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OPEN Odorant ligands for the CO₂ receptor in two Anopheles vectors of malaria

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Exhaled CO₂ is an important host-seeking cue for Anopheles mosquitoes, which is detected by a highly conserved heteromeric receptor consisting of three 7-transmembrane proteins Gr22, Gr23, and Gr24. The CO₂ receptor neuron has been shown to also respond sensitively to a variety of odorants in Aedes aegypti. The detection of CO₂ is important for upwind navigation and for enhancing the attraction to body heat as well as to skin odorants. The orthologs of the CO₂ receptor proteins are present in malaria-transmitting mosquitoes like Anopheles coluzzii and Anopheles sinensis. Activators and inhibitors of the CO₂-neuron were tested on the maxillary palps in these two species by single-sensillum electrophysiology. The electrophysiological testing of three prolonged-activator odorants identified originally in Aedes aegypti also showed varying ability to reduce the CO₂-ellicited increase in spikes. These findings provide a foundation for comparing the functional conservation with the evolutionary conservation of an important class of odorant receptor. The identification of a suite of natural odorants that can be used to modify the CO₂-detection pathway may also contribute to odor-blends that can alter the behavior of these disease transmitting mosquitoes.

Vector-borne diseases cause significant morbidity and mortality throughout the globe. Mosquitoes transmit a variety of pathogenic microorganisms that are responsible for malaria, filariasis, dengue fever, and encephalitis. Anopheline mosquitoes transmit malarial parasites, which infect >216 million people in Africa and Asia, resulting in ~445,000 deaths annually¹. Indoor residual insecticide spraying (IRS) and long-lasting insecticide treated bednets (LLINs) provide protection against endophilic/endophagic mosquitoes, but lack of protection from bites for activities outside a bednet or outdoors from exophilic and diurnal mosquitoes require the development of new tools.

Mosquitoes are attracted long-range to carbon dioxide (CO_2) exhaled from human breath and short-range to skin odor and body temperature^{2,3}. The heteromeric CO_2 -receptor is expressed in the capitate peg sensillum A neuron (cpA) on the maxillary palps and consists of three 7-transmembrane proteins of the Gustatory Receptor family (Gr22, Gr23, and Gr24)^{4,5}. This heteromeric receptor also detects skin odorants in Aedes aegypti and Anopheles coluzzii, participating in attraction when tested in A. aegypti⁶. Several additional classes of odorants have been identified using chemical informatics and electrophysiology that either activate or inhibit the CO₂ receptor neuron in A. aegypti⁶. Given the sequence-level conservation of the orthologous Gr receptors in anopheline species, it would be interesting to study whether or not the physiological responses to known ligands are also conserved. Activators of the CO2 receptor that we identified previously have been shown to lure mosquitoes into a trap when tested in a greenhouse for Ae. aegypti and in semi-field conditions for An. coluzzii^{6,7}. Identification of additional activators of the CO2 receptor in anophelines could therefore assist in development of convenient synthetic odorant lures that can mimic CO₂.

Detection of skin odorants also occurs through other receptors belonging to the Or and Ir families⁸⁻¹⁰, as a Gr3 receptor mutant female Ae. aegypti can still seek out a human inside a greenhouse¹¹. While these alternative pathways exist in mosquitoes to sense mammalian hosts, there is still a reduction in host-seeking behavior of Ae. *aegypti* by interfering with the detection of CO_2 using inhibitory odorants⁶ or genetically when testing mice in a

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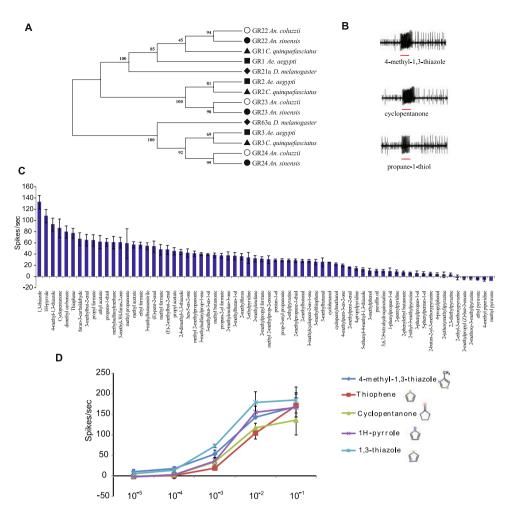


Figure 1. (A) Maximum likelihood tree depicts the phylogenetic relationship among the CO₂ receptors GR22, GR23, and GR24 orthologs in mosquitoes. Mosquito species as labeled as follow: Open circles, *An. coluzzii*; filled circles, *An. sinensis*; filled squares, *Ae. aegypti*, filled triangles, *Cx. quinquefasciatus*; and filled diamond, *D. melanogaster*. Bootstrap support for the branches is also shown. (B) *An. coluzzii* cpA neuron representative traces depicting activation of the cpA neuron upon exposure to 0.5 second pulse (red horizontal bar) of the described odor and (C) mean responses to 68 chemical volatiles (headspace above 1% solution on filter paper). n = 4. error = s.e.m. (D) Dose-responses of the activators of the *An. coluzzii* cpA neuron by five strong activators and their chemical structures. N = 3. error = s.e.m.

large cage arena¹². The identification of odorant ligands of the *Anopheles* CO_2 receptor neuron (cpA) can contribute to the design of attraction masking agents, which could reduce anopheline biting rates.

Using single sensillum recordings, we screened the cpA neuron of two anopheline mosquito species, anthropophilic *An. coluzzii* and facultative anthropophilic *Anopheles sinensis*, that transmit malaria with a large set of odorants that were initially identified by chemical-informatics as putative CO₂ receptor ligands, and subsequently tested in the *Ae. aegypti* mosquitoes⁶. We identify several odorants that show conserved effects as activators, inhibitors, and ultra-prolonged activator of the cpA neuron. Some of these odorants have potential in reducing anopheline-biting rates.

Results

Sequence conservation of CO₂ receptor proteins. There is a close relationship among CO₂ receptors in the Anophelinae (*An. coluzzii* and *An. sinensis*) and Culicinae (*Ae. aegypti* and *Culex quinquefasciatus*) mosquitoes, and orthologs cluster together in distinct branches (Fig. 1A). The percent amino acid identities between the sequences of these mosquito orthologs ranged as follows: GR22 (81–90%), GR23 (83–97%), and GR24 (74–92%). Similarities ranged for GR22 from 88% to 95%, for GR23 between 94% and 98%, and for GR24 from 85% to 91%.

Conservation of response to agonists in *An. coluzzii*. In order to test conservation of the CO_2 -receptor neuron responses to different odorants, we tested a structurally diverse set of 67 ligands of the CO_2 -receptor previously identified in *Ae. aegypti* and *Cx. quinquefasciatus*⁶. The *An. coluzzii* CO_2 neuron responded to several of these odorants with different chemical structures (Fig. 1B). Out of the 67 odors tested 35 (52%) evoked responses \geq 30 spikes/sec whereas 14 odors (21%) showed responses lower than the solvent (paraffin oil; Fig. 1C). The odorants that evoked the strongest activation from the *An. coluzzii* cpA neuron were further evaluated in

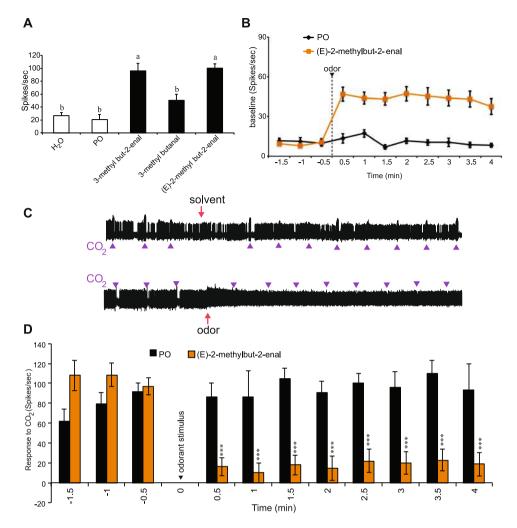


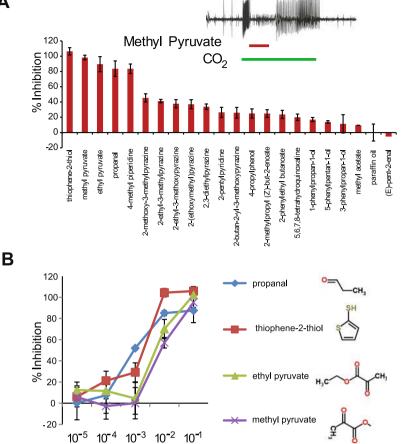
Figure 2. (A) Mean response of *An. coluzzii* cpA neuron in females to the ultraprolonged activators (headspace above 10% solution on filter paper) and the solvent (n = 5-6). (B) cpA baseline activity exposure to odorant. (C) Representative traces from the cp sensillum to 1 s pulses of 0.15% CO₂ prior to and following a 3-s exposure to either solvent (PO-paraffin oil) or (E)-2-methylbut-2-enal (headspace above 10% solution on filter paper). (D) Mean responses of the cpA neuron to 1 s pulses of 0.15% CO₂, calculated by subtracting 1-s of baseline activity prior to each stimulus after exposure to paraffin oil (gray) or (E)-2-methylbut-2-enal (headspace above 10% solution on filter paper) (orange). n = 5-6 individuals; t test, ***p < 0.001. Error = s.e.m.

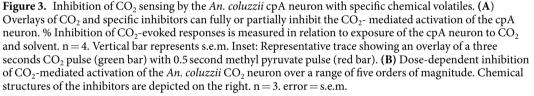
dose-response assays across a range of five orders of magnitude. All odorants showed a dose-response and four out of the five odors still evoked responses \geq 30 spikes/sec with headspace from a 10⁻³ dilution (Fig. 1D).

Conservation of ultra-prolonged activators in *An. coluzzii*. Another class of ligands that have been identified in *A. aegypti* are ultraprolonged activators of the CO_2 neuron^{6,13}. In order to test these longer-term responses, recordings were performed as before with three known odorants (Fig. 2A). This analysis revealed that two of three odorants are conserved in their ability to evoke ultraprolonged activation in *An. coluzzii*. After a 3-s exposure to a (E)-2-methylbut-2-enal stimulus, the cpA neuron continues firing at ~50 spikes/sec for at least 5 min (Fig. 2B,C). Consistent with previous results responses to repeated CO_2 stimuli during this 5-min period after the pre-exposure to (E)-2-methylbut-2-enal were significantly reduced (Fig. 2D).

Conservation of inhibitors in *An. coluzzii*. Among the odorants that induced responses lower than the solvent, six odors actually inhibited the baseline activity of the *An. coluzzii* cpA neuron when tested with the headspace from 10^{-2} concentration odor cartridges (Fig. 1A). In order to test whether some of these odorants could constitute potential antagonists of cpA, we tested the ability of 21 odorants at a higher concentration (head-space from 10^{-1} concentration odor cartridges) to inhibit CO₂-mediated (0.15% concentration) activation of the *An. coluzzii* cpA neuron in overlay assays. Of the 21 odorants tested, 11 were capable of reducing CO₂-mediated cpA activation between 20% and 45%, and 5 odorants inhibited CO₂ activation by >80% (Fig. 3A). Four top inhibitors were selected for dose-response assays. Four odorants were able to inhibit cpA activation by at least 50% when tested at 10^{-2} concentration and propanal evoked similar levels of inhibition when tested at 10^{-3} (Fig. 3B).







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Conservation of inhibitors in *An. sinensis.* In order to test whether inhibitory odorants of the *An. coluzzii* cpA neuron could be of utility in *An. sinensis*, four of the strong inhibitors (propanal, ethyl pyruvate, thiophene-2-thiol, and 4-methyl piperidine) were tested at two concentrations using electrophysiology. All the tested odorants showed some degree of inhibition, but to varying extent. The strongest inhibition was observed for thio-2-thiol and ethyl pyruvate at the higher concentrations (1%) (Fig. 4A). Among the three tested amines, amyl amine (AA) exhibited the highest inhibitory activity followed by butyl amine (BA). Spermidine (SP), on the other hand, acted as a weaker inhibitor in the presence of CO_2 (Fig. 4B). Taken together these results indicate that the *An. sinensis* CO_2 receptors respond similarly to *Ae. aegypti* and *An. coluzzi* when it comes to CO_2 response inhibition.

Discussion

Unlike the complex blend of human skin odor, the CO_2 in exhaled breath provides a simpler cue to study. Most mosquito species are strongly attracted to CO_2 exhaled from human breath². Carbon-dioxide is detected by the cpA neuron upon binding to a receptor, comprised of three members of the gustatory receptor gene family and named GR22, GR23, and GR24 in *An. coluzzii*^{4,5}. The detection of CO_2 plays several important roles in host-seeking behaviors like long-range navigation towards a live animal^{2,14}. The identification of volatile ligands of the CO_2 receptor neuron can contribute to the design of masking agents using inhibitors and trapping-lures using activators, both of which can reduce human contact and prevent disease transmission by mosquitoes⁶. We previously developed a computational approach¹⁵ which we applied to identify novel odorant ligands of the cpA neuron in *A. aegypti*⁶. Here we tested the conservation of the ligand responses in two species of anopheline mosquitoes, *An. coluzzii* and *An. sinensis*, that transmit malaria in Africa and Asia respectively. *An. sinensis* is one of the vectors of malaria in Asia and has Gr receptor orthologs that are closely related to ones in *An. coluzzi*. A high degree of functional conservation was observed amongst the ligands and we identified all three classes of ligands: activators, prolonged activators, and inhibitors.

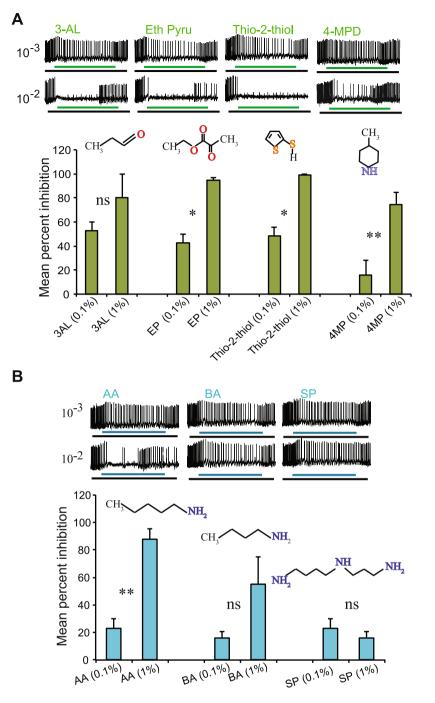


Figure 4. Inhibition of *An. sinensis* CO_2 receptor neuron (cpA). (A) Representative traces, mean percent inhibition, and chemical structure of cpA background for propanal (3-AL), ethyl pyruvate (EP), thiophene-2-thiol (Thio-2-thiol) and 4- methyl piperidine (4-MP), and (B) amyl amine (AA), butyl amine (BA) and spermidine (SP). n = 6 sensilla. Green and blue bars indicate 0.5 s odor stimulus; black bars indicate the duration of the CO_2 stimuli (1 s).

In general, the responses to activators were weaker in *An. coluzzii* than in *Ae. aegypti*⁶, and only two odorants ((E)-pent-2-enal and methyl acetate) evoked stronger responses in *An. coluzzii* than in *Ae. aegypti*⁶. Conversely, inhibition of CO₂-mediated cpA activation is stronger in *An. coluzzii*, as odorants unable to inhibit the cpA neuron in *Ae. aegypti*⁶ were capable of inhibiting the *An. coluzzii* and *An. sinensis* CO₂ neuron counterparts. Amongst the inhibitors we have identified both high and low volatility compounds that act in both *An. coluzzii* and *An. sinensis*.

The detection of CO_2 by the cpA neuron also activates attraction to other cues like skin odorants, visual cues, and importantly to body-warmth of 37 °C which is one of the strongest attraction cues at close-quarters^{12,16,17}. The maxillary palp neuron, cpA also plays an important role in detection of human skin odorants⁶, and inhibitors such ethyl pyruvate and spermidine show reduction in attraction to skin odorants in *Aedes*^{6,18}. Odorants like these

could be useful in reducing host-seeking behavior and transmission of malaria, especially when used alongside others that block receptors detecting skin odorants.

Another approach to modulate behavior is using strong prolonged activators of cpA neurons that make the neuron unresponsive to CO_2 as has been shown with 2,3-butanedione and blends on *An. coluzzii* and *Ae. aegypti*¹³. However, at the higher concentration needed for this effect the unpleasant smell of this odorant and health concerns precluded integration into practical solutions. We were able to demonstrate that *An. coluzzii* showed an ultra-prolonged activation to (E)-2-methylbut-2-enal, which resulted in masking the detection of CO_2 significantly for several minutes after by the maxillary palp cpA neurons suggesting that this odor could disrupt detection of CO_2 and navigation toward its source as has been shown in *Ae. aegypti*¹³ and could be utilized for potential practical applications in preventing mosquito bites and spreading of mosquito-borne diseases¹⁹.

Some of the CO_2 receptor neuron inhibitors have organoleptic and physicochemical properties that are conducive to development into spatial and short-range masking agents for anopheline mosquitoes. However, ultimately for a masking strategy to work, additional odorants will be needed in a blend to block other human skin receptors that are members of the *Odorant receptor (Or)* and *Ionotropic receptor (Ir)* gene families.

Experimental Procedures

Mosquitoes. The M form of *Anopheles gambiae* (Herein *An. coluzzii*, Ngousso strain, Cameroon) were maintained in a 12:12 (Light:Dark) photocycle at 27 °C and 70% RH. The *An. sinensis* mosquitoes were received from MR4 center and then colony was maintained in insectary at the same conditions as *An. coluzzii*. Adult females were fed on bovine blood through a heated membrane feeding system (Hemostat Laboratories, California, USA).

Electrophysiology. Single-sensillum recording was carried out with 4–12 days old female anopheline mosquitoes as described elsewhere^{6,13,20}. All recording replicates were performed in different specimens. All odorants were obtained from Sigma at >98% purity and were diluted to (as indicated) in paraffin oil or water. A filter paper with 50 μ l of the solution was inserted into a Pasteur pipette cartridge and the headspace was injected into a humidified airstream to further dilute it 3-fold as done previously⁶.

For the ultra-prolonged activators (E)-2-methylbut-2-enal, 3-methylbut-2-enal and 3-methylbutanal were dissolved at 10^{-1} in paraffin oil or water, from which 50 µl of the solution was added on a filter paper inside a Pasteur pipette and the headspace was used for odor delivery as indicated in the section above. The odor delivery system was modified as shown in⁶; solvent responses during the same recording session were subtracted. A controlled 3-s stimulus of solvent/stimulus was delivered from a Pasteur pipette into the carrier airstream. Subsequent 1-s of CO₂ (0.15%) stimuli was delivered using a MNJ-D microinjector (Tritech Research). Activity was calculated by subtracting baseline activity 1-s prior to each stimulus. Spike counting was done with Clampfit 10.3.

Phylogenetic analysis. The amino acid sequences of the CO_2 receptor orthologs of *Drosophila melanogaster*, *An. coluzzii, An. sinensis, Ae. aegypti*, and *Cx. quinquefasciatus* (signal peptide removed) were aligned with the ClustalW software, and the phylogenetic tree was constructed with the MEGA6²¹ software, using the Maximum Likelihood method and LG + G matrix-based model²². Reliability of the branches was inferred by 1,000 bootstrap replicates²³.

Sequence access numbers. The GenBank access numbers for the CO₂ receptor are as followed: *An. coluzzii* GR22 (XP_319142), GR23 (XP_312786), and GR24 (ABK97614); *An. sinensis* GR22 (KFB40998), GR23 (KFB38231), and GR24 (KFB40736); *Ae. aegypti* GR1 (XP_001655150), GR2 (XP_001654839), and GR3 (XP_001660602); *Cx. quinquefasciatus* GR1 (XP_001848097), GR2 (XP_001848828), and GR3 (XP_001848689); and *Drosophila melanogaster* GR21a (ABK97615) and GR63a (ABK97613).

References

- 1. WHO. World Malaria Report (2017).
- Carde, R. T. G. G. In *Ecology of Vector-Borne Diseases* (eds Knols, W. & Takken, B. G. J.) 115–141 (Wageningen Academic Publishers, 2010).
- 3. Webster, B., Lacey, E. S. & Carde, R. T. Waiting with bated breath: opportunistic orientation to human odor in the malaria mosquito, Anopheles gambiae, is modulated by minute changes in carbon dioxide concentration. *J Chem Ecol* **41**, 59–66, https://doi. org/10.1007/s10886-014-0542-x (2015).
- 4. Lu, T. et al. Odor coding in the maxillary palp of the malaria vector mosquito Anopheles gambiae. Curr Biol 17, 1533–1544 (2007).
- 5. Robertson, H. M. & Kent, L. B. Evolution of the gene lineage encoding the carbon dioxide receptor in insects. *J Insect Sci* 9, 19, https://doi.org/10.1673/031.009.1901 (2009).
- Tauxe, G. M., MacWilliam, D., Boyle, S. M., Guda, T. & Ray, A. Targeting a dual detector of skin and CO2 to modify mosquito host seeking. *Cell* 155, 1365–1379 (2013).
- Mburu, M. M. et al. 2-Butanone as a carbon dioxide mimic in attractant blends for the Afrotropical malaria mosquitoes Anopheles gambiae and Anopheles funestus. Malar J 16, 351, https://doi.org/10.1186/s12936-017-1998-2 (2017).
- Carey, A. F., Wang, G., Su, C. Y., Zwiebel, L. J. & Carlson, J. R. Odorant reception in the malaria mosquito Anopheles gambiae. *Nature* 464, 66–71, https://doi.org/10.1038/nature08834 (2010).
- 9. Pitts, R. J., Derryberry, S. L., Zhang, Z. & Zwiebel, L. J. Variant Ionotropic Receptors in the Malaria Vector Mosquito Anopheles gambiae Tuned to Amines and Carboxylic Acids. *Sci Rep* 7, 40297 (2017).
- Wang, G., Carey, A. F., Carlson, J. R. & Zwiebel, L. J. Molecular basis of odor coding in the malaria vector mosquito Anopheles gambiae. *Proc Natl Acad Sci USA* 107, 4418–4423 (2010).
- 11. DeGennaro, M. *et al.* orco mutant mosquitoes lose strong preference for humans and are not repelled by volatile DEET. *Nature* **498**, 487–491 (2013).
- McMeniman, C. J., Corfas, R. A., Matthews, B. J., Ritchie, S. A. & Vosshall, L. B. Multimodal integration of carbon dioxide and other sensory cues drives mosquito attraction to humans. *Cell* 156, 1060–1071, https://doi.org/10.1016/j.cell.2013.12.044 (2014).
- 13. Turner, S. L. et al. Ultra-prolonged activation of CO2-sensing neurons disorients mosquitoes. Nature 474, 87-91 (2011).
- 14. Ray, A., van Naters, W. G. & Carlson, J. R. Molecular determinants of odorant receptor function in insects. *J Biosci* **39**, 555–563 (2014).

- 15. Boyle, S. M., McInally, S. & Ray, A. Expanding the olfactory code by in silico decoding of odor-receptor chemical space. *Elife* **2**, e01120 (2013).
- 16. Dekker, T., Geier, M. & Carde, R. T. Carbon dioxide instantly sensitizes female yellow fever mosquitoes to human skin odours. *J Exp Biol* 208, 2963–2972 (2005).
- 17. van Breugel, F., Riffell, J., Fairhall, A. & Dickinson, M. H. Mosquitoes Use Vision to Associate Odor Plumes with Thermal Targets. *Curr Biol* 25, 2123–2129 (2015).
- MacWilliam, D., Kowalewski, J., Kumar, A., Pontrello, C. & Ray, A. Signaling Mode of the Broad-Spectrum Conserved CO2 Receptor Is One of the Important Determinants of Odor Valence in Drosophila. *Neuron* 97, 1153–1167 e1154 (2018).
- 19. Ray, A. Reception of odors and repellents in mosquitoes. Curr Opin Neurobiol 34, 158-164 (2015).
- Turner, S. L. & Ray, A. Modification of CO2 avoidance behaviour in Drosophila by inhibitory odorants. *Nature* 461, 277–281 (2009).
 Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* 30, 2725–2729 (2013).
- 22. Le, S. Q. & Gascuel, O. An improved general amino acid replacement matrix. *Mol Biol Evol* 25, 1307–1320 (2008).
- 23. Felsenstein, J. Confidence Limits on Phylogenies: An Approach Using the Bootstrap. Evolution 39, 783-791 (1985).

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Author Contributions

I.V.C.-A. conducted experiments in Figures 1 and 3 and wrote the first draft of the manuscript; K.S. conducted experiments in Figures 2 and 4 and helped edit the manuscript; L.C. and G.Y. secured funding and helped edit the manuscript; A.R. conceived the project, managed the project and wrote the final version of the manuscript.

Additional Information

Competing Interests: A. Ray is founder and equity holder of Sensorygen Llc. A. Ray and I.V. Coutinho-Abreu are listed as inventors in patents submitted by UC Riverside.

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