

# Osteoclast Activity and Subtypes as a Function of Physiology and Pathology—Implications for Future Treatments of Osteoporosis

K. Henriksen, J. Bollerslev, V. Everts, and M. A. Karsdal

Nordic Bioscience A/S (K.H., M.A.K.), DK-2730 Herlev, Denmark; Section of Endocrinology (J.B.), Department of Medicine, Rikshospitalet, Oslo University Hospital and the University of Oslo, 0450 Oslo, Norway; and Department Oral Cell Biology (V.E.), Academic Centre of Dentistry Amsterdam, University of Amsterdam and VU University Amsterdam, Research Institute Move, 1066 EA Amsterdam, The Netherlands

Osteoclasts have traditionally been associated exclusively with catabolic functions that are a prerequisite for bone resorption. However, emerging data suggest that osteoclasts also carry out functions that are important for optimal bone formation and bone quality. Moreover, recent findings indicate that osteoclasts have different subtypes depending on their location, genotype, and possibly in response to drug intervention.

The aim of the current review is to describe the subtypes of osteoclasts in four different settings: 1) physiological, in relation to turnover of different bone types; 2) pathological, as exemplified by monogenomic disorders; 3) pathological, as identified by different disorders; and 4) in drug-induced situations.

The profiles of these subtypes strongly suggest that these osteoclasts belong to a heterogeneous cell population, namely, a diverse macrophage-associated cell type with bone catabolic and anabolic functions that are dependent on both local and systemic parameters. Further insight into these osteoclast subtypes may be important for understanding cell–cell communication in the bone microenvironment, treatment effects, and ultimately bone quality. (*Endocrine Reviews* 32: 31–63, 2011)

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## I. Introduction

Osteoclasts are multinucleated bone-resorbing cells that are unique in their ability to degrade mineralized matrices, such as bone and calcified cartilage (1). Osteoclasts have for a long time been considered bone-resorbing “machines,” yet some years ago it was demonstrated that not all osteoclasts are the same and that careful elucidation of the osteoclast subtype may prove

Abbreviations: AGE, Advanced glycation end-product; BMD, bone mineral density; CAII, carbonic anhydrase II; CIC-7, chloride channel 7; CT-1, cardiostrophin-1; CTX, C-terminal crosslinked telopeptide of type I collagen; ER, estrogen receptor; GLP, glucagon-like peptide; HRT, hormone replacement therapy; ICTP, carboxyterminal peptide of type I collagen; MMP, matrix metalloproteinase; ONJ, osteonecrosis of the jaw; OPG, osteoprotegerin; RA, rheumatoid arthritis; RANK, receptor activator of nuclear factor  $\kappa$ B; RANKL, RANK ligand; SERM, selective estrogen receptor modulator; TRACP, tartrate-resistant acid phosphatase; V-ATPase, vacuolar type ATPase.

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beneficial (1–4). As illustrated in Table 1, under normal circumstances, osteoclasts are influenced by a complex combination of systemic hormones and local mediators present in the different bones. Under pathological conditions, such as cessation of estrogen production or inflammatory conditions, additional cytokines are present. Even the bone type and age may influence the phenotype of the osteoclast (2, 5–7). Finally and importantly, different classes of drugs used for treatment of osteoporosis and other diseases also influence the osteoclasts significantly.

The aim of this review is to provide a thorough description of the complex nature of osteoclasts under healthy and diseased states and to describe their modulation by drugs that have been approved for use or are under development. The paper will also emphasize the role of osteoclasts in initiating bone formation, a recently discovered activity of these cells that has gained much attention (3, 8–13).

## II. Bone Remodeling

Bone remodeling is required for optimal control of calcium homeostasis and strength of the bones and is essential for the continued maintenance of a healthy skeleton (14). Bone remodeling is performed by three cell types: 1) the osteoclasts, which are the sole cells in the body possessing the ability to degrade both the inorganic calcium matrix and the organic collagen matrix; 2) the osteoblasts, which are the bone-forming cells; and 3) the osteocytes, which appear to regulate the activity of both osteoclasts and osteoblasts (14, 15). In healthy adults, under normal circumstances, bone resorption is always followed by an equal degree of bone formation, a tightly balanced process referred to as coupling (9, 16). The modulation of activities of the cells involved in the remodeling cycle was recently described in detail (15).

Coupling was initially discovered in the 1960s when Frost and co-workers (17, 18) demonstrated that osteoblasts filled the resorption pits created by osteoclasts in more than 97% of the cases (17–21). Since then, coupling has been understood as a coordinated and balanced induction of osteoblastic bone formation in response to prior bone resorption (19). Uncoupling occurs when the balance between resorption and formation is disrupted, which often leads to pathological situations such as osteoporosis or osteopetrosis (3, 9, 14, 22, 23). However, uncoupling also occurs under physiological conditions, *i.e.*, during skeletal growth in children, where bone formation exceeds bone resorption (10).

Hypogonadal osteoporosis is usually caused by a decrease or loss of sex steroid production, which results in accelerated osteoclastogenesis and bone resorption (24)

that cannot be completely countered by an increase in bone formation. This results in low bone mass, in deterioration of the microarchitecture of the skeleton, and often in fractures (25). Osteoporotic fractures are associated with increased morbidity and mortality and give rise to a significant public health problem (24).

Osteopetrosis, on the other hand, is a rare, inherited disease in various species including man, which was originally identified by Albers-Schönberg in 1904 (26). In the majority of cases, it is caused by defective resorption by the osteoclasts, resulting in high bone mass with poor bone quality and increased fracture frequency due to defective bone remodeling (1, 26, 27). However, recent studies also characterized patients with osteopetrosis due to dysfunctional osteoclastogenesis either directly affecting the osteoclast precursors or indirectly through the osteoblasts, and thus the phenotype was caused by the absence of osteoclasts, rather than inactivity of these cells (28–31). Interestingly, the studies of osteopetrotic patients have indicated that the presence of osteoclasts, but not their activity, is essential for bone formation, indicating that some aspects of the coupling principle should be revised (1, 3).



Because hypogonadal osteoporosis is associated with increased numbers and activity of osteoclasts (16), most treatments developed so far, such as bisphosphonates and hormone replacement therapy (HRT)/selective estrogen receptor modulators (SERMs), have focused on eliminating or reducing the number of osteoclasts and thereby reducing bone resorption (32). These treatments are associated with secondary decreases in bone formation due to the coupled nature of the bone remodeling process, which naturally limits their efficacy (3, 24). However, as seen in the osteopetrotic syndromes, there are indications that bone resorption and bone formation can be dissociated, and from recent studies it appears that the osteoclast itself, whether it is a physiological, pathological, or drug-induced subtype, is highly important for a secondary effect on bone formation (1, 3).

In this review, we describe differences in osteoclast activity and subtypes in relation to physiology, pathophysiology, and medication, with special attention to coupling in the bone remodeling process. Ultimately, this review highlights potential directions for new treatment modalities.

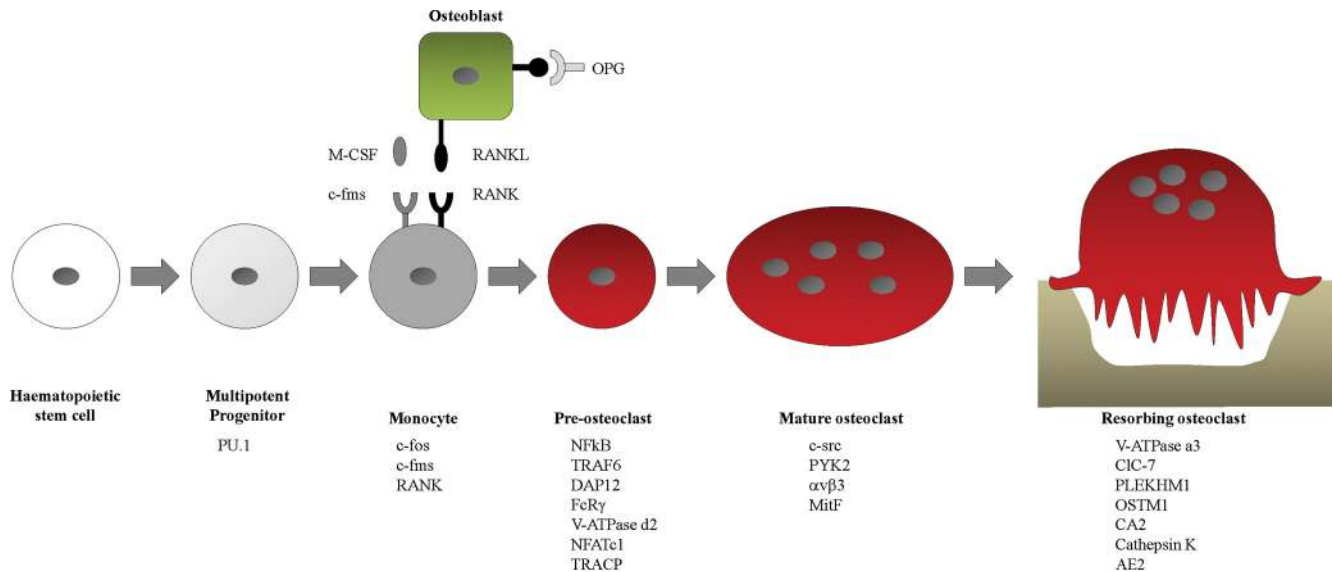
## III. The Classical Osteoclast

Osteoclastogenesis is a complex process requiring both the correct extracellular stimuli and the correct cellular molecules to interact without impediment (22, 33). Osteoclasts arise from hematopoietic stem cells that, in the presence of

**TABLE 1.** A simplified summary of osteoclast phenotypes as a function of physiology, pathology, and drugs, also indicating areas of osteoclast biology that are not well-understood

|                          | Osteoclast no.<br> | Bone resorption          | Resorptive process |       |     | Bone formation<br> |
|--------------------------|-----------------------------------------------------------------------------------------------------|--------------------------|--------------------|-------|-----|-------------------------------------------------------------------------------------------------------|
|                          |                                                                                                     |                          | Acid               | Cat K | MMP |                                                                                                       |
| Classical osteoclast     | Normal                                                                                              | Normal                   | +++                | +++   | +/- | Balanced                                                                                              |
| Physiology               |                                                                                                     |                          |                    |       |     |                                                                                                       |
| Targeted                 | Recruitment to specific areas increased                                                             | Increased                | ++                 | ?     | ?   | Balanced                                                                                              |
| Stochastic               | Not clear                                                                                           | Not clear                | +                  | ?     | ?   | Not clear                                                                                             |
| Night                    | Normal                                                                                              | Increased                | ++                 | ++    | +/- | Minor up-regulation                                                                                   |
| Day                      | Normal                                                                                              | Decreased                | +                  | ?     | ?   | Minor down-regulation                                                                                 |
| Chondroclast             | Normal                                                                                              | Normal                   | +                  | +     | ++  | Balanced                                                                                              |
| Endochondral             | Normal                                                                                              | Normal                   | +                  | +     | -   | Balanced                                                                                              |
| Intramembraneous         | Normal                                                                                              | Normal                   | +                  | -     | +   | Not balanced, opposite side of bone than resorption                                                   |
| Trabecular               | Normal                                                                                              | Normal                   | ++                 | ++    | -   | Balanced                                                                                              |
| Cortical                 | Normal                                                                                              | Normal                   | ++                 | ++    | -   | Balanced                                                                                              |
| Pathology                |                                                                                                     |                          |                    |       |     |                                                                                                       |
| Osteoporosis             | Increased                                                                                           | Increased                |                    | +++   | -   | Increased, but less than resorption                                                                   |
| Age                      | Increased                                                                                           | Increased                | ++                 | ++    | ?   | Decreased                                                                                             |
| OC-rich OP               | Greatly increased                                                                                   | Decreased                | -                  | -     | -   | Increased according to increased OC number                                                            |
| OC-poor OP               | No osteoclasts                                                                                      | Decreased                | -                  | -     | -   | Decreased?                                                                                            |
| Pycnodysostosis          | Unchanged/increased osteoclast size                                                                 | Decreased                | +                  | -     | +++ | Not clear                                                                                             |
| Paget's                  | Greatly increased at local sites                                                                    | Increased at local sites | ++                 | +     | +++ | Increased locally, but does not compensate resorption                                                 |
| RA                       | Greatly increased at local sites                                                                    | Not known                | +                  | +     | +++ | Not known                                                                                             |
| Lytic metastases         | Greatly increased at local sites                                                                    | Increased at local sites | +                  | +     | +++ | Increased locally, but does not compensate resorption                                                 |
| Drug-induced             |                                                                                                     |                          |                    |       |     |                                                                                                       |
| BPs                      | Reduced                                                                                             | Decreased                | -                  | -     | ?   | Decreased secondary to resorption                                                                     |
| HRT/SERMs                | Reduced                                                                                             | Decreased                | -                  | -     | ?   | Decreased secondary to resorption                                                                     |
| Calcitonin               | Unchanged                                                                                           | Decreased                | -                  | -     | ?   | Not changed or minor decrease                                                                         |
| PTH                      | Increased/unchanged                                                                                 | Decreased                | ++                 | ++    | ?   | Increased, but only temporarily                                                                       |
| Strontium ranelate       | Unchanged                                                                                           | Decreased?               | -                  | -     | ?   | Increased                                                                                             |
| GCs                      | Unchanged/increased                                                                                 | Unchanged/increased      | +                  | +     | ?   | Decreased strongly                                                                                    |
| Denosumab                | Greatly reduced                                                                                     | Decreased                | -                  | -     | -   | Decreased secondary to resorption                                                                     |
| Cat K inhibitors         | Unchanged                                                                                           | Decreased                | +                  | -     | +++ | Decreased secondary to resorption                                                                     |
| GLP-2                    | Unchanged                                                                                           | Decreased                | -                  | -     | ?   | Not changed, but long term effects are not known                                                      |
| Acidification inhibitors | Increased                                                                                           | Decreased                | -                  | -     | -   | Increased, but so far only in animal models                                                           |

The table shows the subtype of osteoclasts, the number of osteoclasts, the effect on bone resorption, which part of the resorption machinery that is active/affected, and the effect on bone formation. OC, Osteoclast; OP, osteoporosis; Cat K, cathepsin K; BPs, bisphosphonates; GCs, glucocorticoids; MMP, matrix metalloproteinase; RA, rheumatoid arthritis; PTH, parathyroid hormone; HRT, hormone replacement therapy; SERMs, selective estrogen receptor modulators; GLP-2, glucagon-like peptide 2.



**FIG. 1.** Schematic illustration of the molecules involved in osteoclastogenesis and function. (See Refs. 1, 33, and 42). NFκB, nuclear factor κ B; TRAF6, TNF receptor-associated factor 6; nuclear factor of activated T-cells, cytoplasmic, calcineurin-dependent 1; PYK2, Proline-rich Tyrosine Kinase 2; MitF, microphthalmia-associated transcription factor; CIC-7, Chloride Channel 7; PLEKHM1, pleckstrin homology domain containing, family M (with RUN domain) member 1; osteopetrosis associated transmembrane protein 1; CA2, carbonic anhydrase II; AE2, anion exchanger 2.

receptor activator of nuclear factor κB (RANK) ligand (RANKL) and macrophage-colony stimulating factor, undergo differentiation and fusion resulting in large multinucleated cells characterized by expression of a series of osteoclast markers, such as tartrate-resistant acid phosphatase (TRACP), matrix metalloproteinase (MMP)-9, cathepsin K, carbonic anhydrase II (CAII), the a3 subunit of the vacuolar [H<sup>+</sup>]-ATPase, chloride channel 7 (CIC-7), osteopetrosis-associated transmembrane protein 1, and the calcitonin receptor (34–41). Osteoclastogenesis and the molecules involved in this process are summarized in Fig. 1, but are not discussed in any further detail because several excellent reviews have been published recently on this topic (1, 22, 33, 42).

Polarization and formation of the sealing zone, which is a specialized ring structure containing a high number of β-actin filaments, are the next steps in the life span of an osteoclast (43, 44). These processes require the α<sub>v</sub>β<sub>3</sub> integrin and the intracellular signal transducers c-src, Syk, and proline-rich tyrosine kinase 2, as well as the microphthalmia transcription factor (33, 45, 46), which appears to be an important regulator of osteoclastic gene transcription (47–49).

The final step of osteoclastogenesis is the activation of resorption, a process that is characterized by the formation of a ruffled border that is an intensely convoluted membrane present inside the sealing zone (43, 44). The formation of the ruffled border is not well-characterized; however, the signaling molecule Rab7 is required (50).

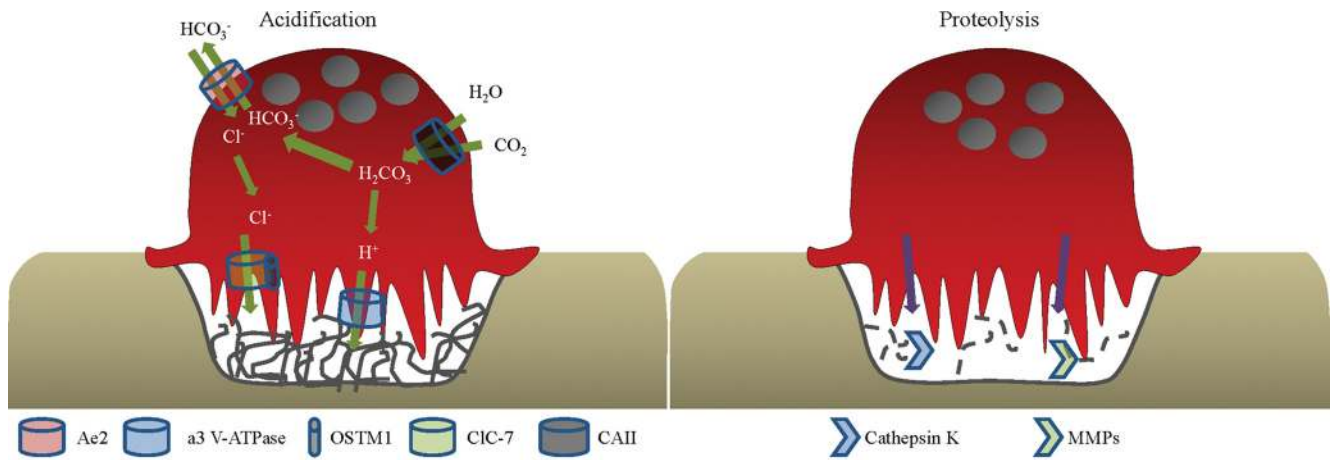
Bone resorption takes place at the ruffled border localized at the apical side of the osteoclasts, and it can be divided into two processes, namely acid secretion and pro-

teolysis, although these processes likely occur at the same time (44, 51).

Bone resorption is initiated by active secretion of protons through a vacuolar type ATPase (V-ATPase) and passive transport of chloride through a chloride channel (52, 53). The secretion of hydrochloric acid leads to dissolution of the inorganic matrix of the bones (54). The osteoclastic V-ATPase is functionally specific and contains the a3 subunit, and accordingly, loss of a3 leads to osteopetrosis (37, 55–57). In both mice and man, the chloride channel CIC-7, has been shown to mediate chloride transport, thereby ensuring the electrochemical balance required for intense acidification (Fig. 2) (8, 39, 58). Recent data showed that CIC-7 functions as a proton-chloride antiporter (59, 60).

To generate the necessary levels of H<sup>+</sup> and Cl<sup>-</sup>, the enzyme CAII catalyzes conversion of CO<sub>2</sub> and H<sub>2</sub>O into H<sub>2</sub>CO<sub>3</sub>, which ionizes into H<sup>+</sup> and HCO<sub>3</sub><sup>-</sup> (35), thereby providing the protons for the V-ATPase (27). Meanwhile, basolateral exchange of HCO<sub>3</sub><sup>-</sup> ions for Cl<sup>-</sup> by anion exchanger 2 (61–63) provides Cl<sup>-</sup> ions required for the intense acidification occurring in the resorption lacuna. Interestingly, long bones differ from flat bones with respect to the molecular nature of the acidification machinery (62).

Proteolysis of the type I collagen matrix in bones is mainly mediated by the cysteine proteinase, cathepsin K. This enzyme is active at low pH in the resorption lacuna (Fig. 2) (64–68). The neutral MMPs also appear to play a minor role during organic matrix degradation; however, the exact role of MMPs is still being investigated (69) and is highly dependent on the bone type (38, 70, 71). The resorbed material is removed from the resorption pit by



| Mutated gene/protein               | Phenotypes                                                | Osteoclast characteristics                                            |
|------------------------------------|-----------------------------------------------------------|-----------------------------------------------------------------------|
| <i>CICN7</i> / <i>CIC-7</i>        | ADOII, IARO, ARO<br>Severe osteopetrosis in mice          | Decreased acid secretion<br>Low resorption<br>High numbers            |
| <i>TCIRG1</i> / $\alpha 3$ subunit | ARO<br>Severe osteopetrosis in mice                       | Decreased acid secretion<br>Low resorption<br>High numbers            |
| <i>OSTM1</i> / OSTM1               | ARO                                                       | Low resorption<br>High numbers                                        |
| <i>CA2</i> / Carbonic anhydrase II | IARO<br>Moderate osteopetrosis in mice                    | Altered internal pH regulation<br>Low resorption<br>Increased numbers |
| <i>SLC4A2</i> / Ae2                | No human phenotype known<br>Severe osteopetrosis in mice  | Altered internal pH regulation<br>Low resorption<br>Normal numbers    |
| <i>CSTK</i> / Cathepsin K          | Pycnodysostosis<br>Osteopetrosis-like disease in mice     | Defective collagenolysis<br>Low resorption<br>Normal numbers          |
| <i>MMP-9</i> / MMP-9               | No human phenotype known<br>Delayed long bone development | Normal resorption<br>Normal numbers                                   |

For references see text

**FIG. 2.** Top, Schematic illustration of the differences between acid secretion and proteolysis during osteoclastic bone resorption, illustrating that the collagen matrix is removed by proteolysis after acidification. Bottom, Mutations/knockout in genes/proteins involved in bone resorption, phenotypes, and effect on osteoclasts. Ae2, Anion exchanger 2;  $\alpha 3$  V-ATPase,  $\alpha 3$  subunit of the osteoclast-specific V-ATPase. CIC-7, chloride channel 7; OSTM1, osteopetrosis associated transmembrane protein 1; CA2, carbonic anhydrase 2; MMP-9, matrix metalloproteinase 9.

uptake and transcytosis through the osteoclast (72, 73). After completing resorption, osteoclasts either undergo apoptosis or perform a further round of resorption (44) (Table 1).

In summary, the osteoclasts are highly specialized for both dissolution of the inorganic matrix and degradation of the organic matrix of the bones. These highly polarized cells are characterized by a unique set of membrane-bound molecules that ensure an efficient resorption of bone and other mineralized tissues. This complex machinery may be affected by a range of important parameters in physiology and pathology, and importantly in drug-induced situations that are important to identify and advance osteoclast research and biology.

#### IV. Osteoclast Subtypes in Physiological Situations

Osteoclast activities are essential for development, as well as remodeling of bone in response to aging and stress (6, 14, 15). Under normal physiological circumstances, the osteoclasts can be categorized into subgroups depending on the matrix on which they are positioned, the time of day, and the type of remodeling in which they participate. These different groups of osteoclasts have provided key information on skeletal maintenance.

##### A. Endochondral vs. intramembranous bone osteoclasts

Anatomically, two types of bones are present in the body, the long bones (e.g., the femur and tibia) and the

flat bones (*e.g.*, the calvarium), with the main difference between these two types of bone being their development (74). Studies also indicate that osteoclasts on these two types of bones are functionally different with respect to both acid secretion and proteases involved in degradation (2).

Evidence that differences between resorption of flat and long bone exist was presented in 1999 (4). However, indications that even the acidification process is different have been published only recently (62). Data from mice deficient in the bicarbonate-anion exchanger *Ae2* (*Slc4a2*) have shown that it is essential for bone resorption in long bones (61, 63), whereas it is not involved in bone resorption in calvariae (62), showing that distinct acid transport mechanisms are present in different subsets of osteoclasts. With respect to acid secretion into the resorption lacunae, it is presently not known whether any differences exist, although the absence of calvarial thickening in patients with defective *Clc-7* strongly suggests that *Clc-7* is not involved in resorption of the flat bones (75).

Extensive research into the proteolytic processes involved in resorption of flat and long bones clearly demonstrates that the proteolytic processes involved in degradation of these two types of bone matrix are distinct (4, 71). Osteoclasts in flat bones preferentially appear to engage in MMP-mediated bone resorption, although cathepsin L seems to be involved, too. Osteoclasts in long bones primarily depend on cysteine proteinases, in particular cathepsin K (4, 71). TRACP also appears to be involved in bone resorption, and more so in calvarial bone (76–78). When osteoclasts generated from human peripheral blood are seeded on cortical bone, they primarily depend on cathepsin K, whereas when cathepsin K activity is blocked, there is some compensatory bone resorption mediated by MMPs (69). How these osteoclasts behave on bone substrates other than cortical bone is presently not known (Table 1).

Data suggesting that the bone matrix could play a role in the control of osteoclastic activities were presented in a study showing compositional differences between long bone and flat bone matrices, including differences in the presence of putative cysteine proteinase inhibitors (79). The functional significance of these data still remains to be fully elucidated, although they clearly illustrate the importance of understanding how a given context affects the osteoclasts.

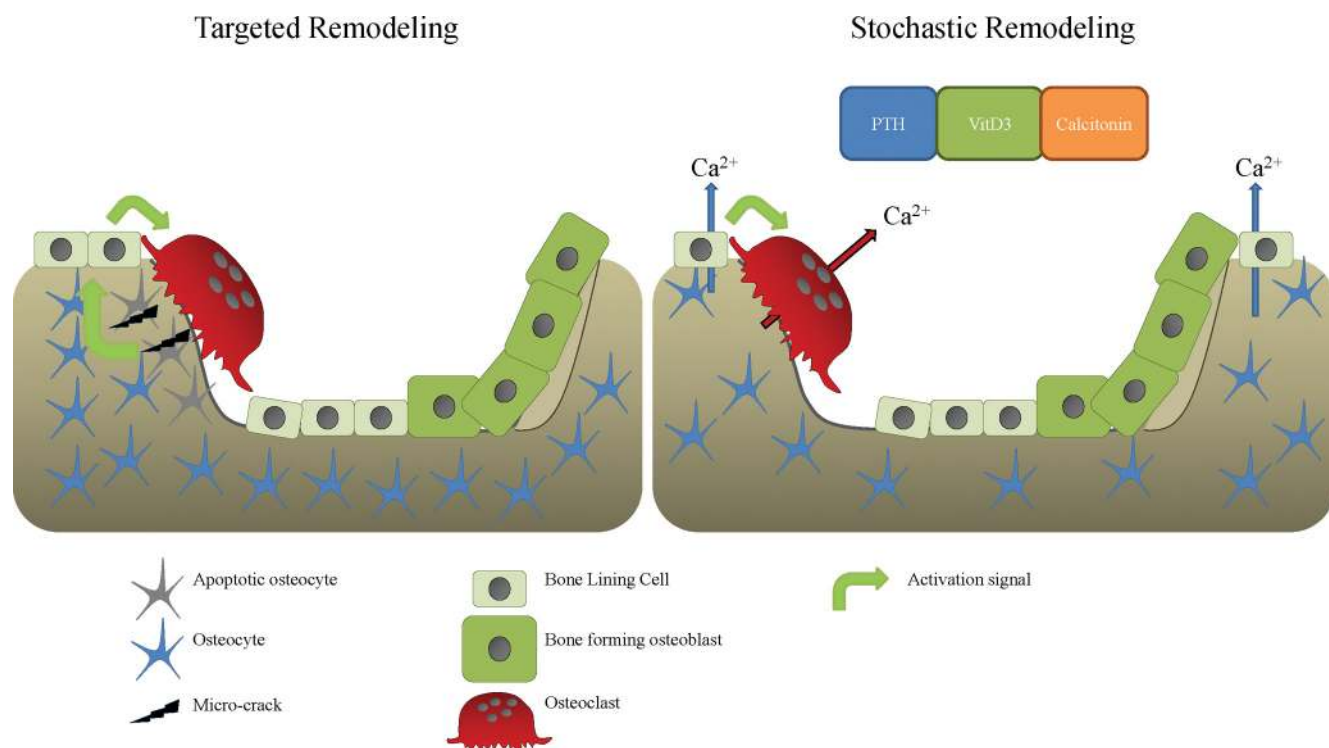
## B. Chondroclasts

It has long been discussed whether chondroclasts are a “real” cell type or whether they simply are osteoclasts that reside on cartilage instead of bone (77, 80–82). Chondroclasts are mainly important in endochondral bone development, and in addition there is some evidence that

chondroclasts may also play a role in both rheumatoid arthritis (RA) and osteoarthritis (83–85). The term chondroclast derives from the localization of these cells on calcified cartilage as seen in the expanding growth plates during endochondral ossification (80, 86). For their formation, these cells are dependent on the presence of macrophage-colony stimulating factor and RANKL, as are bone-resorbing osteoclasts (87–90).

Most of the evidence for the functionality of chondroclasts is derived from studies of the longitudinal growth of long bones, *i.e.*, metatarsals and tibiae isolated from mouse embryos (38, 80, 91–93). First, bone/cartilage resorption in these models is still dependent on acid secretion as evidenced by mice with mutations in the acid secretion process, *i.e.*, *Clc-7*-deficient and  $\alpha 3$  V-ATPase-deficient mice, as well as mice unable to form sealing zones, *i.e.*, *c-src*-deficient mice, in which the massive bones mainly consist of calcified cartilage due to the defective resorption process (56, 94, 95). Similar findings have been noted in the corresponding human disease(s) (27, 96, 97). Interestingly, the ruffled border is less prominent in the chondroclasts than osteoclasts, potentially suggesting that lower levels of acid secretion are required for dissolution of this matrix (98, 99). The main difference between chondroclasts and osteoclasts is in the profiles of enzymes necessary for tissue degradation. Resorption by chondroclasts does not appear to depend as much on cathepsin K as does resorption by osteoclasts. Cathepsin K-deficient mice show no evidence of calcified cartilage in the marrow cavity of long bones, indicating that the removal of calcified cartilage during endochondral ossification occurs, although there are indications that the process is delayed (65, 100). Of importance is the observation of a massive compensation by MMPs in the absence of cathepsin K, which obscures the interpretation of data from cathepsin K-deficient systems (69, 70, 101–103). Finally, one study has indicated that TRACP is activated and secreted at the ruffled border in cathepsin K-deficient osteoclasts, and only in cells that play a role in the removal of calcified cartilage (104) (Table 1).

Although chondroclasts are involved in the degradation of a different matrix than osteoclasts, an interesting observation is that bone formation is tightly coupled to resorption of the mineralized matrix, as has clearly been demonstrated in studies of endochondral ossification (86). It is more likely that bone formation is coupled to chondroclast numbers because release of molecules from degradation of cartilage, which in composition is far from bone, would be expected to be different from molecules released during bone resorption; however, this has never been studied in detail.



**FIG. 3.** Schematic illustration of stochastic vs. targeted remodeling. The figure illustrates the local nature of targeted remodeling, which is activated at specific sites after the formation of microcracks and leads to removal of the microcrack and restoration of the damaged bone. Stochastic remodeling, on the other hand, is of a systemic nature and is activated by low calcium levels in the circulation leading to PTH release. Other hormones, such as vitamin D3 (VitD3), and potentially calcitonin also play roles in stochastic remodeling. It appears that there are two levels of calcium homeostasis—one mediated by osteocytes independent of osteoclasts, and one including the osteoclasts—although the balance between these two ways of releasing calcium still remains to be fully understood.

### C. Osteoclasts involved in targeted and stochastic remodeling

Two different modes of remodeling have been proposed: targeted and stochastic. Targeted bone remodeling takes place at specific sites, whereas stochastic remodeling occurs more randomly (19, 105, 106). The first type is primarily performed to replace microdamaged bone and thus to maintain the load-bearing capacity of the skeleton. The second type of remodeling appears random with respect to localization, although it may be involved in maintaining integrity of the bones, independent of damage. This process is hormonally regulated (105, 107).

The balance between these two modes of bone remodeling has not been fully elucidated yet, but studies in dogs indicate that approximately 30% of all remodeling is targeted and the remaining 70% is stochastic (105). With respect to the osteoclasts mediating these two types of remodeling, most studies have focused on how targeted remodeling is controlled.

It appears that the bone matrix contains signals regulating the activity of the osteoclasts (108). A recent study demonstrated that aged bones were more readily resorbed than young bones, thus supporting the hypothesis that the bone matrix composition influences remodeling rates (7). Furthermore, areas of microdamage, which are character-

ized by high numbers of apoptotic osteocytes, are preferentially and rapidly degraded by osteoclasts. This further supports the possibility that changes in the bone matrix and the balance between live and dead osteocytes determine which areas will be remodeled (108). Finally, a recent study demonstrated that targeted ablation of osteocytes led to a dramatic increase in osteoclast activity (109) (Table 1). These data indicate that death of osteocytes is a key point in induction of osteoclast activity (Fig. 3).

Taken together with the finding that bones of different age lead to different levels of osteoclastogenesis (7), it appears that osteoclast functionality is at least partially controlled by osteocyte-derived molecules, which are sequestered in the bone matrix.

Stochastic remodeling, while occurring at random sites, is centrally regulated by hormones such as PTH, vitamin D3, and potentially calcitonin, and its main role is the regulation of calcium homeostasis (110–112). It has even been questioned to what extent this process depends on the presence of osteoclasts because patients with nonfunctional osteoclasts have normal calcium homeostasis (110–112). Yet, when they are calcium-deprived, osteopetrotic patients fail to correct their calcium levels, indicating that osteoclasts, which are either absent or nonfunctional in

these patients, do play a role in calcium homeostasis, and thus stochastic remodeling (Fig. 3) (113).

In summary, targeted remodeling is beginning to be understood in detail, and it is a tightly regulated and coupled process involving osteocytes, osteoclasts, and cells of the osteoblast lineage. On the other hand, stochastic remodeling and the role it plays in calcium homeostasis are still not very well understood, although there are indications that there is a level of regulation by the activity of osteoclasts.

#### D. Trabecular and cortical osteoclasts

Bone remodeling does not occur with the same frequency in cortical and trabecular bone. Every year, 25% of the trabecular bone matrix, but only 4% of the cortical matrix, is remodeled (114). Interestingly, most *in vitro* osteoclast experiments are based on cortical bone (or dentine) substrates, which are either slowly remodeled or not remodeled at all (7, 102, 114–116). Studies have shown that bones endogenously contain signals regulating osteoclastogenesis and resorption and that these signals appear to be related to the age of the bone (7, 117) (Table 1). Thus, an interesting question is whether osteoclasts themselves are indeed different when derived from different matrices or whether the difference is matrix related. Furthermore, systemic regulation is likely to be involved in controlling which bones are resorbed to some extent.

These data also correlate with evidence indicating that remodeling of different bone compartments can be either primarily targeted, such as in the cortex, or primarily stochastic, as seen in some parts of trabecular bone (106). A further understanding of this could provide directions for the development of novel drugs producing optimal benefit at the sites where it is most needed, *i.e.*, leading to a better fracture reduction than that presently obtained.

#### E. Diurnal variation in osteoclasts or osteoclast activity?

Bone resorption markers measured in serum may be interpreted as indicating the net result of all osteoclast subtypes and activity levels at one particular time. A wide range of factors, known and unknown, may influence the interpretation (15). Diurnal variation is a well-established and important parameter of bone turnover. Postprandially, bone resorption decreases by approximately 50% compared with that of fasting individuals, but during the night, bone resorption increases to an equally large degree (118–120). Several investigations have demonstrated that the circadian variation in bone resorption is induced in part by food intake (121–123), which, at least partially, involves the peptide hormone glucagon-like peptide (GLP) 2 (124). Interestingly, the osteoclast number does not appear to depend on the time of day, further emphasizing differences between osteoclast

number and activity (32) (Table 1). An interesting aspect of this is that targeting nocturnal resorptive activity appears to lead to inhibition of bone resorption, whereas not attenuating bone formation (125–127), thereby highlighting an interesting prospect of reducing bone resorption in a specific, nocturnal manner.

In summary, studies of osteoclasts under different physiological conditions, such as those listed above, have highlighted the heterogeneity of these cells. Furthermore, these studies highlighted the importance of the balance between bone resorption and bone formation, a tightly regulated phenomenon that rarely is disturbed under physiological conditions. Finally, how the heterogeneity of the osteoclasts affects bone formation is presently not well understood, but a further understanding of this process could help optimal treatment of diseases involving alterations in bone remodeling.

### V. Osteoclast Subtypes in Pathological Situations

Changes in osteoclast activity and number have been detected in several diseases, ranging from illnesses involving excessive bone resorption, such as osteoporosis and Paget's disease; to those involving secondary activation of osteoclasts, such as osteolytic metastases and RA; to diseases involving defective osteoclast differentiation and/or function, such as osteopetrosis. These different types of diseases have shed important light on osteoclastic function with respect to obtaining the right type of treatment. They have also shed light on a very central aspect in bone biology, the coupling principle. The coupling principle describes the phenomenon that bone formation follows bone resorption, which leads to a complete restoration of the bone removed during bone resorption (17).

#### A. Osteoporotic osteoclasts

##### 1. Changes in osteoclastogenic potential in osteoporosis

An important aspect of osteoporosis is whether the number of osteoclast precursors in the circulation increases, and, if so, whether the osteoclastogenic potential of these cells is increased. Eghbali-Fatourehchi *et al.* (128) showed that the overall number of cells expressing RANKL is increased in postmenopausal women compared with premenopausal or estrogen-treated women, clearly indicating that the bone marrow microenvironment, including stromal, T, and B cells, changes in a proosteoclastic direction when estrogen is reduced. These data were supported by a recent study from the same authors showing that bone marrow cells isolated from estrogen-treated or control postmenopausal women displayed



reduced osteoclastogenic potential (129) (Table 1). Estrogen was shown to have a dual mode of action—the first leading to overall lower RANKL expression by bone marrow cells, and the second reducing the osteoclastogenic response to RANKL (130). Interestingly, aging of mice was also shown to increase the osteoclastogenic potential of bone marrow cells, both by up-regulation of RANKL production and by increasing precursor sensitivity to RANKL (131).

*In vitro* studies of the changes in cellular activity of osteoclasts from osteoporosis patients are limited, but these have indicated both an accelerated osteoclastogenesis and resorption (132, 133). Furthermore, a key cell in the up-regulation of osteoclastogenesis appears to be the T cell, which responds to lowered estrogen levels by increasing RANKL production (134).

The main issue with all the studies of osteoporotic osteoclasts and their precursors is the use of mixed cell populations, which clouds the interpretation of the results, and therefore these aspects of osteoclastic function still require further investigation. Furthermore, with the recent publication of the possibility of assessing the “anabolic potential” of osteoclasts (11, 13), it would be of interest to investigate the anabolic capacity of osteoporotic osteoclasts and thus shed light on the imbalance between bone resorption and bone formation in osteoporosis.

## 2. A direct role for sex steroids on osteoclasts

The role of estrogen on cells belonging to the osteoclastic lineage has been studied extensively, with several findings indicating that estrogen suppresses osteoclastogenesis but not the resorptive activity of mature osteoclasts (135–137). Androgens, such as dihydrotestosterone, exhibit similar effects to estrogen on osteoclasts *in vitro*, although this has not been studied in great detail (138). Finally, a recent study using mice deficient for the estrogen receptor (ER)- $\alpha$ , specifically in mature osteoclasts, showed bone loss in female, but not male, mice (139). This demonstrated that estrogen likely plays a direct role in bone resorption by even mature osteoclasts (139). Although the authors used the cathepsin K promoter to ensure specific knock-down of the ER- $\alpha$  in osteoclasts, cathepsin K is also expressed in preosteoclasts, albeit to a lower extent (139). More studies are needed to investigate the role of ER- $\alpha$  in mature osteoclasts specifically. Interestingly, osteoclasts in different bone sites preferentially express different ERs, with cortical osteoclasts mainly expressing ER- $\alpha$  and trabecular osteoclasts mainly expressing ER- $\beta$  (140), whereas from a functional point of view ER- $\alpha$  appears to be more relevant for trabecular, not cortical, bone (141). Furthermore, the expression pattern also differs between mature and differentiating osteoclasts; ER- $\alpha$  is mainly expressed in immature cells, and ER- $\beta$  is present at all stages

of osteoclastogenesis (137). Again, there appear to be different osteoclastic subtypes, which also appear to be relevant in the context of bone loss rates in different bone compartments during osteoporosis (114). As indicated above, an important point is the difference between genders (139). In mice, the gender-based difference between cortical apposition and endocortical resorption that becomes more apparent with increasing age might be explained by differences in ER expression (142). With respect to changes in the osteoclasts after menopause, a couple of studies have clearly demonstrated that bone resorption, as well as bone formation, increases in women after menopause (143–145), and these changes become more explicit in high- and low-turnover patients (145). Although bone formation increases as a function of the increased resorption, it does not match bone resorption, thereby illustrating the importance of understanding the interplay between osteoclasts and osteoblasts in detail.

## B. Changes in osteoclast activities with increasing bone matrix age

Numerous studies have investigated the control of osteoclast activity as a function of changes in biochemical properties of the bone matrix. Aging leads to accumulation of different biochemical modifications of the bone matrix, such as advanced glycation end-products (AGEs), homocysteine, increased calcium concentration, as well as some modifications of the collagen matrix (146).

Recent studies have indicated that these modifications of the bone matrix itself actually modulate the activity of the osteoclasts to a certain extent (7, 117). Homocysteine, which accumulates in bone and in circulation with age, was shown to activate osteoclastogenesis and bone resorption (147) (Table 1). AGEs are modifications of proteins that accumulate in various tissues with age, and they have been implicated in the pathology of osteoporosis (146, 148). Some evidence indicating a direct regulation of osteoclast activity by AGEs has been published, but these studies are contradictory. One study shows activation of resorption by AGE-modified proteins (149), whereas the other study shows the opposite (117); however, quite different techniques were used.

Interestingly, AGEs are accumulated in diabetes, and they have been speculated to be involved in the increased fracture rates observed in patients with this disease (150, 151). Another intriguing finding is the induction of apoptosis in osteoblasts by AGEs (152), which potentially could play a role in the imbalance between osteoclast and osteoblast function during osteoporosis and aging. These findings are all preliminary in nature, and they await confirmation from independent research groups. However, once again they illustrate the heterogeneity of osteoclasts,

in this case as a function of matrix age, and the importance of understanding this phenomenon.

### C. Osteoclast-rich osteopetrosis

The most frequently occurring forms of osteopetrosis are those caused by mutations in either the  $\alpha 3$  subunit of the V-ATPase, *CLC-7*, or osteopetrosis-associated transmembrane protein 1. Osteoclasts from patients with mutations in these genes or proteins and from knockout mice have been studied quite extensively (8, 39, 56, 58, 95, 153–156).

Microscopic analyses of cells from patients with defective acid secretion by osteoclasts indicated defective ruffled border formation, but also accumulation of material in vesicles, indicating hampered transcytosis (157). Apart from confirming the defective acid secretion and thereby bone resorption, when either *CLC-7* or the  $\alpha 3$  subunit is mutated (8, 39, 56, 58, 95, 153, 154), these studies also shed light on important aspects of bone remodeling.

*In vitro* studies indicate that osteoclasts with impaired acid secretion have higher survival rates than cells with normal secretion, due to the reduced release of proapoptotic signals during resorption (159). This observation correlates well with the high numbers of osteoclasts observed *in vivo* in this group of patients, as well as with findings in mice with attenuated acidification of the resorption lacunae (95, 97) (Table 1). Furthermore, significantly increased resorbed areas are seen during impaired acid secretion, but the resorption pits are shallow, indicating a disturbed activity of the osteoclasts (97). More importantly, these studies highlighted that bone formation in these patients is ongoing—a process that appears to be correlated to the increased number of osteoclasts rather than bone resorption (96, 154, 160, 161). These findings contrast with the classical perception that bone formation always follows bone resorption in a tightly coordinated manner and illustrate the importance of the actual presence of osteoclasts to maintain bone formation.

### D. Osteoclast-poor osteopetrosis

Several murine forms of osteoclast-poor osteopetrosis have been described in the literature (1, 42). In general, whereas the mutations express a pronounced osteopetrotic phenotype and few or no osteoclasts are present, the phenotypes are less severe than the phenotypes of the different osteoclast-rich osteopetrotic mutations (1). These data strongly suggest that the osteoclasts are indeed involved in the production of anabolic signals for bone formation (3, 15).

Studies of mice deficient in *c-src* and *c-fos*, a key molecule involved in ruffled border formation and a key signal transducer for osteoclastogenesis, clearly demonstrated

that osteopetrosis was due to nonfunctional osteoclasts or the absence of osteoclasts, respectively (162, 163). Interestingly, these two groups of mice have opposing phenotypes with respect to bone formation. The osteoclast-rich *c-src* knockouts have increased bone formation (164), and the osteoclast-poor *c-fos* knockouts have decreased bone formation (165). The anabolic effects of PTH are present in the *c-src*<sup>-/-</sup> mice but are blunted in the *c-fos*<sup>-/-</sup> mice (166), indicating that osteoclasts are central for bone formation (Table 1).

Two recent studies identified mutations in the genes for RANK and RANKL as the causes of osteopetrosis in a novel group of patients (28, 29). No indications of osteoclasts were found in these patients (28, 29), which is consistent with previous observations in mice deficient in both RANKL and RANK (87, 90). Patients with mutations in either RANKL or RANK have a pronounced osteopetrotic phenotype and classical histological hallmarks of osteopetrosis including unresorbed primary spongiosa. However, although limited data have been published, the osteopetrotic phenotype appears to be less severe than the one observed in the osteoclast-rich forms (1). Thus, mutations within the RANK/RANKL/osteoprotegerin (OPG) system can lead to osteoclast-poor osteopetrosis with low bone formation in mice and men.

Interestingly, alterations in osteoblast function, such as changes in the production of RANKL and OPG, may have the same effect. Stabilizing osteoblastic  $\beta$ -catenin in transgenic mice, thus mimicking constitutive activation of the canonical Wnt signaling pathway, was followed by an up-regulation of OPG in relation to RANKL (31, 167). As expected, the mice developed osteoclast-poor osteopetrosis with failure of tooth eruption, a classical phenomenon in murine osteopetrosis. Mutations within *LRP5* related to the Wnt signaling pathway have underscored the fundamental importance of this pathway for regulation of bone mass. The osteoporosis pseudoglioma syndrome was found to be caused by loss of function mutations in the gene for *LRP5* (168). In contrast, mutations affecting the first propeller of the coreceptor, presumed to be followed by chronic activation of the Wnt pathway, were found in various forms of monogenic human osteosclerotic phenotypes (169). Among these, autosomal dominant osteopetrosis type 1 has been well characterized clinically, biochemically, histomorphometrically, and biomechanically (75). Autosomal dominant osteopetrosis type 1 is an osteoclast-poor osteopetrotic phenotype with increased biomechanical competence and no low-energy fractures. Osteoclast profiles are markedly decreased (97), bone formation seems to be normal, and OPG levels in the circulation increased (170). However, when investigating os-

teoclasts *ex vivo* from these patients, they express normal bone resorptive capacity (30).

In summary, osteoclast-poor osteopetrosis can arise in murine mutations/transgenics or humans when the OPG/RANKL/RANK system is affected directly or indirectly. These findings underscore this cytokine system as a key regulator of osteoclastogenesis. Moreover, the phenotypes seem to be less affected than the osteoclast-rich forms, the reason for which is so far unresolved, although there are indications that reductions in bone formation are involved (1, 28, 29).

### E. Pycnodysostotic osteoclasts

An interesting subtype of osteoclasts with defective bone resorption is observed in patients with pycnodysostosis. Pycnodysostosis is caused by loss of function or loss of expression mutations in the cysteine proteinase cathepsin K, which in humans causes dwarfism and poor bone quality due to defective remodeling of the bones (36, 171–173). Few studies examining the phenotype of pycnodysostotic osteoclasts have been published. Microscopic analyses of the osteoclasts have shown significantly increased amounts of demineralized collagen matrix in the resorption pit, but also inside the osteoclasts, indicating disturbed resorption and trafficking of resorbed components (174, 175). A study of biochemical markers of bone turnover showed that C-terminal crosslinked telopeptide of type I collagen (CTX-I) release was reduced, whereas production of the MMP-generated type I collagen fragment carboxyterminal telopeptide of type I collagen (ICTP) was increased (67) (Table 1). Several studies of cathepsin K-deficient mice have been published, and whereas they confirm that cathepsin K is essential for degradation of the organic matrix in bone (65, 66, 176), there are also several differences between the human and mouse phenotypes (103). Furthermore, in cathepsin K-deficient mice, bone formation parameters are highly increased (100). These findings have not been replicated in pycnodysostosis patients in whom the bone matrix is disordered (177), and a clinical case study indicated that anabolic response to PTH was absent (174). Two recently published clinical studies have shown that whereas bone resorption markers are strongly reduced, bone formation is also suppressed in women treated with the cathepsin K inhibitor odanacatib (178). In a monkey study monitoring bone formation by histomorphometry reductions in bone formation, rates were shown in the trabecular bone compartment, whereas bone formation was increased in the cortical compartment (179–181).

In conclusion, cathepsin K mediates cleavage of type I collagen in the resorption lacunae, but its secondary effects on bone formation are bone type-dependent and still need to be investigated further.

### F. Other diseases characterized by increased osteoclast activity

Apart from hypogonadal osteoporosis, several diseases are characterized by accelerated osteoclastogenesis and function. Although the etiology of these diseases is different, there are interesting overlaps and discrepancies that provide highly useful information about osteoclastic function and secondary effects on bone formation under different circumstances (182).

#### 1. Pagetic osteoclasts

Paget's disease is a late-onset disease that is quite common in the elderly Caucasian population, where it affects approximately 3% of individuals (182). The disease is characterized by focal increases in osteoclast numbers, nuclearity, and size, which leads to localized bone destruction, although surrounding osteoblasts also are activated (183) (Table 1). The identified causes of the disease include mutations in four different genes, *TNFRSF11A*, *TNFRSF11B*, *VCP*, and *SQSTM1* (182, 184–186). These genes encode RANK, OPG, p97, and p62, all of which are involved in the regulation of osteoclastogenesis. The mutations all result in different subtypes of Paget's (182, 184–186). These mutations render the osteoclast precursors more sensitive to RANKL stimulation, resulting in a higher number of osteoclasts, and potentially also explaining the presence of giant osteoclasts (185, 187, 188). Interestingly, a recent study in mice indicated that the most common mutation in p62 does not make the osteoclasts Pagetic alone, although it sensitizes them to other yet-to-be-described causes of Paget's (189, 190).

In Paget's patients, biochemical markers of both bone resorption and bone formation are increased, showing an overall increase in bone turnover at the affected sites. However, bone resorption clearly exceeds bone formation (182). Whether the osteoclasts in Paget's behave differently from those in healthy individuals during bone resorption is presently unknown. In particular, it is not known whether osteoclasts in Paget's require acidification to resorb bone, or whether cathepsin K is the main protease, although answers to these questions might be of value in the development of new therapies for Paget's. Furthermore, an explanation for the localized nature of Pagetic lesions has still not been found. Even under the extreme circumstances seen in Pagetic lesions, bone formation is coupled to osteoclastic parameters, although whether this is due to increased osteoclast numbers or activities is not known. Moreover, in this case a treatment type eliminating the activity of both types of cells is most likely to be preferred because the increase in bone formation occurring is part of the pathology, and likely will provide no benefit for the bones if maintained. Thus, bisphosphonate, which strongly attenuated overall bone

turnover, appears to be highly relevant in the context of Paget's disease (191).

## 2. Osteolytic osteoclasts

Several forms of cancer can metastasize to bone and form osteolytic metastases (192–196). Once the cancer has reached the bone, tumor and bone interact in a vicious cycle in which tumor-secreted factors, such as PTHrP, stimulate bone cells, which in turn release growth factors and cytokines that promote further tumor cell growth (192, 197). The activation of osteoclastogenesis induced by tumor cells has been shown to involve a switch in the RANKL/OPG ratio favoring osteoclastogenesis and activation, leading to release of the tumorigenic factor TGF- $\beta$ , and thereby inducing the vicious cycle (198). As a function of the increased numbers of osteoclasts and accelerated bone resorption, a marked up-regulation of osteoblast activities is also observed (199, 200).

The activity of osteoclasts in metastases has been monitored closely using biochemical markers of bone turnover (199, 200), and these studies have indicated that bone resorption by tumor-induced osteoclasts to some extent depends on MMP activity, rather than cathepsin K, because the type I collagen fragment ICTP is released in high amounts (199, 201) (Table 1). Animal models of breast cancer bone metastases are to some extent sensitive to inhibitors of both cathepsin K and MMPs (101, 202–204); however, clinical data for MMP inhibitors have been disappointing (205, 206). An interesting question is whether these agents, to be effective, have to inhibit MMPs before the tumors actually metastasize. For cathepsin K inhibitors, the data indicate a beneficial effect on the release of the bone resorption marker N-terminal crosslinked peptide of type I collagen, and an increase in ICTP levels (204). However, further information is needed to draw reliable conclusions on the usefulness of cathepsin K inhibitors for metastatic bone disease.

In contrast, treatments ablating both osteoclasts and the increased osteoblast activity, such as denosumab and bisphosphonates, reduce the destructive capacity of the metastasis and, importantly, the afflicted pain. However, they do not appear to affect the cancer cells (192, 207–210), although there are some indications that the bisphosphonates affect the life span of the cancer cells as well as reducing osteolysis (211).

In summary, from a treatment point of view, there are several similarities between Paget's and osteolytic metastases. The optimal approach appears to involve a strategy of reducing overall bone turnover toward the normal range, such as with the use of denosumab or bisphosphonates. An intriguing possibility would be to target only the areas undergoing destruction, but whether this is feasible is presently not known.

## 3. Arthritic osteoclasts

Later stages of RA are characterized by massive bone destruction caused by osteoclasts (212, 213). However, there are several indications that these osteoclasts are not classical bone-resorbing osteoclasts but include cells that degrade calcified cartilage (83, 214) (Table 1). Several studies have indicated that TNF- $\alpha$  at least partially drives osteoclastogenesis in RA (215), as exemplified by mice overexpressing human TNF- $\alpha$  with massive joint destruction including bone erosion (216). Furthermore, TNF- $\alpha$ -neutralizing antibodies, such as infliximab, or soluble TNF- $\alpha$  receptor antagonists, such as etanercept, provide amelioration of RA in humans (217, 218). Apart from TNF- $\alpha$ , RANKL is, not surprisingly, a crucial factor in osteoclastogenesis during RA (87), and mice deficient in RANKL are protected against bone, but not cartilage, erosion (219). Treatment with OPG of mice with collagen-induced arthritis also leads to amelioration of bone destruction, while having a markedly lower effect on cartilage degradation (220). In addition, a study in which TNF- $\alpha$  overexpressing mice were crossed with mice deficient in *c-fos* (*i.e.*, deficient in osteoclasts), showed no bone destruction but clear evidence of cartilage destruction (221). Furthermore, human clinical trials using denosumab have demonstrated that the RANKL/RANK axis is a key player in RA and that inhibition of RANKL signaling may provide a useful treatment option (222). Finally, more recent evidence has indicated that IL-1 $\alpha$  and IL-1 $\beta$  both play a partial role in bone resorption and cartilage degradation (223). Anakinra, which is a soluble IL-1 receptor antagonist, is also used for treatment of RA, although it appears to be less effective than the TNF- $\alpha$  inhibitors (217). In addition, tocilizumab (anti-interleukin-6 receptor inhibitor) has shown promise in preventing RA progression through an effect including a reduction in osteoclast numbers (158, 324).

Because osteoclasts play a significant role in RA, bisphosphonates appear to be an attractive treatment option. Early evidence has indicated that zoledronate may be useful (224), although this has not been fully established yet (225). Furthermore, interpretation of the effects of bisphosphonates in RA is often clouded by glucocorticoid treatment of the same patients because glucocorticoids are associated with rapid systemic bone loss, independent of RA (226, 227). However, in both collagen-induced arthritis in rats and in the TNF- $\alpha$  transgenic mouse model, zoledronate was effective in reducing both bone and cartilage destruction (228, 229).

The bone resorption process in RA is still not completely understood despite several studies into the molecular mechanisms. The role of cathepsin K has been extensively studied, and the data are somewhat conflicting

(230–232). Overexpression of cathepsin K has been shown to accelerate joint destruction in mice (231), and overexpression of cathepsin K has been observed in humans with RA (233, 234). However, studies in the TNF- $\alpha$  overexpression model crossed with cathepsin K-deficient mice showed that cathepsin K plays only a marginal role in bone resorption in RA (232), a finding supported by a case study showing severe arthritis in a pycnodysostotic patient (235), although there are still controversies with respect to the role of cathepsin K in RA (236).

Other cathepsins have not been explored in detail, and their expression patterns do not indicate a particular effect on osteoclast function in RA (237).

Under some circumstances, MMPs also play a role in bone resorption (2, 81). Studies showing that the MMP-derived collagen type I fragment, ICTP, is increased in RA could indicate that osteoclasts used MMPs to digest matrix under these circumstances (238, 239). Infliximab treatment has been shown to reduce ICTP levels, as well as osteoclast numbers (240), further indicating that osteoclasts utilize MMP-mediated bone degradation in RA. However, a direct link between the production of ICTP and osteoclasts has not been demonstrated yet.

Whether acid secretion by osteoclasts is needed for bone destruction in RA is also not clear. Because bone destruction is likely to occur as a result of MMP activity, the need for acidification may be reduced when compared with “classical” bone resorption (69), although this is still not fully understood. Another possibility is that MMP-mediated collagen type I degradation is mediated by another cell type, although this still remains to be clarified. A case study of arthritis in a case of autosomal dominant osteopetrosis type II (241) showed a lack of bone degradation, whereas cartilage degradation was abundant, thereby mimicking the situation seen in osteoclast-deficient systems (221) and indicating that bone resorption in RA depends fully on acid secretion.

In summary, development of severe RA involves osteoclasts, and a reduction of bone resorption by these cells is desired. This may be obtained through inhibition of inflammation and thereby bone and cartilage destruction, as seen with anti-TNF- $\alpha$  therapy. Alternatively, therapies such as denosumab that target the osteoclasts directly may also be useful, although these fail to eliminate inflammation and only partially prevent cartilage degradation (220). The optimal therapy could be a combination of antiinflammatory and antiosteoclastic measures, although this is presently not known.

In summary, studies of osteoclasts under pathological circumstances have highlighted some important phenomena. First, osteoclasts themselves, not just their resorptive activity, mediate bone formation and therefore perform an

important secondary role in bone remodeling, which is of importance when developing novel treatments for osteoporosis (15). Second, excessive and local activation of osteoclasts occurs in several diseases, and interestingly the osteoclasts appear to switch subtype with respect to their resorption machinery. These findings highlight the importance of characterizing the function of osteoclasts under pathological circumstances to optimize treatment strategies.

## VI. Drug-Induced Osteoclast Subtypes

### A. Existing drugs

Several antiresorptive drugs for the treatment of osteoporosis, as well as glucocorticoids and PTH treatment, are known to alter osteoclasts in various ways. These drugs all provide critical information on osteoclast function, and furthermore, they have also played a great role in illustrating the interplay between osteoclasts and osteoblasts, as will be described in the following section.

#### 1. Bisphosphonates

Bisphosphonates have long been associated with induction of apoptosis in osteoclasts, and the mechanism of action underlying the apoptotic effect depends on whether or not the bisphosphonates contain nitrogen (242). Both classes of bisphosphonates bind to the bone matrix and are taken up by the osteoclast during bone resorption. The simple bisphosphonates are metabolized into toxic ATP analogs, thereby inducing osteoclast apoptosis *in vitro* (242). The nitrogen-containing bisphosphonates exert their function by inhibiting the mevalonate pathway, which leads to the generation of an ATP analog known to induce apoptosis in osteoclasts *in vitro* (242). The antiresorptive potency of the nitrogen-containing bisphosphonates *in vivo* is controlled by mineral binding affinity and by their ability to inhibit the mevalonate pathway (242).

Although *in vitro* data clearly show that bisphosphonates induce apoptosis, analyses of osteoclast numbers in iliac crest biopsies failed to show a reduction in the number of osteoclasts when patients were treated with bisphosphonates (243–245). On the other hand, bisphosphonates reduce systemic levels of TRACP 5b and cathepsin K, both markers of osteoclast number (32, 246–248), potentially indicating that osteoclasts undergo systemic apoptosis, which correlates well with the expected effects of bisphosphonates (242) (Table 1). Other studies have shown that when bisphosphonate therapy continues for more than 1 yr, the number of circulating osteoclast precursors is reduced, and these reductions are speculated to be related to reduced serum RANKL levels (249, 250).

A recent study of biopsies from alendronate-treated patients showed the presence of giant hypernucleated, detached, and frequently apoptotic osteoclasts, and the number of these abnormal osteoclasts correlated with the cumulative dose of bisphosphonate (251). Although interesting, the biological implications of this finding are not clear yet.

One potential explanation for the discrepancies in scoring osteoclasts in the iliac crest biopsies is the very low number of osteoclasts observed in general. Recent reports have also debated the clinical relevance of studying iliac crest biopsies because they are from non-weight-bearing bones and these are different from weight-bearing bones (252–254), and in general more data are needed to draw a final conclusion on the osteoclastic response to bisphosphonates.

On the other hand, the effect on reduction of bone resorption measured both by biochemical markers and by bone histomorphometry (activation frequency) confirms a potent reduction in bone resorption, and the level of reduction is often down to the lower range of premenopausal levels, although this depends heavily on the efficacy of the individual bisphosphonate (242, 243, 245, 255–258).

With respect to secondary effects on bone formation, measurement of biochemical markers of bone turnover shows a marked reduction in bone formation markers, and the effects are maintained throughout the treatment period, although this again is dependent on the individual bisphosphonate (242, 243, 245, 255–258). Biopsy studies have confirmed that bone formation is reduced when compared with placebo, and although the reduction in bone formation rates is dependent on the individual bisphosphonate, the data indicate that bone formation is not completely suppressed but is reduced to the lower postmenopausal levels (243, 245, 258). The FLEX study (Fracture Intervention Trial Long-term Extension), although showing continued reductions in vertebral fractures, increase in bone mineral density (BMD), and reduction of bone turnover markers with alendronate, did not show a significant reduction in bone formation rates when comparing patients stopping alendronate to patients continuing treatment; however, the numbers of biopsies were low (259).

All in all, there is no doubt about the fracture-preventing effects of bisphosphonates; however, knowledge of the effect of bisphosphonates on osteoclasts *in vivo* is quite limited. Apoptosis of the osteoclasts most likely explains the reduction in bone resorption. Furthermore, although the extent of the secondary reduction in bone formation is still discussed, it appears to be clinically relevant, and it most likely is the explanation for the attenuation of the BMD increase seen after the first year of treatment.

*a. Osteonecrosis of the jaw (ONJ) and bisphosphonates.* Bisphosphonate therapy, especially in the case of malignancy-induced bone loss, has been connected to the occurrence of ONJ, mainly due to the ability of bisphosphonates to strongly suppress bone turnover (260–262). Although the probability of ONJ is very low for the dosing regimens used for treatment of osteoporosis, there has still been a lot of debate about whether ONJ is the result of the massive suppression of bone turnover in the jaw (262). Interestingly, alveolar bone of the jaw is very similar to bone matrix in the long bone, *i.e.*, it contains the classical cell types as well as the lamellar structure (263). Furthermore, bone remodeling occurs normally in alveolar bone, although the rate of remodeling has been estimated to be up to 10-fold higher than the corresponding rate in long bones (263–266). In ONJ, the number of osteoclasts has been investigated, and it appears that the osteoclasts are absent from the lesions (267, 268), although opposing evidence also exists (269) and thus more studies are needed.

It has been speculated that massive suppression of osteoclast function, and thus bone turnover, in this high-turnover compartment is what causes ONJ to occur; however, there are several other factors involved, such as tooth extraction or infections, and the overall causality is still not clear (262). One point of particular interest is whether this phenomenon is specific for bisphosphonates or whether it will happen with other very potent and long-lived antiresorptives; however, this is presently not known.

## **2. Selective estrogen receptor modulators/hormone replacement therapy**

Because cessation of estrogen production is a major cause of osteoporosis (3, 14) and both estrogen and SERMs are used for treatment of osteoporosis, several studies have been conducted to clarify their effect on osteoclasts.

Estrogen has been shown to exert direct antiosteoclastic effects at several stages of osteoclastic differentiation and function, namely osteoclastogenesis, resorption, and apoptosis. Direct inhibition of the formation of multinucleated osteoclasts is thought to be caused by suppression of RANKL-induced *c-Jun* and basal *c-Jun* N-terminal kinase activity in osteoclast precursor cells (135, 136). In nonpurified osteoclast-precursor systems, estrogen was found to inhibit osteoclastic differentiation in a human system (270), possibly via down-regulation of the  $\alpha_v\beta_3$  integrin (271). Two studies of estrogen have been conducted using CD14+ osteoclast precursors. As mentioned earlier, one study showed significant inhibition of osteoclastogenesis (137), whereas the other showed no direct effect on osteoclast precursors (272). To date, there is no explanation for this discrepancy.

Studies of the effects of SERMs on osteoclasts have shown that tamoxifen inhibits osteoclastogenesis directly, whereas raloxifene and ospemifene only inhibited osteoclasts through up-regulation of the expression of OPG by osteoblasts (273). Although early studies showed an effect of raloxifene on osteoclastogenesis, these were conducted using mixed cell populations and therefore most likely reflect the increase in OPG (274).

Mature osteoclasts have also been shown to respond directly to estrogen (275, 276). These studies showed that both the activity and the production of the lysosomal enzymes are down-regulated by estrogen (277, 278), possibly explaining the reduction in resorption by the down-regulation of cathepsin K and TRACP (36, 66, 77, 279) (Table 1).

In summary, *in vitro* data clearly demonstrate that estrogen and SERMs reduce osteoclast numbers via inhibition of osteoclastogenesis, and potential effects on bone resorption and apoptosis might add to the *in vivo* effect.

Although some studies of osteoclasts in patients treated with either HRT or SERMs have been conducted, the effects of both estrogen and SERMs on bone remodeling indices based on histomorphometry are quite modest (280–284). Overall, these studies show a reduction in activation, frequency, and depth of resorption, as well as—where detectable—a small decrease in osteoclast numbers. Reduced bone formation rates were also observed, confirming the coupled nature of inhibition mediated by estrogen and SERMs (280–284). These data are corroborated by biochemical markers of bone turnover, which clearly demonstrated a coupled reduction in bone resorption and bone formation (32, 285–287), and furthermore explain the plateau effect observed in BMD measurements after 1 yr (285).

In summary, many of the numerous studies of the *in vitro* mode of action of HRT and SERMs show a reduction in osteoclastogenesis. In alignment, *in vivo* studies of these therapies on osteoclasts confirm that osteoclastogenesis is lower than in the untreated population, and importantly, these also confirm the secondary decrease in bone formation.

### 3. Calcitonin

Calcitonin is a natural peptide hormone produced by parafollicular cells (C cells) of the thyroid gland. Calcitonin possesses potent antiresorptive effects (288), and binding of calcitonin to the calcitonin receptor on osteoclasts induces a rapid change in the cytoskeletal structure of the osteoclasts *in vitro*, which in turn leads to a reduction in bone resorption without inducing apoptosis of the cells (102, 289, 290). Calcitonin in either an injectable or a nasal form has been approved for treatment of osteoporosis; however, because it only prevents about 35% of

vertebral fractures, most likely due to low exposure, the clinical usefulness is limited (122). Recent studies have indicated that a recently developed oral formulation of salmon calcitonin will lead to improved efficacy because it has been optimized with respect to pharmacokinetic and pharmacodynamic properties. This has led to a 10-fold higher exposure and thereby a greater reduction in bone resorption parameters. Thus, this agent will most likely provide improved efficacy in preventing fractures (291), and although it remains to be proven in long-term clinical trials, the phase II data are promising (127).

The mode of action of oral calcitonin is a transient suppression of the nocturnal rise in bone resorption obtained by giving the treatment at the right time of day—in the evening (292), which results in a reduction in bone resorption, but no effect or very modest secondary effects on bone formation (127) (Table 1). These findings are further supported by other clinical studies showing that calcitonin may inhibit bone resorption without affecting bone formation, a finding observed independent of administration route (293–296).

There are histological indications that calcitonin attenuates ruffled border formation by osteoclasts (296–298), and this appears to be the mode of action underlying the antiresorptive effects of calcitonin *in vivo*, thereby elaborating on the previously described transient reduction in bone resorption (292).

Studies of mice lacking the calcitonin receptor indicated that bone formation was increased, and thus that calcitonin is a suppressor of bone formation (299, 300). These studies were conducted mainly in young mice. A recent study in mice deficient in the calcitonin receptor specifically in osteoclasts failed to reproduce this finding (301, 302). However, considering the very modest, or nonexistent, suppression of bone formation in patients treated with calcitonin, the mice data appear of low relevance in the clinical setting (127, 293–296).

Further studies are needed to understand this potential dissociation of bone resorption and bone formation. It may be that this dissociation occurs because calcitonin disappears quickly from the circulation and thus is a completely reversible treatment (122). An interesting question is whether calcitonin treatment may result in better bone quality than potent antiresorptives due to the lack of effect on bone formation and the lower suppression of bone resorption, which is expected to lead to a slow, yet prolonged increase in BMD (6, 303).

### 4. Parathyroid hormone

Although PTH does not appear to affect osteoclasts directly because these cells do not appear to express the PTH receptor, PTH nonetheless affects osteoclast function on many different levels (10). *In vitro* studies of the

effects of PTH on osteoclasts all show that PTH induces osteoclastogenesis and that induction of a transient RANKL expression is essential for this effect (10). However, PTH has mainly been studied in relation to its powerful anabolic effects on osteoblasts (10). Intermittent dosing of PTH in human subjects results in a marked increase in bone formation markers, and secondarily in activation of bone resorption through increased RANKL expression (9, 304, 305). Bone histomorphometric and biochemical marker studies confirm the increase in bone turnover (306–308) (Table 1).

The anabolic mode of action of PTH has been debated extensively. Studies show that PTH directly activates bone formation by osteoblasts when given intermittently (309, 310). In mouse models that are either deficient in osteoclasts or deficient in bone resorption, data suggest that the anabolic effect of PTH is dependent on the presence of mature osteoclasts, but not on their activity (165, 166, 311). Furthermore, initial clinical trials combining alendronate and PTH showed that alendronate blunted the anabolic effect of PTH (312, 313), and there were indications that even pretreatment with alendronate led to a blunting of the PTH response (314). On the other hand, animal studies indicate that PTH can be combined with a bisphosphonate (315, 316), but, as noted by Johnston *et al.* (315), there are marked differences in the doses of PTH used in rodents and in humans.

Collectively, PTH exerts marked regulation of bone turnover (15), including the activation of osteoclasts. The potential anabolic role of osteoclasts and, especially, how to achieve the right osteoclast subtype are debated intensely. Future studies will most likely explain this complex interplay between bone cells and thus guide the right combination of PTH and antiresorptive.

### 5. Strontium ranelate

Strontium ranelate is approved for treatment of osteoporosis, albeit only in Europe, through its ability to reduce fracture risk in patients (317–321). The mode of action has been studied extensively, and yet it is not fully clear exactly how it works *in vivo*. Bone biopsies have been investigated, and these indicated small increases in bone formation and mineralization rates but no changes in bone resorption or osteoclast parameters, thus indicating that strontium ranelate stimulates novel bone formation (322). These data were supported by analysis of biochemical markers of bone turnover demonstrating increased bone formation (308, 323), while also showing a modest decrease in bone resorption markers (323, 325).

*In vitro* studies support the hypothesis that strontium ranelate has a dual effect, namely inhibition of bone resorption while stimulating bone formation (326–329).

Furthermore, strontium has also been shown to increase OPG expression by osteoblasts (330).

In summary, strontium ranelate is a very interesting molecule with respect to effects on osteoclasts, and several lines of *in vitro* evidence indicate that it reduces osteoclast function (326). However, the relevance of the effect on osteoclasts is still debated, and thus the overall effects of this “uncoupling” molecule are still not fully understood.

### 6. Glucocorticoids

Glucocorticoids are used to overcome inflammatory conditions, such as inflammatory bowel diseases and RA (331). Glucocorticoid use is associated with severe bone loss due to strongly attenuated bone formation (332). This attenuation of bone formation leads to a rapid acceleration in the number of fractures in glucocorticoid-treated patients (332), especially in trabecular bone compartments such as vertebrae (333). Glucocorticoid treatment is the most common cause of secondary osteoporosis (333), and thus patients on glucocorticoids are often treated with antiresorptives (334).

*In vivo*, glucocorticoids inhibit osteoblastogenesis, the generation of bone-forming osteoblasts, and promote apoptosis of osteoblasts and osteocytes, which is consistent with the well-known inhibition of bone formation (335).

In contrast, the cellular effects of glucocorticoids on osteoclasts are a subject of controversy. *In vivo*, the effects appear to fall into two categories, one being a short-lived acceleration of osteoclastogenesis and bone resorption, whereas the other is a reduction in osteoclast numbers, which is not well-characterized with respect to exposure time to glucocorticoids (335–340) (Table 1). Interestingly, a study by Kim *et al.* (340) showed that the detrimental effect of glucocorticoids on bone formation was absent when the glucocorticoid receptor was ablated specifically in osteoclasts in mice.

*In vitro* studies of glucocorticoids are often conducted in the presence of contaminating cells, and because glucocorticoid treatment also promotes RANKL and reduces OPG expression in osteoblasts, it is unclear exactly to what extent they influence the osteoclasts (341, 342). Two recent studies showed that glucocorticoid treatment hyperactivated osteoclasts and thus suggests that glucocorticoids indeed have a direct effect on bone resorption (343, 344). Yet some studies show the opposite (340). Overall, the results appear to be very context-dependent, illustrating the complex nature of the biological effects of glucocorticoids.

Measurements of biochemical markers of bone turnover in human subjects on glucocorticoid therapy provided diverse results, which appeared to be dependent on the dose of glucocorticoid used (331, 345, 346). However, biochemical marker data indicate that bone resorption



increases short term, whereas bone formation is attenuated long term (345) in response to glucocorticoid therapy, which corresponds well to mouse studies (335–339). The short-term increase in bone resorption and long-term suppression of bone formation are also observed with histomorphometry in mice (347, 348).

In summary, glucocorticoids exert detrimental effects on bone, and whereas the effects on osteoclasts are not completely clear yet, further investigation of the effect on the coupling between osteoclasts and osteoblasts could explain the overall beneficial effect of antiresorptives on a syndrome mediated primarily by suppressed bone formation (227, 349). These findings further highlight the importance of understanding the interplay between bone cells to provide the optimal treatment.

## B. Future treatments

A series of interesting targets for osteoporosis treatment are currently under investigation. The targets of these treatments to some extent employ novel modes of action on osteoclasts. These novel modes of action are of importance when investigating whether they may have secondary effects on bone formation, and subsequently on bone quality.

### 1. Denosumab

Denosumab is a fully humanized monoclonal antibody to RANKL; it has gone through a phase III fracture efficacy trial in which it was shown to reduce fracture rates by 68% in vertebrae and 40% in hip (351); and it was recently accepted for treatment of severe osteoporosis in both the United States and Europe.

In line with *in vitro* studies of inhibition of RANKL (210, 352), denosumab prevents osteoclastogenesis, blocks bone resorption, and increases osteoclast apoptosis. It induces a massive reduction of osteoclasts *in vivo* and, thereby, almost complete suppression of bone resorption in both humans and mice (352–354) (Table 1). Denosumab treatment also leads to a marked suppression of bone formation markers in humans (353, 354), as well as a marked suppression in bone formation rates measured by histomorphometry in animal models (352, 355). Thus, denosumab treatment is consistent with the classical perception of coupling.

A key point with respect to denosumab is whether the suppression is too severe and could lead to detrimental effects on bone quality long term (6). However, as is the case with bisphosphonate treatment, this is not clear at present.

### 2. Cathepsin K inhibitors

Cathepsin K is a critical protease for degradation of the type I collagen matrix in the resorption pits during bone

resorption by osteoclasts (36, 65, 66). Studies conducted in pycnodysostosis patients before the final identification of cathepsin K showed massive accumulation of nondigested bone collagen fibers in the resorption pit below the osteoclasts (175). These findings were matched by those from investigations in cathepsin K-deficient mice (66), demonstrating a critical role for cathepsin K in degradation of the organic matrix. Further studies in cathepsin K-deficient systems have indicated that cells of the osteoblast lineage, namely bone-lining cells (68); cells of hematopoietic origin (69); and a general up-regulation of the osteoclastic stimuli, osteoclast numbers, and proteases, especially RANKL and MMPs (103), are involved in compensating for the lack of cathepsin K. Interestingly, a hallmark of the absence of cathepsin K function is the presence of the MMP-derived collagen fragment ICTP, which is seen in pycnodysostosis patients, cathepsin K-deficient mice, and cell cultures (67, 69, 103, 356), strongly indicating a compensation by MMPs in the absence of cathepsin K (Table 1).

An interesting study by Fuller *et al.* (357) showed that inhibition of cathepsin K in cultured osteoclasts led to augmented secretion of IGF-I. Furthermore, increased numbers of osteoclasts, containing granules of matrix proteins, have been observed in monkey studies of cathepsin K inhibitors (175, 181), thus indicating the potential of this protease for anabolic stimulation of the osteoblasts. Cathepsin K-deficient mice have been studied extensively, and recent experiments indicate that bone formation in trabecular bone is increased after cathepsin K administration and thus that bone resorption and bone formation are not coupled (100, 176). However, clinical studies of cathepsin K inhibitors, such as odanacatib, have shown that whereas a robust reduction in CTX and N-terminal crosslinked peptide of type I collagen occurred and no changes in TRACP 5b were observed, a significant decrease in the bone formation marker pro-peptide of collagen type I and nonsignificant reductions in bone formation rates by histomorphometry were seen (178). Furthermore, a study of osteoclast morphology as a function of cathepsin K inhibition in humans indicated increased size of the osteoclasts and the presence of large TRACP-positive vacuoles, yet no increase in osteoclast numbers (358). Studies in monkeys clearly demonstrated that bone formation in the trabecular compartments was dose-dependent and significantly reduced by cathepsin K inhibitors, whereas an induction of bone formation was observed at cortical sites (179, 180). Further studies are needed to clarify whether the osteoclasts in cathepsin K-deficient situations indeed signal to the osteoblasts. An indication came from a pycnodysostosis case study that showed no bone formation response to PTH (174), and

thus indicated that secretion of the coupling signals may be attenuated at least in human systems. A possible explanation for the lack of secondary anabolic effects induced by inhibition of cathepsin K is the presence of demineralized collagen fibers in the resorption pit, which are removed by bone-lining cells (68). Although it is not well understood how the presence of fibers and their subsequent removal affect osteoblasts, a study indicated that RGD sequences, which are numerous in collagen, antagonize osteoblast function (359).

These findings again illustrate the importance of carefully investigating the osteoclast subtype as a function of cathepsin K inhibition to more accurately predict potential secondary effects on bone formation.

### 3. Glucagon-like peptide-2

GLP-2 is a 33-amino acid peptide. GLP-2 is created by specific posttranslational proteolytic cleavage of proglucagon in a process that also liberates the related GLP-1 (124). GLP-2 is produced by the intestinal endocrine L cell and by various neurons in the central nervous system (124). Intestinal GLP-2 is cosecreted along with GLP-1 upon nutrient ingestion.

GLP-2 has in clinical settings been demonstrated to inhibit bone resorption (124–126) (Table 1). Reductions in bone resorption by exogenous GLP-2 require an intact gastrointestinal tract (125, 361, 362). The decreased meal-induced inhibition of bone resorption in jejunostomy patients, who lack a GLP-2 response, supports the view that GLP-2 plays a role in postprandial reduction in bone resorption (361, 362).

GLP-2 has in addition been suggested to inhibit bone resorption without affecting bone formation (125), highlighting this mode of inhibition of resorption for further investigation with respect to osteoclast subtypes.

### 4. Acid secretion inhibitors

Acid secretion by osteoclasts has been an interesting therapeutic target since the discovery that this process is controlled by the  $\alpha 3$  subunit of the V-ATPase and ClC-7, both of which are quite specific to osteoclasts (37, 39, 56). Furthermore, *in vitro* studies of osteoclasts treated with inhibitors of these ion transporters have shown that the osteoclasts are unable to resorb bone and that they therefore survive longer (8, 159, 363), thereby mimicking the elevated numbers of osteoclasts observed in patients with mutations in the genes for  $\alpha 3$  and ClC-7 (37, 97) (Table 1). In aged ovariectomized rats, early low-potency chloride channel inhibitors were able to prevent bone resorption by approximately 50%, as monitored by both BMD and the biochemical markers of bone resorption CTX-I or deoxypyridinoline, while augmenting the number of osteoclasts

and showing no inhibition of bone formation markers (8, 364). Similar findings were published for an inhibitor of the V-ATPase (365). In a study of prosthetic implants coated with bafilomycin, osteoclast numbers were elevated, and indications of increased bone formation were observed (366). These studies were the first to provide proof of concept that inhibition of acidification is a really promising target for osteoporosis treatment. Most interestingly, bone formation levels, as measured by osteocalcin and by evaluation of the dynamic histomorphometry parameters mineral apposition rate and the mineralizing surface *vs.* bone surface, were not affected. These data therefore suggest that inhibition of acidification of the osteoclastic resorption lacunae results in an uncoupling of bone formation and bone resorption, thereby possibly improving the potential efficacy of the treatment. This is in contrast to other antiresorptive treatments where a secondary decrease in bone formation is observed (3, 367). These data also indicate that the subtype of osteoclasts obtained—nonresorbing yet alive—when targeting acid secretion is active with respect to bone formation, and thus might possibly be combined with PTH treatment in the future.

Finally, other compounds that appear to modulate the activity of osteoclasts are in development for osteoporosis. These include calcilytics, PTHrP, and sclerostin, but their effects on osteoclasts, which most likely are indirect, are not clear yet (368–370), and thus these will not be described further.

## VII. The Bone Anabolic Effects of the Osteoclasts

Since the early discovery that osteoclast activities were involved in regulation of bone formation during targeted remodeling (17, 18, 371, 372), a series of studies have investigated the nature of this process.

The early studies focused mainly on the release of molecules from the bone matrix during bone resorption and identified molecules such as IGF-I and TGF- $\beta$  (357, 373, 374). However, with the recent discovery that mature osteoclasts, not osteoclast precursors and not necessarily bone resorption, are needed for stimulation of bone formation (8, 9), a series of studies have investigated this phenomenon.

Zhao *et al.* (12) demonstrated that osteoclast-mediated expression of EphrinB2 and osteoblast-mediated expression of EphB4 were involved in a bidirectional communication between these cell types. EphrinB2 on osteoclasts stimulated bone formation by the osteoblasts via binding to EphB4, while EphB4 expression on osteoblasts in turn inhibited osteoclastogenesis via binding to EphrinB2 (12).

However, ephrin signaling requires close contact between the osteoclasts and their target cells. This has led to the speculation that ephrin signaling could be involved in the interplay between osteoclasts and bone-lining cells, which are found in close contact and appear to regulate the activity of each other (68, 375).

Stimulatory signals from osteoclasts directly to mature bone-forming osteoblasts are, on the other hand, likely to be paracrine because these cell types are not found in close contact (264). Both TGF- $\beta$  and IGF-I are produced by the osteoclasts and are known to stimulate bone formation under various circumstances (376–380). In relation to these findings, it is interesting that the anabolic effect of PTH in mice was shown to be mediated through IGF-I (350), an effect that is absent in the absence of osteoclasts (166). This confirms that IGF-I is a coupling factor.

A recent study demonstrated the mature human osteoclasts, independent of their resorptive activity, secrete factors that activate nodule formation by the osteoblasts (11). This study was followed by a study showing that osteoclasts produce the anabolic factors bone morphogenetic protein 6, Wnt10b, and sphingosine-1-phosphate, again independent of bone resorption (13). Furthermore, inhibition of bone morphogenetic protein 6, Wnt10b, and sphingosine-1-phosphate led to inhibition of the osteoclast-mediated stimulation of bone formation *in vitro*. Finally, osteoclasts have also been shown to produce cardiotrophin-1 (CT-1), which activates bone formation by osteoblasts, although the role of CT-1 was clearly shown to be dependent on age because loss of CT-1 in newborn mice caused osteopenia, whereas in larger mice it caused mild osteopetrosis due to defective bone resorption (360).

In summary, the presence of mature osteoclasts is associated with the secretion of stimulation of bone anabolic signals, and whereas several candidate factors have been identified, a clear demonstration that removal of one of the molecules specifically in the osteoclasts *in vivo* leads to loss of bone formation is still missing.

## VIII. Conclusions and Future Perspectives

Osteoclasts have traditionally been viewed as bone resorption “machines”; however, studies of osteoclasts have highlighted that these cells are highly context-specific, and the context of the individual osteoclasts is important for the continued regulation of bone remodeling.

As described in detail in this review, the osteoclasts possess at least two highly important functions: 1) bone resorption, a process that is highly dependent on a series of external stimuli, such as matrix type, remodeling status, hormones involved in calcium homeostasis, genotype, inflammation, and importantly also on intervention strate-

gies; and 2) stimulation of bone formation by the osteoblasts, a process that as illustrated by studies conducted in osteopetrotic patients is, to a large extent, independent of bone resorption. It is presently not completely clear when the osteoclasts are anabolically active, yet it appears to be related to the presence of large multinuclear osteoclasts because bone anabolic responses are seen under these circumstances (3, 15).

Understanding osteoclast functioning may be useful for developing drugs that not only inhibit bone resorption but also enable bone resorption levels that ensure targeted remodeling and, importantly, support continued anabolic signaling from osteoclasts to osteoblasts in the bone remodeling compartment. This has the triple effect of: 1) maintaining a sufficient resorption level and thereby avoiding excessive aging of the bones; 2) sustaining a local stimulation of bone formation at the resorption site only; and 3) not initiating induction of bone formation in otherwise quiescent sites. Theoretically, this type of inhibition of bone resorption would allow a continuous, ongoing increase in BMD, which is in contrast to the effects of the presently approved antiresorptives where a plateau effect on BMD is observed within the first 12–18 months. This means that even with less powerful suppression of bone resorption, such as that seen with the oral formulation of salmon calcitonin, the long-term effects would surpass those of the bisphosphonates.

A deeper understanding of both the differences in the resorption process depending on circumstances and the knowledge relating to when the osteoclasts are anabolically active will aid in the identification of novel treatment opportunities for bone diseases.

Finally, the use of biochemical markers of bone turnover is becoming increasingly relevant for the continued understanding of osteoclasts. Markers provide systemic information on the outcome of a given treatment and can help answer questions such as whether glucocorticoids exert detrimental effects on bone formation, and whether antiresorptives antagonize bone formation secondary to bone resorption because of suppression of osteoclast numbers or activity.

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Address all correspondence and requests for reprints to: K. Henriksen, Nordic Bioscience A/S, Herlev Hovedgade 207, DK-2730 Herlev, Denmark. E-mail: kh@nordicbioscience.com.

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## References

1. Segovia-Silvestre T, Neutzsky-Wulff AV, Sorensen MG, Christiansen C, Bollerslev J, Karsdal MA, Henriksen K 2009 Advances in osteoclast biology resulting from the study of osteopetrotic mutations. *Hum Genet* 124: 561–577
2. Everts V, de Vries TJ, Helfrich MH 2009 Osteoclast heterogeneity: lessons from osteopetrosis and inflammatory conditions. *Biochim Biophys Acta* 1792:757–765
3. Karsdal MA, Martin TJ, Bollerslev J, Christiansen C, Henriksen K 2007 Are nonresorbing osteoclasts sources of bone anabolic activity? *J Bone Miner Res* 22:487–494
4. Everts V, Korper W, Jansen DC, Steinfort J, Lammerse I, Heera S, Docherty AJ, Beertsen W 1999 Functional heterogeneity of osteoclasts: matrix metalloproteinases participate in osteoclastic resorption of calvarial bone but not in resorption of long bone. *FASEB J* 13:1219–1230
5. Marks Jr SC 1983 The origin of osteoclasts: evidence, clinical implications and investigative challenges of an extra-skeletal source. *J Oral Pathol* 12:226–256
6. Leeming DJ, Henriksen K, Byrjalsen I, Qvist P, Madsen SH, Garnero P, Karsdal MA 2009 Is bone quality associated with collagen age? *Osteoporos Int* 20:1461–1470
7. Henriksen K, Leeming DJ, Byrjalsen I, Nielsen RH, Sorensen MG, Dziegiel MH, Martin TJ, Christiansen C, Qvist P, Karsdal MA 2007 Osteoclasts prefer aged bone. *Osteoporos Int* 18:751–759
8. Karsdal MA, Henriksen K, Sorensen MG, Gram J, Schaller S, Dziegiel MH, Heegaard AM, Christophersen P, Martin TJ, Christiansen C, Bollerslev J 2005 Acidification of the osteoclastic resorption compartment provides insight into the coupling of bone formation to bone resorption. *Am J Pathol* 166:467–476
9. Martin TJ, Sims NA 2005 Osteoclast-derived activity in the coupling of bone formation to resorption. *Trends Mol Med* 11:76–81
10. Martin TJ, Quinn JM, Gillespie MT, Ng KW, Karsdal MA, Sims NA 2006 Mechanisms involved in skeletal anabolic therapies. *Ann NY Acad Sci* 1068:458–470
11. Karsdal MA, Neutzsky-Wulff AV, Dziegiel MH, Christiansen C, Henriksen K 2008 Osteoclasts secrete non-bone derived signals that induce bone formation. *Biochem Biophys Res Commun* 366:483–488
12. Zhao C, Irie N, Takada Y, Shimoda K, Miyamoto T, Nishiwaki T, Suda T, Matsuo K 2006 Bidirectional ephrinB2-EphB4 signaling controls bone homeostasis. *Cell Metab* 4:111–121
13. Pederson L, Ruan M, Westendorf JJ, Khosla S, Oursler MJ 2008 Regulation of bone formation by osteoclasts involves Wnt/BMP signaling and the chemokine sphingosine-1-phosphate. *Proc Natl Acad Sci USA* 105:20764–20769
14. Seeman E, Delmas PD 2006 Bone quality—the material and structural basis of bone strength and fragility. *N Engl J Med* 354:2250–2261
15. Henriksen K, Neutzsky-Wulff AV, Bonewald LF, Karsdal MA 2009 Local communication on and within bone controls bone remodeling. *Bone* 44:1026–1033
16. Chavassieux P, Seeman E, Delmas PD 2007 Insights into material and structural basis of bone fragility from diseases associated with fractures: how determinants of the biomechanical properties of bone are compromised by disease. *Endocr Rev* 28:151–164
17. Takahashi H, Epker B, Frost HM 1964 Resorption precedes formative activity. *Surg Forum* 15:437–438
18. Hattner R, Epker BN, Frost HM 1965 Suggested sequential mode of control of changes in cell behaviour in adult bone remodelling. *Nature* 206:489–490
19. Parfitt AM 1982 The coupling of bone formation to bone resorption: a critical analysis of the concept and of its relevance to the pathogenesis of osteoporosis. *Metab Bone Dis Relat Res* 4:1–6
20. Martin TJ 1993 Hormones in the coupling of bone resorption and formation. *Osteoporos Int* 3(Suppl 1):121–125
21. Nakamura M, Udagawa N, Matsuura S, Mogi M, Nakamura H, Horiuchi H, Saito N, Hiraoka BY, Kobayashi Y, Takaoka K, Ozawa H, Miyazawa H, Takahashi N 2003 Osteoprotegerin regulates bone formation through a coupling mechanism with bone resorption. *Endocrinology* 144:5441–5449
22. Teitelbaum SL, Ross FP 2003 Genetic regulation of osteoclast development and function. *Nat Rev Genet* 4:638–649
23. Goltzman D 2002 Discoveries, drugs and skeletal disorders. *Nat Rev Drug Discov* 1:784–796
24. Martin TJ, Seeman E 2007 New mechanisms and targets in the treatment of bone fragility. *Clin Sci (Lond)* 112:77–91
25. Harvey N, Earl S, Cooper C 2006 Epidemiology of osteoporotic fractures. In: Favus MJ, ed. *Primer on the metabolic bone diseases and disorders of mineral metabolism*. Chap 42. 6th ed. Philadelphia: Lippincott Williams & Wilkins; 244–248
26. Albers-Schönberg HE 1904 Röntgenbilder einer seltenen Knochenkrankung. 5th ed. Munch. Med. Wochenschr, München: Germany; 365–368
27. Tolar J, Teitelbaum SL, Orchard PJ 2004 Osteopetrosis. *N Engl J Med* 351:2839–2849
28. Sobacchi C, Frattini A, Guerrini MM, Abinun M, Pangrazio A, Susani L, Bredius R, Mancini G, Cant A, Bishop N, Grabowski P, Del Fattore A, Messina C, Errigo G, Coxon FP, Scott DI, Teti A, Rogers MJ, Vezzoni P, Villa A, Helfrich MH 2007 Osteoclast-poor human osteopetrosis due to mutations in the gene encoding RANKL. *Nat Genet* 39:960–962
29. Guerrini MM, Sobacchi C, Cassani B, Abinun M, Kilic SS, Pangrazio A, Moratto D, Mazzolari E, Clayton-Smith J, Orchard P, Coxon FP, Helfrich MH, Crockett JC, Mellis D, Vellodi A, Tezcan I, Notarangelo LD, Rogers MJ, Vezzoni P, Villa A, Frattini A 2008 Human osteoclast-poor osteopetrosis with hypogammaglobulinemia due to TNFRSF11A (RANK) mutations. *Am J Hum Genet* 83: 64–76
30. Henriksen K, Gram J, Høegh-Andersen P, Jemtland R, Ueland T, Dziegiel MH, Schaller S, Bollerslev J, Karsdal MA 2005 Osteoclasts from patients with autosomal dominant osteopetrosis type I (ADOI) caused by a T253I mutation in LRP5 are normal *in vitro*, but have decreased resorption capacity *in vivo*. *Am J Pathol* 167:1341–1348
31. Glass 2nd DA, Karsenty G 2006 Canonical Wnt signaling in osteoblasts is required for osteoclast differentiation. *Ann NY Acad Sci* 1068:117–130
32. Henriksen K, Tanko LB, Qvist P, Delmas PD, Christiansen C, Karsdal MA 2007 Assessment of osteoclast number and

- function: application in the development of new and improved treatment modalities for bone diseases. *Osteoporos Int* 18:681–685
33. Teitelbaum SL 2007 Osteoclasts: what do they do and how do they do it? *Am J Pathol* 170:427–435
  34. Hayman AR, Jones SJ, Boyde A, Foster D, Colledge WH, Carlton MB, Evans MJ, Cox TM 1996 Mice lacking tartrate-resistant acid phosphatase (Acp 5) have disrupted endochondral ossification and mild osteopetrosis. *Development* 122:3151–3162
  35. Sly WS, Hewett-Emmett D, Whyte MP, Yu YS, Tashian RE 1983 Carbonic anhydrase II deficiency identified as the primary defect in the autosomal recessive syndrome of osteopetrosis with renal tubular acidosis and cerebral calcification. *Proc Natl Acad Sci USA* 80:2752–2756
  36. Gelb BD, Shi GP, Chapman HA, Desnick RJ 1996 Pycnodysostosis, a lysosomal disease caused by cathepsin K deficiency. *Science* 273:1236–1238
  37. Frattini A, Orchard PJ, Sobacchi C, Giliani S, Abinun M, Mattsson JP, Keeling DJ, Andersson AK, Wallbrandt P, Zecca L, Notarangelo LD, Vezzoni P, Villa A 2000 Defects in TCIRG1 subunit of the vacuolar proton pump are responsible for a subset of human autosomal recessive osteopetrosis. *Nat Genet* 25:343–346
  38. Vu TH, Shipley JM, Bergers G, Berger JE, Helms JA, Hanahan D, Shapiro SD, Senior RM, Werb Z 1998 MMP-9/gelatinase B is a key regulator of growth plate angiogenesis and apoptosis of hypertrophic chondrocytes. *Cell* 93:411–422
  39. Kornak U, Kasper D, Bösl MR, Kaiser E, Schweizer M, Schulz A, Friedrich W, Delling G, Jentsch TJ 2001 Loss of the ClC-7 chloride channel leads to osteopetrosis in mice and man. *Cell* 104:205–215
  40. Chalhoub N, Benachenhou N, Rajapurohitam V, Pata M, Ferron M, Frattini A, Villa A, Vacher J 2003 Grey-lethal mutation induces severe malignant autosomal recessive osteopetrosis in mouse and human. *Nat Med* 9:399–406
  41. Findlay DM, Martin TJ 1997 Receptors of calcitropic hormones. *Horm Metab Res* 29:128–134
  42. Del Fattore A, Cappariello A, Teti A 2008 Genetics, pathogenesis and complications of osteopetrosis. *Bone* 42:19–29
  43. Väänänen HK, Horton M 1995 The osteoclast clear zone is a specialized cell-extracellular matrix adhesion structure. *J Cell Sci* 108:2729–2732
  44. Roodman GD 1999 Cell biology of the osteoclast. *Exp Hematol* 27:1229–1241
  45. Boyle WJ, Simonet WS, Lacey DL 2003 Osteoclast differentiation and activation. *Nature* 423:337–342
  46. Zou W, Kitaura H, Reeve J, Long F, Tybulewicz VL, Shattil SJ, Ginsberg MH, Ross FP, Teitelbaum SL 2007 Syk, c-Src, the  $\alpha\beta 3$  integrin, and ITAM immunoreceptors, in concert, regulate osteoclastic bone resorption. *J Cell Biol* 176:877–888
  47. Meadows NA, Sharma SM, Faulkner GJ, Ostrowski MC, Hume DA, Cassady AI 2007 The expression of Clcn7 and Ostm1 in osteoclasts is coregulated by microphthalmia transcription factor. *J Biol Chem* 282:1891–1904
  48. Motyckova G, Weillbaeher KN, Horstmann M, Rieman DJ, Fisher DZ, Fisher DE 2001 Linking osteopetrosis and pycnodysostosis: regulation of cathepsin K expression by the microphthalmia transcription factor family. *Proc Natl Acad Sci USA* 98:5798–5803
  49. Luchin A, Purdom G, Murphy K, Clark MY, Angel N, Cassady AI, Hume DA, Ostrowski MC 2000 The microphthalmia transcription factor regulates expression of the tartrate-resistant acid phosphatase gene during terminal differentiation of osteoclasts. *J Bone Miner Res* 15:451–460
  50. Zhao H, Laitala-Leinonen T, Parikka V, Väänänen HK 2001 Downregulation of small GTPase Rab7 impairs osteoclast polarization and bone resorption. *J Biol Chem* 276:39295–39302
  51. Rodan GA, Martin TJ 2000 Therapeutic approaches to bone diseases. *Science* 289:1508–1514
  52. Blair HC, Teitelbaum SL, Ghiselli R, Gluck S 1989 Osteoclastic bone resorption by a polarized vacuolar proton pump. *Science* 245:855–857
  53. Blair HC, Teitelbaum SL, Tan HL, Koziol CM, Schlesinger PH 1991 Passive chloride permeability charge coupled to H(+)-ATPase of avian osteoclast ruffled membrane. *Am J Physiol* 260:C1315–C1324
  54. Baron R, Neff L, Louvard D, Courtoy PJ 1985 Cell-mediated extracellular acidification and bone resorption: evidence for a low pH in resorbing lacunae and localization of a 100-kD lysosomal membrane protein at the osteoclast ruffled border. *J Cell Biol* 101:2210–2222
  55. Scimeca JC, Franchi A, Trojani C, Parrinello H, Grosgeorge J, Robert C, Jaillon O, Poirier C, Gaudray P, Carle GF 2000 The gene encoding the mouse homologue of the human osteoclast-specific 116-kDa V-ATPase subunit bears a deletion in osteosclerotic (oc/oc) mutants. *Bone* 26:207–213
  56. Li YP, Chen W, Liang Y, Li E, Stashenko P 1999 Atp6i-deficient mice exhibit severe osteopetrosis due to loss of osteoclast-mediated extracellular acidification. *Nat Genet* 23:447–451
  57. Kornak U, Schulz A, Friedrich W, Uhlhaas S, Kremens B, Voit T, Hasan C, Bode U, Jentsch TJ, Kubisch C 2000 Mutations in the  $\alpha 3$  subunit of the vacuolar H(+)-ATPase cause infantile malignant osteopetrosis. *Hum Mol Genet* 9:2059–2063
  58. Henriksen K, Gram J, Schaller S, Dahl BH, Dziegiel MH, Bollerslev J, Karsdal MA 2004 Characterization of osteoclasts from patients harboring a G215R mutation in ClC-7 causing autosomal dominant osteopetrosis type II. *Am J Pathol* 164:1537–1545
  59. Graves AR, Curran PK, Smith CL, Mindell JA 2008 The Cl(-)/H(+) antiporter ClC-7 is the primary chloride permeation pathway in lysosomes. *Nature* 453:788–792
  60. Weinert S, Jabs S, Supanchart C, Schweizer M, Gimber N, Richter M, Rademann J, Stauber T, Kornak U, Jentsch TJ 2010 Lysosomal pathology and osteopetrosis upon loss of H+-driven lysosomal Cl- accumulation. *Science* 328:1401–1403
  61. Josephsen K, Praetorius J, Frische S, Gawenis LR, Kwon TH, Agre P, Nielsen S, Fejerskov O 2009 Targeted disruption of the Cl-/HCO3- exchanger Ae2 results in osteopetrosis in mice. *Proc Natl Acad Sci USA* 106:1638–1641
  62. Jansen ID, Mardones P, Lecanda F, de Vries TJ, Recalde S, Hoeben KA, Schoenmaker T, Ravesloot JH, van Borren MM, van Eijden TM, Bronckers AL, Kellokumpu S, Medina JF, Everts V, Oude Elferink RP 2009 Ae2a,b-Deficient

- mice exhibit osteopetrosis of long bones but not of calvaria. *FASEB J* 23:3470–3481
63. Wu J, Glimcher LH, Aliprantis AO 2008 HCO<sub>3</sub><sup>-</sup>/Cl<sup>-</sup> anion exchanger SLC4A2 is required for proper osteoclast differentiation and function. *Proc Natl Acad Sci USA* 105:16934–16939
  64. Bossard MJ, Tomaszek TA, Thompson SK, Amegadzie BY, Hanning CR, Jones C, Kurdyla JT, McNulty DE, Drake FH, Gowen M, Levy MA 1996 Proteolytic activity of human osteoclast cathepsin K. Expression, purification, activation, and substrate identification. *J Biol Chem* 271:12517–12524
  65. Saftig P, Hunziker E, Wehmeyer O, Jones S, Boyde A, Rommerskirch W, Moritz JD, Schu P, von Figura K 1998 Impaired osteoclastic bone resorption leads to osteopetrosis in cathepsin-K-deficient mice. *Proc Natl Acad Sci USA* 95:13453–13458
  66. Gowen M, Lazner F, Dodds R, Kapadia R, Feild J, Tavarira M, Bertocello I, Drake F, Zavarselk S, Tellis I, Hertzog P, Debouck C, Kola I 1999 Cathepsin K knockout mice develop osteopetrosis due to a deficit in matrix degradation but not demineralization. *J Bone Miner Res* 14:1654–1663
  67. Nishi Y, Atley L, Eyre DE, Edelson JG, Superti-Furga A, Yasuda T, Desnick RJ, Gelb BD 1999 Determination of bone markers in pycnodysostosis: effects of cathepsin K deficiency on bone matrix degradation. *J Bone Miner Res* 14:1902–1908
  68. Everts V, Delaissé JM, Korper W, Jansen DC, Tigchelaar-Gutter W, Saftig P, Beertsen W 2002 The bone lining cell: its role in cleaning Howship's lacunae and initiating bone formation. *J Bone Miner Res* 17:77–90
  69. Henriksen K, Sørensen MG, Nielsen RH, Gram J, Schaller S, Dziegiel MH, Everts V, Bollerslev J, Karsdal MA 2006 Degradation of the organic phase of bone by osteoclasts: a secondary role for lysosomal acidification. *J Bone Miner Res* 21:58–66
  70. Everts V, Korper W, Hoeben KA, Jansen ID, Bromme D, Cleutjens KB, Heeneman S, Peters C, Reinheckel T, Saftig P, Beertsen W 2006 Osteoclastic bone degradation and the role of different cysteine proteinases and matrix metalloproteinases: differences between calvaria and long bone. *J Bone Miner Res* 21:1399–1408
  71. Shorey S, Heersche JN, Manolson MF 2004 The relative contribution of cysteine proteinases and matrix metalloproteinases to the resorption process in osteoclasts derived from long bone and scapula. *Bone* 35:909–917
  72. Salo J, Lehenkari P, Mulari M, Metsikkö K, Väänänen HK 1997 Removal of osteoclast bone resorption products by transcytosis. *Science* 276:270–273
  73. Nesbitt SA, Horton MA 1997 Trafficking of matrix collagens through bone-resorbing osteoclasts. *Science* 276:266–269
  74. Baron R 2005 General principles of bone biology. In: Rosen CJ, Compston JE, Lian JB, eds. *Primer on the metabolic bone diseases and disorders of mineral metabolism*. Chap 1. 5th ed. Hoboken, NJ: Wiley; 1–8
  75. Bollerslev J 1989 Autosomal dominant osteopetrosis: bone metabolism and epidemiological, clinical, and hormonal aspects. *Endocr Rev* 10:45–67
  76. Perez-Amadio S, Jansen DC, Schoenmaker T, Vogels IM, Reinheckel T, Hayman AR, Cox TM, Saftig P, Beertsen W, Everts V 2006 Calvarial osteoclasts express a higher level of tartrate-resistant acid phosphatase than long bone osteoclasts and activation does not depend on cathepsin K or L activity. *Calcif Tissue Int* 79:245–254
  77. Hollberg K, Hultenby K, Hayman A, Cox T, Andersson G 2002 Osteoclasts from mice deficient in tartrate-resistant acid phosphatase have altered ruffled borders and disturbed intracellular vesicular transport. *Exp Cell Res* 279:227–238
  78. Roberts HC, Knott L, Avery NC, Cox TM, Evans MJ, Hayman AR 2007 Altered collagen in tartrate-resistant acid phosphatase (TRAP)-deficient mice: a role for TRAP in bone collagen metabolism. *Calcif Tissue Int* 80:400–410
  79. van den Bos T, Speijer D, Bank RA, Brömme D, Everts V 2008 Differences in matrix composition between calvaria and long bone in mice suggest differences in biomechanical properties and resorption: special emphasis on collagen. *Bone* 43:459–468
  80. Engsig MT, Chen QJ, Vu TH, Pedersen AC, Therkildsen B, Lund LR, Henriksen K, Lenhard T, Foged NT, Werb Z, Delaissé JM 2000 Matrix metalloproteinase 9 and vascular endothelial growth factor are essential for osteoclast recruitment into developing long bones. *J Cell Biol* 151:879–889
  81. Delaissé JM, Andersen TL, Engsig MT, Henriksen K, Troen T, Blavier L 2003 Matrix metalloproteinases (MMP) and cathepsin K contribute differently to osteoclastic activities. *Microsc Res Tech* 61:504–513
  82. Shibata S, Yamashita Y 2001 An ultrastructural study of osteoclasts and chondroclasts in poorly calcified mandible induced by high doses of strontium diet to fetal mice. *Ann Anat* 183:357–361
  83. Bromley M, Woolley DE 1984 Chondroclasts and osteoclasts at subchondral sites of erosion in the rheumatoid joint. *Arthritis Rheum* 27:968–975
  84. Karsdal MA, Leeming DJ, Dam EB, Henriksen K, Alexandersen P, Pastoureaux P, Altman RD, Christiansen C 2008 Should subchondral bone turnover be targeted when treating osteoarthritis? *Osteoarthritis Cartilage* 16:638–646
  85. Mansell JP, Collins C, Bailey AJ 2007 Bone, not cartilage, should be the major focus in osteoarthritis. *Nat Clin Pract Rheumatol* 3:306–307
  86. Ortega N, Behonick DJ, Werb Z 2004 Matrix remodeling during endochondral ossification. *Trends Cell Biol* 14:86–93
  87. Kong YY, Yoshida H, Sarosi I, Tan HL, Timms E, Capparelli C, Morony S, Oliveira-dos-Santos AJ, Van G, Itie A, Khoo W, Wakeham A, Dunstan CR, Lacey DL, Mak TW, Boyle WJ, Penninger JM 1999 OPGL is a key regulator of osteoclastogenesis, lymphocyte development and lymph-node organogenesis. *Nature* 397:315–323
  88. Wiktor-Jedrzejczak W, Bartocci A, Ferrante Jr AW, Ahmed-Ansari A, Sell KW, Pollard JW, Stanley ER 1990 Total absence of colony-stimulating factor 1 in the macrophage-deficient osteopetrotic (op/op) mouse. *Proc Natl Acad Sci USA* 87:4828–4832
  89. Wiktor-Jedrzejczak W, Urbanowska E, Aukerman SL, Pollard JW, Stanley ER, Ralph P, Ansari AA, Sell KW, Szperl M 1991 Correction by CSF-1 of defects in the osteopetrotic op/op mouse suggests local, developmental, and humoral

- requirements for this growth factor. *Exp Hematol* 19:1049–1054
90. Li J, Sarosi I, Yan XQ, Morony S, Capparelli C, Tan HL, McCabe S, Elliott R, Scully S, Van G, Kaufman S, Juan SC, Sun Y, Tarpley J, Martin L, Christensen K, McCabe J, Kostenuik P, Hsu H, Fletcher F, Dunstan CR, Lacey DL, Boyle WJ 2000 RANK is the intrinsic hematopoietic cell surface receptor that controls osteoclastogenesis and regulation of bone mass and calcium metabolism. *Proc Natl Acad Sci USA* 97:1566–1571
  91. Gerber HP, Vu TH, Ryan AM, Kowalski J, Werb Z, Ferrara N 1999 VEGF couples hypertrophic cartilage remodeling, ossification and angiogenesis during endochondral bone formation. *Nat Med* 5:623–628
  92. Blavier L, Delaissé JM 1995 Matrix metalloproteinases are obligatory for the migration of preosteoclasts to the developing marrow cavity of primitive long bones. *J Cell Sci* 108:3649–3659
  93. Dieudonné SC, Foo P, van Zoelen EJ, Burger EH 1991 Inhibiting and stimulating effects of TGF- $\beta$  1 on osteoclastic bone resorption in fetal mouse bone organ cultures. *J Bone Miner Res* 6:479–487
  94. Lowe C, Yoneda T, Boyce BF, Chen H, Mundy GR, Soriano P 1993 Osteopetrosis in Src-deficient mice is due to an autonomous defect of osteoclasts. *Proc Natl Acad Sci USA* 90:4485–4489
  95. Neutzsky-Wulff AV, Karsdal MA, Henriksen K 2008 Characterization of the bone phenotype in CIC-7-deficient mice. *Calcif Tissue Int* 83:425–437
  96. Bollerslev J, Steiniche T, Melsen F, Mosekilde L 1989 Structural and histomorphometric studies of iliac crest trabecular and cortical bone in autosomal dominant osteopetrosis: a study of two radiological types. *Bone* 10:19–24
  97. Bollerslev J, Marks Jr SC, Pockwinse S, Kassem M, Brixen K, Steiniche T, Mosekilde L 1993 Ultrastructural investigations of bone resorptive cells in two types of autosomal dominant osteopetrosis. *Bone* 14:865–869
  98. Nordahl J, Andersson G, Reinholt FP 1998 Chondroclasts and osteoclasts in bones of young rats: comparison of ultrastructural and functional features. *Calcif Tissue Int* 63:401–408
  99. Sawae Y, Sahara T, Sasaki T 2003 Osteoclast differentiation at growth plate cartilage-trabecular bone junction in newborn rat femur. *J Electron Microscop* (Tokyo) 52:493–502
  100. Pennypacker B, Shea M, Liu Q, Masarachia P, Saftig P, Rodan S, Rodan G, Kimmel D 2009 Bone density, strength, and formation in adult cathepsin K ( $-/-$ ) mice. *Bone* 44:199–207
  101. Garnero P, Ferreras M, Karsdal MA, Nicamhloibh R, Risteli J, Borel O, Qvist P, Delmas PD, Foged NT, Delaissé JM 2003 The type I collagen fragments ICTP and CTX reveal distinct enzymatic pathways of bone collagen degradation. *J Bone Miner Res* 18:859–867
  102. Sørensen MG, Henriksen K, Schaller S, Henriksen DB, Nielsen FC, Dziegiel MH, Karsdal MA 2007 Characterization of osteoclasts derived from CD14 $+$  monocytes isolated from peripheral blood. *J Bone Miner Metab* 25:36–45
  103. Kiviranta R, Morko J, Alatalo SL, NicAmhloibh R, Risteli J, Laitala-Leinonen T, Vuorio E 2005 Impaired bone resorption in cathepsin K-deficient mice is partially compensated for by enhanced osteoclastogenesis and increased expression of other proteases via an increased RANKL/OPG ratio. *Bone* 36:159–172
  104. Zenger S, Hollberg K, Ljusberg J, Norgård M, Ek-Rylander B, Kiviranta R, Andersson G 2007 Proteolytic processing and polarized secretion of tartrate-resistant acid phosphatase is altered in a subpopulation of metaphyseal osteoclasts in cathepsin K-deficient mice. *Bone* 41:820–832
  105. Burr DB 2002 Targeted and nontargeted remodeling. *Bone* 30:2–4
  106. Parfitt AM 2002 Targeted and nontargeted bone remodeling: relationship to basic multicellular unit origination and progression. *Bone* 30:5–7
  107. Noble B 2003 Bone microdamage and cell apoptosis. *Eur Cell Mater* 6:46–55; discussion 55
  108. Verborgt O, Gibson GJ, Schaffler MB 2000 Loss of osteocyte integrity in association with microdamage and bone remodeling after fatigue in vivo. *J Bone Miner Res* 15:60–67
  109. Tatsumi S, Ishii K, Amizuka N, Li M, Kobayashi T, Kohno K, Ito M, Takeshita S, Ikeda K 2007 Targeted ablation of osteocytes induces osteoporosis with defective mechanotransduction. *Cell Metab* 5:464–475
  110. Talmage RV, Talmage DW 2006 Calcium homeostasis: solving the solubility problem. *J Musculoskelet Neuronal Interact* 6:402–407
  111. Parfitt AM 2003 Misconceptions (3): calcium leaves bone only by resorption and enters only by formation. *Bone* 33:259–263
  112. Marenzana M, Shipley AM, Squitiero P, Kunkel JG, Rubinacci A 2005 Bone as an ion exchange organ: evidence for instantaneous cell-dependent calcium efflux from bone not due to resorption. *Bone* 37:545–554
  113. Dent CE, Smellie JM, Watson L 1965 Studies in osteopetrosis. *Arch Dis Child* 40:7–15
  114. Manolagas SC 2000 Birth and death of bone cells: basic regulatory mechanisms and implications for the pathogenesis and treatment of osteoporosis. *Endocr Rev* 21:115–137
  115. Karsdal MA, Hjorth P, Henriksen K, Kirkegaard T, Nielsen KL, Lou H, Delaissé JM, Foged NT 2003 Transforming growth factor- $\beta$  controls human osteoclastogenesis through the p38 MAPK and regulation of RANK expression. *J Biol Chem* 278:44975–44987
  116. Van Wesenbeeck L, Odgren PR, Coxon FP, Frattini A, Moens P, Perdu B, MacKay CA, Van Hul E, Timmermans JP, Vanhoenacker F, Jacobs R, Peruzzi B, Teti A, Helfrich MH, Rogers MJ, Villa A, Van Hul W 2007 Involvement of PLEKHM1 in osteoclastic vesicular transport and osteopetrosis in incisors absent rats and humans. *J Clin Invest* 117:919–930
  117. Valcourt U, Merle B, Gineyts E, Viguet-Carrin S, Delmas PD, Garnero P 2007 Non-enzymatic glycation of bone collagen modifies osteoclastic activity and differentiation. *J Biol Chem* 282:5691–5703
  118. Schlemmer A, Hassager C, Jensen SB, Christiansen C 1992 Marked diurnal variation in urinary excretion of pyridinium cross-links in premenopausal women. *J Clin Endocrinol Metab* 74:476–480
  119. Gertz BJ, Clemens JD, Holland SD, Yuan W, Greenspan S 1998 Application of a new serum assay for type I collagen

- cross-linked N-telopeptides: assessment of diurnal changes in bone turnover with and without alendronate treatment. *Calcif Tissue Int* 63:102–106
120. Qvist P, Christgau S, Pedersen BJ, Schlemmer A, Christiansen C 2002 Circadian variation in the serum concentration of C-terminal telopeptide of type I collagen (serum CTx): effects of gender, age, menopausal status, posture, daylight, serum cortisol, and fasting. *Bone* 31:57–61
  121. Bjarnason NH, Henriksen EE, Alexandersen P, Christgau S, Henriksen DB, Christiansen C 2002 Mechanism of circadian variation in bone resorption. *Bone* 30:307–313
  122. Karsdal MA, Byrjalsen I, Riis BJ, Christiansen C 2008 Optimizing bioavailability of oral administration of small peptides through pharmacokinetic and pharmacodynamic parameters: the effect of water and timing of meal intake on oral delivery of salmon calcitonin. *BMC Clin Pharmacol* 8:5
  123. Karsdal MA, Byrjalsen I, Azria M, Arnold M, Choi L, Riis BJ, Christiansen C 2009 Influence of food intake on the bioavailability and efficacy of oral calcitonin. *Br J Clin Pharmacol* 67:413–420
  124. Henriksen DB, Alexandersen P, Byrjalsen I, Hartmann B, Bone HG, Christiansen C, Holst JJ 2004 Reduction of nocturnal rise in bone resorption by subcutaneous GLP-2. *Bone* 34:140–147
  125. Henriksen DB, Alexandersen P, Hartmann B, Adrian CL, Byrjalsen I, Bone HG, Holst JJ, Christiansen C 2007 Dissociation of bone resorption and formation by GLP-2: a 14-day study in healthy postmenopausal women. *Bone* 40:723–729
  126. Henriksen DB, Alexandersen P, Hartmann B, Adrian CL, Byrjalsen I, Bone HG, Holst JJ, Christiansen C 2009 Four-month treatment with GLP-2 significantly increases hip BMD: a randomized, placebo-controlled, dose-ranging study in postmenopausal women with low BMD. *Bone* 45:833–842
  127. Tankó LB, Bagger YZ, Alexandersen P, Devogelaer JP, Reginster JY, Chick R, Olson M, Benmamar H, Mindholm L, Azria M, Christiansen C 2004 Safety and efficacy of a novel salmon calcitonin (sCT) technology-based oral formulation in healthy postmenopausal women: acute and 3-month effects on biomarkers of bone turnover. *J Bone Miner Res* 19:1531–1538
  128. Eghbali-Fatourehchi G, Khosla S, Sanyal A, Boyle WJ, Lacey DL, Riggs BL 2003 Role of RANK ligand in mediating increased bone resorption in early postmenopausal women. *J Clin Invest* 111:1221–1230
  129. Clowes JA, Eghbali-Fatourehchi GZ, McCready L, Oursler MJ, Khosla S, Riggs BL 2009 Estrogen action on bone marrow osteoclast lineage cells of postmenopausal women in vivo. *Osteoporos Int* 20:761–769
  130. Taxel P, Kaneko H, Lee SK, Aguila HL, Raisz LG, Lorenzo JA 2008 Estradiol rapidly inhibits osteoclastogenesis and RANKL expression in bone marrow cultures in postmenopausal women: a pilot study. *Osteoporos Int* 19:193–199
  131. Cao JJ, Wronski TJ, Iwaniec U, Phleger L, Kurimoto P, Boudignon B, Halloran BP 2005 Aging increases stromal/osteoblastic cell-induced osteoclastogenesis and alters the osteoclast precursor pool in the mouse. *J Bone Miner Res* 20:1659–1668
  132. D'Amelio P, Grimaldi A, Pescarmona GP, Tamone C, Roato I, Isaia G 2005 Spontaneous osteoclast formation from peripheral blood mononuclear cells in postmenopausal osteoporosis. *FASEB J* 19:410–412
  133. Jevon M, Hirayama T, Brown MA, Wass JA, Sabokbar A, Ostelere S, Athenasou NA 2003 Osteoclast formation from circulating precursors in osteoporosis. *Scand J Rheumatol* 32:95–100
  134. D'Amelio P, Grimaldi A, Di Bella S, Brianza SZ, Cristofaro MA, Tamone C, Giribaldi G, Ulliers D, Pescarmona GP, Isaia G 2008 Estrogen deficiency increases osteoclastogenesis up-regulating T cells activity: a key mechanism in osteoporosis. *Bone* 43:92–100
  135. Shevde NK, Bendixen AC, Dienger KM, Pike JW 2000 Estrogens suppress RANK ligand-induced osteoclast differentiation via a stromal cell independent mechanism involving c-Jun repression. *Proc Natl Acad Sci USA* 97:7829–7834
  136. Srivastava S, Toraldo G, Weitzmann MN, Cenci S, Ross FP, Pacifici R 2001 Estrogen decreases osteoclast formation by down-regulating receptor activator of NF- $\kappa$ B ligand (RANKL)-induced JNK activation. *J Biol Chem* 276:8836–8840
  137. Sørensen MG, Henriksen K, Dziegiel MH, Tankó LB, Karsdal MA 2006 Estrogen directly attenuates human osteoclastogenesis, but has no effect on resorption by mature osteoclasts. *DNA Cell Biol* 25:475–483
  138. Huber DM, Bendixen AC, Pathrose P, Srivastava S, Dienger KM, Shevde NK, Pike JW 2001 Androgens suppress osteoclast formation induced by RANKL and macrophage-colony stimulating factor. *Endocrinology* 142:3800–3808
  139. Nakamura T, Imai Y, Matsumoto T, Sato S, Takeuchi K, Igarashi K, Harada Y, Azuma Y, Krust A, Yamamoto Y, Nishina H, Takeda S, Takayanagi H, Metzger D, Kanno J, Takaoka K, Martin TJ, Chambon P, Kato S 2007 Estrogen prevents bone loss via estrogen receptor  $\alpha$  and induction of Fas ligand in osteoclasts. *Cell* 130:811–823
  140. Bord S, Horner A, Beavan S, Compston J 2001 Estrogen receptors  $\alpha$  and  $\beta$  are differentially expressed in developing human bone. *J Clin Endocrinol Metab* 86:2309–2314
  141. Martin-Millan M, Almeida M, Ambrogini E, Han L, Zhao H, Weinstein RS, Jilka RL, O'Brien CA, Manolagas SC 2010 The estrogen receptor- $\alpha$  in osteoclasts mediates the protective effects of estrogens on cancellous but not cortical bone. *Mol Endocrinol* 24:323–334
  142. Riggs BL, Melton Iii 3rd LJ, Robb RA, Camp JJ, Atkinson EJ, Peterson JM, Rouleau PA, McCollough CH, Bouxsein ML, Khosla S 2004 Population-based study of age and sex differences in bone volumetric density, size, geometry, and structure at different skeletal sites. *J Bone Miner Res* 19:1945–1954
  143. Recker R, Lappe J, Davies KM, Heaney R 2004 Bone remodeling increases substantially in the years after menopause and remains increased in older osteoporosis patients. *J Bone Miner Res* 19:1628–1633
  144. Garnero P, Sornay-Rendu E, Chapuy MC, Delmas PD 1996 Increased bone turnover in late postmenopausal women is a major determinant of osteoporosis. *J Bone Miner Res* 11:337–349
  145. Arlot ME, Delmas PD, Chappard D, Meunier PJ 1990 Trabecular and endocortical bone remodeling in postmenopausal osteoporosis: comparison with normal postmenopausal women. *Osteoporos Int* 1:41–49



146. Viguet-Carrin S, Garnero P, Delmas PD 2006 The role of collagen in bone strength. *Osteoporos Int* 17:319–336
147. Herrmann M, Widmann T, Colaianni G, Colucci S, Zalzone A, Herrmann W 2005 Increased osteoclast activity in the presence of increased homocysteine concentrations. *Clin Chem* 51:2348–2353
148. Saito M, Marumo K 2010 Collagen cross-links as a determinant of bone quality: a possible explanation for bone fragility in aging, osteoporosis, and diabetes mellitus. *Osteoporos Int* 21:195–214
149. Miyata T, Notoya K, Yoshida K, Horie K, Maeda K, Kurokawa K, Taketomi S 1997 Advanced glycation end products enhance osteoclast-induced bone resorption in cultured mouse unfractionated bone cells and in rats implanted subcutaneously with devitalized bone particles. *J Am Soc Nephrol* 8:260–270
150. Schwartz AV, Garnero P, Hillier TA, Sellmeyer DE, Strotmeyer ES, Feingold KR, Resnick HE, Tylavsky FA, Black DM, Cummings SR, Harris TB, Bauer DC 2009 Pentosidine and increased fracture risk in older adults with type 2 diabetes. *J Clin Endocrinol Metab* 94:2380–2386
151. Raska Jr I, Broulík P 2005 The impact of diabetes mellitus on skeletal health: an established phenomenon with in-established causes? *Prague Med Rep* 106:137–148
152. Sanguineti R, Storace D, Monacelli F, Federici A, Odetti P 2008 Pentosidine effects on human osteoblasts in vitro. *Ann NY Acad Sci* 1126:166–172
153. Henriksen K, Gram J, Neutzsky-Wulff AV, Jensen VK, Dziegiel MH, Bollerslev J, Karsdal MA 2009 Characterization of acid flux in osteoclasts from patients harboring a G215R mutation in *CLC-7*. *Biochem Biophys Res Commun* 378:804–809
154. Taranta A, Migliaccio S, Recchia I, Caniglia M, Luciani M, De Rossi G, Dionisi-Vici C, Pinto RM, Francalanci P, Boldrini R, Lanino E, Dini G, Morreale G, Ralston SH, Villa A, Vezzoni P, Del Principe D, Cassiani F, Palumbo G, Teti A 2003 Genotype-phenotype relationship in human ATP6i-dependent autosomal recessive osteopetrosis. *Am J Pathol* 162:57–68
155. Maranda B, Chabot G, Décarie JC, Pata M, Azeddine B, Moreau A, Vacher J 2008 Clinical and cellular manifestations of *OSTM1*-related infantile osteopetrosis. *J Bone Miner Res* 23:296–300
156. Rajapurohitam V, Chalhoub N, Benachenhou N, Neff L, Baron R, Vacher J 2001 The mouse osteopetrotic greylethal mutation induces a defect in osteoclast maturation/function. *Bone* 28:513–523
157. Semba I, Ishigami T, Sugihara K, Kitano M 2000 Higher osteoclastic demineralization and highly mineralized cement lines with osteocalcin deposition in a mandibular cortical bone of autosomal dominant osteopetrosis type II: ultrastructural and undecalcified histological investigations. *Bone* 27:389–395
158. Garnero P, Thompson E, Woodworth T, Smolen JS 2010 Rapid and sustained improvement in bone and cartilage turnover markers with the anti-interleukin-6 receptor inhibitor tocilizumab plus methotrexate in rheumatoid arthritis patients with an inadequate response to methotrexate: results from a substudy of the multicenter double-blind, placebo-controlled trial of tocilizumab in inadequate responders to methotrexate alone. *Arthritis Rheum*. 62:33–43
159. Nielsen RH, Karsdal MA, Sørensen MG, Dziegiel MH, Henriksen K 2007 Dissolution of the inorganic phase of bone leading to release of calcium regulates osteoclast survival. *Biochem Biophys Res Commun* 360:834–839
160. Alatalo SL, Ivaska KK, Waguespack SG, Econs MJ, Väänänen HK, Halleen JM 2004 Osteoclast-derived serum tartrate-resistant acid phosphatase 5b in Albers-Schonberg disease (type II autosomal dominant osteopetrosis). *Clin Chem* 50:883–890
161. Del Fattore A, Peruzzi B, Rucci N, Recchia I, Cappariello A, Longo M, Fortunati D, Ballanti P, Iacobini M, Luciani M, Devito R, Pinto R, Caniglia M, Lanino E, Messina C, Cesaro S, Letizia C, Bianchini G, Fryssira H, Grabowski P, Shaw N, Bishop N, Hughes D, Kapur RP, Datta HK, Taranta A, Fornari R, Migliaccio S, Teti A 2006 Clinical, genetic, and cellular analysis of 49 osteopetrotic patients: implications for diagnosis and treatment. *J Med Genet* 43:315–325
162. Soriano P, Montgomery C, Geske R, Bradley A 1991 Targeted disruption of the *c-src* proto-oncogene leads to osteopetrosis in mice. *Cell* 64:693–702
163. Grigoriadis AE, Wang ZQ, Cecchini MG, Hofstetter W, Felix R, Fleisch HA, Wagner EF 1994 *c-Fos*: a key regulator of osteoclast-macrophage lineage determination and bone remodeling. *Science* 266:443–448
164. Marzia M, Sims NA, Voit S, Migliaccio S, Taranta A, Bernardini S, Faraggiana T, Yoneda T, Mundy GR, Boyce BF, Baron R, Teti A 2000 Decreased *c-Src* expression enhances osteoblast differentiation and bone formation. *J Cell Biol* 151:311–320
165. Demiralp B, Chen HL, Koh AJ, Keller ET, McCauley LK 2002 Anabolic actions of parathyroid hormone during bone growth are dependent on *c-fos*. *Endocrinology* 143:4038–4047
166. Koh AJ, Demiralp B, Neiva KG, Hooten J, Nohutcu RM, Shim H, Datta NS, Taichman RS, McCauley LK 2005 Cells of the osteoclast lineage as mediators of the anabolic actions of parathyroid hormone in bone. *Endocrinology* 146:4584–4596
167. Holmen SL, Zylstra CR, Mukherjee A, Sigler RE, Faugere MC, Bouxsein ML, Deng L, Clemens TL, Williams BO 2005 Essential role of  $\beta$ -catenin in postnatal bone acquisition. *J Biol Chem* 280:21162–21168
168. Gong Y, Slee RB, Fukai N, Rawadi G, Roman-Roman S, Reginato AM, Wang H, Cundy T, Glorieux FH, Lev D, Zacharin M, Oexle K, Marcelino J, Suwairi W, Heeger S, Sabatakos G, Apte S, Adkins WN, Allgrove J, Arslan-Kirchner M, Batch JA, Beighton P, Black GC, Boles RG, Boon LM, Borrone C, Brunner HG, Carle GF, Dallapiccola B, De Paepe A, Floege B, Halfhide ML, Hall B, Hennekam RC, Hirose T, Jans A, Jüppner H, Kim CA, Keppler-Noreuil K, Kohlschütter A, LaCombe D, Lambert M, Lemyre E, Letteboer T, Peltonen L, Ramesar RS, Romanengo M, Somer H, Steichen-Gersdorf E, Steinmann B, Sullivan B, Superti-Furga A, Swoboda W, van den Boogaard MJ, Van Hul W, Vikkula M, Votruba M, Zabel B, Garcia T, Baron R, Olsen BR, Warman ML 2001 LDL receptor-related protein 5 (*LRP5*) affects bone accrual and eye development. *Cell* 107:513–523
169. Van Wesenbeeck L, Cleiren E, Gram J, Beals RK, Bénichou O, Scopelliti D, Key L, Renton T, Bartels C, Gong Y, Warman ML, De Vernejoul MC, Bollerslev J, Van Hul W 2003

- Six novel missense mutations in the LDL receptor-related protein 5 (LRP5) gene in different conditions with an increased bone density. *Am J Hum Genet* 72:763–771
170. **Bollerslev J, Ueland T, Odgren PR** 2003 Serum levels of TGF- $\beta$  and fibronectin in autosomal dominant osteopetrosis in relation to underlying mutations and well-described murine counterparts. *Crit Rev Eukaryot Gene Expr* 13:163–171
  171. **Sarnethsiri P, Hitt OK, Eyring EJ, Frost HM** 1971 Tetracycline-based study of bone dynamics in pycnodysostosis. *Clin Orthop Relat Res* 74:301–312
  172. **Ho N, Punturieri A, Wilkin D, Szabo J, Johnson M, Whalley J, Davis J, Clark A, Weiss S, Francomano C** 1999 Mutations of CTSK result in pycnodysostosis via a reduction in cathepsin K protein. *J Bone Miner Res* 14:1649–1653
  173. **Hou WS, Brömme D, Zhao Y, Mehler E, Dushey C, Weinstein H, Miranda CS, Fraga C, Greig F, Carey C, Rimoind DL, Desnick RJ, Gelb BD** 1999 Characterization of novel cathepsin K mutations in the pro and mature polypeptide regions causing pycnodysostosis. *J Clin Invest* 103:731–738
  174. **Chavassieux P, Asser Karsdal M, Segovia-Silvestre T, Neutzsky-Wulff AV, Chapurlat R, Boivin G, Delmas PD** 2008 Mechanisms of the anabolic effects of teriparatide on bone: insight from the treatment of a patient with pycnodysostosis. *J Bone Miner Res* 23:1076–1083
  175. **Everts V, Aronson DC, Beertsen W** 1985 Phagocytosis of bone collagen by osteoclasts in two cases of pycnodysostosis. *Calcif Tissue Int* 37:25–31
  176. **Li CY, Jepsen KJ, Majeska RJ, Zhang J, Ni R, Gelb BD, Schaffler MB** 2006 Mice lacking cathepsin K maintain bone remodeling but develop bone fragility despite high bone mass. *J Bone Miner Res* 21:865–875
  177. **Fratzl-Zelman N, Valenta A, Roschger P, Nader A, Gelb BD, Fratzl P, Klaushofer K** 2004 Decreased bone turnover and deterioration of bone structure in two cases of pycnodysostosis. *J Clin Endocrinol Metab* 89:1538–1547
  178. **Bone HG, McClung MR, Roux C, Recker RR, Eisman JA, Verbruggen N, Hustad CM, DaSilva C, Santora AC, Ince BA** 2010 Odanacatib, a cathepsin-K inhibitor for osteoporosis: a two-year study in postmenopausal women with low bone density. *J Bone Miner Res* 25:937–947
  179. **Pennypacker B, Wesolowski G, Heo J, Duong LT** 2009 Effects of odanacatib on central femur cortical bone in estrogen-deficient adult rhesus monkeys. *J Bone Miner Res* 24(Suppl 1):1171 (Abstract)
  180. **Cusick T, Pennypacker B, Scott K, Duong LT, Kimmel D** 2009 Effects of odanacatib on bone mass, turnover and strength in the femoral neck of estrogen deficient adult rhesus monkeys. *J Bone Miner Res* 24(Suppl 1):FR0416 (Abstract)
  181. **Scott K, Cusick T, Duong LT, Pennypacker B, Kimmel D** 2009 Effects of odanacatib on bone turnover and osteoclast morphology in the lumbar vertebra of ovariectomized adult rhesus monkeys. *J Bone Miner Res* 24(Suppl 1):SU0227 (Abstract)
  182. **Helfrich M, Crockett JC, Hocking LJ, Coxon FP** 2007 The pathogenesis of osteoclast diseases: some knowns, but still many unknowns. *BoneKEy-Osteovision* 4:61–77
  183. **Roodman GD** 1996 Paget's disease and osteoclast biology. *Bone* 19:209–212
  184. **Goode A, Layfield R** 2010 Recent advances in understanding the molecular basis of Paget's disease of bone. *J Clin Pathol* 63:199–203
  185. **Roodman GD, Windle JJ** 2005 Paget disease of bone. *J Clin Invest* 115:200–208
  186. **Ralston SH, Langston AL, Reid IR** 2008 Pathogenesis and management of Paget's disease of bone. *Lancet* 372:155–163
  187. **Neale SD, Smith R, Wass JA, Athanasou NA** 2000 Osteoclast differentiation from circulating mononuclear precursors in Paget's disease is hypersensitive to 1,25-dihydroxyvitamin D(3) and RANKL. *Bone* 27:409–416
  188. **Singer FR, Mills BG, Gruber HE, Windle JJ, Roodman GD** 2006 Ultrastructure of bone cells in Paget's disease of bone. *J Bone Miner Res* 21(Suppl 2):P51–P54
  189. **Kurihara N, Hiruma Y, Zhou H, Subler MA, Dempster DW, Singer FR, Reddy SV, Gruber HE, Windle JJ, Roodman GD** 2007 Mutation of the sequestosome 1 (p62) gene increases osteoclastogenesis but does not induce Paget disease. *J Clin Invest* 117:133–142
  190. **Hiruma Y, Kurihara N, Subler MA, Zhou H, Boykin CS, Zhang H, Ishizuka S, Dempster DW, Roodman GD, Windle JJ** 2008 A SQSTM1/p62 mutation linked to Paget's disease increases the osteoclastogenic potential of the bone microenvironment. *Hum Mol Genet* 17:3708–3719
  191. **Gennari L, Merlotti D, Mossetti G, Rendina D, De Paola V, Martini G, Nuti R** 2009 The use of intravenous aminobisphosphonates for the treatment of Paget's disease of bone. *Mini Rev Med Chem* 9:1052–1063
  192. **Clines GA, Guise TA** 2008 Molecular mechanisms and treatment of bone metastasis. *Expert Rev Mol Med* 10:e7
  193. **Guise TA, Mohammad KS, Clines G, Stebbins EG, Wong DH, Higgins LS, Vessella R, Corey E, Padalecki S, Suva L, Chirgwin JM** 2006 Basic mechanisms responsible for osteolytic and osteoblastic bone metastases. *Clin Cancer Res* 12:6213s–6216s
  194. **Hu MI, Lu H, Gage RF** 2010 Cancer therapies and bone health. *Curr Rheumatol Rep* 12:177–185
  195. **Akhtari M, Mansuri J, Newman KA, Guise TM, Seth P** 2008 Biology of breast cancer bone metastasis. *Cancer Biol Ther* 7:3–9
  196. **Lipton A** 2006 Future treatment of bone metastases. *Clin Cancer Res* 12:6305s–6308s
  197. **Cleazardin P, Teti A** 2007 Bone metastasis: pathogenesis and therapeutic implications. *Clin Exp Metastasis* 24:599–608
  198. **Ciccek M, Oursler MJ** 2006 Breast cancer bone metastasis and current small therapeutics. *Cancer Metastasis Rev* 25:635–644
  199. **Leeming DJ, Koizumi M, Byrjalsen I, Li B, Qvist P, Tankó LB** 2006 The relative use of eight collagenous and noncollagenous markers for diagnosis of skeletal metastases in breast, prostate, or lung cancer patients. *Cancer Epidemiol Biomarkers Prev* 15:32–38
  200. **Leeming DJ, Delling G, Koizumi M, Henriksen K, Karsdal MA, Li B, Qvist P, Tankó LB, Byrjalsen I** 2006  $\alpha$  CTX as a biomarker of skeletal invasion of breast cancer: immunolocalization and the load dependency of urinary excretion. *Cancer Epidemiol Biomarkers Prev* 15:1392–1395
  201. **Koopmans N, de Jong IJ, Breeuwsma AJ, van der Veer E** 2007 Serum bone turnover markers (PINP and ICTP) for the early detection of bone metastases in patients with

- prostate cancer: a longitudinal approach. *J Urol* 178:849–853
202. Winding B, NicAmhlaibh R, Misander H, Høegh-Andersen P, Andersen TL, Holst-Hansen C, Heegaard AM, Foged NT, Brüner N, Delaissé JM 2002 Synthetic matrix metalloproteinase inhibitors inhibit growth of established breast cancer osteolytic lesions and prolong survival in mice. *Clin Cancer Res* 8:1932–1939
  203. Le Gall C, Bonnelye E, Clézardin P 2008 Cathepsin K inhibitors as treatment of bone metastasis. *Curr Opin Support Palliat Care* 2:218–222
  204. Le Gall C, Bellahcène A, Bonnelye E, Gasser JA, Castronovo V, Green J, Zimmermann J, Clézardin P 2007 A cathepsin K inhibitor reduces breast cancer induced osteolysis and skeletal tumor burden. *Cancer Res* 67:9894–9902
  205. Coussens LM, Fingleton B, Matrisian LM 2002 Matrix metalloproteinase inhibitors and cancer: trials and tribulations. *Science* 295:2387–2392
  206. Pavlaki M, Zucker S 2003 Matrix metalloproteinase inhibitors (MMPi): the beginning of phase I or the termination of phase III clinical trials. *Cancer Metastasis Rev* 22:177–203
  207. Coleman R, Gnant M 2009 New results from the use of bisphosphonates in cancer patients. *Curr Opin Support Palliat Care* 3:213–218
  208. Machado M, Cruz LS, Tannus G, Fonseca M 2009 Efficacy of clodronate, pamidronate, and zoledronate in reducing morbidity and mortality in cancer patients with bone metastasis: a meta-analysis of randomized clinical trials. *Clin Ther* 31:962–979
  209. Fizazi K, Lipton A, Mariette X, Body JJ, Rahim Y, Gralow JR, Gao G, Wu L, Sohn W, Jun S 2009 Randomized phase II trial of denosumab in patients with bone metastases from prostate cancer, breast cancer, or other neoplasms after intravenous bisphosphonates. *J Clin Oncol* 27:1564–1571
  210. Kearns AE, Khosla S, Kostenuik PJ 2008 Receptor activator of nuclear factor  $\kappa$ B ligand and osteoprotegerin regulation of bone remodeling in health and disease. *Endocr Rev* 29:155–192
  211. Rachner TD, Singh SK, Schoppet M, Benad P, Bornhäuser M, Ellenrieder V, Ebert R, Jakob F, Hofbauer LC 2010 Zoledronic acid induces apoptosis and changes the TRAIL/OPG ratio in breast cancer cells. *Cancer Lett* 287:109–116
  212. Schett G, Teitelbaum SL 2009 Osteoclasts and arthritis. *J Bone Miner Res* 24:1142–1146
  213. Schett G 2007 Erosive arthritis. *Arthritis Res Ther* 9(Suppl 1):S2
  214. Gravalles EM, Harada Y, Wang JT, Gorn AH, Thornhill TS, Goldring SR 1998 Identification of cell types responsible for bone resorption in rheumatoid arthritis and juvenile rheumatoid arthritis. *Am J Pathol* 152:943–951
  215. Feldmann M, Brennan FM, Maini RN 1996 Role of cytokines in rheumatoid arthritis. *Annu Rev Immunol* 14:397–440
  216. Keffer J, Probert L, Cazlaris H, Georgopoulos S, Kaslaris E, Kioussis D, Kollias G 1991 Transgenic mice expressing human tumour necrosis factor: a predictive genetic model of arthritis. *EMBO J* 10:4025–4031
  217. Singh JA, Christensen R, Wells GA, Suarez-Almazor ME, Buchbinder R, Lopez-Olivo MA, Ghogomu ET, Tugwell P 2009 A network meta-analysis of randomized controlled trials of biologics for rheumatoid arthritis: a Cochrane overview. *CMAJ* 181:787–796
  218. Licastro F, Chiappelli M, Ianni M, Porcellini E 2009 Tumor necrosis factor- $\alpha$  antagonists: differential clinical effects by different biotechnological molecules. *Int J Immunopathol Pharmacol* 22:567–572
  219. Pettit AR, Ji H, von Stechow D, Müller R, Goldring SR, Choi Y, Benoist C, Gravalles EM 2001 TRANCE/RANKL knockout mice are protected from bone erosion in a serum transfer model of arthritis. *Am J Pathol* 159:1689–1699
  220. Romas E, Sims NA, Hards DK, Lindsay M, Quinn JW, Ryan PF, Dunstan CR, Martin TJ, Gillespie MT 2002 Osteoprotegerin reduces osteoclast numbers and prevents bone erosion in collagen-induced arthritis. *Am J Pathol* 161:1419–1427
  221. Redlich K, Hayer S, Ricci R, David JP, Tohidast-Akrad M, Kollias G, Steiner G, Smolen JS, Wagner EF, Schett G 2002 Osteoclasts are essential for TNF- $\alpha$ -mediated joint destruction. *J Clin Invest* 110:1419–1427
  222. Cohen SB, Dore RK, Lane NE, Ory PA, Peterfy CG, Sharp JT, van der Heijde D, Zhou L, Tsuji W, Newmark R 2008 Denosumab treatment effects on structural damage, bone mineral density, and bone turnover in rheumatoid arthritis: a twelve-month, multicenter, randomized, double-blind, placebo-controlled, phase II clinical trial. *Arthritis Rheum* 58:1299–1309
  223. Zwerina J, Redlich K, Polzer K, Joosten L, Krönke G, Distler J, Hess A, Pundt N, Pap T, Hoffmann O, Gasser J, Scheinecker C, Smolen JS, van den Berg W, Schett G 2007 TNF-induced structural joint damage is mediated by IL-1. *Proc Natl Acad Sci USA* 104:11742–11747
  224. Jarrett SJ, Conaghan PG, Sloan VS, Papanastasiou P, Ortmann CE, O'Connor PJ, Grainger AJ, Emery P 2006 Preliminary evidence for a structural benefit of the new bisphosphonate zoledronic acid in early rheumatoid arthritis. *Arthritis Rheum* 54:1410–1414
  225. Breuil V, Euler-Ziegler L 2006 Bisphosphonate therapy in rheumatoid arthritis. *Joint Bone Spine* 73:349–354
  226. Lems WF, Lodder MC, Lips P, Bijlsma JW, Geusens P, Schrameijer N, van de Ven CM, Dijkmans BA 2006 Positive effect of alendronate on bone mineral density and markers of bone turnover in patients with rheumatoid arthritis on chronic treatment with low-dose prednisone: a randomized, double-blind, placebo-controlled trial. *Osteoporos Int* 17:716–723
  227. Reid DM, Devogelaer JP, Saag K, Roux C, Lau CS, Reingster JY, Papanastasiou P, Ferreira A, Hartl F, Fashola T, Mesenbrink P, Sambrook PN 2009 Zoledronic acid and risedronate in the prevention and treatment of glucocorticoid-induced osteoporosis (HORIZON): a multicentre, double-blind, double-dummy, randomised controlled trial. *Lancet* 373:1253–1263
  228. Sims NA, Green JR, Glatt M, Schlicht S, Martin TJ, Gillespie MT, Romas E 2004 Targeting osteoclasts with zoledronic acid prevents bone destruction in collagen-induced arthritis. *Arthritis Rheum* 50:2338–2346
  229. Herrak P, Görtz B, Hayer S, Redlich K, Reiter E, Gasser J, Bergmeister H, Kollias G, Smolen JS, Schett G 2004 Zoledronic acid protects against local and systemic bone loss in tumor necrosis factor-mediated arthritis. *Arthritis Rheum* 50:2327–2337

230. Morko JP, Söderström M, Säämänen AM, Salminen HJ, Vuorio EI 2004 Up regulation of cathepsin K expression in articular chondrocytes in a transgenic mouse model for osteoarthritis. *Ann Rheum Dis* 63:649–655
231. Morko J, Kiviranta R, Joronen K, Säämänen AM, Vuorio E, Salminen-Mankonen H 2005 Spontaneous development of synovitis and cartilage degeneration in transgenic mice overexpressing cathepsin K. *Arthritis Rheum* 52:3713–3717
232. Schurigt U, Hummel KM, Petrow PK, Gajda M, Stöckigt R, Middel P, Zwerina J, Janik T, Bernhardt R, Schüler S, Scharnweber D, Beckmann F, Saftig P, Kollias G, Schett G, Wiederanders B, Bräuer R 2008 Cathepsin K deficiency partially inhibits, but does not prevent, bone destruction in human tumor necrosis factor-transgenic mice. *Arthritis Rheum* 58:422–434
233. Hou WS, Li Z, Gordon RE, Chan K, Klein MJ, Levy R, Keysser M, Keyszer G, Brömme D 2001 Cathepsin K is a critical protease in synovial fibroblast-mediated collagen degradation. *Am J Pathol* 159:2167–2177
234. Yasuda Y, Kaleta J, Brömme D 2005 The role of cathepsins in osteoporosis and arthritis: rationale for the design of new therapeutics. *Adv Drug Deliv Rev* 57:973–993
235. Ainola M, Valleala H, Nykänen P, Risteli J, Hanemaaijer R, Konttinen YT 2008 Erosive arthritis in a patient with pycnodysostosis: an experiment of nature. *Arthritis Rheum* 58:3394–3401
236. Svelander L, Erlandsson-Harris H, Astner L, Grabowska U, Klareskog L, Lindstrom E, Hewitt E 2009 Inhibition of cathepsin K reduces bone erosion, cartilage degradation and inflammation evoked by collagen-induced arthritis in mice. *Eur J Pharmacol* 613:155–162
237. Salminen-Mankonen HJ, Morko J, Vuorio E 2007 Role of cathepsin K in normal joints and in the development of arthritis. *Curr Drug Targets* 8:315–323
238. Hakala M, Risteli J, Aman S, Kautiainen H, Korpela M, Hannonen P, Leirisalo-Repo M, Laasonen L, Paimela L, Möttönen T 2008 Combination drug strategy in recent-onset rheumatoid arthritis suppresses collagen I degradation and is associated with retardation of radiological progression. *Scand J Rheumatol* 37:90–93
239. Sassi ML, Aman S, Hakala M, Luukkainen R, Risteli J 2003 Assay for cross-linked carboxyterminal telopeptide of type I collagen (ICTP) unlike CrossLaps assay reflects increased pathological degradation of type I collagen in rheumatoid arthritis. *Clin Chem Lab Med* 41:1038–1044
240. Chopin F, Garnero P, le Henanff A, Debiais F, Daragon A, Roux C, Sany J, Wendling D, Zarnitsky C, Ravaud P, Thomas T 2008 Long-term effects of infliximab on bone and cartilage turnover markers in patients with rheumatoid arthritis. *Ann Rheum Dis* 67:353–357
241. Kadono Y, Tanaka S, Nishino J, Nishimura K, Nakamura I, Miyazaki T, Takayanagi H, Nakamura K 2009 Rheumatoid arthritis associated with osteopetrosis. *Mod Rheumatol* 19:687–690
242. Russell RG, Watts NB, Ebetino FH, Rogers MJ 2008 Mechanisms of action of bisphosphonates: similarities and differences and their potential influence on clinical efficacy. *Osteoporos Int* 19:733–759
243. Chavassieux PM, Arlot ME, Reda C, Wei L, Yates AJ, Meunier PJ 1997 Histomorphometric assessment of the long-term effects of alendronate on bone quality and re-modeling in patients with osteoporosis. *J Clin Invest* 100:1475–1480
244. Recker RR, Weinstein RS, Chesnut 3rd CH, Schimmer RC, Mahoney P, Hughes C, Bonvoisin B, Meunier PJ 2004 Histomorphometric evaluation of daily and intermittent oral ibandronate in women with postmenopausal osteoporosis: results from the BONE study. *Osteoporos Int* 15:231–237
245. Recker RR, Delmas PD, Halse J, Reid IR, Boonen S, García-Hernandez PA, Supronik J, Lewiecki EM, Ochoa L, Miller P, Hu H, Mesenbrink P, Hartl F, Gasser J, Eriksen EF 2008 Effects of intravenous zoledronic acid once yearly on bone remodeling and bone structure. *J Bone Miner Res* 23:6–16
246. Nenonen A, Cheng S, Ivaska KK, Alatalo SL, Lehtimäki T, Schmidt-Gayk H, Uusi-Rasi K, Heinonen A, Kannus P, Sievanen H, Vuori I, Vaananen HK, Halleen JM 2005 Serum TRACP 5b is a useful marker for monitoring alendronate treatment: comparison with other markers of bone turnover. *J Bone Miner Res* 20:1804–1812
247. Hannon RA, Clowes JA, Eagleton AC, Al Hadari A, Eastell R, Blumsohn A 2004 Clinical performance of immunoreactive tartrate-resistant acid phosphatase isoform 5b as a marker of bone resorption. *Bone* 34:187–194
248. Muñoz-Torres M, Reyes-García R, Mezquita-Raya P, Fernández-García D, Alonso G, Luna Jde D, Ruiz-Requena ME, Escobar-Jiménez F 2009 Serum cathepsin K as a marker of bone metabolism in postmenopausal women treated with alendronate. *Maturitas* 64:188–192
249. D’Amelio P, Grimaldi A, Di Bella S, Tamone C, Brianza SZ, Ravazzoli MG, Bernabei P, Cristofaro MA, Pescarmona GP, Isaia G 2008 Risedronate reduces osteoclast precursors and cytokine production in postmenopausal osteoporotic women. *J Bone Miner Res* 23:373–379
250. D’Amelio P, Grimaldi A, Cristofaro MA, Ravazzoli M, Molinatti PA, Pescarmona GP, Isaia GC 1 December 2009 Alendronate reduces osteoclast precursors in osteoporosis. *Osteoporos Int* 10.1007/s00198-009-1129-1
251. Weinstein RS, Roberson PK, Manolagas SC 2009 Giant osteoclast formation and long-term oral bisphosphonate therapy. *N Engl J Med* 360:53–62
252. Mori S, Harruff R, Ambrosius W, Burr DB 1997 Trabecular bone volume and microdamage accumulation in the femoral heads of women with and without femoral neck fractures. *Bone* 21:521–526
253. Chapurlat RD, Arlot M, Burt-Pichat B, Chavassieux P, Roux JP, Portero-Muzy N, Delmas PD 2007 Microcrack frequency and bone remodeling in postmenopausal osteoporotic women on long-term bisphosphonates: a bone biopsy study. *J Bone Miner Res* 22:1502–1509
254. Burr DB, Allen MR 2008 Low bone turnover and microdamage? How and where to assess it? *J Bone Miner Res* 23:1150–1151; author reply 1152–1153
255. Ravn P, Hosking D, Thompson D, Cizza G, Wasnich RD, McClung M, Yates AJ, Bjarnason NH, Christiansen C 1999 Monitoring of alendronate treatment and prediction of effect on bone mass by biochemical markers in the early postmenopausal intervention cohort study. *J Clin Endocrinol Metab* 84:2363–2368
256. Ravn P, Clemmesen B, Christiansen C 1999 Biochemical markers can predict the response in bone mass during alendronate treatment in early postmenopausal women. Alen-

- dronate Osteoporosis Prevention Study Group. *Bone* 24: 237–244
257. Ravn P, Thompson DE, Ross PD, Christiansen C 2003 Biochemical markers for prediction of 4-year response in bone mass during bisphosphonate treatment for prevention of postmenopausal osteoporosis. *Bone* 33:150–158
  258. Eriksen EF, Melsen F, Sod E, Barton I, Chines A 2002 Effects of long-term risedronate on bone quality and bone turnover in women with postmenopausal osteoporosis. *Bone* 31:620–625
  259. Black DM, Schwartz AV, Ensrud KE, Cauley JA, Levis S, Quandt SA, Satterfield S, Wallace RB, Bauer DC, Palermo L, Wehren LE, Lombardi A, Santora AC, Cummings SR 2006 Effects of continuing or stopping alendronate after 5 years of treatment: the Fracture Intervention Trial Long-term Extension (FLEX): a randomized trial. *JAMA* 296: 2927–2938
  260. Durie BG, Katz M, Crowley J 2005 Osteonecrosis of the jaw and bisphosphonates. *N Engl J Med* 353:99–102
  261. Watts NB, Diab DL 2010 Long-term use of bisphosphonates in osteoporosis. *J Clin Endocrinol Metab* 95:1555–1565
  262. Khosla S, Burr D, Cauley J, Dempster DW, Ebeling PR, Felsenberg D, Gagel RF, Gilsanz V, Guise T, Koka S, McCauley LK, McGowan J, McKee MD, Mohla S, Pendrys DG, Raisz LG, Ruggiero SL, Shafer DM, Shum L, Silverman SL, Van Poznak CH, Watts N, Woo SB, Shane E 2007 Bisphosphonate-associated osteonecrosis of the jaw: report of a task force of the American Society for Bone and Mineral Research. *J Bone Miner Res* 22:1479–1491
  263. Cheng A, Daly CG, Logan RM, Stein B, Goss AN 2009 Alveolar bone and the bisphosphonates. *Aust Dent J* 54(Suppl 1):S51–S61
  264. Tran Van PT, Vignery A, Baron R 1982 Cellular kinetics of the bone remodeling sequence in the rat. *Anat Rec* 202: 445–451
  265. Tran Van P, Vignery A, Baron R 1982 An electron-microscopic study of the bone-remodeling sequence in the rat. *Cell Tissue Res* 225:283–292
  266. Baron R, Neff L, Tran Van P, Nefussi JR, Vignery A 1986 Kinetic and cytochemical identification of osteoclast precursors and their differentiation into multinucleated osteoclasts. *Am J Pathol* 122:363–378
  267. Favia G, Pilolli GP, Maiorano E 2009 Histologic and histomorphometric features of bisphosphonate-related osteonecrosis of the jaws: an analysis of 31 cases with confocal laser scanning microscopy. *Bone* 45:406–413
  268. Bedogni A, Blandamura S, Lokmic Z, Palumbo C, Ragazzo M, Ferrari F, Tregnaighi A, Pietrogrande F, Procopio O, Saia G, Ferretti M, Bedogni G, Chiarini L, Ferronato G, Ninfo V, Lo Russo L, Lo Muzio L, Nocini PF 2008 Bisphosphonate-associated jawbone osteonecrosis: a correlation between imaging techniques and histopathology. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 105: 358–364
  269. Hansen T, Kunkel M, Weber A, James Kirkpatrick C 2006 Osteonecrosis of the jaws in patients treated with bisphosphonates—histomorphologic analysis in comparison with infected osteoradionecrosis. *J Oral Pathol Med* 35:155–160
  270. Ramalho AC, Couttet P, Baudoin C, Morieux C, Graulet AM, de Vernejoul MC, Cohen-Solal ME 2002 Estradiol and raloxifene decrease the formation of multinucleate cells in human bone marrow cultures. *Eur Cytokine Netw* 13:39–45
  271. Saintier D, Burde MA, Rey JM, Maudelonde T, de Vernejoul MC, Cohen-Solal ME 2004  $17\beta$ -Estradiol down-regulates  $\beta 3$ -integrin expression in differentiating and mature human osteoclasts. *J Cell Physiol* 198:269–276
  272. Michael H, Härkönen PL, Väänänen HK, Hentunen TA 2005 Estrogen and testosterone use different cellular pathways to inhibit osteoclastogenesis and bone resorption. *J Bone Miner Res* 20:2224–2232
  273. Michael H, Härkönen PL, Kangas L, Väänänen HK, Hentunen TA 2007 Differential effects of selective oestrogen receptor modulators (SERMs) tamoxifen, ospemifene and raloxifene on human osteoclasts in vitro. *Br J Pharmacol* 151:384–395
  274. Taranta A, Brama M, Teti A, De luca V, Scandurra R, Spera G, Agnusdei D, Termine JD, Migliaccio S 2002 The selective estrogen receptor modulator raloxifene regulates osteoclast and osteoblast activity in vitro. *Bone* 30:368–376
  275. Oursler MJ, Osdoby P, Pyfferoen J, Riggs BL, Spelsberg TC 1991 Avian osteoclasts as estrogen target cells. *Proc Natl Acad Sci USA* 88:6613–6617
  276. Oursler MJ, Pederson L, Fitzpatrick L, Riggs BL, Spelsberg T 1994 Human giant cell tumors of the bone (osteoclastomas) are estrogen target cells. *Proc Natl Acad Sci USA* 91:5227–5231
  277. Oursler MJ, Pederson L, Pyfferoen J, Osdoby P, Fitzpatrick L, Spelsberg TC 1993 Estrogen modulation of avian osteoclast lysosomal gene expression. *Endocrinology* 132: 1373–1380
  278. Kremer M, Judd J, Rifkin B, Auszmann J, Oursler MJ 1995 Estrogen modulation of osteoclast lysosomal enzyme secretion. *J Cell Biochem* 57:271–279
  279. Parikka V, Lehenkari P, Sassi ML, Halleen J, Risteli J, Härkönen P, Väänänen HK 2001 Estrogen reduces the depth of resorption pits by disturbing the organic bone matrix degradation activity of mature osteoclasts. *Endocrinology* 142:5371–5378
  280. Ott SM, Oleksik A, Lu Y, Harper K, Lips P 2002 Bone histomorphometric and biochemical marker results of a 2-year placebo-controlled trial of raloxifene in postmenopausal women. *J Bone Miner Res* 17:341–348
  281. Steiniche T, Hasling C, Charles P, Eriksen EF, Mosekilde L, Melsen F 1989 A randomized study on the effects of estrogen/gestagen or high dose oral calcium on trabecular bone remodeling in postmenopausal osteoporosis. *Bone* 10:313–320
  282. Patel S, Pazianas M, Tobias J, Chambers TJ, Fox S, Chow J 1999 Early effects of hormone replacement therapy on bone. *Bone* 24:245–248
  283. Eriksen EF, Langdahl B, Vesterby A, Rungby J, Kassem M 1999 Hormone replacement therapy prevents osteoclastic hyperactivity: a histomorphometric study in early postmenopausal women. *J Bone Miner Res* 14:1217–1221
  284. Vedi S, Purdie DW, Ballard P, Bord S, Cooper AC, Compston JE 1999 Bone remodeling and structure in postmenopausal women treated with long-term, high-dose estrogen therapy. *Osteoporos Int* 10:52–58
  285. Ettinger B, Black DM, Mitlak BH, Knickerbocker RK, Nickelsen T, Genant HK, Christiansen C, Delmas PD,

- Zanchetta JR, Stakkestad J, Glüer CC, Krueger K, Cohen FJ, Eckert S, Ensrud KE, Avioli LV, Lips P, Cummings SR 1999 Reduction of vertebral fracture risk in postmenopausal women with osteoporosis treated with raloxifene: results from a 3-year randomized clinical trial. Multiple Outcomes of Raloxifene Evaluation (MORE) Investigators. *JAMA* 282:637–645
286. Meunier PJ, Vignot E, Garnero P, Confavreux E, Paris E, Liu-Leage S, Sarkar S, Liu T, Wong M, Draper MW 1999 Treatment of postmenopausal women with osteoporosis or low bone density with raloxifene. Raloxifene Study Group. *Osteoporos Int* 10:330–336
287. Lufkin EG, Whitaker MD, Nickelsen T, Argueta R, Caplan RH, Knickerbocker RK, Riggs BL 1998 Treatment of established postmenopausal osteoporosis with raloxifene: a randomized trial. *J Bone Miner Res* 13:1747–1754
288. Chambers TJ, Moore A 1983 The sensitivity of isolated osteoclasts to morphological transformation by calcitonin. *J Clin Endocrinol Metab* 57:819–824
289. Suzuki H, Nakamura I, Takahashi N, Ikuhara T, Matsuzaki K, Isogai Y, Hori M, Suda T 1996 Calcitonin-induced changes in the cytoskeleton are mediated by a signal pathway associated with protein kinase A in osteoclasts. *Endocrinology* 137:4685–4690
290. Shyu JF, Shih C, Tseng CY, Lin CH, Sun DT, Liu HT, Tsung HC, Chen TH, Lu RB 2007 Calcitonin induces podosome disassembly and detachment of osteoclasts by modulating Pyk2 and Src activities. *Bone* 40:1329–1342
291. Karsdal MA, Henriksen K, Bay-Jensen AC, Molloy B, Arnold M, John MR, Byrjalsen I, Azria M, Riis BJ, Qvist P, Christiansen C 2010 Lessons learned from the development of oral calcitonin: the first tablet formulation of a peptide in phase III clinical trials. *J Clin Pharmacol* 10.1177/0091270010372625
292. Karsdal MA, Byrjalsen I, Riis BJ, Christiansen C 2008 Investigation of the diurnal variation in bone resorption for optimal drug delivery and efficacy in osteoporosis with oral calcitonin. *BMC Clin Pharmacol* 8:12
293. Kung AW, Pasion EG, Sofiyan M, Lau EM, Tay BK, Lam KS, Wilawan K, Ongphiphadhanakul B, Thiebaud D 2006 A comparison of teriparatide and calcitonin therapy in postmenopausal Asian women with osteoporosis: a 6-month study. *Curr Med Res Opin* 22:929–937
294. Hwang JS, Tu ST, Yang TS, Chen JF, Wang CJ, Tsai KS 2006 Teriparatide vs. calcitonin in the treatment of Asian postmenopausal women with established osteoporosis. *Osteoporos Int* 17:373–378
295. Trovas GP, Lyritis GP, Galanos A, Raptou P, Constantelou E 2002 A randomized trial of nasal spray salmon calcitonin in men with idiopathic osteoporosis: effects on bone mineral density and bone markers. *J Bone Miner Res* 17:521–527
296. Chesnut 3rd CH, Majumdar S, Newitt DC, Shields A, Van Pelt J, Laschansky E, Azria M, Kriegman A, Olson M, Eriksen EF, Mindeholm L 2005 Effects of salmon calcitonin on trabecular microarchitecture as determined by magnetic resonance imaging: results from the QUEST study. *J Bone Miner Res* 20:1548–1561
297. Ikegame M, Ejiri S, Ozawa H 2004 Calcitonin-induced change in serum calcium levels and its relationship to osteoclast morphology and number of calcitonin receptors. *Bone* 35:27–33
298. Jiang Y, Zhao J, Geusens P, Liao EY, Adriaensens P, Gelan J, Azria M, Boonen S, Caulin F, Lynch JA, Ouyang X, Genant HK 2005 Femoral neck trabecular microstructure in ovariectomized ewes treated with calcitonin: MRI microscopic evaluation. *J Bone Miner Res* 20:125–130
299. Hoff AO, Catala-Lehnen P, Thomas PM, Priemel M, Rueger JM, Nasonkin I, Bradley A, Hughes MR, Ordonez N, Cote GJ, Amling M, Gagel RF 2002 Increased bone mass is an unexpected phenotype associated with deletion of the calcitonin gene. *J Clin Invest* 110:1849–1857
300. Dacquin R, Davey RA, Laplace C, Lévassieur R, Morris HA, Goldring SR, Gebre-Medhin S, Galson DL, Zajac JD, Karsenty G 2004 Amylin inhibits bone resorption while the calcitonin receptor controls bone formation *in vivo*. *J Cell Biol* 164:509–514
301. Davey RA, Turner AG, McManus JF, Chiu WS, Tjahjono F, Moore AJ, Atkins GJ, Anderson PH, Ma C, Glatt V, MacLean HE, Vincent C, Bouxsein M, Morris HA, Findlay DM, Zajac JD 2008 Calcitonin receptor plays a physiological role to protect against hypercalcemia in mice. *J Bone Miner Res* 23:1182–1193
302. Turner C, Tjahjono A, Moore A, Findlay D, Morris H, Zajac J, Davey R 2009 The calcitonin receptor expressed by osteoclasts plays a biological role to protect against induced hypercalcemia in mice. *J Bone Miner Res* 24(Suppl 1):1049 (Abstract)
303. Karsdal MA, Byrjalsen I, Leeming DJ, Delmas PD, Christiansen C 2008 The effects of oral calcitonin on bone collagen maturation: implications for bone turnover and quality. *Osteoporos Int* 19:1355–1361
304. Holtrop ME, King GJ, Cox KA, Reit B 1979 Time-related changes in the ultrastructure of osteoclasts after injection of parathyroid hormone in young rats. *Calcif Tissue Int* 27:129–135
305. Ma YL, Cain RL, Halladay DL, Yang X, Zeng Q, Miles RR, Chandrasekhar S, Martin TJ, Onyia JE 2001 Catabolic effects of continuous human PTH (1–38) *in vivo* is associated with sustained stimulation of RANKL and inhibition of osteoprotegerin and gene-associated bone formation. *Endocrinology* 142:4047–4054
306. Hodsman AB, Kiesel M, Adachi JD, Fraher LJ, Watson PH 2000 Histomorphometric evidence for increased bone turnover without change in cortical thickness or porosity after 2 years of cyclical hPTH(1–34) therapy in women with severe osteoporosis. *Bone* 27:311–318
307. Arlot M, Meunier PJ, Boivin G, Haddock L, Tamayo J, Correa-Rotter R, Jasqui S, Donley DW, Dalsky GP, Martin JS, Eriksen EF 2005 Differential effects of teriparatide and alendronate on bone remodeling in postmenopausal women assessed by histomorphometric parameters. *J Bone Miner Res* 20:1244–1253
308. Recker RR, Marin F, Ish-Shalom S, Mörnicke R, Hawkins F, Kapetanios G, de la Peña MP, Kekow J, Farrerons J, Sanz B, Oertel H, Stepan J 2009 Comparative effects of teriparatide and strontium ranelate on bone biopsies and biochemical markers of bone turnover in postmenopausal women with osteoporosis. *J Bone Miner Res* 24:1358–1368
309. Dobnig H, Turner RT 1995 Evidence that intermittent treatment with parathyroid hormone increases bone formation in adult rats by activation of bone lining cells. *Endocrinology* 136:3632–3638

310. Jilka RL, Weinstein RS, Bellido T, Roberson P, Parfitt AM, Manolagas SC 1999 Increased bone formation by prevention of osteoblast apoptosis with parathyroid hormone. *J Clin Invest* 104:439–446
311. Luiz de Freitas PH, Li M, Ninomiya T, Nakamura M, Ubaidus S, Oda K, Udagawa N, Maeda T, Takagi R, Amizuka N 2009 Intermittent PTH administration stimulates pre-osteoblastic proliferation without leading to enhanced bone formation in osteoclast-less *c-fos*( $-/-$ ) mice. *J Bone Miner Res* 24:1586–1597
312. Black DM, Greenspan SL, Ensrud KE, Palermo L, McGowan JA, Lang TF, Garner P, Bouxsein ML, Bilezikian JP, Rosen CJ 2003 The effects of parathyroid hormone and alendronate alone or in combination in postmenopausal osteoporosis. *N Engl J Med* 349:1207–1215
313. Finkelstein JS, Hayes A, Hunzelman JL, Wyland JJ, Lee H, Neer RM 2003 The effects of parathyroid hormone, alendronate, or both in men with osteoporosis. *N Engl J Med* 349:1216–1226
314. Ettinger B, San Martin J, Crans G, Pavo I 2004 Differential effects of teriparatide on BMD after treatment with raloxifene or alendronate. *J Bone Miner Res* 19:745–751
315. Johnston S, Andrews S, Shen V, Cosman F, Lindsay R, Dempster DW, Iida-Klein A 2007 The effects of combination of alendronate and human parathyroid hormone(1-34) on bone strength are synergistic in the lumbar vertebra and additive in the femur of C57BL/6J mice. *Endocrinology* 148:4466–4474
316. Samadfam R, Xia Q, Goltzman D 2007 Co-treatment of PTH with osteoprotegerin or alendronate increases its anabolic effect on the skeleton of oophorectomized mice. *J Bone Miner Res* 22:55–63
317. Meunier PJ, Roux C, Seeman E, Ortolani S, Badurski JE, Spector TD, Cannata J, Balogh A, Lemmel EM, Pors-Nielsen S, Rizzoli R, Genant HK, Reginster JY 2004 The effects of strontium ranelate on the risk of vertebral fracture in women with postmenopausal osteoporosis. *N Engl J Med* 350:459–468
318. Seeman E, Vellas B, Benhamou C, Aquino JP, Semler J, Kaufman JM, Hozzowski K, Varela AR, Fiore C, Brixen K, Reginster JY, Boonen S 2006 Strontium ranelate reduces the risk of vertebral and nonvertebral fractures in women eighty years of age and older. *J Bone Miner Res* 21:1113–1120
319. Reginster JY, Seeman E, De Vernejoul MC, Adami S, Compston J, Phenekos C, Devogelaer JP, Curiel MD, Sawicki A, Goemaere S, Sorensen OH, Felsenberg D, Meunier PJ 2005 Strontium ranelate reduces the risk of nonvertebral fractures in postmenopausal women with osteoporosis: Treatment of Peripheral Osteoporosis (TROPOS) study. *J Clin Endocrinol Metab* 90:2816–2822
320. Seeman E, Boonen S, Borgström F, Vellas B, Aquino JP, Semler J, Benhamou CL, Kaufman JM, Reginster JY 2010 Five years treatment with strontium ranelate reduces vertebral and nonvertebral fractures and increases the number and quality of remaining life-years in women over 80 years of age. *Bone* 46:1038–1042
321. Ferrari S 2009 Continuous broad protection against osteoporotic fractures with strontium ranelate. *Rheumatology (Oxford)* 48(Suppl 4):iv20–iv24
322. Arlot ME, Jiang Y, Genant HK, Zhao J, Burt-Pichat B, Roux JP, Delmas PD, Meunier PJ 2008 Histomorphometric and microCT analysis of bone biopsies from postmenopausal osteoporotic women treated with strontium ranelate. *J Bone Miner Res* 23:215–222
323. Meunier PJ, Slosman DO, Delmas PD, Sebert JL, Brandi ML, Albanese C, Lorenc R, Pors-Nielsen S, De Vernejoul MC, Roces A, Reginster JY 2002 Strontium ranelate: dose-dependent effects in established postmenopausal vertebral osteoporosis—a 2-year randomized placebo controlled trial. *J Clin Endocrinol Metab* 87:2060–2066
324. Axmann R, Böhm C, Krönke G, Zwerina J, Smolen J, Schett G 2009 Inhibition of interleukin-6 receptor directly blocks osteoclast formation in vitro and in vivo. *Arthritis Rheum* 60:2747–2756
325. Bruyère O, Collette J, Rizzoli R, Decock C, Ortolani S, Cormier C, Detilleux J, Reginster JY 2010 Relationship between 3-month changes in biochemical markers of bone remodelling and changes in bone mineral density and fracture incidence in patients treated with strontium ranelate for 3 years. *Osteoporos Int* 21:1031–1036
326. Bonnelye E, Chabadel A, Saltel F, Jurdic P 2008 Dual effect of strontium ranelate: stimulation of osteoblast differentiation and inhibition of osteoclast formation and resorption in vitro. *Bone* 42:129–138
327. Canalis E, Hott M, Deloffre P, Tsouderos Y, Marie PJ 1996 The divalent strontium salt S12911 enhances bone cell replication and bone formation in vitro. *Bone* 18:517–523
328. Takahashi N, Sasaki T, Tsouderos Y, Suda T 2003 S 12911-2 inhibits osteoclastic bone resorption in vitro. *J Bone Miner Res* 18:1082–1087
329. Hurtel-Lemaire AS, Mentaverri R, Caudrillier A, Cournaire F, Wattel A, Kamel S, Terwilliger EF, Brown EM, Brazier M 2009 The calcium-sensing receptor is involved in strontium ranelate-induced osteoclast apoptosis. New insights into the associated signaling pathways. *J Biol Chem* 284:575–584
330. Brennan TC, Rybchyn MS, Green W, Atwa S, Conigrave AD, Mason RS 2009 Osteoblasts play key roles in the mechanisms of action of strontium ranelate. *Br J Pharmacol* 157:1291–1300
331. Engvall IL, Svensson B, Tengstrand B, Brismar K, Hafström I 2008 Impact of low-dose prednisolone on bone synthesis and resorption in early rheumatoid arthritis: experiences from a two-year randomized study. *Arthritis Res Ther* 10:R128
332. Caplan L, Saag KG 2009 Glucocorticoids and the risk of osteoporosis. *Expert Opin Drug Saf* 8:33–47
333. Canalis E, Mazziotti G, Giustina A, Bilezikian JP 2007 Glucocorticoid-induced osteoporosis: pathophysiology and therapy. *Osteoporos Int* 18:1319–1328
334. van Brussel MS, Bultink IE, Lems WF 2009 Prevention of glucocorticoid-induced osteoporosis. *Expert Opin Pharmacother* 10:997–1005
335. Weinstein RS, Jilka RL, Parfitt AM, Manolagas SC 1998 Inhibition of osteoblastogenesis and promotion of apoptosis of osteoblasts and osteocytes by glucocorticoids. Potential mechanisms of their deleterious effects on bone. *J Clin Invest* 102:274–282
336. Hofbauer LC, Zeitz U, Schoppert M, Skalicky M, Schüller C, Stolina M, Kostenuik PJ, Erben RG 2009 Prevention of glucocorticoid-induced bone loss in mice by inhibition of RANKL. *Arthritis Rheum* 60:1427–1437
337. Jia D, O'Brien CA, Stewart SA, Manolagas SC, Weinstein

- RS 2006 Glucocorticoids act directly on osteoclasts to increase their life span and reduce bone density. *Endocrinology* 147:5592–5599
338. Yao W, Cheng Z, Busse C, Pham A, Nakamura MC, Lane NE 2008 Glucocorticoid excess in mice results in early activation of osteoclastogenesis and adipogenesis and prolonged suppression of osteogenesis: a longitudinal study of gene expression in bone tissue from glucocorticoid-treated mice. *Arthritis Rheum* 58:1674–1686
339. Weinstein RS, Chen JR, Powers CC, Stewart SA, Landes RD, Bellido T, Jilka RL, Parfitt AM, Manolagas SC 2002 Promotion of osteoclast survival and antagonism of bisphosphonate-induced osteoclast apoptosis by glucocorticoids. *J Clin Invest* 109:1041–1048
340. Kim HJ, Zhao H, Kitaura H, Bhattacharyya S, Brewer JA, Muglia LJ, Ross FP, Teitelbaum SL 2006 Glucocorticoids suppress bone formation via the osteoclast. *J Clin Invest* 116:2152–2160
341. Sivagurunathan S, Muir MM, Brennan TC, Seale JP, Mason RS 2005 Influence of glucocorticoids on human osteoclast generation and activity. *J Bone Miner Res* 20:390–398
342. Hofbauer LC, Gori F, Riggs BL, Lacey DL, Dunstan CR, Spelsberg TC, Khosla S 1999 Stimulation of osteoprotegerin ligand and inhibition of osteoprotegerin production by glucocorticoids in human osteoblastic lineage cells: potential paracrine mechanisms of glucocorticoid-induced osteoporosis. *Endocrinology* 140:4382–4389
343. Soares-Schanoski A, Gómez-Piña V, del Fresno C, Rodríguez-Rojas A, García F, Glaría A, Sánchez M, Vallejo-Cremades MT, Baos R, Fuentes-Prior P, Arnalich F, López-Collazo E 2007 6-Methylprednisolone down-regulates IRAK-M in human and murine osteoclasts and boosts bone-resorbing activity: a putative mechanism for corticoid-induced osteoporosis. *J Leukoc Biol* 82:700–709
344. Søb K, Delaissé JM 30 April 2010 Glucocorticoids maintain human osteoclasts in the active mode of their resorption cycle. *J Bone Miner Res* 10.1002/jbmr.113
345. Dovio A, Perazzolo L, Osella G, Ventura M, Termine A, Milano E, Bertolotto A, Angeli A 2004 Immediate fall of bone formation and transient increase of bone resorption in the course of high-dose, short-term glucocorticoid therapy in young patients with multiple sclerosis. *J Clin Endocrinol Metab* 89:4923–4928
346. Minisola S, Del Fiacco R, Piemonte S, Iorio M, Mascia ML, Fidanza F, Cipriani C, Raso I, Porfiri ML, Francucci CM, D'Erasmo E, Romagnoli E 2008 Biochemical markers in glucocorticoid-induced osteoporosis. *J Endocrinol Invest* 31:28–32
347. Dalle Carbonare L, Bertoldo F, Valenti MT, Zenari S, Zanatta M, Sella S, Giannini S, Cascio VL 2005 Histomorphometric analysis of glucocorticoid-induced osteoporosis. *Micron* 36:645–652
348. Stellan AJ, Webb A, Compston JE 1988 Bone histomorphometry and structure in corticosteroid treated chronic active hepatitis. *Gut* 29:378–384
349. Stoch SA, Saag KG, Greenwald M, Sebba AI, Cohen S, Verbruggen N, Giezek H, West J, Schnitzer TJ 2009 Once-weekly oral alendronate 70 mg in patients with glucocorticoid-induced bone loss: a 12-month randomized, placebo-controlled clinical trial. *J Rheumatol* 36:1705–1714
350. Bikle DD, Sakata T, Leary C, Elalieh H, Ginzinger D, Rosen CJ, Beamer W, Majumdar S, Halloran BP 2002 Insulin-like growth factor I is required for the anabolic actions of parathyroid hormone on mouse bone. *J Bone Miner Res* 17:1570–1578
351. Cummings SR, San Martin J, McClung MR, Siris ES, Eastell R, Reid IR, Delmas P, Zoog HB, Austin M, Wang A, Kutilek S, Adami S, Zanchetta J, Libanati C, Siddhanti S, Christiansen C 2009 Denosumab for prevention of fractures in postmenopausal women with osteoporosis. *N Engl J Med* 361:756–765
352. Kostenuik PJ, Nguyen HQ, McCabe J, Warmington KS, Kurahara C, Sun N, Chen C, Li L, Cattley RC, Van G, Scully S, Elliott R, Grisanti M, Morony S, Tan HL, Asuncion F, Li X, Ominsky MS, Stolina M, Dwyer D, Dougall WC, Hawkins N, Boyle WJ, Simonet WS, Sullivan JK 2009 Denosumab, a fully human monoclonal antibody to RANKL, inhibits bone resorption and increases BMD in knock-in mice that express chimeric (murine/human) RANKL. *J Bone Miner Res* 24:182–195
353. Bone HG, Bolognese MA, Yuen CK, Kendler DL, Wang H, Liu Y, San Martin J 2008 Effects of denosumab on bone mineral density and bone turnover in postmenopausal women. *J Clin Endocrinol Metab* 93:2149–2157
354. McClung MR, Lewiecki EM, Cohen SB, Bolognese MA, Woodson GC, Moffett AH, Peacock M, Miller PD, Lederer SN, Chesnut CH, Lain D, Kivitz AJ, Holloway DL, Zhang C, Peterson MC, Bekker PJ 2006 Denosumab in postmenopausal women with low bone mineral density. *N Engl J Med* 354:821–831
355. Ominsky MS, Schroeder J, Jollette J, Smith SY, Farrell DJ, Atkinson JE, Kostenuik PJ 2007 Decreased bone turnover and porosity are associated with improved bone strength in ovariectomized (OVX) cynomolgus monkeys treated with denosumab, a fully human RANKL antibody. *J Bone Miner Res* 22:S126 (Abstract)
356. Sassi ML, Eriksen H, Risteli L, Niemi S, Mansell J, Gowen M, Risteli J 2000 Immunochemical characterization of assay for carboxyterminal telopeptide of human type I collagen: loss of antigenicity by treatment with cathepsin K. *Bone* 26:367–373
357. Fuller K, Lawrence KM, Ross JL, Grabowska UB, Shiroo M, Samuelsson B, Chambers TJ 2008 Cathepsin K inhibitors prevent matrix-derived growth factor degradation by human osteoclasts. *Bone* 42:200–211
358. Chappard D, Libouban H, Mindeholm L, Baslé MF, Legendrand E, Audran M 2010 The cathepsin K inhibitor AAE581 induces morphological changes in osteoclasts of treated patients. *Microsc Res Tech* 73:726–732
359. Adams CS, Shapiro IM 2003 Mechanisms by which extracellular matrix components induce osteoblast apoptosis. *Connect Tissue Res* 44(Suppl 1):230–239
360. Walker EC, McGregor NE, Poulton IJ, Pompolo S, Allan EH, Quinn JM, Gillespie MT, Martin TJ, Sims NA 2008 Cardiotrophin-1 is an osteoclast-derived stimulus of bone formation required for normal bone remodeling. *J Bone Miner Res* 23:2025–2032
361. Gottschalk IB, Jeppesen PB, Hartmann B, Holst JJ, Henriksen DB 2008 Effects of treatment with glucagon-like peptide-2 on bone resorption in colectomized patients with distal ileostomy or jejunostomy and short-bowel syndrome. *Scand J Gastroenterol* 43:1304–1310
362. Gottschalk IB, Jeppesen PB, Holst JJ, Henriksen DB 2008



- Reduction in bone resorption by exogenous glucagon-like peptide-2 administration requires an intact gastrointestinal tract. *Scand J Gastroenterol* 43:929–937
363. **Sørensen MG, Henriksen K, Neutzsky-Wulff AV, Dziegiel MH, Karsdal MA** 2007 Diphyllin, a novel and naturally potent V-ATPase inhibitor, abrogates acidification of the osteoclastic resorption lacunae and bone resorption. *J Bone Miner Res* 22:1640–1648
  364. **Schaller S, Henriksen K, Sveigaard C, Heegaard AM, Hélix N, Stahlhut M, Ovejero MC, Johansen JV, Solberg H, Andersen TL, Hougaard D, Berryman M, Shiødt CB, Sørensen BH, Lichtenberg J, Christophersen P, Foged NT, Delaissé JM, Engsig MT, Karsdal MA** 2004 The chloride channel inhibitor n53736 prevents bone resorption in ovariectomized rats without changing bone formation. *J Bone Miner Res* 19:1144–1153
  365. **Visentin L, Dodds RA, Valente M, Misiano P, Bradbeer JN, Oneta S, Liang X, Gowen M, Farina C** 2000 A selective inhibitor of the osteoclastic V-H(+)-ATPase prevents bone loss in both thyroparathyroidectomized and ovariectomized rats. *J Clin Invest* 106:309–318
  366. **Rzeszutek K, Sarraf F, Davies JE** 2003 Proton pump inhibitors control osteoclastic resorption of calcium phosphate implants and stimulate increased local reparative bone growth. *J Craniofac Surg* 14:301–307
  367. **Schaller S, Henriksen K, Sørensen MG, Karsdal MA** 2005 The role of chloride channels in osteoclasts: ClC-7 as a target for osteoporosis treatment. *Drug News Perspect* 18:489–495
  368. **Brown EM** 2007 The calcium-sensing receptor: physiology, pathophysiology and CaR-based therapeutics. *Subcell Biochem* 45:139–167
  369. **Martin TJ** 2005 Osteoblast-derived PTHrP is a physiological regulator of bone formation. *J Clin Invest* 115:2322–2324
  370. **Kramer I, Keller H, Leupin O, Kneissel M** 2010 Does osteocytic SOST suppression mediate PTH bone anabolism? *Trends Endocrinol Metab* 21:237–244
  371. **Thompson ER, Baylink DJ, Wergedal JE** 1975 Increases in number and size of osteoclasts in response to calcium or phosphorus deficiency in the rat. *Endocrinology* 97:283–289
  372. **Howard GA, Bottemiller BL, Turner RT, Rader JI, Baylink DJ** 1981 Parathyroid hormone stimulates bone formation and resorption in organ culture: evidence for a coupling mechanism. *Proc Natl Acad Sci USA* 78:3204–3208
  373. **Lazowski DA, Fraher LJ, Hodsman A, Steer B, Modrowski D, Han VK** 1994 Regional variation of insulin-like growth factor-I gene expression in mature rat bone and cartilage. *Bone* 15:563–576
  374. **Robinson JA, Riggs BL, Spelsberg TC, Oursler MJ** 1996 Osteoclasts and transforming growth factor- $\beta$ : estrogen-mediated isoform-specific regulation of production. *Endocrinology* 137:615–621
  375. **Karsdal MA, Fjording MS, Foged NT, Delaissé JM, Lochter A** 2001 Transforming growth factor- $\beta$ -induced osteoblast elongation regulates osteoclastic bone resorption through a p38 mitogen-activated protein kinase- and matrix metalloproteinase-dependent pathway. *J Biol Chem* 276:39350–39358
  376. **Mundy GR, Bonewald LF** 1990 Role of TGF  $\beta$  in bone remodeling. *Ann NY Acad Sci* 593:91–97
  377. **Baylink DJ, Finkelman RD, Mohan S** 1993 Growth factors to stimulate bone formation. *J Bone Miner Res* 8(Suppl 2):S565–S572
  378. **Hayden JM, Mohan S, Baylink DJ** 1995 The insulin-like growth factor system and the coupling of formation to resorption. *Bone* 17:93S–98S
  379. **Janssens K, ten Dijke P, Janssens S, Van Hul W** 2005 Transforming growth factor- $\beta$ 1 to the bone. *Endocr Rev* 26:743–774
  380. **Zhang M, Xuan S, Bouxsein ML, von Stechow D, Akeno N, Faugere MC, Malluche H, Zhao G, Rosen CJ, Efstratiadis A, Clemens TL** 2002 Osteoblast-specific knockout of the insulin-like growth factor (IGF) receptor gene reveals an essential role of IGF signaling in bone matrix mineralization. *J Biol Chem* 277:44005–44012