

Optimization and Characterization of Bioactive Biocomposite Film Based on Orange Peel Incorporated With Gum Arabic Reinforced by Cr2O3 Nanoparticles

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Research Article

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26 Abstract

In this paper, the effect of adding gum Arabic at levels of 0-5%, and chromium oxide 27 nanoparticles (Cr₂O₃ NPs) at levels of 0-3%, are investigated on orange peel-based films. The 28 29 obtained results reveal a significant increase (p<0.05) in water vapor permeability, weight loss, tensile strength, and Young's modulus of film samples by increasing the percentage of 30 both gum and nanoparticles. Moreover, the addition of gum Arabic and Cr₂O₃ NPs decreases 31 the thickness, water-solubility, L*, a*, b* indexes while increasing the elongation to the 32 33 breaking point. Furthermore, the moisture content of the film samples was decreased by the 34 addition of nanoparticles, however, the addition of gum Arabic increased this parameter. The obtained results from the morphology of the samples indicated an increase in both roughness 35 and cracks by increasing the percentage of nanoparticles as well as creating a smooth surface 36 with the addition of gum Arabic. Besides, the results of FTIR revealed no new peak in the 37 prepared samples, as compared to the control sample. The results of XRD indicated that the 38 39 addition of gum Arabic and nanoparticles simultaneously caused the formation of new crystals and increasing the crystallinity of the films. Based on TGA results, the thermal 40 41 stability of films containing the nanoparticles increased, as compared to the control sample. In the meantime, the addition of gum and nanoparticles increased the antimicrobial properties 42 of the film samples, as compared to the control. Overall, those films created by the orange 43 peel including gum Arabic and Cr₂O₃ NPs could enhance the mechanical properties and 44 water vapor permeability of the samples. 45

46

47 Keywords: Biodegradable film; Orange peel; Gum Arabic; Cr₂O₃ nanoparticles; Response
48 surface methodology

49

50 Introduction

Plastic, well-known as one of the best human products, has now become a major challenge for both the environment and humans since it is an indestructible material with a shelf life of approximately 300 years. Here, it should be mentioned that the big volume of discarded plastics resultant from the packaging of food and hygiene products, now has created several problems, the most obvious of which is the irreparable damage to water, soil, air, and creatures [1-3].

By increasing the population as well as pressure on the restricted resources and the 57 environment, the use of renewable resources to produce edible and biodegradable films has 58 59 become increasingly important, which can improve product quality or decrease the waste disposal problems [4, 5]. Antimicrobial packaging as an active packaging type is one of the 60 packages, which has recently been widely used to increase the shelf life of food [6, 7]. 61 Antimicrobial food packaging acts to inhibit or delay the growth of microorganisms possibly 62 existing in the food package or the packaged food. Metal oxide nanoparticles are among the 63 64 compounds used in the antimicrobial packaging, among which, special attention has been paid to the formation and properties of chromium (Cr₂O₃), as an important heterogeneous 65 66 coating material, abrasion-resistant catalyst, solar energy storage, and high coloring.

Gums are one of the natural polymers that are recently utilized to synthase the packaging 67 materials. Gums or hydrocolloids have been used since 5000 years ago [8, 9]. They are 68 generally consumed in the food industry to change the texture, rheological properties and 69 preserve the appearance of food as a result of their ability to stabilize emulsions and water 70 storage [10]. Gum Arabic is one of the types of hydrocolloids, which is the best type of gum 71 because of its emulsification and encapsulation properties for use in oil-in-water emulsion 72 systems [11]. Other advantages of gum Arabic include its cost-effectiveness, high 73 concentration, and widespread use of various products [12]. 74

75 Orange fruit is widely used around the world as fresh produce and juice. Its peel is often 76 discarded as waste, which includes various secondary compounds with significant antioxidant 77 properties, as compared to other parts of the fruit [13]. Moreover, the orange peel is a good 78 source of molasses, pectin, and limonene, which are usually dried and mixed with dried pulp and sold as animal feed [14]. In this paper, the orange peel powder was used as another 79 component to prepare the packaging film. For this purpose, the biodegradable film based on 80 81 orange peel incorporated with gum Arabic containing chromium oxide nanoparticles (Cr₂O₃ NPs) was optimized and characterized. 82

83

84 Materials and methods

85 Materials

86 The oranges (Valencia cultivar) were bought from the local market of Urmia, Iran. Gum 87 Arabic, Cr₂O₃ NPs, corn starch, glycerol, calcium nitrate, calcium sulfate, potassium sulfate, sodium hydroxide, sodium chloride, and methanol 99.8% were from Merck (Darmstadt, 88 Germany). 2,2-diphenyl-1-picrylhydrazyl (DPPH) was made by Sigma-Aldrich (St. Louis, 89 MO, USA). The Nutrient agar culture medium was prepared by the Merck (Darmstadt, 90 91 Germany). Moreover, the standard strains of Staphylococcus aureus ATCC 25923 and Escherichia coli H₇:O₁₅₇ ATCC 700728 were prepared from the Iran Industrial and Scientific 92 Research Organization, Tehran, Iran. 93

94 Preparing orange peel powder (OPP)

95 The orange peel was dried at room temperature after washing and ground. After sieving, the 96 resulting powder was covered in a plastic bag and was stored in the refrigerator.

97 Film preparation

At first, two grams of orange peel were dissolved in 100 ml of distilled water and then stirred
for 20 h on a heated mixer at 30 °C at 250 RPM. Then, after 20 h, 40% glycerol (based on the

100 dry weight of orange peel) was added and stirred again for 30 min. In the end, after filtration,

101 the prepared solution was poured into 35 ml plates and then dried at room temperature.

102 Film characteristics

103 Thickness

The film thickness was measured using a digital micrometer at five random points of each film (around and center of each film). Afterward, the average thickness of different points of each film was utilized to calculate the mechanical properties and water vapor permeability [15].

108 Weight loss percentage

To measure the weight loss of the films, small pieces of the film were dried in an oven at 105
°C for 24 h. Then, the weight of the films was recorded before and after drying in the oven.
The amount of weight loss was then calculated as the percentage of initial weight loss as
follows:

113 Weight loss =
$$\frac{(W_0 - W)}{W_0}$$

where W_0 means the initial dry weight, W_1 represents the final dry weight [16].

115 Measuring the moisture content of the films

The films were cut into 2×2 cm and then weighed carefully. Afterward, they were placed in aluminum dishes and dried in an oven at 105 °C for 24 h. The moisture content was calculated based on the difference between the initial and final weight of the samples [17].

119 Water solubility

To measure the water solubility of the films, the film samples were prepared in 2×2 cm. They were placed in an oven at 110 °C for 6 h to obtain the initial dry weight. After weighing (W₁), the samples were immersed in the sealed dishes containing 50 ml of distilled water. The dishes were stirred as cross-sectional at 25 °C for 18 h. Then, the films were removed from the water and again placed in an oven at 110 °C for 6 h to achieve a constant weight. By reweighing, the samples, the final dry weight (W₂) was obtained. The percentage of watersolubility was calculated as follows [18]:

127
$$\%WS = \frac{W_1 - W_2}{W_1} \times 100$$

128 where W_1 is the initial dry weight and W_2 denotes the final dry weight.

129 Measuring water vapor permeability (WVP)

ASTM E96-05 method is employed to measure water vapor transfer (ASTM, 2005) using the 130 special vials. There was a 5 mm diameter hole in the lid of these vials, in which a piece of 131 film was placed. Then, 3 g of calcium sulfate was placed in vials. A piece of film was cut and 132 then placed in the lid of the vial, and closed on the vial. The vials were weighed with all the 133 134 contents and then placed in a desiccator containing a saturated solution of potassium sulfate. To ensure saturation, some precipitate of potassium sulfate was allowed to form on the 135 bottom of the desiccator. Saturated potassium sulfate at 25 °C produces a relative humidity 136 of 97%. Then, the weight of the vials was measured every 4 days for several hours. 137

The amount of water vapor transferred from the films was determined by increasing the weight of the vials. The weight gain curve of the vials over time was plotted, and after calculating the linear regression, the slope of the resulting line was calculated. Dividing the slope of the line associated with each vial by the total surface area of the film exposed to water vapor transfer, the water vapor transfer rate (WVTR) is obtained. The water vapor permeability (WVP) was then calculated as follows:

144
$$WVP = \frac{WVTR}{P(R_1 - R_2)}.X$$

where X means the film thickness (m), P denotes the pure water vapor pressure at 25 °C (3169 Pa), R_1 refers to the relative moisture in the desiccator (97%), and R_2 is the relative moisture inside the vial (0%). The test was implemented on each sample in three repetitions.

148 Measuring color properties

A colorimetric device was employed to specify the surface color of the film samples. The
results were exhibited in light-dark (L*), green-red (a*), and blue-yellow (b*) [19]. Besides,
the whiteness index (WI) and Chroma (C*) were calculated using the following equations :

152
$$WI = 100 - \sqrt{(100 - L^*)^2 + {a^*}^2 + {b^*}^2}$$

Chroma =
$$\sqrt{(a^*)^2 + (b^*)^2}$$

154

155 Measuring mechanical properties

156 The mechanical properties of the film samples were specified using the tensile tests by a Texture Analyzer based on the instructions of the ASTM D882-10 standard method. To 157 implement the tensile test, the samples were conditioned in a desiccator including the 158 magnesium nitrate saturated solution with relative moisture of 50±5% for 24 h. Then, they 159 were cut into rectangular strips, and both longitudinal ends of each film were placed between 160 161 the two jaws of the device [20]. By initiating the test operation, the film was pulled between the two jaws until it was torn, based on which the work process appeared as a stress-strain 162 diagram. The tensile strength, elongation at the breaking point, and the modulus of elasticity 163 properties of the films were calculated as follows: 164

165 Ultimate tensile strength =
$$\frac{F_{\text{max}}}{A}$$

166 Elongation at break =
$$\frac{L_{max}}{L_0} \times 100$$

167 Young's modulus
$$= \frac{(F. L_0)}{(A. \Delta L)}$$

where A represents the film cross-sectional area (m²), F_{max} denotes the maximum force at the breakpoint (N), L_{max} means the film elongation at the breakpoint (m), L_0 is the initial length of film sample (m), F is the force (N) and ΔL represents the changes in the length of the sample to the breaking point (m).

172 Field emission scanning electron microscopy (FE-SEM)

To evaluate the morphology, the surface of the film samples was examined using Field emission scanning electron microscopy (FE-SEM) at room temperature. Before scanning, a gold coating in several nanometers thickness was coated on the fracture surfaces [21].

176 Fourier transform infrared spectroscopy (FTIR) test

Fourier transform infrared spectroscopy (FTIR) was utilized in the range of 400-4000 cm⁻¹ to survey the interactions in the film matrix. First, the device was zeroed with KBr tablet as a control sample, and then the samples were prepared for FT-IR analysis by mixing 1 mg of completely dried samples via 150 mg of dry KBr powder. The thin tablets were prepared from each sample by compressing the mixture in a press device [22].

182 X-ray diffraction test (XRD)

183 X-ray diffraction pattern of the film samples was employed to investigate the crystal structure
184 of the prepared films and determine the distribution of components in the polymer matrix
185 [23].

186 Thermogravimetric analysis (TGA) change test

The dried film samples were analyzed from 20 to 500 °C at a heating rate of 10 °C/min along
with the pure nitrogen gas at a rate of 20 mm/min.

189 Investigating antimicrobial properties of the films

To determine the antimicrobial properties of active films, the method of penetration of antimicrobial compounds in agar medium (every 4 days) was utilized. In this method, the films were cut into circular plates and then transferred to the nutrient agar culture medium, which was previously inoculated with 10^5 - 10^6 cfu/ml of *Escherichia coli* (*E. coli*) or *Staphylococcus aureus* (*S. aureus*) microorganisms. After that, petri dishes comprising the contaminated culture medium along with antimicrobial films were kept in an incubator at 37 °C for 24 h. To obtain the degree of the microorganisms growth inhibition by the film, the diameter of the growth inhibition zone formed around the films was measured using a caliper[24].

199 Statistical analysis

200 In this paper, the effect of two numerical factors on the concentration of gum Arabic and Cr₂O₃ NPs are investigated using the Response Surface Methodology (RSM) in the form of a 201 central composite design. In this design, 13 samples (including 4 factorial points, 4 axial 202 203 points, and 5 central points are considered to estimate the mismatching and reproducibility). After implementing the experiments and data collection to test the significance of the factors 204 205 and their interactions, the method of analysis of variance (ANOVA) and Fisher distribution was employed at the significance level of α =0.05. The Design expert software V.11 was 206 utilized to analyze the data and draw the graphs. 207

208

209 Results and discussion

210 Thickness

The results indicated that by increasing the percentage of Cr₂O₃ NPs, the film thickness 211 decreased, whereas the addition of gum Arabic had little effect on the film sample thickness 212 (Fig. 1). Overall, those films via the highest levels of nanoparticles and gum had the highest 213 thickness, which might be due to the increase in the solid materials [25]. The reason for the 214 thickness reduction behavior through increasing the nanoparticle percentage is also related to 215 the increased density in the polymer film structure by the Cr₂O₃ NPs. According to the results 216 of several researchers, it can be concluded that the thickness of the films changes based on 217 218 the type of polymer and the added nanoparticles, and can cause the thickness to decrease, increase, or remain constant. 219

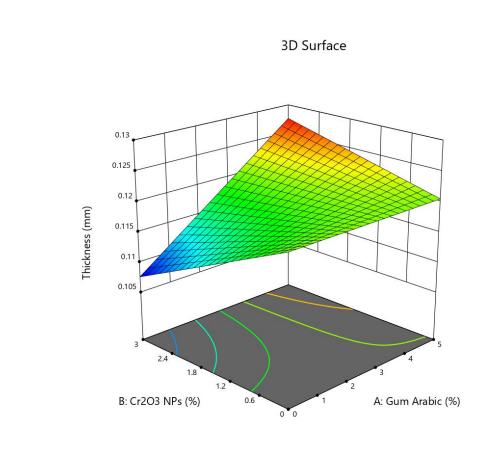




Fig. 1: The three-dimensional curve of the effect of nanoparticle-gum percentage on the thickness of films

225 Weight loss

FactoFedibeliAguActual

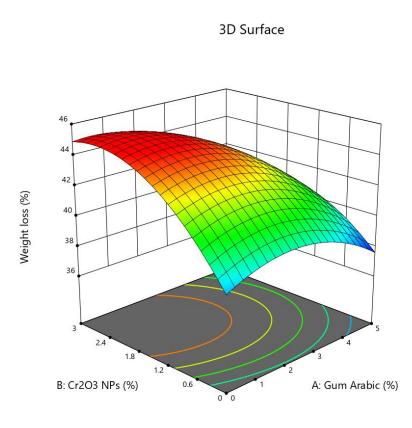
0.128

Thickness (mm) T**his (mm)**

X2 = B: Cr2O3 NPs X1 = A: Gum Arabic X2 = B: Cr2O3 NPs

0.108

Fig. 2 illustrates the three-dimensional curve of the film weight loss percentage. By increasing the gum content percentage, the film weight loss percentage reduced while increasing the Cr_2O_3 NPs, it was increased significantly (p<0.05). It should be noted that those samples containing the highest levels of nanoparticles and no gum Arabic had the highest weight loss. Moreover, the effect of the second-degree percentage of gum and Cr_2O_3 NPs on weight loss was also significant.





Factor Contractual

37.5 X1 = A: Gum Arabic X2 = B: Cr2O3 NPs X1 = A: Gum Arabic X2 = B: Cr2O3 NPs

Weight loss (%) Weight states

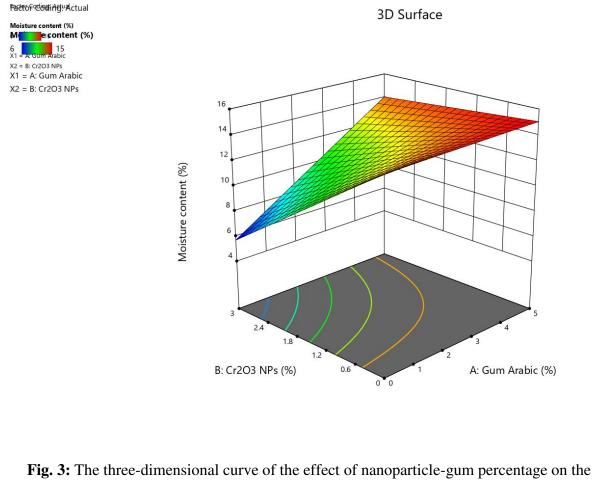
Fig. 2: The three-dimensional curve of the effect of nanoparticle-gum percentage on the
 percentage of weight loss of films

236

237 Measuring the moisture content of the films

Fig. 3 depicts the three-dimensional curve of the effect of gum and Cr_2O_3 NPs percentage on the amount of moisture. It should be mentioned that by increasing the percentage of gum, the moisture content of the film increased, while by increasing the percentage of Cr_2O_3 NPs, the moisture content of the film significantly decreased (p<0.05).

Besides, the moisture content of the film increased by increasing the gum. This seems to be due to the increased compression of the film matrix and the entrapment of more water in the film matrix structure. On the other hand, high hydrophilicity of the gum cause more water absorption by the gum and thus increasing the moisture content of the films [26]. Furthermore, the reduction in the moisture content with the addition of nanoparticles is likely due to the weak interaction of nanoparticles with the hydroxyl polymer group [25]. Our results in terms of the effect of adding nanoparticles on reducing the moisture content of films are in accordance with the results of Li et al. [27] who added zinc oxide nanoparticles to chitosan, as well as Bahrami et al. [25] adding the silver nanoparticles to the films based on hydroxypropyl methylcellulose and Tragacanth.



255

254

252 253

moisture content of the films

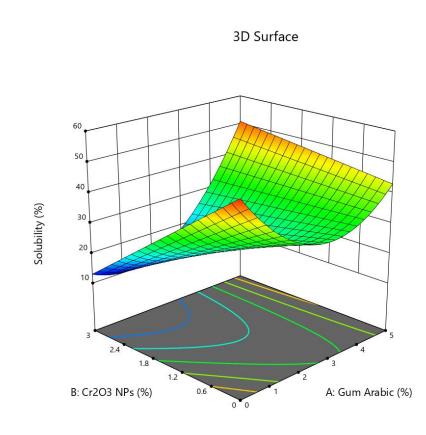
256

257 Water solubility

Fig. 4 exhibits the three-dimensional curve of the film solubility as a function of two variables of the gum and Cr_2O_3 NPs percentages. The obtained results revealed that by increasing the percentage of gum and Cr_2O_3 NPs, the solubility of the film decreased. Note 261 that the second-degree effect of gum and the linear effect of Cr_2O_3 NPs were also 262 significant.

Here, it is worth noting that solubility is an essential feature in biodegradable films because it 263 can resist the film compared to water, especially in the environment containing moisture such 264 265 as meat products, and determine the release speed of antioxidant and antimicrobial compounds when in contact with the material food. Adding the nanoparticles increases the 266 electrostatic bonding between film polymer, and thus decreasing the water solubility of the 267 film. Some similar results are provided by adding clay nanoparticles to the film based on 268 pectin [28]. Rezaei et al. [29] reported that the addition of zinc oxide nanoparticles reduced 269 270 the water solubility of the films. Our results in terms of the effect of gum on reducing water 271 solubility were in accordance with the result of Sui et al. [30]. They found reduced water solubility by increasing the amount of Tragacanth. Khoirunnisa et al. [31] expressed that the 272 addition of zinc nanoxide led to a significant change in the solubility of the films, which was 273 consistent with our findings in terms of the effect of nanoparticles on reducing the water 274 solubility of film. 275

They described this phenomenon attributed to the possible bonding between nanoparticles and film matrix. Some studies revealed that the addition of nanoparticles causes hydrogen bonding between nanoparticles and polymer matrix. As such, the bonding of free molecules of water and hydrophilic biopolymer groups reduced, and subsequently, the film solubility decreased [32].





FattoFediogiAguActual

X2 = B: Cr2O3 NPs X1 = A: Gum Arabic X2 = B: Cr2O3 NPs

55.2

Solubility (%)

Fig. 4: A three-dimensional curve of the percentage of nanoparticle-gum on the watersolubility of films

285

286 Measurement of water vapor permeability (WVP)

Fig.5 illustrates the three-dimensional curve of water vapor permeability (WVP) as a function of two variables of gum and Cr_2O_3 NPs. The results of the statistical analysis confirmed that by increasing Cr_2O_3 NPs, the amount of WVP increases significantly. Moreover, by increasing the gum surfaces, the amount of WVP of the samples decreased, and then slightly increased.

Furthermore, the second-degree effect of gum and Cr_2O_3 NPs was significant on WVP (p<0.05). Note that increasing the WVP of the films by adding nanoparticles, is due to porous structure and gaps caused by nanoparticles [33]. Similar results were obtained for films comprising Gracilaria vermiculophylla extract and zinc oxide nanoparticles [33].

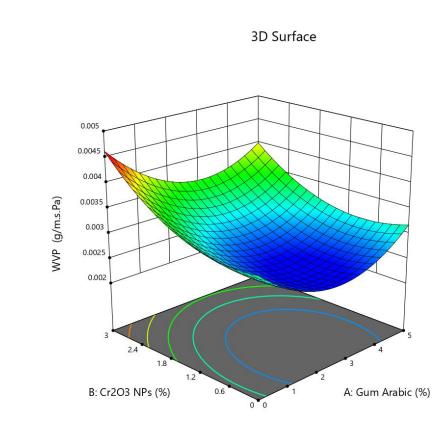




Fig. 5: A three-dimensional curve of the percentage of nanoparticle-gum on the water vapor
 permeability of the films

301 Measuring color properties

FattoFediogiAguActual

0.0045

WVP (g/m.s.Pa) WWB

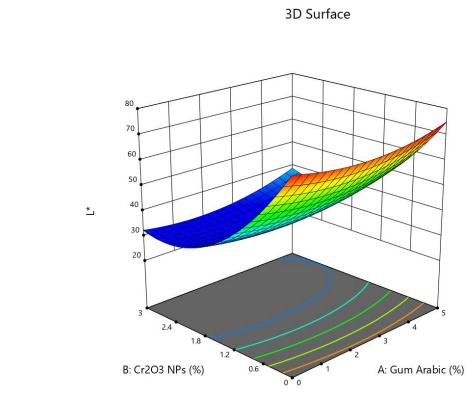
X2 = B: Cr2O3 NPs X1 = A: Gum Arabic X2 = B: Cr2O3 NPs

0.0025

The color and appearance of the polymer used in food packaging is an essential and effective factor for choosing and accepting the product by the consumer. Most food packaging films are transparent and colorless. Nevertheless, in some cases, the use of the inhibitor compounds of light, and generating the color in the matrix of the packaging material are necessary, due to the sensitivity of food to light, the loss of its nutritional compounds by optical oxidation, and color matching contents with packaging material to attract the consumer attention [34].

Fig. 6 depicts the three-dimensional curve of film lightness (L*) as a function of two variables of gum and Cr_2O_3 NPs percentages. The obtained results of the statistical analysis revealed that by increasing the gum percentage, the film L* increased; however, by increasing the percentage of Cr_2O_3 NPs, the amount of the film L* decreases significantly.

Sui et al. [30] indicated that the higher the ratio of gum to soy protein isolate, the brighter the color of the film will be. This is in line with our results revealing the increased L* along with increasing the gum. As can be observed, by increasing nanoparticles, the transparency of the films decreases, which was in accordance with the results of Asdagh et al. [35]. They described that by increasing the number of nanofibers, the transparency of polylactic acid films decreased. Besides, the decrease in L* with the addition of nanoparticles may be due to the matte appearance of Cr_2O_3 NPs [25].



Factor Coding Actual





Fig. 6: A three-dimensional curve of the percentage of nanoparticle-gum on the L* index



323

322

of films

- Fig. 7 exhibits the three-dimensional curve of the a^* index of the film as a function of two variables of gum and Cr₂O₃ NPs. The results indicated that by increasing the gum percentage, the value of a^* index increased, while by increasing the Cr₂O₃ NPs, the a^* index content of the films decreased. In terms of the effect of nanoparticles, it could be seen that by adding nanoparticles, the value
- of a* was reduced, which was consistent with the results of Oleyaei et al. [26].



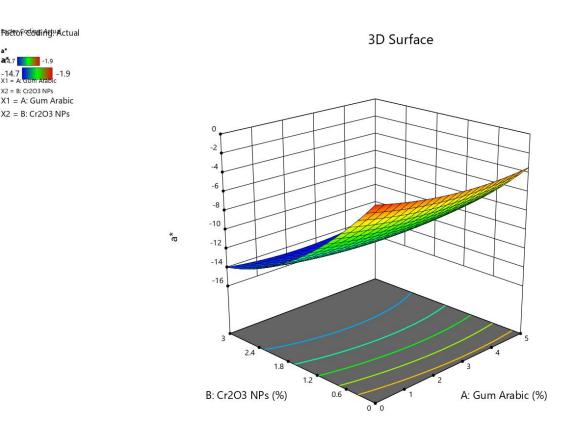


Fig. 7: A three-dimensional curve of the percentage of nanoparticle-gum on the parameter

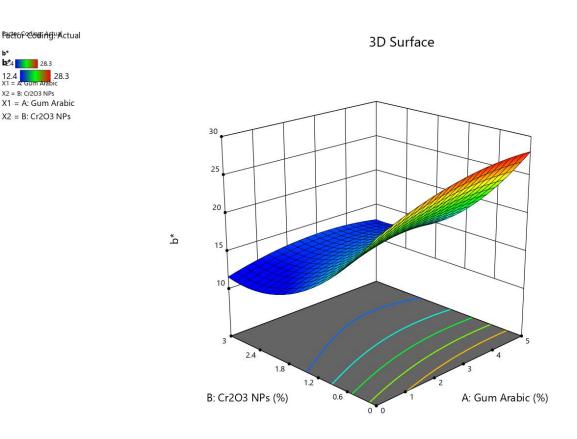
a* index films

334

Fig. 8 illustrates the three-dimensional curve of the b* index of the film as a function of two variables of gum and Cr_2O_3 NPs. The results of statistical analysis indicated that by increasing the percentage of gum, the amount of b* index of the film increased, while by

increasing the percentage of Cr_2O_3 NPs, the b* index of the film significantly decreased. Considering the effect of nanoparticles, it can be observed that by the addition of nanoparticles, the b* was decreased, which was in line with the results of Oleyaei et al. [26].





342

Fig. 8: A three-dimensional curve of the percentage of nanoparticle-gum on the parameter
b* index films

345

Fig. 9 depicts a three-dimensional whiteness index (WI) as a function of two variables of gum and Cr_2O_3 NPs percentage. The obtained results confirmed that the WI rate increased by increasing the percentage of gum, whereas by increasing the Cr_2O_3 NPs, the WI decreased.

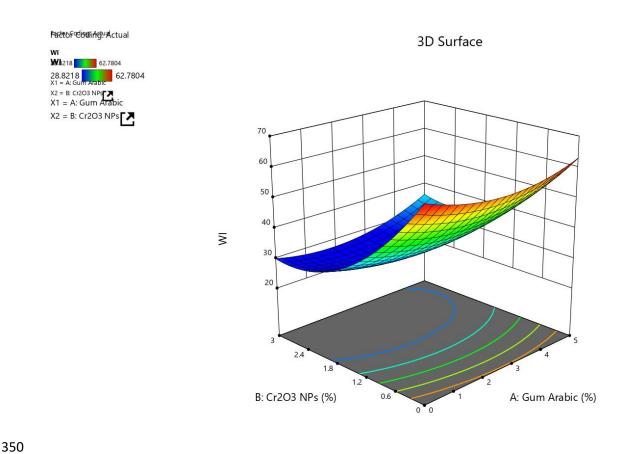
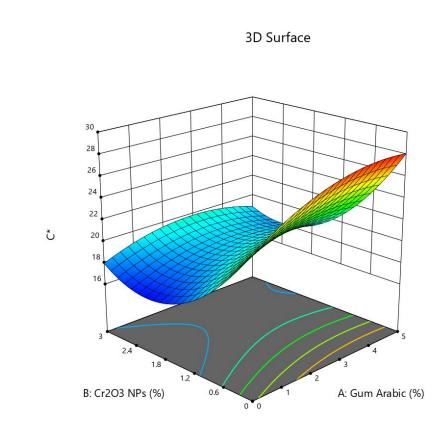


Fig. 9: A three-dimensional curve for the effect of the percentage of nanoparticle-gum on the
WI of films

Fig. 10 exhibits the three-dimensional curve of the film as a function of two variables of 354 gum and Cr₂O₃ NPs percentage. The statistical analysis revealed that by increasing the gum 355 percentage, the amount of C* index in the film increased, while by increasing the percentage 356 of Cr₂O₃ NPs, the film C* significantly decreased (p<0.05). The C* denotes a measure of a 357 color difference of gray, which can be defined as color purity. The calculation of the amount 358 of C* in the samples shows the highest amount of purity in the film samples. The results in 359 this regard were consistent with the results of He et al. [36]. They provided that with the 360 addition of oxide nanoparticles, the amount of C* decreased. 361



363 Fig. 10: The three-dimensional curve of the nanoparticle-gum percentage on C* index of 364 films 365

Measuring mechanical properties 367

Factor Coal Age Actual

28.554 16.2265 28.554 X2 = B: Cr2O3 NPs X1 = A: Gum Arabic X2 = B: Cr2O3 NPs

G.2265

368 Mechanical properties are considered the most important properties of materials used in food packaging. Among the important mechanical properties of biodegradable films are the tensile 369 strength and tensilability to the breaking point to determine their resistances in different 370 processes, transportation, and warehousing. 371

Table 1 lists the percentage of elongation, tensile strength, and elastic modulus of samples 6, 372 8, 12 against the control sample (sample 13). As can be observed, by adding the gum Arabic 373 and nanoparticles in samples 6 and 12, the tensile resistance parameter has a significant 374 increase, as compared to the control sample. This can be associated with the creation of 375 appropriate interactions between film matrix and additive materials (such as gum Arabic and 376 Cr₂O₃ NPs), where this parameter increased by forming new hydrogen bonding. 377

Furthermore, the uniformity of nanoparticles in the films increases the tensile strength, as compared to the control sample [37]. Moreover, the modulus of the elasticity reveals the same process and indicates the creation of interactions in the film matrix. This is in line with the results of Xu et al. [11] in terms of the addition of the ZnO-CMC combined nanoparticles to the pea starch film. Besides, He et al. [36] proposed that two parameters of elasticity modulus and tensile strength were increased by adding the zinc nanoxide to the gelatin fish.

In this matter, the addition of titanium oxide and silver nanoparticles to the Carboxymethyl 384 cellulose film increased both the tensile strength and modulus of elasticity [37]. However, in 385 sample 8 (i.e. the sample including the highest percentage of gum Arabic and chromium 386 oxide), it can be seen the decrease in these two parameters can be associated with the lack of 387 adequate interactions with the film matrix and the formation of bonding between 388 nanoparticles and gum. In other words, instead of the formation of abundant hydrogen 389 bonding with film matrix, nanoparticles and gum create abundant hydrogen bonding with 390 391 each other, which finally would result in reducing the tensile strength elasticity modulus. 392 Meanwhile, there was no logical process for the percentage of elongation (% E).

Table 1: The data on the mechanical properties of films based on orange peel and gum

394

Arabic

% E	YM (MPa)	TS (MPa)
\pm 0.88 ^b	34.07 ±1.21 ª	7.34 ±0.10 ^a
21.62		
17.69 ±0.67 ^a	60.41 ± 4.36 °	10.63 ± 0.39 ^c
\pm 0.13 ^b	$50.54~\pm~0.73$ ^b	10.76 ± 0.18 °
21.29		
\pm 0.83 ^b	41.98 ± 2.10^{a}	8.76 ± 0.20 ^b
20.95		
	$\pm 0.88^{b}$ 21.62 17.69 $\pm 0.67^{a}$ $\pm 0.13^{b}$ 21.29 $\pm 0.83^{b}$	$\begin{array}{c} \pm \ 0.88^{b} & 34.07 \ \pm 1.21^{a} \\ 21.62 \\ 17.69 \ \pm 0.67^{a} & 60.41 \ \pm 4.36^{c} \\ \pm \ 0.13^{b} & 50.54 \ \pm \ 0.73^{b} \\ 21.29 \\ \pm \ 0.83^{b} & 41.98 \ \pm \ 2.10^{a} \end{array}$

395 Different letters in each column indicate significance at the level of p < 0.05.

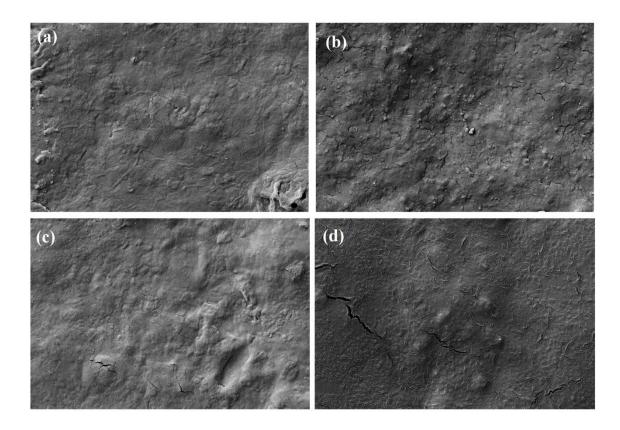
396

397 Field emission scanning electron microscopy (FE-SEM)

398 The analysis of the film's morphology provides information on the spatial layout of different 399 components in the film, which helps understand the mechanical properties and water vapor transfer mechanisms [38]. In this regard, the SEM images resulting from the sample surface 400 401 of the films are depicted in Fig. 11. The SEM images show the control film based on the dense and uniform surface of orange peel (Fig. 11a). As can be observed from Fig. 11b, 402 Cr₂O₃ NPs were uniformly distributed without agglomeration of particles at the surface of the 403 film sample. Besides, as compared to the control sample, adding nanoparticles to films 404 caused rough surfaces, more cracks, and a break at the surface of the film samples. 405

406 Oun and Rhim [39] reported the uniform distribution of zinc oxide nanoparticles in 407 carrageenan films. Based on Fig. 11c, the addition of gum Arabic to orange peel films caused 408 a uniform surface with no fracture. This condition may be resultant due to intra-molecular 409 interaction between polymer matrix. In this regard, the results of coordination were achieved 410 for those films based on carrageenan gum, Xanthan Gum, and Gellan gum, which were 411 prepared with different proportions [40].

In the images related to films containing 5% gum and 3% of Cr₂O₃ NPs, surface with more 412 and deeper cracks and fractures, and more non-uniformity can be seen, as compared to the 413 control sample. Besides, they had a non-homogeneous surface than the control film (Fig. 414 11d). These results are probably due to the interaction between polymer, gum, and 415 nanoparticles. Martins et al. [41] reported that films based on the carrageenan and lacuste 416 417 gum had a uniform and dense surface and the addition of clay nanoparticles increased the roughness of the samples. Moreover, the addition of TiO₂ nanoparticles to films based on 418 carrageenan gum, Xanthan Gum, and Gellan gum, caused the roughness of the sample [40]. 419



421 Fig. 11: The field emission scanning electron microscopy images (a) OPP, (b) OPP / Cr₂O₃,
422 (c) OPP / AG, and (d) OPP / Cr₂O₃ / AG

420

424 Fourier transform infrared spectroscopy (FTIR) test

FTIR analysis is a useful and practical method to study and identify the intra-molecular 425 interactions of the film samples. FTIR spectra of the film samples are provided in Fig. 12. 426 The peaks of the film are in the range of 600 cm⁻¹ to 4000 cm⁻¹. The FTIR spectra of the 427 control films were almost similar to other film samples and there was no significant change in 428 their functional groups. Meanwhile, there was no new peak in the film samples, indicating no 429 change in the pectin-based film of the orange peel comprising gum Arabic and nanoparticles. 430 Overall, the position and severity of changed peaks are related to the interaction between 431 nanoparticles and gum Arabic with a polymer matrix. The peak index in 325 cm⁻¹ reveals the 432 tensile vibrations of O-H and CH₂-OH groups, which is related to the presence of starch, 433 glycerol, and water compounds. Besides, the peak at 2923 cm⁻¹ is related to the C-H 434

435 alkankins and compounds in the polymer film matrix [42]. In this way, the absorbed bands of 2870-2960 cm⁻¹ are attributed to the symmetric and asymmetric C=H groups [42, 43]. The 436 peak in the range of 1605 cm⁻¹ is related to the N-H Amide groups. Moreover, the absorption 437 band in 1740 cm⁻¹ represents the presence of tensile bonding C=O in the Amide groups [44]. 438 The absorption peak in the range of 1410 cm⁻¹ is related to the vibrational group of O=H. 439 Meanwhile, the absorption peak in the range of -1200 cm⁻¹ to 1350 cm⁻¹ represents the 440 existence of the C-O tensile group in the Polysaccharide complex. The peak existing in the 441 range of 1015 cm⁻¹ to 1950 cm⁻¹ denotes C=O tensile and vibrating groups. Moreover, the 442 peaks observed in the range of 650 cm⁻¹ to 950 cm⁻¹ are related to the C=C and C-H bonding 443 of the aromatic ring [45]. Overall, the addition of gum Arabic and Cr₂O₃ NPs did not create a 444 new peak in the FTIR spectra. 445

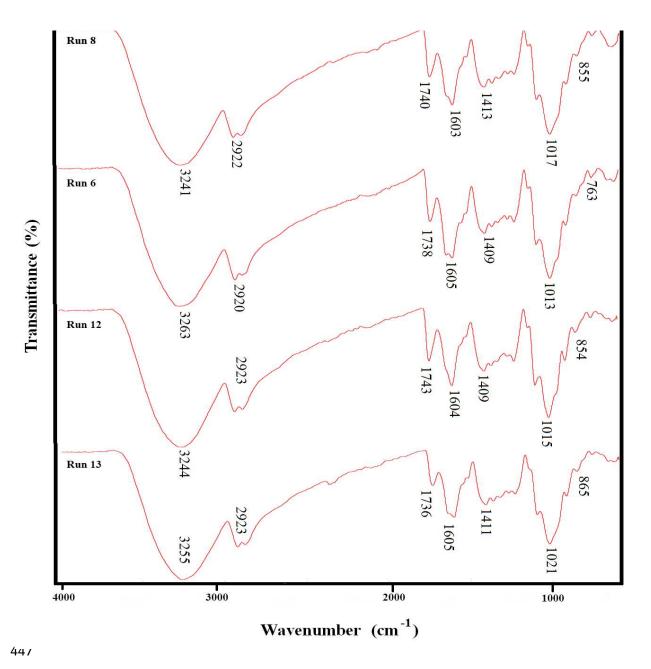


Fig. 12: The FTIR spectroscopy (RUN 13) OPP, (RUN 6) OPP / Cr₂O₃, (RUN12) OPP / AG 448 and (RUN 8) OPP / Cr₂O₃ / AG

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- 450

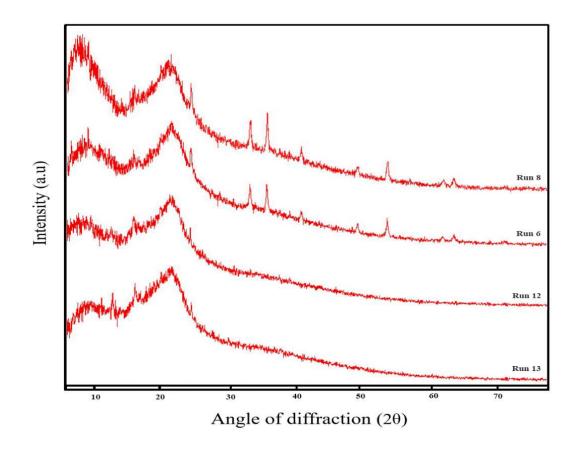
X-ray diffraction test (XRD) 451

The X-ray diffraction pattern is utilized to investigate the crystalline behavior of edible films 452 453 and their morphological properties [23]. The structure of compound films containing gum and nanoparticles was compared with the control sample (RNU 13) using the XRD test, 454 which is shown in Fig. 13. The XRD spectrum of the film indicated that the films in 2θ of 455

456 12.3, 15.8, and 24.5 have sharp peaks, indicating the existence of crystalline structures in 457 these films. Besides, those peaks with relatively broad heads in 2 θ of 8 and 12, indicate the 458 existence of an amorphous structure or irregularity in the film.

By adding gum Arabic in sample 12, the peak of $2\theta=12.3$ was removed while the peak height 459 460 of $2\theta=15.8$ was decreased. Meanwhile, the head of peak $2\theta=8$ was also relatively wider. All these changes including reduced crystalline structure, the presence of crystals, and the 461 increased amorphous structure in the film structure are caused by gum Arabic, as compared 462 to the control sample. In the XRD spectrum of samples 6 with the highest percentage of 463 nanoparticles and Arab gum, some new peaks in 20 of 33.7, 36.2, 41.6, 50.3, 58.9, 63.5, and 464 65 are created. All these sharp peaks indicate crystallization and the formation of new 465 crystals in sample 6, as compared to sample 13 as a result of the crystallization of 466 nanoparticles added to the film structure. 467

In sample 8 with the highest percentage of gum and nanoparticles, in comparison with sample 468 13, a displacement was created in the peak from $2\theta=8$ to $2\theta=7$, such that the height of this 469 470 peak was increased, indicating an increase in the crystallization degree of this sample, as compared to sample 13. In comparison with sample 6, a shift was created in sample 8 from 471 472 $2\theta = 58.9$ to $2\theta = 54.8$, in which the height of the peak was increased. Furthermore, in 2θ of 33.7, 36.2, and 41.6, the height of the peaks was increased, while at 2θ =50.3, the height of the 473 peak was decreased. Overall, the crystallization degree of sample 8 was more than other 474 tested samples, indicating that the addition of gum Arabic and nanoparticles simultaneously 475 caused new crystals, finally increasing the crystallization degree of the film. 476



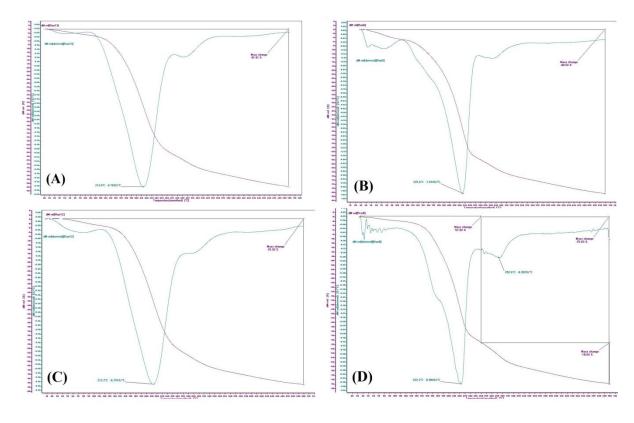
478 Fig. 13: The XRD curve (RUN 13) curve OPP, (Run 6) OPP / Cr₂O₃, (RUN12) OPP /
 479 AG, and (RUN 8) OPP / Cr₂O₃ / AG

477

481 Thermogravimetric analysis (TGA) change test

TGA analysis is employed to investigate the thermal stability of the films. TGA and DTG thermal diagrams are provided in Fig. 14. In all film samples, the initial weight drop was observed at 70-120 °C because of the evaporation of moisture from the surface of the films [38]. The next weight drop was observed at around 150-250 °C because of the thermal decomposition of polymer and glycerol evaporation used as a plasticizer [46].

In this regard, similar results were proposed by Slavutsky et al. [47] for the film based on Montmorillonite nanoparticles and BREA gum. The maximum decomposition temperature specified from the temperature peak of the DTG curve is 213° C for the control and film samples containing 5% gum Arabic, whereas it is 225 °C for film samples containing 3% of 491 the chromium oxidized nanoparticles, gum, and nanoparticles. As can be observed, by adding the nanoparticles, the maximum decomposition temperature increases, as compared to the 492 493 film samples without the nanoparticles. Furthermore, these results indicate that the thermal stability of orange peel film increases by adding the Cr₂O₃ NPs. Such behavior is likely due 494 495 to the increased bending in the path of gas emissions as well as the restriction of oxygen and 496 gases emission from the pyrolysis [47]. Oun and Rhim [39] proposed similar results reporting that the addition of zinc oxide nanoparticles to Carrageenans-based films increased the 497 498 stability of films.



500 Fig. 14: The TGA curve of (A) OPP, (B) OPP / Cr_2O_3 , (C) OPP / AG and (D) OPP / Cr_2O_3 /

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AG

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503 Investigating antimicrobial properties of films

The disk diffusion method was employed to specify the antimicrobial properties of gum and Cr_2O_3 NPs. In this technique, a piece of antimicrobial film is placed in a solid culture medium

506 containing the target microorganisms. After incubation at a specific temperature, the bright 507 halo around the film represents the emission of the antimicrobial agent from the film and thus 508 inhibition of the target microorganisms' growth. The disk diffusion method simulates 509 packaging the food, indicating the performance of the film in contact with the contaminated 510 surfaces as well as the immigration of the antimicrobial agent from the film to food [48].

The results of antimicrobial activity of gum and nanoparticles samples are illustrated in 511 512 comparison with the control sample on the gram-positive bacteria of S. aureus and gramnegative bacteria of E. coli in Fig. 15. As can be observed, the control sample does not show 513 514 the antimicrobial properties against both bacteria, and the halo is not observed around the film disk. Based on the obtained image, with the addition of gum and nanoparticles, the 515 growth halo was observed around the films, indicating the release of antimicrobial agents. 516 517 Nevertheless, as seen, the most halo was created in sample 8. However, there was no synergistic effect of gum and nanoparticles to destruct the tested microorganisms. 518

d gum Arabic

Diameter of non-growth area (mm)		
Staphylococcus aureus	Escherichia coli	
0 ^a	0 ^a	
19.59 ±1.24 ^b	0 ^a	
33.03 ±1.34 °	20.42 ±1.64 ^b	
$36.30 \pm 2.30^{\circ}$	18.99 ±1.02 ^b	
	Staphylococcus aureus 0 ^a 19.59 ±1.24 ^b 33.03 ±1.34 ^c	

520 Different letters in each column indicate the significance at the level of p < 0.05.

521

522 Moreover, as seen, adding gum alone did not have a specific antimicrobial effect on the 523 gram-negative bacteria of *E. coli*, which can be due to the presence of liposaccharide walls 524 around the Peptidoglycan wall of gram-negative bacteria resulting in more bacterial resistance against the antibacterial agents [49]. The destruction of microorganisms due to the addition of nanoparticles can be associated with the leakage of intracellular materials by the holes generated on the cell wall. These holes are formed by oxidation of wall liposaccharides of cells owing to the nanoparticles-caused changes and differences.

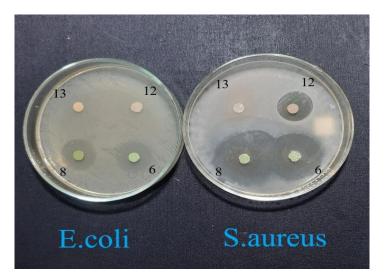


Fig. 15: The images of antimicrobial properties of films (RUN 13) OPP, (RUN 6) OPP/
Cr₂O₃, (RUN12) OPP/ AG, and (RUN 8) OPP/ Cr₂O₃ / AG

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529

533 Conclusion

In this paper, OPP was utilized as an available, abundant, and cheap substance, compared to 534 gum Arabic to prepare the edible films. Examining these films indicated that they had 535 mechanical properties, inhibition, and good water solubility. The results revealed that the film 536 solubility decreased by increasing the percentage of gum and Cr₂O₃ NPs. Furthermore, by 537 adding gum Arabic and nanoparticles, the tensile strength parameter had a significant 538 increase, as compared to the control sample, which could be associated with the creation of 539 540 appropriate interactions between film matrix and additive materials (such as gum Arabic and Cr₂O₃ NPs). Besides, the creation of new hydrogen bonding increased the parameter. 541 Moreover, the transparency of the films increased significantly by increasing the percentage 542 of gum. The antimicrobial properties of the films increased significantly by increasing the 543

- 544 nanoparticle percentage. Furthermore, the results of X-ray diffraction analysis confirmed a
- 545 decreased crystalline structure and increased Amorphous structure because of the addition of
- 546 gum Arabic.
- 547
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