

1 **Title: Parental care influences social immunity in burying beetle larvae**

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12

13 **Abstract**

14 1. We provide evidence for social immunity in offspring of a sub-social species, the burying
15 beetle, *Nicrophorus vespilloides*.

16 2. *N. vespilloides* is a carrion breeder, and in a similar fashion to the adult beetles, the
17 offspring produce exudates that exhibit lytic activity, which are used to coat the breeding
18 resource. This strategy defends against the microbial community.

19 3. The lytic activity in larval exudates declines as the brood develops, perhaps being most
20 beneficial at the start of the breeding bout.

21 4. Changing levels of parental care through widowing/orphaning affects lytic activity in the
22 larval exudates, with levels decreasing in the absence of both parents.

23 **Introduction**

24 The burying beetle, *N. vespilloides* (Figure S1, Supplementary Material), breeds on
25 the carcass of small mammals and exhibits bi-parental care, one component of which is a
26 social immune response (Cotter & Kilner 2010b). Cotter & Kilner suggest that “any type of
27 immune response that has been selected to increase the fitness of the challenged individual
28 and one or more recipients should be classified as social immunity” (2010b). The beetles
29 coat the carcass with antimicrobial anal exudates (Cotter & Kilner 2010a) in order to
30 minimise competition from the microbial community, delaying decomposition of the carcass
31 (Rozen et al. 2008). This social immune response is costly; antibacterial activity is only
32 upregulated in the presence of a carcass (Cotter & Kilner 2010a) and forced upregulation
33 reduces lifetime reproductive success (Cotter et al. 2010). In *N. vespilloides*, both sexes
34 produce exudates, however, the females’ exudates show higher levels of antibacterial activity

35 than the males (Cotter & Kilner 2010a). The beetles can also flexibly adjust the level of
36 antibacterial activity in response to mate loss.

37 Observations have shown that the larvae also produce exudates throughout their
38 development, though their potential antibacterial activity is unknown. With production of
39 these exudates being so costly to the parents, it would benefit them if their offspring were
40 also able to partake in carcass preservation. Furthermore, we know that larvae of this species
41 can survive without parental care, albeit at a reduced rate and with a resulting poorer quality
42 (Eggert et al. 1998). Their ability to survive the loss of both parents may be due in part to the
43 production of antibacterial substances. Here we ask whether 1) antibacterial activity is
44 present, 2) how it changes during larval development and 3) whether larvae, like adults, can
45 flexibly alter levels of antibacterial activity in response to changing conditions, for example,
46 presence/absence of parents.

47 **Materials and Methods**

48 ***N. vespilloides* colony**

49 The colony was established from a pedigreed *N. vespilloides* colony at the
50 Department of Zoology, University of Cambridge. Beetles were maintained as described
51 previously (Cotter & Kilner 2010a).

52 **Experiment 1 – characterising antibacterial activity across the larval stage**

53 Exudates were collected from all larvae in a brood using a capillary tube and pooled
54 in a single eppendorf tube. This enabled us to use a known volume of exudate in the later
55 analysis (1 μ l). Larvae were sampled from day 1 (hatching) to day 5 (dispersal). Day 1 and 2
56 larvae were so small that exudate collection with a capillary tube was not possible. Instead,
57 the tip of the abdomen was gently pressed against a single punched circle of filter paper for

58 every larva in the brood. Whilst volume was impossible to control for in these samples, we
59 collected <1 µl of exudate sample on each filter paper circle, and so the level of lytic activity
60 would be a slight under-estimate compared to day 3, 4 and 5 larvae. All samples were stored
61 at -20°C until testing. Larvae were sampled from fifty-five families, though some were
62 sampled on more than one day, giving 75 samples in total. Forty-two families were sampled
63 from once, nine families were sampled from twice and five families were sampled from three
64 times. The numbers of larvae from which samples were collected and pooled on each day
65 were as follows: day 1 = 10.33 larvae +/-2.01, day 2 = 14.63 larvae +/- 1.49, day 3 = 9.65
66 larvae +/-1.24, day 4 = 14.27 larvae +/-1.88 and day 5 = 7.17 larvae +/- 2.29.

67 **Experiment 2 – the effect of reduced parental care on larval antibacterial activity**

68 Sixty pairs were established and were assigned to one of four treatment groups, male
69 removed, female removed, both removed, or neither removed (control). On the day of
70 hatching, parents were removed according to the treatment and the larval exudate sampled
71 daily as described above. Exudate collection was more successful for this experiment, sixty
72 families were sampled daily for five days giving 300 samples in total. Breeding success was
73 recorded for all families.

74 **Antibacterial activity**

75 Lysozyme-like antibacterial activity was measured as described previously (Cotter &
76 Kilner 2010a). In brief, 1 µl of each sample was pipetted into a hole in an agar plate
77 inoculated with freeze-dried *Micrococcus lysodeikticus* cells and incubated for 24 hours at
78 33°C. We selected *Micrococcus lysodeikticus* as it is a soil bacterium, which is the breeding
79 environment of the burying beetle. It is one of the microorganisms analysed in Hall et al.
80 (2011) and in that study was inhibited by secretions from *Nicrophorus*. It is also the main
81 bacteria used for lytic plates in studies on burying beetles (Cotter et al. 2010, Steiger et al.
82 2011, Arce et al. 2012) and many other insect species e.g. locusts (Wilson et al. 2002). Filter

83 paper circles were placed directly onto the surface of the agar. Clear zones in the agar were
84 measured with digital calipers and calibrated against a lysozyme standard (Figure S2,
85 Supplementary Material). In order to test for the potential mechanical differences between
86 using filter paper and punched holes, we compared samples tested using both methods and
87 found no difference between the size of the clear zone produced by 1 μ l of sample pipetted
88 into a hole and that produced using filter paper placed on the surface of the agar ($F_{1,25} = 0.27$,
89 $P = 0.61$). We were therefore able to reliably compare samples measured using the different
90 techniques. Whilst other methods are now available to measure antimicrobial activity, the
91 majority of analyses to date in this and many other insect species have been carried out using
92 zones of inhibition on agar plates. Using this method allows us to compare across studies to
93 some extent.

94 **Statistical analyses**

95 Antibacterial activity was log-transformed to approximate normality. Lytic data from
96 both experiments was analysed with Restricted Estimate Maximum Likelihood (REML)
97 models in Genstat 15 (VSN International, Hemel Hempstead, UK) with family included as a
98 random effect to account for multiple testing from each family. Breeding data were analysed
99 using a generalised linear model (GLM). Carcass weight and interaction terms were included
100 in all models but were removed due to non-significance. Figures were produced in R 2.15.1
101 (Development Core Team, 2013).

102 **Results and Discussion**

103 We show for the first time in this study that the larval exudates contain antibacterial
104 substances. It has been shown previously that insect larvae have the ability to produce
105 antibacterial secretions e.g. blowfly larvae, *L. sericata* (Kerridge et al. 2005), however this
106 strategy has not been widely documented. Levels of lytic activity were highest in the newly-

107 hatched larvae and declined throughout the brood (REML: $F_{1,49} = 51.27$, $P < 0.001$; Figure 1).
108 Indeed, as the exudate volume collected from day 1 and day 2 larvae was $< 1 \mu\text{l}$, the lytic
109 activity at this stage is actually slightly underestimated, and so in reality should be even
110 higher than observed. If the trend had been in the other direction i.e. larval exudate activity
111 increasing with age, we could not have drawn reliable conclusions as in this case a lower
112 activity in day 1 and day 2 larvae could have been due to a smaller exudate volume. As we
113 did not collect a fixed amount of exudate from day 1 and 2 larvae (although volume $< 1 \mu\text{l}$),
114 we tested whether activity was dependent on the number of larvae from which it was
115 collected, however there was no effect (REML: $F_{1,27} = 0.51$, $P = 0.48$). The pattern of decline
116 in larval lytic activity mirrors that of the parents; highest when larvae arrive on the carcass
117 (Cotter et al. 2013). The need for lytic exudates may fall after a high amount of initial
118 preservation, resulting in increasing sterility. For the parents, assistance from the larvae may
119 mean that they do not need to invest as heavily in lytic activity, which could promote further
120 reproductive success and longevity.

121 With regards parental removal experiments, preliminary data exploration showed that
122 the effects of removing males and females were very similar but different from the other two
123 treatment groups. Therefore we grouped these two treatments into a single treatment
124 representing “one parent removed”. Lytic activity was much lower in larvae where both
125 parents were removed (REML: $F_{2,46} = 3.66$, $P = 0.033$; Figure 2a) and there was a significant
126 effect of brood age, with the pattern being very similar to experiment 1 (REML: $F_{1,210} =$
127 119.95 , $P < 0.001$). These results illustrate that, like the parents (Cotter & Kilner 2010a), the
128 larvae exhibit plasticity with regards lytic activity levels. Whilst we hypothesised that this
129 antibacterial activity may contribute to their survival in orphaned conditions, activity actually
130 decreased. This drop may be due to the fact that the larvae must now invest in other
131 activities, for example self-feeding, at the detriment to lytic investment. Their condition is

132 also likely to be compromised even at this stage, which may cause a decline in lytic activity.
133 A different experimental set-up may be required to unveil an upregulation of lytic activity in
134 the larvae, for example using a rotten carcass but maintaining parental assistance i.e. a greater
135 requirement for combatting microorganisms but without the strain of self-feeding. The
136 number of larvae produced was lower when both parents were removed post-hatching (GLM:
137 $F_{2,51} = 4.30$, $P = 0.019$; Figure 2b), but the mean weight of larvae was not affected (GLM:
138 $F_{2,49} = 2.07$, $P = 0.137$). This is consistent with findings from previous studies (Rozen et al.
139 2008; Arce et al. 2012).

140 Future experiments should consider the cost of lytic activity from the larval
141 perspective. The larvae may be constrained developmentally if the mechanism for
142 production in the parents also drives production in the larval stage. In light of plasticity from
143 parents and their offspring, future studies should consider the scope for conflict. For
144 example, if the larvae didn't produce antibacterial exudates, would the parents be forced to
145 produce more? To conclude, we show that larvae produce antibacterial exudates and that
146 parental care influences the extent of social immunity.

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150 analysed the data, and co-wrote the paper; LB co-designed the experiments and collected the
151 data; CER co-designed the experiments, collected the data and co-wrote the paper.

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183 **Figure legends**

184 **Figure 1**

185 The decline in the antibacterial activity of larval exudate over time. Larvae hatch on
186 day 1 and disperse from the carcass on day 5.

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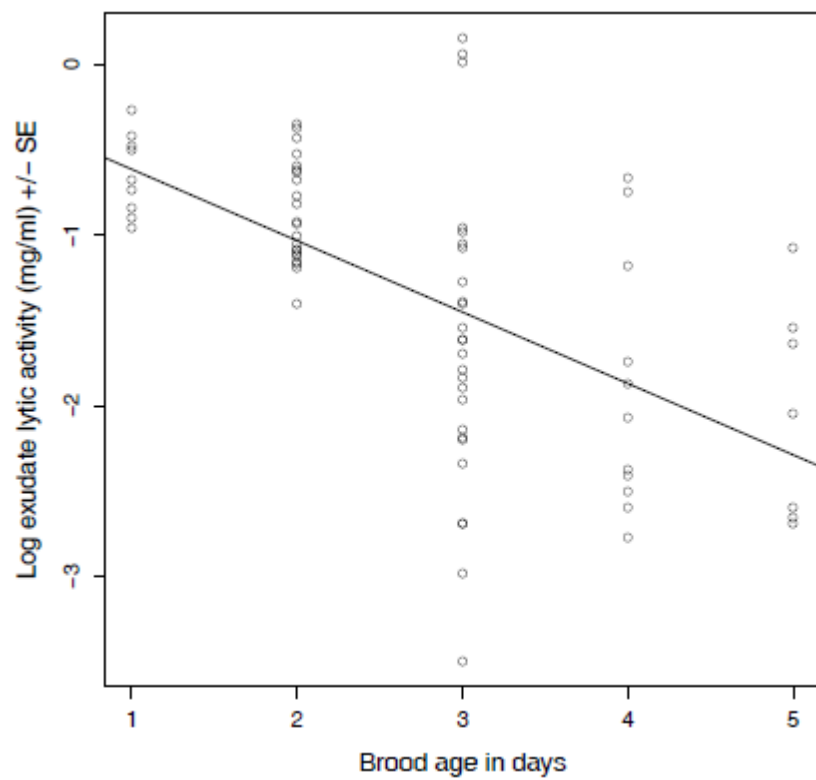
188 **Figure 2**

189 The effect of removing one or both parents on a) the antibacterial activity of larval
190 exudate and b) the number of larvae dispersing from the carcass. Means and SEs in a) are
191 predicted values from a REML model controlling for family.

192 **Figures**

193 **Figure 1**

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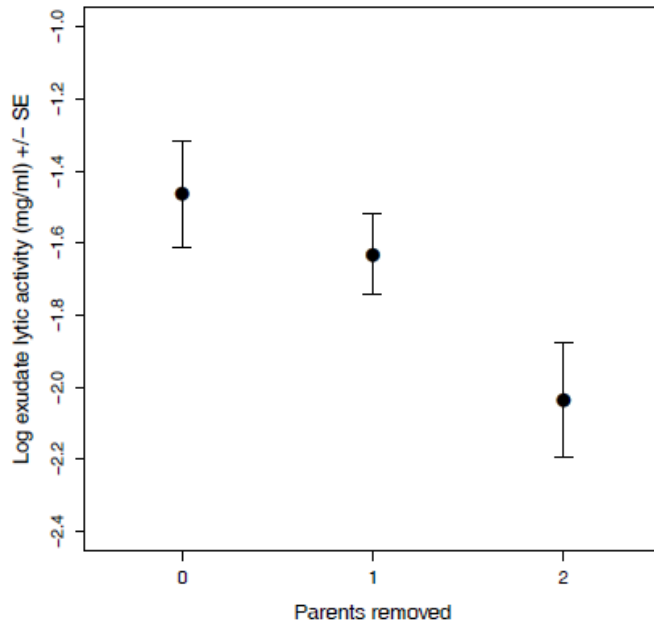
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197 **Figure 2**

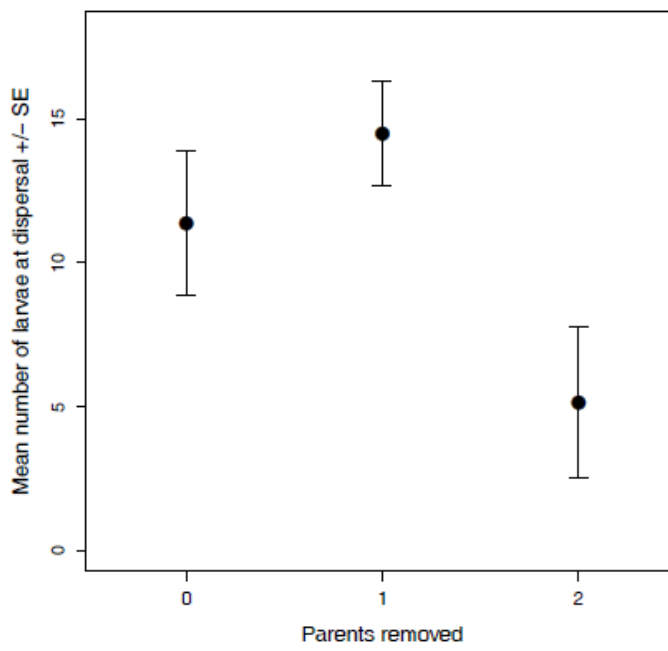
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199 a)



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201 b)



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