

Supporting Information

pH-Induced Mechanistic Changeover From Hydroxyl Radicals to Iron(IV) in the Fenton Reaction

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Additional experimental detail

The following chemicals were obtained commercially and used as received: iron(II) perchlorate hydrate (98%), iron(III) perchlorate hydrate (low chloride < 0.005%), deuterio perchloric acid DCIO₄ (68 wt% in D₂O, 99+ atom % D), titanium(IV) oxysulfate (99.99% purity, 15 wt% solution in dilute sulfuric acid), deuterium oxide (99.9 atom %D), (R)-(+)-methyl *p*-tolyl sulfoxide (99%), and 1,10-phenanthroline (99+ %) (all Aldrich), dimethyl sulfoxide (≥ 99.9%, A.C.S spectrophotometric grade, Sigma-Aldrich), sodium hydroxide (99.1%), hydrogen peroxide (30 wt%), perchloric acid (70 wt%), anhydrous dibasic sodium phosphate (all Fisher), monobasic

sodium phosphate monohydrate (Baker) piperazine-N,N'-bis(4-butanefulfonic acid) (PIPBS), 4-(N-morpholino)butanesulfonic acid (MOBS), 3-(N-morpholino)-propanesulfonic acid (MOPS), and 2-(N-morpholino)ethanesulfonic acid (MES) (all GFS).

Stock solutions of iron(II) perchlorate in H₂O or D₂O were prepared freshly before each set of experiments and standardized with phenanthroline. Solutions of H₂O₂ were standardized with titanium oxysulfate¹ which places a reliable detection limit at 0.02 mM H₂O₂. Deionized water was further purified by passage through a Barnstead Easy Pure II -UV/UF water purifier.

UV-Vis absorbance measurements and kinetic studies used a Shimadzu UV-3101 PC spectrophotometer, Olis RSM-1000 stopped-flow instrument, and Applied Photophysics (APP) sequential stopped-flow DX-17MV at 25.0 ± 0.1 °C. Gaseous products were analyzed by an Agilent Technologies 7890A GC gas chromatograph equipped with a FID detector and a 15-m capillary column (GS-GASPRO), 0.320 mm I.D. Nitrogen flow rate was constant at 25 cm³/s. Both the split injector (1:40) and FID detector were held at 200⁰ C. The initial oven temperature was 40⁰ C, and was increased by 10⁰ C/min. ¹H-NMR spectra were recorded with a 400 MHz Bruker DRX-400 spectrometer at room temperature. The pH was measured with an Accumet AP71 pH meter. Kinetic experiments were performed at 25.1 ± 0.1⁰ C.

Procedures All of the solutions were purged with argon before mixing and appeared clear to the eye in both tertiary amine and phosphate buffers prior to and during the reaction. The solvent was H₂O for kinetic studies, and D₂O for NMR product analysis. The pH (pD) was controlled with noncoordinating tertiary amine buffers MES (pK_a 6.06), MOPS (pK_a 7.09), MOBS (pK_a 7.48) and PIPBS (pK_{a1} 4.29, pK_{a2} 8.55)² or with phosphate buffers (pH 6-8). Buffer concentrations were typically 8-50 times greater than the concentration of the limiting reagents, i. e. at the level required to hold the pH at the desired value without interfering with the NMR

spectra or altering the chemistry. The pH decrease in kinetics experiments was less than 0.2 units. In the NMR experiments, which typically used 1-2 mM of Fe(II) and H₂O₂, the pH decrease was typically 0.3-0.5 units. Solutions were acidified before the NMR spectra were recorded. Concentrations of Fe(II) in spent reaction solutions were determined with phenanthroline. A correction was applied for the absorbance of iron(III)-phenanthroline as previously described.³

The stoichiometry was determined from absorbance changes at 270 nm using $\Delta\epsilon_{270} = 2650 \pm 50 \text{ M}^{-1} \text{ cm}^{-1}$ at pH 6-7. This value was determined by oxidizing a solution of Fe(II) (0.010 - 0.020 mM) with excess H₂O₂ (0.1 - 1.5 mM) in the stopped-flow and monitoring the absorbance for up to 30 seconds after the completion of the reaction. During this time, the reading remained constant and yielded $\epsilon_{270} = 2600 \pm 50 \text{ M}^{-1} \text{ cm}^{-1}$. Several experiments were also performed on a longer time scale by mixing the reagents (0.02 mM Fe(II) and 0.2 mM H₂O₂ in 0.6 mM MES or MOPS buffer) in a spectrophotometric cell and monitoring the absorbance with a conventional spectrophotometer. The absorbance after the completion of the reaction remained constant for at least five minutes and yielded $\epsilon_{270} = 2700 \pm 50 \text{ M}^{-1} \text{ cm}^{-1}$. The constancy of ϵ_{270} on millisecond-to-minute time scale confirms that the turbidity or precipitation of iron(III) did not affect absorbance readings. This point is also illustrated in the kinetic plots in Figures S6-S8. The three experiments had identical initial concentrations of Fe(II) and H₂O₂ (in excess), while the concentration of (CH₃)₂SO was varied. The reaction times varied over a factor of 14, but the overall absorbance change was the same within the experimental error, showing again that changes in the degree of hydrolysis or agglomeration of the Fe(III) product either did not take place or had no effect on the absorbance around 270 nm.

Product analysis by NMR. A solution of hydrogen peroxide was added to a magnetically stirred solution of $\text{Fe}(\text{H}_2\text{O})_6^{2+}/\text{Fe}(\text{H}_2\text{O})_5\text{OH}^+$ ($\text{pK}_a = 9.5$)⁴ and substrate in D_2O buffered at the desired pD under argon. The pH was measured before and after the reaction, and converted to pD by adding 0.4 to the measured value. Immediately after the completion of the reaction, the solutions were acidified to pD 1 with perchloric acid to avoid agglomeration and precipitation of iron(III) hydroxide(s), and the NMR spectrum was run within minutes. Such solutions contained small amounts of unreacted Fe(II) or H_2O_2 , but not both, so that no additional oxidation of the substrate could take place after the acidification. Acetonitrile (0.79 mM) was used as internal standard to quantify the products. No shift or broadening of the product resonances was observed in acidified solutions.

In phosphate buffer at pH 7, the products are ethane (60% yield by GC) and methylsulfinate (25% by NMR), indicative of HO^\bullet radicals. The resonances were somewhat broadened, most likely because solutions of iron(III) became slightly inhomogeneous during the time required to record the spectrum. The reasons for the lower yields of methylsulfinate are not fully understood, but a portion of the anion may be complexed to the Fe(III) product and thus not detectable by NMR. In acidic solutions, where the binding would be much weaker (pK_a for $\text{CH}_3\text{S}(\text{O})\text{OH}$ is 2.35), the methyl signal for methylsulfinic acid is shifted to 2.50 ppm and appears on the side of the strong $(\text{CH}_3)_2\text{SO}$ signal (2.55 ppm) which makes the integration imprecise. Under these conditions, a signal for sulfonic acid (2.63 ppm) was also detected, but it was not well separated from DMSO. Control experiments with samples of genuine sulfinic acid confirmed that significant oxidation to sulfonic acid takes place in the time required for (aerobic) manipulation of the solutions and the recording of the spectrum. Despite the difficulties in detecting all of the expected sulfinic acid, the majority of the reaction in phosphate buffers

appears to involve hydroxyl radicals based on the data and arguments in the main text, and on the finding that the yields of the accompanying product, ethane, were much higher, about 60% (Table 1).



Stopped-flow experiments. Aqueous solutions of iron(II) were mixed with the buffer and dmsO immediately before the experiment and loaded into one of the stopped-flow syringes. The other syringe was filled with the H₂O₂ solution. The formation of Fe(III) was monitored either at 270 nm with the Applied Photophysics APP DX-17MV instrument or in the 260-320 nm range with an Olis RSM-1000 rapid scan instrument.

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Figure S1. ¹H NMR spectrum of the products of the reaction of 1.2 mM Fe(ClO₄)₂, 1.1 mM H₂O₂ and 36 mM DMSO in MOPS buffer (11 mM). Initial pD 7.6, final pD 7.2. Spectrum was recorded after acidifying reaction mixture to 0.1 M DClO₄. The reaction generated (50 ± 5 μM) DMSO₂. ¹³C satellite peaks are denoted with an asterisk.

Figure S2. a) ¹H NMR spectrum showing TMSO₂ (0.65 ± 0.05 mM) generated in the reaction of 1.8 mM Fe(ClO₄)₂, 2.4 mM H₂O₂ and 29 mM TMSO in PIPBS buffer (15 mM). Initial pD 7.1, final pD 5.7. **b)** Control experiment: ¹H NMR spectrum of a mixture of H₂O₂ (2.0 mM), TMSO (43 mM) and PIPBS buffer (22 mM) in D₂O, pD 7.1. The spectra were recorded without acidifying the reaction mixtures. ¹³C satellite peaks are denoted with an asterisk.

Figure S3. ¹H NMR spectrum after the completion of the reaction of 1.0 mM Fe(ClO₄)₂, 1.0 mM H₂O₂ and 50 mM TMSO in D₂O, phosphate buffer (20 mM). Initial pD 6.6, final pD 6.0. Spectrum was recorded after acidifying reaction mixture to 0.1 M DClO₄. ¹³C satellite peaks are denoted with an asterisk.

Figure S4. ¹H NMR spectrum showing (CH₃)₂SO₂ (0.26 mM) generated from the reaction of Fe(ClO₄)₂ (0.49 mM), H₂O₂ (0.49 mM) and (CH₃)₂SO (190 mM) in D₂O, MES buffer (9.6 mM),

pD 6.7. Spectrum was recorded after acidifying the reaction mixture to 0.1 M DClO₄. ¹³C satellite peaks are denoted with an asterisk.

Figure S5 ¹H NMR spectrum showing (CH₃)₂SO₂ (0.49 mM) generated from the reaction of Fe(ClO₄)₂ (0.49 mM), H₂O₂ (0.49 mM) and (CH₃)₂SO (950 mM) in MES buffer (9.6 mM), pD 6.70. Spectrum was recorded after acidifying the reaction mixture to 0.1 M DClO₄. ¹³C satellite peaks are denoted with an asterisk.

Figure S6. Kinetic (stopped flow) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ in the absence of (CH₃)₂SO, pH 6.2 (0.56 mM MES)

Figure S7. Kinetic (stopped flow) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ at 0.50 M (CH₃)₂SO, pH 6.2 (0.56 mM MES)

Figure S8. Kinetic (conventional spectrophotometer) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ at 0.98 M DMSO, pH 6.2 (0.6 mM MES)

Figure S9. Plot of *k*_{obs} vs [H₂O₂] for the Fe(II)/ H₂O₂ reaction at pH 6.1 (MES buffer). Conditions: [Fe(ClO₄)₂]₀ = 0.005 – 0.035 mM, [MES] = 0.25 – 0.55 mM, 25 °C. pH decreased during the reaction by 0.2 units, see text.

Figure S10. Plot of *k*_{obs} vs [H₂O₂] for the Fe(II)/ H₂O₂ reaction at pH 7 in a MOPS buffer. Conditions: [Fe(ClO₄)₂]₀ = 0.008 – 0.015 mM, [MOPS] = 0.5 - 0.6 mM, 25 °C. pH decreased during the reaction by 0.2 units, see text.

Figure S11. Plot of *k*_{obs} vs [Fe(II)] for the Fe(II)/ H₂O₂ reaction at pH 6.1 in a MES buffer. Conditions: [H₂O₂]₀ = 0.005 - 0.02 mM, [MES] = 0.5-0.6 mM, 25 °C. pH decreased during the reaction by 0.2 units, see text.

Figure S12. Representative kinetic trace in phosphate buffer. Conditions: 0.031 mM Fe(II), 1.0 mM H₂O₂, pH 6.1 (0.6 mM phosphate buffer)

Figure S13. Plot of pseudo-first-order rate constants for the reaction of Fe(ClO₄)₂ with H₂O₂ against the concentration of H₂O₂ in phosphate buffer. Conditions: [Fe(ClO₄)₂] = 0.018 – 0.030 mM, [phosphate] = 0.60 mM, pH = 6.2.

Figure S14. (a) ¹H NMR spectrum of the reaction mixture after the completion of the reaction between Fe(ClO₄)₂ (1.0 mM), H₂O₂ (1.0 mM), and (CH₃)₂SO (5 mM) at pD 1 in the presence of 5 mM MES. The reaction generated CH₃SO₂H and C₂H₆ in amounts comparable to those

obtained in the absence of MES. b) Mixture of MES (5 mM) and H₂O₂ (1 mM), pD 1. c) MES (5 mM), pD 1.

Figures S1-S11

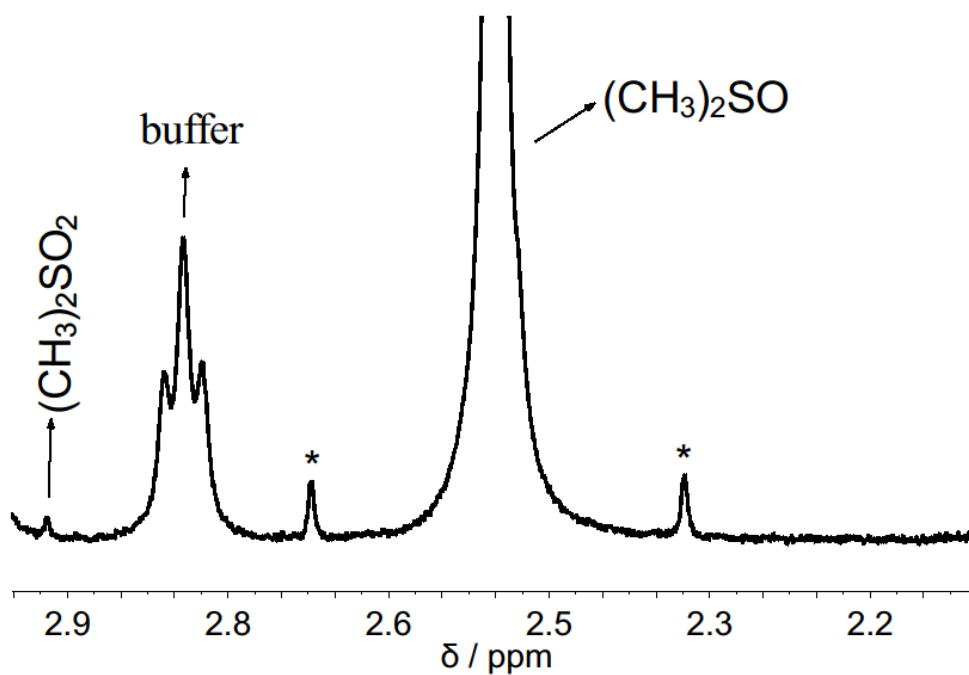


Figure S1. ¹H NMR spectrum of the products of the reaction of 1.2 mM Fe(ClO₄)₂, 1.1 mM H₂O₂ and 36 mM DMSO in MOPS buffer (11 mM). Initial pD 7.6, final pD 7.2. Spectrum was

recorded after acidifying reaction mixture to 0.1 M DClO_4 . The reaction generated $(50 \pm 5 \mu\text{M})$ DMSO_2 . ^{13}C satellites are denoted with an asterisk.

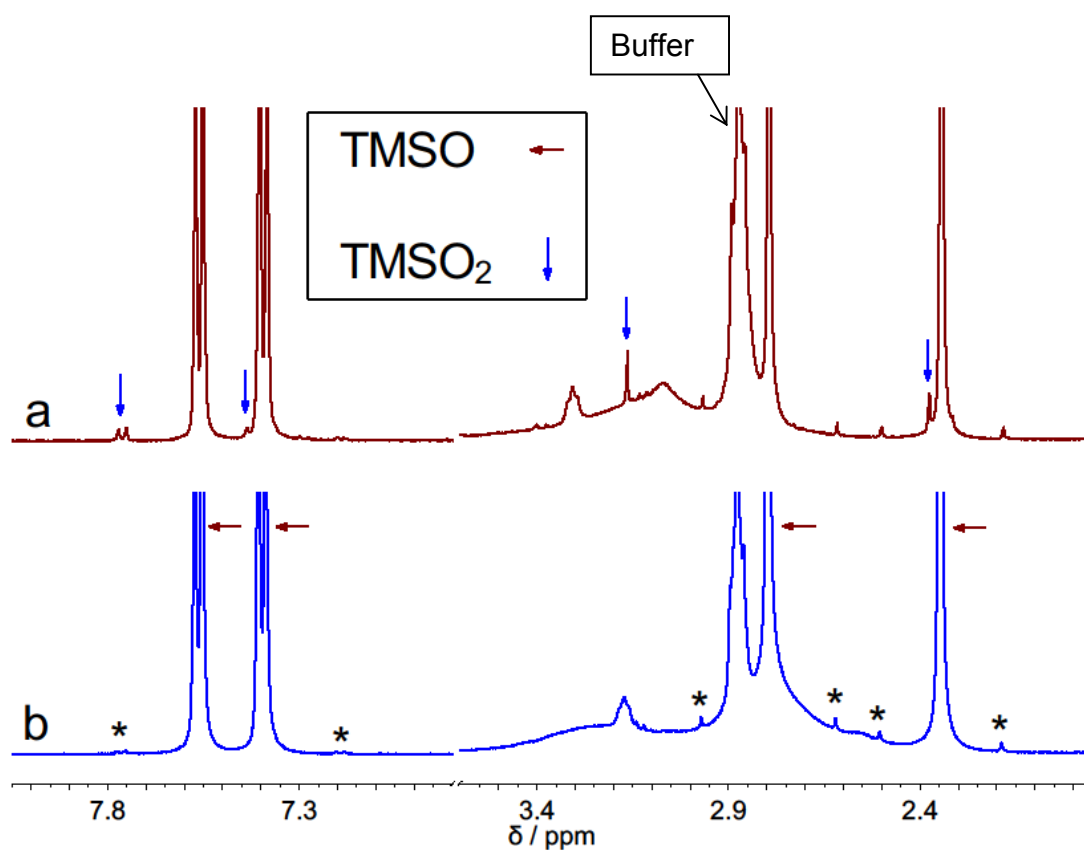


Figure S2. **a)** ^1H NMR spectrum showing TMSO_2 ($0.65 \pm 0.05 \text{ mM}$) generated in the reaction of $1.8 \text{ mM Fe}(\text{ClO}_4)_2$, $2.4 \text{ mM H}_2\text{O}_2$ and 29 mM TMSO in PIPBS buffer (15 mM). Initial pD 7.1 , final pD 5.7 . **b)** Control experiment: ^1H NMR spectrum of a mixture of H_2O_2 (2.0 mM), TMSO (43 mM) and PIPBS buffer (22 mM) in D_2O , pD 7.1 . The spectra were recorded without acidifying the reaction mixtures. ^{13}C satellite peaks are denoted with an asterisk.

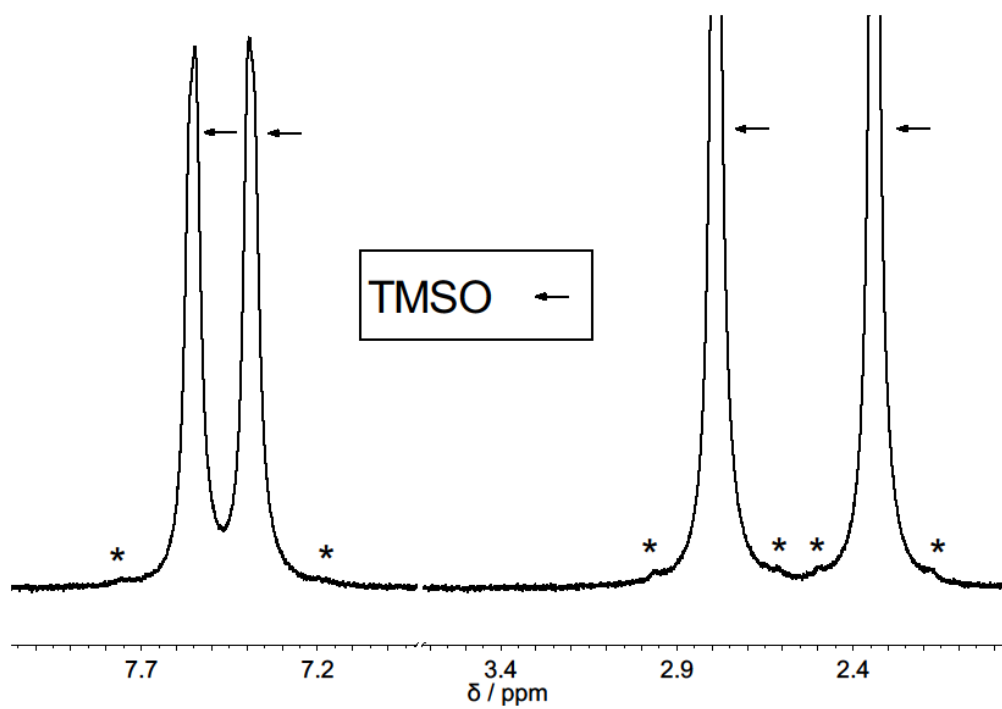


Figure S3. ^1H NMR spectrum after the completion of the reaction between 1.0 mM $\text{Fe}(\text{ClO}_4)_2$, 1.0 mM H_2O_2 and 50 mM TMSO in D_2O , phosphate buffer (20 mM). Initial pD 6.6, final pD 6.0. ^{13}C satellites are denoted with an asterisk.

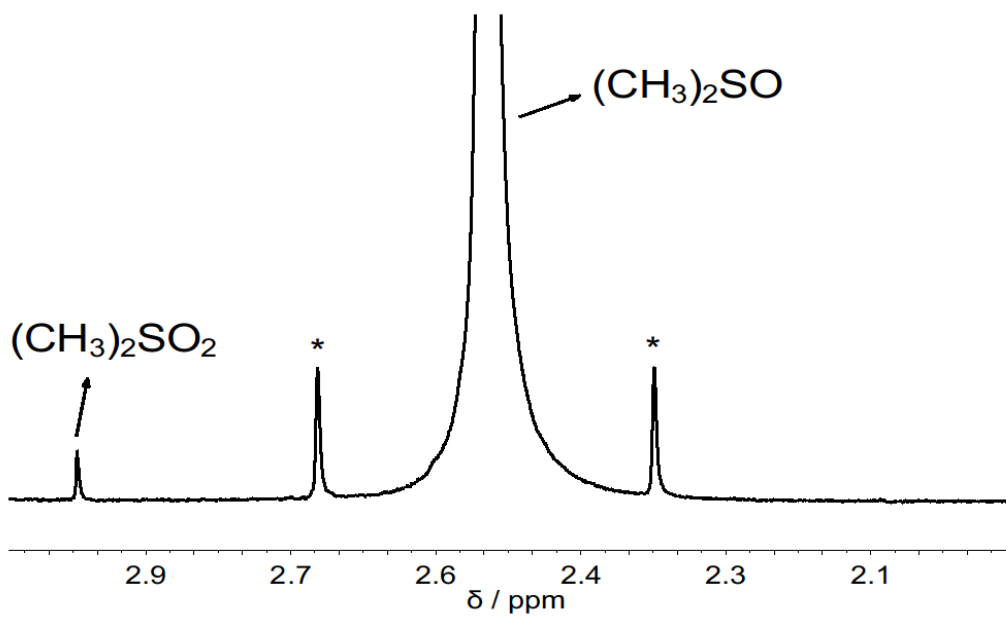


Figure S4. ¹H NMR spectrum showing (CH₃)₂SO₂ (0.26 mM) generated from the reaction of Fe(ClO₄)₂ (0.49 mM), H₂O₂ (0.49 mM) and (CH₃)₂SO (190 mM) in D₂O, MES buffer (9.6 mM), pD 6.7. Spectrum was recorded after acidifying the reaction mixture to 0.1 M DClO₄. ¹³C satellites are denoted with an asterisk.

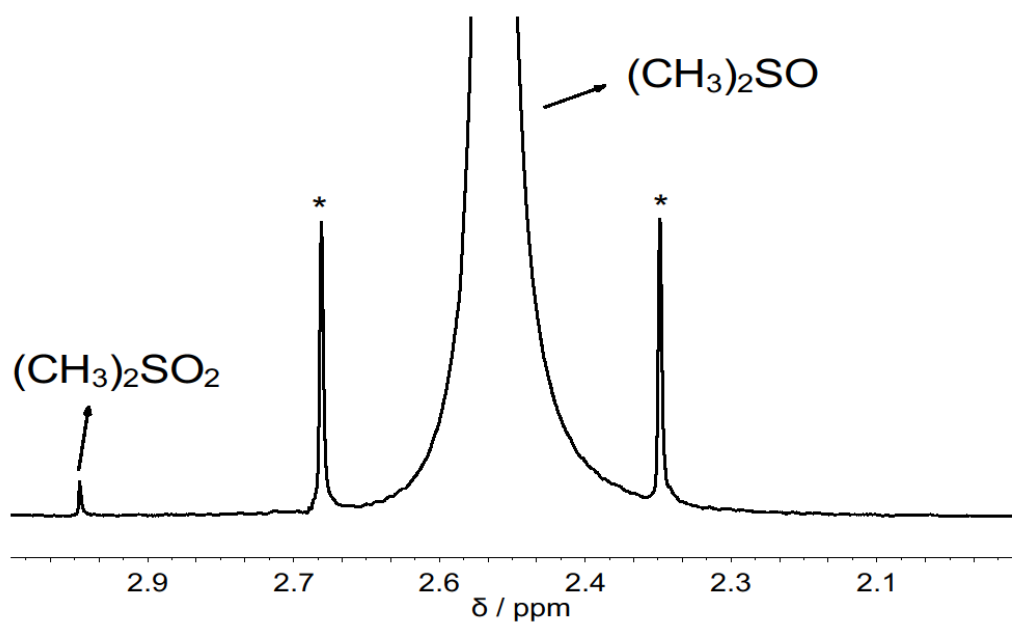


Figure S5 ^1H NMR spectrum showing $(\text{CH}_3)_2\text{SO}_2$ (0.49 mM) generated from the reaction of $\text{Fe}(\text{ClO}_4)_2$ (0.49 mM), H_2O_2 (0.49 mM) and $(\text{CH}_3)_2\text{SO}$ (950 mM) in MES buffer (9.6 mM), pD 6.70. Spectrum was recorded after acidifying the reaction mixture to 0.1 M DClO_4 . ^{13}C satellite peaks are denoted with an asterisk.

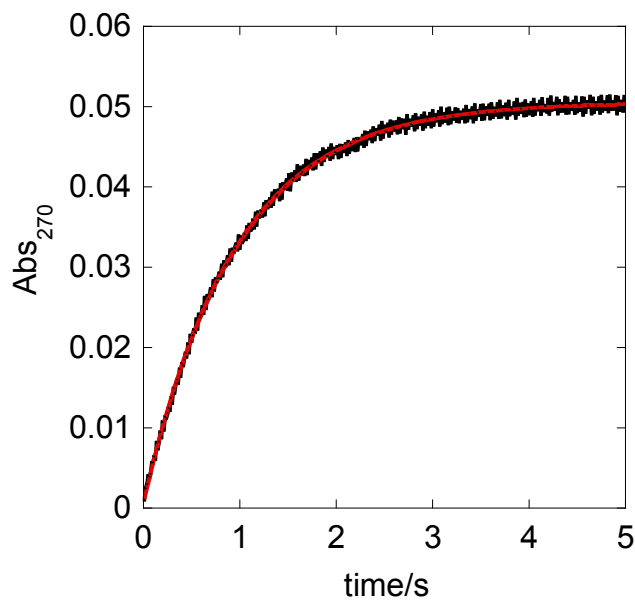


Figure S6. Kinetic (stopped flow) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ in the absence of DMSO, pH 6.2 (0.56 mM MES)

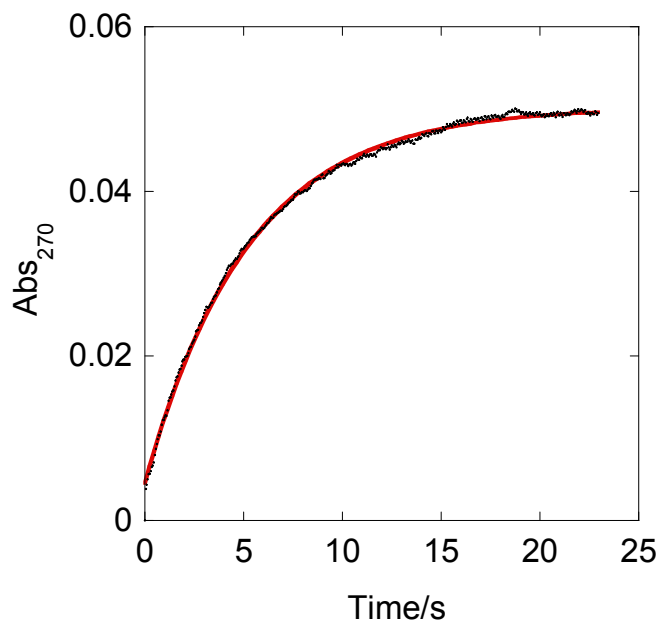


Figure S7. Kinetic (stopped flow) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ at 0.50 M DMSO, pH 6.2 (0.56 mM MES)

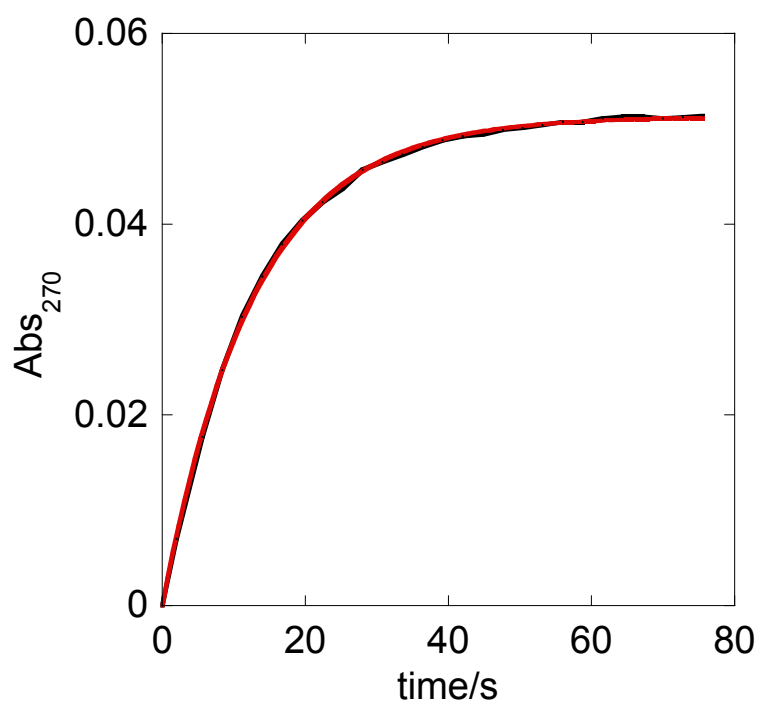


Figure S8. Kinetic (conventional spectrophotometer) trace for a reaction between 0.020 mM Fe(II) and 0.78 mM H₂O₂ at 0.98 M DMSO, pH 6.2 (0.6 mM MES)

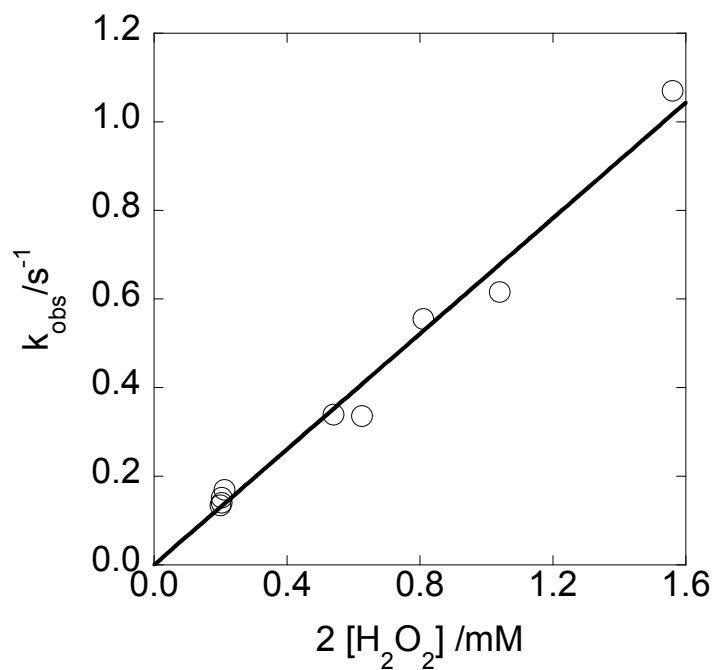


Figure S9. Plot of k_{obs} vs $2[\text{H}_2\text{O}_2]$ for the Fe(II)/ H_2O_2 reaction at pH 6.1 in MES buffer. Conditions: $[\text{Fe}(\text{ClO}_4)_2]_0 = 0.005 - 0.035 \text{ mM}$, $[\text{MES}] = 0.25 - 0.55 \text{ mM}$, $25 \text{ }^\circ\text{C}$. The pH decreased during the reaction by 0.2 units, see text.

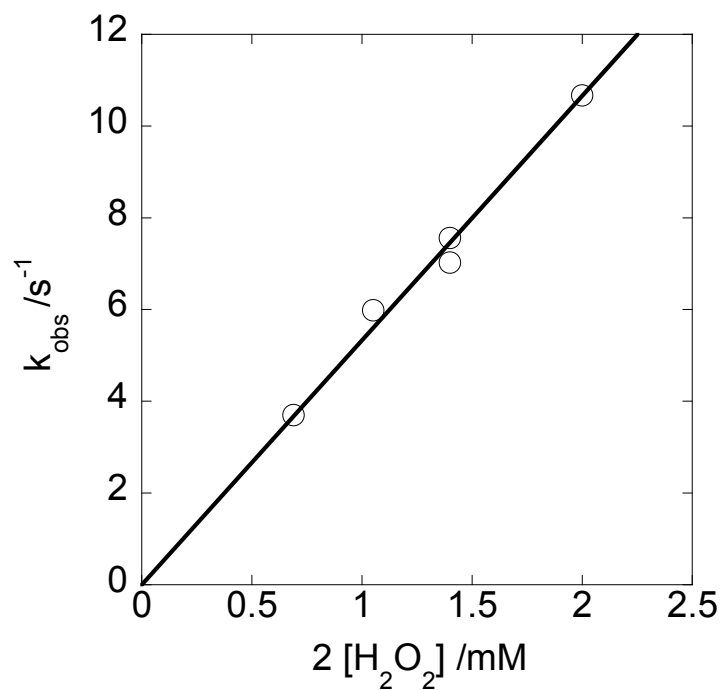


Figure S10. Plot of k_{obs} vs $[\text{H}_2\text{O}_2]$ for the Fe(II)/ H_2O_2 reaction at pH 7 in a MOPS buffer. Conditions: $[\text{Fe}(\text{ClO}_4)_2]_0 = 0.008 - 0.015 \text{ mM}$, $[\text{MOPS}] = 0.5 - 0.6 \text{ mM}$, $25 \text{ }^\circ\text{C}$. pH decreased during the reaction by 0.2 units, see text.

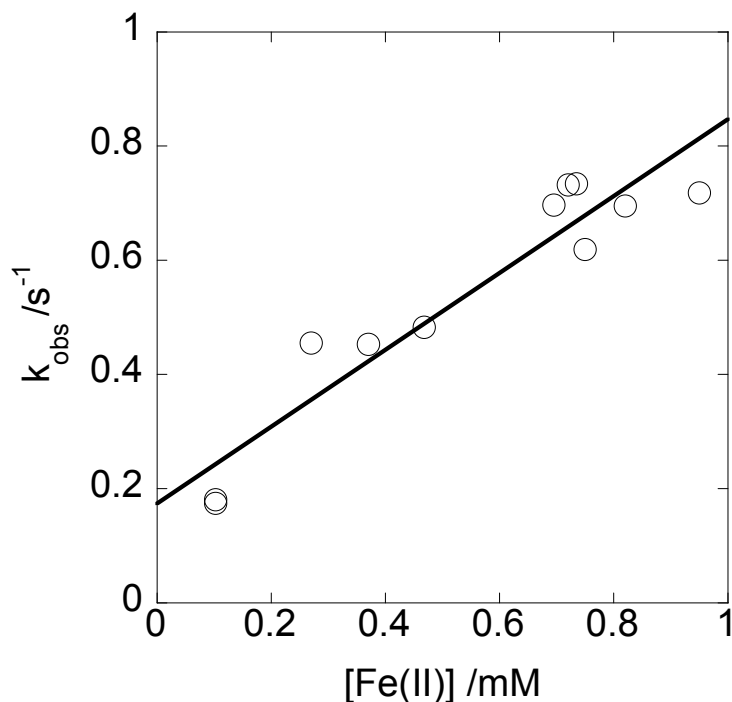


Figure S11. Plot of k_{obs} vs $[\text{Fe(II)}]$ for the $\text{Fe(II)}/\text{H}_2\text{O}_2$ reaction at pH 6.1 in a MES buffer. Conditions: $[\text{H}_2\text{O}_2]_0 = 0.005 - 0.02$ mM, $[\text{MES}] = 0.5\text{-}0.6$ mM, 25 °C. pH decreased during the reaction by 0.2 units, see text.

Comment on Figure S11. The scatter of the points in Figure S11 is larger than in experiments with excess H_2O_2 (Figs S9 - S10). The plot also exhibits an apparent intercept, but overall the data are in agreement with those obtained with excess H_2O_2 in that the slope of the line in Figure S11 ($k_{\text{Fe}} = 670 \pm 70$ $\text{M}^{-1}\text{s}^{-1}$) is comparable to that obtained by plotting k_{obs} against $2[\text{H}_2\text{O}_2]$ in Figure S10 ($k_{\text{Fe}} = 652 \pm 19$ $\text{M}^{-1}\text{s}^{-1}$) for experiments using excess H_2O_2 . These results are as expected for the 2:1 $[\text{Fe(II)}]/[\text{H}_2\text{O}_2]$ stoichiometry that was also calculated directly from the observed absorbance changes in these experiments. All of the experiments on catalytic oxidation of sulfoxides required (by definition) excess H_2O_2 , conditions that exhibited clean kinetic behavior as shown in Figures S9 and S10.

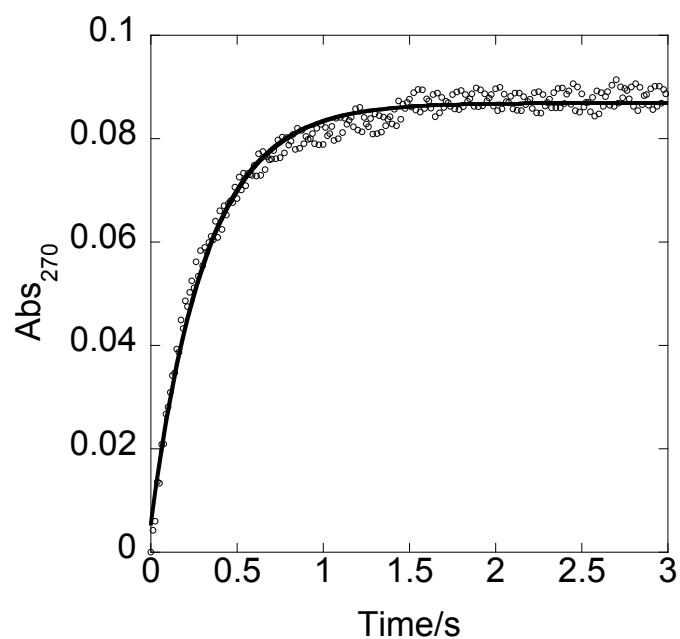


Figure S12. A representative kinetic trace in phosphate buffer. Conditions: 0.031 mM Fe(II), 1.0 mM H₂O₂, pH 6.1 (0.6 mM phosphate buffer)

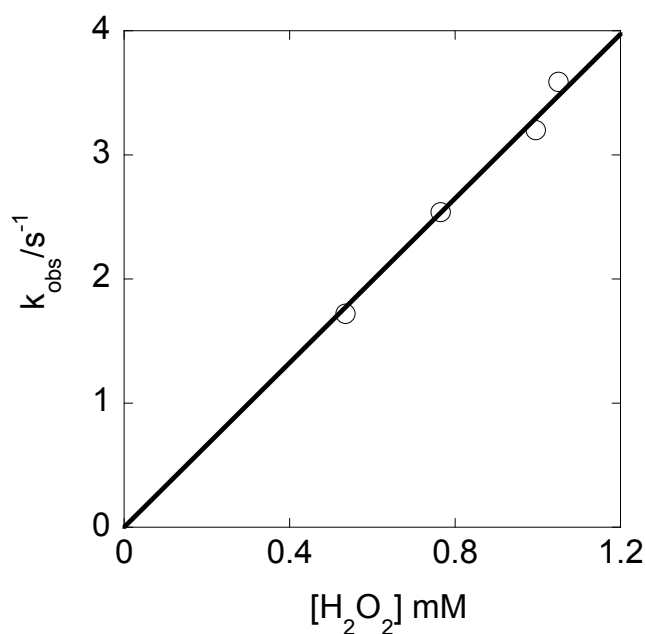


Figure S13. Plot of pseudo-first-order rate constants for the reaction of Fe(ClO₄)₂ with H₂O₂ against the concentration of H₂O₂ in phosphate buffer. Conditions: [Fe(ClO₄)₂] = 0.018 – 0.030 mM, [phosphate] = 0.60 mM, pH = 6.1-6.2.

Comment on noninterference of buffers.

Despite the large rate constant reported for the reaction of HO[•] with tertiary amines $k = (2-3) \times 10^9 \text{ M}^{-1}\text{s}^{-1}$,^{S5} this reaction cannot be important under our conditions even if HO[•] radicals were involved. This is so because the rate constant for the (CH₃)₂SO/HO[•] reaction is even larger, $k_{\text{DMSO}} = 7 \times 10^9 \text{ M}^{-1}\text{s}^{-1}$, and the concentration of (CH₃)₂SO was also always much higher (and never less than four times higher) than the buffer concentration, so that inequality $k_{\text{DMSO}}[(\text{CH}_3)_2\text{SO}] \gg k_{\text{buffer}}[\text{buffer}]$ held true in every experiment.

In acidic solutions, where the Fenton reaction generates HO[•], we observed no change in the products of Fe(H₂O)₆²⁺/H₂O₂/(CH₃)₂SO reaction in the presence of added MES, as shown in Figure S14a [In addition to sulfinic acid and ethane, the reaction in acidic solutions also produces small amounts of the sulfone in a minor parallel path that does not involve Fe(IV)].^{S6} The lack of reactivity at nitrogen in acidic solutions is caused by protonation, but the rate constants for hydrogen atom abstraction from C-H bonds are not expected to change significantly between pH 1 and pH 6. The addition of H₂O₂ to MES in the absence of Fe(ClO₄)₂ also had no effect on the NMR spectrum of MES (Figure S14b), as expected on the basis of literature data.^{S7}

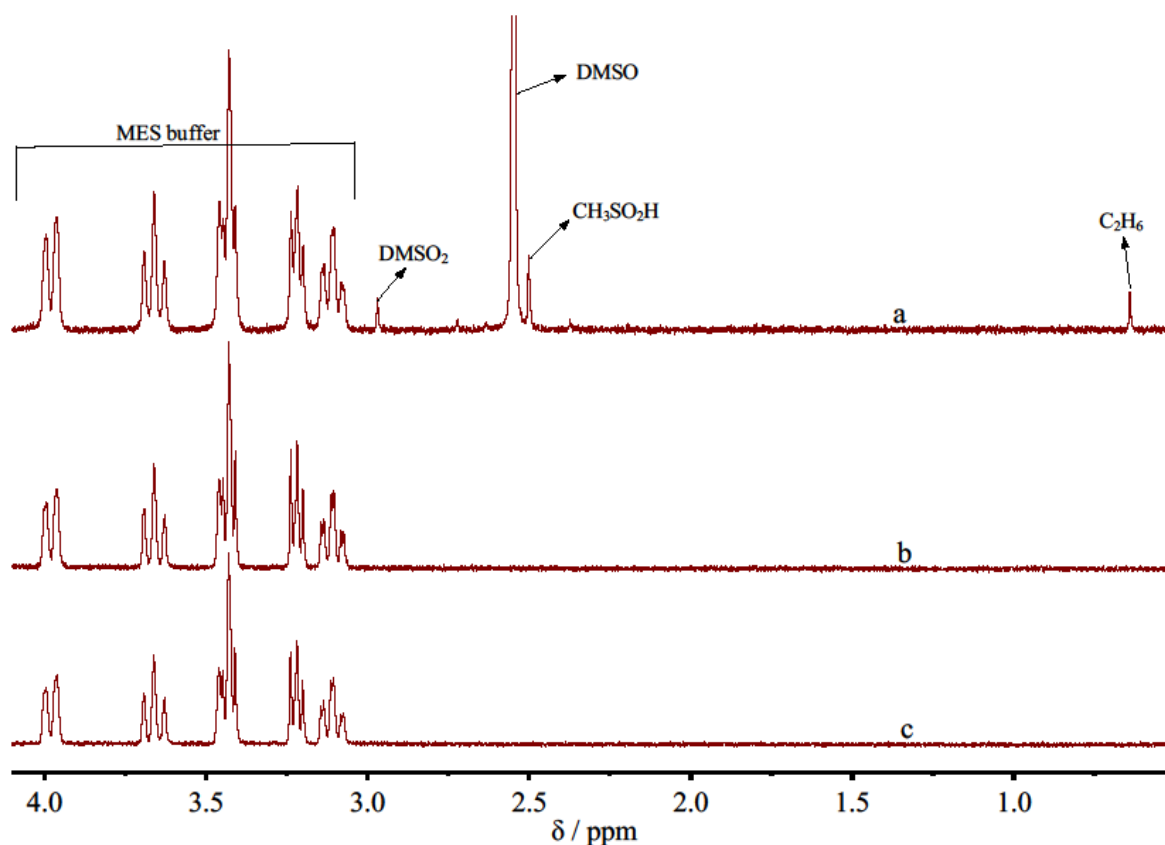


Figure S14. (a) ¹H NMR spectrum of the reaction mixture after the completion of the reaction between Fe(ClO₄)₂ (1.0 mM), H₂O₂ (1.0 mM), and (CH₃)₂SO (5 mM) at pD 1 in the presence of 5 mM MES. The reaction generated CH₃SO₂H and C₂H₆ in amounts comparable to those obtained in the absence of MES. b) Mixture of MES (5 mM) and H₂O₂ (1 mM), pD 1. c) MES (5 mM), pD 1.

We also ruled out any reaction between the buffers and Fe(IV), the actual intermediate in the Fenton reaction at pH ≥ 6, by observing that the change in buffer concentration (MES, 10 mM - 24 mM, pD 6.7) under standard conditions (1.2 mM Fe(II), 1.1 mM H₂O₂, 36 mM (CH₃)₂SO) had no effect on the yield of sulfone.

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