

Phase I Safety, Pharmacokinetics, and Pharmacogenetics Study of the Antituberculosis Drug PA-824 with Concomitant Lopinavir-Ritonavir, Efavirenz, or Rifampin

Kelly E. Dooley,^a Anne F. Luetkemeyer,^b Jeong-Gun Park,^c Reena Allen,^d Yoninah Cramer,^c Stephen Murray,^e Deborah Sutherland,^f Francesca Aweeka,^b Susan L. Koletar,^g Florence Marzan,^b Jing Bao,^h Rada Savic,^b David W. Haas,^f the AIDS Clinical Trials Group A5306 Study Team

Johns Hopkins University School of Medicine, Baltimore, Maryland, USA^a; University of California, San Francisco, California, USA^b; Statistical and Data Analysis Center, Harvard School of Public Health, Boston, Massachusetts, USA^c; Social & Scientific Systems, Inc., Silver Spring, Maryland, USA^d; Global Alliance for TB Drug Development, New York, New York, USA^e; Vanderbilt University School of Medicine, Nashville, Tennessee, USA^f; Ohio State University, Columbus, Ohio, USA^g; Henry M. Jackson Foundation, Division of AIDS, National Institutes of Health, Bethesda, Maryland, USA^h

There is an urgent need for new antituberculosis (anti-TB) drugs, including agents that are safe and effective with concomitant antiretrovirals (ARV) and first-line TB drugs. PA-824 is a novel antituberculosis nitroimidazole in late-phase clinical development. Cytochrome P450 (CYP) 3A, which can be induced or inhibited by ARV and antituberculosis drugs, is a minor (~20%) metabolic pathway for PA-824. In a phase I clinical trial, we characterized interactions between PA-824 and efavirenz (arm 1), lopinavir/ritonavir (arm 2), and rifampin (arm 3) in healthy, HIV-uninfected volunteers without TB disease. Participants in arms 1 and 2 were randomized to receive drugs via sequence 1 (PA-824 alone, washout, ARV, and ARV plus PA-824) or sequence 2 (ARV, ARV with PA-824, washout, and PA-824 alone). In arm 3, participants received PA-824 and then rifampin and then both. Pharmacokinetic sampling occurred at the end of each dosing period. Fifty-two individuals participated. Compared to PA-824 alone, plasma PA-824 values (based on geometric mean ratios) for maximum concentration (C_{max}), area under the concentration-time curve from 0 to 24 h (AUC_{0-24}), and trough concentration (C_{min}) were reduced 28%, 35%, and 46% with efavirenz, 13%, 17%, and 21% with lopinavir-ritonavir (lopinavir/r) and 53%, 66%, and 85% with rifampin, respectively. Medications were well tolerated. In conclusion, lopinavir/r had minimal effect on PA-824 exposures, supporting PA-824 use with lopinavir/r without dose adjustment. PA-824 exposures, though, were reduced more than expected when given with efavirenz or rifampin. The clinical implications of these reductions will depend upon data from current clinical trials defining PA-824 concentration-effect relationships. (This study has been registered at ClinicalTrials.gov under registration no. NCT01571414.)

In 2012 there were 8.6 million cases of tuberculosis (TB) and 1.3 million tuberculosis-related deaths (1). Short-course treatment of drug-sensitive tuberculosis requires 6 months of therapy. Multidrug-resistant (MDR) tuberculosis (i.e., resistant to isoniazid and rifampin) is a growing public health threat, with therapeutic options limited by drug availability, acceptability, and efficacy (2). Current MDR-tuberculosis treatment requires ≥ 18 months of multidrug therapy with at least 6 months of an injectable agent (3), is poorly tolerated, and is successful in only 48% of patients (2). Almost one-third of tuberculosis-related deaths globally are in patients with HIV coinfection (1). There is an urgent need for novel antituberculosis regimens to shorten treatment duration for drug-sensitive tuberculosis and to improve the efficacy and safety for MDR tuberculosis. The utility of novel drugs will be significantly enhanced if they are safe and effective among patients who require anti-HIV therapy.

The investigational nitroimidazole PA-824 has potent *in vitro* activity against *Mycobacterium tuberculosis* and no cross-resistance with marketed antituberculosis drugs (4, 5). Its activity against metabolically active and nonreplicating *M. tuberculosis* (5) suggests likely bactericidal and sterilizing activity. The latter is critical for treatment shortening. In mouse models of tuberculosis, PA-824 given with rifampin and pyrazinamide reduced curative treatment duration from 6 months to 4 months (6). In mice, PA-824 with moxifloxacin and pyrazinamide was similarly potent; in humans, this same three-drug combination reduced spu-

tum mycobacterial colony counts more effectively than standard treatment over 2 weeks' time when PA-824 was given at a dose of 200 mg once daily (7, 8). A phase 2B trial of PA-824 over 8 weeks at doses of 100 mg and 200 mg with moxifloxacin and pyrazinamide finished recently (results pending). PA-824 has not been tested clinically in a combination with rifampin plus pyrazinamide, the key sterilizing drugs in antituberculosis therapy. It is not known whether PA-824 can reduce treatment duration when it is given with first-line or second-line antituberculosis drugs.

To include PA-824 in antituberculosis regimens in HIV-infected patients, the safety and pharmacokinetics (PK) of drug combinations must be assessed. PA-824 is extensively metabolized via a combination of reductive metabolism and oxidative metabolism with no one single metabolic path that can be considered major. *In vitro* studies suggest that cytochrome P450 (CYP) 3A

Received 12 May 2014 Returned for modification 8 June 2014

Accepted 15 June 2014

Published ahead of print 23 June 2014

Address correspondence to Kelly E. Dooley, kdooley1@jhmi.edu.

Supplemental material for this article may be found at <http://dx.doi.org/10.1128/AAC.03332-14>.

Copyright © 2014, American Society for Microbiology. All Rights Reserved.

doi:10.1128/AAC.03332-14

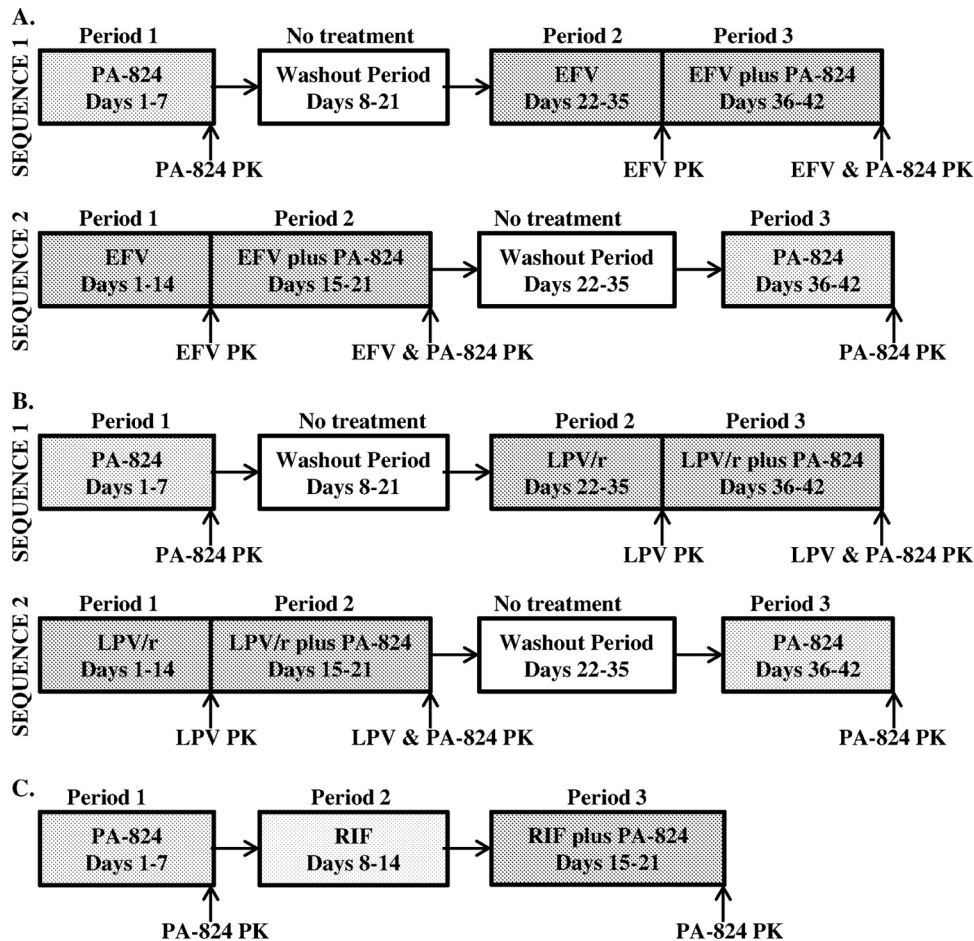


FIG 1 Schematic of the dosing regimen and pharmacokinetic sample collection: arm 1, PA-824 with efavirenz (EFV) (A); arm 2, PA-824 with lopinavir/ritonavir (LPV/r) (B); arm 3, PA-824 with rifampin (RIF) (C).

contributes up to 20% to overall metabolism; PA-824 is not a substrate of CYP2C9, -2C19, or -2D6 metabolizing enzymes (Stephen Murray, TB Alliance, personal communication). Rifampin induces many metabolizing enzymes including CYP3A (9). Efavirenz (EFV) is included in first-line regimens for HIV in many settings, and lopinavir (LPV; with ritonavir, lopinavir/r) is the most widely prescribed HIV-1 protease inhibitor globally. Efavirenz induces CYP3A enzymes, while lopinavir and ritonavir can inhibit or induce CYP3A (10). In addition, the *CYP2B6* genotype is a key determinant of efavirenz concentrations (11–15), while *SLCO1B1* polymorphisms impact lopinavir exposures (16–18). In this phase I trial, we investigated the safety and PK interactions of PA-824 with efavirenz, lopinavir/r, and rifampin, taking into account pharmacogenetics.

MATERIALS AND METHODS

Study population. Healthy adults 18 to 65 years were recruited at AIDS Clinical Trials Group (ACTG) sites in the United States. Eligible participants had negative HIV and hepatitis C antibody tests, normal alanine aminotransferase (ALT) levels, and creatinine clearance values of >50 ml/min. Volunteers were excluded for hemoglobin of ≤ 12.0 g/dl (male) or ≤ 11.0 g/dl (female), absolute neutrophil count of $<1,250$ cells/mm³, platelet counts of $<125,000$ cells/mm³, electrocardiogram (ECG) with a corrected QT (QTc) of >450 or PR of >200 ms, or active tuberculosis.

Frequent headaches was another exclusion criterion. The study was approved by institutional review boards of the participating sites. All participants provided written informed consent. ACTG study A5306 was registered at ClinicalTrials.gov under registration number NCT01571414.

Experimental protocol. (i) Study design of the phase I, open-label PK and safety study. PA-824 was dosed 200 mg once daily, efavirenz was given at 600 mg once daily, rifampin was dosed at 600 mg once daily, and lopinavir/r was given at 400/100 mg every 12 h. Efavirenz was taken in the evenings. PA-824 and rifampin were taken in the mornings. All medications were taken on an empty stomach. Participants were sequentially assigned to arm 1 (efavirenz), arm 2 (lopinavir/r), or arm 3 (rifampin) (Fig. 1). In arm 1, participants were randomized to sequence 1, consisting of PA-824 for 7 days, a 2-week washout period, efavirenz for 14 days, and then efavirenz with PA-824 for 7 days, or sequence 2, consisting of efavirenz for 14 days, efavirenz with PA-824 for 7 days, a 2-week washout period, and then PA-824 alone for 7 days. Arm 2 participants were randomized to two sequences as follows: sequence 1, consisting of PA-824 for 7 days, a 2-week washout period, lopinavir/r for 14 days, and then lopinavir/r with PA-824 for 7 days, or sequence 2, consisting of lopinavir/r for 14 days, lopinavir/r with PA-824 for 7 days, a two-week washout period, and PA-824 alone for 7 days. In arm 3, participants received PA-824 for 7 days, rifampin for 7 days, and then PA-824 with rifampin for 7 days. Adherence was assessed by pill counts and medication diaries. All doses prior to PK sampling were observed by study staff. Serial plasma sampling for PK was performed at the end of each dosing period for PA-824, efa-

virenz, and lopinavir. For PA-824, plasma was obtained predose and at 1, 2, 3, 4, 5, 6, 8, 10, 12, and 24 h postdose. For efavirenz, plasma was obtained predose and at 1, 2, 3, 4, 8, 12, and 24 h postdose. For lopinavir, plasma was obtained predose and at 1, 2, 3, 4, 5, 6, 8, 10, and 12 h postdose. Rifampin concentrations were not measured in this small PK study because the effects of PA-824 on rifampin PK were expected to be small. In addition, since rifampin concentrations are highly variable and since that variability is not well explained by known genetic polymorphisms, even large changes would be unlikely to be detected.

(ii) Safety monitoring. Participants underwent weekly safety evaluations. ECG evaluations were performed at baseline and on the final day of PA-824 dosing periods, given that some nitroimidazole antibiotics can cause QT prolongation. Adverse events were graded according to the Division of AIDS Table for Grading the Severity of Adult and Pediatric Adverse Events, version 1.0 (19).

Drug concentration analysis. (i) Plasma assay for PA-824. PA-824 and the internal standard, triazolam, were isolated from EDTA-plasma by liquid-liquid extraction. The organic phase was removed, transferred to a clean test tube, and evaporated under nitrogen. The residue was reconstituted in MeOH-water (1:1), transferred to autosampler vials for injection onto a Chromolith SpeedROD-18 high-performance liquid chromatograph (HPLC) column, and eluted with a linear gradient of 10 mM ammonium acetate and methanol (30:70). The ion pairs 359.8/174.7 for PA-824 and 342.8/307.7 for triazolam were selected for tandem mass detection in multiple reaction monitoring (MRM) mode. Quantification of PA-824 was performed with a liquid chromatography-tandem mass spectrometer (LC-MS/MS) system comprising two PerkinElmer series 200 micro-LC pumps and a series 200 autosampler coupled with an AB Sciex API 2000 tandem mass spectrometer. For calibration curves, spiked concentrations and peak area ratios of PA-824 and the internal standard were fitted by linear least-squares regression, weighted by $1/x$. The method was validated over a linear range of 10 to 10,000 ng/ml with a correlation value, R , of 0.9984. For the measurements of PA-824, the interassay precision (percent coefficient of variation [CV]) ranged from 2.65 to 4.71% and the percent deviation ranged from 0.8 to 5.2% of the nominal values of the control concentrations. The intra-assay precision (percent CV) ranged from 1.58 to 6.29%, and the percent deviation ranged from -0.40 to 11.19% of the nominal values of the control concentrations. The lower limit of quantification (LLOQ) and upper limit of quantification (ULOQ) were 10 and 10,000 ng/ml, respectively. Plasma aliquots of 50 μ l were sufficient for analysis.

(ii) Plasma assays for efavirenz and lopinavir. Efavirenz was quantified by reversed-phase HPLC following extraction from human plasma by simple protein precipitation. Detection involved a photodiode array detector, scanning at a wavelength of 247 nm, and reserpine as the internal standard. The calibration curve concentration range was 100 ng/ml to 6,000 ng/ml. For calibration curves, spiked concentrations and peak height ratios of efavirenz and the internal standard were fitted by linear least-squares regression, weighted $1/x$. Efavirenz concentrations were calculated from regression parameters using peak height ratios. The method was validated over a linear range of 100 to 10,000 ng/ml with a correlation value, R , of 0.9995. For the measurements of EFV, the interassay precision (percent CV) ranged from 2.4 to 4.5%, and the percent deviation ranged from -0.4 to 3.3% of the nominal values of the control concentrations. The intra-assay precision (percent CV) ranged from 0.6 to 5.4%, and the percent deviation ranged from -1.7 to 6.1% of the nominal values of the control concentrations. The lower limit of quantification (LLOQ) and upper limit of quantification (ULOQ) were 100 and 10,000 ng/ml, respectively.

Lopinavir and its deuterated internal standards were extracted from 50 μ l of EDTA-human plasma by protein precipitation with acetonitrile, followed by centrifugation. The clear supernatant was transferred into autosampler vials for a 10- μ l injection onto an Agilent Zorbax XDB-C₈ (5- μ m particle size; 2.1- by 50-mm HPLC column). The mobile phase comprised 10 mM ammonium formate buffer (pH 4.0) and acetonitrile

containing 0.1% formic acid. Elution was performed using a gradient flow rate of 400 μ l/minute with MS/MS detection on an ABCIEEX API 2000 mass spectrometer (MS) using electrospray in positive-ion mode. The ion pairs 629.2/429.0 for lopinavir and 637.2/429.0 for LPV-D⁸ were selected for tandem mass detection. For calibration curves, spiked concentrations using peak area ratios of lopinavir were fitted by $1/x$ linear regression. The method was validated over a linear range of 50 to 8,000 ng/ml with a correlation R of 0.9988. For the measurements of LPV, the interassay precision (percent CV) ranged from 4.50 to 5.11%, and the percent deviation ranged from -8.15 to 0.037% of the nominal values of the control concentrations. The intra-assay precision (percent CV) ranged from 2.35 to 6.39%, and the percent deviation ranged from -9.33 to 2.72% of the nominal values of the control concentrations. The lower limit of quantification (LLOQ) and upper limit of quantification (ULOQ) were 50 and 8,000 ng/ml, respectively.

Pharmacogenetic testing. Genetic polymorphisms reported to predict plasma PK of efavirenz (11–15), rifampin (20, 21), and lopinavir (17, 18) were genotyped in duplicate. For efavirenz, *CYP2B6* 516 G→T (rs3745274), 983 T→C (rs28399499), and 15582 C→T (rs4803419) were assayed using MassARRAY iPLEX Gold (Sequenom Inc., San Diego, CA, USA). The composite *CYP2B6* genotype was defined as follows (15): an extensive metabolizer contains 516 G/G and 983 T/T, with either 15582 C/C or C/T; an intermediate metabolizer contains 516 G/T or 983 T/C but not both, or homozygosity for 15582 T/T; and a slow metabolizer contains 516 T/T, 983 C/C, or the combination of 516 G/T and 983 T/C. *SLCO1B1* C→T (rs4149032) (for rifampin) and *SLCO1B1* 521 T→C (rs4149056) (for lopinavir/r) were genotyped by TaqMan (Applied Biosystems, Inc., Foster City, CA). The *CYP3A5**3 6986 A→G variant (rs776746) (22) was genotyped by MassARRAY iPLEX Gold.

Pharmacokinetic and statistical analyses. (i) Sample size. Thirteen volunteers per arm were estimated to provide 80% power to detect a 20% mean difference in the area under the concentration-time curve from 0 to 24 h (AUC_{0-24h}) for PA-824 when it was coadministered with companion drug versus being given alone, using a two-sided t test at a significance level of 0.05. We targeted enrollment of 16 participants per arm.

(ii) Pharmacokinetic and statistical evaluation. PK parameters for PA-824, efavirenz, and lopinavir, including AUC, maximum plasma concentration (C_{max}), time of maximum plasma concentration (T_{max}), half-life ($t_{1/2}$), and oral clearance (CL/F) were determined using standard non-compartmental methods performed in SAS (SAS Institute, Inc., Cary, NC). Statistical analyses were based on nonparametric tests. The P values evaluating changes in PK of PA-824 coadministered with efavirenz, lopinavir/r, or rifampin to PA-824 alone determined using a Wilcoxon signed-rank test and comparing changes in PK parameters of these drugs among metabolizer groups using a Wilcoxon rank sum test or Kruskal-Wallis test are reported. Calculated geometric means of ratios (GMR) and 90% confidence intervals based on log-transformed PK parameters were also used for PK comparisons. Associations between genotypes and PK parameters were assessed using the Jonckheere-Terpstra trend test and assuming additive genetic models.

RESULTS

Study subjects. Fifty-two participants enrolled. Median age was 34 years (range, 19 to 63 years), median weight was 83 kg (range, 47 to 119 kg), median body mass index was 27 kg/m² (range, 18 to 41 kg/m²), and 30 (58%) were male. Thirty-one (60%) were white, 17 (33%) were African-American or black, and 2 were Asian, and 2 were not reported. Of 52 participants, 48 completed all PK visits. There were two early discontinuations each in arms 1 and 2. In arm 1, one individual had efavirenz-related side effects, and another was a passenger in a motor vehicle accident. In arm 2, two participants self-administered a lower-than-prescribed lopinavir/r dose and so were discontinued. Thus, 48 participants were eligible for PK analyses.

TABLE 1 Pharmacokinetic parameters of PA-824 when PA-824 is administered alone or coadministered with steady-state efavirenz, lopinavir/r, or rifampin

Companion drug (treatment group) ^a	Pharmacokinetic parameter for PA-824	Median (IQR) for the parameter ^c		GMR ^b	90% CI ^d	P value ^e
		PA-824 alone	PA-824 and companion drug			
Efavirenz (arm 1)	AUC ₀₋₂₄ (ng · h/ml)	36,495 (30,853, 53,857)	24,917 (19,094, 34,257)	0.65	(0.56, 0.76)	<0.001
	C _{max} (ng/ml)	2,035 (1,805, 2,840)	1,510 (1,225, 2,025)	0.72	(0.62, 0.83)	0.001
	C _{min} (ng/ml)	1,110 (892, 1,650)	653 (502, 936)	0.54	(0.45, 0.64)	<0.001
	T _{max} (h)	4.0 (4.0, 5.0)	4.0 (3.5-5.0)	0.88	(0.65, 1.20)	0.383
	t _{1/2} (h)	24.8 (18.9, 27.1)	16.2 (14.9, 21.0)	0.74	(0.68, 0.80)	<0.001
	CL/F (liters/h)	5.48 (3.71, 6.48)	8.03 (5.87, 10.6)	1.53	(1.31, 1.78)	<0.001
Lopinavir/r (arm 2)	AUC ₀₋₂₄ (ng · h/ml)	39,035 (24,295, 42,187)	29,899 (20,691, 37,949)	0.83	(0.71, 0.98)	0.02
	C _{max} (ng/ml)	2,130 (1,440, 2,425)	1,770 (1,285, 2,245)	0.87	(0.75, 1.0)	0.03
	C _{min} (ng/ml)	1,085 (708, 1,320)	838 (509, 1,155)	0.79	(0.66, 0.93)	0.01
	T _{max} (h)	4.0 (3.5, 5.0)	4.5 (3.5, 5.0)	1.10	(0.88, 1.38)	0.47
	t _{1/2} (h)	21.6 (18.4, 28.7)	16.7 (15.1, 23.7)	0.83	(0.73, 0.94)	0.04
	CL/F (liters/h)	5.13 (4.74, 8.23)	6.69 (5.28, 9.67)	1.20	(1.03, 1.41)	0.03
Rifampin (arm 3)	AUC ₀₋₂₄ (ng · h/ml)	42,495 (29,501, 48,661)	13,659 (9,981, 19,070)	0.34	(0.27, 0.42)	<0.001
	C _{max} (ng/ml)	2,490 (1,925, 2,885)	1,165 (769, 1,520)	0.47	(0.39, 0.56)	<0.001
	C _{min} (ng/ml)	1,080 (722, 1,300)	173 (93, 320)	0.15	(0.11, 0.21)	<0.001
	T _{max} (h)	4.0 (3.0, 6.0)	4.0 (3.5, 4.5)	1.00	(0.81, 1.22)	0.58
	t _{1/2} (h)	19.25 (15.66, 20.78)	8.07 (6.28, 9.22)	0.41	(0.36, 0.46)	<0.001
	CL/F (liters/h)	4.72 (4.12, 6.78)	14.67 (10.52, 20.31)	2.97	(2.41, 3.67)	<0.001

^a Dosing was as follows: PA-824, 200 mg once daily; efavirenz, 600 mg once daily; lopinavir/r, 400 mg/100 mg twice daily; rifampin, 600 mg once daily.

^b Geometric mean of ratios (GMR) of pharmacokinetics of PA-824 coadministered with a companion drug to PA-824 alone.

^c IQR, interquartile range.

^d CI, confidence interval.

^e P value of Wilcoxon signed-rank test comparing pharmacokinetics of PA-824 coadministered with the companion drug to PA-824 alone.

Pharmacokinetics of PA-824, efavirenz, and lopinavir. Compared to PA-824 alone, plasma exposures (AUC₀₋₂₄) of PA-824 (based on GMR) were reduced by 35% with efavirenz, 17% with lopinavir/r, and 66% with rifampin (Table 1). Plasma concentration-time curves for PA-824 alone versus PA-824 with efavirenz, lopinavir/r, and rifampin are shown in Fig. 2 and in color in Fig. S1 in the supplemental material. Plasma efavirenz and lopinavir con-

centrations were not appreciably affected by PA-824 (Table 2). In a *post hoc* nonlinear mixed-effects modeling analysis of our phase 1 trial data plus raw data from phase 1 and 2 trials of PA-824 supplied by TB Alliance, we found that PA-824 exposures were similar for a 200-mg dose taken together with efavirenz and a 100-mg dose taken without efavirenz. The same was true for PA-824 coadministered with rifampin, except that while overall expo-

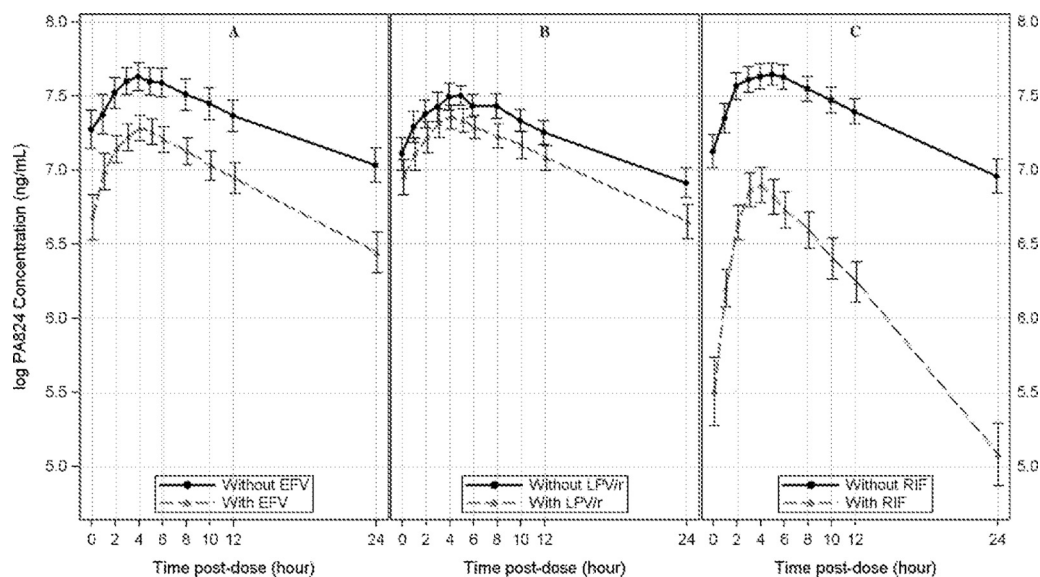


FIG 2 Mean log₁₀ PA-824 plasma concentration-versus-time curve of PA-824 at 200 mg once daily alone (solid lines) or together (dotted lines) with steady-state efavirenz (EFV) (A), lopinavir/ritonavir (LPV/r) (B), or rifampin (RIF) (C). Values shown represent means with standard errors.

TABLE 2 Pharmacokinetic parameters of efavirenz and lopinavir when administered alone or coadministered with PA-824

Drug (treatment group) ^a	Pharmacokinetic parameter	Median (IQR) for the parameter ^c				
		Drug alone	Drug with PA-824	GMR ^b	90% CI ^d	P value
Efavirenz (arm 1)	AUC _{0–24} (ng · h/ml)	62,112 (48,568, 72,301)	55,835 (44,347, 73,023)	0.96	(0.91, 1.02)	0.25
	C _{max} (ng/ml)	4,380 (3,712, 4,773)	3,945 (2,616, 4,897)	0.86	(0.72, 1.02)	0.21
	C _{min} (ng/ml)	1,868 (1,359, 2,275)	1,768 (1,193, 2,270)	0.96	(0.90, 1.04)	0.82
	t _{1/2} (h)	20.2 (15.6, 24.3)	18.5 (16.5, 33.3)	1.14	(0.93, 1.40)	0.38
	CL/F (liters/h)	3.23 (2.77, 4.12)	3.61 (2.74, 4.51)	1.04	(0.98, 1.11)	0.19
Lopinavir/r (arm 2)	AUC _{0–24} (ng · h/ml)	95,689 (71,321, 117,600)	87,092 (58,858, 98,231)	0.86	(0.77, 0.96)	0.005
	C _{max} (ng/ml)	11,450 (8,955, 13,400)	10,150 (7,400, 10,850)	0.83	(0.76, 0.92)	0.004
	C _{min} (ng/ml)	4,095 (2,245, 6,255)	2,925 (2,200, 5,110)	1.03	(0.60, 1.80)	0.009
	t _{1/2} (h)	6.71 (4.67, 8.76)	6.54 (4.95, 8.22)	0.96	(0.84, 1.09)	0.63
	CL/F (liters/h)	4.18 (3.41, 5.64)	4.59 (4.07, 6.80)	1.17	(1.05, 1.30)	0.005

^a Efavirenz was given at a dose of 600 mg daily; lopinavir/r was given at a dose of 400 mg/100 mg twice daily.

^b Geometric mean of ratios (GMR) of pharmacokinetics of PA-824 coadministered with a companion drug to PA-824 alone.

^c IQR, interquartile range.

^d CI, confidence interval.

tures were similar, trough concentrations remained modestly reduced with rifampin coadministration (data not shown).

Pharmacogenetic associations. Of 16 subjects evaluable for PK in arm 1 (efavirenz), 6 (38%) were *CYP2B6* extensive metabolizers, 10 (63%) were intermediate metabolizers, and none were slow metabolizers. Changes in PK parameter values for PA-824 (based on geometric means of ratios [GMR]) with concomitant efavirenz did not differ significantly between *CYP2B6* intermediate and extensive metabolizers (e.g., PA-824 C_{min} reduced by 44% and 49%, respectively; *P* = 0.692). Of 16 subjects evaluable for PK in arm 2 (lopinavir/r), 12 (75%) were homozygous for *SLCO1B1* T/T and 4 (25%) were heterozygous for *SLCO1B1* 521 C/T. Regarding *CYP3A5* 36986A→G (rs776746), 5 subjects (31%) had extensive, 3 (19%) had intermediate, and 8 (50%) had slow metabolizer genotypes. Changes in PK parameter values for PA-824 (based on GMR) with concomitant lopinavir/r did not differ consistently between *SLCO1B1* C/T and *SLCO1B1* T/T or by *CYP3A5* genotype. Of 16 subjects evaluable for PK in arm 3 (rifampin), 4 (25%) were homozygous for *SLCO1B1* 38664 (rs4149032), 5 (31%) were heterozygous for *SLCO1B1* 38664 (rs4149032), and 7 (44%) were homozygous for *SLCO1B1* 38664 (rs4149032). Regarding *CYP3A5* 36986A→G (rs776746), 4 subjects (25%) had extensive, 6 (38%) had intermediate, and 6 (38%) had slow metabolizer genotypes. Changes in PK parameter values for PA-824 (based on GMR) with concomitant rifampin did not differ significantly by *SLCO1B1* rs4149032 or by *CYP3A5* genotype. Relationships between polymorphisms and PK parameters of efavirenz and lopinavir as well as relationships between polymorphisms and magnitude of drug-drug interactions when PA-824 was administered with efavirenz, lopinavir/r, or rifampin are described in the supplemental material.

To explore genetic associations, we considered PA-824 PK data without concomitant efavirenz, lopinavir/r, or rifampin in all 48 subjects. We found no apparent association between *CYP2B6*, *SLCO1B1*, and *CYP3A5* polymorphisms and PA-824 PK parameters. The relationship between these polymorphisms and C_{min} is shown in the supplemental material.

Safety and tolerability. PA-824 was well tolerated alone and with concomitant efavirenz, lopinavir/r, and rifampin. There were two adverse events of grade ≥3. In arm 2, one participant

had an asymptomatic elevation of aspartate transaminase (AST) following vigorous exercise (PA-824 alone, sequence 2). One subject in arm 3 experienced grade 3 neutropenia on the last day of dosing, likely due to rifampin. Both adverse events resolved quickly after drug discontinuation. There were no QTc events of grade ≥2.

DISCUSSION

For the first time in decades, there is a robust drug development pipeline for tuberculosis. The nitroimidazole PA-824 is poised to enter phase 3 clinical trials. Anticipating the need to treat patients coinfecting with HIV-1, we examined the safety, tolerability, and PK of PA-824 given with commonly used antiretrovirals that induce or inhibit P450 metabolizing enzymes (efavirenz and lopinavir/r) and with the essential first-line tuberculosis drug, rifampin. We showed substantial reduction of plasma PA-824 exposure by efavirenz and rifampin but modest changes with lopinavir/r. The combinations were safe and well tolerated, and PA-824 did not affect plasma efavirenz or lopinavir exposures.

Concurrent treatment of HIV and tuberculosis reduces mortality and new AIDS-defining illnesses (23–25). Because cotreatment can be complicated by drug-drug interactions, overlapping drug toxicities, immune reconstitution syndrome, and high pill burden (26), late-phase clinical trials of antituberculosis drugs typically exclude patients requiring antiretroviral therapy. In the present study, coadministration of PA-824 with lopinavir/ritonavir did not increase PA-824 concentrations; rather, exposures were modestly reduced. Ritonavir is a mixed inducer and inhibitor, and with regard to interactions with PA-824, induction apparently dominated. This demonstrates the importance of empirical data when ritonavir is used with other drugs as it is difficult to make *a priori* predictions about its likely effects on companion drugs (10, 27). The modest reductions in PA-824 with lopinavir/r were statistically significant but are likely not clinically relevant. In contrast, efavirenz substantially decreased PA-824 exposure, likely by upregulating *CYP3A* or other metabolizing enzymes. While *CYP3A* contributes only 20% to overall metabolism of PA-824, this percentage may increase when *CYP3A* is induced. Whether

interactions of efavirenz and rifampin with PA-824 will be clinically important can only be answered by PK/pharmacodynamics (PD) analysis of trial data in which different doses of PA-824 are tested for longer durations in patients with tuberculosis. PK and outcome data from an 8-week phase 2 clinical trial are expected soon, and with these data in hand, concentration-effect relationships can be explored more fully.

Rifampin has unique sterilizing activity against *M. tuberculosis*, making it a mainstay of first-line tuberculosis treatment. To date, there are no drugs clinically proven to have sterilizing activity equal to rifampin. Rifampin is, however, a potent inducer of metabolizing enzymes and drug transporters (28). Rifampin-induced drug interactions complicate drug development efforts for drug-sensitive tuberculosis in two ways. First, promising investigational drugs cannot be added to first-line antituberculosis regimens without evaluating PK effects of rifampin and other coadministered antituberculosis drugs. For example, rifampin reduces concentrations of the newest TB drug, bedaquiline, by 50% (29). Conversely, isoniazid can inhibit metabolizing enzymes, and unexpected effects may occur when rifampin and isoniazid are coadministered with a third drug (30, 31). Second, rifamycin antibiotics like rifampin and rifapentine have dose-dependent treatment-shortening potential, but evaluating high-dose rifamycins is challenging because the magnitude of drug interactions at increased rifamycin doses is unknown. That is, while it is generally believed that rifamycins' inductive capabilities are maximized at currently used doses, recent preliminary studies in human hepatocytes suggest that mRNA expression of CYP3A increases with higher rifamycin concentrations, within clinically relevant ranges (32). Whether or not higher mRNA expression will lead to greater enzyme activity or higher risk for clinically meaningful drug interactions is unknown.

Interpretation of drug interaction study results requires understanding of study drug pharmacodynamics (PD; i.e., correlations between PK parameters and efficacy). For tuberculosis, lack of a reliable biomarker of treatment response makes it difficult to define PK-PD relationships. PA-824 is being tested in a 2-month phase 2B treatment trial at doses of 100 mg and 200 mg daily because doses from 100 mg to 1,000 mg had similar activities in the 2-week dose-ranging phase 2A monotherapy studies; only at 50 mg daily was early bactericidal activity (EBA) decreased (33, 34). Preclinical studies suggest time-dependent activity of PA-824 against *M. tuberculosis* (35), suggesting that the AUC_{0-24} may be a key pharmacodynamic parameter; however, target AUC values have not been defined. The phase 2B study may help define dose-effect or concentration-effect relationships that will give our results context.

There are well-replicated associations between *CYP2B6* polymorphisms and efavirenz PK (11–15) and between an *SLCO1B1* polymorphism and lopinavir PK (16–18). An association has been reported between an *SLCO1B1* polymorphism and rifampin PK (20, 21). It is important to consider whether these polymorphisms affect drug interactions. In addition, because PA-824 is metabolized in part by CYP3A, we assessed a *CYP3A5* loss-of-function polymorphism. Our study did not show magnitudes of effects of efavirenz, lopinavir, and rifampin on PA-824 PK parameters to differ by the above genetic polymorphisms. In addition, we found no apparent associations between these polymorphisms and PA-824 C_{min} though the sample size was limited. We suspect that the

apparent association between *CYP3A5**3 and lopinavir PK is spurious since this association has not been seen elsewhere (16–18).

There were limitations to the present study. Because study drugs were given for relatively brief intervals, the full safety profile could not be assessed. The effect of rifampin on PA-824 was not assessed in the context of full, multidrug first-line antituberculosis treatment. It is possible that effects of rifampin alone differ from effects when it is combined with other first-line antituberculosis drugs. None of the multiple metabolites of PA-824 were measured in this study, and the specific metabolizing enzyme(s) that mediates reductions in PA-824 seen in this study is not known.

In conclusion, PA-824 was well tolerated when given with efavirenz, lopinavir/r, or rifampin. Concomitant lopinavir/r only modestly reduced PA-824 plasma exposures, suggesting that the drugs can be coadministered without dose adjustment. Efavirenz reduced PA-824 exposures more substantially, and rifampin reduced PA-824 exposure even more. The clinical implications of these findings should be interpreted in light of results of ongoing phase 2 dose-ranging trials that will define dose-response relationships and identify target concentrations for maximal PA-824 effect so that use of PA-824 in first-line regimens and in patients requiring HIV-1 therapy can be optimized.

ACKNOWLEDGMENTS

We thank the individuals who volunteered for and participated in this study. We acknowledge the support of the analytical laboratory at University of California at San Francisco. We also thank Danielle Richardson, Melissa Potter, and Cara Sutcliffe in the Vanderbilt DNA Resources Core for genotyping. In particular, we appreciate the contributions of study team members Liz Barr, Erica Egizi, Laura Hovind, Jennifer Janik, Patricia Lizak, Carl Mendel, and Thucuma Size. The Global Alliance for TB Drug Development provided PA-824 for this study.

We appreciate the efforts of the investigators and research staff at the following ACTG Clinical Trial Units (CTU) and Clinical Research Sites (CRS): Michael Leonard and Fred Nicotera, Vanderbilt Therapeutics Clinical Research Site (site 3652) (ACTG CTU grant U01-AI069439-01; Vanderbilt CTSA grant UL1RR024975 from NCRR/NIH); John Dwyer, Anna Smith, and Jacqueline Clark, UCSF AIDS CRS (site 801) (ACTG CTU grant U01-AI069502-01 and UCSF CTSI grant UL1TR000004); Heather Harber and Jan Clark, The Ohio State University AIDS CRS (site 2301) (ACTG CTU grant 1U01AI069474-01 and grant UL1TR001070); Ilene Wiggins and Andrea Weiss, Johns Hopkins Adult AIDS CRS (site 201) (ACTG CTU grant 1U01AI069465-01 and grant UL1 TR001079; Johns Hopkins University Institute for Clinical and Translational Research).

This study was supported by the AIDS Clinical Trials Group sponsored by the National Institute of Allergy and Infectious Diseases and was supported by the following grants: UM1AI068636 (NIAID), UM1AI68634 (ACTG Statistical and Data Management Center), U01 AI069465 (K.E.D.), TR-000445, AI-077505 (D.W.H.), K23AI080842 (K.E.D.), and U01 AI068634 (J.-G.P. and Y.C.). This study was also funded in part by the National Institute of Allergy and Infectious Diseases, National Institutes of Health, Department of Health and Human Services contract HHSN272200800014C (J.B.).

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institute of Allergy and Infectious Diseases or the National Institutes of Health.

REFERENCES

1. World Health Organization. 2013. Global tuberculosis report 2013. WHO/HTM/TB/2013.11. World Health Organization, Geneva, Switzerland. http://www.who.int/tb/publications/global_report/en/.

2. World Health Organization. 2013. Multidrug-resistant tuberculosis (MDR-TB), 2013 update. World Health Organization, Geneva, Switzerland. http://www.who.int/tb/challenges/mdr/MDR_TB_FactSheet.pdf.
3. World Health Organization. 2011. Guidelines for the programmatic management of drug-resistant tuberculosis, 2011 update. WHO/HTM/TB/2011.6. World Health Organization, Geneva, Switzerland. http://www.who.int/tb/challenges/mdr/programmatic_guidelines_for_mdrtb/en/index.html.
4. Stover CK, Warren P, VanDevanter DR, Sherman DR, Arain TM, Langhorne MH, Anderson SW, Towell JA, Yuan Y, McMurray DN, Kreiswirth BN, Barry CE, Baker WR. 2000. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. *Nature* 405: 962–966. <http://dx.doi.org/10.1038/35016103>.
5. Lenaerts AJ, Gruppo V, Marietta KS, Johnson CM, Driscoll DK, Tompkins NM, Rose JD, Reynolds RC, Orme IM. 2005. Preclinical testing of the nitroimidazopyran PA-824 for activity against *Mycobacterium tuberculosis* in a series of *in vitro* and *in vivo* models. *Antimicrob. Agents Chemother.* 49:2294–2301. <http://dx.doi.org/10.1128/AAC.49.6.2294-2301.2005>.
6. Tasneen R, Tyagi S, Williams B, Grosset J, Nuermberger E. 2008. Enhanced bactericidal activity of rifampin and/or pyrazinamide when combined with PA-824 in a murine model of tuberculosis. *Antimicrob. Agents Chemother.* 52:3664–3668. <http://dx.doi.org/10.1128/AAC.00686-08>.
7. Nuermberger E, Tyagi S, Tasneen R, Williams KN, Almeida D, Rosenthal I, Grosset JH. 2008. Powerful bactericidal and sterilizing activity of a regimen containing PA-824, moxifloxacin, and pyrazinamide in a murine model of tuberculosis. *Antimicrob. Agents Chemother.* 52: 1522–1524. <http://dx.doi.org/10.1128/AAC.00074-08>.
8. Diacon AH, Dawson R, von Groote-Bidlingmaier F, Symons G, Venter A, Donald PR, van Niekerk C, Everitt D, Winter H, Becker P, Mendel CM, Spigelman MK. 2012. 14-Day bactericidal activity of PA-824, bedaquiline, pyrazinamide, and moxifloxacin combinations: a randomised trial. *Lancet* 380:986–993. [http://dx.doi.org/10.1016/S0140-6736\(12\)61080-0](http://dx.doi.org/10.1016/S0140-6736(12)61080-0).
9. Dooley KE, Flexner C, Andrade AS. 2008. Drug interactions involving combination antiretroviral therapy and other anti-infective agents: repercussions for resource-limited countries. *J. Infect. Dis.* 198:948–961. <http://dx.doi.org/10.1086/591459>.
10. Kirby BJ, Collier AC, Kharasch ED, Whittington D, Thummel KE, Unadkat JD. 2011. Complex drug interactions of HIV protease inhibitors 1: inactivation, induction, and inhibition of cytochrome P450 3A by ritonavir or nelfinavir. *Drug Metab. Dispos.* 39:1070–1078. <http://dx.doi.org/10.1124/dmd.110.037523>.
11. Haas DW, Ribaldo HJ, Kim RB, Tierney C, Wilkinson GR, Gulick RM, Clifford DB, Hulgath T, Marzolini C, Acosta EP. 2004. Pharmacogenetics of efavirenz and central nervous system side effects: an Adult AIDS Clinical Trials Group study. *AIDS* 18:2391–2400.
12. Rotger M, Colombo S, Furrer H, Bleiber G, Buclin T, Lee BL, Keiser O, Biollaz J, Decosterd L, Telenti A, Swiss HIV Cohort Study. 2005. Influence of CYP2B6 polymorphism on plasma and intracellular concentrations and toxicity of efavirenz and nevirapine in HIV-infected patients. *Pharmacogenet. Genomics* 15:1–5. <http://dx.doi.org/10.1097/01213011-200501000-00001>.
13. Wyen C, Hendra H, Vogel M, Hoffmann C, Knechten H, Brockmeyer NH, Bogner JR, Rockstroh J, Esser S, Jaeger H, Harrer T, Mauss S, van Lunzen J, Skoetz N, Jetter A, Gronauer C, Fatkenheuer G, Khoo SH, Egan D, Back DJ, Owen A, and German Competence Network for HIV/AIDS. 2008. Impact of CYP2B6 983T>C polymorphism on non-nucleoside reverse transcriptase inhibitor plasma concentrations in HIV-infected patients. *J. Antimicrob. Chemother.* 61:914–918. <http://dx.doi.org/10.1093/jac/dkn029>.
14. Ribaldo HJ, Liu H, Schwab M, Schaeffeler E, Eichelbaum M, Motsinger-Reif AA, Ritchie MD, Zanger UM, Acosta EP, Morse GD, Gulick RM, Robbins GK, Clifford D, Haas DW. 2010. Effect of CYP2B6, ABCB1, and CYP3A5 polymorphisms on efavirenz pharmacokinetics and treatment response: an AIDS Clinical Trials Group Study. *J. Infect. Dis.* 202:717–722. <http://dx.doi.org/10.1086/655470>.
15. Holzinger ER, Grady B, Ritchie MD, Ribaldo HJ, Acosta EP, Morse GD, Gulick RM, Robbins GK, Clifford DB, Daar ES, McLaren P, Haas DW. 2012. Genome-wide association study of plasma efavirenz pharmacokinetics in AIDS Clinical Trials Group protocols implicates several CYP2B6 variants. *Pharmacogenet. Genomics* 22:858–867. <http://dx.doi.org/10.1097/FPC.0b013e32835a450b>.
16. Lubomirov R, di Iulio J, Fayet A, Colombo S, Martinez R, Marzolini C, Furrer H, Vernazza P, Calmy A, Cavassini M, Ledergerber B, Rentsch K, Descombes P, Buclin T, Decosterd LA, Csajka C, Telenti A, Swiss HIV Cohort Study. 2010. ADME pharmacogenetics: investigation of the pharmacokinetics of the antiretroviral agent lopinavir coformulated with ritonavir. *Pharmacogenet. Genomics* 20:217–230. <http://dx.doi.org/10.1097/FPC.0b013e328336eee4>.
17. Hartkoorn RC, Kwan WS, Shallcross V, Chaikan A, Liptrott N, Egan D, Sora ES, James CE, Gibbons S, Bray PG, Back DJ, Khoo SH, Owen A. 2010. HIV protease inhibitors are substrates for OATP1A2, OATP1B1 and OATP1B3 and lopinavir plasma concentrations are influenced by SLCO1B1 polymorphisms. *Pharmacogenet. Genomics* 20:112–120. <http://dx.doi.org/10.1097/FPC.0b013e328335b02d>.
18. Kohlrausch FB, de Cassia Estrela R, Barroso PF, Suarez-Kurtz G. 2010. The impact of SLCO1B1 polymorphisms on the plasma concentration of lopinavir and ritonavir in HIV-infected men. *Br. J. Clin. Pharmacol.* 69: 95–98. <http://dx.doi.org/10.1111/j.1365-2125.2009.03551.x>.
19. Division of AIDS, NIAID. 2009. Division of AIDS table for grading the severity of adult and pediatric adverse events, version 1.0, December 2004; clarification August 2009. Division of AIDS, NIAID, National Institutes of Health, Bethesda, MD. http://www.mtnstopshiv.org/sites/default/files/attachments/DAIDS_AE_GradingTable_ClarificationAug2009_Final_%5B1%5D.pdf.
20. Chigutsa E, Visser ME, Swart EC, Denti P, Pushpakom S, Egan D, Holford NH, Smith PJ, Maartens G, Owen A, McIlleron H. 2011. The SLCO1B1 rs4149032 polymorphism is highly prevalent in South Africans and is associated with reduced rifampin concentrations: dosing implications. *Antimicrob. Agents Chemother.* 55:4122–4127. <http://dx.doi.org/10.1128/AAC.01833-10>.
21. Weiner M, Peloquin C, Burman W, Luo CC, Engle M, Prihoda TJ, MacKenzie WR, Bliven-Sizemore E, Johnson JL, Vernon A. 2010. Effects of tuberculosis, race, and human gene SLCO1B1 polymorphisms on rifampin concentrations. *Antimicrob. Agents Chemother.* 54:4192–4200. <http://dx.doi.org/10.1128/AAC.00353-10>.
22. Kuehl P, Zhang J, Lin Y, Lamba J, Assem M, Schuetz J, Watkins PB, Daly A, Wrighton SA, Hall SD, Maurel P, Relling M, Brimer C, Yasuda K, Venkataramanan R, Strom S, Thummel K, Boguski MS, Schuetz E. 2001. Sequence diversity in CYP3A promoters and characterization of the genetic basis of polymorphic CYP3A5 expression. *Nat. Genet.* 27:383–391. <http://dx.doi.org/10.1038/86882>.
23. Havlir DV, Kendall MA, Ive P, Kumwenda J, Swindells S, Qasba SS, Luetkemeyer AF, Hogg E, Rooney JF, Wu X, Hosseinipour MC, Laloo U, Veloso VG, Some FF, Kumarasamy N, Padayatchi N, Santos BR, Reid S, Hakim J, Mohapi L, Mugenyi P, Sanchez J, Lama JR, Pape JW, Sanchez A, Amelash A, Moko E, Sawe F, Andersen J, Sanne I, AIDS Clinical Trials Group Study A5221. 2011. Timing of antiretroviral therapy for HIV-1 infection and tuberculosis. *N. Engl. J. Med.* 365:1482–1491. <http://dx.doi.org/10.1056/NEJMoa1013607>.
24. Blanc F, Sok T, Laureillard D, Borand L, Rekeciewicz C, Nerrienet E, Madec Y, Marcy O, Chan S, Prak N, Kim C, Lak KK, Hak C, Dim B, Sin CI, Sun S, Guillard B, Sar B, Vong S, Fernandez M, Fox L, Delfraissy J, Goldfeld AE, CAMELIA (ANRS 1295-CIPRA KH001) Study Team. 2011. Earlier versus later start of antiretroviral therapy in HIV-infected adults with tuberculosis. *N. Engl. J. Med.* 365:1471–1481. <http://dx.doi.org/10.1056/NEJMoa1013911>.
25. Abdool Karim SS, Naidoo K, Grobler A, Padayatchi N, Baxter C, Gray AL, Gengiah T, Gengiah S, Naidoo A, Jithoo N, Nair G, El-Sadr WM, Friedland G, Abdool Karim Q. 2011. Integration of antiretroviral therapy with tuberculosis. *N. Engl. J. Med.* 365:1492–1501. <http://dx.doi.org/10.1056/NEJMoa1014181>.
26. CDC. 2013. Managing drug interactions in the treatment of HIV-related tuberculosis. Centers for Disease Control and Prevention, Atlanta, GA. http://www.cdc.gov/tb/HIV_Drugs/default.htm.
27. Kharasch ED, Stubbert K. 2013. Cytochrome P4503A does not mediate the interaction between methadone and ritonavir/lopinavir. *Drug Metab. Dispos.* 41:2166–2174. <http://dx.doi.org/10.1124/dmd.113.053991>.
28. Burman W, Dooley KE, Nuermberger E. 2011. The rifamycins: renewed interest in an old drug class, p 18–24. *In* Donald PR, van Helden PD (ed), *Antituberculosis chemotherapy*. Karger, Basel, Switzerland.
29. Janssen Therapeutics. 2012. Sirturo package insert. Janssen Therapeutics, Titusville, NJ. http://www.accessdata.fda.gov/drugsatfda_docs/label/2012/204384s000lbl.pdf.
30. Lee L, Soon GH, Chew N, Else L, Amara A, Knoo S. 2013. Differential

- induction of efavirenz metabolism by rifampin without and with isoniazid in healthy volunteers with CYP2B6 516GG and TT genotypes, abstr 516. Abstr. 20th Conf. Retroviruses Opportunistic Infect., Atlanta, GA, 3 to 6 March 2013.
31. Bertrand J, Verstuyft C, Chou M, Borand L, Chea P, Nay KH, Blanc FX, Mentre F, Taburet AM, CAMELIA (ANRS 1295-CIPRA KH001) Study Group. 2014. Dependence of efavirenz- and rifampicin-isoniazid-based antituberculosis treatment drug-drug interaction on CYP2B6 and NAT2 genetic polymorphisms: ANRS 12154 study in Cambodia. *J. Infect. Dis.* 209:399–408. <http://dx.doi.org/10.1093/infdis/jit466>.
 32. Williamson B, Dooley KE, Zhang Y, Back DJ, Owen A. 2013. Induction of influx and efflux transporters and cytochrome P450 3A4 in primary human hepatocytes by rifampin, rifabutin, and rifapentine. *Antimicrob. Agents Chemother.* 57:6366–6369. <http://dx.doi.org/10.1128/AAC.01124-13>.
 33. Diacon AH, Dawson R, Hanekom M, Narunsky K, Maritz SJ, Venter A, Donald PR, van Niekerk C, Whitney K, Rouse DJ, Laurenzi MW, Ginsberg AM, Spigelman MK. 2010. Early bactericidal activity and pharmacokinetics of PA-824 in smear-positive tuberculosis patients. *Antimicrob. Agents Chemother.* 54:3402–3407. <http://dx.doi.org/10.1128/AAC.01354-09>.
 34. Diacon AH, Dawson R, du Bois J, Narunsky K, Venter A, Donald PR, van Niekerk C, Erondu N, Ginsberg AM, Becker P, Spigelman MK. 2012. Phase II dose-ranging trial of the early bactericidal activity of PA-824. *Antimicrob. Agents Chemother.* 56:3027–3031. <http://dx.doi.org/10.1128/AAC.06125-11>.
 35. Ahmad Z, Peloquin CA, Singh RP, Derendorf H, Tyagi S, Ginsberg A, Grosset JH, Nuermberger EL. 2011. PA-824 exhibits time-dependent activity in a murine model of tuberculosis. *Antimicrob. Agents Chemother.* 55:239–245. <http://dx.doi.org/10.1128/AAC.00849-10>.