

Review Article

Overview of Alzheimer's Disease and Some Therapeutic Approaches Targeting A β by Using Several Synthetic and Herbal Compounds

Sandeep Kumar Singh,¹ Saurabh Srivastav,¹ Amarish Kumar Yadav,¹ Saripella Srikrishna,¹ and George Perry²

¹Department of Biochemistry, Faculty of Science, Banaras Hindu University, Varanasi 221 005, India

²Department of Biology, The University of Texas at San Antonio, San Antonio, TX 78249, USA

Correspondence should be addressed to Saripella Srikrishna; skrishna@bhu.ac.in and George Perry; george.perry@utsa.edu

Received 12 August 2015; Accepted 5 November 2015

Academic Editor: Anne-Laure Bulteau

Copyright © 2016 Sandeep Kumar Singh et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Alzheimer's disease (AD) is a complex age-related neurodegenerative disease. In this review, we carefully detail amyloid- β metabolism and its role in AD. We also consider the various genetic animal models used to evaluate therapeutics. Finally, we consider the role of synthetic and plant-based compounds in therapeutics.

1. Alzheimer's Disease (Overview)

Alzheimer's disease (AD) is the most common neurodegenerative disorder characterized by progressive memory loss. In 1907, Alois Alzheimer was the first to report a case of intellectual deterioration with the histological findings of senile plaques and neurofibrillary tangles [1]. An estimated 4.5 million Americans have AD and, as the elderly population continues to grow, the prevalence could increase by threefold to 13.2 million by 2050 [2]. The scenario seems more alarming, as it is estimated that by the year 2020, approximately 70% of the world's population aged 60 and above will be living in developing countries, with 14.2% in India. Previous reports suggest that age-adjusted prevalence of AD to be 1.91% in a community residing population in a southern Indian province of Kerala, as a part of the cognition in older adults in Trivandrum (COAT) study [3]. The reported incidence rates for AD have been lower in Asian countries than in the industrialized world [4, 5]. The impact of AD on health care costs, including direct and indirect medical and social services, is currently estimated to be greater than \$100 billion per year [6]. In addition, there is currently no cure for AD; therefore, the major challenges for the near future will be

the development of new therapies and therapeutic targets for disease modification and prevention.

To date, there are two major neuropathological features for the diagnosis of AD, namely, the extracellular plaque formation and neurofibrillary tangles (NFTs) formation intracellularly. The former comprises amyloid- β protein (A β) while the latter involves neurofibrillary tangles (NFTs) consisting of paired helical filaments of hyperphosphorylated tau protein. These histopathological lesions are mainly confined in the hippocampus region of the brain and in the cerebral cortex, the two large forebrain domains related to memory and other higher cognitive functions. The characteristic pathology in due course leads to the typical clinical symptoms, for example memory impairment, general cognitive decline, and personality changes associated with AD. The causes of AD are still rather poorly known, with different etiologies (e.g., A β overproduction, genetics, A β impaired clearance, and NFT formation) leading to senile plaques, neurofibrillary tangle (NFT) formation, and extensive neuronal death. However, several studies and evidence point to A β as critical in the pathogenesis of AD. According to the amyloid cascade hypothesis, A β peptides form aggregates and toxic assemblies which initiate several processes leading

to neuronal dysfunction and ultimately large-scale cell death [7]. The prevalence of AD varies among several different factors, including age, genetics, comorbidities, and education level. There is no way to absolutely diagnose AD without performing an autopsy. There is no cure for AD; however promising research and development for early detection and treatment is underway.

1.1. History. Alzheimer's disease was discovered by a German Neurobiologist and Psychiatrist named Alois Alzheimer in 1906 [9]. Before Alzheimer's 1906 discovery, both scientists and the nonscience community viewed dementia as a "natural" progression of age, and "senility was accepted as a part of aging" according to Natalie Whaley in her honours thesis on the social history of Alzheimer's disease. First time, AD was observed in Auguste D., a 51-year-old woman. Her family observed some unusual behavioural changes in her personality and then they brought her to Dr. Alois Alzheimer in 1901. The family reported problems with memory loss, speaking difficulty, loss of good judgement, disorientation to time and place, and problem with abstract thinking. Later on, Dr. Alzheimer described that she is having an aggressive form of dementia, memory impairment, problem using language, and behavioural changes [10]. Dr. Alzheimer also noted many other abnormal symptoms, including rapid mood swing, personality changes, loss of initiative, sleeping longer than usual, and loss of interest in usual activity [11]. Dr. Alois followed her for about five years, until her death, in 1906. After death, he performed an autopsy and found dramatic shrinkage of the cerebral cortex, deposition of fat bodies in blood vessels, and atrophied brain cells [9]. He discovered neurofibrillary tangles and senile plaques, which have become indicative of AD [11]. The condition was first discussed in the medical literature in 1907 and named after Alzheimer in 1910.

1.2. Pathology and Pathophysiology of Alzheimer's Disease. The brain of AD patient often shows marked atrophy, with broadened sulci and shrinkage of the gyri. In the majority of cases, every part of the cerebral cortex is involved. However, the occipital lobe is often relatively spared. The cortical ribbon may be thinned and ventricular dilatation is apparent, especially in the temporal horn, due to atrophy of the amygdala and hippocampus. During the last couple of decades, the pathology of AD has been extensively studied whereby animal models have provided valuable information in understanding of the pathogenic mechanisms of AD. Furthermore, AD pathology can be divided into three broad sections: (a) positive lesions (lesions related to accumulation), (b) negative lesions (those that are due to losses), and (c) inflammation and plasticity (those that are due to the reactive processes). The first category involving positive lesions is very common and easy to detect and constitutes the basis of the diagnosis. Both the neuronal and synapse loss are difficult to evaluate, as they do not belong to the diagnostic criteria but could be the alterations that are more directly related to the cognitive deficit. Furthermore, microscopic studies on AD brain revealed significant neuronal loss, in addition to

shrinkage of large cortical neurons. Many neuropathologists believe that loss of synapses neurons in association with shrinkage of the dendritic arbor of large neurons is the critical pathological substrate. The main neuropathological hallmarks of AD are senile plaques and neurofibrillary tangles, although these two are not unique to AD and can be found in other human neurodegenerative disorders and in clinically normal individuals as well. Apart from senile plaques, two other types of amyloid-related plaques are found in the brains of AD patients: burnt-out plaques, which consist of an isolated dense amyloid core and diffuse plaques, which contain poorly defined amyloid but no well-circumscribed amyloid core. It is believed that the abnormal processing of the amyloid- β protein precursor through amyloidogenic pathway results in different fragments, the most toxic of which is the $A\beta_{42}$ peptide [12]. $A\beta_{42}$ readily self-aggregates and forms clumps of insoluble fibrils in the brain, thereby triggering the formation of senile plaques. It has been postulated that $A\beta_{42}$ is mainly responsible for initiating a cascade of events leading to neuronal dysfunction, later followed by death. Although increasing evidence supports the hypothesis that the accumulation of $A\beta$ is very decisive to the pathogenesis of AD [13], some investigators believe that $A\beta$ is not exclusively responsible for the neuronal alterations that underlie its symptoms [14].

Neurofibrillary tangles (NFTs) are the other important characteristic histopathological features of AD. Neurofibrillary tangles are found inside neurons and are composed of paired helical filaments of hyperphosphorylated microtubule-associated protein tau (MAPT). Accumulation of NFTs intracellularly may cause dysfunction of the normal cytoskeletal architecture of neurons with subsequent death. Senile plaques ($A\beta$) and neurofibrillary tangles (NFTs) are not distributed evenly across the brain in AD but are confined to vulnerable neural systems.

Other pathological modifications commonly discovered in the brains of AD patients include granulovacuolar degeneration, neuropil threads, and amyloid angiopathy. The latter one is a distinct vascular lesion and found in many AD brains, consisting of amyloid deposition in the walls of small- to medium-sized cortical and leptomeningeal arteries due to which the involved vessels may become compromised with resultant hemorrhage.

After microscopic examination, observation of sufficient amount of senile plaques and neurofibrillary tangles suggests important pathological criteria for the diagnosis of AD. Because of the presence of amyloid- β in senile plaques and to a variable degree in cerebral blood vessels in the AD brain, the roles of this important protein and its precursor peptide, amyloid- β protein precursor, have been widely investigated [15], although the exact nature of their roles in the pathogenesis of AD remains unclear. Increasingly, the importance of differential neuronal vulnerability and the relationship of this to the morphological and biochemical characteristics of AD are being recognized. The most consistent neurochemical change associated with AD has been the well-documented decline in cholinergic activity that has inspired many attempts to treat AD with cholinergic drugs. However, additional deficiencies

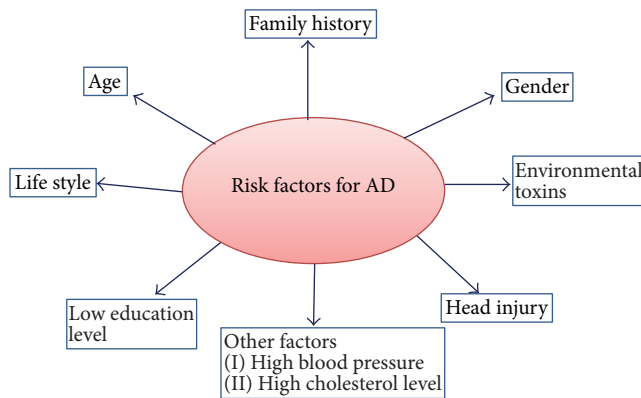


FIGURE 1: Showing different causal and risk factors for Alzheimer's disease.

in glutamate, norepinephrine, serotonin, somatostatin, and corticotrophin-releasing factors have also been described.

1.3. Risk Factors for AD. While scientists know that distortion of nerve cells in case of Alzheimer's disease occurs, why this happens is still unknown. However, they have discovered certain risk factors that increase the likelihood of developing Alzheimer's (Figure 1).

1.3.1. Age. With increasing age, the risk of developing AD becomes higher. Most patients develop AD after the age of 65 years. The risk of developing AD reaches 50% for individuals above the age of 85 years. Statistically speaking, about 5% of men and women between the ages of 65 and 74 have Alzheimer's disease, and nearly half of those aged 85 and older may have the disease. Despite its prevalence, Alzheimer's disease is not a normal part of aging. The age-specific incidence rates for Alzheimer's disease demonstrate a doubling of incidence for about every six years of added life, which indicates an exponential increasing risk with increasing age of individuals. This exponential risk is somewhat similar across studies, regardless of geographic region, even if the underlying absolute incidence rate differs.

1.3.2. Familial History and Genetics. Another risk factor involves family history. Researchers have shown that those who have a parent, brother, or sister with AD are more likely to develop the disease than individuals who do not have a first-degree relative with AD. The vast majority of AD cases are not genetically inherited, although some genes may act as risk factors [16]. The risk increases if more than one family member have the illness. Genetically identified forms of AD, which usually have an onset before the age of 65, have been identified and account for 0.1% of disease cases [17]. Scientists have identified three genes that described people who will develop Alzheimer's, but only a very small percentage of individuals with AD (about 1%) carry these genes. The apolipoprotein E (APOE- ϵ 4) is carried by about 25% of individuals and increases the risk of developing AD, but it is not sure that individuals with APOE-E4 will develop

the disease. Scientists believe that the vast majority of AD cases are caused by a complex combination of genetic and nongenetic determinants.

1.3.3. Other Risk Factors. Besides age, family history, and genetics, other important risk factors exist that may contribute to AD risk. In this context, some promising research suggests that strategies for keeping and living overall healthy aging may help maintain brain health and may even provide some protection against AD. These factors include eating habits, healthy lifestyle, staying socially and physically active, and avoiding excess alcohol and tobacco.

Some of the strongest evidence links brain health to heart health. The risk of developing Alzheimer's disease or vascular dementia appears to be increased by many conditions that damage the heart and blood vessels. These include heart disease, diabetes, stroke, high blood pressure, and high cholesterol. It is a common saying in medical practice that "work with your doctor to monitor your heart health and treat any problems that arise." Further, studies of donated brain tissue provide additional evidence for the heart-head connection. These studies suggest that plaques and tangles are more likely to cause Alzheimer's symptoms if strokes or damage to the brain's blood vessels is also present.

2. Different Hypotheses Postulated Related to Alzheimer's Disease

2.1. The Tau Hypothesis of Alzheimer's Disease. Tau protein plays a critical role in pathophysiology of AD. The tau hypothesis focuses primarily on the role of the microtubule binding tau protein, which is the main component of NFTs in AD. Hyperphosphorylation of tau protein results in NFT formation. This hypothesis proposes a mechanism for neurotoxicity based on the loss of microtubule-stabilizing tau protein that leads to degradation of the cytoskeleton [18]. However, it is not clear whether tau hyperphosphorylation is responsible or is caused by the formation of abnormal helical filaments [19]. Tauopathy-like diseases also support the tau hypothesis in which the same protein is significantly misfolded [20]. However, a majority of research groups support the alternative hypothesis that amyloid- β is the primary causative agent for AD [19].

2.2. The Cholinergic Hypothesis of Alzheimer's Disease. The cholinergic hypothesis is the oldest AD hypothesis [21]. This hypothesis proposed that AD is caused by the reduced synthesis of a neurotransmitter called acetylcholine in neurons. The cholinergic hypothesis was formulated over 30 years ago and suggests that a dysfunction of acetylcholine-containing neurons in the basal forebrain contributes substantially to the cognitive decline observed in AD patients [22]. In addition to the dysfunction and neuronal loss in basal forebrain regions, confirmation of cholinergic losses comes from studies that report deterioration in the activity of acetylcholine esterase (AChE) and choline acetyltransferase (ChAT), reduced acetylcholine (ACh) release, and decreased level of nicotinic and muscarinic receptors in the AD affected

brain [23]. This observation led to the formation of the cholinergic hypothesis, considered the oldest hypothesis of AD [24, 25]. The cholinergic hypothesis has not had widespread support, largely because medications intended to treat acetylcholine deficiency have not been very effective, although 4 of the 5 approved drugs work on just this mechanism. Other cholinergic effects have also been proposed, for example, initiation of large-scale aggregation of amyloid leading to generalized neuroinflammation [26].

2.3. The Mitochondrial Cascade Hypothesis. The mitochondrial cascade hypothesis was first proposed by Swerdlow and Khan, 2004, which postulates that mitochondrial dysfunction is the primary cause of $A\beta$ deposition, neurofibrillary tangle (NFT) formation, and synaptic degeneration in AD [27]. The mitochondrial cascade hypothesis takes several conceptual liberties. It assumes that similar physiologic mechanisms underlie AD and brain aging. It postulates that because AD mitochondrial dysfunction is systemic, it cannot simply represent a consequence of neurodegeneration. The mitochondrial cascade hypothesis argues that non-Mendelian genetic factors contribute to nonautosomal dominant AD. Finally, it posits that AD brain mitochondrial dysfunction drives to amyloidosis, tau phosphorylation, and cell cycle reentry. Mitochondrial dysfunction is observed in several AD tissues [28], including platelets, fibroblast, mitochondria, and brain. There are basically three mitochondrial enzymes that are found to be defective. This includes reduced activities of α -ketoglutarate dehydrogenase complex, cytochrome oxidase, and pyruvate dehydrogenase complex [29]. Through special analysis of AD brains, level of cytochrome oxidase is found to be normal, but the enzyme itself is structurally altered [30]. In AD, oxidative stress and proteasome dysfunction have been postulated to facilitate mitochondrial dysfunction [31]. Also, studies on cytoplasmic hybrid (cybrid) indicate that mtDNA at least, in part, accounts for reduced cytochrome oxidase activity in AD [32].

2.4. Amyloid Cascade Hypothesis. Although the exact cause of AD is still a matter of debate, the amyloid cascade hypothesis is the best accepted and most studied hypothesis among those mentioned above. The presence of amyloid plaques is considered to be the main characteristic of AD pathology. The primary constituent of senile plaques identified so far is $A\beta$ peptide, which is produced on account of proteolytic processing of the amyloid- β protein precursor ($A\beta$ PP) by β - and γ -secretases [33, 34]. Furthermore, cloning of the $A\beta$ PP gene [35] has allowed the disease to be examined at molecular and biochemical levels. Subsequently, mapping of several familial forms of AD (fAD) mutations in the $A\beta$ PP gene [36], the association of AD with Down's syndrome, and higher prevalence of AD with increased numbers of $A\beta$ PP all established the critical role of $A\beta$ PP in AD pathogenesis [33, 37, 38]. The central role of $A\beta$ PP in AD etiology is further supported by the identification of fAD mutations in presenilin 1 (PS1), which involves $A\beta$ PP cleavage and generates $A\beta$ and AICD fragments.

In the early 1990s, it was proposed that the main essence of the amyloid cascade hypothesis is increased production or decreased clearance of $A\beta$ peptide, the culprit behind AD [39, 40]. Accumulation of the hydrophobic $A\beta$ peptide ($A\beta_{40}$ and $A\beta_{42}$) results in its self-aggregation and formation of insoluble plaques, triggering a cascade of events resulting in death of the neuronal cells and thus causing AD (Figure 2).

A large portion of fAD cases is accounted for by mutations in the presenilin 1 (PS1) gene [41]. PS1 is one of the four main membrane proteins in presenilin complex which associates with three other membrane proteins to form the γ -secretase complex. Unlike in $A\beta$ PP, fAD mutations in PS1 are scattered throughout the length of molecules. Many of these mutations result in modified cleavage of $A\beta$ PP, causing enhanced production of the longer $A\beta_{42}$ peptide, which is more prone to self-aggregate as compared to the smaller $A\beta_{40}$ [42] and is shown to be more toxic *in vitro*. Since the level of $A\beta_{42}$ is found to be much higher in AD patients, it was postulated that the rise in levels of $A\beta_{42}$ triggered the cascade of the distorting events resulting in AD [43]. Although there is increase in $A\beta_{42}$ due to many fAD mutations, some mutations in PS1 do not elevate $A\beta_{42}$ levels but rather decrease $A\beta_{40}$ levels. This has led to yet another possibility that an enhancement in the $A\beta_{42/40}$ ratio, instead of the absolute levels of $A\beta_{42}$, is pathogenic and triggers the deleterious events leading to the disease. This view is supported by the observation that increased $A\beta_{42/40}$ ratio is generally inversely related to the age of onset of AD [44].

3. Amyloid- β Protein Precursor ($A\beta$ PP) and Its Function

The amyloid- β protein precursor ($A\beta$ PP) gene is located on chromosome 21 in humans. Alternative splicing of the $A\beta$ PP transcript generates 8 isoforms, of which 3 are most common: $A\beta$ PP695, $A\beta$ PP751, and $A\beta$ PP770. The 695 amino acid form predominantly expresses in the CNS, and 751 and 770 amino acid forms express ubiquitously [45]. $A\beta$ PP belongs to type I transmembrane proteins that include the amyloid precursor-like proteins (APLP1 and APLP2) in mammals and the amyloid precursor protein-like (APPL) in *Drosophila*.

The exact physiological function of $A\beta$ PP is not well known and remains an important issue in AD research. In many studies, overexpression of $A\beta$ PP shows a positive effect on cell health and growth. This effect is epitomized in transgenic mice that overexpress wild-type $A\beta$ PP and have enlarged neurons [46]. $A\beta$ PP knockout mice are viable and fertile, showing a comparatively subtle abnormal phenotype [47, 48]. APLP1 and APLP2 knockout mice also survive and are fertile. However, double null mice $A\beta$ PP/APLP2 and APLP1/APLP2 and triple null mice $A\beta$ PP/APLP1/APLP2 show early postnatal lethality [49, 50]. Interestingly, $A\beta$ PP/APLP1 mice are viable [50], suggesting that APLP2 is critical when either $A\beta$ PP or APLP1 is absent.

Further, the similarity in proteolytic processing and topology between Notch and $A\beta$ PP suggest that $A\beta$ PP may function as a membrane receptor like Notch. Indeed, different $A\beta$ PP ligands have been identified, such as $A\beta$ [51],

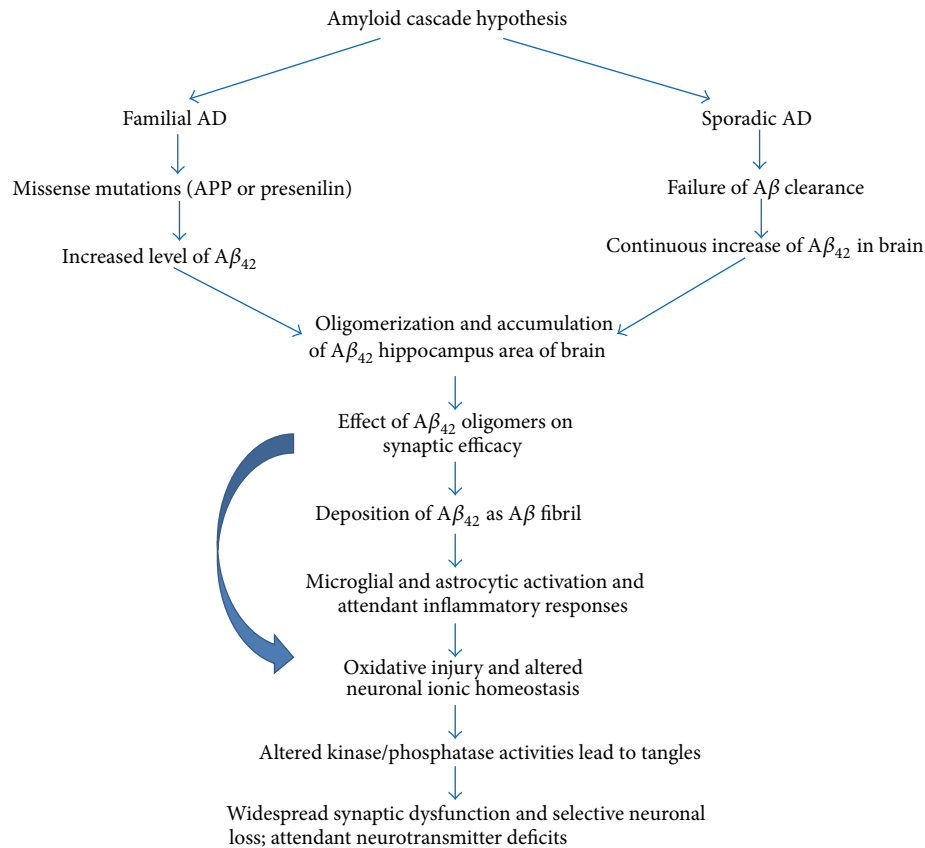


FIGURE 2: The amyloid cascade hypothesis of Alzheimer's disease. This hypothesis represents the classic theory of the origins of AD. Both familial forms of Alzheimer's (fAD) and later-onset forms with no known etiology (sporadic AD) lead to the production of excess Aβ₄₂. Once this toxic peptide begins to aggregate, a cascade of events is triggered that produces the biological and neurological symptoms of Alzheimer's disease.

netrin-1 [52], and F-spondin [53]. However, while binding of AβPP by these ligands can affect AβPP processing, the exact downstream signalling events triggered by such binding remain to be clarified and an authentic membrane receptor function for APP remains speculative.

Although AβPP has been the subject of much study since its identification, its physiological function remains unclear. AβPP has important roles in neurites' outgrowth and synaptogenesis, cell adhesion, calcium metabolism, neuronal protein trafficking along the axon, and transmembrane signal transduction, among others, all requiring additional *in vivo* experimental evidence [54]. AβPP generates various fragments during proteolytic processing and these AβPP metabolites serve various functions. Therefore, the net effect of full-length AβPP on biological activity may be a combination of its metabolites' functions, depending on the proportion of levels of each AβPP metabolite. In adult animals, intracerebral injections of the AβPP ectodomain can improve cognitive function and synaptic density [55, 56]. The sites most responsible for the bioactivity of the AβPP ectodomain appear to be its two heparin-binding domains [57]. Overall, studies from various research groups suggest that AβPP plays an important role in protein trafficking regulation.

4. Proteolytic Processing of AβPP and Generation of Aβ Peptide

As described above, AβPP is a type I transmembrane protein. It is synthesized in the endoplasmic reticulum (ER) and transported to the trans-Golgi-network (TGN) through the Golgi apparatus where the highest concentration of AβPP is found in neurons at steady state [58–60]. One of the most prominent areas of AD research is the study of the generation of Aβ after AβPP processing. Aβ generation takes place in ER and Golgi/TGN [60]. Further, AβPP can be transported from the TGN to TGN-derived secretory vesicles to the cell surface where it is either reinternalized via an endosomal/lysosomal degradation pathway [61, 62] or cleaved by α-secretase to produce a soluble molecule, AβPPs_α [63]. Some reports also suggest involvement of endosomal/lysosomal system in Aβ generation [64]. Unlike Aβ which is neurotoxic, studies suggest that AβPPs_α is neuroprotective, making the subcellular distribution of AβPP an important factor in neurodegeneration [65]. Therefore the characterization of the mechanisms involved in APP transport and trafficking are crucial to understanding the pathogenesis of AD.

Processing of AβPP takes place by two different pathways: amyloidogenic pathway, which results in the generation of

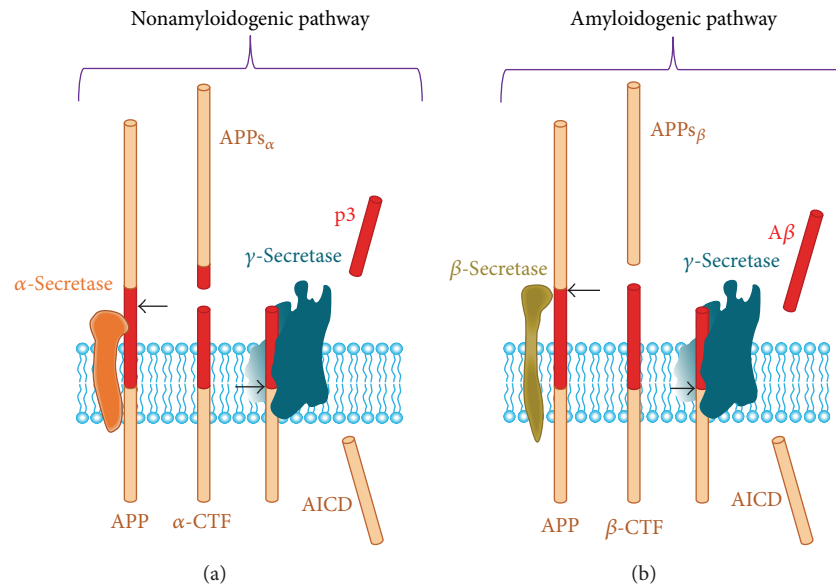


FIGURE 3: Processing of the amyloid- β protein precursor ($A\beta$ PP) occurs by two pathways. (a) Nonamyloidogenic processing of $A\beta$ PP involving action of α -secretase followed by γ -secretase as shown in the figure. (b) Amyloidogenic processing of $A\beta$ PP involving β -secretase followed by the action of γ -secretase. Both processes generate soluble ectodomains ($A\beta$ PPs _{α} and $A\beta$ PPs _{β}) and a similar intracellular C-terminal fragment (AICD). The $A\beta$ peptide starts within the ectodomain and continues into the transmembrane region (red). Adapted from Thinkaran and Koo [8].

toxic $A\beta$ fragment of 42 amino acid, and nonamyloidogenic pathway, which is required for normal functioning of neurons. There is involvement of three different types of serine proteases in the $A\beta$ PP processing which are α -, β -, and γ -secretases. The quantitatively and functionally most important proteolytic processing of $A\beta$ PP is mediated through nonamyloidogenic pathway, that is, cleavage of $A\beta$ PP by the action of α - and γ -secretases. Action of α -secretase releases the $A\beta$ PPs _{α} ectodomain and a carboxy terminal fragment ($A\beta$ PP-CTF _{α}). Action of γ -secretase later generates a small p3 and AICD fragment (Figure 3). $A\beta$ PPs _{α} has been suggested to show neuroprotective and synapse-promoting activities [66], but the mechanism behind this and identification of the receptor mediating these effects has not yet been identified.

Cleavage of $A\beta$ PP by the action of β -secretase and γ -secretase results in $A\beta_{42}$ generation, called amyloidogenic pathway (Figure 3). Action of β -secretase releases the ectodomain $A\beta$ PPs _{β} and rests $A\beta$ PP carboxy-terminal fragment ($A\beta$ PP-CTF _{β}) which is further cleaved by the γ -secretase producing the $A\beta$ peptide(s) and the $A\beta$ PP intracellular domain (AICD) fragment. The biological function of all the above fragments ($A\beta$ PPs _{β} , $A\beta$, and AICD) generated through amyloidogenic pathways is still to be explored, although $A\beta$ release is associated with synaptic activity and synaptic transmission onto neurons [67]. The AICD fragment is thought to be a nuclear signalling molecule [68], but this is also not fully explored [69].

5. Definition of Amyloid- β Peptide

Definition of amyloid- β follows the guidelines of nomenclature established in the November 2006 meeting of

The Nomenclature Committee of the International Society of Amyloidosis. Amyloid- β is defined as peptides of 36–43 amino acids that are primarily involved in AD as the main component of the amyloid plaques found in the brains of AD patients. Amyloid- β , also defined as protein deposits found *in vivo*, can be distinguished from nonamyloid protein deposits by observing under an electron microscope. Amyloid- β has a characteristic fibril appearance, a unique pattern of X-ray diffraction, and an affinity for the dye Congo red of histological samples, which results in an apple green birefringence under plane-polarized light [70]. The term amyloid- β was initially reported to restrict as extracellular deposits only. However, many types of amyloid- β have since been reported to begin intracellularly, resulting in the characteristic extracellular amyloid deposits found upon cell death. Therefore, amyloid is no longer restricted to extracellular inclusion but also includes those intracellular inclusions having typical amyloid appearance [70].

6. Physiological Function of $A\beta$ Peptide

Multiple lines of evidence reveal that overproduction of $A\beta$ through $A\beta$ PP processing results in a neurodegenerative cascade leading to self-aggregation, synaptic dysfunction, formation of intraneuronal fibrillary tangles, and gradual neuron loss in the hippocampus [71]. There are two main toxic species, $A\beta_{40}$ and $A\beta_{42}$, with $A\beta_{42}$ being more hydrophobic in nature and more prone to self-aggregation which results in $A\beta$ fibril formation [72]. Previous studies on familial form of AD (fAD) mutations consistently show rise in the ratio of $A\beta_{42/40}$ [73], thereby indicating that elevated levels of $A\beta_{42}$ relative to $A\beta_{40}$ are crucial for AD pathogenesis,

likely achieved by providing the core for $A\beta$ assembly into oligomers, fibrils, and amyloidogenic plaques [74]. Although the majority of $A\beta$ peptides are secreted from the cell, $A\beta$ can be generated in several subcellular compartments within the cell, such as the ER, Golgi/TGN, and endosome/lysosome. In addition, internalization of extracellular $A\beta$ can be done by a cell for its degradation. The presence of intracellular $A\beta$ implies that $A\beta$ may accumulate within the neuronal cell and contribute to AD pathogenesis. Confirming the presence of intracellular $A\beta$, intraneuronal $A\beta$ immunoreactivity has been found in the hippocampal and entorhinal cortical regions which are more prone to early AD pathology with mild cognitive impairment (MCI) in AD patients [75]. The accumulation of intracellular $A\beta$ paves the way for extracellular plaque formation in Down's syndrome (DS) patients [76] and the level of intraneuronal $A\beta$ reduced as the extracellular $A\beta$ plaques accumulate [77]. Studies of transgenic mouse models consistently confirm evidence for intracellular accumulation of $A\beta$ as an early event in the neuropathological phenotype, with decreasing intraneuronal levels of $A\beta$ as extracellular plaques build up [78–80]. Intraneuronal $A\beta$ can also disrupt amygdala-dependent emotional responses by modulating the ERK/MAPK signalling pathway [81]. Previous reports also suggest reduction in $A\beta$ neurotoxicity due to inhibition of dynamin-mediated but not clathrin-mediated $A\beta$ internalization [82]. One recent study by Friedrich et al. suggests that intracellular $A\beta$ can self-aggregate within the cell and disrupt the vesicular membrane, contributing to its pathological effect [83].

$A\beta$ was originally considered a neurotoxic species confined to the brain of aged or demented persons. Later findings suggest that the presence of soluble $A\beta$ species in the bodily fluids of many species [84] and in the conditional cell culture media [85] has disproved this concept and inferred a physiological function for $A\beta$. Low levels of $A\beta$ enhance hippocampal long-term potentiation and improve memory, indicating its novel positive, modulatory role in neurotransmission and memory [86], while excessive $A\beta$ causes neuronal loss as well as synaptic dysfunction. One study using a transgenic *Caenorhabditis elegans* model found that intracellular $A\beta$ aggregation in muscle cells may trap excess free copper to reduce copper-mediated cytotoxic effects [87]. However, whether $A\beta$ can form intracellular aggregates in human peripheral cells to exert a physiologically protective function remains to be determined.

7. Mechanism of Formation of $A\beta$ Fibril

The mechanism of formation of $A\beta$ oligomer *in vivo* remains unclear. In this context, Glabe suggests that the complexity of the oligomer formation can be assumed by the fact that multiple $A\beta$ oligomer conformations are produced via different pathways [88]. The mechanisms of fibril formation of extracellular and intracellular oligomers may also vary. The fibrillization of $A\beta$ into senile plaques is a complex process involving several steps [89–91]. After $A\beta$ is released from cells, it can bind to several proteins: for example, albumin, α 1-antichymotrypsin, apolipoprotein E, and complement proteins [92]. Presence of $A\beta$ as stable soluble

dimers is detected in both cell culture media and brain homogenates [93]. Total $A\beta$ concentration may be the critical determinant of fibril formation. In normal brain, breakdown of $A\beta$ takes place immediately after its production from the cells before fibrillization or deposition, while, in the aging brain, increased production of $A\beta$ and its reduced rate of clearance may lead to $A\beta$ fibrillization, further leading to disease condition and AD pathogenesis. Recent studies reveal three different types of $A\beta$ oligomers: (a) very short oligomers ranging from dimer to hexamer size [94, 95]; (b) small oligomers ranging from 17 to 42 kDa which are $A\beta$ -derived diffusible ligands (ADDLs) [96]; and (c) protofibrils that can be seen in electron microscopy as short fibril intermediates of less than 8 nm in diameter and less than 150 nm in length. Protofibrils are short-lived structures detected during *in vitro* formation of mature amyloid fibrils [97–99]. However, relationships between the aforementioned oligomers remain unclear. Moreover, all oligomeric forms of $A\beta$ derived intermediates, that is, oligomers, ADDLs, protofibrils, and mature $A\beta$ fibrils, are potentially neurotoxic and may be a key cause of neurotoxicity in AD. $A\beta$ exists mainly in two alloforms: $A\beta_{40}$ and $A\beta_{42}$, which follow distinct oligomerization pathways [94, 100]. Each peptide showed different behaviour at the earliest stage of assembly and monomer oligomerization. Kinetic studies of $A\beta$ fibril formation have shown that formation of $A\beta_{42}$ self-aggregates is faster than $A\beta_{40}$ and forms fibril [98, 101]. It is also well reported that the fibrillogenic and neurotoxic property of $A\beta_{42}$ is higher than that of $A\beta_{40}$. The initial phase of fibrillization of $A\beta_{42}$ monomers involves formation of pentamer/hexamer units called paranuclei (Figure 4). These paranuclei are initial structures that can further oligomerize to larger units and form large oligomers, protofibrils, and fibrils. Monomers, paranuclei, and large oligomers are predominantly unstructured with only short β -sheet/ β -turn and helical elements. During formation of protofibrils, essential conformational changes take place when the unstructured, α -helix, and β -strand elements convert into β -sheet/ β -turn structures. Paranuclei could not be detected for $A\beta_{40}$ at similar concentrations of the peptide. Aggregate-free $A\beta_{40}$, when carefully prepared, existed as monomers, dimers, trimers, and tetramers, in rapid equilibrium [94]. The important residue promoting the pentamer/hexamer formation is Ile41. Addition of later residue to $A\beta_{40}$ is sufficient to induce paranuclei formation [94]. A natural propensity to form paranuclei is the only feature of $A\beta_{42}$. This important finding may explain the predominantly strong association of $A\beta_{42}$ with AD. Paranuclei formation in $A\beta_{42}$ is blocked by oxidation of Met35 and produces oligomers indistinguishable in morphology and size and from those produced by $A\beta_{40}$ [95]. Preventing the fibrillization of toxic $A\beta_{42}$ paranuclei through selective Met35 oxidation thus represents a potential therapeutic target for AD treatment. The most important feature of controlling early oligomerization of $A\beta$ is the length of the C-terminal as compared to 34 physiological relevant alloforms of $A\beta$ [95]. The primary amino acid residue in $A\beta_{42}$ is a side chain of residue 41 which is crucial for effective paranuclei formation and self-aggregation into oligomer formation. $A\beta_{40}$ self-aggregation is particularly

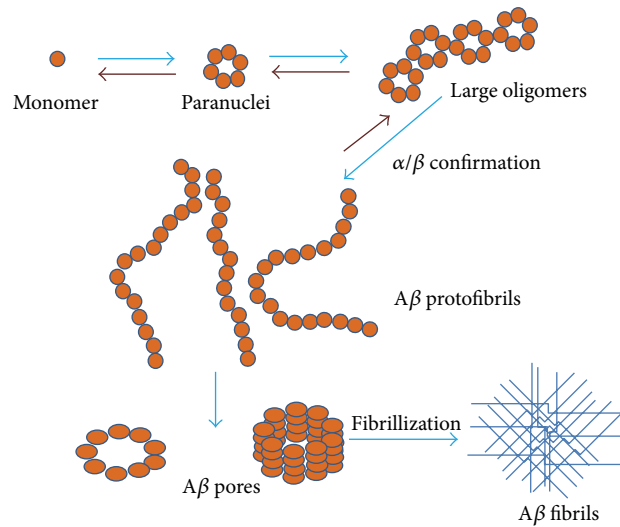


FIGURE 4: A model showing $A\beta_{42}$ oligomerization and fibrillization: the equilibrium between monomer to paranuclei and from paranuclei to large oligomers is rapid and reversible. The conversion of oligomers to protofibrils is slower but also reversible. Conversion of protofibrils into fibrils is an irreversible step. Basically, the monomers, paranuclei, and large oligomers do not have any definite structure instead of some β -turn/ β -sheet and helical (α) elements. Essential conformational changes occur during protofibril formation where the unstructured, α -helix, and β -strand elements transform into β -sheet/ β -turn structures.

critical to substitutions of Glu22 or Asp23 and to truncation of the N terminus [94]. Whereas $A\beta_{42}$ oligomerization is largely unaffected by substitutions at positions 22 or 23 or by N-terminal truncations, it is significantly affected by Phe19 or Ala21 substitutions. The above statement reveals that $A\beta$ oligomerization differs between $A\beta_{40}$ and $A\beta_{42}$ which are controlled by specific regions and residues.

8. $A\beta$ Toxicity

Alzheimer's disease is considered by some researchers to be a disease of the synapses and has been termed "synaptic failure" [102]. While $A\beta$ can destroy neurons, synaptotoxicity may be more appropriate for earlier stages of AD that are best categorized by synaptic loss rather than neuronal death. Loss of dendritic spines or synaptic terminals may cause the associated deterioration in cognitive functions that characterizes AD. However, it is still unclear whether the synaptotoxic and neurotoxic actions of $A\beta$ are a separate mechanistic process or if the actions follow a common mechanism [103]. As discussed above, the pentameric and hexameric oligomers may be the building blocks of the more toxic decameric and dodecameric complexes. Both cross-linked oligomeric forms of $A\beta$ and $A\beta$ fibril were significantly more toxic than disaggregated $A\beta$ (dimers threefold and tetramers 13-fold more toxic than monomers). One of the main results is the fact that monomers have very low toxicity, while toxicity rises substantially only when $A\beta$ self-associates, although it is still challenging to establish a degree of increasing toxicity with the number of monomers in the given oligomer because of the decrease in occurrence frequency of higher order oligomers. Monomeric $A\beta$ has the propensity to adopt different conformations in water solutions, including momentarily extended

β -sheet conformations in the central and C-terminal regions, connected by turn between them, or β -hairpin [104].

Several other lines of study support a role for oligomeric form of $A\beta$ as the toxic entity in AD patients. Human brain shows soluble oligomers with similar structural properties as observed *in vitro* by antioligomeric antibodies staining; the same oligomers were also observed *in vivo* [105]. $A\beta$ oligomer toxicity *in vitro* has been attributed to several distinct mechanisms, including but not limited to membrane disruption and direct formation of ion channels. There have been numerous reports of increased membrane conductance or leakage in the presence of $A\beta$ oligomers ranging from small globulomers to large prefibrillar assemblies [106, 107], with some evidence presented to support formation of discrete ion channels of pores [108, 109].

Fibrillar $A\beta$, on the other hand, has been shown to bind to a wide array of cell surface proteins, including the receptor for advanced glycation end products (RAGE) complex and $A\beta$ PP [110], leading in some cases to increased free radical formation and oxidative stress. Similarly, binding to the α -7 nicotinic receptor can mediate N-methyl-D-aspartate (NMDA) receptor activity with broad effects on cellular metabolism [111]. Any or all of these effects may play a role in loss of synaptic function, leading to symptomatic AD. Other proposed interactions, such as dysregulation of calcium channels, may be confounded by membrane disruption effects, making them harder to confirm.

It is important to note that since $A\beta$ exists *in vitro* and *in vivo* as a continuum of different oligomeric states, none of which are particularly stable, it is difficult to distinguish biological effects induced by one specific type of nonfibrillar oligomer. Therefore, it is entirely feasible that $A\beta$ has significantly different physiological effects when in different oligomeric forms. Thus, it is difficult to exclude any of

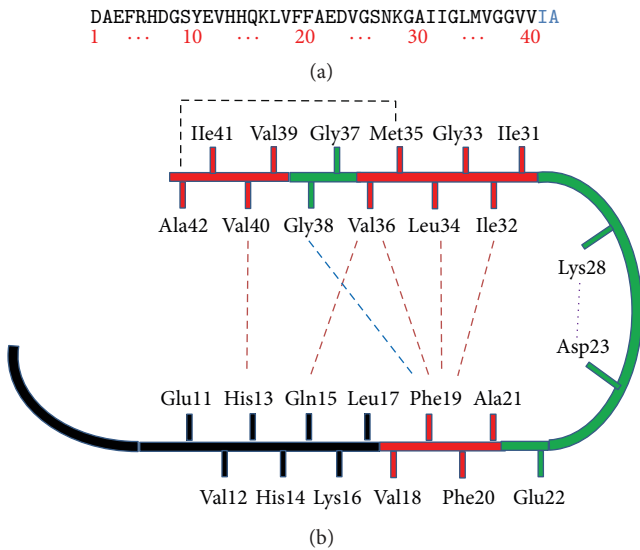


FIGURE 5: (a) Sequence of $A\beta_{42}$ that is derived from human $A\beta$ PP. (b) Structural constraints in $A\beta_{40}$ and $A\beta_{42}$ fibrils. NMR measurements of $A\beta_{40}$ fibrils have shown that residues 1–10 are unstructured and residues 11–40 adopt a β -turn- β fold. Side chain packing is observed between Phe19 and Ile32, Leu34 and Val36, Gln15 and Val36, and His13 and Val40 (orange dashed line). In $A\beta_{42}$ fibrils, residues 1–17 may be unstructured (in black), with residues 18–42 forming a β -turn- β fold. Molecular contacts have been reported within the monomer unit of $A\beta_{42}$ fibrils between Phe19 and Gly38 (blue dashed line) and between Met35 and Ala42 (black dashed line). In both $A\beta_{40}$ and $A\beta_{42}$, the turn conformation is stabilized by hydrophobic interactions (red residues) and by a salt bridge between Asp23 and Lys28 (purple dashed line).

the putative mechanisms for involvement of $A\beta$ oligomers in progression of AD without further study.

$A\beta_{40}$ is the common, more soluble form of $A\beta$. $A\beta_{42}$ has two extra amino acids on the end of the peptide (Figure 5(a)). One of these, 42nd amino acid, is an alanine, which can loop back to form a salt bridge with 35th amino acid, methionine (Met35) (Figure 5(b)). This extra hairpin turn of $A\beta_{42}$ makes it less soluble and more toxic. The toxicity of $A\beta_{42}$ is much greater than $A\beta_{40}$.

9. Role of $A\beta$ in AD Pathogenesis

As discussed so far, it is clear that $A\beta$ is one of the hallmarks for Alzheimer's disease. $A\beta$ is generated from $A\beta$ PP processing by β - and γ -secretases through the amyloid cascade pathway. $A\beta$ is one of the main toxic peptides which has a critical role in AD pathogenesis. $A\beta$ normally has a propensity for self-aggregation, resulting in $A\beta$ fibril formation which ultimately form senile plaques extracellularly, causing neuronal damage and synaptic dysfunction. Although $A\beta$ aggregates are mainly found in the hippocampus area of postmortem brain of AD patients, they are also distributed to some extent in the cortex area of brain. As we know, the hippocampus is the prime memory storage part of the brain, so these aggregates affect the surrounding neurons

in hippocampus area and are responsible for AD pathology. The temporal profile of pathological features, together with genetic risk factors for AD, has led to the hypothesis that accumulation of $A\beta$ oligomers during early, preclinical stages of the disease initiates a cascade of events resulting in synaptic dysfunction, neuronal loss, and atrophy within the temporoparietal and hippocampal regions. This neurodegeneration, in turn, causes neuronal dysfunction, cognitive decline, and ultimately complete loss of memory [112].

10. Therapeutic Approaches for AD

There is no cure for AD; however, drug treatments are available to help relieve symptoms in several aspects of the disease, and researchers around the world are focusing on finding better treatments, preventive strategies, and ultimately a cure. A variety of cellular mechanisms can lead to the generation of Alzheimer's disease. Along with $A\beta$, microtubule-associated protein tau is another hallmark of AD [113]. In the case of AD, tau becomes hyperphosphorylated, aggregated, and finally accumulated as NFT [114]. Tau plays an important role, not only as axonal protein but also as regulator of dendritic function, particularly mediating early $A\beta$ toxicity during AD [115]. Therefore, $A\beta$ and tau became targets in drug development for AD. Many clinical trials targeting these two proteins have been implemented; several lines of research are still under investigations. There are several therapeutic approaches being investigated for the treatment of AD (Figure 6). Therapeutic strategies are basically categorized in the following three ways: (i) treatments that prevent the onset of the disease by sequestering the primary progenitors; (ii) disease-modifying therapies termination or the reversal of disease progression; and (iii) symptomatic treatments that treat the cognitive symptoms of the disease and protect from further cognitive decline. Among the therapeutic strategies mentioned in Figure 6, amyloid-based therapies using synthetic- as well as herbal-based anti-amyloid approach are highlighted here.

10.1. Amyloid-Focused Therapies. Along with tau-focused therapeutic approaches, amyloid-focused treatment strategies are also in development in order to prevent the aggregation and accumulation of insoluble $A\beta$ and/or clear $A\beta$ plaques postformation. Still, studies have reported that soluble $A\beta$ peptides may similarly be protective *in vivo* as an ameliorative response to free radical toxicity [116, 117].

10.1.1. Inhibition of $A\beta$ Aggregation. Since $A\beta$ aggregation is hypothesized to be the most crucial step of the pathogenic process of AD, the strategy to inhibit $A\beta$ aggregation has emerged as a promising approach to treat AD. Numerous synthetic as well as herbal compounds have been identified as inhibitors of $A\beta$ aggregation; however, the mechanistic interaction between $A\beta$ and these compounds is still not clear [118]. To gain insight into the mechanism of inhibition, it is necessary to understand the structure of $A\beta$. While the structure of $A\beta$ has not been resolved by crystallography,

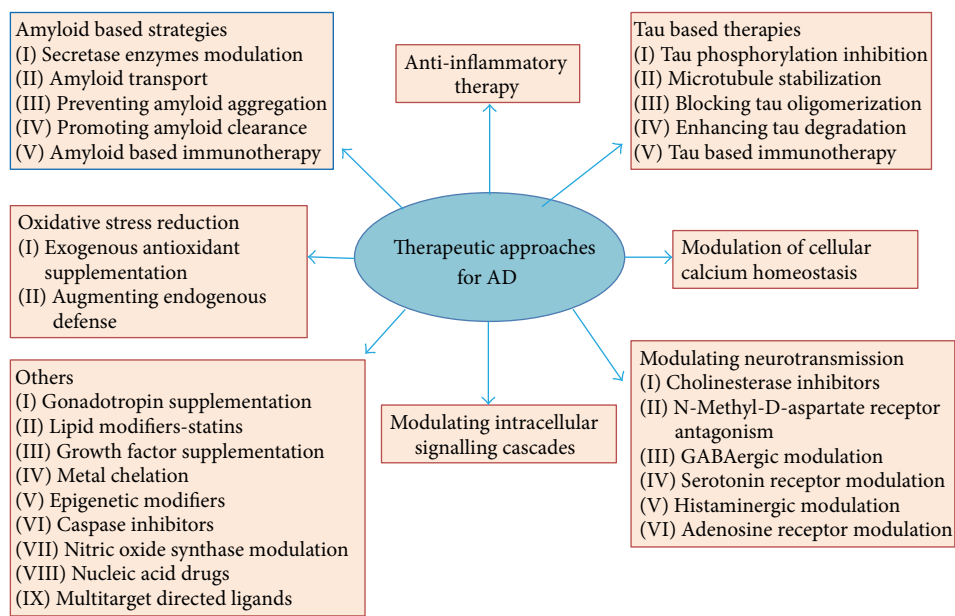


FIGURE 6: Different therapeutic approaches for the treatment of AD.

several structures have been predicted by different techniques, including nuclear magnetic resonance (NMR) spectroscopy and computer simulation [119–121]. Petkova et al. proposed a broadly used structural model for $A\beta$ fibrils using solid state NMR (SS-NMR), as shown in Figure 5(b) [122]. In this model, residues 1–10 are structurally disordered, while residues 12–24 and 30–40 adopt β -strand conformations and form parallel β -sheets by internal hydrogen bonding. Figure 5(b) shows the secondary structure for a single $A\beta_{42}$ monomer within the fibril. Residues 25–29 contain a 180° bend of the protein backbone that brings the two β -sheets in contact through side chain-side chain interactions. A single cross- β unit is a double-layered β -sheet structure with a hydrophobic core and a hydrophobic face. The only charged residues in the core are Asp23 and Lys28, which form a salt bridge to stabilize the β -sheet structure.

This model contains information relevant to the design of inhibitors for $A\beta$ aggregation. For example, compounds that have interaction propensity with the hydrophobic core of $A\beta$ peptide would disrupt monomer-monomer interaction, thereby destabilizing the formation of small oligomeric aggregates or nuclei. Compounds that recognize residues 12–24, which are included in the interaction between fibril units, would interfere with lateral association, while compounds interacting through hydrogen bonds formation with amino or carboxyl groups of residues 12–24 or 30–40 are expected to inhibit soluble aggregate elongation.

Most compounds showing inhibitory capability toward $A\beta$ aggregation are aromatic in nature, such as resveratrol, coumarin, and nicotine [123–125]. It is hypothesized that the aromaticity plays an important role by breaking the hydrophobic interaction between $A\beta$ monomers. Aromatic compounds can interact with residues Phe19 and Phe20 of $A\beta$ peptide via π - π stacking interactions.

Another finding that the derivatives of penta peptide, KLVFF (residues 16–20 of $A\beta$), can inhibit $A\beta$ aggregation supports this speculation [126, 127]. Therefore, to achieve better inhibitory capability toward $A\beta$ aggregation, aromatic compounds have been modified and functionalized on their aromatic centre.

11. Different Model Organisms to Study Alzheimer's Disease

As the rate of occurrences of AD is growing continuously, pressure is mounting on the research community to develop a suitable and effective drug treatment. While other age-related diseases like heart disease and cancer can now be successfully studied, treated, and to a certain extent cured, AD, and other age-related human neurodegenerative diseases such as Parkinson's disease, is still not curable. This is not only due to a poorer understanding of AD, the complexity of the brain, and its relative inaccessibility, but it is also due to a lack of "natural" disease models. For example, the dog naturally mimics some AD features including $A\beta$ cortical pathology, loss of neuronal cells, and learning and memory deficits, but it does not develop neuritic plaques and NFTs [128]. Primates do develop forms of both but are not well studied. Also, though rodents will readily develop cancer, the senile plaques and NFTs formation have never been reported [129].

Thus, transgenic AD mouse and rat models have enabled the scientific community to overcome the lack of a suitable natural model for the study of AD. The major limitation is that rodents do not naturally have anything close to AD. The AD in these models is imposed. Since the study of AD in humans is methodologically and ethically complex and critical, AD transgenic models provide an approach to understanding

AD pathogenesis, so as to recognize new biomarkers and to design new therapeutics, although they do not utilize normal physiology. In addition, transgenic AD models allow investigation of the early stages of the disease, something that is problematic with human postmortem tissue [130]. In contrast numerous drug targets have “cured” these mouse models while having no benefit in clinical trials of AD patients [131]. The problem is that AD pathology is a response whose removal has limited, if any, benefit for humans but is an imposed abnormality for the transgenic rodents.

11.1. Transgenic Mice Model of AD. The mouse model of AD was established in the mid-1990s with the development of the PDAPP model [132], followed by the Tg2576 in subsequent years [133] and APP23 [134] models, being currently the most extensively used amyloidosis models in AD research. The PDAPP model expresses human A β PP carrying the Indiana familial AD mutation (V717F) driven by the platelet-derived growth factor- β promoter, whereas both Tg2576 and A β PP23 models express human A β PP with the Swedish mutation (K670N/M671L) driven by the hamster prion protein and murine Thy-1 promoters, respectively. All the above-mentioned models support the amyloid cascade hypothesis; they show progressive A β deposition in both diffuse and neuritic plaques, cerebral amyloid angiopathy, microgliosis, (limited) hippocampal atrophy, astrogliosis, synaptic and neurotransmitter alterations, and cognitive and behavioural deficits, relevant to human AD neuropathological profile [135–138]. A β PP-based models confirm the principal role of A β PP and A β in the Alzheimer disease process and allow target identification and subsequent preclinical evaluation of various symptomatic and disease-modifying drugs, primarily targeting the amyloid cascade. The major drawback of these models, however, is the lack of NFT formation, although hyperphosphorylated tau may be present.

The discovery of early-onset mutations in the PSEN genes aids in development of PSEN1 and PSEN2 transgenic mouse models. Even though an increased A β ₄₂/A β ₄₀ ratio in some of these models has been observed, they are void of plaque pathology. Further, few behavioural and cognitive discrepancies are present in these models; they lack NFT development like A β PP-based models as well. They are mainly useful for the development of double transgenic A β PP/PSEN mice, which display an elevated A β ₄₂/A β ₄₀ ratio and accelerated A β pathology compared to the single A β PP model they are based on, thereby supporting the modifying role of PSEN. In addition, these A β PP/PSEN mice display amyloid-associated inflammation, neuronal loss, cognitive decline, and BPSD-like behavioural alterations [139, 140]. The major loophole of all the above-mentioned models—lack of NFT formation—was moderately overcome by the development of transgenic mice having human tau insertion and the subsequent crossing of tau and A β PP models, the latter including enhanced amyloid deposition accompanied by tau hyperphosphorylation, NFT-like formation, and obvious death of neurons, thereby supporting the amyloid cascade hypothesis affirming that A β pathology mediates tau pathology. However, there is no colocalization of plaques and NFT in AD brain of A β PP/tau mice. This limitation was

compensated with the development of the triple transgenic (3xTg) mouse [78]. Instead of crossing independent mutant mouse lines, two transgenic constructs (mutant A β PP and tau) were microinjected into single-cell embryos of homozygous mutant PSEN1 mice, thereby preventing segregation of A β PP and tau genes in succeeding generations. In accordance with the amyloid cascade theory, these 3xTg mice develop A β plaques prior to NFT pathology with a temporal and spatial profile equivalent to AD, in addition to inflammation, synaptic dysfunction, and cognitive decline [141].

The generation of transgenic rodent research models that develop some of the pathological hallmarks of AD has given a substantial boost to drug discovery efforts and has also raised many intriguing questions about the underlying disease process. However, one should never neglect the potential danger of uncritical extrapolating from mouse/rat to humans. The fact that at the moment no animal model recapitulates all aspects of human AD reflects the limitations of using a rodent system to model a human condition that takes decades to develop and primarily involves higher cognitive functions.

11.2. *Caenorhabditis elegans* Transgenic AD Model. *Caenorhabditis elegans*, a free-living small nematode of approximately 1.2 mm in length, has several characteristics that make it useful as a model organism. The nematodes are transparent, which allows the study of embryonic development and gene expression in living animals under the microscope. It was first used to study molecular and developmental biology by Sydney Brenner in the 1970s [142]. This invertebrate was the first animal for which an entire genome was sequenced and has become one of the most popular model organisms to study neurodegenerative disease, as demonstrated by the development of numerous transgenic disease models, including for Alzheimer's disease (discussed below).

Several AD-related genes and pathways found in humans have orthologues in *C. elegans*. The nematode genome encodes three orthologues for PSEN1: (i) sel-12, (ii) hop-1, and (iii) spe-4. The first has been found in a screen for suppressors of the egg-laying defective phenotype in lin-12 gain-of-function worms [143]; it facilitates Notch/lin-12 signalling, functional mostly during embryonic development. The second, that is hop-1, homolog of PSEN1 [144], in fact shares more homology to human PSEN2; and the third, spe-4, has no clear human counterpart [144]. Three genes, aph-1, pen-2, and aph-2, combine together to form a functional complex of γ -secretase. In addition, an orthologue of A β (apl-1) has been described in *C. elegans* [145]. Similar to *Drosophila*, the APL-1 protein does not contain the A β sequence; neither does *C. elegans* display BACE1-like activity.

There are basically three A β -expressing nematode models which have been developed. When expressed in muscle cells, A β _{1–42} induced the formation of amyloid-immunoreactive inclusions. A subset of these deposits also binds the A β -specific dye thioflavin S, showing that amyloid fibrils are formed similar to human AD. In addition, paralysis of the nematodes occurred, thereby indicating a muscle cell specific toxicity of A β [146]. Nematodes expressing A β _{1–42} in neuronal cells also develop A β deposits but display only a very

subtle phenotype [147]. Interestingly, oligomeric $A\beta$ were detected in these strains that might be similar to the neurotoxic $A\beta$ -derived diffusible ligands [148]. These transgenic models provide important insight into the toxicity of specific $A\beta$ species but do not allow screening of chemical or genetic modifiers of $A\beta$ PP processing.

To create tauopathy models of nematode, both wild-type and mutated human tau proteins were expressed in neurons of *C. elegans*, inducing a progressive phenotype of defective motility (uncoordinated phenotype), which was more deceptive in the mutants. Interestingly, these transgenic lines also display hyperphosphorylation of tau [149], which is linked to GSK-3 β activation. Future genome-wide screens will display which modifier genes are linked to the complex disease process and characterize diagnostic or therapeutic drug targets.

11.3. *Drosophila melanogaster* as a Transgenic Model of AD. *Drosophila melanogaster* are the most commonly used species of *Drosophila* in the laboratory worldwide for research purposes. Their use in the modelling of human neurodegenerative disease is based on the inherent assumption that the fundamental aspects of cell biology are conserved throughout evolution in higher organisms. This is supported by the fact that ~75% of human disease-related genes have homologs in *Drosophila*, suggesting that molecular mechanisms of any disease in humans may be conserved in the fly.

There are many compelling reasons to study AD in the *Drosophila* model. The *Drosophila* brain has approximate 300,000 neurons and is organized into the area with separate, specialized functions such as memory, learning, olfaction, and vision, similar to human. *Drosophila*, on account of its very short generation time (10–12 days) and easy maintenance, is a popular model in genetic research. Although one could argue that the *Drosophila*'s maximum lifespan of 55–80 days is significantly greater than that of the worm (~18 days), it is still much shorter than that of the mouse (2–3 years), making it ideal for studying a progressive age-related disease such as AD.

In addition, the *Drosophila* has an unrivalled battery of genetic tools, including a fully sequenced genome; an extensive library of mutant stocks including RNA interference (RNAi) and knockout (KO) lines; sophisticated transposon-based methods for gene manipulation; systems for spatial and temporal specific ectopic gene expression; and balancer chromosomes. Balancer chromosomes are unique: composed of multiple inversions that prevent recombination, together with dominant, lethal, and visible markers. They allow the maintenance in long-term culture of lethal or deleterious mutations in heterozygotes, without the necessity to set up specific crosses.

The combination of such extensive genetic tools and practicality makes the *Drosophila* ideal for genetic screening. A variety of screening methods are available in the *Drosophila*, involving chemical mutagenesis (EMS), genetic deletion kits, or mobile genetic elements (P, EP, and GS elements). Genetic screens are powerful experiments providing an unbiased forward genetic approach, which allows the discovery of

genes or metabolic pathways not immediately apparent in the pathogenesis of AD.

11.4. Drug Screens Using *Drosophila* AD Models. Another potential use for a characterized *Drosophila* disease model is its use for novel drug screening. The secreted $A\beta$ peptide fly model was verified as a platform for drug discovery by testing the efficacy of a drug used to treat human AD patients and was shown to slow progression of AD [150]. The drug memantine, a noncompetitive glutamate antagonist, is effective in slowing progression of human AD [151]. In addition, the life span of flies expressing two copies of $A\beta_{42}$ or one copy of $A\beta_{42}$ -arctic was increased when flies were treated with MK-801, an inhibitor of the excitatory action of glutamate on the NMDA receptor [150]. A therapeutic intervention that is effective in human AD patients is, therefore, also effective in the fly; thus the fly AD model is useful for testing novel human drugs. Congo red, which binds to $A\beta$ and has been shown to reduce neurodegeneration in a fly model of polyQ disease [152] and a mouse model of Huntington's disease [153], has also been shown to reverse the reduced life span of flies expressing two copies of $A\beta_{42}$ or one copy of $A\beta_{42}$ -arctic [150].

The $A\beta$ PP processing model in *Drosophila* has also been used in drug validation studies. Ubiquitous expression of $A\beta$ PP, BACE, and DPSn resulted in reduced longevity and a visible wing phenotype, which were used for screening β - and γ -secretase inhibitors [154]. Feeding flies with either β - or γ -secretase inhibitors resulted in an increased survival of $A\beta$ PP/BACE/DPSn expressing transgenic flies, making this fly model useful for investigating drugs that modulate $A\beta$ PP processing and have the potential to decrease $A\beta$ -induced cellular degeneration [154]. Singh in his doctoral thesis showed neuroprotective effect of some herbal compounds targeting anti- $A\beta$ therapeutic approach using *Drosophila* model of AD [155].

12. $A\beta$ and Metallosis

12.1. Metal Ions and $A\beta$ Toxicity. In case of AD, elevated metal ions concentrations have been demonstrated in several studies [156–159], in particular copper and zinc which are associated with both the aggregation and the neurotoxicity of $A\beta$ peptides, and proposed as an important factor in neuropathology of AD [156, 159–162].

An extensive number of reports have provided empirical data showing metal mediated toxicity of amyloid- β , but a detailed NMR or X-ray diffraction atomic structure is yet to be described [163, 164]. However, Azimi and Rauk [165] were able to use MD simulations to demonstrate that $A\beta$ -copper coordinated structures can form both parallel and antiparallel conformations. Zinc ions have been shown to form intermolecular complexes while copper ions tend to form intramolecular complexes cross-linking multiple peptides [166–168]. The schematic in Figure 7(a) shows the details of metal ion mediated $A\beta$ toxicity which results in fibril formation and leads to AD. Figure 7(b) depicts $A\beta$ -copper interactions [162]. Interaction of $A\beta$ amino acid

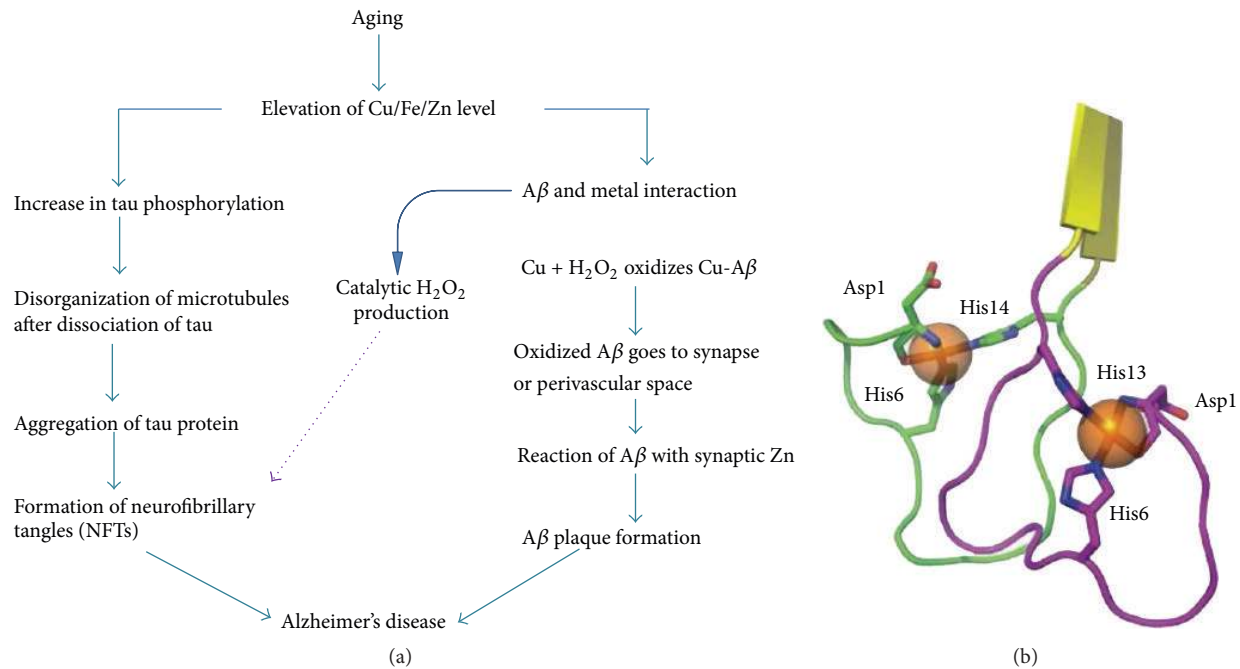


FIGURE 7: (a) Proposed model for AD pathology based on abnormal metal interaction. Cu and Fe levels increase during aging in the CNS and result in increase in metal and A β interaction. Cu binding to A β results in ROS production and autooxidation of A β peptide. Oxidized A β contributes to synaptic pathology and plaque formation. Metals may also promote phosphorylation of tau and hence enhance formation of NFT which further contribute to AD pathology. (b) Model showing the amyloid-copper interaction. Notice the coordination sites at His 6 from one peptide together with His13 and His14 from the second peptide.

residues with copper is shown in figure which leads to Cu mediated A β toxicity. Copper has been shown to interact with amyloid- β at the His13 and His14 residues on one peptide with the His6 residue on the other peptide.

Histidine is well known as Zn²⁺ and Cu²⁺ ligand for many other proteins and peptides [169, 170]. Coordination of Zn²⁺ to His13 and His14 but not His6 has been found to be critical to induce A β ₁₋₄₀ aggregation [171, 172]. Cu²⁺ is able to compete with Zn²⁺ for binding to histidine residues of A β and at low concentration it inhibits the ability of Zn²⁺ to induce aggregation, but at higher Cu²⁺ concentrations aggregation does occur [173].

It has been reported that the levels of copper (0.4 ± 0.1 mg/g of wet weight of plaque) and zinc (1.2 ± 0.2 mg/g) are found to be high in the senile plaques found within AD brains [157]. Use of a Cu-Zn chelator, such as clioquinol, inhibits A β accumulation in AD transgenic mice [174, 175], which highlights the importance of studying copper and zinc binding to A β .

12.2. ROS Generation by Metal Mediated A β Toxicity. The coordination of metal ions such as copper, iron, and zinc to A β also results in the chemical reduction of these metals and the subsequent generation of hydrogen peroxide from molecular oxygen together with other available biological reducing agents such as cholesterol, in a catalytic manner [176–178]. In the case of A β ₄₂, the reduction of copper is independent of the aggregation state of the peptide, as both soluble and fibrillar forms show copper-reducing ability.

The generation of hydrogen peroxide in the presence of reduced metals, and in the absence of sufficient detoxifying enzymes such as catalase and glutathione peroxidase, gives rise to the toxic hydroxyl radical via Fenton chemistry [176]. The generation of hydrogen peroxide contributes to A β toxicity. In support of this, cellular toxicity can be rescued by the addition of catalase [178, 179]. Further, resistance to A β toxicity is associated with an enhanced ability to degrade hydrogen peroxide [180], and catalase inhibitors can enhance A β toxicity. The potentiation of A β toxicity by copper is the greatest for A β ₄₂ > A β ₄₀ > rodent A β ₄₀, which corresponds to the peptide relative activities in reducing copper(II) to copper(I) [176]. These data support a role for A β in the generation of hydrogen peroxide via metal ion reduction and for oxidative processes in the augmentation of A β to potentiate the AD cascade. A summary of the proposed role of metal ions in AD has already been shown in Figure 7(a).

12.3. Neuroprotective Role of Metal Chelator against A β Induced Toxicity. To prevent metal mediated neurotoxicity of A β , researchers are focusing on chelation therapy. Chelation therapy is the use of metal specific chelators which are able to chelate extra metal ions present in brain; hence reduced possibility of interaction of these metals with A β , and ultimately a slowdown of metal, mediated A β -toxicity.

Traditional metal chelators have been used to sequester or redistribute metal ions from metal-bound A β species in order to suppress metal mediated A β neurotoxicity *in vitro* and *in vivo* [181–184]. Cherny et al. [174] initially reported that

Cu/Zn chelators solubilize A β from tissues of postmortem AD brain. They choose the clioquinol (CQ, 5-chloro-7-iodo-8-hydroxyquinoline) based on its ability to cross the blood-brain barrier as tested in AD transgenic mice (Tg2576) and found significantly reduced levels (~65%) of A β -aggregates as well as ROS generation in CQ treated mice as compared to control.

Ongoing research in this area focuses on the prevention of metal mediated A β neurotoxicity and ROS production by metal chelating therapy, which is an emerging trend in current research. There is immense need to develop such a suitable metal chelator that can prevent A β aggregation by effectively sequestering extra metal ions. Several groups focused on developing such type of new molecules [182, 185, 186]. More particularly, in a pioneering work, Lakatos et al. developed two carbohydrate-containing compounds, N,N'-bis[(5- β -D-glucopyranosyloxy-2-hydroxy)benzyl]-N,N'-dimethyl-ethane-1,2-diamine (H2GL1) and N,N'-bis[(5- β -D-glucopyranosyloxy-3-tert-butyl-2-hydroxy)benzyl]-N,N'-dimethyl-ethane-1,2-diamine (H2GL2), that are shown to be promising therapeutic tools against AD, based on *in vitro* studies [185]. In this context, we designed and synthesized novel compound L, 2,6-pyridinedicarboxylic acid, 2,6-bis[2-[(4-carboxyphenyl)methylene]hydrazide], to test the *in vivo* neuroprotective efficacy in a well-established *Drosophila* transgenic model system. Recently Singh et al. reported the neuroprotective role of a novel copper chelator against copper mediated A β toxicity [162].

13. Neuroprotective Role of Flavonoid against AD

Naturally occurring as well as synthetically synthesized dietary flavonoids have been extensively used as alternative candidates for Alzheimer's treatment, taking into account their antioxidative, anti-amyloidogenic, and anti-inflammatory properties. Experimental evidence from different studies supports the hypothesis that certain flavonoids may protect against AD, in part by interfering with the generation and assembly of amyloid- β peptides into neurotoxic oligomeric aggregates and also by reducing tau aggregation. Dietary supplementation studies using flavonoid-rich plant or food extracts have shown their ability to influence cognition and learning in humans and also in animal models of diseases [187–192]. Presently, there is no direct association between flavonoid consumption and improvement in neurological health. Nevertheless, the potential beneficial effect of flavonoids in the brain seems to be related to their ability to interact with intracellular neuronal and glial signalling pathways, thus influencing the peripheral and cerebral vascular system, protecting vulnerable neurons, enhancing existing neuronal function, or stimulating neuronal regeneration.

Flavonoids are naturally occurring polyphenolic compounds widely spread in plants. They are present in foods and beverages of plant origin such as a variety of fruits, vegetables, cocoa, cereals, tea, and wine [193]. The six main subclasses of flavonoids include (1) flavanols (e.g., kaempferol, quercetin), present in onions, leeks, and broccoli; (2) isoflavones (e.g., daidzein, genistein), found mainly in soy and soy products;

(3) flavones (e.g., apigenin, luteolin), present in parsley and celery; (4) flavanols (e.g., catechin, epicatechin, epigallocatechin, and epigallocatechin gallate (EGCG)), abundant in green tea, red wine, and chocolate; (5) flavanones (e.g., hesperetin, naringenin), primarily found in citrus fruit and tomatoes; and finally (6) anthocyanidins (e.g., pelargonidin, cyanidin, and malvidin), sources of which include berry fruits and red wine.

It was thought that the ability of flavonoids to promote memory, learning, and cognitive function was mediated by their antioxidant capacity [194]. Nevertheless, due to their limited absorption and their low bioavailability in the brain, increasing evidence demonstrates that they are able to interact with the cellular and molecular components of the brain responsible for memory, having the potential to protect vulnerable neurons, enhance existing neuronal function, stimulate neuronal regeneration, and induce neurogenesis [194, 195]. Recent study from our lab showed the neuroprotective property of a novel synthetic flavonoid derivative against A β -induce neurotoxicity in *Drosophila* model of AD [196].

14. Neuroprotective Role of Natural Polyphenols in AD

Nature has gifted mankind with a plethora of vegetables, flora-bearing fruits, and nuts. Natural polyphenols are the most commonly found chemical compounds in consumable herbal beverages and food worldwide [197, 198]. They constitute a large group of phytochemicals with more than 8000 identified compounds. The variety of bioactive nutrients present in these natural products play a central role in prevention and cure of various human neurodegenerative diseases, such as Alzheimer's disease (AD), Parkinson's disease, and other kinds of neuronal damage. Plants have a long history as a rich source of new bioactive compounds for drug discovery and may have advantages in relation to efficacy. Several reports documented the effectiveness of herbal extracts over isolated material, in protection against lipid peroxidation [199], and anticancer effects [200]. For example, a mixture of carotenoids has been found to be more effective than any one single carotenoid in protecting liposomes against lipid peroxidation [199].

Polyphenolic compounds from medicinal plants are key sources of neuroprotective agents against AD. Using the structure of these bioactive ingredients as templates for synthetic drugs offers a wide range of potential neuroprotective compounds [201]. In the past few decades, several studies attempted to measure the effect of total plant extract on AD and to isolate the active component responsible for the neuroprotective effects [202, 203].

Natural polyphenols reveal their antioxidant effect by reducing free radical species and/or encouraging endogenous antioxidant capacity. Thus, the antioxidant properties positively contribute to their neuroprotective effects. Furthermore, some of them influence synthesis of endogenous antioxidant molecules in cells via activating Nrf/ARE pathway [204]. Apart from antioxidant property, most of them appear to have a number of different molecular targets, affecting several signalling pathways and showing pleiotropic

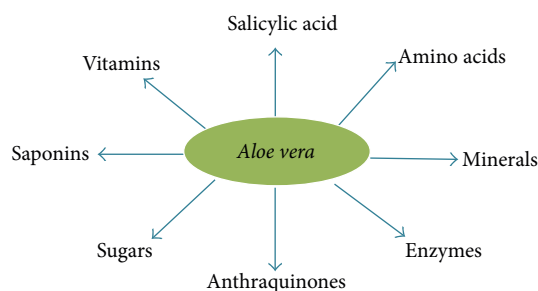


FIGURE 8: Illustration of the different components present in *Aloe vera*.

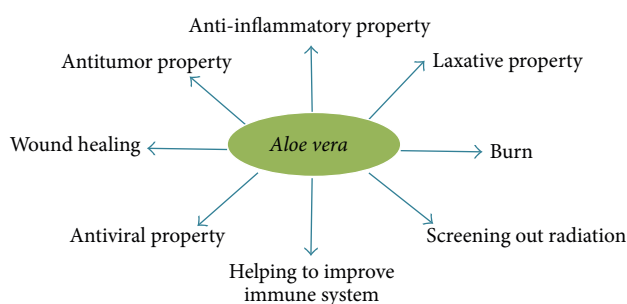


FIGURE 9: Figure showing the medicinal properties of *Aloe vera*.

activity on cells [205]. For instance, polyphenolic compounds can modulate activity of NF- κ B or SIRT1 exerting neuroprotective effects. Recent studies have shown anti-A β activity of compounds from natural sources *in vitro* and *in vivo* [206–208]. Still, evidence for the capability of common edible elements to inhibit A β oligomerization *in vivo* remains a challenge.

Aloe vera has been used as medicinal agent since Roman times [209]. *A. vera* contains different bioactive components (Figure 8) harbouring over 75 biologically active compounds [210] known to have a wide range of pharmacological activities (Figure 9), including anti-inflammatory, wound healing, antioxidative, antiarthritic, antidiabetic, and antitumorigenic effects [211].

Aloe vera has always been preferred as a herbal remedy and is one of the most popular herbal plants. Major value added products from *Aloe* are gel and juice.

Recently, it has been reported that *Aloe vera*, supplemented orally to mice, is effective on wound healing. *Aloe vera* acts as a free radical scavenger and has other antioxidant properties on diabetic patients by controlling elevated anions in an alloxan- or STZ-induced diabetic animal models [212, 213].

15. Conclusion

Currently the accumulated experimental evidence leans toward strongly supporting the toxic role of A β within the pathophysiology of AD. However, the existence of some data regarding the role of A β in the normal physiology of the brain suggests that this peptide may act in different modes

at different times, according to diverse conditions. So far, it appears that at the initial stages of development and in the young brain, when in physiological doses (i.e., picomolar to nanomolar range) and in soluble, oligomeric forms, A β can show neuroprotective, antioxidant, and trophic properties, even facilitating synaptic plasticity. On the contrary, in many potentially adverse conditions, A β may deploy multiple toxic effects, contributing significantly to neuronal damage, as seen in AD. Some of these conditions appear to be associated with A β itself, such as high concentrations and fibrillar or aggregated states, presence of free metals, brain tissue previously injured or aged, and decreased antioxidative mechanisms. Moreover, it is necessary to remark that both trophic and toxic effects may not be mutually exclusive. In other words, they might coexist and cross-modulate each other, even throughout advanced stages of AD, complicating an approach based upon anti-amyloidogenic therapy, at least theoretically. This functional duality may also underlie the modest success and the high rate of collateral consequences of such therapies. In summary, blockade, inhibition, or modulation of those sites, effects, and negative processes in which A β is involved, but simultaneously respecting those sites and physiologic processes in which A β is also taking part, remain a major challenge for therapeutic research in the future.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

Support was provided by the Indian Council of Medical Research and Banaras Hindu University as UGC-RET and SRF Fellowship, respectively, to Sandeep Kumar Singh. Also support was provided by the Semmes Foundation and by a grant from the National Institute on Minority Health and Health Disparities (G12MD007591) from the National Institutes of Health.

References

- [1] D. J. Selkoe, "Alzheimer's disease: genes, proteins, and therapy," *Physiological Reviews*, vol. 81, no. 2, pp. 741–766, 2001.
- [2] L. E. Hebert, P. A. Scherr, J. L. Bienias, D. A. Bennett, and D. A. Evans, "Alzheimer disease in the US population: prevalence estimates using the 2000 census," *Archives of Neurology*, vol. 60, no. 8, pp. 1119–1122, 2003.
- [3] P. S. Mathuranath, P. J. Cherian, R. Mathew et al., "Dementia in Kerala, South India: prevalence and influence of age, education and gender," *International Journal of Geriatric Psychiatry*, vol. 25, no. 3, pp. 290–297, 2010.
- [4] V. Chandra, R. Pandav, H. H. Dodge et al., "Incidence of Alzheimer's disease in a rural community in India. The Indo-US study," *Neurology*, vol. 57, no. 6, pp. 985–989, 2001.
- [5] T. Yoshitake, Y. Kiyohara, I. Kato et al., "Incidence and risk factors of vascular dementia and Alzheimer's disease in a defined elderly Japanese population: the Hisayama study," *Neurology*, vol. 45, no. 6, pp. 1161–1168, 1995.

- [6] R. L. Ernst and J. W. Hay, "Economic research on Alzheimer disease: a review of the literature," *Alzheimer Disease and Associated Disorders*, vol. 11, supplement 6, pp. 135–145, 1997.
- [7] C. Haass and D. J. Selkoe, "Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid β -peptide," *Nature Reviews Molecular Cell Biology*, vol. 8, no. 2, pp. 101–112, 2007.
- [8] G. Thinakaran and E. H. Koo, "Amyloid precursor protein trafficking, processing, and function," *The Journal of Biological Chemistry*, vol. 283, no. 44, pp. 29615–29619, 2008.
- [9] Alzheimer's Association 2010, Web, 2010, <http://alz.org>.
- [10] Z. S. Khachaturian and T. S. Radebaugh, *Alzheimer's Disease: Cause(s), Diagnosis, Treatment, and Care*, CRC, Boca Raton, Fla, USA, 1996.
- [11] M. Goedert and M. G. Spillantini, "A century of Alzheimer's disease," *Science*, vol. 314, no. 5800, pp. 777–781, 2006.
- [12] T. A. Bayer, O. Wirths, K. Majtényi et al., "Key factors in Alzheimer's disease: beta-amyloid precursor protein processing, metabolism and intraneuronal transport," *Brain Pathology*, vol. 11, no. 1, pp. 1–11, 2001.
- [13] J. Hardy and D. J. Selkoe, "The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics," *Science*, vol. 297, no. 5580, pp. 353–356, 2002.
- [14] J. Carter and C. F. Lippa, " β -amyloid, neuronal death and Alzheimer's disease," *Current Molecular Medicine*, vol. 1, no. 6, pp. 733–737, 2001.
- [15] J. L. Cummings, H. V. Vinters, G. M. Cole, and Z. S. Khachaturian, "Alzheimer's disease: etiologies, pathophysiology, cognitive reserve, and treatment opportunities," *Neurology*, vol. 51, supplement 1, pp. S2–S17, 1998.
- [16] Alzheimer Europe, March 2004, <http://www.alzheimer-europe.org/?lm1=D8105B21BD2C>.
- [17] K. Blennow, M. J. de Leon, and H. Zetterberg, "Alzheimer's disease," *The Lancet*, vol. 368, no. 9533, pp. 387–403, 2006.
- [18] E. G. Gray, M. Paula-Barbosa, and A. Roher, "Alzheimer's disease: paired helical filaments and cytomembranes," *Neuropathology and Applied Neurobiology*, vol. 13, no. 2, pp. 91–110, 1987.
- [19] A. Mudher and S. Lovestone, "Alzheimer's disease—do taoists and baptists finally shake hands?" *Trends in Neurosciences*, vol. 25, no. 1, pp. 22–26, 2002.
- [20] D. R. Williams, "Tauopathies: classification and clinical update on neurodegenerative diseases associated with microtubule-associated protein tau," *Internal Medicine Journal*, vol. 36, no. 10, pp. 652–660, 2006.
- [21] P. T. Francis, A. M. Palmer, M. Snape, and G. K. Wilcock, "The cholinergic hypothesis of Alzheimer's disease: a review of progress," *Journal of Neurology Neurosurgery and Psychiatry*, vol. 66, no. 2, pp. 137–147, 1999.
- [22] A. V. Terry Jr. and J. J. Buccafusco, "The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development," *Journal of Pharmacology and Experimental Therapeutics*, vol. 306, no. 3, pp. 821–827, 2003.
- [23] S. Auld Daniel, "Amyloid β peptides as direct cholinergic neuromodulators: a missing link?" *Trends in Neurosciences*, vol. 21, pp. 43–49, 1998.
- [24] R. T. Bartus, R. L. Dean III, B. Beer, and A. S. Lippa, "The cholinergic hypothesis of geriatric memory dysfunction," *Science*, vol. 217, no. 4558, pp. 408–414, 1982.
- [25] P. Davies and A. J. F. Maloney, "Selective loss of central cholinergic neurons in Alzheimer's disease," *The Lancet*, vol. 308, no. 8000, p. 1403, 1976.
- [26] Z. X. Shen, "Brain cholinesterases: II, The molecular and cellular basis of Alzheimer's disease," *Medical Hypotheses*, vol. 63, no. 2, pp. 308–321, 2004.
- [27] R. H. Swerdlow and S. M. Khan, "A 'mitochondrial cascade hypothesis' for sporadic Alzheimer's disease," *Medical Hypotheses*, vol. 63, no. 1, pp. 8–20, 2004.
- [28] R. H. Swerdlow and S. J. Kish, "Mitochondria in Alzheimer's disease," *International Review of Neurobiology*, vol. 53, pp. 341–385, 2002.
- [29] G. E. Gibson, K.-F. R. Sheu, and J. P. Blass, "Abnormalities of mitochondrial enzymes in Alzheimer disease," *Journal of Neural Transmission*, vol. 105, no. 8–9, pp. 855–870, 1998.
- [30] W. D. Parker Jr. and J. K. Parks, "Cytochrome c oxidase in Alzheimer's disease brain: purification and characterization," *Neurology*, vol. 45, no. 31, pp. 482–486, 1995.
- [31] Q. Ding, S. Martin, E. Dimayuga, A. J. Bruce-Keller, and J. N. Keller, "LMP2 knock-out mice have reduced proteasome activities and increased levels of oxidatively damaged proteins," *Antioxidants & Redox Signaling*, vol. 8, no. 1–2, pp. 130–135, 2006.
- [32] R. H. Swerdlow, "Mitochondria in cybrids containing mtDNA from persons with mitochondriopathies," *Journal of Neuroscience Research*, vol. 85, no. 15, pp. 3416–3428, 2007.
- [33] G. G. Glenner and C. W. Wong, "Alzheimer's disease and Down's syndrome: sharing of a unique cerebrovascular amyloid fibril protein," *Biochemical and Biophysical Research Communications*, vol. 122, no. 3, pp. 1131–1135, 1984.
- [34] G. G. Glenner and C. W. Wong, "Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein," *Biochemical and Biophysical Research Communications*, vol. 120, no. 3, pp. 885–890, 1984.
- [35] J. Kang, H.-G. Lemaire, A. Unterbeck et al., "The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor," *Nature*, vol. 325, no. 6106, pp. 733–736, 1987.
- [36] M. J. Owen, L. A. James, J. A. Hardy, R. Williamson, and A. M. Goate, "Physical mapping around the Alzheimer disease locus on the proximal long arm of chromosome 21," *The American Journal of Human Genetics*, vol. 46, no. 2, pp. 316–322, 1990.
- [37] A. Rovelet-Lecrux, D. Hannequin, G. Raux et al., "APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy," *Nature Genetics*, vol. 38, no. 1, pp. 24–26, 2006.
- [38] K. Sleegers, N. Brouwers, I. Gijssels et al., "APP duplication is sufficient to cause early onset Alzheimer's dementia with cerebral amyloid angiopathy," *Brain*, vol. 129, no. 11, pp. 2977–2983, 2006.
- [39] J. A. Hardy and G. A. Higgins, "Alzheimer's disease: the amyloid cascade hypothesis," *Science*, vol. 256, no. 5054, pp. 184–185, 1992.
- [40] D. J. Selkoe, "The molecular pathology of Alzheimer's disease," *Neuron*, vol. 6, no. 4, pp. 487–498, 1991.
- [41] M. Cruts and C. Van Broeckhoven, "Presenilin mutations in Alzheimer's disease," *Human Mutation*, vol. 11, no. 3, pp. 183–190, 1998.
- [42] N. Suzuki, T. T. Cheung, X.-D. Cai et al., "An increased percentage of long amyloid beta protein secreted by familial amyloid beta protein precursor (beta APP717) mutants," *Science*, vol. 264, no. 5163, pp. 1336–1340, 1994.
- [43] S. G. Younkin, "Evidence that A β 42 is the real culprit in Alzheimer's disease," *Annals of Neurology*, vol. 37, no. 3, pp. 287–288, 1995.

- [44] M. Bentahir, O. Nyabi, J. Verhamme et al., "Presenilin clinical mutations can affect gamma-secretase activity by different mechanisms," *Journal of Neurochemistry*, vol. 96, no. 3, pp. 732–742, 2006.
- [45] T. A. Bayer, R. Cappai, C. L. Masters, K. Beyreuther, and G. Multhaup, "It all sticks together—the APP-related family of proteins and Alzheimer's disease," *Molecular Psychiatry*, vol. 4, no. 6, pp. 524–528, 1999.
- [46] E. S. Oh, A. V. Savonenko, J. F. King et al., "Amyloid precursor protein increases cortical neuron size in transgenic mice," *Neurobiology of Aging*, vol. 30, no. 8, pp. 1238–1244, 2009.
- [47] H. Zheng, M. Jiang, M. E. Trumbauer et al., " β -amyloid precursor protein-deficient mice show reactive gliosis and decreased locomotor activity," *Cell*, vol. 81, no. 4, pp. 525–531, 1995.
- [48] G. R. Dawson, G. R. Seabrook, H. Zheng et al., "Age-related cognitive deficits, impaired long-term potentiation and reduction in synaptic marker density in mice lacking the β -amyloid precursor protein," *Neuroscience*, vol. 90, no. 1, pp. 1–13, 1999.
- [49] C. S. Von Koch, H. Zheng, H. Chen et al., "Generation of APLP2 KO mice and early postnatal lethality in APLP2/APP double KO mice," *Neurobiology of Aging*, vol. 18, no. 6, pp. 661–669, 1997.
- [50] S. Heber, J. Herms, V. Gajic et al., "Mice with combined gene knock-outs reveal essential and partially redundant functions of amyloid precursor protein family members," *Journal of Neuroscience*, vol. 20, no. 21, pp. 7951–7963, 2000.
- [51] A. Lorenzo, M. Yuan, Z. Zhang et al., "Amyloid β interacts with the amyloid precursor protein: a potential toxic mechanism in Alzheimer's disease," *Nature Neuroscience*, vol. 3, no. 5, pp. 460–464, 2000.
- [52] F. C. Lourenço, V. Galvan, J. Fombonne et al., "Netrin-1 interacts with amyloid precursor protein and regulates amyloid- β production," *Cell Death & Differentiation*, vol. 16, no. 5, pp. 655–663, 2009.
- [53] A. Ho and T. C. Südhof, "Binding of F-spondin to amyloid- β precursor protein: A candidate amyloid- β precursor protein ligand that modulates amyloid- β precursor protein cleavage," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 8, pp. 2548–2553, 2004.
- [54] H. Zheng and E. H. Koo, "The amyloid precursor protein: beyond amyloid," *Molecular Neurodegeneration*, vol. 1, article 5, 2006.
- [55] H. Meziane, J.-C. Dodart, C. Mathis et al., "Memory-enhancing effects of secreted forms of the β -amyloid precursor protein in normal and amnesic mice," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 21, pp. 12683–12688, 1998.
- [56] J.-M. Roch, E. Masliah, A.-C. Roch-Levecq et al., "Increase of synaptic density and memory retention by a peptide representing the trophic domain of the amyloid β /A4 protein precursor," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 91, no. 16, pp. 7450–7454, 1994.
- [57] S. S. Mok, G. Sberna, D. Heffernan et al., "Expression and analysis of heparin-binding regions of the amyloid precursor protein of Alzheimer's disease," *FEBS Letters*, vol. 415, no. 3, pp. 303–307, 1997.
- [58] H. Xu, D. Sweeney, R. Wang et al., "Generation of Alzheimer β -amyloid protein in the trans-Golgi network in the apparent absence of vesicle formation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 8, pp. 3748–3752, 1997.
- [59] T. Hartmann, S. C. Bieger, B. Brühl et al., "Distinct sites of intracellular production for Alzheimer's disease A β 40/42 amyloid peptides," *Nature Medicine*, vol. 3, no. 9, pp. 1016–1020, 1997.
- [60] J. P. Greenfield, J. Tsai, G. K. Gouras et al., "Endoplasmic reticulum and trans-Golgi network generate distinct populations of Alzheimer β -amyloid peptides," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 2, pp. 742–747, 1999.
- [61] S. S. Sisodia, "Beta-amyloid precursor protein cleavage by a membrane-bound protease," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 89, no. 13, pp. 6075–6079, 1992.
- [62] C. Nordstedt, G. L. Caporaso, J. Thyberg, S. E. Gandy, and P. Greengard, "Identification of the Alzheimer β /A4 amyloid precursor protein in clathrin-coated vesicles purified from PC12 cells," *Journal of Biological Chemistry*, vol. 268, no. 1, pp. 608–612, 1993.
- [63] G. L. Caporaso, K. Takei, S. E. Gandy et al., "Morphologic and biochemical analysis of the intracellular trafficking of the Alzheimer β /A4 amyloid precursor protein," *The Journal of Neuroscience*, vol. 14, no. 5, pp. 3122–3138, 1994.
- [64] C. Haass, A. Y. Hung, M. G. Schlossmacher, D. B. Teplow, and D. J. Selkoe, " β -amyloid peptide and a 3-kDa fragment are derived by distinct cellular mechanisms," *Journal of Biological Chemistry*, vol. 268, no. 5, pp. 3021–3024, 1993.
- [65] K. Furukawa, B. L. Sopher, R. E. Rydel et al., "Increased activity-regulating and neuroprotective efficacy of α -secretase-derived secreted amyloid precursor protein conferred by a C-terminal heparin-binding domain," *Journal of Neurochemistry*, vol. 67, no. 5, pp. 1882–1896, 1996.
- [66] S. Bandyopadhyay, L. E. Goldstein, D. K. Lahiri, and J. T. Rogers, "Role of the APP non-amyloidogenic signaling pathway and targeting α -secretase as an alternative drug target for treatment of Alzheimer's diseases," *Current Medicinal Chemistry*, vol. 14, no. 27, pp. 2848–2864, 2007.
- [67] F. Kamenetz, T. Tomita, H. Hsieh et al., "APP processing and synaptic function," *Neuron*, vol. 37, no. 6, pp. 925–937, 2003.
- [68] P. Cupers, I. Orlans, K. Craessaerts, W. Annaert, and B. De Strooper, "The amyloid precursor protein (APP)-cytoplasmic fragment generated by γ -secretase is rapidly degraded but distributes partially in a nuclear fraction of neurones in culture," *Journal of Neurochemistry*, vol. 78, no. 5, pp. 1168–1178, 2001.
- [69] S. S. Hébert, L. Serneels, A. Tolia et al., "Regulated intramembrane proteolysis of amyloid precursor protein and regulation of expression of putative target genes," *EMBO Reports*, vol. 7, no. 7, pp. 739–745, 2006.
- [70] P. Westermark, M. D. Benson, J. N. Buxbaum et al., "A primer of amyloid nomenclature," *Amyloid*, vol. 14, no. 3, pp. 179–183, 2007.
- [71] G. M. Shankar, S. Li, T. H. Mehta et al., "Amyloid- β protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory," *Nature Medicine*, vol. 14, no. 8, pp. 837–842, 2008.
- [72] D. Burdick, B. Soreghan, M. Kwon et al., "Assembly and aggregation properties of synthetic Alzheimer's A4/ β amyloid peptide analogs," *Journal of Biological Chemistry*, vol. 267, no. 1, pp. 546–554, 1992.
- [73] D. R. Borchelt, G. Thinakaran, C. B. Eckman et al., "Familial Alzheimer's disease linked presenilin 1 variants elevate Abeta1-42/1-40 ratio in vitro and in-vivo," *Neuron*, vol. 17, pp. 1005–1013, 1996.

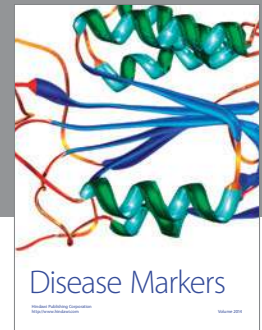
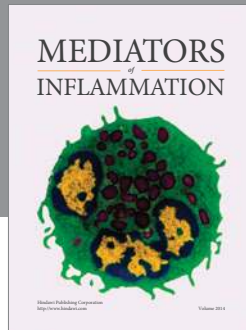
- [74] J. T. Jarrett, E. P. Berger, and P. T. Lansbury Jr., "The carboxy terminus of the beta amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease," *Biochemistry*, vol. 32, no. 18, pp. 4693–4697, 1993.
- [75] G. K. Gouras, J. Tsai, J. Naslund et al., "Intraneuronal A β 42 accumulation in human brain," *American Journal of Pathology*, vol. 156, no. 1, pp. 15–20, 2000.
- [76] K. A. Gyure, R. Durham, W. F. Stewart, J. E. Smialek, and J. C. Troncoso, "Intraneuronal A β -amyloid precedes development of amyloid plaques in Down syndrome," *Archives of Pathology and Laboratory Medicine*, vol. 125, no. 4, pp. 489–492, 2001.
- [77] C. Mori, E. T. Spooner, K. E. Wisniewski et al., "Intraneuronal A β 42 accumulation in Down syndrome brain," *Amyloid*, vol. 9, no. 2, pp. 88–102, 2002.
- [78] S. Oddo, A. Caccamo, J. D. Shepherd et al., "Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular A β and synaptic dysfunction," *Neuron*, vol. 39, no. 3, pp. 409–421, 2003.
- [79] S. Oddo, A. Caccamo, I. F. Smith, K. N. Green, and F. M. LaFerla, "A dynamic relationship between intracellular and extracellular pools of A β ," *American Journal of Pathology*, vol. 168, no. 1, pp. 184–194, 2006.
- [80] H. Oakley, S. L. Cole, S. Logan et al., "Intraneuronal β -amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation," *Journal of Neuroscience*, vol. 26, no. 40, pp. 10129–10140, 2006.
- [81] J. España, L. Giménez-Llort, J. Valero et al., "Intraneuronal β -amyloid accumulation in the amygdala enhances fear and anxiety in Alzheimer's disease transgenic mice," *Biological Psychiatry*, vol. 67, no. 6, pp. 513–521, 2010.
- [82] C. Yu, E. Nwabuisi-Heath, K. Laxton, and M. J. Ladu, "Endocytic pathways mediating oligomeric A β 42 neurotoxicity," *Molecular Neurodegeneration*, vol. 5, article 19, 2010.
- [83] R. P. Friedrich, K. Tepper, R. Röncke et al., "Mechanism of amyloid plaque formation suggests an intracellular basis of A β pathogenicity," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 5, pp. 1942–1947, 2010.
- [84] P. Seubert, C. Vigo-Pelfrey, F. Esch et al., "Isolation and quantification of soluble Alzheimer's beta-peptide from biological fluids," *Nature*, vol. 359, no. 6393, pp. 325–327, 1992.
- [85] C. Haass, M. G. Schlossmacher, A. Y. Hung et al., "Amyloid β -peptide is produced by cultured cells during normal metabolism," *Nature*, vol. 359, no. 6393, pp. 322–325, 1992.
- [86] D. Puzzo, L. Privitera, E. Leznik et al., "Picomolar amyloid- β positively modulates synaptic plasticity and memory in hippocampus," *The Journal of Neuroscience*, vol. 28, no. 53, pp. 14537–14545, 2008.
- [87] A. N. Minniti, D. L. Rebolledo, P. M. Grez et al., "Intracellular amyloid formation in muscle cells of A β -transgenic *Caenorhabditis elegans*: determinants and physiological role in copper detoxification," *Molecular Neurodegeneration*, vol. 4, article 2, 2009.
- [88] C. G. Glabe, "Structural classification of toxic amyloid oligomers," *Journal of Biological Chemistry*, vol. 283, no. 44, pp. 29639–29643, 2008.
- [89] D. B. Teplow, "Structural and kinetic features of amyloid β -protein fibrillogenesis," *Amyloid*, vol. 5, no. 2, pp. 121–142, 1998.
- [90] B. P. Tseng, W. P. Esler, C. B. Clish et al., "Deposition of monomeric, not oligomeric, A β mediates growth of Alzheimer's disease amyloid plaques in human brain preparations," *Biochemistry*, vol. 38, no. 32, pp. 10424–10431, 1999.
- [91] D. M. Walsh, B. P. Tseng, R. E. Rydel, M. B. Podlisny, and D. J. Selkoe, "The oligomerization of amyloid β -protein begins intracellularly in cells derived from human brain," *Biochemistry*, vol. 39, no. 35, pp. 10831–10839, 2000.
- [92] J. Ma, A. Yee, H. B. Brewer Jr., S. Das, and H. Potter, "Amyloid-associated proteins α 1-antichymotrypsin and apolipoprotein E promote assembly of Alzheimer β -protein into filaments," *Nature*, vol. 372, no. 6501, pp. 92–94, 1994.
- [93] M. Enya, M. Morishima-Kawashima, M. Yoshimura et al., "Appearance of sodium dodecyl sulfate-stable amyloid β -protein (A β) dimer in the cortex during aging," *American Journal of Pathology*, vol. 154, no. 1, pp. 271–279, 1999.
- [94] G. Bitan, B. Tarus, S. S. Vollers et al., "A molecular switch in amyloid assembly: Met35 and amyloid β -protein oligomerization," *Journal of the American Chemical Society*, vol. 125, no. 50, pp. 15359–15365, 2003.
- [95] G. Bitan, S. S. Vollers, and D. B. Teplow, "Elucidation of primary structure elements controlling early amyloid β -protein oligomerization," *Journal of Biological Chemistry*, vol. 278, no. 37, pp. 34882–34889, 2003.
- [96] M. P. Lambert, A. K. Barlow, B. A. Chromy et al., "Diffusible, nonfibrillar ligands derived from A β 1-42 are potent central nervous system neurotoxins," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 11, pp. 6448–6453, 1998.
- [97] J. D. Harper, S. S. Wong, C. M. Lieber, and P. T. Lansbury Jr., "Observation of metastable A β amyloid protofibrils by atomic force microscopy," *Chemistry and Biology*, vol. 4, no. 2, pp. 119–125, 1997.
- [98] D. M. Walsh, A. Lomakin, G. B. Benedek, M. M. Condron, and D. B. Teplow, "Amyloid β -protein fibrillogenesis. Detection of a protofibrillar intermediate," *The Journal of Biological Chemistry*, vol. 272, no. 35, pp. 22364–22372, 1997.
- [99] W. Yong, A. Lomakin, M. D. Kirkitadze et al., "Structure determination of micelle-like intermediates in amyloid β -protein fibril assembly by using small angle neutron scattering," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 1, pp. 150–154, 2002.
- [100] M. D. Kirkitadze, G. Bitan, and D. B. Teplow, "Paradigm shifts in Alzheimer's disease and other neurodegenerative disorders: the emerging role of oligomeric assemblies," *Journal of Neuroscience Research*, vol. 69, no. 5, pp. 567–577, 2002.
- [101] M. D. Kirkitadze, M. M. Condron, and D. B. Teplow, "Identification and characterization of key kinetic intermediates in amyloid β -protein fibrillogenesis," *Journal of Molecular Biology*, vol. 312, no. 5, pp. 1103–1119, 2001.
- [102] D. J. Selkoe, "Alzheimer's disease is a synaptic failure," *Science*, vol. 298, no. 5594, pp. 789–791, 2002.
- [103] R. Cappai and K. J. Barnham, "Delineating the mechanism of Alzheimer's disease A β peptide neurotoxicity," *Neurochemical Research*, vol. 33, no. 3, pp. 526–532, 2008.
- [104] A. Sandberg, L. M. Luheshi, S. Söllvander et al., "Stabilization of neurotoxic Alzheimer amyloid-beta oligomers by protein engineering," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 35, pp. 15595–15600, 2010.
- [105] R. Kaye, E. Head, F. Sarsoza et al., "Fibril specific, conformation dependent antibodies recognize a generic epitope common

- to amyloid fibrils and fibrillar oligomers that is absent in prefibrillar oligomers,” *Molecular Neurodegeneration*, vol. 2, no. 2, article 18, 2007.
- [106] S. Chimon and Y. Ishii, “Capturing intermediate structures of Alzheimer’s β -amyloid, $A\beta(1-40)$, by solid-state NMR spectroscopy,” *Journal of the American Chemical Society*, vol. 127, no. 39, pp. 13472–13473, 2005.
- [107] L. Yu, R. Edalji, J. E. Harlan et al., “Structural characterization of a soluble amyloid β -peptide oligomer,” *Biochemistry*, vol. 48, no. 9, pp. 1870–1877, 2009.
- [108] N. Arispe, “Architecture of the Alzheimer’s $A\beta$ P ion channel pore,” *Journal of Membrane Biology*, vol. 197, no. 1, pp. 33–48, 2004.
- [109] A. Quist, I. Doudevski, H. Lin et al., “Amyloid ion channels: a common structural link for protein-misfolding disease,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 30, pp. 10427–10432, 2005.
- [110] Y. Verdier and B. Penke, “Binding sites of amyloid beta-peptide in cell plasma membrane and implications for Alzheimer’s disease,” *Current Protein & Peptide Science*, vol. 5, no. 1, pp. 19–31, 2004.
- [111] E. M. Snyder, Y. Nong, C. G. Almeida et al., “Regulation of NMDA receptor trafficking by amyloid- β ,” *Nature Neuroscience*, vol. 8, no. 8, pp. 1051–1058, 2005.
- [112] C. R. Jack Jr., D. S. Knopman, W. J. Jagust et al., “Hypothetical model of dynamic biomarkers of the Alzheimer’s pathological cascade,” *The Lancet Neurology*, vol. 9, no. 1, pp. 119–128, 2010.
- [113] E.-M. Mandelkow and E. Mandelkow, “Tau in Alzheimer’s disease,” *Trends in Cell Biology*, vol. 8, no. 11, pp. 425–427, 1998.
- [114] D. H. Geschwind, “Tau phosphorylation, tangles, and neurodegeneration: the chicken or the egg?” *Neuron*, vol. 40, no. 3, pp. 457–460, 2003.
- [115] L. M. Ittner, Y. D. Ke, F. Delerue et al., “Dendritic function of tau mediates amyloid- β toxicity in Alzheimer’s disease mouse models,” *Cell*, vol. 142, no. 3, pp. 387–397, 2010.
- [116] A. Nunomura, G. Perry, G. Aliev et al., “Oxidative damage is the earliest event in Alzheimer disease,” *Journal of Neuropathology and Experimental Neurology*, vol. 60, no. 8, pp. 759–767, 2001.
- [117] C. L. Masters, G. Simms, and N. A. Weinman, “Amyloid plaque core protein in Alzheimer disease and Down syndrome,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 82, no. 12, pp. 4245–4249, 1985.
- [118] B. Cheng, H. Gong, H. Xiao, R. B. Petersen, L. Zheng, and K. Huang, “Inhibiting toxic aggregation of amyloidogenic proteins: a therapeutic strategy for protein misfolding diseases,” *Biochimica et Biophysica Acta—General Subjects*, vol. 1830, no. 10, pp. 4860–4871, 2013.
- [119] O. O. Olubiyi and B. Strodel, “Structures of the amyloid β -peptides $A\beta(1-40)$ and $A\beta(1-42)$ as influenced by pH and a D-peptide,” *Journal of Physical Chemistry B*, vol. 116, no. 10, pp. 3280–3291, 2012.
- [120] T. Luhrs, C. Ritter, M. Adrian et al., “3D structure of Alzheimer’s amyloid-beta(1-42) fibrils,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 48, pp. 17342–17347, 2005.
- [121] C. Velez-Vega and F. A. Escobedo, “Characterizing the structural behavior of selected $A\beta$ -42 monomers with different solubilities,” *Journal of Physical Chemistry B*, vol. 115, no. 17, pp. 4900–4910, 2011.
- [122] A. T. Petkova, R. D. Leapman, Z. Guo, W.-M. Yau, M. P. Mattson, and R. Tycko, “Self-propagating, molecular-level polymorphism in Alzheimer’s β -amyloid fibrils,” *Science*, vol. 307, no. 5707, pp. 262–265, 2005.
- [123] A. R. A. Ladiwala, J. C. Lin, S. S. Bale et al., “Resveratrol selectively remodels soluble oligomers and fibrils of amyloid $A\beta$ into off-pathway conformers,” *The Journal of Biological Chemistry*, vol. 285, no. 31, pp. 24228–24237, 2010.
- [124] D. D. Soto-Ortega, B. P. Murphy, F. J. Gonzalez-Velasquez et al., “Inhibition of amyloid- β aggregation by coumarin analogs can be manipulated by functionalization of the aromatic center,” *Bioorganic and Medicinal Chemistry*, vol. 19, no. 8, pp. 2596–2602, 2011.
- [125] S. A. Moore, T. N. Huckerby, G. L. Gibson et al., “Both the D-(+) and L-(–) enantiomers of nicotine inhibit $A\beta$ aggregation and cytotoxicity,” *Biochemistry*, vol. 43, no. 3, pp. 819–826, 2004.
- [126] T. L. Lowe, A. Strzelec, L. L. Kiessling, and R. M. Murphy, “Structure-function relationships for inhibitors of β -amyloid toxicity containing the recognition sequence KLVFF,” *Biochemistry*, vol. 40, no. 26, pp. 7882–7889, 2001.
- [127] M. H. Viet, S. T. Ngo, N. S. Lam, and M. S. Li, “Inhibition of aggregation of amyloid peptides by beta-sheet breaker peptides and their binding affinity,” *The Journal of Physical Chemistry B*, vol. 115, no. 22, pp. 7433–7446, 2011.
- [128] M. Sarasa and P. Pesini, “Natural non-transgenic animal models for research in Alzheimer’s disease,” *Current Alzheimer Research*, vol. 6, no. 2, pp. 171–178, 2009.
- [129] C. D. Link, “Invertebrate models of Alzheimer’s disease,” *Genes, Brain and Behavior*, vol. 4, no. 3, pp. 147–156, 2005.
- [130] J. Wentzell and D. Kretzschmar, “Alzheimer’s Disease and tauopathy studies in flies and worms,” *Neurobiology of Disease*, vol. 40, no. 1, pp. 21–28, 2010.
- [131] J. L. Cummings, T. Morstorf, and K. Zhong, “Alzheimer’s disease drug-development pipeline: few candidates, frequent failures,” *Alzheimer’s Research & Therapy*, vol. 6, article 37, 2014.
- [132] D. Games, D. Adams, R. Alessandrini et al., “Alzheimer-type neuropathology in transgenic mice overexpressing V717F β -amyloid precursor protein,” *Nature*, vol. 373, no. 6514, pp. 523–527, 1995.
- [133] K. Hsiao, P. Chapman, S. Nilsen et al., “Correlative memory deficits, $A\beta$ elevation, and amyloid plaques in transgenic mice,” *Science*, vol. 274, no. 5284, pp. 99–102, 1996.
- [134] C. Stürchler-Pierrat, D. Abramowski, M. Duke et al., “Two amyloid precursor protein transgenic mouse models with Alzheimer disease-like pathology,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 24, pp. 13287–13292, 1997.
- [135] D. Van Dam, E. Vloeberghs, D. Abramowski, M. Staufenbiel, and P. P. De Deyn, “APP23 mice as a model of Alzheimer’s disease: an example of a transgenic approach to modeling a CNS disorder,” *CNS Spectrums*, vol. 10, no. 3, pp. 207–222, 2005.
- [136] J. M. Basak and D. M. Holtzman, “APP-based transgenic models of Alzheimer’s dementia: the PDAPP model,” in *Animal Models of Dementia*, P. P. De Deyn and D. Van Dam, Eds., pp. 371–385, Springer, New York, NY, USA, 2010.
- [137] R. M. J. Deacon, “APP-based transgenic models of Alzheimer dementia: the Tg2576 mouse,” in *Animal Models of Dementia*, P. P. De Deyn and D. Van Dam, Eds., pp. 387–398, Springer, New York, NY, USA, 2010.
- [138] D. Van Dam and P. P. De Deyn, “The APP23 mouse model for Alzheimer’s disease,” in *Animal Models of Dementia*, P.

- P. De Deyn and D. Van Dam, Eds., pp. 399–413, Springer Science+Business Media, New York, NY, USA, 2010.
- [139] E. McGowan, J. Eriksen, and M. Hutton, “A decade of modeling Alzheimer’s disease in transgenic mice,” *Trends in Genetics*, vol. 22, no. 5, pp. 281–289, 2006.
- [140] D. Van Dam and P. P. De Deyn, “Drug discovery in dementia: the role of rodent models,” *Nature Reviews Drug Discovery*, vol. 5, no. 11, pp. 956–970, 2006.
- [141] M. Sy, M. Kitazawa, and F. LaFerla, “The 3xTg-AD mouse model: reproducing and modulating plaque and tangle pathology,” in *Animal Models of Dementia*, P. P. De Deyn and D. Van Dam, Eds., vol. 48 of *Neuromethods*, pp. 469–482, Springer Science+Business Media, New York, NY, USA, 2011.
- [142] S. Brenner, “The genetics of *Caenorhabditis elegans*,” *Genetics*, vol. 77, no. 1, pp. 71–94, 1974.
- [143] D. Levitan and I. Greenwald, “Facilitation of lin-12-mediated signalling by sel-12, a *Caenorhabditis elegans* s182 Alzheimer’s disease gene,” *Nature*, vol. 377, no. 6547, pp. 351–354, 1995.
- [144] X. Li and I. Greenwald, “HOP-1, a *Caenorhabditis elegans* presenilin, appears to be functionally redundant with SEL-12 presenilin and to facilitate LIN-12 and GLP-1 signaling,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 22, pp. 12204–12209, 1997.
- [145] I. Daigle and C. Li, “apl-1, a *Caenorhabditis elegans* gene encoding a protein related to the human β -amyloid protein precursor,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 90, no. 24, pp. 12045–12049, 1993.
- [146] C. D. Link, “Expression of human beta-amyloid peptide in transgenic *Caenorhabditis elegans*,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 92, no. 20, pp. 9368–9372, 1995.
- [147] C. D. Link, “*C. elegans* models of age-associated neurodegenerative diseases: lessons from transgenic worm models of Alzheimer’s disease,” *Experimental Gerontology*, vol. 41, no. 10, pp. 1007–1013, 2006.
- [148] Y. Wu, Z. Wu, P. Butko et al., “Amyloid- β -induced pathological behaviors are suppressed by *Ginkgo biloba* extract EGB 761 and ginkgolides in transgenic *Caenorhabditis elegans*,” *Journal of Neuroscience*, vol. 26, no. 50, pp. 13102–13113, 2006.
- [149] B. C. Kraemer, B. Zhang, J. B. Leverenz, J. H. Thomas, J. Q. Trojanowski, and G. D. Schellenberg, “Neurodegeneration and defective neurotransmission in a *Caenorhabditis elegans* model of tauopathy,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 17, pp. 9980–9985, 2003.
- [150] D. C. Crowther, K. J. Kinghorn, E. Miranda et al., “Intraneuronal A β , non-amyloid aggregates and neurodegeneration in a *Drosophila* model of Alzheimer’s disease,” *Neuroscience*, vol. 132, no. 1, pp. 123–135, 2005.
- [151] B. Reisberg, R. Doody, A. Stöfler, F. Schmitt, S. Ferris, and H. J. Möbius, “Memantine in moderate-to-severe Alzheimer’s disease,” *The New England Journal of Medicine*, vol. 348, no. 14, pp. 1333–1341, 2003.
- [152] B. L. Apostol, A. Kazantsev, S. Raffioni et al., “A cell-based assay for aggregation inhibitors as therapeutics of polyglutamine-repeat disease and validation in *Drosophila*,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 100, no. 10, pp. 5950–5955, 2003.
- [153] I. Sánchez, C. Mahlke, and J. Yuan, “Pivotal role of oligomerization in expanded polyglutamine neurodegenerative disorders,” *Nature*, vol. 421, no. 6921, pp. 373–379, 2003.
- [154] I. Greeve, D. Kretschmar, J.-A. Tschäpe et al., “Age-dependent neurodegeneration and Alzheimer-amyloid plaque formation in transgenic *Drosophila*,” *Journal of Neuroscience*, vol. 24, no. 16, pp. 3899–3906, 2004.
- [155] S. K. Singh, *Studies on therapeutic potential of synthetic as well as herbal compounds for Alzheimer’s disease using transgenic Drosophila model [Doctoral thesis]*, Banaras Hindu University, Varanasi, India, 2015.
- [156] M. P. Cuajungco and G. J. Lees, “Zinc and Alzheimer’s disease: is there a direct link?” *Brain Research Reviews*, vol. 23, no. 3, pp. 219–236, 1997.
- [157] D. Beauchemin and R. Kisilevsky, “A method based on ICP-MS for the analysis of Alzheimer’s amyloid plaques,” *Analytical Chemistry*, vol. 70, no. 5, pp. 1026–1029, 1998.
- [158] M. A. Lovell, J. D. Robertson, W. J. Teesdale, J. L. Campbell, and W. R. Markesbery, “Copper, iron and zinc in Alzheimer’s disease senile plaques,” *Journal of the Neurological Sciences*, vol. 158, no. 1, pp. 47–52, 1998.
- [159] X. Huang, M. P. Cuajungco, C. S. Atwood, R. D. Moir, R. E. Tanzi, and A. I. Bush, “Alzheimer’s disease, β -amyloid protein and zinc,” *The Journal of Nutrition*, vol. 130, no. 5, pp. 1488S–1492S, 2000.
- [160] J. Constantinidis, *Mineral and Metal Neurotoxicology*, edited by: M. Yasui, M. Strong, K. Ota, A. Verity, CRC Press, Boca Raton, Fla, USA, 1997.
- [161] A. Rolfs and M. A. Hediger, “Metal ion transporters in mammals: structure, function and pathological implications,” *Journal of Physiology*, vol. 518, no. 1, pp. 1–12, 1999.
- [162] S. K. Singh, P. Sinha, L. Mishra, and S. Srikrishna, “Neuroprotective role of a novel copper chelator against A β_{42} induced neurotoxicity,” *International Journal of Alzheimer’s Disease*, vol. 2013, Article ID 567128, 9 pages, 2013.
- [163] F. Hane, G. Tran, S. J. Attwood, and Z. Leonenko, “Cu²⁺ affects amyloid- β (1–42) aggregation by increasing peptide-peptide binding forces,” *PLoS ONE*, vol. 8, no. 3, Article ID e59005, 2013.
- [164] M. Innocenti, E. Salvietti, M. Guidotti et al., “Trace copper(II) or zinc(II) ions drastically modify the aggregation behavior of amyloid- β 1–42: an AFM study,” *Journal of Alzheimer’s Disease*, vol. 19, no. 4, pp. 1323–1329, 2010.
- [165] S. Azimi and A. Rauk, “On the involvement of copper binding to the N-terminus of the amyloid beta peptide of Alzheimer’s disease: a computational study on model systems,” *International Journal of Alzheimer’s Disease*, vol. 2011, Article ID 539762, 15 pages, 2011.
- [166] V. Minicozzi, F. Stellato, M. Comai et al., “Identifying the minimal copper- and zinc-binding site sequence in amyloid- β peptides,” *Journal of Biological Chemistry*, vol. 283, no. 16, pp. 10784–10792, 2008.
- [167] P. Faller and C. Hureau, “Bioinorganic chemistry of copper and zinc ions coordinated to amyloid- β peptide,” *Dalton Transactions*, vol. 7, pp. 1080–1094, 2009.
- [168] C. J. Sarell, S. R. Wilkinson, and J. H. Viles, “Substoichiometric levels of Cu²⁺ ions accelerate the kinetics of fiber formation and promote cell toxicity of amyloid-beta from Alzheimer disease,” *The Journal of Biological Chemistry*, vol. 285, no. 53, pp. 41533–41540, 2010.
- [169] C. D. Syme and J. H. Viles, “Solution ¹H NMR investigation of Zn²⁺ and Cd²⁺ binding to amyloid-beta peptide (A β) of Alzheimer’s disease,” *Biochimica et Biophysica Acta—Proteins and Proteomics*, vol. 1764, no. 2, pp. 246–256, 2006.

- [170] Y. Zhang, S. Akilesh, and D. E. Wilcox, "Isothermal titration calorimetry measurements of Ni(II) and Cu(II) binding to His, GlyGlyHis, HisGlyHis, and bovine serum albumin: a critical evaluation," *Inorganic Chemistry*, vol. 39, no. 14, pp. 3057–3064, 2000.
- [171] S.-T. Liu, G. Howlett, and C. J. Barrow, "Histidine-13 is a crucial residue in the zinc ion-induced aggregation of the A β peptide of Alzheimer's disease," *Biochemistry*, vol. 38, no. 29, pp. 9373–9378, 1999.
- [172] D.-S. Yang, J. McLaurin, K. Qin, D. Westaway, and P. E. Fraser, "Examining the zinc binding site of the amyloid- β peptide," *European Journal of Biochemistry*, vol. 267, no. 22, pp. 6692–6698, 2000.
- [173] K. Suzuki, T. Miura, and H. Takeuchi, "Inhibitory effect of copper(II) on zinc(II)-induced aggregation of amyloid β -peptide," *Biochemical and Biophysical Research Communications*, vol. 285, no. 4, pp. 991–996, 2001.
- [174] R. A. Cherny, C. S. Atwood, M. E. Xilinas et al., "Treatment with a copper-zinc chelator markedly and rapidly inhibits β -amyloid accumulation in Alzheimer's disease transgenic mice," *Neuron*, vol. 30, no. 3, pp. 665–676, 2001.
- [175] A. K. Tickler, J. D. Wade, and F. Separovic, "The role of A β peptides in Alzheimer's disease," *Protein and Peptide Letters*, vol. 12, no. 6, pp. 513–519, 2005.
- [176] X. Huang, C. S. Atwood, M. A. Hartshorn et al., "The A β peptide of Alzheimer's disease directly produces hydrogen peroxide through metal ion reduction," *Biochemistry*, vol. 38, no. 24, pp. 7609–7616, 1999.
- [177] M. P. Cuajungco, L. E. Goldstein, A. Nunomura et al., "Evidence that the β -amyloid plaques of Alzheimer's disease represent the redox-silencing and entombment of A β by zinc," *The Journal of Biological Chemistry*, vol. 275, no. 26, pp. 19439–19442, 2000.
- [178] C. Opazo, X. Huang, R. A. Cherny et al., "Metalloenzyme-like activity of Alzheimer's disease β -amyloid: Cu-dependent catalytic conversion of dopamine, cholesterol, and biological reducing agents to neurotoxic H₂O₂," *The Journal of Biological Chemistry*, vol. 277, no. 43, pp. 40302–40308, 2002.
- [179] G. D. Ciccotosto, D. Tew, C. C. J. Curtain et al., "Enhanced toxicity and cellular binding of a modified amyloid β peptide with a methionine to valine substitution," *Journal of Biological Chemistry*, vol. 279, no. 41, pp. 42528–42534, 2004.
- [180] Y. Sagara, R. Dargusch, F. G. Klier, D. Schubert, and C. Behl, "Increased antioxidant enzyme activity in amyloid β protein-resistant cells," *Journal of Neuroscience*, vol. 16, no. 2, pp. 497–505, 1996.
- [181] L. E. Scott and C. Orvig, "Medicinal inorganic chemistry approaches to passivation and removal of aberrant metal ions in disease," *Chemical Reviews*, vol. 109, no. 10, pp. 4885–4910, 2009.
- [182] C. Hureau, I. Sasaki, E. Gras, and P. Faller, "Two functions, one molecule: a metal-binding and a targeting moiety to combat Alzheimer's disease," *ChemBioChem*, vol. 11, no. 7, pp. 838–953, 2010.
- [183] J. J. Braymer, A. S. DeToma, J.-S. Choi, K. S. Ko, and M. H. Lim, "Recent development of bifunctional small molecules to study metal-amyloid- β species in Alzheimer's disease," *International Journal of Alzheimer's Disease*, vol. 2011, Article ID 623051, 9 pages, 2011.
- [184] L. R. Perez and K. J. Franz, "Minding metals: tailoring multifunctional chelating agents for neurodegenerative disease," *Dalton Transactions*, vol. 39, no. 9, pp. 2177–2187, 2010.
- [185] A. Lakatos, É. Zsigó, O. D. Hollender et al., "Two pyridine derivatives as potential Cu(II) and Zn(II) chelators in therapy for Alzheimer's disease," *Dalton Transactions*, vol. 39, no. 5, pp. 1302–1315, 2010.
- [186] D. E. Green, M. L. Bowen, L. E. Scott et al., "In vitro studies of 3-hydroxy-4-pyridinones and their glycosylated derivatives as potential agents for Alzheimer's disease," *Dalton Transactions*, vol. 39, no. 6, pp. 1604–1615, 2010.
- [187] A. M. Haque, M. Hashimoto, M. Katakura, Y. Tanabe, Y. Hara, and O. Shido, "Long-term administration of green tea catechins improves spatial cognition learning ability in rats," *Journal of Nutrition*, vol. 136, no. 4, pp. 1043–1047, 2006.
- [188] S. Kuriyama, A. Hozawa, K. Ohmori et al., "Green tea consumption and cognitive function: a cross-sectional study from the Tsurugaya Project," *The American Journal of Clinical Nutrition*, vol. 83, no. 2, pp. 355–361, 2006.
- [189] K. Unno, F. Takabayashi, T. Kishido, and N. Oku, "Suppressive effect of green tea catechins on morphologic and functional regression of the brain in aged mice with accelerated senescence (SAMP10)," *Experimental Gerontology*, vol. 39, no. 7, pp. 1027–1034, 2004.
- [190] Y. Wang, L. Wang, J. Wu, and J. Cai, "The in vivo synaptic plasticity mechanism of EGb 761-induced enhancement of spatial learning and memory in aged rats," *British Journal of Pharmacology*, vol. 148, no. 2, pp. 147–153, 2006.
- [191] K. A. Youdim and J. A. Joseph, "A possible emerging role of phytochemicals in improving age-related neurological dysfunctions: a multiplicity of effects," *Free Radical Biology and Medicine*, vol. 30, no. 6, pp. 583–594, 2001.
- [192] A. N. Sokolov, M. A. Pavlova, S. Klosterhalfen, and P. Enck, "Chocolate and the brain: neurobiological impact of cocoa flavanols on cognition and behavior," *Neuroscience & Biobehavioral Reviews*, vol. 37, no. 10, part 2, pp. 2445–2453, 2013.
- [193] G. R. Beecher, "Overview of dietary flavonoids: nomenclature, occurrence and intake," *Journal of Nutrition*, vol. 133, no. 10, pp. 3248S–3254S, 2003.
- [194] J. P. E. Spencer, "Beyond antioxidants: the cellular and molecular interactions of flavonoids and how these underpin their actions on the brain," *Proceedings of the Nutrition Society*, vol. 69, no. 2, pp. 244–260, 2010.
- [195] J. P. E. Spencer, "The impact of flavonoids on memory: physiological and molecular considerations," *Chemical Society Reviews*, vol. 38, no. 4, pp. 1152–1161, 2009.
- [196] S. K. Singh, R. Gaur, A. Kumar, R. Fatima, L. Mishra, and S. Srikrishna, "The flavonoid derivative 2-(4'-benzyloxyphenyl)-3-hydroxy-chromen-4-one protects against A β 42-induced neurodegeneration in transgenic drosophila: insights from in silico and in vivo studies," *Neurotoxicity Research*, vol. 26, no. 4, pp. 331–350, 2014.
- [197] T. B. Joseph, S. W. J. Wang, X. Liu et al., "Disposition of flavonoids via enteric recycling: enzyme stability affects characterization of prunetin glucuronidation across species, organs, and UGT isoforms," *Molecular Pharmacology*, vol. 4, no. 6, pp. 883–894, 2007.
- [198] S. Ramos, "Effects of dietary flavonoids on apoptotic pathways related to cancer chemoprevention," *Journal of Nutritional Biochemistry*, vol. 18, no. 7, pp. 427–442, 2007.
- [199] S. A. R. Paiva and R. M. Russell, " β -carotene and other carotenoids as antioxidants," *Journal of the American College of Nutrition*, vol. 18, no. 5, pp. 426–433, 1999.
- [200] M. Y. Hong, N. P. Seeram, Y. Zhang, and D. Heber, "Anticancer effects of Chinese red yeast rice versus monacolin K alone on

- colon cancer cells," *Journal of Nutritional Biochemistry*, vol. 19, no. 7, pp. 448–458, 2008.
- [201] D. J. Newman and G. M. Cragg, "Natural products as sources of new drugs over the 30 years from 1981 to 2010," *Journal of Natural Products*, vol. 75, no. 3, pp. 311–335, 2012.
- [202] S. Asadi, A. Ahmadiani, M. A. Esmaeili, A. Sonboli, N. Ansari, and F. Khodaghali, "In vitro antioxidant activities and an investigation of neuroprotection by six *Salvia* species from Iran: a comparative study," *Food and Chemical Toxicology*, vol. 48, no. 5, pp. 1341–1349, 2010.
- [203] A. Kuruuzum-Uz, H. Suleyman, E. Cadirci, Z. Guvenalp, and L. Omur Demirezer, "Investigation on anti-inflammatory and antiulcer activities of *Anchusa azurea* extracts and their major constituent rosmarinic acid," *Zeitschrift fur Naturforschung C: Journal of Biosciences*, vol. 67, no. 7-8, pp. 360–366, 2012.
- [204] M. Dumont and M. F. Beal, "Neuroprotective strategies involving ROS in Alzheimer disease," *Free Radical Biology and Medicine*, vol. 51, no. 5, pp. 1014–1026, 2011.
- [205] J. Kim, H. J. Lee, and K. W. Lee, "Naturally occurring phytochemicals for the prevention of Alzheimer's disease," *Journal of Neurochemistry*, vol. 112, no. 6, pp. 1415–1430, 2010.
- [206] N. B. Chauhan and J. Sandoval, "Amelioration of early cognitive deficits by aged garlic extract in Alzheimer's transgenic mice," *Phytotherapy Research*, vol. 21, no. 7, pp. 629–640, 2007.
- [207] K. Ono, M. M. Condron, L. Ho et al., "Effects of grape seed-derived polyphenols on amyloid beta-protein self-assembly and cytotoxicity," *The Journal of Biological Chemistry*, vol. 283, no. 47, pp. 32176–32187, 2008.
- [208] D. S. Kim, J.-Y. Kim, and Y. S. Han, "Alzheimer's disease drug discovery from herbs: neuroprotectivity from β -amyloid (1–42) insult," *Journal of Alternative and Complementary Medicine*, vol. 13, no. 3, pp. 333–340, 2007.
- [209] J. F. Morton, "Folk uses and commercial exploitation of Aloe leaf pulp," *Economic Botany*, vol. 15, no. 4, pp. 311–319, 1961.
- [210] D. Yim, S. S. Kang, D. W. Kim, S. H. Kim, H. S. Lillehoj, and W. Min, "Protective effects of *Aloe vera*-based diets in *Eimeria maxima*-infected broiler chickens," *Experimental Parasitology*, vol. 127, no. 1, pp. 322–325, 2011.
- [211] T. Reynolds and A. C. Dweck, "Aloe vera leaf gel: a review update," *Journal of Ethnopharmacology*, vol. 68, no. 1–3, pp. 3–37, 1999.
- [212] F. Borrelli and A. A. Izzo, "The plant kingdom as a source of anti-ulcer remedies," *Phytotherapy Research*, vol. 14, no. 8, pp. 581–591, 2000.
- [213] S.-A. Im, Y.-R. Lee, Y.-H. Lee et al., "In vivo evidence of the immunomodulatory activity of orally administered *Aloe vera* gel," *Archives of Pharmacal Research*, vol. 33, no. 3, pp. 451–456, 2010.



Hindawi
Submit your manuscripts at
<http://www.hindawi.com>

