Organocatalytic asymmetric biomimetic transamination of aromatic ketone to optically active amine

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Supporting Information

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All commercially available reagents were used without **General Methods.** further purification. Toluene distilled from sodium-benzophenone. was Dichloromethane was distilled from CaH₂. N,N-Dimethylformamide was dried over 4 Å molecular sieves (activated at 180 °C in vacuo over 8 h in vacuum). Column chromatography was performed on silica gel (200-300 mesh). ¹H NMR spectra were recorded on a 400 MHz NMR spectrometer and ¹³C NMR spectra were recorded on a 100 MHz NMR spectrometer. IR spectra were recorded on a FT-IR spectrometer. Melting points were uncorrected. o-HOPhCH₂NH₂ was prepared from 2-methoxybenzylamine through demethylation using BBr₃¹ and hydroquinine was prepared according to the reported procedure.²

(1) A. J. Hallett, G. J. Kwant and J. G. Vries, Chem.-Eur. J., 2009, 15, 2111.

(2) C. Palacio and S. J. Connon, Org. Lett., 2011, 13, 1298.

Preparation of Catalyst C4



To a solution of hydroquinine (2.0 g, 6.1 mmol) in DMF (20 mL) was added NaH (70% suspension in mineral oil) (0.627 g, 18.3 mmol). After the reaction mixture was stirred at rt for 1 h, 1-bromopentane (1.84 g, 12.2 mmol) was added in one portion. Upon stirring at 40 °C overnight, the reaction mixture was quenched with H₂O (20 mL), extracted with EtOAc (40 mL x 3), washed with brine (20 mL x 3), dried over MgSO₄, filtered, and concentrated to give a yellow oil, which was used directly for the next step.

To a suspension of NaH (70% suspension in mineral oil) (2.1 g, 61.3 mmol) (washed with *n*-hexane, dried in vacuo) in DMF (20 mL) was added EtSH (7.58 g, 122.0 mmol) dropwise at 0 $^{\circ}$ C under N₂ over 10 min. After the reaction mixture was

stirred at rt for 20 min, a solution of the crude product from the previous step in DMF (10 mL) was added in one portion. Upon stirring at 110 °C overnight, the reaction mixture was cooled to rt, acidified with concentrated HCl, washed with n-hexane (40 mL x 3), brought to pH = 10 with NH₄OH, extracted with EtOAc (60 mL x 3), washed with H₂O and brine, dried over MgSO₄, filtered, concentrated, and purified by flash column chromatography (silica gel, packed with EtOAc containing 1% Et₃N) (eluent: EtOAc/MeOH = 40/1 to 10/1) to give compound C4 as a yellow solid (1.26 g, 54% overall). mp. 82-85 °C; $[\alpha]^{20}_{D} = -66.8$ (c 0.99, CHCl₃); IR (film) 3072, 1618, 1464, 1241, 1118 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 11.38 (br s, 1H), 8.67 (d, J = 4.4 Hz, 1H), 8.23 (s, 1H), 7.99 (d, J = 9.2 Hz, 1H), 7.38 (d, J = 4.4 Hz, 1H),7.30 (dd, J = 9.2, 1.6 Hz, 1H), 5.34 (s, 1H), 3.71-3.56 (m, 1H), 3.30-3.16 (m, 3H), 3.05-2.80 (m, 2H), 2.42-2.26 (m, 1H), 2.12-1.98 (m, 1H), 1.98-1.86 (m, 1H), 1.86-1.78 (m, 1H), 1.66-1.48 (m, 4H), 1.45-1.22 (m, 5H), 1.22-1.08 (m, 2H), 0.88 (t, J = 7.2 Hz, 3H), 0.75 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.7, 146.4, 144.0, 143.5, 131.4, 128.0, 123.6, 117.8, 106.1, 78.7, 69.4, 59.3, 58.0, 43.5, 37.0, 29.8, 28.5, 27.5, 27.3, 25.4, 22.5, 19.7, 14.1, 11.9; HRMS Calcd for C₂₄H₃₅N₂O₂ (M+H): 383.2693; Found: 383.2686.

X. Xiao, Y. Xie, C. Su, M. Liu and Y. Shi, J. Am. Chem. Soc., 2011, 133, 12914.

Representative procedure for transamination of ketones (Table 2, entry 1).

To a Schlenk tube was added ketone **14a** (0.120 g, 1.00 mmol), *o*-HOPhCH₂NH₂ (0.185 g, 1.50 mmol), catalyst **C4** (0.077 g, 0.20 mmol), and toluene (5 mL). After stirring at 110 °C for 72 h, the reaction mixture was concentrated and subjected to flash column chromatography (silica gel, eluent: PE/EtOAc = 30/1) to remove *o*-HOPhCH₂NH₂, catalyst, and other byproducts. The resulting aldimine along with small amounts of ketone **14a** was dissolved in THF (1 mL) and 1N HCl (4 mL). Upon stirring at 20 °C for 24 h, the reaction mixture was washed with *n*-hexane (15 mL x 3), brought to pH > 7 with solid K₂CO₃, extracted with CH₂Cl₂ (30 mL x 3),

dried over MgSO₄, filtered, and concentrated to give amine **15a** as a yellow oil (0.080 g, 66%).

Preparation of *N*-benzoyl derivative of amine for the determination of the enantiomeric excess.

To a solution of amine **15a** (0.012 g, 0.10 mmol) in $CH_2Cl_2(1 \text{ mL})$ were added Et_3N (0.018 g, 0.18 mmol) and PhCOCl (0.018 g, 0.15 mmol). Upon stirring at rt for 30 min, the reaction mixture was purified by flash column chromatography (silica gel, eluent: PE/EtOAc = 8/1) to give *N*-benzoyl amine **16a** as a white solid (0.019 g, 82%). The sample was subjected to chiral HPLC (Chiralpak AD-H column) to determine the enantiomeric excess.

Table 2, entry 1



Yellow oil; $[\alpha]^{20}{}_{D} = -26.5 \ (c \ 0.33, \text{CHCl}_3) \ (80\% \ \text{ee});$ IR (film) 3442, 1629, 1453 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.28 (m, 4H), 7.26-7.19 (m, 1H), 4.10 (q, J = 6.8 Hz, 1H), 1.47 (br s, 2H), 1.38 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 148.0, 128.6, 126.9, 125.8, 51.5, 25.9.

- (1) Y. Chu, Z. Shan, D. Liu and N. Sun, J. Org. Chem., 2006, 71, 3998.
- (2) D. Guijarro, Ó. Pablo and M. Yus, J. Org. Chem., 2010, 75, 5265.

Table 2, entry 2



Light yellow oil; $[\alpha]_{D}^{20} = -23.1$ (*c* 1.08, CHCl₃) (84% ee); IR (film) 3360, 1584, 1489 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.43-7.38 (m, 1H), 7.23-7.16 (m, 1H),

7.15-7.08 (m, 1H), 7.04-6.97 (m, 1H), 4.38 (q, J = 6.4 Hz, 1H), 1.59 (br s, 2H), 1.42 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 161.8, 159.3, 134.7, 134.5, 128.3, 128.2, 126.89, 126.85, 124.40, 124.37, 115.7, 115.5, 45.59, 45.56, 24.2; HRMS Calcd for C₈H₁₁FN (M+H): 140.0870; Found: 140.0868.

L. M. Klingensmith, K. A. Nadeau and G. A. Moniz, *Tetrahedron Lett.*, 2007, 48, 4589.

Table 2, entry 3



Yellow oil; $[\alpha]^{20}{}_{\rm D} = -21.1 \ (c \ 1.19, \text{CHCl}_3) \ (71\% \text{ ee});$ IR (film) 3441, 1631, 1468 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.54-7.47 (m, 2H), 7.35-7.27 (m, 1H), 7.12-7.04 (m, 1H), 4.49 (q, J = 6.4 Hz, 1H), 1.76 (br s, 2H), 1.38 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 146.3, 133.1, 128.4, 128.0, 126.7, 123.3, 50.2, 23.9; HRMS Calcd for C₈H₁₁BrN (M+H): 200.0069; Found: 200.0069.

L. M. Klingensmith, K. A. Nadeau and G. A. Moniz, *Tetrahedron Lett.*, 2007, 48, 4589.

Table 2, entry 4



Yellow oil; $[\alpha]^{20}{}_{\rm D} = -19.7 (c \ 0.95, \text{CHCl}_3) (78\% \text{ ee});$ IR (film) 3374, 1594, 1567, 1474 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.50 (s, 1H), 7.35 (d, J = 7.6 Hz, 1H), 7.26 (d, J = 8.0 Hz, 1H), 7.21-7.14 (m, 1H), 4.12-4.03 (m, 1H), 1.66 (br s, 2H), 1.36 (d, J = 6.0 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 150.2, 130.2, 130.0, 129.1, 124.6, 122.7, 51.1, 25.8; HRMS Calcd for C₈H₁₁BrN (M+H): 200.0069; Found: 200.0065.

Table 2, entry 5



Yellow oil; $[\alpha]^{20}{}_{\rm D} = -29.2$ (*c* 1.15, CHCl₃) (82% ee); IR (film) 3441, 1608, 1450 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.26-7.17 (m, 1H), 7.17-7.09 (m, 2H), 7.07-7.01 (m, 1H), 4.06 (q, *J* = 6.8 Hz, 1H), 2.34 (s, 3H), 1.75 (br s, 2H), 1.37 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 147.8, 138.2, 128.5, 127.7, 126.6, 122.9, 51.4, 25.7, 21.6; HRMS Calcd for C₉H₁₃NNa (M+Na): 158.0940; Found: 158.0937.

M. Pallavicini, E. Valoti, L. Villa and O. Piccolo, *Tetrahedron: Asymmetry*, 2001, **12**, 1071.

Table 2, entry 6

Yellow oil; $[\alpha]^{20}{}_{D} = -26.2$ (*c* 0.99, CHCl₃) (79% ee); IR (film) 3361, 1592, 1492 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.30-7.26 (m, 4H), 4.10 (q, *J* = 6.4 Hz, 1H), 1.88 (br s, 2H), 1.36 (d, *J* = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 146.0, 132.6, 128.8, 127.4, 50.9, 25.7; HRMS Calcd for C₈H₁₁ClN (M+H): 156.0575; Found: 156.0571.

D. Guijarro, Ó. Pablo and M. Yus, J. Org. Chem., 2010, 75, 5265.

Table 2, entry 7



Yellow oil; $[\alpha]^{20}{}_{D} = -36.3$ (*c* 1.02, CHCl₃) (83% ee); IR (film) 3286, 1514, 1455 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.27-7.20 (m, 2H), 7.17-7.11 (m, 2H), 4.13-4.03 (m, 1H), 2.33 (s, 3H), 1.78 (br s, 2H), 1.37 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.0, 136.5, 129.3, 125.8, 51.2, 25.8, 21.2; HRMS Calcd for C₉H₁₃NNa (M+Na): 158.0940; Found: 158.0936.

D. R. J. Hose, M. F. Mahon, K. C. Molloy, T. Raynham and M. Wills, J. Chem. Soc., Perkin Trans. 1, 1996, 691.

Table 2, entry 8



Yellow oil; $[\alpha]^{20}{}_{D} = -23.2 \ (c \ 1.12, \text{CHCl}_3) \ (78\% \text{ ee});$ IR (film) 3290, 1512, 1456 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.24 (d, J = 8.0 Hz, 2H), 7.13 (d, J = 8.0 Hz, 2H), 4.07 (q, J = 6.4 Hz, 1H), 2.59 (t, J = 7.6 Hz, 2H), 1.72 (br s, 2H), 1.64-1.53 (m, 2H), 1.41-1.29 (m, 5H), 0.92 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.1, 141.6, 128.6, 125.7, 51.2, 35.4, 33.8, 25.7, 22.5, 14.1; HRMS Calcd for C₁₂H₂₀N (M+H): 178.1590; Found: 178.1590.

Table 2, entry 9



Yellow oil; $[\alpha]^{20}{}_{\rm D}$ = -20.0 (*c* 1.28, CHCl₃) (76% ee); IR (film) 3291, 1509, 1461 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.37-7.33 (m, 2H), 7.29-7.23 (m, 2H), 4.08 (q, J = 6.4 Hz, 1H), 1.73 (br s, 2H), 1.38 (d, J = 6.4 Hz, 3H), 1.31 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 149.8, 144.9, 125.50, 125.49, 51.1, 34.6, 31.5, 25.7; HRMS

Calcd for $C_{12}H_{20}N$ (M+H): 178.1590; Found: 178.1586.

Table 2, entry 10



Yellow oil; $[\alpha]^{20}{}_{D} = -25.5$ (*c* 1.17, CHCl₃) (85% ee); IR (film) 3291, 1580, 1505 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.15-7.08 (m, 1H), 7.04-6.97 (m, 2H), 4.08 (q, J = 6.4 Hz, 1H), 2.24 (d, J = 1.2 Hz, 3H), 1.83 (br s, 2H), 1.36 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 162.8, 160.4, 147.9, 131.6, 131.5, 123.3, 123.1, 121.20, 121.17, 112.5, 112.3, 50.9, 25.8, 14.39, 14.35; HRMS Calcd for C₉H₁₃FN (M+H): 154.1027; Found: 154.1023.

Table 2, entry 11



Yellow oil; $[\alpha]^{20}{}_{D} = -27.0 \ (c \ 1.21, CHCl_3) \ (83\% \ ee);$ IR (film) 3291, 1504, 1451 cm⁻¹; ¹H NMR (400 MHz, CDCl_3) δ 7.15-7.04 (m, 3H), 4.12-4.00 (m, 1H), 2.27 (s, 3H), 2.25 (s, 3H), 1.67 (br s, 2H), 1.38 (d, $J = 6.0 \ Hz$, 3H); ¹³C NMR (100 MHz, CDCl_3) δ 145.4, 136.8, 135.2, 129.9, 127.2, 123.2, 51.2, 25.8, 20.0, 19.5; HRMS Calcd for C₁₀H₁₅NNa (M+Na): 172.1097; Found: 172.1093.

Table 2, entry 12



Light red oil; $[\alpha]^{20}_{D} = -21.0 \ (c \ 0.98, CHCl_3) \ (81\% \ ee);$ IR (film) 3364, 3287, 1601, 1507, 1451 cm⁻¹; ¹H NMR (400 MHz, CDCl_3) δ 7.86-7.76 (m, 4H), 7.52-7.41 (m, 3H), 4.29 (q, J = 6.4 Hz, 1H), 1.74 (br s, 2H), 1.48 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 145.2, 133.7, 132.8, 128.4, 127.9, 127.8, 126.2, 125.7, 124.7, 123.9, 51.6, 25.7; HRMS Calcd for C₁₂H₁₃NNa (M+Na): 194.0940; Found: 194.0938.

D. Guijarro, Ó. Pablo and M. Yus, J. Org. Chem., 2010, 75, 5265.

Table 2, entry 13

NH₂ 15m

Yellow oil; $[\alpha]^{20}{}_{D} = -8.6 \ (c \ 0.79, \text{CHCl}_3) \ (70\% \text{ ee});$ IR (film) 3441, 1630, 1450, 1372, 1309 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.18-7.14 (m, 1H), 6.95-6.88 (m, 2H), 4.36 (q, J = 6.8 Hz, 1H), 1.73 (br s, 2H), 1.48 (d, J = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 153.0, 126.7, 123.5, 122.2, 47.5, 26.5.

D. Guijarro, Ó. Pablo and M. Yus, J. Org. Chem., 2010, 75, 5265.

Table 2, entry 14

Yellow oil; $[\alpha]^{20}{}_{D} = -14.5$ (*c* 0.60, CHCl₃) (77% ee); IR (film) 3418, 1628, 1454 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.36-7.28 (m, 4H), 7.26-7.20 (m, 1H), 3.80 (t, *J* = 6.4 Hz, 1H), 1.80-1.60 (m, 4H), 0.87 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 146.6, 128.6, 127.1, 126.6, 58.0, 32.6, 11.1; HRMS Calcd for C₉H₁₃NNa (M+Na): 158.0940; Found: 158.0937.

(1) Y. Chu, Z. Shan, D. Liu and N. Sun, J. Org. Chem., 2006, 71, 3998.

(2) D. Guijarro, Ó. Pablo and M. Yus, J. Org. Chem., 2010, 75, 5265.

The chromatograms for determination of enantioselectivity

 Table 2, entry 1



HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 2









HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 4









HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (90/10); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 6









HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 8





Table 2, entry 9



HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (90/10); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 10









HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 12





Table 2, entry 13

S HN-Bz 16m

HPLC Condition: Column: Chiralpak AD-H, Daicel Chemical Industries, Ltd.; **Eluent:** Hexanes/IPA (85/15); **Flow rate:** 0.5 mL/min; **Detection:** UV230 nm.



Table 2, entry 14





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S-17













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