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PCA-based bootstrap confidence interval tests for gene-disease association involving multiple SNPs

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Abstract

Background: Genetic association study is currently the primary vehicle for identification and characterization of disease-predisposing variant(s) which usually involves multiple single-nucleotide polymorphisms (SNPs) available. However, SNP-wise association tests raise concerns over multiple testing. Haplotype-based methods have the advantage of being able to account for correlations between neighbouring SNPs, yet assuming Hardy-Weinberg equilibrium (*HWE*) and potentially large number degrees of freedom can harm its statistical power and robustness. Approaches based on principal component analysis (*PCA*) are preferable in this regard but their performance varies with methods of extracting principal components (*PCs*).

Results: *PCA*-based bootstrap confidence interval test (*PCA-BCIT*), which directly uses the *PC* scores to assess gene-disease association, was developed and evaluated for three ways of extracting *PCs*, i.e., cases only(*CAES*), controls only(*COES*) and cases and controls combined(*CES*). Extraction of *PCs* with *COES* is preferred to that with *CAES* and *CES*. Performance of the test was examined via simulations as well as analyses on data of rheumatoid arthritis and heroin addiction, which maintains nominal level under null hypothesis and showed comparable performance with permutation test.

Conclusions: *PCA-BCIT* is a valid and powerful method for assessing gene-disease association involving multiple SNPs.

Background

Genetic association studies now customarily involve multiple SNPs in candidate genes or genomic regions and have a significant role in identifying and characterizing disease-predisposing variant(s). A critical challenge in their statistical analysis is how to make optimal use of all available information. Population-based case-control studies have been very popular[1] and typically involve contingency table tests of SNP-disease association[2]. Notably, the genotype-wise Armitage trend test does not require *HWE* and has equivalent power to its allele-wise counterpart under *HWE*[3,4]. A thorny issue with individual tests of SNPs for linkage disequilibrium (*LD*) in such setting is multiple testing, however, methods for multiple testing adjustment assuming independence such as Bonferroni's[5,6] is knowingly

conservative[7]. It is therefore necessary to seek alternative approaches which can utilize multiple SNPs simultaneously. The genotype-wise Armitage trend test is appealing since it is equivalent to the score test from logistic regression[8] of case-control status on dosage of disease-predisposing alleles of SNP. However, testing for the effects of multiple SNPs simultaneously via logistic regression is no cure for difficulty with multicollinearity and curse of dimensionality[9]. Haplotype-based methods have many desirable properties[10] and could possibly alleviate the problem[11-14], but assumption of *HWE* is usually required and a potentially large number of degrees of freedom are involved[7,11,15-18].

It has recently been proposed that *PCA* can be combined with logistic regression test (*LRT*)[7,16,17] in a unified framework so that *PCA* is conducted first to account for between-SNP correlations in a candidate region, then *LRT* is applied as a formal test for the association between *PC* scores (linear combinations of the original SNPs) and disease. Since *PCs* are orthogonal, it avoids multicollinearity and at the meantime is

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less computer-intensive than haplotype-based methods. Studies have shown that *PCA-LRT* is at least as powerful as genotype- and haplotype-based methods[7,16,17]. Nevertheless, the power of *PCA*-based approaches vary with ways by which *PCs* are extracted, e.g., from genotype correlation, LD, or other kinds of metrics[17], and in principle can be employed in frameworks other than logistic regression[7,16,17]. Here we investigate ways of extracting *PCs* using genotype correlation matrix from different types of samples in a case-control study, while presenting a new approach testing for gene-disease association by direct use of *PC* scores in a *PCA*-based bootstrap confidence interval test (*PCA-BCIT*). We evaluated its performance via simulations and compared it with *PCA-LRT* and permutation test using real data.

Methods

PCA

Assume that p SNPs in a candidate region of interest have coded values (X_1, X_2, \dots, X_p) according to a given genetic model (e.g., additive model) whose correlation matrix is C . *PCA* solves the following equation,

$$Cl_i - \lambda l_i = 0 \quad (1)$$

where $l_i^T l_i = 1, i = 1, 2, \dots, p, l_i = (l_{i1}, l_{i2}, \dots, l_{ip})^T$ are loadings of *PCs*. The score for an individual subject is

$$F_i = l_{i1}X_1 + l_{i2}X_2 + \dots + l_{ip}X_p, \quad i = 1, 2, \dots, p, \quad (2)$$

where $\text{cov}(F_i, F_j) = 0, i \neq j$, and $\text{var}(F_1) \geq \text{var}(F_2) \geq \dots \geq \text{var}(F_p)$.

Methods of extracting *PCs*

Potentially, *PCA* can be conducted via four distinct extracting strategies (*ES*) using case-control data, i.e., 0. Calculate *PC* scores of individuals in cases and controls separately (*SES*), 1. Use cases only (*CAES*) to obtain loadings for calculation of *PC* scores for subjects in both cases and controls, 2. Use controls only (*COES*) to obtain the loadings for both groups, and 3. Use combined cases and controls (*CES*) to obtain the loadings for both groups. It is likely that in a case-control association study, loadings calculated from cases and controls can have different connotations and hence we only consider scenarios 1-3 hereafter. More formally, let (X_1, X_2, \dots, X_p) and (Y_1, Y_2, \dots, Y_p) be p -dimension vectors of SNPs at a given candidate region for cases and controls respectively, then we have,

Strategy 1 (*CAES*):

$$C_{XX}l_i^1 - \lambda l_i^1 = 0 \quad (3)$$

where C_{XX} is the correlation matrix of (X_1, X_2, \dots, X_p) , $l_i^1 = (l_{i1}^1, l_{i2}^1, \dots, l_{ip}^1)^T$ and $l_i^1 l_i^1 = 1, i = 1, 2, \dots, p$. The i^{th} *PC* for cases is calculated by

$$F_i^D = l_{i1}^1 X_1 + l_{i2}^1 X_2 + \dots + l_{ip}^1 X_p \quad (4)$$

and for controls

$$F_i^C = l_{i1}^1 Y_1 + l_{i2}^1 Y_2 + \dots + l_{ip}^1 Y_p \quad (5)$$

Strategy 2 (*COES*):

$$C_{YY}l_i - \lambda l_i = 0 \quad (6)$$

where C_{YY} is the correlation matrix of (Y_1, Y_2, \dots, Y_p) . The i^{th} *PC* for controls is calculated by

$$F_i^C = l_{i1} Y_1 + l_{i2} Y_2 + \dots + l_{ip} Y_p \quad (7)$$

And for cases, the i^{th} *PC*, $i = 1, 2, \dots, p$, is calculated by

$$F_i^D = l_{i1} X_1 + l_{i2} X_2 + \dots + l_{ip} X_p \quad (8)$$

Strategy 3 (*CES*):

$$C\tilde{l}_i - \lambda \tilde{l}_i = 0 \quad (9)$$

where C is the correlation matrix obtained from the pooled data of cases and controls, $\tilde{l}_i = (\tilde{l}_{i1}, \tilde{l}_{i2}, \dots, \tilde{l}_{ip})^T$ and $\tilde{l}_i^T \tilde{l}_i = 1, i = 1, 2, \dots, p$. The i^{th} *PC* of cases is calculated by

$$F_i^D = \tilde{l}_{i1} X_1 + \tilde{l}_{i2} X_2 + \dots + \tilde{l}_{ip} X_p \quad (10)$$

The i^{th} *PC* of controls is calculated by

$$F_i^C = \tilde{l}_{i1} Y_1 + \tilde{l}_{i2} Y_2 + \dots + \tilde{l}_{ip} Y_p \quad (11)$$

PCA-BCIT

Given a sample of N cases and M controls with p -SNP genotypes $(X_1, X_2, \dots, X_N)^T, (Y_1, Y_2, \dots, Y_M)^T$, and $X_i = (X_{1i}, X_{2i}, \dots, X_{pi})$ for the i^{th} case, $Y_i = (Y_{1i}, Y_{2i}, \dots, Y_{pi})$ for the i^{th} control, a *PCA-BCIT* is furnished in three steps:

Step 1: Sampling

Replicate samples of cases and controls are obtained with replacement separately from $(X_1^{(b)}, X_2^{(b)}, \dots, X_N^{(b)})^T$ and $(Y_1^{(b)}, Y_2^{(b)}, \dots, Y_M^{(b)})^T, b = 1, 2, \dots, B (B = 1000)$.

Step 2: PCA

For each replicate sample obtained at Step 1, *PCA* is conducted and a given number of *PCs* retained with a

threshold of 80% explained variance for all three strategies[16], expressed as $(F_1^D, F_2^D, \dots, F_k^D)^{(b)}$ and $(F_1^C, F_2^C, \dots, F_k^C)^{(b)}$.

Step 3: PCA-BCIT

3a) For each replicate, the mean of the k^{th} PC in cases is calculated by

$$mean(F_k^D)^{(b)} = \frac{1}{N} \sum_{i=1}^N F_{ki}^D \quad (12)$$

and that of the k^{th} PC in controls is calculated by

$$mean(F_k^C)^{(b)} = \frac{1}{M} \sum_{j=1}^M F_{kj}^C \quad (13)$$

3b) Given confidence level $(1 - \alpha)$, the confidence interval of $mean(F_k^D)^{(b)}$ is estimated by percentile method, with form

$$(P_{k\frac{\alpha}{2}}^D, P_{k(1-\frac{\alpha}{2})}^D) \text{ for case} \quad (14)$$

where $P_{k\frac{\alpha}{2}}^D$ is the $100\frac{\alpha}{2}^{th}$ percentile of $mean(F_k^D)^{(b)}$, and $P_{k(1-\frac{\alpha}{2})}^D$ is the $100(1-\frac{\alpha}{2})^{th}$ percentile.

The confidence interval of $mean(F_k^C)^{(b)}$ is estimated by

$$(P_{k\frac{\alpha}{2}}^C, P_{k(1-\frac{\alpha}{2})}^C) \text{ for control} \quad (15)$$

where $P_{k\frac{\alpha}{2}}^C$ is the $100\frac{\alpha}{2}^{th}$ percentile of $mean(F_k^C)^{(b)}$, and $P_{k(1-\frac{\alpha}{2})}^C$ is the $100(1-\frac{\alpha}{2})^{th}$ percentile.

3c) Confidence intervals of cases and controls are compared. The null hypothesis is rejected if $(P_{k\frac{\alpha}{2}}^D, P_{k(1-\frac{\alpha}{2})}^D)$ and $(P_{k\frac{\alpha}{2}}^C, P_{k(1-\frac{\alpha}{2})}^C)$ do not overlap, which is $mean(F_k^D)^{(b)}$ and $mean(F_k^C)^{(b)}$ are statistically different[19], indicating the candidate region is significantly associated with disease at level α . Otherwise, the candidate region is not significantly associated with disease at level α .

Simulation studies

We examine the performance of PCA-BCIT through simulations with data from the North American Rheumatoid Arthritis (RA) Consortium (NARAC) (868 cases and 1194 controls)[20], taking advantage of the fact that association between protein tyrosine phosphatase non-receptor type 22 (PTPN22) and the development of RA

has been established[21-24]. Nine SNPs have been selected from the PNP22 region (114157960-114215857), and most of the SNPs are within the same LD block (Figure 1). Females are more predisposed (73.85%) and are used in our simulation to ensure homogeneity. The corresponding steps for the simulation are as follows.

Step 1: Sampling

The observed genotype frequencies in the study sample are taken to be their true frequencies in populations of infinite sizes. Replicate samples of cases and controls of given size ($N, N = 100, 200, \dots, 1000$) are generated whose estimated genotype frequencies are expected to be close to the true population frequencies while both the allele frequencies and LD structure are maintained. Under null hypothesis, replicate cases and controls are sampled with replacement from the controls. Under alternative hypothesis, replicate cases and controls are sampled with replacement from the cases and controls respectively.

Step 2: PCA-BCITing

For each replicate sample, PCA-BCITs are conducted through the three strategies of extracting PCs as outlined above on association between PC scores and disease (RA).

Step 3: Evaluating performance of PCA-BCITs

Repeat steps 1 and 2 for K ($K = 1000$) times under both null and alternative hypotheses, and obtain the frequencies (P_α) of rejecting null hypothesis at level α ($\alpha = 0.05$).

Applications

PCA-BCITs are applied to both the NARAC data on PTPN22 in 1493 females (641 cases and 852 controls) described above and a data containing nine SNPs near μ -opioid receptor gene (OPRM1) in Han Chinese from Shanghai (91 cases and 245 controls) with endophenotype of heroin-induced positive responses on first use [25]. There are two LD blocks in the region of gene OPRM1 (Figure 2).

Results

Simulation study

The performance of PCA-BCIT is shown in Table 1 for the three strategies given a range of sample sizes. It can be seen that strategies 2 and 3 both have type I error rates approaching the nominal level ($\alpha = 0.05$), but those from strategy 1 deviate heavily. When sample size larger than 800, the power of PCA-BCIT is above 0.8, and strategies 2 and 3 outperform strategy 1 slightly.

Applications

For the NARAC data, Armitage trend test reveals none of the SNPs in significant association with RA using Bonferroni correction (Table 2), but the results of PCA-BCIT with strategies 2 and 3 show that the first PC

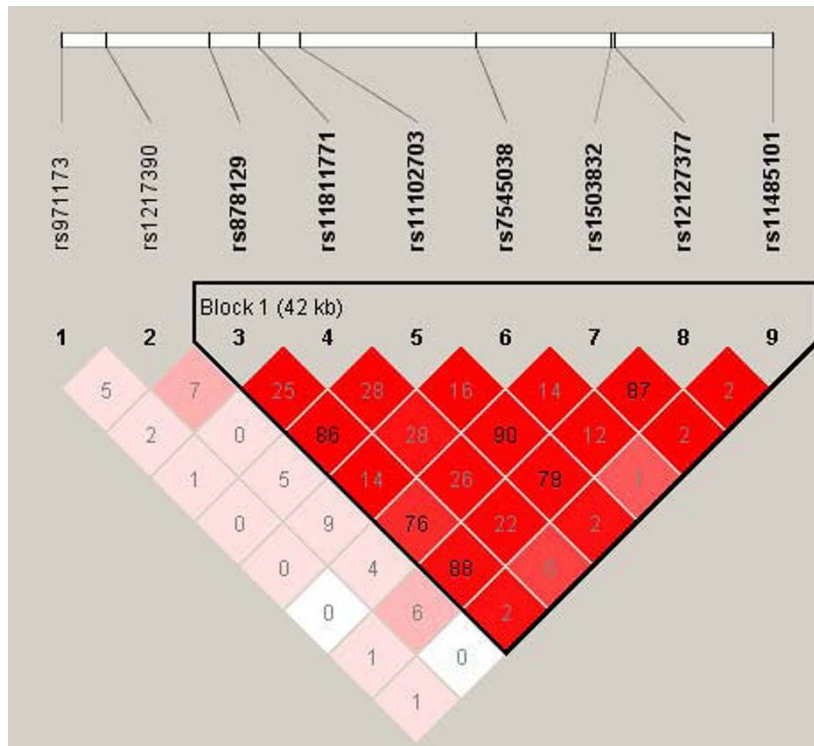


Figure 1 LD (r^2) among nine *PTPN22* SNPs. The nine *PTPN22* SNPs are rs971173, rs1217390, rs878129, rs11811771, rs11102703, rs7545038, rs1503832, rs12127377, rs11485101. The triangle marks a single LD block within this region: (rs878129, rs11811771, rs11102703, rs7545038, rs1503832, rs12127377, rs11485101).

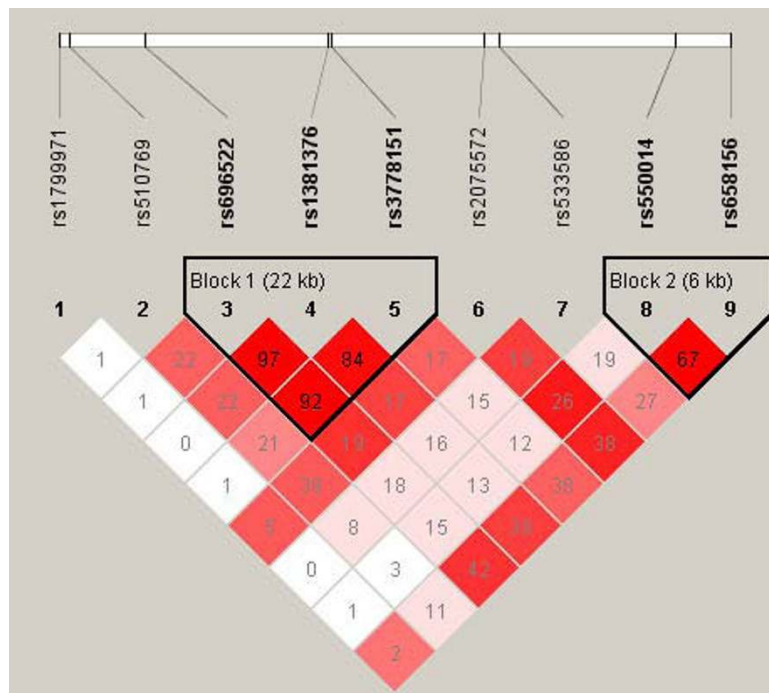


Figure 2 LD (r^2) among nine *OPRM1* SNPs. The nine *OPRM1* SNPs are rs1799971, rs510769, rs696522, rs1381376, rs3778151, rs2075572, rs533586, rs550014, rs658156. The triangles mark the LD block 1 (rs696522, rs1381376, rs3778151) and LD block 2 (rs550014, rs658156).

Table 1 Performance of PCA-BCIT at level 0.05 with strategies 1-3†

Sample size	Type I error			Power		
	1	2	3	1	2	3
100	0.014	0.036	0.037	0.156	0.163	0.176
200	0.016	0.044	0.036	0.249	0.278	0.292
300	0.017	0.028	0.029	0.383	0.426	0.368
400	0.014	0.04	0.02	0.508	0.485	0.516
500	0.009	0.035	0.042	0.613	0.595	0.597
600	0.006	0.032	0.042	0.677	0.662	0.683
700	0.007	0.061	0.04	0.733	0.758	0.73
800	0.004	0.043	0.045	0.801	0.791	0.819
900	0.005	0.057	0.051	0.826	0.855	0.858
1000	0.01	0.056	0.05	0.871	0.901	0.889

†1 case-only extracting strategy (CAES), 2 control-only extracting strategy (COES), 3 case-control extracting strategy (CES)

extracted in region of *PTPN22* is significantly associated with RA. The results are similar to that from permutation test (Table 3).

For the *OPRM1* data, the sample characteristics are comparable between cases and controls (Table 4), and three SNPs (rs696522, rs1381376 and rs3778151) are showed significant association with the endophenotype (Table 5). The results of *PCA-BCIT* with strategies 2 and 3 and permutation test are all significant at level $\alpha = 0.01$. In contrast, result from *PCA-LRT* is not significant at level $\alpha = 0.05$ with strategy 2 (Table 3). The apparent separation of cases and controls are shown in Figure 3 for *PCA-BCIT* with strategy 3, suggesting an intuitive interpretation.

Discussion

In this study, a *PCA*-based bootstrap confidence interval test[19,26-28] (*PCA-BCIT*) is developed to study gene-disease association using all SNPs genotyped in a given region. There are several attractive features of *PCA-*

Table 2 Armitage trend test on nine *PTPN22* SNPs and RA susceptibility

SNP	Genotype	Female			Male		
		Case	Control	P-value	Case	control	P-value
rs971173	CC	334	381	0.025	116	169	0.779
	AC	236	363		85	134	
	AA	71	106		26	39	
rs1217390	AA	268	319	0.333	99	112	0.108
	AG	272	392		89	175	
	GG	98	138		38	55	
rs878129	GG	338	507	0.009	131	187	0.384
	AG	251	291		83	130	
	AA	52	54		13	25	
rs11811771	AA	224	272	0.090	78	111	0.717
	AG	303	411		104	168	
	GG	112	169		45	62	
rs11102703	CC	312	469	0.024	121	174	0.418
	AC	269	314		90	137	
	AA	60	69		16	31	
rs7545038	GG	321	428	0.696	109	186	0.417
	AG	265	342		98	114	
	AA	52	80		20	40	
rs1503832	AA	324	487	0.013	129	185	0.249
	AG	262	306		86	127	
	GG	55	59		12	30	
rs12127377	AA	349	521	0.017	139	197	0.230
	AG	243	282		78	121	
	GG	49	48		10	24	
rs11485101	AA	564	738	0.656	206	305	0.430
	AG	72	112		21	35	
	GG	5	2		0	2	

None of the P-values is significant after Bonferroni Correction.

Table 3 PCA-BCIT, PCA-LRT and permutation test on real data

Study	Strategy†	99%CI	95%CI	P-value‡	
				PCA-LRT	Permutation test
PTPN22	2	(-5.4E-01,-4.7E-03)** (-7.5E-16,6.9E-16)	(-4.8E-01,-8.6E-02)* (-4.6E-16,4.2E-16)	0.006**	0.002**
	3	(1.7E-02,3.3E-01)** (-2.5E-01,-1.3E-02)	(4.9E-02,3.0E-01)* (-2.2E-01,-3.7E-02)	0.007**	0.002**
OPRM1	2	(-1.2E+00,-1.1E-02)** (-4.7E-16,5.0E-16)	(-1.1E+00,-1.8E-01)* (-3.7E-16,3.4E-16)	0.107	0.002**
	3	(5.3E-02,1.4E+00)** (-4.9E-01,-1.7E-02)	(2.4E-01,1.2E+00)* (-4.2E-01,-8.0E-02)	0.012*	0.004**

†2 control-only extracting strategy (COES), 3 case-control extracting strategy (CES)
 ‡* significant at levels $\alpha = 0.05$ (*) and $\alpha = 0.01$ (**).

based approaches. First of all, they are at least as powerful as genotype- and haplotype-based methods[7,16,17]. Secondly, they are able to capture LD information between correlated SNPs and easy to compute with needless consideration of multicollinearity and multiple testing. Thirdly, BCIT integrates point estimation and hypothesis testing as a single inferential statement of great intuitive appeal[29] and does not rely on the distributional assumption of the statistic used to calculate confidence interval[19,26-29].

While there have been several different but closely related forms of bootstrap confidence interval calculations[28], we focus on percentiles of the asymptotic distribution of PCs for given confidence levels to estimate the confidence interval. PCA-BCIT is a data-learning method[29], and shown to be valid and powerful for sufficiently large number of replicates in our study. Our investigation involving three strategies of extracting PCs reveals that strategy 1 is invalid, while strategies 2 and 3 are acceptable. From analyses of real data we find that PCA-BCIT is more favourable compared with PCA-LRT and permutation test. It is suggested that a practical advantage of PCA-BCIT is that it offers an intuitive measure of difference between cases and controls by using the set of SNPs (PC scores) in a candidate region (Figure 3). As extraction of PCs through COES is more in line with the

principle of a case-control study, it will be our method of choice given that it has a comparable performance with CES. Nevertheless, PCA-BCIT has the limitation that it does not directly handle covariates as is usually done in a regression model.

Conclusions

PCA-BCIT is both a valid and a powerful PCA-based method which captures multi-SNP information in study of gene-disease association. While extracting PCs based on CAES, COES and CES all have good performances, it appears that COES is more appropriate to use.

Abbreviations

SNP: single nucleotide polymorphism; HWE: Hardy-Weinberg Equilibrium; LD: linkage disequilibrium; LRT: logistic regression test; PCA: principle component analysis; PC: principle component; ES: extracting strategy; SES: separate case and control extracting strategy (strategy 0); CAES: case-based extracting strategy (strategy 1); COES: control-based extracting strategy (strategy 2); CES: combined case and control extracting strategy (strategy 3); BCIT: bootstrap confidence interval test.

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Table 4 Sample characteristics of heroin-induced positive responses on first use

	Cases (N = 91)	Controls (N = 245)	P-value
Age (yrs)	30.42 ± 7.65	30.93 ± 8.18	0.6057
Women (%)	26.4	29.8	0.5384
Age at onset (yrs)	26.29 ± 7.41	26.97 ± 7.89	0.4760
Reason for first use of heroin			0.7173
Curiousness	79.1	75.1	
Peer pressure	6.6	4.9	
Physical disease	7.7	10.2	
Trouble	5.5	6.1	
Other reasons	1.1	3.8	

Table 5 Armitage trend tests on nine *OPRM1* SNPs and heroin-induced positive responses on first use

SNP	Genotype	Count and frequency				Armitage trend test	
		Cases		Controls		Chi-square	P-value
rs1799971	AA	55	0.604	150	0.622	0.003	0.9537
	AG	27	0.297	64	0.266		
	GG	9	0.099	24	0.112		
rs510769	TT	56	0.667	167	0.749	2.744	0.0976
	TC	24	0.286	53	0.237		
	CC	4	0.048	4	0.018		
rs696522	AA	64	0.762	215	0.907	11.097	0.0009*
	AG	19	0.226	21	0.089		
	GG	1	0.012	1	0.004		
rs1381376	CC	70	0.769	221	0.913	13.409	0.0003*
	CT	20	0.220	21	0.087		
	TT	1	0.011	0	0.000		
rs3778151	GG	66	0.733	215	0.896	14.655	0.0001*
	GA	23	0.256	25	0.104		
	AA	1	0.011	0	0.000		
rs2075572	GG	50	0.556	149	0.642	1.574	0.2096
	GC	33	0.367	82	0.353		
	CC	7	0.078	11	0.047		
rs533586	TT	68	0.840	203	0.868	0.761	0.3830
	TC	12	0.148	31	0.132		
	CC	1	0.012	0	0.000		
rs550014	TT	78	0.857	203	0.832	0.093	0.7602
	TC	12	0.132	41	0.168		
	CC	1	0.011	0	0.000		
rs658156	GG	65	0.714	192	0.787	2.041	0.1531
	GA	24	0.264	52	0.213		
	AA	1	0.011	0	0.000		

* significant after Bonferroni Correction.

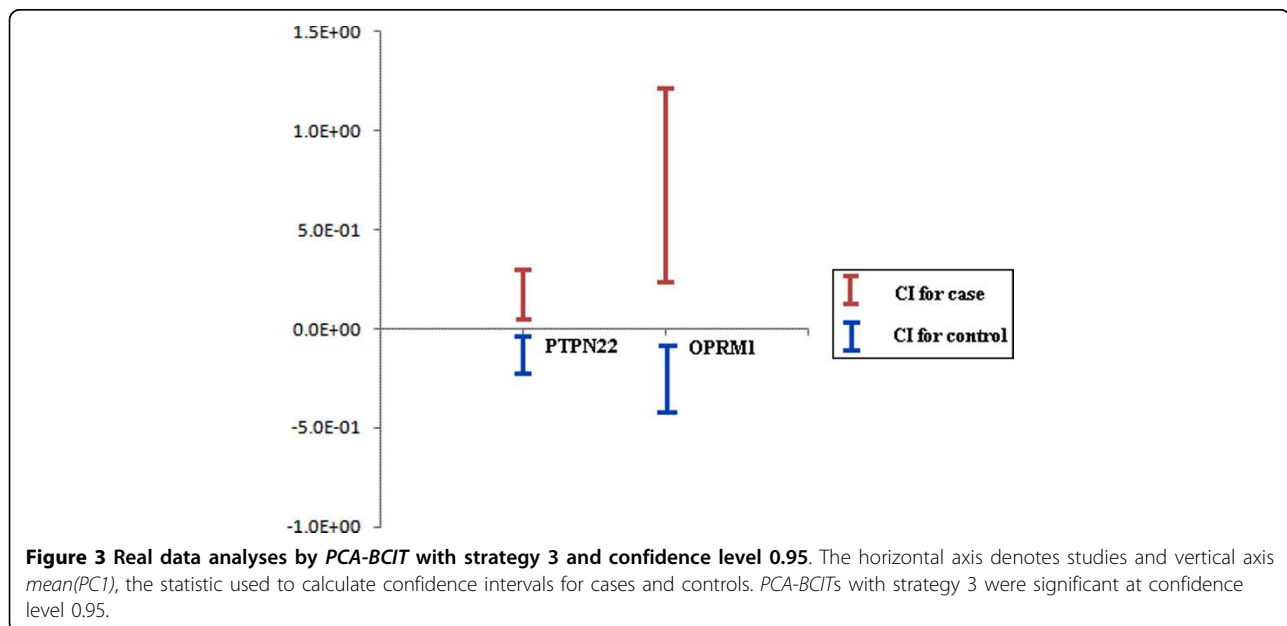


Figure 3 Real data analyses by PCA-BCIT with strategy 3 and confidence level 0.95. The horizontal axis denotes studies and vertical axis *mean(PC1)*, the statistic used to calculate confidence intervals for cases and controls. *PCA-BCITs* with strategy 3 were significant at confidence level 0.95.

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Authors' contributions

QQP, JHZ, and FZX conceptualized the study, acquired and analyzed the data and prepared for the manuscript. All authors approved the final manuscript.

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References

- Morton NE, Collins A: Tests and estimates of allelic association in complexes. *Proc Natl Acad Sci USA* 1998, **95**:11389-11393.
- Sasieni PD: From genotypes to genes: doubling the sample size. *Biometrics* 1997, **53**:1253-1261.
- Gordon D, Haynes C, Yang Y, Kramer PL, Finch SJ: Linear trend tests for case-control genetic association that incorporate random phenotype and genotype misclassification error. *Genet Epidemiol* 2007, **31**:853-870.
- Slager SL, Schaid DJ: Case-control studies of genetic markers: Power and sample size approximations for Armitage's test for trend. *Human Heredity* 2001, **52**:149-153.
- Sidak Z: On Multivariate Normal Probabilities of Rectangles: Their Dependence on Correlations. *The Annals of Mathematical Statistics* 1968, **39**:1425-1434.
- Sidak Z: On Probabilities of Rectangles in Multivariate Student Distributions: Their Dependence on Correlations. *The Annals of Mathematical Statistics* 1971, **42**:169-175.
- Zhang FY, Wagener D: An approach to incorporate linkage disequilibrium structure into genomic association analysis. *Journal of Genetics and Genomics* 2008, **35**:381-385.
- Balding DJ: A tutorial on statistical methods for population association studies. *Nature Reviews Genetics* 2006, **7**:781-791.
- Schaid DJ, McDonnell SK, Hebring SJ, Cunningham JM, Thibodeau SN: Nonparametric tests of association of multiple genes with human disease. *American Journal of Human Genetics* 2005, **76**:780-793.
- Becker T, Schumacher J, Cichon S, Baur MP, Knapp M: Haplotype interaction analysis of unlinked regions. *Genetic Epidemiology* 2005, **29**:313-322.
- Chapman JM, Cooper JD, Todd JA, Clayton DG: Detecting disease associations due to linkage disequilibrium using haplotype tags: A class of tests and the determinants of statistical power. *Human Heredity* 2003, **56**:18-31.
- Epstein MP, Satten GA: Inference on haplotype effects in case-control studies using unphased genotype data. *American Journal of Human Genetics* 2003, **73**:1316-1329.
- Fallin D, Cohen A, Essioux L, Chumakov I, Blumenfeld M, Cohen D, Schork NJ: Genetic analysis of case/control data using estimated haplotype frequencies: Application to APOE locus variation and Alzheimer's disease. *Genome Research* 2001, **11**:143-151.
- Stram DO, Pearce CL, Bretsky P, Freedman M, Hirschhorn JN, Altshuler D, Kolonel LN, Henderson BE, Thomas DC: Modeling and E-M estimation of haplotype-specific relative risks from genotype data for a case-control study of unrelated individuals. *Human Heredity* 2003, **55**:179-190.
- Clayton D, Chapman J, Cooper J: Use of unphased multilocus genotype data in indirect association studies. *Genetic Epidemiology* 2004, **27**:415-428.
- Gauderman WJ, Murcray C, Gilliland F, Conti DV: Testing association between disease and multiple SNPs in a candidate gene. *Genetic Epidemiology* 2007, **31**:383-395.
- Oh S, Park T: Association tests based on the principal-component analysis. *BMC Proc* 2007, **1**(Suppl 1):S130.
- Wang T, Elston RC: Improved power by use of a weighted score test for linkage disequilibrium mapping. *American Journal of Human Genetics* 2007, **80**:353-360.
- Heller G, Venkatraman ES: Resampling procedures to compare two survival distributions in the presence of right-censored data. *Biometrics* 1996, **52**:1204-1213.
- Plenge RM, Seielstad M, Padyukov L, Lee AT, Remmers EF, Ding B, Liew A, Khalili H, Chandrasekaran A, Davies LRL, et al: TRAF1-C5 as a risk locus for rheumatoid arthritis - A genomewide study. *New England Journal of Medicine* 2007, **357**:1199-1209.
- Begovich AB, Carlton VE, Honigberg LA, Schrodi SJ, Chokkalingam AP, Alexander HC, Ardlie KG, Huang Q, Smith AM, Spoecke JM, et al: A missense single-nucleotide polymorphism in a gene encoding a protein tyrosine phosphatase (PTPN22) is associated with rheumatoid arthritis. *Am J Hum Genet* 2004, **75**:330-337.
- Carlton VE, Hu XL, Chokkalingam AP, Schrodi SJ, Brandon R, Alexander HC, Chang M, Catanese JJ, Leong DU, Ardlie KG, et al: PTPN22 genetic variation: Evidence for multiple variants associated with rheumatoid arthritis. *American Journal of Human Genetics* 2005, **77**:567-581.
- Kallberg H, Padyukov L, Plenge RM, Ronnelid J, Gregersen PK, Helm-van Mil van der AHM, Toes REM, Huizinga TW, Klareskog L, Alfredsson L, et al: Gene-gene and gene-environment interactions involving HLA-DRB1, PTPN22, and smoking in two subsets of rheumatoid arthritis. *American Journal of Human Genetics* 2007, **80**:867-875.
- Plenge RM, Padyukov L, Remmers EF, Purcell S, Lee AT, Karlson EW, Wolfe F, Kastner DL, Alfredsson L, Altshuler D, et al: Replication of putative candidate-gene associations with rheumatoid arthritis in > 4,000 samples from North America and Sweden: Association of susceptibility with PTPN22, CTLA4, and PADI4. *American Journal of Human Genetics* 2005, **77**:1044-1060.
- Zhang D, Shao C, Shao M, Yan P, Wang Y, Liu Y, Liu W, Lin T, Xie Y, Zhao Y, et al: Effect of mu-opioid receptor gene polymorphisms on heroin-induced subjective responses in a Chinese population. *Biol Psychiatry* 2007, **61**:1244-1251.
- Carpenter J: Test Inversion Bootstrap Confidence Intervals. *Journal of the Royal Statistical Society Series B (Statistical Methodology)* 1999, **61**:159-172.
- Davison AC, Hinkley DV, Young GA: Recent developments in bootstrap methodology. *Statistical Science* 2003, **18**:141-157.
- DiCiccio TJ, Efron B: Bootstrap confidence intervals. *Statistical Science* 1996, **11**:189-212.
- Efron B: Bootstrap Methods: Another Look at the Jackknife. *The Annals of Statistics* 1979, **7**:1-26.

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