 2011.
  114. McKnight JR, Satterfield MC, Jobgen WS, Smith SB, Spencer TE, Meininger CJ, McNeal, CJ, and Wu G. Beneficial effects of L-arginine on reducing obesity: Potential mechanisms and important implications for human health. *Amino Acids* 39: 349–357, 2010.
  115. McKnight JR, Satterfield MC, Li XL, Gao HJ, Wang JJ, Li DF, and Wu G. Obesity in pregnancy: Problems and potential solutions. *Front Biosci* E3: 442–452, 2011.



116. Melzner I, Scott V, Dorsch K, Fischer P, Wabitsch M, Bruderlein S, Hasel C, and Moller P. Leptin gene expression in human preadipocytes is switched on by maturation-induced demethylation of distinct CpGs in its proximal promoter. *J Biol Chem* 277: 45420–45427, 2002.
117. Miao F, Gonzalo IG, Lanting L, and Natarajan R. *In vivo* chromatin remodeling events leading to inflammatory gene transcription under diabetic conditions. *J Biol Chem* 279: 18091–18097, 2004.
118. Miao F, Smith DD, Zhang L, Min A, Feng W, and Natarajan R. Lymphocytes from patients with type 1 diabetes display a distinct profile of chromatin histone H3 lysine 9 dimethylation: An epigenetic study in diabetes. *Diabetes* 57: 3189–3198, 2008.
119. Milagro FI, Campion J, Garcia-Diaz DF, Goyenechea E, Paternain L, and Martinez JA. High fat diet-induced obesity modifies the methylation pattern of leptin promoter in rats. *J Physiol Biochem* 65: 1–9, 2009.
120. Mouillet JF, Chu T, and Sadovsky Y. Expression patterns of placental microRNAs. *Birth Defects Res A* 91: 737–743, 2011.
121. Muhlhausler BS, Duffield JA, and McMillen IC. Increased maternal nutrition increases leptin expression in perirenal and subcutaneous adipose tissue in the postnatal lamb. *Endocrinology* 148: 6157–6163, 2007.
122. Nakanishi N, Nakagawa Y, Tokushige N, Aoki N, Matsuzaka T, Ishii K, Yahagi N, Kobayashi K, Yatoh S, Takahashi A, Suzuki H, Urayama O, Yamada N, and Shimano H. The upregulation of microRNA-335 is associated with lipid metabolism in liver and white adipose tissue of genetically obese mice. *Biochem Biophys Res Commun* 385: 492–496, 2009.
123. Ng SF, Lin RC, Laybutt DR, Barres R, Owens JA, and Morris MJ. Chronic high-fat diet in fathers programs  $\beta$ -cell dysfunction in female rat offspring. *Nature* 467: 963–966, 2010.
124. Nijland MJ, Mitsuya K, Li C, Ford SP, McDonald TJ, Nathanielsz PW, and Cox LA. Epigenetic modification of fetal baboon hepatic phosphoenolpyruvate carboxykinase following exposure to moderately reduced nutrient availability. *J Physiol* 588: 1349–1359, 2010.
125. Nolan CJ, Damm P, and Prentki M. Type 2 diabetes across generations: From pathophysiology to prevention and management. *Lancet* 378: 169–181, 2011.
126. Ogden CL, Yanovski SZ, Carroll MD, and Flegal KM. The epidemiology of obesity. *Gastroenterology* 132: 2087–2102, 2007.
127. Oommen AM, Griffin JB, Sarath G, and Zempleni J. Roles for nutrients in epigenetic events. *J Nutr Biochem* 16: 74–77, 2005.
128. Ovesen P, Rasmussen S, and Kesmodel U. Effect of pregnancy maternal overweight and obesity on pregnancy outcome. *Obstet Gynecol* 118: 305–312, 2011.
129. Parikh H, Nilsson E, Ling C, Poulsen P, Almgren P, Nittby H, Eriksson KF, Vaag A, and Croop LC. Molecular correlates for maximal oxygen uptake and type 1 fibers. *Am J Physiol Endocrinol Metab* 294: E1152–E1159, 2008.
130. Park JH, Stoffers DA, Nicholls RD, and Simmons RA. Development of type 2 diabetes following intrauterine growth retardation in rats is associated with progressive epigenetic silencing of Pdx1. *J Clin Invest* 118: 2316–2324, 2008.
131. Peterson CL and Lanier MA. Histones and histone modifications. *Curr Biol* 14: R546–R551, 2004.
132. Philbin VJ and Levy O. Developmental biology of the innate immune response: Implications for neonatal and infant vaccine development. *Pediatr Res* 65: 98R–105R, 2009.
133. Phillips DIW. Programming of the stress response: A fundamental mechanism underlying the long-term effects of the fetal environment. *J Intern Med* 261: 453–460, 2007.
134. Philp LK, Muhlhausler BS, Janovska A, Wittert GA, Duffield JA, and McMillen IC. Maternal overnutrition suppresses the phosphorylation of 5'-AMP-activated protein kinase in liver, but not skeletal muscle, in the fetal and neonatal sheep. *Am J Physiol Regul Integr Comp Physiol* 295: R1982–1990, 2008.
135. Pickart CM. Mechanisms underlying ubiquitination. *Annu Rev Biochem* 70: 503–533, 2001.
136. Pokholok DK, Harbison CT, Levine S, Cole M, Hannett NM, Lee TI, Bell GW, Walker K, Rolfe PA, Herbolsheimer E, Zeitlinger J, Lewitter F, Gifford DK, and Young RA. Genome-wide map of nucleosome acetylation and methylation in yeast. *Cell* 122: 517–527, 2005.
137. Poston L, Harthoorn LF, and Van Der Beek EM. Contributors to the ILSI Europe Workshop. Obesity in pregnancy: Implications for the mother and lifelong health of the child. A consensus statement. *Pediatr Res* 69: 175–180, 2011.
138. Poy MN, Eliasson L, Krutzfeldt J, Kuwajima S, Ma X, Macdonald PE, Pfeffer S, Tuschl T, Rajewsky N, Rorsman P, and Stoffel M. A pancreatic islet-specific microRNA regulates insulin secretion. *Nature* 432: 226–230, 2004.
139. Qipshidze N, Metreveli N, Lominadze D, and Tyagi SC. Folic acid improves acetylcholine-induced vasoconstriction of coronary vessels isolated from hyperhomocysteinemic mice: An implication to coronary vasospasm. *J Cell Physiol* 226: 2712–2720, 2011.
140. Raychaudhuri N, Raychaudhuri S, Thamocharan M, and Devaskar SU. Histone code modifications repress glucose transporter 4 expression in the intrauterine growth-restricted offspring. *J Biol Chem* 283: 13611–13626, 2008.
141. Rayner KJ, Sheedy FJ, Esau CC, Hussain FN, Temel RE, Parathath S, van Gils JM, Rayner AJ, Chang AN, Suarez Y, Fernandez-Hernando C, Fisher EA, and Moore KJ. Antagonism of miR-33 in mice promotes reverse cholesterol transport and regression of atherosclerosis. *J Clin Invest* 121: 2921–2931, 2011.
142. Rayner KJ, Esau CC, Hussain FN, McDaniel AL, Marshall SM, van Gils JM, Ray TD, Sheedy FJ, Goedeke L, Liu X, Khatsenko OG, Kaimal V, Lees CJ, Fernandez-Hernando C, Fisher EA, Temel RE, Moore KJ. Inhibition of miR-33a/b in nonhuman primates raises plasma HDL and lowers VLDL triglycerides. *Nature* 478: 404–407, 2011.
143. Reynolds LP, Caton JS, Redmer DA, Grazul-Bilska AT, Vonnahme KA, Borowicz PB, Luther JS, Wallace JM, Wu G, and Spencer TE. Evidence for altered placental blood flow and vascularity in compromised pregnancies. *J Physiol* 572: 51–58, 2006.
144. Ronn T, Poulsen P, Hansson O, Holmkvist J, Almgren P, Nilsson P, Tuomi T, Isomaa B, Groop L, Vaag A, and Ling C. Age influences DNA methylation and gene expression of COX7A1 in human skeletal muscle. *Diabetologia* 51: 1159–1168, 2008.
145. Rooney K and Ozanne SE. Maternal over-nutrition and offspring obesity predisposition: Targets for preventative interventions. *Int J Obes* 35: 883–890, 2011.
146. Rush D. Nutritional and maternal mortality in the developing world. *Am J Clin Nutr* 72: 212S–240S, 2000.
147. Sakabe K, Wang ZH, and Hart GW.  $\beta$ -N-acetylglucosamine (O-GlcNAc) is part of the histone code. *Proc Natl Acad Sci USA* 107: 19915–19920, 2010.

148. Samuelsson AM, Matthews PA, Argenton M, Christie MR, McConnell JM, Jansen EH, Piersma AH, Ozanne SE, Twinn DF, Remacle C, Rowlerson A, Poston L, and Taylor PD. Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: A novel murine model of developmental programming. *Hypertension* 51: 383–392, 2008.
149. Sathishkumar K, Elkins R, Yallampalli U, and Yallampalli C. Protein restriction during pregnancy induces hypertension and impairs endothelium-dependent vascular function in adult female offspring. *J Vasc Res* 46: 229–239, 2009.
150. Satterfield MC, McKnight JR, Li XL, and Wu G. Nutrition, epigenetics, and vascular function. In: *Nutrition, Epigenetic Mechanisms, and Human Disease*, edited by Maulik N and Maulik G. New York: CRC Press, 2011, pp. 125–139.
151. Satterfield MC, Bazer FW, Spencer TE, and Wu G. Sildenafil citrate treatment enhances amino acid availability in the conceptus and fetal growth in an ovine model of intrauterine growth restriction. *J Nutr* 140: 251–258, 2010.
152. Satterfield MC and Wu G. Growth and development of brown adipose tissue: Significance and nutritional regulation. *Front Biosci* 16: 1589–1608, 2011.
153. Schlambritz-Loutsevitch N, Farley D, Tejero ME, Comuzzie AG, Higgins PB, Cox L, Werner SL, Jenkins SL, Li C, Choj J, Dick EJ, Hubbard GB, Frost P, Dudley DD, Wu G, and Nathanielsz PW. Feto-placental adaptations to maternal obesity in the baboon. *Placenta* 30: 752–760, 2009.
154. Schwer B and Verdin E. Conserved metabolic regulatory functions of sirtuins. *Cell Metab* 7: 104–112, 2008.
155. Sharkey D, Gardner DS, Symonds ME, and Budge H. Maternal nutrient restriction during early fetal kidney development attenuates the renal innate inflammatory response in obese young adult offspring. *Am J Physiol Renal Physiol* 297: F1199–1207, 2009.
156. Simmons R. Epigenetics and maternal nutrition: Nature v. nurture. *Proc Nutr Soc* 70: 73–81, 2011.
157. Sinclair KD, Allegrucci C, Singh R, Gardner DS, Sebastian S, Bispham J, Thurston A, Huntley JF, Rees WD, Maloney CA, Lea RG, Craigon J, McEvoy TG, and Young LE. DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. *Proc Natl Acad Sci USA* 104: 19351–19356, 2007.
158. Schroen B and Heymans S. Small but smart—MicroRNAs in the center of inflammatory processes during cardiovascular diseases, the metabolic syndrome, and aging. *Cardiovasc Res* doi:10.1093/cvr/cvr268, 2011.
159. Smith AD, Kim YI, Refsum H. Is folic acid good for everyone? *Am J Clin Nutr* 87: 517–533, 2008.
160. Sullivan EL, Smith MS, and Grove KL. Perinatal exposure to high-fat diet programs energy balance, metabolism and behavior in adulthood. *Neuroendocrinol* 93: 1–8, 2011.
161. Sun LD, Zhao HB, Xu ZB, Liu QL, Liang W, Wang LT, Cai X, Zhang L, Hu L, Wang G, and Zha X. Phosphatidylinositol 3-kinase/protein kinase B pathway stabilizes DNA methyltransferase I protein and maintains DNA methylation. *Cell Signal* 19: 2255–2263, 2007.
162. Symonds ME, Sebert SP, Hyatt MA, and Budge H. Nutritional programming of the metabolic syndrome. *Nat Rev Endocrinol* 5: 604–610, 2009.
163. Szic KSV, Ndlovu MN, Haegeman G, and Vanden Berghe W. Nature or nurture: Let food be your epigenetic medicine in chronic inflammatory disorders. *Biochem Pharmacol* 80: 1816–1832, 2010.
164. Szyf M. Epigenetics, DNA methylation, and chromatin modifying drugs. *Annu Rev Pharmacol Toxicol* 49: 243–263, 2009.
165. Tateishi K, Okada Y, Kallin EM, and Zhang Y. Role of Jhdm2a in regulating metabolic gene expression and obesity resistance. *Nature* 458: 757–761, 2009.
166. Taylor PD, McConnell J, Khan IY, Holemans K, Lawrence KM, Asare-Anane H, Persaud SJ, Jones PM, Petrie L, Hanson MA, and Poston L. Impaired glucose homeostasis and mitochondrial abnormalities in offspring of rats fed a fat-rich diet in pregnancy. *Am J Physiol Regul Integr Comp Physiol* 288: R134–R139, 2005.
167. Temple IK and Shield JP. Transient neonatal diabetes, a disorder of imprinting. *J Med Genet* 39: 872–875, 2002.
168. Thamocharan M, Garg M, Oak S, Rogers LM, Pan A, Sangiorgi F, Lee PN, and Devaskar SU. Transgenerational inheritance of the insulin resistant phenotypes in embryo-transferred intrauterine growth-restricted adult female rat offspring. *Am J Physiol Endocrinol Metab* 292: E1270–E1279, 2007.
169. Thambirajah AA, Li A, Ishibashi T, and Ausio J. New developments in post-translational modifications and functions of histone H2A variants. *Biochem Cell Biol* 87: 7–17, 2009.
170. Timp W, Levchenko A, and Feinberg AP. A new link between epigenetic progenitor lesions in cancer and the dynamics of signal transduction. *Cell Cycle* 8: 383–390, 2009.
171. Tobi EW, Lumey LH, Talens RP, Kremer D, Putter H, Stein AD, Slagboom PE, and Heijmans BT. DNA methylation differences after exposure to prenatal famine are common and timing- and sex-specific. *Hum Mol Genet* 18: 4046–4053, 2009.
172. Todeschini AL, Teyssset L, Delmarre V, and Ronsseray S. The epigenetic trans-silencing effect in *Drosophila* involves maternally-transmitted small RNAs whose production depends on the piRNA pathway and HP1. *PLoS One* 5: e11032, 2010.
173. Tong JF, Yan X, Zhu MJ, Ford SP, Nathanielsz PW, and Du M. Maternal obesity downregulates myogenesis and beta-catenin signaling in fetal skeletal muscle. *Am J Physiol Endocrinol Metab* 296: E917–E924, 2009.
174. Turner BM. Histone acetylation as an epigenetic determinant of long-term transcriptional competence. *Cell Mol Life Sci* 54: 21–31, 1998.
175. Turunen MP, Aavik E, and Yla-Herttuala S. Epigenetics and atherosclerosis. *Biochim Biophys Acta* 1790: 886–891, 2009.
176. Unterberger A, Szyf M, Nathanielsz PW, and Cox LA. Organ and gestational age effects of maternal nutrient restriction on global methylation in fetal baboons. *J Med Primatol* 38: 219–227, 2009.
177. USDA. Dietary Guidance. 2010. DRI Tables. <http://fnic.nal.usda.gov>. Accessed on July 26, 2011.
178. Vaxillaire M and Froguel P. Monogenic diabetes in the young, pharmacogenetics and relevance to multifactorial forms of type 2 diabetes. *Endocr Rev* 29: 254–264, 2008.
179. Villeneuve LM, Reddy MA, Lanting LL, Wang M, Meng L, and Natarajan R. Epigenetic histone H3 lysine 9 methylation in metabolic memory and inflammatory phenotype of vascular smooth muscle cells in diabetes. *Proc Natl Acad Sci USA* 105: 9047–9052, 2008.
180. Vonnahme KA, Hess BW, Hansen TR, McCormick RJ, Rule DC, Moss GE, Murdoch WJ, Nijland MJ, Skinner DC, Nathanielsz PW, and Ford SP. Maternal undernutrition from early- to mid-gestation leads to growth retardation, cardiac ventricular hypertrophy, and increased liver weight in the fetal sheep. *Biol Reprod* 69: 133–140, 2003.

181. Vucetic Z, Kimmel J, Totoki K, Hollenbeck E, and Reyes TM. Maternal high-fat diet alters methylation and gene expression of dopamine and opioid-related genes. *Endocrinology* 151: 4756–4764, 2010.
182. Waddington CH. *Organizers and Genes*. Cambridge University Press. 1940.
183. Waechter DE and Baserga R. Effect of methylation on expression of microinjected genes. *Proc Natl Acad Sci USA* 79: 1106–1110, 1982.
184. Wallace DC and Fan WW. Energetics, epigenetics, mitochondrial genetics. *Mitochondrion* 10: 12–31, 2010.
185. Wallace JM, Milne JS, and Aitken RP. The effect of over-nourishing singleton-bearing adult ewes on nutrient partitioning to the gravid uterus. *Br J Nutr* 94: 533–539, 2005.
186. Wallace JM, Luther JS, Milne JS, Aitken RP, Redmer DA, Reynolds LP, and Hay WW. Nutritional modulation of adolescent pregnancy outcome. A review. *Placenta* 27: S61–S68, 2006.
187. Warner MJ and Ozanne SE. Mechanisms involved in the developmental programming of adulthood disease. *Biochem J* 427: 333–347, 2010.
188. Weake VM and Workman JL. Histone ubiquitination: Triggering gene activity. *Mol Cell* 29: 653–663, 2008.
189. Weems J and Olson AL. Class II histone deacetylases limit GLUT4 gene expression during adipocyte differentiation. *J Biol Chem* 286: 460–468, 2011.
190. WHO. United Nations Food and Agriculture Organization. <http://www.fao.org/publications/sofi/en.2010>. Accessed on July 20, 2011.
191. Williams KT and Schalinske KL. Homocysteine metabolism and its relation to health and disease. *Biofactors* 36: 19–24, 2010.
192. Williams PJ, Kurlak LO, Perkins AC, Budge H, Stephenson T, Keisler D, Symonds ME, and Gardner DS. Hypertension and impaired renal function accompany juvenile obesity: The effect of prenatal diet. *Kidney Int* 72: 279–289, 2007.
193. Wilson VL and Jones PA. DNA methylation decreases in aging but not in immortal cells. *Science* 220: 1055–1057, 1983.
194. Wilson MJ, Shivapurkar N, and Poirier LA. Hypomethylation of hepatic nuclear DNA in rats fed with a carcinogenic methyl-deficient diet. *Biochem J* 218: 987–990, 1984.
195. Winzerling JJ and Law JH. Comparative nutrition of iron and copper. *Annu Rev Nutr* 17: 501–526, 1997.
196. Wu G. Amino acids: Metabolism, functions, and nutrition. *Amino Acids* 37: 1–17, 2009.
197. Wu G. Functional amino acids in growth, reproduction and health. *Adv Nutr* 1: 31–37, 2010.
198. Wu G and Meininger CJ. Nitric oxide and vascular insulin resistance. *BioFactors* 35: 21–27, 2009.
199. Wu G, Bazer FW, Cudd TA, Meininger CJ, and Spencer TE. Maternal nutrition and fetal development. *J Nutr* 134: 2169–2172, 2004.
200. Wu G, Bazer FW, Wallace JM, and Spencer TE. Board-invited review: Intrauterine growth retardation: implications for the animal sciences. *J Anim Sci* 84: 2316–2337, 2006.
201. Wu G, Bazer FW, Datta S, Johnson GA, Li P, Satterfield MC, and Spencer TE. Proline metabolism in the conceptus: Implications for fetal growth and development. *Amino Acids* 35: 691–702, 2008.
202. Wu G, Bazer FW, Davis TA, Kim SW, Li P, Rhoads JM, Satterfield MC, Smith SB, Spencer TE, and Yin YL. Arginine metabolism and nutrition in growth, health and disease. *Amino Acids* 37: 153–168, 2009.
203. Wu G, Bazer FW, Burghardt RC, Johnson GA, Kim SW, Li XL, Satterfield MC, and Spencer TE. Impacts of amino acid nutrition on pregnancy outcome in pigs: Mechanisms and implications for swine production. *J Anim Sci* 88: E195–E204, 2010.
204. Wu G, Bazer FW, Burghardt RC, Johnson GA, Kim SW, Knabe DA, Li P, Li XL, McKnight JR, Satterfield MC, and Spencer TE. Proline and hydroxyproline metabolism: Implications for animal and human nutrition. *Amino Acids* 40: 1053–1063, 2011.
205. Xiao XM and Li LP. L-arginine treatment for asymmetric fetal growth restriction. *Int J Gynecol Obstet* 88: 15–18, 2005.
206. Yajnik CS and Deshmukh US. Maternal nutrition, intrauterine programming and consequential risks in the offspring. *Rev Endocr Metab Disord* 9: 203–211, 2008.
207. Yamamoto Y, Verma UN, Prajapati S, Kwak YT, and Gaynor RB. Histone H3 phosphorylation by IKK- $\alpha$  is critical for cytokine-induced gene expression. *Nature* 423: 655–659, 2003.
208. Yokomori N, Tawata M, and Onaya T. DNA demethylation modulates mouse leptin promoter activity during the differentiation of 3T3-L1 cells. *Diabetologia* 45: 140–148, 2002.
209. Youngson NA and Whitelaw E. Transgenerational epigenetic effects. *Annu Rev Genomics Hum Genet* 9: 233–257, 2008.
210. Zambrano E, Martinez-Samayoá PM, Bautista CJ, Deas M, Guillen L, Rodriguez-Gonzales GL, Guzmán C, Larrea F, and Nathanielsz PW. Sex differences in transgenerational alterations of growth and metabolism in progeny (F2) of female offspring (F1) of rats fed a low protein diet during pregnancy and lactation. *J Physiol* 566: 225–236, 2005.
211. Zeng XF, Wang FL, Fan X, Yang WJ, Zhou B, Li PF, Yin YL, Wu G, and Wang JJ. Dietary arginine supplementation during early pregnancy enhances embryonic survival in rats. *J Nutr* 138: 1421–1425, 2008.
212. Zhu MJ, Du M, Nijland MJ, Nathanielsz PW, Hess BW, Moss GE, and Ford SP. Down-regulation of growth signaling pathways linked to a reduced cotyledonary vascularity in placentomes of over-nourished, obese pregnant ewes. *Placenta* 30: 405–410, 2009.
213. Zimmermann MB. The influence of iron status on iodine utilization and thyroid function. *Annu Rev Nutr* 26: 367–389, 2006.

Address correspondence to:  
 Dr. Guoyao Wu  
 Room 212, Kleberg Center  
 2471 TAMU  
 Texas A&M University  
 College Station, TX 77843-2471  
 E-mail: g-wu@tamu.edu

Date of first submission to ARS Central, October 31, 2011; date of acceptance, November 1, 2011.

#### Abbreviations Used

DNMT = DNA methyltransferase  
 EAA = essential amino acids  
 H3K4 = histone 3 lysine residue 4  
 HAT = histone acetyltransferase  
 HDAC = histone deacetyltransferase  
 HMT = histone methyltransferase  
 IUGR = intrauterine growth restriction  
 LCFA = long-chain fatty acids  
 miR = microRNA  
 NEAA = nonessential amino acids