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# Open chromatin defined by DNaseI and FAIRE identifies regulatory elements that shape cell-type identity

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The human body contains thousands of unique cell types, each with specialized functions. Cell identity is governed in large part by gene transcription programs, which are determined by regulatory elements encoded in DNA. To identify regulatory elements active in seven cell lines representative of diverse human cell types, we used DNase-seq and FAIRE-seq (Formaldehyde Assisted Isolation of Regulatory Elements) to map “open chromatin.” Over 870,000 DNaseI or FAIRE sites, which correspond tightly to nucleosome-depleted regions, were identified across the seven cell lines, covering nearly 9% of the genome. The combination of DNaseI and FAIRE is more effective than either assay alone in identifying likely regulatory elements, as judged by coincidence with transcription factor binding locations determined in the same cells. Open chromatin common to all seven cell types tended to be at or near transcription start sites and to be coincident with CTCF binding sites, while open chromatin sites found in only one cell type were typically located away from transcription start sites and contained DNA motifs recognized by regulators of cell-type identity. We show that open chromatin regions bound by CTCF are potent insulators. We identified clusters of open regulatory elements (COREs) that were physically near each other and whose appearance was coordinated among one or more cell types. Gene expression and RNA Pol II binding data support the hypothesis that COREs control gene activity required for the maintenance of cell-type identity. This publicly available atlas of regulatory elements may prove valuable in identifying noncoding DNA sequence variants that are causally linked to human disease.

[Supplemental material is available for this article.]

A single genome gives rise to a multitude of cell types, each with its own specialized pattern of gene expression. These programs are partly governed by DNA-encoded regulatory elements. Unlike protein coding genes, DNA regulatory elements are not easy to identify in linear DNA sequence. While nearly 70% of bases in protein-

coding DNA are evolutionary constrained, only half of all the regulatory elements identified in the ENCODE pilot project harbored constrained bases at all, and among these, only 10% of the bases were constrained (The ENCODE Project Consortium 2007). As part of the ENCODE effort (The ENCODE Project Consortium 2007, 2011), we have continued our development of DNase-seq (Crawford et al. 2006b; Boyle et al. 2008a; Song and Crawford 2010) and FAIRE-seq (Formaldehyde Assisted Isolation of Regulatory Elements)(Giresi et al. 2007; Giresi and Lieb 2009) to identify regulatory sites across the genome. DNase-seq employs the DNaseI enzyme to preferentially digest nucleosome-depleted sites, also known as DNaseI hypersensitive (HS) sites (Wu et al. 1979). FAIRE-seq enriches nucleosome-depleted DNA using formaldehyde fixation and phenol-chloroform extraction.

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Here, we call the regions identified by DNaseI or FAIRE “open chromatin.” These open chromatin regions often correspond to nucleosome-depleted regions (NDRs) (Hogan et al. 2006; Giresi et al. 2007; Kim et al. 2007), which are often associated with regulatory factor binding. Several studies have shown that open chromatin is associated with all known classes of active DNA regulatory elements, including promoters, enhancers, silencers, insulators, and locus control regions (Gross and Garrard 1988; Cockerill 2011). We used DNase-seq and FAIRE-seq (Giresi et al. 2007; Xi et al. 2007; Boyle et al. 2008a; Gaulton et al. 2010; Stitzel et al. 2010) to generate genome-wide open chromatin maps spanning seven diverse human cell types, thereby greatly expanding the number of human regulatory elements with experimental support.

## Results

### A coordinated mapping pipeline for data generation, processing, and quality control

DNase-seq and FAIRE-seq were performed on seven cell lines in duplicate or triplicate (Table 1) using material from cells grown in a single batch at the same location (see Methods). DNA libraries were sequenced on an Illumina sequencer, and the resulting data were collected and processed using a standard pipeline (see Methods). ChIP-seq data were generated using the same fixed cells used for FAIRE-seq and analyzed using the same pipeline. In this way, experimental and data processing differences among the assays were minimized (Fig. 1A). Comparisons of multiple independent growths and with results from tiled microarrays on the same material support the quality of these data (Supplemental Methods; Supplemental Table S1).

A continuous range of signal intensities from DNase-seq and FAIRE-seq was observed across the genome (Fig. 1B), and from this signal, we identified discrete peaks (see Methods). For each cell type, the number of DNase-seq or FAIRE-seq peaks ranged from ~100,000 to 225,000, covering between 0.65% and 2.99% of the genome (Table 1). In total, over 870,000 DNaseI and FAIRE sites were identified from the seven cell lines, covering nearly 9% of the genome. While more FAIRE peaks than DNaseI peaks were identified, FAIRE peaks tended to span fewer bases (Table 1).

### DNase-seq and FAIRE-seq identify an overlapping set of open chromatin sites, but each also identifies unique chromatin features

DNaseI and FAIRE assays both identify sites in the genome that tend to be nucleosome-free or nucleosome-depleted (Hogan et al.

2006; Giresi et al. 2007; Cockerill 2011). To quantitatively evaluate the overlap between these assays, we compared rank-ordered peaks from each assay, with rank based on the maximum signal intensity within each peak. We calculated the number of overlapping DNaseI and FAIRE sites for four different peak intensity cutoffs (Fig. 2A; Supplemental Fig. S1). In each cell type, ~30%–40% of the top 100,000 (100K) DNaseI and FAIRE peaks overlapped. The amount of overlap did not change substantially when comparing the top 10,000 (10K) peaks from each assay (20%–40% overlap). About 80% of the top 10K peaks from either assay are found within the top 100K of the alternate assay (Fig. 2A; Supplemental Fig. S1).

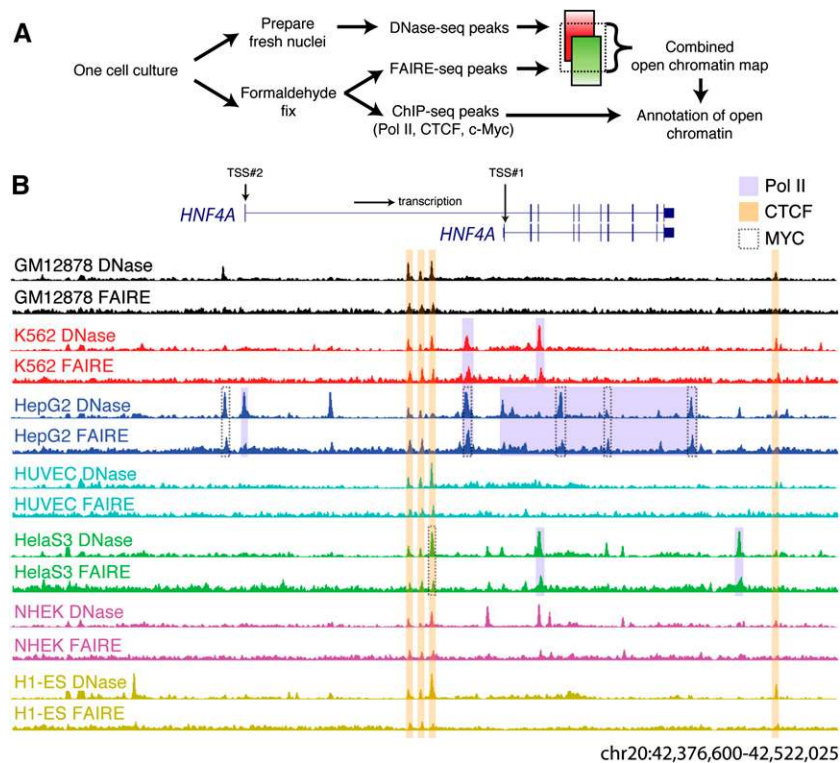
We consider sites detected by both DNase-seq and FAIRE-seq to be “cross-validated” and to represent high-confidence open chromatin sites. Across the seven cell lines, there are more than 180,000 high-confidence sites covering nearly 4.5% of the genome (Table 1). Though we found a significant enrichment of these high-confidence sites within 2 kb of an annotated transcription start site (TSS) ( $P$ -value  $< 2 \times 10^{-5}$ ), most (80%) were far from gene starts (see Methods) (Fig. 2B).

In a given cell type, many sites were detected by only one of the two assays. Multiple lines of evidence suggest that these DNase-only and FAIRE-only sites are biologically relevant and that the assays differentially detect real chromatin features. First, ~50% of DNase-only and 40% of FAIRE-only sites were detected in multiple cell types (Supplemental Fig. S2) and across multiple independent growths (Supplemental Table S1), which would be unlikely if the peaks were spurious. Second, the differences between DNase-seq and FAIRE-seq arise from distinct genomic regions, which may be interrogated differentially by each method. DNaseI-only sites were enriched within 2 kb of a TSS ( $P$ -value  $< 4 \times 10^{-6}$ ) and within 5' exons and introns ( $P$ -value  $< 3 \times 10^{-6}$ ) compared to FAIRE-only sites, while FAIRE-only sites were found preferentially in internal introns and exons and nonpromoter intergenic regions ( $P$ -value  $< 3 \times 10^{-6}$  and  $2 \times 10^{-5}$ , respectively) relative to DNase-only sites (Fig. 2B; Supplemental Table S2). Third, we found that both DNase-only and FAIRE-only sites were enriched for H3K4 monomethylation (H3K4me1), a mark associated with enhancers (Heintzman et al. 2009; Ernst et al. 2011), along with H3K4 trimethylation (H3K4me3) and H3K9 acetylation (H3K9ac); both marks of a TSS (Supplemental Fig. S3; Heintzman et al. 2009; Ernst et al. 2011). DNase-only sites were more strongly associated with H3K4me3 and H3K9ac, and FAIRE-only sites with H3K4me1. Lastly, DNase-only and FAIRE-only sites corresponded differentially to specific transcription factor binding sites, as explained in the next section.

**Table 1. Cell line descriptions**

Cell line	Description	DNaseI sites	FAIRE sites	Union	Open chromatin	Validated sites
GM12878	Lymphoblast	103,075 (1.528%)	146,147 (0.728%)	192,891 (1.865%)	94,517 (1.566%)	47,891 (0.984%)
K562	Chromic myeloid leukemia	139,121 (2.034%)	185,705 (1.261%)	260,340 (2.746)	118,915 (2.170%)	55,177 (1.274%)
HepG2	Hepatocellular carcinoma	125,631 (1.950%)	122,188 (0.871%)	189,957 (2.349%)	104,335 (1.962%)	51,538 (1.135%)
HeLa-S3	Cervical carcinoma	142,403 (2.174%)	131,935 (0.694%)	220,999 (2.481%)	124,093 (2.096%)	57,312 (1.347%)
HUVEC	Human umbilical vein endothelial cells	133,091 (2.259%)	225,564 (1.723%)	271,789 (3.096)	125,427 (2.546%)	68,245 (1.815%)
NHEK	Keratinocyte, normal epidermal cells	141,190 (1.964%)	204,280 (1.443%)	262,941 (2.749%)	137,218 (2.269%)	72,676 (1.433%)
H1-ES	Human embryonic stem cells	138,025 (3.224%)	126,439 (0.695%)	222,265 (3.660%)	114,915 (3.147%)	37,433 (1.032)
	All cell lines	360,950 (6.126%)	767,795 (5.065%)	872,394 (8.844%)	356,625(6.97%)	184,569 (4.400%)

Number of sites found and fraction of genome covered by each assay. Open chromatin sites are a high-confidence ( $P < 0.05$ ) subset of the union of DNaseI and FAIRE sites (see Methods). Validated sites are the strict intersection of DNaseI and FAIRE sites.



**Figure 1.** Identification of open chromatin in seven human cell lines. (A) A schematic representation of the experiment and analysis design. (B) DNaseI ( $y$ -axis fixed at Parzen signal value 0.15) and FAIRE ( $y$ -axis fixed at 0.04) data from seven cell lines surrounding the *HNF4A* locus (145 kb; UCSC Genome Browser) shows both ubiquitous and cell-type selective open sites that are especially prevalent in HepG2 cells. Pol II, CTCF, and MYC ChIP-seq peaks that overlap open chromatin are highlighted.

### Together, DNase-seq and FAIRE-seq identify most of the sites bound by regulatory factors

DNase-seq and FAIRE-seq data were compared to ChIP-seq data generated from the same cell lines using antibodies to CTCF, MYC, and Pol II (see Methods) (Supplemental Fig. S4). Over 96% of the strongest CTCF and MYC ChIP sites were identified by one or both assays (Fig. 2C,D). About 30% of CTCF and 15% of MYC sites were captured by DNase-only or FAIRE-only sites. At any given ChIP-seq peak cut-off, ChIP-seq signal intensity was the strongest for peaks detected by both DNaseI and FAIRE, was weaker in sites detected by only one assay, and the weakest for sites that overlapped neither assay (Supplemental Fig. S5).

We examined the correspondence of published ChIP-seq data in matching cell types (Fujiwara et al. 2009; Motallebipour et al. 2009; Fritze et al. 2010; Kouwenhoven et al. 2010; Raha et al. 2010) with our open chromatin data. DNase-seq and FAIRE-seq captured >80% of sites (>90% of the strongest sites) for TP63 in NHEK, FOXA1, and FOXA3 in HepG2, and GATA1 in K562 (Supplemental Fig. S6), and ~70% of the ZNF263 sites in K562. We note that FOXA1, FOXA3, and GATA1 were better identified by FAIRE-seq, while ZNF263 was found more often by DNase-seq.

We next evaluated our Pol II ChIP-seq data in conjunction with RNA expression data generated from the same cells. For each gene with RNA data in each cell line, we determined whether there was a significant signal for Pol II binding and/or open chromatin in the region 1000 bases upstream of and 500 bases downstream from an annotated transcription start site. We found that 81% of all TSSs harbored accessible chromatin, consistent with previous estimates

that 70%–80% of all genes are either active or poised (Guenther et al. 2007). We divided genes into highly expressed (46% of genes,  $\log_2$  RNA > 7; Methods), moderately expressed (29%,  $\log_2$  RNA between 5 and 7), and lowly or not expressed (25%,  $\log_2$  RNA < 5). We found that nearly all highly expressed genes had Pol II binding and open chromatin at their TSS (Fig. 2E). About 60% of the moderately expressed genes showed Pol II and open chromatin signals, while an additional 30% showed just open chromatin signal. About half of the lowly or nonexpressed genes showed evidence of Pol II or open chromatin, while the remaining half had no evidence of either signal. In all, open chromatin identifies the TSS of nearly all of expressed genes and indicates that a large fraction of the remaining genes may be poised for transcription.

### A combined open chromatin atlas reveals chromatin similarities between functionally related cell types

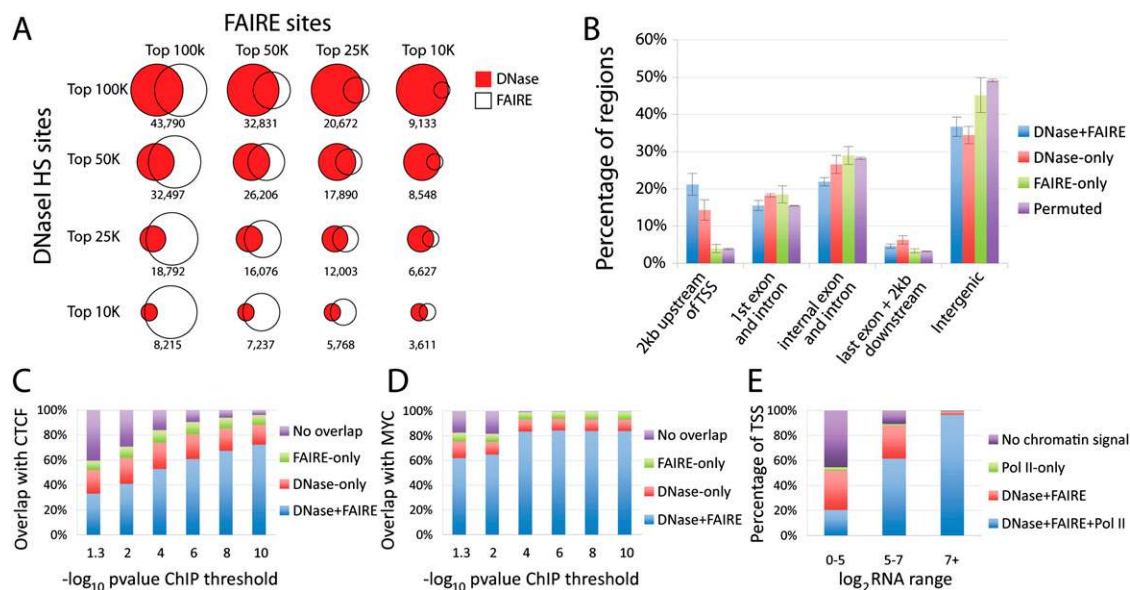
To take advantage of the strengths of each assay, we created a combined annotation for each of the seven cell lines by integrating data from DNaseI and FAIRE (see Methods). Our open chromatin atlas contains sites strongly identified by both assays, high confidence peaks present in only one assay, and lower confidence peaks supported by both assays (Table 1). The number of combined significant open chromatin sites ranged from 100,000 to 125,000 ( $P < 0.05$ ; Methods) for each cell line. Between any two cell types, ~30%–40% of open chromatin sites are shared (Supplemental Table S3).

Using open chromatin sites, we performed hierarchical clustering of the cell lines (see Methods) (Supplemental Fig. S7A). The clustering appears to reflect functional and lineage similarities in cell types and almost perfectly matches cell-line clustering based on gene expression data (Supplemental Fig. S7B). For example, we find that the two cell types of hematopoietic lineage, GM12878 (lymphoblastoid cell line) and K562 (chronic myeloid leukemia), clustered together using either expression or chromatin data. Embryonic stem cells do not have a considerably different number of open chromatin sites and do not contain a superset of open chromatin sites found in other more differentiated cell types. However, embryonic stem cell open chromatin sites tended to be larger and covered a greater fraction of the genome than other cell types (Table 1).

### The discovery of human regulatory elements by open chromatin mapping is far from saturation

We created union sets for every possible combination of 2, 3, 4, 5, 6, and 7 cell types and plotted the rate at which new sites appeared (Fig. 3A). Regardless of the threshold used to call the sites, the number of new sites identified does not abate as the number of cell lines analyzed increases. In contrast, performing the same analysis





**Figure 2.** DNase-seq and FAIRE-seq identify overlapping and unique sets of open chromatin. (A) Comparisons of the top 10K, 25K, 50K, and 100K DNase-seq and FAIRE-seq peaks from a single cell line (GM12878), with overlap indicated *below* each Venn diagram. (B) Average percentage of DNase and/or FAIRE peaks, as well as permuted coordinates, in defined positional categories based on their relationship to annotated genes. Error bars represent the standard deviation over seven cell types. Several categories deviated significantly from random (Supplemental Table S2). (C) The percentage of CTCF ChIP-seq peaks that overlap DNase and/or FAIRE sites in all seven cell types. The x-axis values indicate different signal thresholds for calling sets of CTCF peaks, where the threshold is increasingly more stringent from *left to right*. (D) The same as C, except for MYC ChIP-seq data. (E) Percentage of TSSs with overlapping Pol II ChIP-seq, DNase, and/or FAIRE peaks in seven cell types. x-axis represents expression values for corresponding genes indicating high (7+), medium (5–7), or low/no (0–5) expression.

on DNase-seq data from lymphoblastoid lines derived from seven different individuals (McDaniell et al. 2010) shows clear signs of saturation after the third cell line (Supplemental Fig. S8). This indicates that testing additional cell types is necessary and will continue to uncover new open chromatin sites.

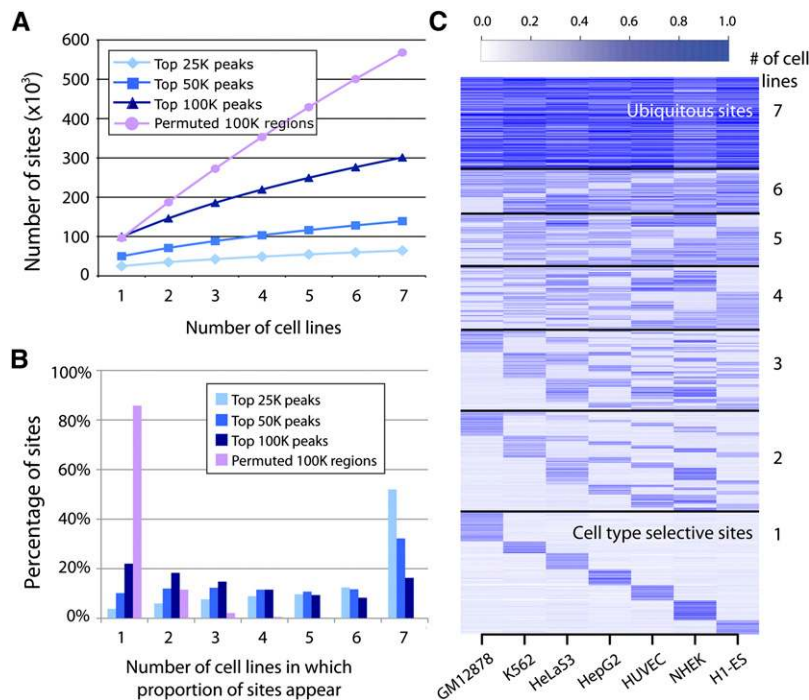
### Open chromatin sites found in all seven cell types are characterized by high signal

The number of open chromatin sites held in common among these seven cell lines, referred to as ubiquitous sites, was much higher than would be expected by chance (Fig. 3B). These sites tended to have the strongest signals in some or all cell types. For example, of the union set of the 25K strongest combined open chromatin peaks in each cell type (64,400 sites total), nearly 52% (33,466) were present in each of the seven cell lines (Fig. 3B; Supplemental Table S4). Of the union set of the top 50K (139,133 total) and 100K (301,235 total) sites, 32% (44,750) and nearly 16% (49,009) were ubiquitous, respectively (Fig. 3B; Supplemental Table S4). In contrast, when we randomly permuted the genomic coordinates of the top 100K sites for each cell line across the genome, over 85% appeared in only a single cell type, while <0.02% were ubiquitous (Fig. 3B; Supplemental Table S4). Overall, open chromatin sites with the strongest signals tend to be detected across cell types, while cell-type selective sites tend to produce weaker signals (Fig. 3C). Indeed, the distributions of  $-\log_{10}$  (*P*-values) for open chromatin sites categorized by the number of cell lines in which the sites appeared were found to be correlated (Supplemental Fig. S9), with ubiquitous sites having significantly higher amplitude than all other categories ( $P < 10^{-16}$ ; pairwise *T*-tests). No single cell type dominates the cell-type selective signal (Fig. 3C).

### Ubiquitous open chromatin sites tend to be near TSSs and are often bound by CTCF, while cell-type selective sites are distal with little CTCF binding

We examined the location of ubiquitous and cell-type selective open chromatin relative to genes, considering for this analysis the union of the top 100K combined open chromatin sites across all cell types. We found that ~30% of ubiquitous sites was located near transcription start sites, ~35% was within intergenic regions, and ~35% was within transcribed regions (see Methods for definitions of categories) (Fig. 4A). Ubiquitous open chromatin sites were much more likely to occur near TSSs, while cell-type selective sites were rarely found near TSSs (Fig. 4A). This suggests that cell-type selective gene regulation is controlled through distal regulatory elements.

CTCF has been shown to perform diverse regulatory functions, but it is primarily identified as an insulator that blocks interaction between promoters and enhancers. Overall, CTCF binding occurred in 28% of this union set of open chromatin sites but was bound to >55% of ubiquitous open chromatin sites (Fig. 4A). In contrast, CTCF bound to <5% of cell-type selective open chromatin sites. We hypothesized that open chromatin sites with evidence of CTCF binding should be more likely to function as insulators than open chromatin sites without CTCF. We tested 15 open chromatin sites with strong CTCF ChIP-seq signals for insulator activity (see Methods) (Supplemental Table S5). Seven of the sites were previously described (Xi et al. 2007). Five were chosen specifically based on the absence of the 20-bp CTCF binding motif to test for insulator function even in the absence of this motif. We also tested three open chromatin sites that lacked evidence of CTCF binding and six sites with no strong evidence of CTCF binding or open chromatin in K562 cells (Supplemental Table S5).



**Figure 3.** Distribution of open chromatin regions across cell types. (A) Saturation of total open chromatin sites discovered as a function of the number of cell types tested ( $x$ -axis). The rate of new top 25K sites per cell type was lower than for top 50K and 100K sites, likely reflecting more ubiquitous sites in this top fraction. (B) Percentage of the top 25K, 50K, and 100K combined open chromatin sites ( $y$ -axis) detected in one to seven of the cell types tested ( $x$ -axis). Over 50% of the top 25K open chromatin sites were ubiquitous, while more top 50K and 100K peaks were cell type selective. (C) Top 100K combined open chromatin sites partitioned by number of cell types in which they appear ( $y$ -axis). Color intensity indicates strength of open chromatin signal in that cell type.

The majority of open chromatin sites also bound by CTCF displayed insulator activity (Fig. 4B). The absence of a canonical CTCF binding motif did not appear to impact insulator function. To support this finding, we deleted the CTCF motif(s) from three potent insulators to determine the effect of removing the known binding site on insulator activity. We found that for one construct, the deletion of the CTCF site completely ablated insulator activity. However, deleting the CTCF binding sequence from the other two sites had no significant effect on insulator activity (Supplemental Fig. S10). At these sites, CTCF may bind to DNA through an unknown motif or protein intermediate. It is also possible that another unknown protein or chromatin configuration provides insulator function at these sites.

### Cell-type selective open chromatin harbors DNA sequence motifs corresponding to master regulators of cell identity

Regulatory elements far from the TSS have been shown, typically through single-gene experiments, to control tissue-specific and cell-type selective gene expression (Deal et al. 2006). Using the top 100K combined open chromatin sites, we used two approaches to characterize and discover DNA sequence motifs in cell-type-selective DNase-seq and FAIRE-seq sites located away from the TSS.

Known transcription factor binding motifs were identified genome-wide using publicly available position weight matrices (PWMs) from the Transfac database (Matys et al. 2006). For each cell type, we determined the top 10 PWMs that were significantly enriched in distal open chromatin sites specific to that single cell

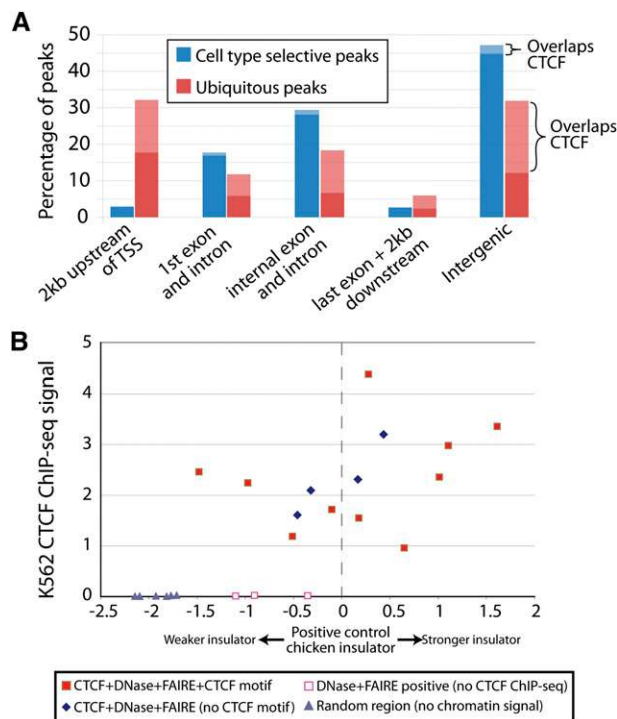
type (see Methods) (Supplemental Table S6). Among these are PWMs that match the binding specificities of factors known to function in processes that occur in the corresponding cell type. For example, the binding sites for IRF1 and IRF7 and the interferon-stimulated responsive element ISRE are enriched in lymphoblasts, which are interferon-responsive. Other examples include GATA family members, which regulate hematopoiesis, in K562; BACH1, necessary for DNA repair and linked to several cancers, in HeLaS3; HNF1A and HNF4A, transcription factors critical for liver development, in HepG2; ELK1, involved in endothelial cell differentiation, in HUVEC; and PLXNA2 family members critical for pluripotency and SP1 in embryonic stem cells.

We also performed de novo motif finding using the cERMIT (Georgiev et al. 2010) and CisFinder (Sharov and Ko 2009). We analyzed the top five motifs returned from each cell line by each algorithm. Using the software STAMP (Mahony and Benos 2007), we searched for motifs in the Transfac (Matys et al. 2006), JASPAR (Bryne et al. 2008), UniPROBE (Newburger and Bulyk 2009), and hPDI (Xie et al. 2010) databases that corresponded to each discovered motif ( $P$ -value  $< 10^{-9}$ ) (Fig. 5A). Consistent with our analysis above, both cERMIT and CisFinder detected the GATA1 motif in

K562, HNF1/4A motifs in HepG2, FOS and TP53 in NHEK, and ETS/ELK motifs in HUVEC. CisFinder identified a highly enriched POU5F1 motif in the embryonic stem cell line.

For each of the transcription factors corresponding to the discovered motifs, we determined their RNA expression levels in each of the seven cell types (Supplemental Table S7). In 10 of 13 instances, the cell type in which the motif was identified expressed that gene at the highest or second highest level among all seven cell types (Fig. 5A). In nearly all cases, the associated transcription factor has been functionally linked to the corresponding cell type (Fig. 5A; Hall et al. 1995; Nichols et al. 1998; Jessen et al. 2000; Taniguchi et al. 2001; Dejana et al. 2007; Shimizu et al. 2008; Bolotin et al. 2010). Both cERMIT and CisFinder also identified motifs that did not match any motifs in the databases we used—for example, a CCCCT motif in H1-ES (a stress-responsive element in yeast) and a CCAGCCTGG motif in HeLaS3 cells, a core sequence in *Alu* repeats.

We repeated the above motif analyses in distal FAIRE-only or DNase-only sites and found biologically relevant motifs. For example, DNase-only sites are most enriched for the same motifs as the combined open chromatin set in K562, GM12878, NHEK, and HepG2 (GATA1, STAT1, AP-1, and HNF4A, respectively). Likewise, FAIRE-only sites are enriched for many of the same motifs detected in the combined open chromatin set in HepG2, HeLaS3, and H1-ES (HNFs, PITX2, and POU5F1, respectively). In ES cells, the POU5F1 motif was found in FAIRE-only sites but not in DNase-only sites. Furthermore, not every motif found in the FAIRE-only or DNase-only sets matched those from the union set.



**Figure 4.** Ubiquitous and cell-type selective sites differ related to transcription start sites and presence of CTCF. (A) Percentage of ubiquitous and cell-type selective open chromatin sites in positional categories relative to annotated genes. Light bars represent open sites overlapping CTCF. (B) Insulator assays performed on sites with (1) DNase-seq, FAIRE-seq, and CTCF ChIP-seq signal and a CTCF motif (filled red squares); (2) signal in all three assays but without a CTCF motif (blue diamonds); (3) DNase-seq and FAIRE-seq signal, but not CTCF ChIP-seq (open red squares); and (4) no signal in any assay (gray triangles).  $y$ -axis indicates the signal from CTCF ChIP-seq in K562 cells. Enhancer blocking values ( $x$ -axis) were calculated as described (Supplemental Methods), with a value of zero equaling the measured activity of a known insulator.

For example, the K562 FAIRE-only sites are enriched for a well-defined NFE2L2 motif (Chen et al. 2010) that does not get picked up by DNase-only or the union set. In summary, the most highly enriched motifs are held in common between the assays, but each assay provides independent, biologically relevant information.

#### Open chromatin specific to a cell type occurs near genes that are expressed specifically in that cell type

If a substantial fraction of open chromatin sites function as positive regulators of transcription, one would expect increased expression of genes near open chromatin sites. To test this, we first identified distal cell-type selective sites present in only one of the tested cell types that were further than 2 kb from a TSS of a nearby gene. For each cell type, we calculated the average and median expression values of all genes mapped to each distal site. In all cases, expression levels of genes linked to distal cell-type selective sites for that cell type were significantly higher than in the remaining cell types (pairwise  $T$ -tests) (Fig. 5B). This was similarly true for six of seven cell types when considering proximal cell-type selective open chromatin sites within 2 kb of a TSS (Supplemental Fig. S11).

#### Cell-type selective open chromatin often occurs near genes that govern cellular identity and function

A number of cell-type selective open chromatin sites reside near genes encoding transcription factors that play critical roles in cell-type specific gene expression and function. In many cases, the motifs recognized by these proteins were themselves enriched in cell-type selective open chromatin (Fig. 5A). For example, in H1 embryonic stem cells, we identified a number of ES-cell specific open chromatin sites around the *POU5F1* and *NANOG* genes, which are known to control pluripotency (Supplemental Fig. S12). The upstream *NANOG* open chromatin site was recently identified as a poised enhancer, which is consistent with our assays (Rada-Iglesias et al. 2010). Other notable examples include a K562 cell-type selective open chromatin site upstream of the *GATA1* transcription factor gene (Supplemental Fig. S13) and a number of HepG2 cell-type selective open chromatin sites within and around the *HNF4A* transcription factor gene (Fig. 1B). In this latter case, we detect open chromatin at the two annotated TSSs (Fig. 1B), both of which are utilized in FT0-2B, another hepatocellular carcinoma cell line (Thomas et al. 2001). These are singular examples, but our chromatin atlas identifies tens of thousands of such putative regulatory elements in each of the cell types we have studied.

#### Clusters of open regulatory elements coordinate cell-type selective gene expression

Our analyses above focused on individual cell-type selective open chromatin sites and their relation to genes with a known function. However, we noticed regions in which multiple open chromatin sites in a given genomic region were coordinately present or absent across one or more cell types (Fig. 1B; Supplemental Figs. 12, 13). This was consistent with previous reports of clustered FAIRE sites called clusters of open regulatory elements (COREs) (Gaulton et al. 2010). To detect COREs systematically in our data set, we calculated pairwise correlations of open chromatin signals using data from all seven cell types (see Methods) (Fig. 6A,B). Using these correlations, we designed a hidden-Markov model to define 181 high-confidence COREs (see Methods) (Supplemental Table S9; Supplemental COREs figures; Supplemental COREs table). COREs varied in size from 32 kb to 6.6 Mb.

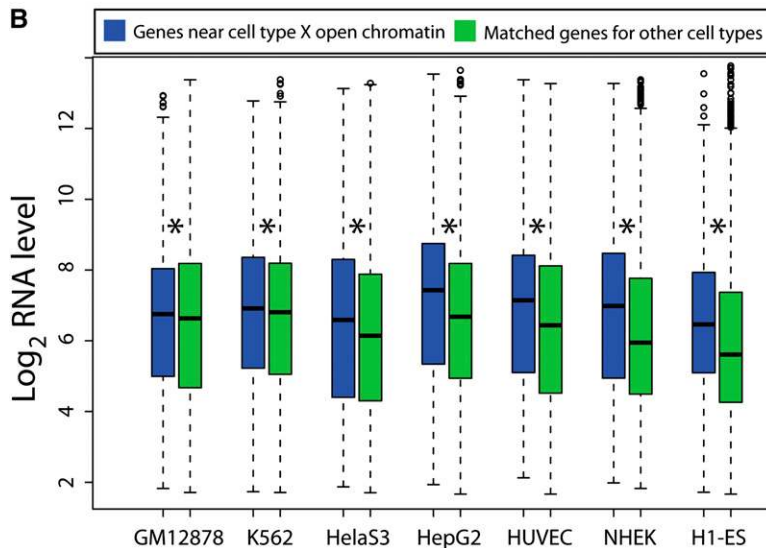
We hypothesized that COREs represent coordinated nucleosome depletion events caused by multiple regulatory elements participating in the regulation of a nearby gene or genes. We determined in which cell line(s) each CORE was active using the Mann-Whitney Wilcoxon rank sum test (see Methods) (Fig. 6C). Ninety-five COREs (52%) had increased open chromatin within primarily one cell-type, while 78 COREs (43%) were characterized by increased open chromatin within at least two cell types (Supplemental Table S9). In the remaining eight COREs (5%), no cell type had significantly more open chromatin signal relative to the others, as defined by our threshold. We examined genes inside or within 10 kb of each CORE and found that, for 75 of the 114 COREs with at least one gene with expression data, the cell type with the highest expression also contained significantly more open chromatin (Supplemental Table S9). Among the 67 COREs not associated with any genes for which we had expression data, the aggregate Pol II signal was greatest in a cell type with significantly enriched open chromatin (Supplemental Table S9). Considering all COREs, the highest cumulative CTCF signal was in a cell type with enriched open chromatin 77% (140/181) of the time. These relations with expression and Pol II and CTCF binding also hold when COREs are analyzed in aggregate (Supplemental



A

Cell line	Motif name	CisFinder/cERMIT rank	Discovered motif	Expression rank	Literature support
K562	GATA1	1/1		1	GATA1 has been linked to leukemias [22]
GM 12878	IRF1	1/1		1	IRFs are involved in the development of lymphocytes [23]
	HOXB13	5/ND		2	
NHEK	AP-1	1/1		1	AP-1 responsive elements are contained within keratinocyte structural genes [24].
	TP53	2/2		4	
HepG2	HNF4A	ND/1		1	HNF4A is essential for liver function [25].
	NR2F1	1/2		2	NR2F1/COUP-TF and HNF4 can function as accessory factors [26]. They have highly similar motifs.
	HNF1A	3/5		1	
	NR6A1	4/ND		2	NR6A1 is highly similar to HNF4A and NR2F1.
HUVEC	c-Ets-2	1/1		4	ETS involved in endothelial cell differentiation and angiogenesis [28].
	STAT5B	2/ND		4	
HeLaS3	TOPORS1	4/ND		1	
H1-ES	POU5F1	1/ND		1	POU5F1 is a well-known ESC pluripotency factor [29]

B



**Figure 5.** Distal cell-type selective open chromatin contains functionally relevant motifs and is linked to cell-type specific expression. (A) Top motifs enriched ( $P$ -value  $< 1 \times 10^{-9}$ ) in cell-type selective open chromatin. Expression rank reflects the transcription factor's expression level in that cell type relative to all other cell types. (B) Distribution of expression values for genes closest to distal cell-type selective open chromatin sites ( $>2$  kb from a TSS) from each cell type ( $x$ -axis) for that cell type (blue box plots). Similar distributions were calculated for these genes in the six other cell types lacking the distal open chromatin sites (green box plots). Asterisk indicates significant difference (pairwise  $T$ -tests).

Fig. S14). In 18 of 175 COREs, open chromatin levels did not correspond to gene expression, CTCF binding, or Pol II binding. For these cases, we may not have correctly identified a distant target gene associated with the CORE, or the open chromatin regions may not be acting as enhancers.

To demonstrate the utility of identifying COREs, we show one typical CORE in detail (Fig. 6A). This CORE extends over 1.2 Mb, but *GYPC* is the only annotated gene found in this region, so we focused on the 90 kb surrounding the gene (Fig. 6B). *GYPC* encodes both glycophorin-C and glycophorin-D, which function in membrane stability of human erythrocytes and lymphocytes (Walker and Reid 2010). Open chromatin is detected at the *GYPC* TSS in nearly all cell types. However, nucleosome-depletion

events unique to GM12878 and K562 occur  $\sim 10$  kb upstream of the TSS and within the transcribed region of *GYPC* (Fig. 6B). These are accompanied by increased FAIRE and DNaseI signal (Fig. 6C), expression of *GYPC* (Fig. 6D), and higher Pol II signal (Fig. 6E–G). Thus, this CORE is typical, showing coordinated nucleosome depletion, gene expression, and transcription factor binding, and identifies several key regions that may be responsible for the cell-type specific regulation of this gene.

Many other COREs reveal putative regulatory elements surrounding genes with cell-type specific functions. For example, CORE 70 defines a 600-kb region on chromosome 1 that contains several genes with specific functions in keratinocytes, including “late cornified envelope” (LCE) genes and small proline rich proteins (SPRR). Within CORE 70, NHEK cells exhibit significantly more nucleosome depletion, higher expression of these genes, greatly increased amounts of Pol II signal, and greater CTCF signal (Supplemental COREs figures). In other cases, unannotated genes within COREs can be associated with a specific cell type. For example, CORE 60 is a 325-kb region encompassing the sparsely annotated *RNF152* (ring finger protein 152) gene. Increased nucleosome depletion, expression of this gene, and Pol II signal are observed specifically in HUVEC and NHEK cells (Supplemental COREs figures.).

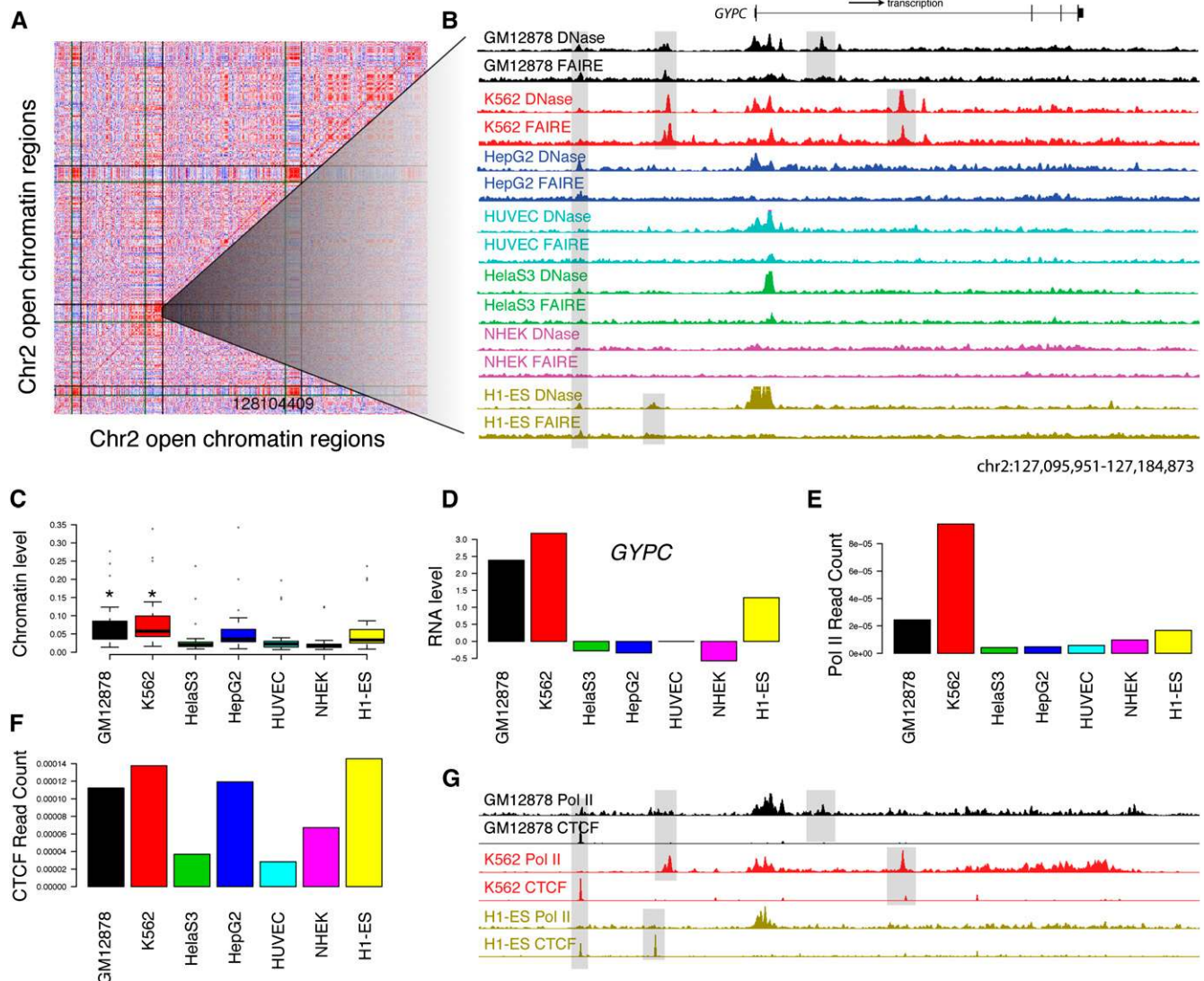
## Discussion

We produced maps of open chromatin in seven diverse human cell types using DNase-seq and FAIRE-seq. DNase-seq and FAIRE-seq are independent methods that provide strong cross-validation. Performing both assays on cells collected from the same culture in each replicate helps to ensure that the differences we observe are due to the assay specificities, rather than experimental variation. We present evidence that sites detected by a single assay are biologically relevant and functional. In each cell type, we identify 100,000–200,000 open chromatin regions covering 1%–2% of the genome.

Differences in DNase-seq and FAIRE-seq may be due to the specific regulatory complexes bound at each open chromatin site, which could affect the ability of DNaseI to cut or formaldehyde to crosslink. DNase-only sites tended to occur at transcription start sites while FAIRE-only sites were more often found in distal regions.

It is possible that FAIRE cannot detect some nucleosome-depleted regions that are bound very tightly by nonhistone proteins if those complexes support a level of crosslinking similar to that of a nucleosome. However, FAIRE appears to capture chromatin structures away from promoters that the DNaseI enzyme cannot easily cut.





**Figure 6.** Open chromatin patterns form clusters of open regulatory elements (COREs). (A) Pairwise correlations between 500 open chromatin sites from chromosome 2 show three blocks of correlated sites (see Methods). Each row and column represents an open chromatin region found by both DNase-seq and FAIRE-seq in at least one of the seven cell types. Red indicates high correlation, white indicates no correlation, and blue indicates negative correlation. Vertical and horizontal lines show CORE boundaries. (B) DNase-seq (y-axis fixed at 0.1) and FAIRE-seq (y-axis fixed at 0.04) signals for a 90-kb subsection of CORE 98 containing the *GYPC* gene. *GYPC* is the only gene in this CORE. Highlighted are open chromatin sites found in all cell types, only GM12878 and K562 together, and GM12878 and K562 individually. (C) Boxplots show the distributions of open chromatin levels within open chromatin sites with CORE 98. GM12878 and K562 both have significantly higher levels of open chromatin (\*; Mann-Whitney Wilcoxon rank sum test). (D) Relative expression levels (y-axis) of *GYPC* show increased expression in GM12878 and K562 cell lines. (E) Open chromatin sites within CORE 98 also show higher normalized Pol II ChIP-seq read counts in GM12878 and K562 cell types. (F) Normalized CTCF ChIP-seq read counts do not show significant differences between GM12878 and K562 and other cell types CORE98. (G) Pol II and CTCF signals in this 90-kb region (shown in B) provide preliminary annotations of similar and differential open chromatin sites.

Most binding sites of regulatory proteins we examined were within open chromatin sites, suggesting that, in general, open chromatin sites are indicators of regulatory proteins operating in each cell type. Between any two cell types, 23%–48% of open chromatin sites are shared. Among the seven cell types, nearly 9% of the genome was identified as an open chromatin site by DNase-seq and/or FAIRE-seq, and the identification of open chromatin sites is not complete. Continued experimentation on new cell types is a cost-effective strategy to find new regulatory elements and determine the fraction of the genome associated with a regulatory function.

DNase-seq and FAIRE-seq cannot directly reveal the function of the identified nucleosome-depleted regions, or the regulatory proteins that are bound to them. ChIP-seq, such as we performed for Pol II, MYC, and CTCF, provides a degree of functional annotation, as does association with certain histone modifications. Motif enrichment can provide guidance for selecting specific transcription factors to confirm by ChIP. We and others have shown that DNase-seq identifies DNaseI footprints, which reveal the location and identity of a bound motif with great accuracy (Hesselberth et al. 2009; Boyle et al. 2010; Pique-Regi et al. 2010). DNaseI footprinting will be critical in inferring binding at

individual open chromatin sites, but it cannot identify proteins that associate indirectly with DNA. As more proteins are mapped by ChIP-seq in relevant cell types, we can more fully annotate the open chromatin sites we report here.

Another challenge is mapping regulatory elements to the genes they regulate. Assuming that the nearest gene is the most likely target is clearly naïve, as demonstrated by counter-examples in the literature (i.e., Spilianakis et al. 2005). We showed that clusters of open regulatory elements defined by multiple sites spanning tens of thousands of bases show good correspondence between open chromatin and levels of gene expression, Pol II signal, and/or CTCF signal. Analysis suggests that COREs encompass noncoding DNA elements that act coordinately to regulate genes important for cell type identity and function. In many cases, the identification of COREs can guide candidate target gene selection, and methods like 3C, 4C, 5C, Hi-C, and ChIA-PET (van Steensel and Dekker 2010) will continue to be important in solving this difficult problem. Finally, low-throughput functional assays (similar to our insulator assays) will continue to be critical for understanding the biological activity of open chromatin sites, but assays that can test thousands of DNA segments in parallel will be required to make any significant progress in characterizing how these regulatory sites work together in a given biological context.

There is a nearly inexhaustible number of combinations of human cell types, genotypes, disease states, and environmental conditions. Genome-wide association studies (GWAS) have linked variation in numerous noncoding regions with different diseases (Gaulton et al. 2010; Hindorf et al. 2009). It is likely that many of these associations are due to polymorphisms that affect gene regulation. Our atlas can be used immediately to guide further characterization of regulatory elements that may be causally linked to disease risk (Gaulton et al. 2010; Stitzel et al. 2010). Furthermore, we have shown previously that identification of open chromatin sites in the same cell type derived from different individuals can identify individual-specific gene regulatory elements (McDaniell et al. 2010). Larger studies with hundreds or thousands of individuals will allow identification of connections between DNA sequence variation, chromatin organization, transcriptional regulation, and disease risk on a population level.

## Methods

### Cell culture

Vendor information and standard cell growth protocols can be found at the UCSC ENCODE site (<http://genome.ucsc.edu/ENCODE/cellTypes.html>) (see Supplemental Methods).

### Experimental protocols

DNase-seq (Song and Crawford 2010), DNase-ChIP (Crawford et al 2006a; Shibata and Crawford 2009), FAIRE (Giresi et al. 2007; Giresi and Lieb 2009), ChIP (Bhinge et al. 2007; The ENCODE Project Consortium 2007), and the insulator/enhancer blocking (Bell et al. 1999) assays were performed as previously described with slight modifications (see Supplemental Methods). Exon arrays were processed following a standard protocol for the ENCODE Consortium (see Supplemental Methods).

### Data processing

For sequence data from all experiments, (1) sequences were aligned to the human reference genome (NCBI Build 36) using MAQ (Li

et al. 2008), and (2) filtered to remove artifacts, (3) replicates were compared for reproducibility, then combined, and (4) base-pair signal was generated using F-seq (Boyle et al. 2008b) and discrete peaks called (see Supplemental Methods). Gene-relative categories were defined as follows: (1) promoter: overlaps 2 kb upstream of any TSS; (2) 5': overlaps first exon or first intron; (3) intragenic region: overlaps internal exon or intron; (4) 3': overlaps last exon or 2 kb downstream from end of transcription; and (5) intergenic: not within any previous category. Sites were assigned to the first category whose criterion was satisfied. Cell type selective and ubiquitous open chromatin sites were calculated using the top 100K sites from each cell type. Combined union sets of DNase-seq and FAIRE-seq sites were created with significance calculated using Fisher's combined probability test (Fisher 1925). Affymetrix Exon 1.0 ST array data (available at GEO, GSE15805) was processed using the Affymetrix Expression Console (see Supplemental Methods).

### Motif finding

Cell-type selective distal peaks were defined as present in a single cell type, at least 2 kb upstream of any TSS, and downstream from 5' exons and introns. Enriched motifs were determined in two ways: (1) Motif scanning was performed using public transcription factor position weight matrices from Transfac (version 7.0; (Matys et al. 2006) with enrichment defined as the ratio of predicted binding site frequency per kb in peaks from one cell type vs. cell-type selective peaks in the other six cells, and significance determined using a  $\chi^2$  test ( $P$ -value  $< 6.6 \times 10^{-3}$ ); and (2) de novo motif finding was performed using cERMIT (Georgiev et al. 2010) and CisFinder (Sharov and Ko 2009). For cERMIT, the combined open chromatin  $-\log(P$ -value) was used as experimental evidence. The union set of cell-type selective peaks from the other six cell lines served as the background. The online version of CisFinder (<http://lgsun.grc.nia.nih.gov/CisFinder/>) was used with default parameter settings, except "clustered" motifs rather than the "elementary" ones. FASTA files of cell-type selective distal peaks were the foreground set and the union set of distal peaks from the other six cell lines was the background set. Top motifs from cERMIT and CisFinder were annotated using the STAMP web server (<http://www.benoslab.pitt.edu/stamp/>) [Mahony and Benos 2007], with the "selected eukaryotic" option, and included Transfac (Matys et al. 2006), JASPAR 2010 (Portales-Casamar et al. 2009), UniPROBE ([http://the\\_brain.bwh.harvard.edu/uniprobe/](http://the_brain.bwh.harvard.edu/uniprobe/)), and a few organism-specific databases. Additional comparisons were done to the hPDI database, (Xie et al. 2010) using a custom PWM file, and to the "predicted human" database in STAMP (Mahony and Benos 2007).

### Clusters of open regulatory elements

Open chromatin sites found by both DNase-seq and FAIRE-seq in at least one cell type were considered. Each site was represented by a vector of seven combined open chromatin values, one for each cell type and quantile-normalized across cell types. Pairwise correlations were calculated between each site and 500 surrounding sites. One hundred and eighty-one high-confidence COREs were defined by a two-state HMM, using the average correlation across five adjacent open regions as the observable. Transition and emission probabilities were set manually. In each CORE, cell types with significantly more open chromatin signal were determined using pairwise Mann-Whitney tests ( $P$ -value  $< 0.05$  vs. at least four cell types). Genes (UCSC knownGene annotation) overlapping and within 10 kb were assigned to each CORE for expression comparisons. CTCF and Pol II ChIP-seq signal were calculated as normalized sequence read counts mapped inside COREs. To

eliminate background signals, only reads located within the top 20K Pol II or top 10K CTCF peaks were considered.

## Data access

All data from this atlas are publicly available on the UCSC Genome Browser (<http://genome.ucsc.edu>; [Kent et al. 2002]), the Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>), GSE30227 [Edgar et al. 2002]), and the NCBI Sequence Read Archive (SRA, <http://www.ncbi.nlm.nih.gov/Traces/sra/>, SRP007348, SRP007349, SRP007350, SRP002002, SRP004453 [Wheeler et al. 2008]).

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