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# Pharmacokinetic and Pharmacodynamic Properties of Calaspargase Pegol *Escherichia coli* L-Asparaginase in the Treatment of Patients With Acute Lymphoblastic Leukemia: Results From Children's Oncology Group Study AALL07P4

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### Purpose

Asparaginase is a critical agent used to treat acute lymphoblastic leukemia (ALL). Pegaspargase (SS-PEG), a pegylated form of *Escherichia coli* L-asparaginase with a succinimidyl succinate (SS) linker, is the first-line asparaginase product used in Children's Oncology Group (COG) ALL trials. Calaspargase pegol (SC-PEG) replaces the SS linker in SS-PEG with a succinimidyl carbamate linker, creating a more stable molecule. COG AALL07P4 was designed to determine the pharmacokinetic and pharmacodynamic comparability of SC-PEG to SS-PEG in patients with newly diagnosed high-risk (HR) B-cell ALL.

### **Patients and Methods**

A total of 165 evaluable patients were randomly assigned at a 2:1 ratio to receive SC-PEG at 2,100 (SC-PEG2100; n = 69) or 2,500 IU/m<sup>2</sup> (SC-PEG2500; n = 42) versus SS-PEG 2,500 IU/m<sup>2</sup> (SS-PEG2500; n = 54) as part of an otherwise identical chemotherapy regimen. The groups were similar demographically, except more female patients received SC-PEG2500.

### Results

The mean half-life of plasma asparaginase activity for both SC-PEG doses was approximately  $2.5 \times$  longer than that of SS-PEG2500. The total systemic exposure, as defined by induction area under the curve from time 0 to 25 days, was greater with SC-PEG2500 than with SS-PEG2500 or SC-PEG2100. The proportion of patients with plasma asparaginase activity  $\geq$  100 mIU/mL and  $\geq$  400 mIU/mL was higher in patients who received SC-PEG as compared with SS-PEG2500. After one dose of pegylated asparaginase on induction day 4, plasma asparagine was undetectable for 11 days for SS-PEG2500 and 18 days for both SC-PEG groups.

### Conclusion

SC-PEG2500 achieves a significantly longer period of asparaginase activity above defined thresholds and asparagine depletion compared with SS-PEG2500 and has a comparable toxicity profile in children with HR B-cell ALL.

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# INTRODUCTION

Asparaginase is a critical agent in the treatment of acute lymphoblastic leukemia (ALL). This enzyme deaminates asparagine and glutamine, interfering with protein synthesis and resulting in cell death as lymphoblasts are deficient in asparagine synthetase.<sup>1,2</sup> Native *Escherichia coli* (*E coli*) L-asparaginase and a pegylated version thereof, pegaspargase (SS-PEG), are major components of ALL treatment regimens. SS-PEG is currently the first-line asparaginase preparation used in Children's Oncology Group (COG) ALL trials.

Calaspargase pegol (SC-PEG) is a newly developed form of pegylated asparaginase that uses the identical enzyme and polyethylene glycol moiety present in SS-PEG; however, SC-PEG uses a succinimidyl carbonate linker that is more stable than the SS-PEG succinimidyl succinate linker. Specifically, SC-PEG urethane linkages formed with lysine groups are more hydrolytically stable.<sup>3</sup> SC-PEG and SS-PEG have comparable pharmacokinetic (PK) and pharmacodynamics (PD) properties in preclinical studies.<sup>3</sup>

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COG AALL07P4 was designed to assess the PK and PD comparability<sup>4</sup> of SC-PEG to SS-PEG when administered intravenously during induction and consolidation therapy in newly diagnosed patients with National Cancer Institute–designated high-risk (HR) B-cell ALL receiving an otherwise identical COG augmented Berlin-Frankfurt-Münster (aBFM) –based chemotherapy regimen.

# **PATIENTS AND METHODS**

### Patients

Eligible patients (age 1 to 30.99 years) had newly diagnosed HR B-cell ALL (age  $\geq$  10 years and/or initial WBC count  $\geq$  50,000/microliter).<sup>5</sup> Key

gimens at Time of Study Initiation
Dose
60 mg/m <sup>2</sup> per day orally on days 1 to 28
IT $\times$ 1 on days -2 to 1
1.5 mg/m <sup>2</sup> (maximum, 2 mg) per dose on days 1, 8, 15, and 22
25 mg/m <sup>2</sup> per dose on days 1, 8, 15, and 22
IT on days 8, 15,° 22,° and 29
2,500 IU/m <sup>2</sup> per dose IV on day 4
60 mg/m <sup>2</sup> per day orally on days 1 to 14
1.5 mg/m <sup>2</sup> (maximum, 2 mg) per dose on days 1 and 8
25 mg/m <sup>2</sup> per dose on day 1
2,500 IU/m <sup>2</sup> per day IV on day 4
1,000 mg/m <sup>2</sup> per day IV on days 1 and 29
75 mg/m <sup>2</sup> per day SQ IV on days 1 to 4, 8 to 11, 29 to 32, and 36 to 39
60 mg/m <sup>2</sup> per day orally on days 1 to 14 and 29 to 42
IT on days 1, 8, 15, <sup>f</sup> and 22 <sup>f</sup>
2,500 IU/m <sup>2</sup> per day IV on days 15 and 43
1.5 mg/m <sup>2</sup> (maximum, 2 mg) per dose on days 15, 22, 43, and 50
1.5 mg/m <sup>2</sup> (maximum, 2 mg) per dose IV on days 1, 11, 21, 31, and 41
100 mg/m <sup>2</sup> per day IV on days 1, 11, 21, 31, and 41 (escalate by 50 mg/m <sup>2</sup> per dose)
2,500 IU/m <sup>2</sup> per day IM on days 2 and 22
IT on days 1 and 31
10 mg/m <sup>2</sup> per day orally on days 1 to 7 and 15 to 21
1.5 mg/m² (maximum, 2 mg) per dose IV on days 1, 8, 15, 43, and 50
25 mg/m <sup>2</sup> per day IV on days 1, 8, and 15
2,500 IU/m <sup>2</sup> per day IV on days 4 and 43
IT on days 1, 29, and 36
1,000 mg/m <sup>2</sup> per day IV on day 29
60 mg/m <sup>2</sup> per day orally on days 29 to 42
75 mg/m <sup>2</sup> per day SQ or IV on days 29 to 32 and 36 to 39
Starting dose of IV methotrexate 50 mg/m <sup>2</sup> < MTD in interim maintenance I, with same escalation rules
-
1.5 mg/m <sup>2</sup> per day IV on days 1, 29, and 57
40 mg/m <sup>2</sup> per day orally on days 1 to 5, 29 to 33, and 57 to 61
75 mg/m <sup>2</sup> per day orally on days 1 to 84
20 mg/m <sup>2</sup> per day orally on days 8, 15, 22, 29, <sup>1</sup> 36, 43, 50, 57, 64, 71, and 78
IT on day 1 (and day 29 in cycles one to four for RER patients)
disease; MTD, maximum-tolerated dose; PEG, pegylated; RER, rapid early responder; usly; SS-PEG, pegaspargase. t to 2.99 years, 50 mg; and age $\geq$ 3 years, 70 mg. to 2.99 years, 10 mg; age 3 to 8.99 years, 12 mg; and age $\geq$ 9 years, 15 mg. one marrow or $\geq$ 1% MRD. gnosis received 18 Gy to cranial midplane in 10 fractions. Patients classified as SERs received 12 tion II; day-36 IT methotrexate and day-29 to -42 thioguanine were omitted.

exclusion criteria included patients with Down syndrome, testicular leukemia, prior cytotoxic chemotherapy, pregnancy, and breastfeeding women.

Institutional review board approval was obtained at each participating institution before patient enrollment. Written informed consent and assent (where appropriate) were obtained from each patient and/or his or her parent or guardian before initiation of therapy.

### Study Design

This pilot multicenter, open-label, randomized trial opened to accrual in July 2008 at 23 COG centers. All patients received a modified COG aBFM chemotherapy regimen based on Children's Cancer Group 1961 arm C,<sup>6</sup> with the asparaginase preparation assigned at study enrollment. Patients were initially randomly assigned at a ratio of 2:1 to SC-PEG 2,500 IU/m<sup>2</sup> (SC-PEG2500; calaspargase pegol [EZN-2285]) versus SS-PEG (SS-PEG2500; pegaspargase). The regimens included a total of seven, 11, or 12 scheduled doses, depending on rapid early responders (RERs), slow early responders (SERs), and extended induction status, respectively, in the first 7 to 11 months of therapy (Table 1).

The primary end point was to determine the PK comparability<sup>4</sup> of SC-PEG and SS-PEG administered intravenously during induction and consolidation. Secondary end points included: safety, serum and CSF asparagine levels, immunogenicity, end-induction minimal residual disease (MRD), percentage of patients who were RERs, and complete remission and event-free survival rates. MRD was determined by multiparameter flow cytometry performed in a central laboratory.<sup>7,8</sup> A cutoff of < 0.1% was used for treatment stratification; however, for outcome analysis, positive MRD was defined as  $\geq$  0.01%, because multivariable analyses found this to be the most important prognostic variable in other COG trials.<sup>9</sup> Patients with < 5% blasts by morphologic bone marrow analysis on day 8 or 15 and day 29 and MRD < 0.1%were RERs; all others were SERs. The area under the curve (AUC) of asparaginase was based on plasma asparaginase activity from time 0 to 25 days  $(\mathrm{AUC}_{\text{0-25 days}})$  after induction day-4 dose of study drug (mIU  $\times$  hr/mL) and was calculated using linear trapezoidal summation. The PK-evaluable analysis population was defined as patients who received 94% to 106% of the induction day-4 asparaginase dose and had samples available to calculate AUC<sub>0-25 days</sub>. Additional PK parameters examined the percentage of patients with asparaginase activity  $\geq$  100 and  $\geq$  400 mIU/mL at specified time points. After planned interim PK analysis, the trial was amended in September 2009 to randomly assign patients between SC-PEG 2100 IU/m<sup>2</sup> (SC-PEG2100) versus SS-PEG2500, because this dose was predicted to achieve comparability of AUC<sub>0-25 days</sub>. In December 2010, after data safety monitoring committee review of MRD suggested inferior results with SC-PEG2100, having crossed predefined response monitoring boundaries, the trial was closed to accrual.

Patients in the SC-PEG2100 arm were nonrandomly changed to SS-PEG2500 for the duration of therapy. In January 2011, results of COG AALL0232, conducted in an identical risk group of patients with ALL, demonstrated the superiority of high-dose methotrexate over Capizzi methotrexate in interim maintenance,<sup>10</sup> and the protocol was amended to incorporate this change. Adverse events (AEs) were graded according to the National Cancer Institute Common Terminology Criteria for Adverse Events (CTCAE; version 3.0) before July 2011 and converted into CTCAE (version 4.0) codes, with subsequent AEs collected in CTCAE (version 4.0).

### Laboratory Tests

PKs, PDs, and immunogenicity were performed at central laboratories (Frontage Laboratories, Malvern, PA, and Commonwealth Biotechnologies, Richmond, VA) at specified time points. PKs were determined by a validated enzymatic coupled activity–modified assay.<sup>11</sup> PDs were determined using validated reverse-phase high-performance liquid chromatography with double mass spectrometry. Immunogenicity assessment included the detection of both binding antibodies and neutralizing antibodies, determined by a validated direct enzyme-linked immunosorbent assay and an enzymatic coupled activity assay, respectively (Data Supplement).

### Statistical Analysis

Clinical data frozen as of September 30, 2012, are included in this report. This report focuses on PK, PD, and AE data up to the end of delayed intensification I, and all patients were past that point in therapy as of that date. All analyses of clinical data were performed by intention to treat using validated SAS software (version 9.2; SAS Institute, Cary, NC).  $\chi^2$  and Fisher's exact tests were used for comparison of proportions between the three randomly assigned cohorts. All continuous data were summarized using medians and were compared among the three cohorts using the nonparametric Kruskal-Wallis test. The PK and PD data were computed from individual asparaginase activity and asparagine concentrations by applying a noncompartmental model approach using Phoenix WINNonlin software (version 6.1; Scientific Consultant, Apex, NC; Pharsight Corporation, St Louis, MO). For maximum plasma concentration ( $C_{max}$ ) and AUC values (AUC<sub>0-t</sub>, AUC<sub>0-25 days</sub>, and AUC<sub>0</sub>.), 90% CIs were calculated. For other parameters, the differences were described descriptively.

# RESULTS

### **Patient Characteristics**

Between October 2008 and December 2010, 166 patients were enrolled and randomly assigned (Fig 1). One patient was inevaluable



Fig 1. CONSORT diagram for Children's Oncology Group AALL07P4. PK, pharmacokinetic; SC-PEG, calaspargase pegol; SS-PEG, pegaspargase.

1	Table 2. Demographic a	nd Clinical Chara	acteristics of Eligible,	Evaluable Patien	ts (N = 165)				
	Regimen								
	SS-PEG 2,50 (n = 5	0 IU/m <sup>2</sup> 4)	SC-PEG 2,50 (n = 4	00 IU/m <sup>2</sup> 12)	SC-PEG 2,100 IU/m <sup>2</sup> (n = 69)				
Characteristic	No.	%	No.	%	No.	%	Р		
Age at diagnosis, years							.13		
< 10	18	33	18	43	19	28			
10 to 15	28	52	19	45	30	43			
≥ 16	8	15	5	12	20	29			
Sex							.04		
Male	31	57	14	33	38	55			
Female	23	43	28	67	31	45			
Race							.55		
White	42	78	35	83	56	81			
African American	6	11	1	2	4	6			
Other	6	11	6	15	9	13			
Ethnicity							.55		
Non-Hispanic	34	63	28	67	53	77			
Hispanic	18	33	12	28	14	20			
Unknown	2	4	2	5	2	3			
CNS status							.20		
1	46	85	36	86	50	72			
2	8	15	5	12	15	22			
3	0	0	1	2	4	6			
Initial WBC, $ imes$ 1,000/ $\mu$ L							.46		
< 50	32	59	20	48	35	51			
≥ 50	22	41	22	52	34	49			
BSA at study entry, m <sup>2</sup>							.23		
Median	1.3		1.2		1.4				
Range	0.5-2.	9	0.5-2	.4	0.4-2.	8			
MRD day 29									
Percent negative (< 0.01%)	36 of 50	72	29 of 39	74	36 of 64	56	.10		
Percent negative (< 0.1%)	39 of 50	78	33 of 39	85	44 of 64	69	.18		
Early response							.15		
RER	38 of 50	76	32 of 39	82	41 of 63	65			
SER	12 of 50	24	7 of 39	18	22 of 63	35			

Abbreviations: BSA, body-surface area; MRD, minimal residual disease; RER, rapid early responder; SC-PEG, calaspargase pegol; SER, slow early responder; SS-PEG, pegaspargase.

because of a major protocol violation and was excluded. Three patients discontinued the trial before receiving the randomly assigned study drug, and 162 received treatment with the randomly assigned study drug. Demographic characteristics (Table 2) of the three groups were similar, except for a higher percentage of female patients in the SC-PEG2500 arm as compared with the SC-PEG2100 and SS-PEG arms (67% v 45% and 43%, respectively; P = .04).

# PKs

Mean plasma asparaginase activity versus asparagine concentration over time during induction and consolidation therapies is shown in Figure 2. After completion of the induction day-4 intravenous dose of asparaginase, high levels of plasma asparaginase activity were observed for all treatment arms 5 minutes after infusion. Mean plasma asparaginase activity in all arms decreased based on the respective half-lives ( $t_{1/2}$ ; Table 3). By 18 days after administration, patients treated with SC-PEG maintained higher levels of asparaginase activity when compared with SS-PEG–treated patients (Fig 2A). Before the consolidation day-15 dose, which is at least 46 days after the induction day-4 asparaginase dose, 20% to 30% of patients in both SC-PEG groups had as paraginase activity  $\geq$  100 mIU/mL, as compared with none in the SS-PEG group.

Similar to induction, after the day-15 consolidation dose (Fig 2B), high levels of plasma asparaginase activity were observed for all treatment arms at 5 minutes afater infusion; these decreased based on their respective  $t_{1/2}$  (Table 3). Twenty-one days after administration, patients treated with SC-PEG maintained higher levels of asparaginase activity when compared with SS-PEG-treated patients, and at 28 days after dosing, mean SS-PEG plasma asparaginase activity was below the therapeutic threshold of 100 mIU/mL, whereas those in the SC-PEG2100 and SC-PEG2500 groups had mean values of 333.8 and 412.9 mIU/mL, respectively.

PK results for the PK-evaluable patients for induction and consolidation are summarized in Table 3. Mean  $t_{1/2}$  of plasma asparaginase activity for both the SC-PEG doses was approximately  $2.5 \times$ longer than that of SS-PEG2500 in induction and consolidation.

When administered at the 2,500 IU/m<sup>2</sup> dose, the total systemic exposure to SC-PEG, as defined by induction  $AUC_{0-25 \text{ days}}$  (404 IU × h/mL) and  $AUC_{0-inf}$  (574 IU × h/mL), was greater than that observed with SS-PEG2500 (365 and 387 IU × h/mL, respectively). For



**Fig 2.** Mean plasma asparaginase activity versus asparagine concentration by treatment group over time during (A) induction (IND) and (B) consolidation (CON), and (C) mean CSF asparagine concentration by treatment group over time during IND and CON. SC-PEG, calaspargase pegol; SS-PEG, pegaspargase.

SC-PEG2500 compared with SS-PEG2500, the ratio of geometric means and corresponding 90% CIs for AUC<sub>0-25 days</sub> was 111.0% (90% CI, 100.7% to 122.3%). This difference in total systemic exposure as defined by Induction AUC<sub>0-25 days</sub> was not seen with SC-PEG2100 group. A similar pattern of  $C_{max}$ ,  $t_{1/2}$ , systemic exposure, and elimination constant was also observed during consolidation (Table 3).

Plasma asparaginase activity presented as the percentage of PKevaluable patients with asparaginase activity  $\geq 100$  and  $\geq 400$  mIU/mL by treatment group during induction and consolidation is shown in Figure 3. During induction, SS-PEG2500 and both doses of SC-PEG achieved comparable results in the proportion of patients who maintained asparaginase activity levels  $\geq 100$  mIU/mL through 18 days after the induction dose. Twenty-five days after the induction dose, both doses of SC-PEG maintained asparaginase activity levels  $\geq 100$  mIU/mL in 95% of patients, as compared with only 28.6% of patients receiving SS-PEG2500 (Fig 3A; P < .001). By the time of the first consolidation dose of asparaginase (approximately 46 days after induction dose), approximately one quarter of the patients receiving SC-PEG continued to have asparaginase activity levels  $\geq$  100 mIU/ mL, whereas no patients in the SS-PEG2500 arm did (P = .002).

Using an asparaginase activity threshold level of  $\geq$  400 mIU/ mL, both SC-PEG2500 and SS-PEG2500 achieved comparable results in the proportion of patients who maintained this level through 11 days, with SC-PEG2100 being slightly lower. After that time point, the proportion of patients maintaining a level of asparaginase activity  $\geq$  400 mIU/mL was higher for both doses of SC-PEG compared with SS-PEG2500. By the end of induction, none of the SS-PEG2500 patients maintained asparaginase activity level  $\geq$  400 mIU/mL, compared with 27.5% in the SC-PEG2500 group and 13.6% in the SC-PEG2100 group (P < .001). During consolidation (Fig 3B), results were observed similar to those seen during the induction phase.

### PDs

There was a direct correlation between the reduction in plasma asparagine and plasma asparaginase activity during induction and consolidation (Figs 2A and 2B). Plasma asparagine was completely reduced with higher levels of asparaginase activity and began to rebound once plasma asparaginase activity declined < 400 mIU/mL. Thus, the longer  $t_{1/2}$  of SC-PEG resulted in a prolonged suppression of plasma asparagine.

Baseline mean plasma asparagine concentrations were similar for all three groups. After the induction dose, plasma asparagine concentrations decreased and were not detectable ( $< 0.05 \ \mu g/mL$ ) for all treatment groups at the 5 minutes postdose time point and remained undetectable for 11 days for SS-PEG2500 and 18 days for both SC-PEG groups. Twenty-five days after administration, plasma asparagine levels were undetectable in some patients in all three groups (88%, 95%, and 96% in SS-PEG2500, SC-PEG2500, and SC-PEG2100 groups, respectively). After this time point, the rate of plasma asparagine rise was greatest in the SS-PEG group.

Before consolidation day-15 dosing, the residual plasma asparagine level in patients in the SS-PEG2500 group was 5.33  $\mu$ g/mL (72.4% of preinduction baseline, with only 3% of patients < lower limit of detection [LLD]). By comparison, the residual plasma asparagine levels in patients treated with SC-PEG2500 and SC-PEG2100 were 0.64 (7.6% of preinduction baseline, with 72% of patients < LLD) and 0.88  $\mu$ g/mL (11.8% of preinduction baseline, with 76% of patients < LLD), respectively. Similar levels were observed after the consolidation dose.

## **CSF** Asparagine Concentration

Mean CSF asparagine concentration by treatment group during induction and consolidation is shown in Figure 2C. All three treatment regimens were effective in reducing CSF asparagine levels. Four days after the induction dose, mean CSF asparagine concentration in all treatment groups was diminished to approximately 25% to 30% of the preinduction dose values. Mean CSF asparagine levels returned to prebaseline levels by the time of the consolidation dose on day 15 in SS-PEG2500–treated patients, which was earlier than occurred for SC-PEG–treated patients (60% of baseline in SC-PEG2500 group; 40% of baseline in SC-PEG2100 group).

	1	<b>fable 3.</b> PK	(Paramete	rs During	Induction an	d Consol	idation in Pł	K-Evaluab	le Populati	on		
			Indu	iction					C	Consolidation	n	
	SS-PEG IU/m² (n	2,500 = 43)*	SC-PEC IU/m <sup>2</sup> (r	6 2,500 n = 40)*	SC-PEG IU/m² (n	2,100 = 62)*	SS-PEG IU/m² (r	6 2,500 n = 30)*	SC-PE IU/m <sup>2</sup>	EG 2,500 (n = 21)*	SC-PEG IU/m <sup>2</sup> (n	6 2,100 1 = 30)*
Parameter	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
C <sub>max</sub> , mIU/mL	1,650	474	1,655	366	1,291	379	1,480†	292	1,610	424	1,280†	274
t <sub>max</sub> , hours	3.8	6.2	6.0	17.0	2.9	6.8	7.7†	11.8	5.1	11.2	7.5†	11.1
CL, mL/hr	9.1	5.0	6.4	4.8	7.9‡	6.6	7.8§	5.2	7.5	0.015	4.9§	0.0045
t <sub>1/2</sub> , hours	127	51	322	118	305‡	98	117§	49	356	133	416§	149
V <sub>ss</sub> , L	2.0	1.2	2.7	2.02	3.1‡	2.0	1.8§	1.4	2.4	2.4	2.5§	1.3
$\rm AUC_{0-25\ days},\ IU  imes hr/mL$	365	77	404	94.1	324	86.4	NA		NA		NA	
$\rm AUC_{0-inf},~IU  imes hr/mL$	387	85.8	574	15.9	454‡	144	441§	109	777	303	687§	197

Abbreviations: AUC, area under the curve; CL, clearance; C<sub>max</sub>, maximum plasma concentration; NA, not applicable; PK, pharmacokinetic; SC-PEG, calaspargase pegol; SD, standard deviation; SS-PEG, pegaspargase; t<sub>1/2</sub>, half-life; t<sub>max</sub>, time to maximum plasma concentration; V, volume. \*n indicates No. of PK-evaluable patients.

tn = 29.

‡n = 58.

§n = 24.

∥n = 17.

### Immunogenicity

Antiasparaginase binding antibodies were confirmed in eight patients: four in the SS-PEG2500 group, two in the SC-PEG2500 group, and two in the SC-PEG2100 group. None of these patients had positive neutralizing antibody assays; however, three patients treated with SS-PEG and one treated with SC-PEG2500 were noted to have



**Fig 3.** Plasma asparaginase activity presented as percentage of pharmacokinetically evaluable patients with asparaginase activity  $\geq$  100 and  $\geq$  400 mIU/mL by treatment group over time during (A) induction and (B) consolidation. SC-PEG, calaspargase pegol; SS-PEG, pegaspargase. (\*) P < .001. (†) P = .006. (‡) P = .008. (§) P = .002.

more rapid clearance of asparaginase activity compared with their treatment groups. Two of these eight patients (one in SC-PEG2500 group and one in SC-PEG2100 group) had positive binding antibodies in the preinduction dose sample and had no subsequent positive tests, and no effect on asparaginase activity was seen. One SC-PEG2100 patient was noted to have a neutralizing antibody to SC-PEG and a false-positive binding assay but was noted to have more rapid clearance of asparaginase activity. In the patients with positive binding antibodies, allergic or hypersensitivity reactions were reported in two of four SS-PEG2500 patients, one of two SC-PEG2500 patients, and no patients in the SC-PEG2100 group.

### Asparaginase-Specific AEs

Protocol-specified asparaginase-specific AEs (regardless of grade or attribution to study drug; allergic reactions, coagulopathy, hyperbilirubinemia, hyperglycemia, hyperlipidemia, ketoacidosis, pancreatitis, thrombosis, and CNS events [bleeding, thrombosis, or infarction]) are listed in Table 4. AEs were similar between the three different treatment arms, with the exception of hyperglycemia in induction and hyperbilirubinemia in delayed intensification I, which were higher for the SC-PEG groups than for the SS-PEG2500 group (P = .001 and .01, respectively). When restricted to grade 3 and 4 events, the incidence of hyperbilirubinemia in delayed intensification I was not significant (P = .71; Appendix Table A1, online only).

# **End-of-Induction Treatment Response**

The end-of-induction treatment responses are listed in Table 2. The rates of RERs and MRD negativity were similar in the SC-PEG2500 (82% and 74%, respectively) and SS-PEG2500 groups (76% and 72%, respectively). However, the SC-PEG2100 group had a lower rate of RERs and MRD negativity (65% and 56%, respectively) compared with the SC-PEG2500 treatment group (P = .15 and .18, respectively). Although these differences were not statistically significant, at the time of interim analysis, the available values had crossed predefined response monitoring boundaries, leading to suspension of enrollment and transition of all SC-PEG2100 patients to SS-PEG2500.

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Table 4. Incidence of Protocol-Specified Asparaginase-Specific Grade 1 to 4 AEs by Regimen and Course								
	SS-PEG 2,5	00 IU/m <sup>2</sup>	SC-PEG 2,5	00 IU/m <sup>2</sup>	SC-PEG 2,1	00 IU/m <sup>2</sup>		
AE and Course	No.	%	No.	%	No.	%	Р	
Allergic reaction*								
Induction	4 of 54	7.4	2 of 42	4.8	1 of 69	1.4	.26	
Consolidation	10 of 43	23.3	9 of 33	27.3	10 of 49	20.4	.77	
Interim maintenance I	0 of 39	0.0	0 of 29	0.0	1 of 46	2.1	1.00	
Delayed intensification I	0 of 38	0.0	0 of 26	0.0	2 of 45	4.4	.34	
Hyperbilirubinemia								
Induction	20 of 54	37.0	14 of 42	33.3	28 of 69	40.6	.75	
Consolidation	13 of 43	30.2	15 of 33	45.5	26 of 49	53.1	.09	
Interim maintenance I	13 of 39	33.3	8 of 29	27.6	19 of 46	41.3	.49	
Delayed intensification I	4 of 38	10.5	10 of 26	38.5	13 of 45	28.9	.01	
Hyperlipidemia								
Induction	4 of 54	7.4	4 of 42	9.5	2 of 69	2.9	.31	
Consolidation	3 of 43	7.0	1 of 33	3.0	1 of 49	2.0	.53	
Interim maintenance I	1 of 39	2.6	1 of 29	3.4	1 of 46	2.2	1.00	
Delayed intensification I	0 of 38	0.0	0 of 26	0.0	1 of 45	2.2	1.00	
Hyperglycemia								
Induction	25 of 54	46.3	34 of 42	81.0	47 of 69	68.1	.001	
Consolidation	20 of 43	46.5	14 of 33	42.4	22 of 49	44.9	.97	
Interim maintenance I	13 of 39	33.3	13 of 29	44.8	16 of 46	34.8	.60	
Delayed intensification I	14 of 38	36.8	16 of 26	61.5	20 of 45	44.4	.13	
CNS								
Induction	0 of 54	0.0	0 of 42	0.0	4 of 69	5.8	.09	
Consolidation	0 of 43	0.0	0 of 33	0.0	0 of 49	0.0	1.00	
Interim maintenance I	0 of 39	0.0	0 of 29	0.0	0 of 46	0.0	1.00	
Delayed intensification I	1 of 38	2.6	1 of 26	3.8	0 of 45	0.0	.34	
Pancreatitis								
Induction	2 of 54	3.7	3 of 42	7.1	4 of 69	5.8	.76	
Consolidation	3 of 43	7.0	2 of 33	6.1	5 of 49	10.2	.85	
Interim maintenance I	1 of 39	2.6	1 of 29	3.4	1 of 46	2.2	1.00	
Delayed intensification I	0 of 38	0.0	2 of 26	7.7	1 of 45	2.2	.25	
Thrombosis								
Induction	0 of 54	0.0	0 of 42	0.0	4 of 69	5.8	.09	
Consolidation	1 of 43	2.3	0 of 33	0.0	0 of 49	0.0	.61	
Interim maintenance I	0 of 39	0.0	0 of 29	0.0	0 of 46	0.0	1.00	
Delayed intensification I	0 of 38	0.0	0 of 26	0.0	1 of 45	2.2	1.00	
Prolongation of activated partial thromboplastin time								
Induction	3 of 54	5.6	3 of 42	7.1	9 of 69	13.0	.35	
Consolidation	3 of 43	7.0	3 of 33	9.1	4 of 49	8.2	.92	
Interim maintenance I	3 of 39	7.7	2 of 29	6.9	3 of 46	6.5	1.00	
Delaved intensification I	7 of 38	18.4	7 of 26	26.9	4 of 45	8.9	.13	
INR increase		-						
Induction	2 of 54	3.7	4 of 42	9.5	9 of 69	13.0	.23	
Consolidation	3 of 43	7.0	1 of 33	3.0	3 of 49	6.1	.79	
Interim maintenance I	1 of 39	2.6	0 of 29	0.0	1 of 46	2.2	1.00	
Delayed intensification I	0 of 38	0.0	1 of 26	3.8	3 of 45	6.7	.29	
-								

Abbreviations: AE, adverse event; INR, international normalized ratio; SC-PEG, calaspargase pegol; SS-PEG, pegaspargase. \*Allergic reactions limited to events attributed as possibly, probably, or definitely related to SS-PEG or SC-PEG treatment.

# DISCUSSION

L-asparaginase is an essential component of standard combination therapy for ALL, and most children with ALL treated in North America and Western Europe now receive SS-PEG.<sup>12</sup> This study investigated a new formulation of pegylated asparaginase conjugated with a more stable SC linker that also imparts improved drug product shelf life compared with SS-PEG.<sup>3</sup> Early studies in nonhuman primates as well as previously published reports have concluded that  $\geq 100 \text{ mIU/mL}$  is an appropriate target level of asparaginase activity associated with asparagine depletion when a native asparagine depletion can still be observed even with levels < 100 mIU/mL.<sup>17,18</sup> Other investigators have suggested that a higher level of asparaginase activity ( $\geq 400 \text{ mIU/mL}$ ) is needed to achieve optimal asparagine depletion.<sup>19</sup> Our

data suggest that asparaginase activity level  $\geq$  400 mIU/mL is associated with more significant asparagine depletion, which could translate to improved outcomes.

We found that SC-PEG2500 was comparable in PKs to SS-PEG2500 with regard to AUC<sub>0-25 days</sub> and C<sub>max</sub>, whereas SC-PEG2100 was comparable in PKs to SS-PEG2500 with regard to  $AUC_{0-25 \text{ days}}$ . The  $t_{1/2}$  of asparaginase activity for both doses of SC-PEG was more than twice that of SS-PEG2500. After a single infusion at day 4, both SC-PEG doses were effective in maintaining asparaginase activity  $\geq$  100 mIU/mL in > 95% of patients at the end of the 4-week induction phase, and 20% to 30% of patients still had levels above this threshold 3 weeks later (consolidation day 15). By comparison, < 30% of patients treated with SS-PEG2500 maintained asparaginase activity  $\geq 100 \text{ mIU/mL}$  at end of induction, and none had levels above this threshold at consolidation day 15. Similarly, patients treated with SC-PEG maintained asparaginase activity level  $\geq$  400 mIU/mL for longer than those who received SS-PEG2500. Both doses of SC-PEG were effective in reducing plasma asparagine levels below detectable levels for a longer duration than SS-PEG2500 during induction and consolidation.

End-of-induction complete remission rates, RERs, and MRD negativity were similar among the three groups. However, the SC-PEG2100 group trended toward inferior responses, with lower rates of RERs and MRD negativity (65% and 56%, respectively) compared with the SS-PEG and SC-PEG2500 groups. This difference crossed predefined statistical monitoring boundaries during conduct of the study, but ultimately, it was not statistically significant. The reason for this difference is unclear and may be an artifact resulting from small sample size or the higher number of older patients in the SC-PEG2100 group (P = .13). Superior responses might be explained by enhanced asparagine depletion having an effect on the clearance of leukemia blasts. There was a slightly higher percentage of patients age  $\geq 16$  years in the SC-PEG2100 group as compared with the other two groups, which did not reach conventional levels of statistical significance (P = .06), and older adolescents have been shown to have inferior end-of-induction MRD responses compared with younger patients with HR B-cell ALL,<sup>20</sup> but this likely does not fully explain the observed differences.

Few patients in all groups had confirmed antiasparaginase antibodies, even though clinical allergic or hypersensitivity reactions were reported at a higher rate. Half of patients with antiasparaginase antibodies did not have clinical allergic or hypersensitivity reactions reported, and numerous patients had allergic or hypersensitivity reactions reported but no antibodies detected. This may be because of assay limitations. Some of the reported allergic reactions may have actually been allergic or hypersensitivity reactions related to other medications or blood products or nonallergic infusion reactions.

The limitations of this study include small numbers of patients, the challenge of measuring plasma asparagine in the presence of ex vivo asparaginase activity, and possibly reporting bias in this openlabel trial. Although COG phase III studies do not require reporting of grade 1 or 2 AEs, for this trial, the US Food and Drug Administration mandated reporting of asparaginase-specific events of all grades regardless of attribution; thus, the rates attributed to asparaginase may have been overestimated and cannot be compared directly with those from other COG ALL trials.

COG AALL07P4 randomly assigned patients with newly diagnosed HR B-cell ALL at a 2:1 ratio to receive SC-PEG (2100 or 2500  $IU/m^2$ ) or SS-PEG (2,500 IU/m<sup>2</sup>) during aBFM multiagent chemotherapy. Because of its increased t<sub>1/2</sub>, SC-PEG2500 produced a significantly longer period of asparagine depletion than SS-PEG2500 and had a comparable toxicity profile in children with HR B-cell ALL. Additional studies are needed to determine whether the superior PK and PD properties of SC-PEG translate to improved treatment outcomes.

# AUTHORS' DISCLOSURES OF POTENTIAL CONFLICTS OF INTEREST

Although all authors completed the disclosure declaration, the following author(s) and/or an author's immediate family member(s) indicated a financial or other interest that is relevant to the subject matter under consideration in this article. Certain relationships marked with a "U" are those for which no compensation was received; those relationships marked with a "C" were compensated. For a detailed description of the disclosure categories, or for more information about ASCO's conflict of interest policy, please refer to the Author Disclosure Declaration and the Disclosures of Potential Conflicts of Interest section in Information for Contributors. Employment or Leadership Position: Taha Keilani, Sigma Tau Pharmaceuticals (C) Consultant or Advisory Role: Stephen P. Hunger, Sigma Tau Pharmaceuticals (C), Jazz Pharmaceuticals (C) Stock Ownership: None Honoraria: Stephen P. Hunger, Jazz Pharmaceuticals, Sigma Tau Pharmaceuticals Research Funding: Julie M. Gastier-Foster, Bristol-Myers Squibb Expert Testimony: None Patents, Royalties, and Licenses: None Other Remuneration: None

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# REFERENCES

1. Broome JD: Evidence that the L-asparaginase of guinea pig serum is responsible for its antilymphoma effects: II. Lymphoma 6C3HED cells cultured in a medium devoid of L-asparagine lose their susceptibility to the effects of guinea pig serum in vivo. J Exp Med 118:121-148, 1963

**2.** Broome JD: Evidence that the L-asparaginase of guinea pig serum is responsible for its antilymphoma effects: I. Properties of the L-asparaginase

of guinea pig serum in relation to those of the antilymphoma substance. J Exp Med 118:99-120, 1963

**3.** Investigator brochure for EZN-2285 (SC-PEG *E coli* L-asparaginase), version 1. Sigma Tau Pharmaceuticals, Gaithersburg, MD, 2007

### Angiolillo et al

4. US Department of Health and Human Services: Guidance for Industry: Statistical Approaches to Establishing Bioequivalence. http://www.fda.gov/downloads/Drugs/Guidances/ucm070244.pdf

5. Smith M, Arthur D, Camitta B, et al: Uniform approach to risk classification and treatment assignment for children with acute lymphoblastic leukemia. J Clin Oncol 14:18-24, 1996

 Seibel NL, Steinherz PG, Sather HN, et al: Early postinduction intensification therapy improves survival for children and adolescents with high-risk acute lymphoblastic leukemia: A report from the Children's Oncology Group. Blood 111:2548-2555, 2008

7. Borowitz MJ, Pullen DJ, Shuster JJ, et al: Minimal residual disease detection in childhood precursor-B-cell acute lymphoblastic leukemia: Relation to other risk factors. A Children's Oncology Group study. Leukemia 17:1566-1572, 2003

8. Weir EG, Borowitz MJ: Flow cytometry in the diagnosis of acute leukemia. Semin Hematol 38: 124-138, 2001

**9.** Borowitz MJ, Devidas M, Hunger SP, et al: Clinical significance of minimal residual disease in childhood acute lymphoblastic leukemia and its relationship to other prognostic factors: A Children's Oncology Group study. Blood 111:5477-5485, 2008 **10.** Larsen EC, Salzer WL, Devidas M, et al: Comparison of high-dose methotrexate (HD-MTX) with Capizzi methotrexate plus asparaginase (C-MTX/ASNase) in children and young adults with high-risk acute lymphoblastic leukemia (HR-ALL): A report from the Children's Oncology Group study AALL0232. J Clin Oncol 29:6s, 2011 (suppl; abstr 3)

**11.** Jayaram HN, Cooney DA, Jayaram S, et al: A simple and rapid method for the estimation of L-asparaginase in chromatographic and electrophoretic effluents: Comparison with other methods. Anal Biochem 59:327-346, 1974

12. Pui CH, Evans WE: Treatment of acute lymphoblastic leukemia. N Engl J Med 354:166-178, 2006

**13.** Ahlke E, Nowak-Göttl U, Schulze-Westhoff P, et al: Dose reduction of asparaginase under pharmacokinetic and pharmacodynamic control during induction therapy in children with acute lymphoblastic leukaemia. Br J Haematol 96:675-681, 1997

**14.** Avramis VI, Sencer S, Periclou AP, et al: A randomized comparison of native Escherichia coli asparaginase and polyethylene glycol conjugated asparaginase for treatment of children with newly diagnosed standard-risk acute lymphoblastic leukemia: A Children's Cancer Group study. Blood 99: 1986-1994, 2002

**15.** Boos J, Werber G, Ahlke E, et al: Monitoring of asparaginase activity and asparagine levels in children on different asparaginase preparations. Eur J Cancer 32A:1544-1550, 1996

**16.** Riccardi R, Holcenberg JS, Glaubiger DL, et al: L-asparaginase pharmacokinetics and asparagine levels in cerebrospinal fluid of rhesus monkeys and humans. Cancer Res 41:4554-4558, 1981

**17.** Klug Albertsen B, Schmiegelow K, Schrøder H, et al: Anti-Erwinia asparaginase antibodies during treatment of childhood acute lymphoblastic leukemia and their relationship to outcome: A case-control study. Cancer Chemother Pharmacol 50: 117-120, 2002

**18.** Vieira Pinheiro JP, Ahlke E, Nowak-Göttl U, et al: Pharmacokinetic dose adjustment of Erwinia asparaginase in protocol II of the paediatric ALL/ NHL-BFM treatment protocols. Br J Haematol 104: 313-320, 1999

19. Avramis VI, Panosyan EH: Pharmacokinetic/pharmacodynamic relationships of asparaginase formulations: The past, the present and recommendations for the future. Clin Pharmacokinet 44:367-393, 2005

**20.** Raetz EA, Devidas M, Carroll AJ, et al: Cytogenetic and early-response characteristics of adolescents and young adults with acute lymphoblastic leukemia (ALL): A Children's Oncology Group (COG) study. J Clin Oncol 28:680s, 2010 (suppl; abstr 9509)

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# **GLOSSARY TERMS**

**pharmacokinetics:** a branch of pharmacology that studies the relationship between drug exposure level, time course of exposure, and the overall response of an organism. Although pharmacokinetics is largely applied to drugs, it is also applicable to other compounds such as nutrients, toxins, hormones, etc. Pharmacokinetics is subdivided into absorption and disposition (distribution, metabolism, and excretion) and is generally referred to as ADME (absorption, distribution, metabolism, excretion). With respect to drugs administered, all processes occur in tandem once a drug dose is administered. In clinical trials, phase I studies will typically study pharmacokinetics and safety of the drug. **pharmacodynamics:** the study of the biochemical and physiologic effects of a drug on the body.

### PKs and PDs of SC-PEG E coli L-Asparaginase in ALL

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# Appendix

AE and Course	SS-PEG 2,5	00 IU/m <sup>2</sup>	SC-PEG 2,5	00 IU/m <sup>2</sup>	SC-PEG 2,10	00 IU/m <sup>2</sup>	
	No.	%	No.	%	No.	%	Р
Pancreatitis							
Induction	2 of 54	3.7	2 of 42	4.8	4 of 69	5.8	.90
Delayed intensification I	0 of 38	0.0	2 of 26	7.7	1 of 45	2.2	.25
Hyperlipidemia							
Induction	2 of 54	3.7	2 of 42	4.8	1 of 69	1.4	.62
Delayed intensification I	0 of 38	0.0	0 of 26	0.0	1 of 45	2.2	1.00
Hyperbilirubinemia							
Induction	4 of 54	7.4	5 of 42	11.9	6 of 69	8.7	.74
Delayed intensification I	0 of 38	0.0	1 of 26	3.9	1 of 45	2.2	.71
Hyperglycemia							
Induction	8 of 54	14.8	11 of 42	26.2	24 of 69	34.8	.04
Delayed intensification I	3 of 38	7.9	6 of 26	23.1	7 of 45	15.6	.24