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PARASHIFT Probes: Solution NMR and X-ray Structural Studies of Macrocyclic Ytterbium and Yttrium Complexes

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ABSTRACT

Ytterbium and yttrium complexes of octadentate ligands based on 1,4,7,10-tetraazacyclododecane with a coordinated pyridyl group and either tricarboxylate (L^1) or triphosphinate (L^2) donors form twisted square antiprismatic structures. The former crystallises in the centrosymmetric group, $P2_1/c$, with the two molecules related by an inversion centre, whereas the latter was found as an unusual kryptoracemate in the chiral space group, $P2_1$. Pure shift NMR and EXSY spectroscopy allowed the dynamic exchange between the $(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ and $(RRR)\text{-}\Lambda\text{-}(\lambda\lambda\lambda\lambda)$ TSAP diastereomers of the $[\text{Y}\cdot L^2]$ complex to be detected. The rate-limiting step in the exchange between Δ and Λ isomers involves cooperative ligand arm rotation, which is much faster for $[\text{Ln}\cdot L^1]$ than $[\text{Ln}\cdot L^2]$. Detailed analysis of NOESY, COSY, HSQC and HMBC spectra confirms that the major conformer in solution is $(RRR)\text{-}\Lambda\text{-}(\lambda\lambda\lambda\lambda)$, consistent with crystal structure analysis and DFT calculations. The magnetic susceptibility tensors for $[\text{Yb}\cdot L^1]$ and $[\text{Yb}\cdot L^2]$, obtained from a full pseudocontact chemical shift analysis, are very different, in agreement with a CASSCF calculation. The remarkably different pseudocontact shift behavior is explained by the change in the orientation of the pseudocontact shift field, as defined by the Euler angles of the susceptibility tensor.

INTRODUCTION

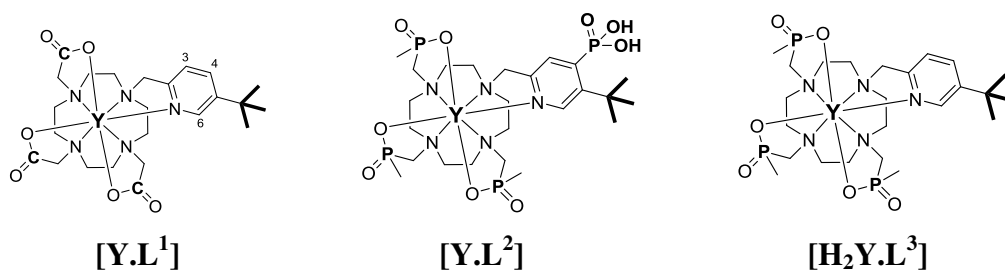
Polyaza macrocyclic ligands with eight or nine donor atoms are known to form kinetically stable complexes with rare earth elements.^{1,2} Over the past 30 years, they have found use in analytical, biochemical and clinical applications, ranging from the widespread adoption of gadolinium contrast agents in magnetic resonance imaging,³ to their role in radioimmunotherapy and radiosciintigraphy using complexes or conjugates labelled with ^{90}Y , ^{153}Sm , ^{149}Tb or ^{177}Lu .⁴ In each of these cases, premature release of the metal ion must be avoided at all costs – both the free ligands and the lan-

thanide ions may give rise to an acute or chronic toxic response.

In recent work, examining the scope and utility of paramagnetic metal complexes in magnetic resonance shift imaging, examples of temperature probes based on complexes of Fe(II), Co(II) or lanthanide(III) ions have been described.^{5, 6-12} A key property of these complexes is that the chemical shift of a reporter nucleus in the ligand gives information not only about its position relative to the paramagnetic ion but also about local temperature.⁵⁻¹² By careful ligand design, the chemical shift of a given reporter nucleus can be made to report local pH simultaneously,¹³⁻¹⁵ or indeed pM or pX, in the general cases,¹⁶ provided that care is taken to design the coordinating ligand rationally.

A key design objective is to create a metal complex that exists as one preferred stereoisomer that is conformationally rigid on the NMR timescale, so that signal intensity is maximised and exchange broadening minimised. Strategies that lead to preferential formation of one solution isomer, or slow down intramolecular motion, have been considered before.¹⁷ For example, the series of octadentate ligands based on mono-substituted triphosphate derivatives of 1,4,7,10-tetra-azacyclododecane (*i.e.* cyclen or 12-N₄) have been studied in detail.¹⁸⁻²⁰ Preferred formation of one relatively rigid isomer (out of 32 possible isomers) was established back in 1995,¹⁸ and the reasons for this selectivity can be traced to the need to minimise steric hindrance around the metal centre, in which the relative position of the three phosphinate substituents (*e.g.* Me, Bn or Ph) and the large bite angle associated with the three N-C-P-O-Ln 5-membered chelate rings are of paramount importance. No directly relevant crystal structures have been reported of monomeric lanthanide(III) triphosphate complexes, based on octadentate ligands.

Chart 1 Structures of the yttrium and ytterbium complexes examined in this study



Very recently, the later lanthanide (Tb, Dy, Ho, Er, Tm) complexes have been created of such ligands, substituted with a “reporter” *t*-butyl group located just over 6 Å away from the metal ion (Chart 1). Their scope as PARASHIFT probes is being evaluated^{21,22}, driven by the prospect of using them, or their simple conjugates with a targeting vector, for the assessment of local temperature and pH in dual or triple proton MRI imaging studies.

The complexes exhibit ¹H NMR paramagnetic shifts of the *t*-butyl resonance in [Ln.L²] that are 3 to 4 times larger than in the [Ln.L¹] complexes. Thus, the reporter *t*-butyl signal in [Ln.L²] resonates over the range +76 ppm (Tm) to –83 ppm (Dy/Tb) at 295 K, with the longitudinal rates of relaxation of the ¹Bu reporter group varying from 80 to 200 s⁻¹ at 7 Tesla, permitting the use of fast pulse repetition rates in imaging and chemical shift imaging experiments. A 15 to 25-fold signal intensity gain over the diamagnetic controls can then be obtained in imaging experiments. The paramagnetic probes can be administered at doses of the order of 0.1 mmole/kg, giving sufficient signal to allow the fate of the complex *in vivo* to be tracked, or to obtain regional temperature or pH maps. These doses are similar to those of the ubiquitous Gd complexes that are used in millions of MRI scans every year.

Here, we present X-ray structures for [Yb.L¹] and [Yb.L²], and examine their diamagnetic Y(III) analogues to establish solution-state structures and conformational exchange dynamics. We examine the origins of the differing shift behaviour of [Yb.L¹] and [Yb.L²] as models for the remaining series, by studying the pseudocontact shift field in detail, relating behaviour to the nature of the magnetic susceptibility tensor. We also report the properties of Yb and Y complexes of the 4-phosphonate derivative of L² – those are important examples of complexes exhibiting temperature- and pH-dependent chemical shifts, for which parallel administration of the Tm and Dy analogues has recently allowed triple imaging studies *in vivo*, simultaneously reporting pH and temperature, as well as detailing anatomy *via* co-observation of the water signal.²²

RESULTS AND DISCUSSION

The ligands L¹⁻³ and their complexes were prepared using previously reported methods.^{19,22} Crystals of the Yb complexes were grown by diffusion of diethyl ether into a methanol solution for [Yb.L¹], and into an ethanol solution for [Yb.L²], (Figures 1

and 2, and ESI: Figures S16, S19, S20). Crystal structures show a common twisted square anti-prismatic coordination geometry with the Yb ion bound to the four ring nitrogen atoms, the pyridine nitrogen atom and the three anionic oxygen atoms.

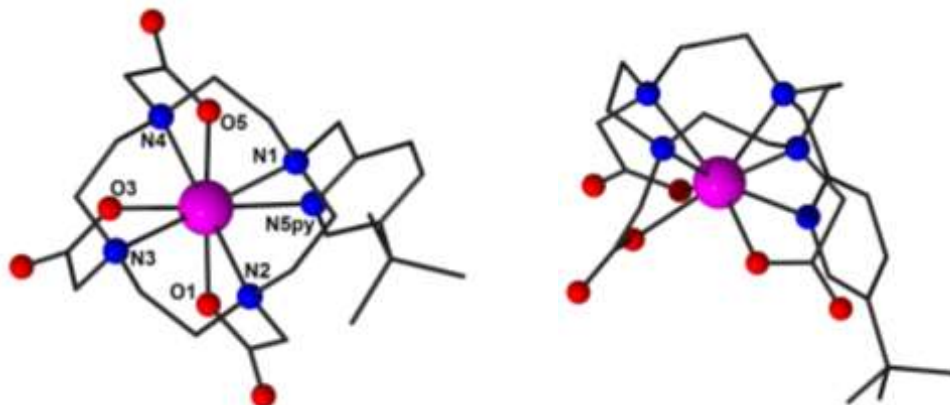


Figure 1 Molecular structure of Δ -($\delta\delta\delta\delta$)-[Yb.L¹], viewed down the 12-N₄ ring (*left*) and from the side (*right*). H atoms omitted for clarity; the enantiomer was present in the unit cell, CCDC-1502162.

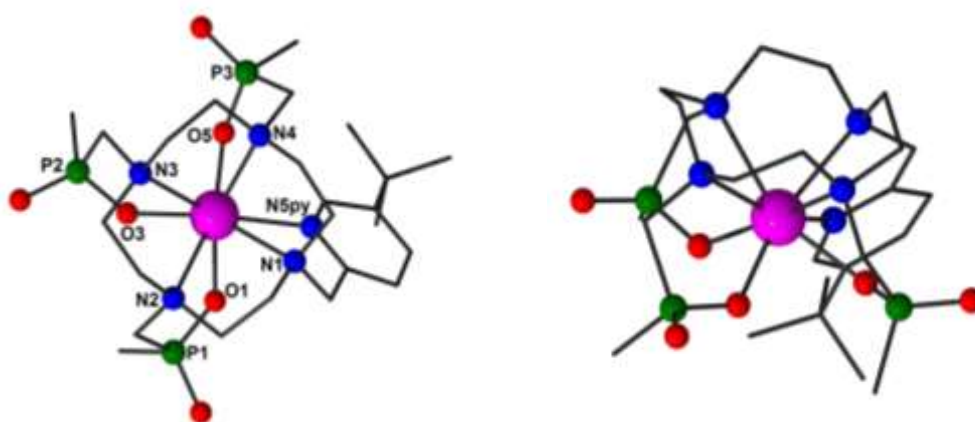


Figure 2 Molecular structure of (RRR)- Λ -($\lambda\lambda\lambda\lambda$) [Yb.L²], viewed down the 12-N₄ ring (*left*) and from the side (*right*). Hydrogen atoms and disorder in the *tert*-butyl pyridine group are omitted for clarity. The complex (SSS)- Λ -($\delta\delta\delta\delta$) [Yb.L²] was also present in the unit cell. The bond distances in the two enantiomeric molecules are, respectively: Yb1-P1, 3.3312(16) & 3.3410(15); Yb1-P2, 3.3856(15) & 3.3714(14); Yb1-P3, 3.3548(16) & 3.3710(16) Å, in agreement with values calculated in related structures, by examining the field dependence of their ³¹P NMR relaxation rates,¹⁸ CCDC-1501473.

Bond distances were systematically shorter between Yb and oxygen in the triphosphate compared to the tricarboxylate complex, and were correspondingly longer to each of the nitrogen donor atoms. Such behaviour reflects both the preference for the

anionic phosphinate oxygen donor and the need for the Yb ion to sit higher above the 12-ring N_4 plane, in order to allow cooperative ligation of the four nitrogen substituents.²³

Table 1. Bond distances in the N_5O_3 coordination sphere for $[Yb.L^1]$ and $[Yb.L^2]$

Bond	Bond distance (Å)	
	$[Yb.L^1]$	$[Yb.L^2]^a$
Yb1-O1	2.242(3)	2.212(4)
Yb1-O3	2.249(3)	2.221(4)
Yb1-O5	2.243(3)	2.206(4)
Yb1-N1	2.521(3)	2.557(6)
Yb1-N2	2.515(3)	2.592(6)
Yb1-N3	2.507(3)	2.613(4)
Yb1-N4	2.515(3)	2.638(5)
Yb1-N5py	2.485(3)	2.569(3)

a) given as the average for the two molecules in the unit cell.

The torsion angles in the four NCCN chelate rings associated with the 12- N_4 ligand are of the same sign in each case, defining the λ/δ configurations reported in Tables 2 and 3. Similarly, NCCN_{py} and NCCO torsion angles define the Δ/Λ configuration (Δ is equivalent to *P* helicity). The smaller twist angle of the “twisted” square-antiprism, (Table 3, averaging $\sim 26^\circ$) compared to the “regular” square antiprism (typically $\sim 40^\circ$) has been observed many times before, notably in the four-fold symmetric N_4O_4 complexes of the analogous carboxylate and phosphinate ligands, that lack a coordinating pyridine group.²⁴

The complex $[Yb.L^1]$ crystallised in a centrosymmetric space group ($P2_1/c$), so that both enantiomers were present and symmetry related. Examples of tricarboxylate and triphosphinate derivatives of 12- N_4 have been reported previously, but are restricted to dimeric species based on heptadentate ligands for triphosphinate derivatives. Tri-carboxylate examples are more prevalent and varied, but only three octadentate ligands crystallise as 8-coordinate tricarboxylate species.^{25,26} $[Yb.L^2]$ is an interesting case of a kryptoracemate,²⁷ crystallizing in a chiral (Sohncke) space group $P2_1$, but with both enantiomers present as symmetrically positioned independent molecules.

The structure also resembles space group $P2_1/c$, in which it can be crudely refined, by assuming extensive disorder.^{28,29}

Table 2. Torsion angles in the 12-N₄ ring (N-C-C-N) for [Yb.L¹] and [Yb.L²].

Torsion chain	Torsion angle (°)		
	[Yb.L ¹] ^a	[Yb.L ²] ($\lambda\lambda\lambda$)	[Yb.L ²] ($\delta\delta\delta\delta$)
N1-C1-C2-N2	±56.7(5)	-58.4(8)	62.9(7)
N2-C3-C4-N3	±55.7(5)	-58.3(7)	59.2(7)
N3-C5-C6-N4	±56.9(5)	-55.3(8)	57.5(7)
N4-C7-C8-N1	±58.3(5)	-59.2(7)	57.7(7)

^a + and -, respectively, for $\delta\delta\delta\delta$ and $\lambda\lambda\lambda$ enantiomers, inversion-related in space group $P2_1/c$

Table 3. Torsion angles around the exocyclic groups for [Yb.L¹] and [Yb.L²].

Torsion chain	Torsion angle (°)		
	[Yb.L ¹] ^a	[Yb.L ²] (Δ)	[Yb.L ²] (Δ)
N1-C15-C16-N5py	25.9(6)	-28.5(9)	30.0(9)
N2-C9-C10(P1)-O1	21.8(6)	-29.2(5)	27.3(6)
N3-C11-C12(P2)-O3	21.6(5)	-31.2(4)	33.7(4)
N4-C13-C14(P3)-O5	21.0(6)	-48.0(4)	46.8(4)

^a + and -, respectively, for Δ and Δ enantiomers, inversion-related in space group $P2_1/c$

Table 4. Twist angles between the NO₃ and N₄ coordination faces of [Yb.L¹] and [Yb.L²].

Plane	Angle (°)	
	[Yb.L ¹]	[Yb.L ²] ^a
N1-Yb-N3 vs N5py-Yb-O3	25.1	25.7
N5py-Yb-O3 vs N4-Yb-N2	65.0	65.4
N4-Yb-N2 vs O5-Yb-O1	26.6	27.0
O5-Yb-O1 vs N1-Yb-N3	63.6	64.6

a) average for the two independent molecules

DFT Studies

The relative free energies of the possible isomeric yttrium complexes of $[Y.L^2]$ were computed using DFT methods. In gas phase geometry optimisations, the two lowest energy structures found (Table 5) were the square anti-prismatic $(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ and the twisted square anti-prismatic $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda\lambda)$ isomers. The other two diastereomers with an $(RRR)\text{-}\Delta$ configuration were about 20 kcal/mol higher in energy, reflecting the destabilisation of the overall structure as a result of unfavourable steric interactions involving the P-methyl groups, pointing towards the 12-N₄ ligand, with the more exposed phosphorus oxygen groups.

Table 5. Calculated relative enthalpies and free energies for diastereoisomers of $[Y.L^2]$

Complex isomer	BP86 (gas phase)		M06-2X (gas phase)		M06-2X (water SMD)	
	ΔH_0 kcal/mol	ΔG_{298} kcal/mol	ΔH_0 kcal/mol	ΔG_{298} kcal/mol	ΔH_0 kcal/mol	ΔG_{298} kcal/mol
$(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ SAP	0	0	2	3	5	7
$(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda\lambda)$ TSAP	0	1	0	0	0	0
$(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ TSAP	18	18	17	17	3	3
$(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda\lambda)$ SAP	23	23	24	25	12	11

However, the presence of a polarisable continuum solvent (M06-2X with SMD water) stabilises the pair of $(RRR)\text{-}\Delta$ isomers: the phosphinate oxygen atoms gain more negative charge (Mulliken point charges increase from -0.5 to -0.65) and the average Ln-O bond length increases from ~ 2.2 to ~ 2.3 Å, causing the methyl groups to be further away from the 12-N₄ ligand. In the $(RRR)\text{-}\Delta$ pair of diastereomers, solvation stabilisation is less pronounced, with the phosphinate groups partially shielded by the macrocyclic 12-N₄ ligand.

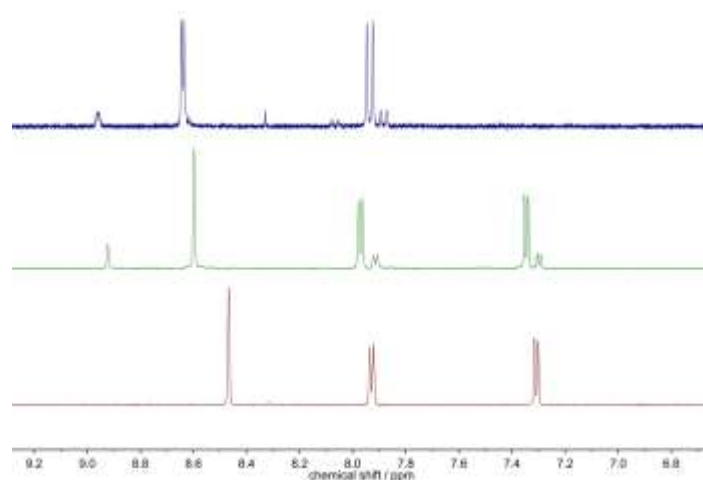
DFT Calculations in a polarisable continuum solvent were also run for $[Y.L^1]$, comparing the stability of the SAP $\Delta\text{-}(\lambda\lambda\lambda\lambda)$ and the TSAP $\Delta\text{-}(\lambda\lambda\lambda\lambda)$ diastereomers, both in the absence (CN = 8) and presence (CN = 9) of an axial coordinated water molecule. The SAP isomer preferred to be 9-coordinate, with a water-Y(III) distance of 2.56 Å. With the TSAP isomer, the complex was also formally 9-coordinate, but the nearest water ended up being hydrogen bonded to the coordinated carboxylate ox-

ygen, and the distance of 3.23 Å from the Y(III) ion is consistent with the idea of “partial hydration”^{18b} in which there is a rather long Y(III)-water distance, that corresponds to a q value of the order of 0.25. This study therefore indirectly reaffirms the early hypothesis concerning partial hydration numbers, in which both short Ln-O bonds and classical $q = 1$ behaviour, and long Ln-O bonds, *i.e.* partial (non-classical) hydration states may occur, determined by the nature of the local coordination environment.^{18b, 18c} In each case, the overall free energy of binding associated with hydration is relatively small, ~2 kcal/mol. The lowest energy form was calculated to be the TSAP form (as also observed by crystallography) but the nine-coordinate SAP structure for [Y.L¹] was only 1.5 kcal/mol higher in energy.

The DFT studies have thus suggested that the favoured solution isomers for [Y.L²] and [Y.L¹] are the 8-coordinate (*RRR*)- Δ -($\lambda\lambda\lambda\lambda$) twisted square anti-prismatic (TSAP) structures, that are also observed in the crystal structures. For [Y.L²], the (*RRR*)- Δ -($\delta\delta\delta\delta$) TSAP diastereomer is of comparable energy. The error in these calculations is of the order of a few kcal/mol,³⁰ and so several isomeric complexes may be expected to be observed in solution by NMR spectroscopic methods.

Solution NMR of yttrium(III) complexes: static and dynamic analysis

¹H and ³¹P NMR spectra were analysed for the diamagnetic yttrium(III) complexes prior to the examination of the ytterbium(III) complexes, whose structure had been determined by X-ray crystallography.



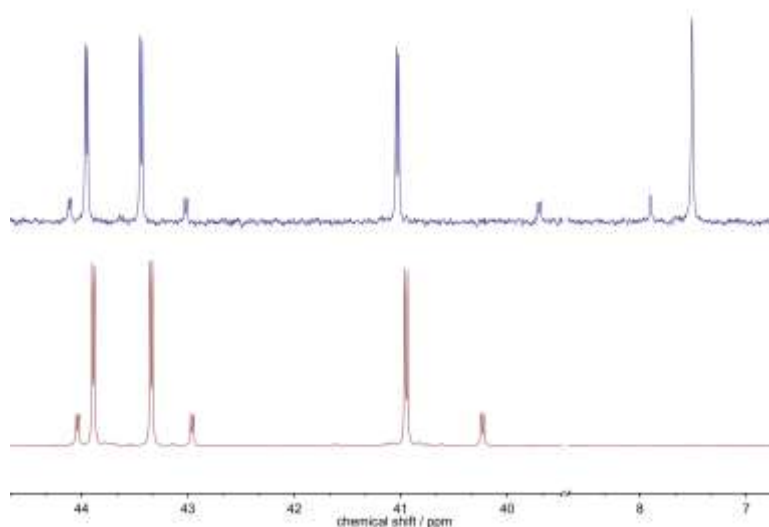


Figure 3. (*upper*) ^1H NMR spectra (600 MHz, D_2O , pH 7.4, 295 K), of the pyridyl ring protons of $[\text{Y.L}^1]$ (*red*), $[\text{Y.L}^2]$ (*green*), and $[\text{Y.L}^3]^{2-}$ (*blue*), revealing two major isomers for each phosphinate complex; (*lower*) $^{31}\text{P}\{^1\text{H}\}$ NMR spectra (14.1 T, D_2O , 298 K), of $[\text{Y.L}^2]$ (*red*), and $[\text{Y.L}^3]^{2-}$ (*blue*), showing the major and minor isomers and the small ^{89}Y - ^{31}P couplings.

^1H NMR analysis of $[\text{Y.L}^2]$ and $[\text{Y.L}^3]^{2-}$ in D_2O solution revealed that there are two sets of resonances for each pyridine ring proton in the triphosphinate complexes, in a ratio of 1:5 and 1:7, respectively (Figure 3). In contrast, analysis of the ^1H NMR spectrum of the tricarboxylate complex, $[\text{Y.L}^1]$, was consistent with the presence of one major species in solution. Alternatively, two or more species may be present in solution, that must be in relatively fast exchange on the NMR timescale (see also ESI: Figures S8, S10, S11 and *vide infra*). Indeed, the NOESY spectrum for $[\text{Y.L}^1]$ at 295 K showed that the diastereotopic pyridyl NCH_2 resonances at 4.26 and 3.84 ppm were undergoing chemical exchange, as the sign of the ^1H - ^1H cross peaks was the same as the sign of the diagonal, consistent with a pair of resonances undergoing chemical exchange. In contrast the ^1H - ^1H cross-peak was of opposite sign for the pyridyl hydrogens, H_3 and H_4 , resonating at 7.93 and 7.31 ppm (ESI, Figure S11), consistent with a classical nuclear Overhauser through-space effect.

By correlating the observed NOE signals with H-H distances in the structure, it is possible to distinguish between SAP and TSAP structural types. Di Bari has shown that two strong NOE cross-peaks are expected between exocyclic NCH_2 protons and endocyclic ring protons expected for a SAP structure, whereas there should only be one present for TSAP.³¹ These cross-peaks are more intense between protons with

an H-H distance below 2.2 Å and are much stronger than other cross-peaks for H-H distances >2.5 Å. The closest H-H distances for exocyclic and endocyclic protons were determined (Table 6) for the structures of [Ln.L¹] and [Ln.L²]. Two strong NOE cross-peaks were observed for [Y.L¹] and one for [Y.L²]. There is a clear reduction in intensity when the distance increases above 2.3 Å.

Such behaviour is consistent with the hypothesis that the major isomers present in solution for [Y.L¹] and [Y.L²] are SAP and TSAP structures respectively.

Table 6. NOESY spectral analysis and inter-proton distances in [Y.L¹] and [Y.L²].

[Y.L ¹] ^a				[Y.L ²]			
Protons	NOE cross-peak /ppm	Relative intensity	Distance / Å ^b	Protons	NOE cross-peak / ppm	Relative intensity	Distance / Å ^c
apax→c8ax	4.26→2.55	1	1.98	apax→c2ax	4.55→3.58	1	2.12
apeq→c1eq	3.84→2.82	0.8	2.20	apax→c1eq	4.55→2.47	0.35	2.58
apeq→c8eq	3.84→2.72	0.2	2.68	apeq→c1eq	3.65→2.47 ^d	0.4	2.39
				apeq→c8eq	3.65→2.43 ^d	0.4	2.47

^aexchange evidenced by EXSY cross-peaks renders equivalent the apax and apeq signals; ^b from the DFDT structure of Δ-λλλλ, i.e. a SAP structure with a coordinated H₂O optimized geometry ESI S1; ^cfrom the [Yb.L²] X-ray crystal structure; ^d overlapping resonances, so the same relative intensity value is given here, averaged over each signal; normally expect relative NOE intensities to vary as r³/r'³.

It has been established that cooperative 12-N₄ ring inversion occurs at a rate of about 50 to 200 s⁻¹ (298 K), and leads to formal exchange between ‘axial’ and ‘equatorial’ ring hydrogens and ‘opposite’ ring carbons (e.g., C₁/C₈; C₁/C₇) in the λλλλ and δδδδ isomers.^{17,18} Moreover, cooperative arm rotation of the exocyclic N substituents interchanges Δ and Λ isomers and usually occurs more slowly, and exchanges the exocyclic methylene hydrogens, i.e. the CH₂CO, CH₂PO or CH₂py hydrogen atoms in this case.^{23,24} Thus, in [Y.L¹], there is direct evidence that cooperative arm rotation is occurring.

The three phosphinate atoms in [Y.L²] and [Y.L³]²⁻ are chemically non-equivalent, and a major and minor isomer was also observed, in each case, by ³¹P NMR spectroscopy.

copy. These isomers were in slow exchange on the NMR timescale (Figure 3 and SI Figure S9). Through-bond J coupling was observed from ^{89}Y ($I = 1/2$, 100% abundance) to each phosphinate ^{31}P nucleus, ($^2J_{\text{YP}} = 5$ Hz), giving rise to a series of doublets in the range 45 to 39 ppm, whereas the phosphonate ^{31}P resonance in $[\text{Y.L}^3]$ is too far away (5 bonds) from Y for any coupling to be observed (singlets at 7.5 ppm and 7.9 ppm for the major and minor isomer respectively). The presence of the phosphonate group at the 4-position of the pyridine ring in $[\text{Y.L}^3]^{2-}$ has little effect on the ^{31}P chemical shifts of the phosphinate groups, except for the minor isomer peak at 39.7 ppm (shifted from 40.2 ppm in $[\text{Y.L}^2]$). Taken together, these data, along with the pH dependence of the 41.0/39.7 ppm pair of ^{31}P resonances, suggest that the major ^{31}P peak at ~41 ppm is the one that is closest to the pyridine ring (ESI: Figures S6 and S7).

Detailed proton NMR assignments for each yttrium complex were made with the aid of ^1H - ^{13}C HSQC, ^1H - ^{31}P HMBC, ^1H - ^{13}C HMBC as well as pure shift NMR experiments (ESI; Figures S1–5, S8). The resonances associated with each of the three phosphinate arms for the triphosphinate complexes, $[\text{Y.L}^2]$ and $[\text{Y.L}^3]^{2-}$, were assigned (SI Table S2, and Experimental section). The pH dependence of the single PCH_2N arm in $[\text{Y.L}^3]^{2-}$ is associated with protonation of the phosphonate group on the pyridine moiety, for which the $\text{p}K_{\text{a}}$ has been established to be 7.15(6) (295K, $I = 0$)²² in the Dy, Ho, Er and Tm(III) complexes. The ionic radius of Y(III) falls between that of Ho(III) and Er(III).

Full assignment of all 31 chemically distinct protons of $[\text{Y.L}^2]$ (Table S2) was undertaken by detailed analysis of all available NMR data. Pure shift ^1H NMR techniques^{32,33} proved to be very useful, and in the present case the removal of the ^1H - ^1H splitting was achieved using the PSYCHE pulse sequence;³⁴ ^1H - ^{31}P splitting was removed by standard ^{31}P decoupling methods (Figure 4).

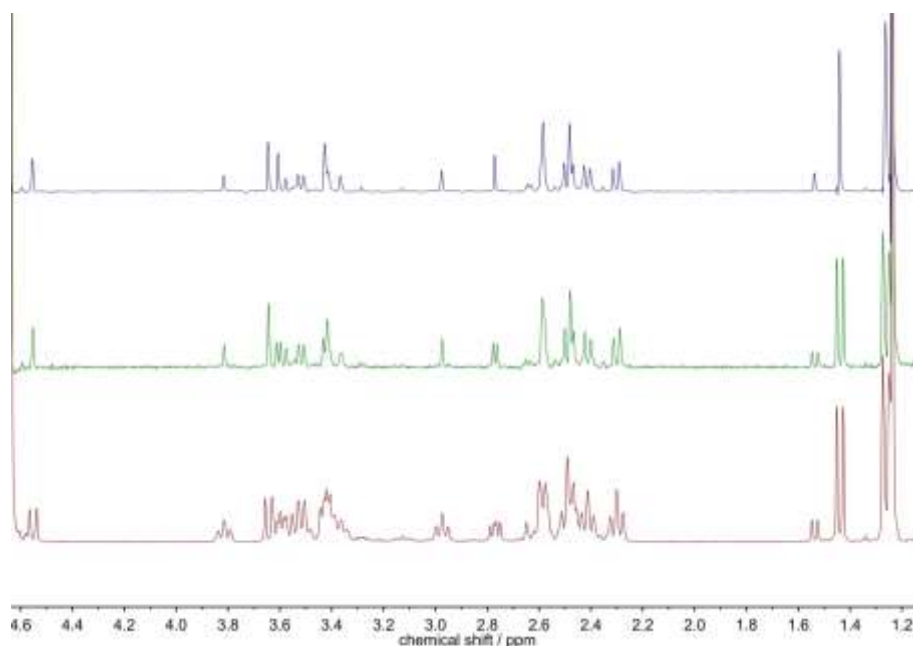


Figure 4. ^1H (*red*), ^1H { ^1H decoupled} (*green*), and ^1H { ^1H and ^{31}P decoupled} (*blue*) NMR spectra (14.1 T, D_2O , pD 6, 295 K), for the triphosphate complex, $[\text{Y.L}^2]$, in the shift range 1.1 to 4.6 ppm.

There is only one $[\text{Y.L}^2]$ conformer namely $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda)$ that does not contradict the observed NOESY spectrum (Figure). Each methyl group has only two NOE cross-peaks with axial and equatorial protons of corresponding arms. The absence of additional cross-peaks with cyclen protons rules out $(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ and $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda)$ conformers because in these cases, the methyl groups are closer to cyclen than to the exocyclic CH_2 protons (ESI Figure S7). The other important feature in the NOESY spectrum is the presence of two cross-peaks for the axial proton of the pyridine arm at 4.55 ppm with cyclen protons attached to different carbons; such a feature rules out the presence of the $(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ conformer, because it has only neighbouring cyclen protons that are attached to the same carbon (ESI Figure S8).

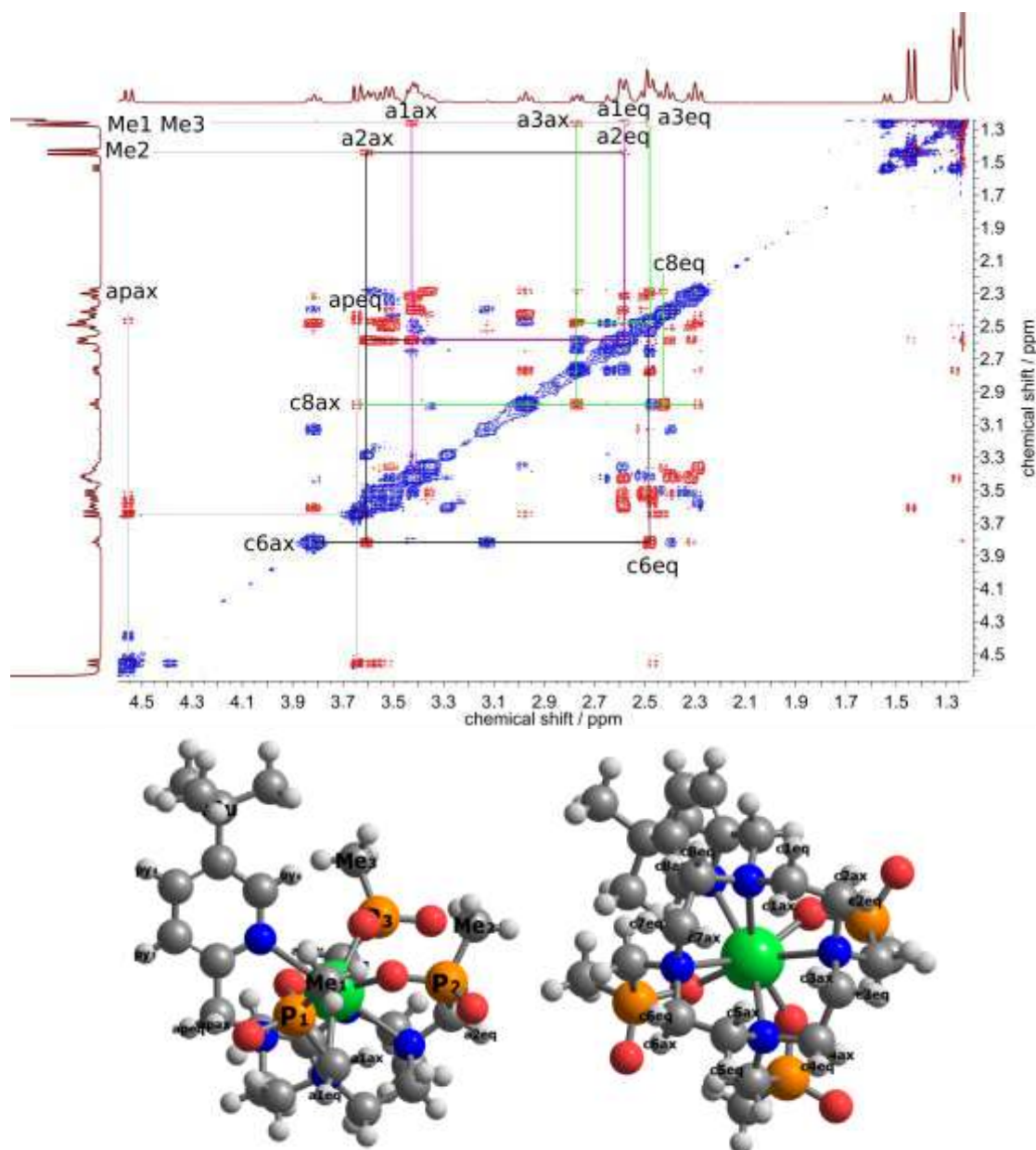


Figure 5. (*upper*): ^1H - ^1H 2D NOESY/EXSY NMR spectrum (14.1 T, D_2O , 298 K, mixing time = 300 ms), of the aromatic protons of $[\text{Y.L}^2]$, showing exchange correlations (*blue*) as well as through-space correlations (*red*). Important NOE cross-peaks that suggest $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda)$ conformation are highlighted (see text for the discussion); (*lower*) Labels for atoms used in the assignment tables. The cyclen protons and carbons are labeled with 'c' and ordered clockwise, where 'ax' stands for axial and 'eq' stands for equatorial protons. Exocyclic methylene protons are labeled with 'a' and numbered the same way; pyridine arm is labeled 'ap'.

Through-space NOE correlations were observed between H_4 and H_3 , out-of-phase with the diagonal, due to the close proximity of these two protons. Evidence for exchange was observed between the singlets at 8.92 (H_6 , minor) and 8.60 (H_6 , major), doublets at 7.97 (H_4 major) and 7.91 (H_4 minor), and doublets at 7.35 (H_3 major) and

7.30 (H₃ minor), (Figure 6), showing that these are the same pyridine protons in the two different conformational isomers of [Y.L²]. Similar observations were seen in [HY.L³]⁻: H₆ = 8.64 ppm (major) and 8.96 ppm (minor); H₃ = 7.93 ppm (major), and 7.88 ppm (minor). Additionally, ³¹P exchange was observed for [²HY.L³]⁻, (ESI: Figure S12).

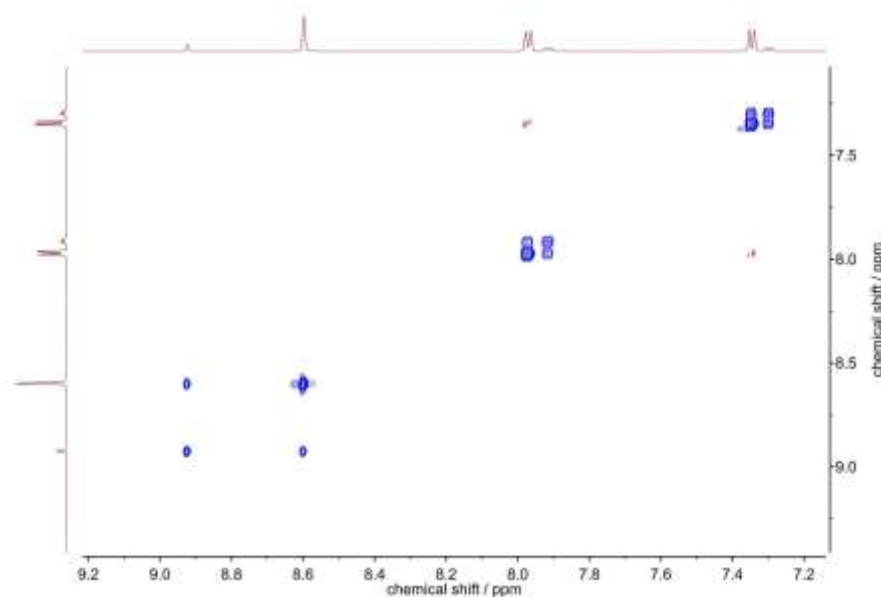


Figure 6. ¹H-¹H 2D NOESY/EXSY NMR spectrum (14.1 T, D₂O, 298 K, mixing time = 300 ms), of the aromatic protons of [Y.L²], showing exchange correlations (*blue*) as well as through-space correlations (*red*).

Exchange spectroscopy (EXSY) calculations were performed using the MestreNova EXSYCalc programme, using the data obtained from two EXSY spectra, one with a mixing time of zero ms and another with a mixing time of 300 ms. The rate of exchange between the minor and the major isomer was of the order of 1.1 to 1.4 s⁻¹. The reverse rates were slower, and fell in the range 0.17 to 0.23 s⁻¹, (Table 7).

Table 7 EXSY exchange rates between isomers for [Y.L²] and [HY.L³]⁻ (295K, D₂O, pD 7)

Complex	Exchange pair	k / s^{-1}	k^{-1} / s^{-1}	K_{eq}
[Y.L ²]	H ⁶ (pyridine ¹ H)	1.13(3)	0.23(2)	4.9(3)
[Y.L ³]	H ⁶ (pyridine ¹ H)	1.38(3)	0.18(2)	7.7(5)
[Y.L ³]	³¹ P phosphinate (41.0-39.7 ppm pair)	1.34(4)	0.16(2)	8.4(5)

Functionalisation at the 4'-position of the pyridine ring does not significantly alter the rate at which the two conformational isomers interconvert. The equilibrium constant for $[\text{Y}.\text{L}^2]$ is 4.9(3), whereas that of $[\text{HY}.\text{L}^3]^-$ is larger, averaging 8.0(5), in reasonable agreement with the different isomer ratios observed (5:1 and 7:1) for the major and minor species by integration of the fully relaxed NMR spectra of each complex.

Pseudocontact shift analysis Two conformers are observed for each complex with a ratio of 3:1 (Figure 7) for $[\text{Yb}.\text{L}^1]$ and 5:1 for $[\text{Yb}.\text{L}^2]$. Several exchange pairs were observed in the EXSY spectra of $[\text{Yb}.\text{L}^1]$, (ESI: Figure S10), whereas cross peaks were more difficult to observe with $[\text{Yb}.\text{L}^2]$, emphasising the difference in their dynamic exchange behaviour. Using EXSYCalc for two EXSY spectra with 5 ms and 100 ms mixing times (looking at the pyridine H^4 signal), it was possible to estimate a k_1 rate for $[\text{Yb}.\text{L}^1]$ of 54 s^{-1} , 50 times faster than for $[\text{Y}.\text{L}^2]$, and a k_{-1} value of 16 s^{-1} . These rate values are consistent with reported rates of SAP to TSAP isomerisation observed in related 12- N_4 based lanthanide complexes.²⁴

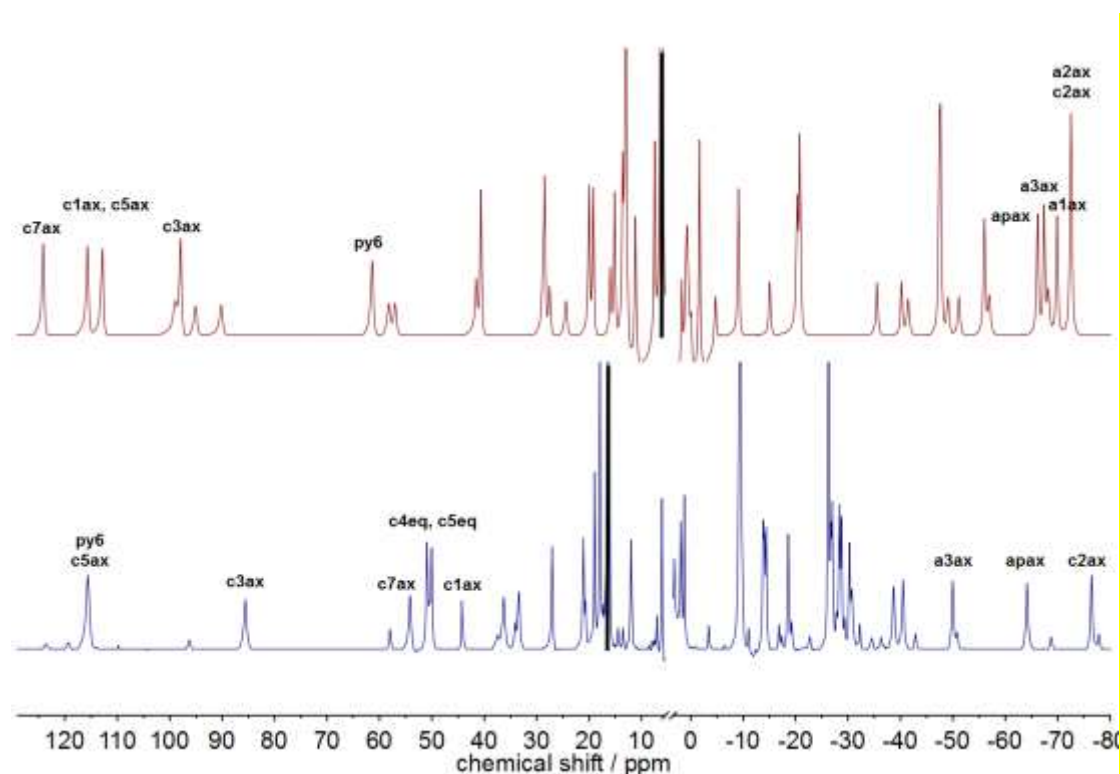


Figure 7. ^1H NMR spectrum of $[\text{Yb}.\text{L}^1]$ (red) and $[\text{Yb}.\text{L}^2]$ (blue) (4.7 T, D_2O , 295 K); spectra have been vertically clipped to aid clarity; (full assignments are given in ESI Tables S3 and S6). The more intense ^1Bu signals of the major isomer are marked with vertical bars. The labels are given in Figure 2 and Fig. S5.

Simultaneous assignment and fitting of the proton and phosphorus paramagnetic shifts to the susceptibility tensor was carried out using the DFT optimised structures of (*RRR*)-*A*-($\lambda\lambda\lambda$) conformers (ESI: Table S8) and an automated combinatorial fitting procedure developed and implemented in *Spinach 1.9*. Longitudinal relaxation rates and the linewidths have also been taken into account during the assignment process. The final assignments (Table S3, Figure S18) are in agreement with the observed COSY cross-peaks. Notably, the cyclen protons of the axial positions ('c1ax', 'c3ax', 'c5ax' and 'c7ax') form a well separated group in the case of [Yb.L¹], but they are more separated in the case of [Yb.L²]; a similar effect is observed for the 'axial' protons in the exocyclic CH₂ groups.

The 'best-fit' traceless susceptibility tensors, (ESI: Tables S4 and S7: Figures S22 and S18), can be converted to axially and rhombicity parameters and three Euler angles, with respect to the molecular frame. The z-axis of molecular frame points towards the centre of four nitrogen atoms of cyclen and the x-axis points towards the N1 atom. Comparison of the CASSCF computed susceptibility anisotropy and the PCS fitted values are shown below (Table 8).

Table 8. Anisotropy of the susceptibility tensors of [Yb.L¹] and [Yb.L²], extracted from paramagnetic shift fitting and computed with CASSCF. Axiality is shown in the Å³ in SI units and Euler angles in zy'z'' convention; the maximum value of rhombicity/axiality is defined as 1/3 (see computational details and ESI).

	axiality, Å ³	rhombicity/axiality	α °	β °	γ °
[Yb.L ¹] NMR	0.127	0.163	58	4	54
[Yb.L ¹] CASSCF	0.268	0.135	26	86	167
[Yb.L ²] NMR	0.113	0.140	185	23	300
[Yb.L ²] CASSCF	0.139	0.114	204	29	111

Computed axiality is larger than that extracted from PCS by about a factor of two for [Yb.L¹] and by 20% for [Yb.L²]. In each case, it can be ascribed to systematic overestimation of the axiality, owing to the lack of dynamical correlation in CASSCF. The modulation of the ligand field by the surrounding water molecule is also not taken

into account. To tackle the latter effect, we have also computed susceptibility for each complex with one explicit water molecule; indeed, for $[\text{Yb.L}^1]$ it led to a reduction of axially by about 20% (Table S5). The reconstructed PCS fields based on the best-fit susceptibility tensors are presented at the Figure 8. Since the amplitude of the anisotropy is almost the same for both compounds the radial part of the PCS field is very similar. The intermediate ratio of rhombicity/axiality manifests itself in the flattening of the otherwise axially symmetric blue region of the PCS field (Figure 8). The most important difference between $[\text{Yb.L}^1]$ and $[\text{Yb.L}^2]$ is the orientation of the PCS field that is mainly defined by the β angle of the susceptibility tensor (Table 8).

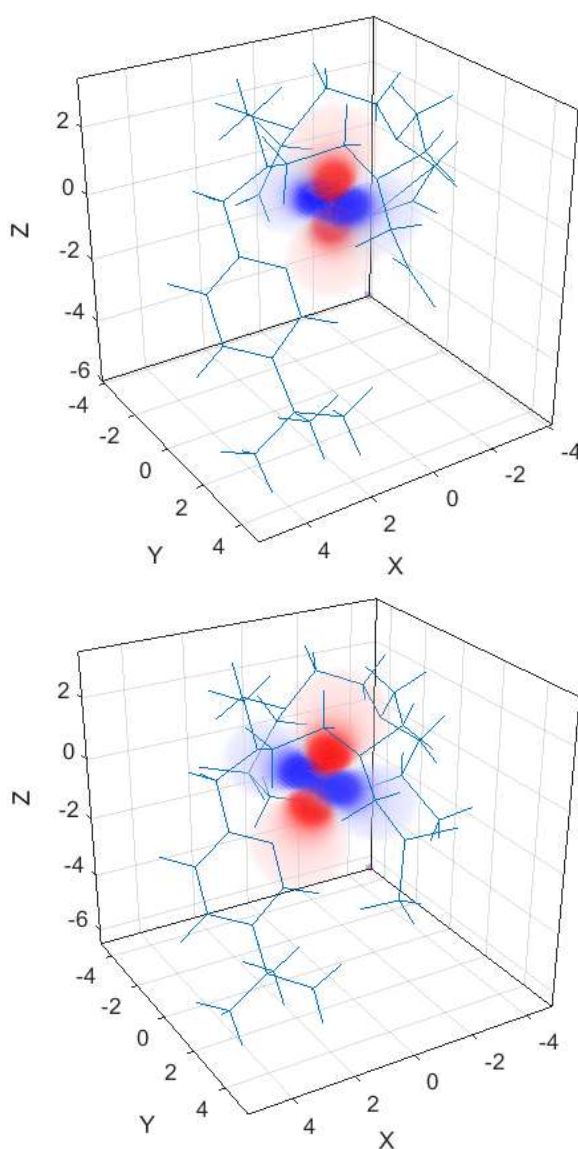


Figure 8. Pseudocontact shift (PCS) field reconstructed using the best fit susceptibility tensor for $[\text{Yb.L}^1]$ (top) and $[\text{Yb.L}^2]$ (bottom) created in *Spinach*.

The difference in the orientation of the PCS field is also responsible for the interchange in the order of the paramagnetic shifts observed by ^1H NMR (Figure 6).

SUMMARY AND CONCLUSIONS

The lanthanide(III) complexes of L^2 are prototypical examples of PARASHIFT NMR contrast agents that are being developed for use in magnetic resonance spectral imaging *in vivo*. Understanding their structure and solution dynamics is an important part of appreciating why they behave so well for this role.¹⁹⁻²²

The twisted square anti-prismatic (TSAP) lanthanide complexes of L^2 exist as two isomers differing in free energy in solution by only a few kJ/mol and they undergo exchange by slow cooperative arm rotation. These structures, specifying a common chirality at P, are the $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda\lambda)$ and $(RRR)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ diastereoisomers, i.e. TSAP isomers. In the solid-state structure, only the lower energy $(RRR)\text{-}\Delta\text{-}(\lambda\lambda\lambda\lambda)$ and $(SSS)\text{-}\Delta\text{-}(\delta\delta\delta\delta)$ enantiomers were found, as a krypto-racemate. Every octadentate, tri- or tetra-phosphinate lanthanide complex, of the second half of the lanthanide series, reported to date has been found to be 8-coordinate (Y(III) has the same ionic radius as Er(III)) and lacks a short bond to water that would make it formally 9-coordinate.²⁴

In the Yb complex of L^1 the crystal structure reveals an 8-coordinate TSAP coordination geometry. In solution, two species are observed undergoing chemical exchange. The computed pseudocontact shift field fitted best to a CASSCF calculation by introducing an axially coordinated water molecule. Such a 9-coordinate structure is a SAP isomer, exchanging with the TSAP structure in solution by fast dissociative water exchange, (rates typically $>10^6\text{ s}^{-1}$),²⁴ and a ring inversion process with a rate of the order of 50 s^{-1} . For the $[\text{Y.L}^1]$ complex, evidence for dynamic exchange was found from EXSY NMR studies, but only one solution species was observed, suggesting that in this case, the solution species are 9-coordinate enantiomeric SAP structures, i.e. $\Delta\text{-}(\lambda\lambda\lambda\lambda)$ and $\Delta\text{-}(\delta\delta\delta\delta)$, that interconvert by sequential ring inversion and arm rotation.

The pseudocontact shifts of the ^1Bu reporter resonance in $[\text{Ln.L}^2]$ are 50 or 60 ppm greater than the tricarboxylate analogues, based on L^1 , for Dy, Tb and Tm complexes. In terms of the distance between the paramagnetic centre and the average position of the ^1Bu hydrogen nuclei, the X-ray studies reveal that in $[\text{Yb.L}^1]$ the distances range

from 5.28 to 7.95 Å ($10^3/\langle r^3 \rangle = 3.76 \text{ \AA}^{-3}$), whereas for [Yb.L²], corresponding values are 5.46 to 8.05 Å ($10^3/\langle r^3 \rangle = 3.82 \text{ \AA}^{-3}$). Evidently, the greater ¹Bu shift found in [Ln.L²] compared to [Ln.L¹] is not simply due to a shorter distance. Detailed analysis of all PCS shifts shows that it is the difference in the orientation of the magnetic susceptibility tensor for the two complexes, rather than its anisotropy that accounts very well for their markedly different NMR spectral behaviour.¹⁹

EXPERIMENTAL SECTION

X-ray crystallography

The X-ray single crystal data for $[\text{Yb.L}^1] \cdot 2\text{MeOH} \cdot \text{H}_2\text{O}$ and $[\text{Yb.L}^2] \cdot \text{EtOH} \cdot 2\text{H}_2\text{O}$ were collected using Mo- $K\alpha$ radiation ($\lambda = 0.71073 \text{ \AA}$) on a Bruker D8 Venture (Photon100 CMOS detector, μS -microsource, focusing mirrors) diffractometer equipped with a Cryostream (Oxford Cryosystems) open-flow N_2 cryostat. Both structures were solved by direct method and refined by full-matrix least squares on F^2 for all data using Olex2³⁵ and SHELXL³⁶ software. All non-hydrogen atoms were refined anisotropically, hydrogen atoms were placed in the calculated positions and refined in riding mode. $[\text{Yb.L}^1]$ was a 2-component non-merohedral twin with component ratio 0.930:0.070(1), $[\text{Yb.L}^2]$ as an inversion twin with component ratio 0.489:0.511(7).

Crystal data: $\text{C}_{24}\text{H}_{36}\text{N}_5\text{O}_6\text{Yb} \cdot 2\text{CH}_3\text{OH} \cdot \text{H}_2\text{O}$, $M = 745.72$, monoclinic, space group $P2_1/c$ (no. 14), $a = 13.4134(7)$, $b = 16.7937(9)$, $c = 14.9527(8) \text{ \AA}$, $\beta = 119.464(2)^\circ$, $V = 2932.6(3) \text{ \AA}^3$, $Z = 4$, $D_{\text{calc}} = 1.689 \text{ g cm}^{-3}$, $\mu = 3.25 \text{ mm}^{-1}$, $T = 120 \text{ K}$, 69043 reflections with $\theta \leq 30.5^\circ$ ($R_{\text{merg}} = 0.082$), 381 refined parameters, final $wR_2(F^2) = 0.063$ (on 7804 unique data), conventional $R_1(F) = 0.035$ on 4783 data with $I \geq 2\sigma$, CCDC-1502162.

$\text{C}_{24}\text{H}_{45}\text{N}_5\text{O}_6\text{P}_3\text{Yb} \cdot \text{C}_2\text{H}_6\text{O} \cdot 2\text{H}_2\text{O}$, $M = 847.70$, monoclinic, space group $P2_1$ (no. 4), $a = 9.5975(4)$, $b = 13.8670(6)$, $c = 26.4618(11) \text{ \AA}$, $\beta = 99.837(2)^\circ$, $V = 3470.0(3) \text{ \AA}^3$, $Z = 4$, $D_{\text{calc}} = 1.623 \text{ g cm}^{-3}$, $\mu = 2.89 \text{ mm}^{-1}$, $T = 120 \text{ K}$, 94524 reflections with $\theta \leq 34.9^\circ$ ($R_{\text{merg}} = 0.032$), 820 refined parameters, final $wR_2(F^2) = 0.060$ (on 28857 unique data), conventional $R_1(F) = 0.032$ on 25209 data with $I \geq 2\sigma$, CCDC-1501473.

NMR and mass spectroscopy

^1H , ^{13}C and ^{31}P NMR spectra were recorded in commercially available deuterated solvents on a Bruker Avance-400 (^1H at 400.06 MHz, ^{13}C at 100.61 MHz and ^{31}P at 161.95 MHz), a Mercury 400 (^1H at 399.95 MHz), a Varian VNMRS-600 (^1H at 599.67 MHz, ^{13}C at 150.79 MHz and ^{31}P at 242.75 MHz), or a Varian VNMRS-700 (^1H at 699.73 MHz, ^{13}C at 175.95 MHz and ^{31}P at 283.26 MHz). All chemical shifts are given in ppm, with coupling constants are in Hz.

Electrospray mass spectra were obtained on a TQD mass spectrometer equipped with an Acquity UPLC system, an electrospray ion source and an Acquity photodiode array detector (Waters Ltd., UK). Accurate masses were recorded on an LCT Premier XE mass spectrometer or a QToF Premier Mass spectrometer, both equipped with an Acquity UPLC, a lock-mass electrospray ion source and an Acquity photodiode array detector (Waters Ltd., UK). Methanol was used as the carrier solvent.

Electronic structure theory

Initial geometry optimization was done using BP86³⁷ exchange-correlation functional followed by re-optimization and thermochemical analysis with M06-2X.³⁸ Solvent effects were included using the SMD model (water).³⁹ All calculations used cc-pVDZ⁴⁰ basis set with Stuttgart ECP⁴¹ on the lanthanide. D3 dispersion correction was used to account for weak interactions.⁴² The resolution of identity approximation with the corresponding auxiliary basis sets was used to speed up the calculations.⁴³ DFT calculations were carried out with the Gaussian09.⁴⁴

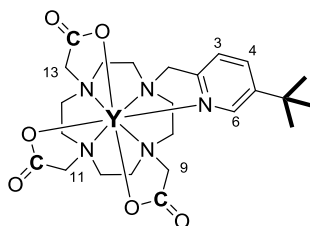
Magnetic susceptibility tensors were computed in MOLCAS 8.0⁴⁵ using CASSCF method⁴⁶ and relativistic ANO-RCC-VDZP basis set⁴⁷. Spin-orbit coupling of 7 spin-free states of $4f^{13}$ configuration has been taken into account in RASSI module⁴⁸ followed by SINGLE_ANISO calculation of the magnetic susceptibility tensors.

Fitting of the paramagnetic shifts to the magnetic susceptibility tensors was carried out in *Spinach* 1.9 package⁴⁹. Fermi contact terms were assumed to be negligible for all protons and phosphorous atoms. Point model for pseudocontact shift (PCS) has been used for all simulations⁵⁰. Eigenvalues of the fitted susceptibility tensors were ordered to satisfy $|\chi_x| < |\chi_y| < |\chi_z|$, then axiality was computed as $3/2\chi_z$ and rhombicity as $(\chi_x - \chi_y)/2$. With this definition, the ratio between the rhombicity and the axiality falls between 0 and 1/3, the former yielding a d_{z^2} orbital like spatial profile and the latter yielding a d_{yz} orbital like spatial profile. ZYZ convention was used for the Euler angles.

Characterisation

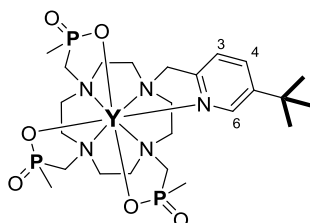
The synthesis of the ligands L^{1-3} and the method of preparation of their lanthanide(III) and yttrium complexes has been reported earlier.^{19, 22}

[Y.L¹] (see ESI Table S3 for detail)



¹H NMR (600 MHz, D₂O, pD 6) δ (ppm) 8.47 (s, H^{6py}), 7.93 (d, ³J_{HH} = 8.0 Hz, H^{4py}), 7.31 (d, ³J_{HH} = 8.0 Hz, H^{3py}), 4.26 (d, ²J_{HH} = -15.0 Hz, pyCH₂N), 3.84 (d, ²J_{HH} = -15.0 Hz, pyCH₂N), 3.60-3.50 (m, NCH₂NCOO⁻), 3.45-3.25 (m, cyclen-CH₂), 3.22-3.11 (m, NCH₂NCOO⁻), 2.96 (d, ²J_{HH} = -17.5 Hz, NCH₂C⁹OO⁻) 2.87-2.78 (m, cyclen-CH₂), 2.76-2.51 (m, cyclen-CH₂), 2.47-2.31 (m, NCH₂C⁹OO⁻/cyclen-CH₂), 2.24-2.17 (m, cyclen-CH₂), 1.63-1.55 (m, cyclen-CH₂), 1.19 (s, C(CH₃)₃). ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 146.4 (C^{6py}), 137.7 (C^{4py}), 121.9 (C^{3py}), 66.2 (pyCH₂N), 66.0 (C⁹), 65.8 (C¹¹ and C¹³), 56.8-54.8 (cyclen-CH₂), 33.1 (C(CH₃)₃). ESI-LRMS (+) *m/z* 580.2 [M+H]⁺; ESI-HRMS (+) *m/z* calcd. C₂₄H₃₇N₅O₆Y 580.1802, found 580.1815.

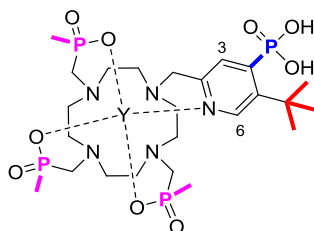
[Y.L²] (ESI Table S6, for detail)



¹H NMR (600 MHz, D₂O) δ (ppm) 8.92 (s, H^{6py} minor), 8.60 (s, H^{6py} major), 7.97 (d, ³J_{HH} = 8.0 Hz, H^{4py} major), 7.91 (d, ³J_{HH} = 8.0 Hz, H^{4py} minor), 7.35 (d, ³J_{HH} = 8.0 Hz, H^{3py} major), 7.30 (d, ³J_{HH} = 8.0 Hz, H^{3py} minor), 4.55 (d, ²J_{HH} = -16.0 Hz, pyCH₂N), 3.85-3.77 (m, cyclen-CH₂), 3.64 (d, ²J_{HH} = -16.0 Hz, pyCH₂N), 3.61-3.32 (m, NCH₂P/cyclen-CH₂), 3.01-2.94 (m, cyclen-CH₂), 2.80-2.74 (m, NCH₂P), 2.66-2.35 (m, NCH₂P/cyclen-CH₂), 2.34-2.26 (m, cyclen-CH₂), 1.53 (d, ²J_{PH} = 13.0 Hz, PCH₃ minor), 1.46-1.42 (m, PCH₃), 1.29-1.22 (m, PCH₃/C(CH₃)₃), 1.24 (C(CH₃)₃). ¹³C NMR (150 MHz, CDCl₃) δ (ppm) 148.0 (C^{6py} minor), 145.2 (C^{6py} major), 138.4 (C^{4py} major), 137.8 (C^{4py} minor), 123.6 (C^{3py} major), 123.0 (C^{3py} minor), 59.3 (pyCH₂N), 55.5/56.3/56.0 (3×d, ²J_{PC} = 95 Hz PCH₂N major),

53.9/53.4/53.2/53.6/51.7/51.6/51.5/50.1 (cyclen- $\underline{\text{C}}\text{H}_2$ major), 18.7 (d, $^2J_{\text{PC}} = 95$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 16.3/15.7/15.6 (3×d, $^2J_{\text{PC}} = 95$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 33.3 ($\underline{\text{C}}(\text{CH}_3)_3$), 29.9 ($\text{C}(\underline{\text{C}}\text{H}_3)_3$ major). $^{31}\text{P}\{^1\text{H}\}$ NMR (243 MHz, D_2O ,) 44.0 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 43.9 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 43.3 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 43.0 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 41.0 ($^2J_{\text{YP}} = 6$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 40.2 ($^2J_{\text{YP}} = 6$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor). ESI-LRMS (+) m/z 682.2 $[\text{M}+\text{H}]^+$; ESI-HRMS (+) m/z calcd. for $\text{C}_{24}\text{H}_{45}\text{N}_5\text{O}_6\text{P}_3\text{Y}$ 682.1720, found 682.1714.

$[\text{H}_2\text{Y}\cdot\text{L}^3]$



^1H NMR (600 MHz, D_2O) δ (ppm) 8.96 (d, $^4J_{\text{HP}} = 5.5$ Hz, $\text{H}^{6\text{py}}$ minor), 8.64 (d, $^4J_{\text{HP}} = 5.5$ Hz, $\text{H}^{6\text{py}}$ major), 7.93 (d, $^3J_{\text{HP}} = 13.5$ Hz, $\text{H}^{3\text{py}}$ major), 7.88 (d, $^3J_{\text{HP}} = 13.5$ Hz, $\text{H}^{3\text{py}}$ minor), 4.55 (d, $^2J_{\text{HH}} = -15.5$ Hz, $\text{py}\underline{\text{C}}\text{H}_2\text{N}$ major), 3.80 (m, cyclen- $\underline{\text{C}}\text{H}_2$), 3.66 (d, $^2J_{\text{HH}} = -15.5$ Hz, $\text{py}\underline{\text{C}}\text{H}_2\text{N}$ major), 3.63-3.33 (m, cyclen- $\underline{\text{C}}\text{H}_2/\text{N}\underline{\text{C}}\text{H}_2\text{P}$), 3.22-3.14 (m, cyclen- $\underline{\text{C}}\text{H}_2$), 3.04-2.94 (m, cyclen- $\underline{\text{C}}\text{H}_2$), 2.82-2.75 (m, $\text{N}\underline{\text{C}}\text{H}_2\text{P}$), 2.66-2.24 (m, $\text{N}\underline{\text{C}}\text{H}_2\text{P}/\text{cyclen-}\underline{\text{C}}\text{H}_2$), 2.34-2.26 (m, cyclen- $\underline{\text{C}}\text{H}_2$), 1.53 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 1.45 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 1.28 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 1.26 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 1.23 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 1.19 (d, $^2J_{\text{PH}} = 14.0$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 1.46 (s, $\text{C}(\underline{\text{C}}\text{H}_3)_3$). ^{13}C NMR (150 MHz, CDCl_3 ,) δ (ppm) 146.7 ($\text{C}^{6\text{py}}$ major), 129.0 ($\text{C}^{3\text{py}}$ major), 59.6 ($\text{py}\underline{\text{C}}\text{H}_2\text{N}$ major), 55.6/56.3/56.0 (3×d, $^2J_{\text{PC}} = 95$ Hz $\text{P}\underline{\text{C}}\text{H}_2\text{N}$ major), 53.9/53.5/53.2/52.6/51.8/51.7/51.6/50.1 (cyclen- $\underline{\text{C}}\text{H}_2$ major), 35.3 ($\underline{\text{C}}(\text{CH}_3)_3$), 30.7 ($\text{C}(\underline{\text{C}}\text{H}_3)_3$ major), 13.0/16.4/15.7 (3×d, $^2J_{\text{PC}} = 95$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major). $^{31}\text{P}\{^1\text{H}\}$ NMR (243 MHz, D_2O ,) 44.1 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 44.0 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 43.4 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 43.0 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 41.0 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ major), 39.7 ($^2J_{\text{YP}} = 5$ Hz, $\text{P}\underline{\text{C}}\text{H}_3$ minor), 7.9 (P(O)OOH minor), 7.5 (P(O)OOH minor); ESI-LRMS (+) m/z 762.6 $[\text{M}+\text{H}]^+$; ESI-HRMS (+) m/z calcd. for $\text{C}_{24}\text{H}_{46}\text{N}_5\text{O}_9\text{P}_4\text{Y}$ 762.1383, found 762.1388.

ASSOCIATED CONTENT

Supporting Information.

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.inorg-chem.xxxxxx

Crystallographic data (CIF) . The data have been deposited at the Cambridge Crystallographic Data Centre as supplementary publication CCDC-1502162 and CCDC-1501473.

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Notes

The authors declare no competing financial interest.

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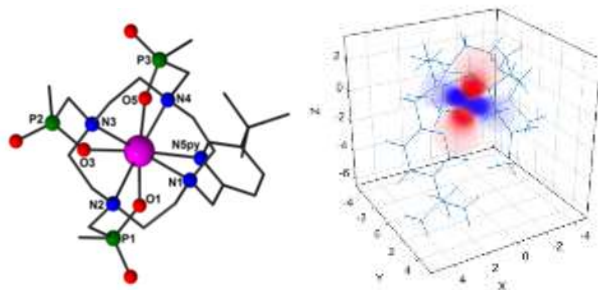
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Eight-coordinate ytterbium complexes with a bound pyridine group adopt a twisted square antiprismatic coordination geometry; the orientation of the pseudocontact shift field is mainly defined by the β angle of the magnetic susceptibility tensor.