Phenotypic variation in progenies from somatic hybrids between *Brassica napus* and *Sinapis alba*

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Abstract A number of variant phenotypes, such as long siliques, high density of pods, increased seed number per pod and yellow seed color resembling Sinapis alba, were selected from intergeneric somatic hybrids between Brassica napus and Sinapis alba through successive backcrosses. Resistance to Sclerotinia sclerotiorum among 24 BCF₄ lines was also tested by inoculation with mycelia, and four lines showed stronger resistance than the resistant rapeseed variety 'Zhongshuang 9'. Comparative anatomy studies on mature seed revealed that the seed coat pigments were mainly distributed in the palisade layer, which is considerably thinner in S. alba compared to B. napus. The area index of protein bodies in cotyledon cells was highest in S. alba, lowest in B. napus and intermediate in the intergeneric progenies. This study demonstrates that wide hybridization can enable exploitation of valuable trait

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Institute for Plant Breeding and Crop Science I, Justus Liebig University, Heinrich-Buff-Ring 26-32, 35392 Giessen, Germany diversity from *S. alba* for broadening the genetic basis for rapeseed breeding.

Keywords Brassica napus L. · Intergeneric somatic hybrid · Sclerotinia sclerotiorum · Sinapis alba L. · Yellow seed

Introduction

The genetic basis of Brassica napus L., one of the most important oilseed crops worldwide, is quite narrow. The species originated in Europe and was introduced into China as an oil crop in the 1950s (Liu 1984; Wang 2005). Intergeneric hybridization with related crucifer species is often used to broaden genetic variability in B. napus (Snowdon et al. 2006), however, sexual incompatibility limits the use of such approaches (Wang et al. 2005a, b, c). On the other hand, somatic hybridization allows not only the creation of hybrid and cybrid combinations of species that are sexually incompatible, but also recombination and transfer of cytoplasmic genes from both parents, with the aim of improving the characteristics controlled by cytoplasmic genes (Ge and Chen 2004; Liu et al. 2005; Prakash et al. 2009). Protoplast fusion was also used by Ren et al. (2000) and Hu et al. (2002) to recover novel rapeseed germplasm resistant to bacterial soft rot and Leptosphaeria maculans.

Sinapis alba L., a member of the Brassicaceae, possesses desirable agronomic characteristics such as yellow seed color, tolerance to drought stress, reduced pod shattering (Downey 1987), resistance to virus diseases, blackleg disease, black spot and beet cyst nematodes (Bodnaryk and Lamb 1991; Lelivelt et al. 1993; Brown et al. 1997; Hansen and Earle 1997). Intergeneric hybrids resistant to black spot were produced by ovary culture of B. rapa \times S. alba sexual hybrids (Gong et al. 1994). Somatic hybrids of B. napus and S. alba obtained by electrofusion have been described previously (Wang et al. 2005b, c). For creation of high yielding rapeseed lines with improved disease resistance and seed quality, these hybrids were subsequently backcrossed with B. napus and self-pollinated to obtain a BCF₄ generation with valuable agronomic characteristics derived from S. alba. This material contains various interesting phenotypes that may be useful for rapeseed breeding. The objective of this study was to characterize phenotypic variants observed in the BCF₄ progenies.

Materials and methods

Materials

Intergeneric somatic hybrids (AACCSS, 2n = 62) between *B. napus* and *S. alba* obtained by electrofusion were subsequently backcrossed with *B. napus* cv. 'Yangyou 6' (AACC, 2n = 38) and self-pollinated. Figure 1 shows the backcrossing scheme for

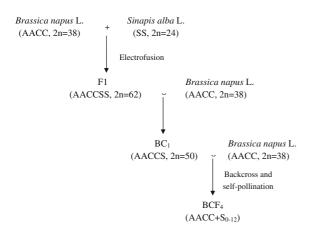


Fig. 1 Backcrossing scheme for the development of BCF₄ progenies

the development of BCF_4 progeny during 2005 and 2008.

Observation of agronomic characteristics

Field tests of the somatic hybrid backcross progeny were carried out at the experimental station of Jiangsu Lixiahe Region Agricultural Research Institute, China. Morphological variations observed in BCF_4 included main axis length, number of pods per plant, number of seeds per silique and pod length, as well as the beak length of pods. Pod density was described by the ratio between the number of pods and the length of the main raceme.

Pigment distribution in seed coat and thickness of the palisade layer

Experimental samples were taken from the adaxial side of seeds, then 15 μ m thick frozen sections were cut using a Leica CM1100 microtome at -15° C. Sections were photographed using an Olympus CX 41 light microscope without staining. The thickness of the palisade layer was measured on three seeds from each line, using the parental lines as a control.

Anatomical structure of seeds

After soaking in ddH₂O for 1 h at room temperature, seed capsules from the adaxial side and cotyledon embryos were used as experimental samples for transmission electronic microscopy. Samples were embedded in Spurr's resin, then semi-thin resin sections (1 µm) were cut on a Leica EM UC6 microtome using a glass blade, while ultra-thin sections (70 nm) were generated using a diamond blade. The semi-thin sections were counterstained with PAS-sudan black B-coomassie blue R according to Hu and Xu (1990). Observations and photographs were made via light microscopy. The number of protein bodies was counted in ten adjacent cells on each section, and the cumulative area of protein bodies along with the total area of the ten cells was measured using the morphological analysis software JD801. Subsequently, the mean area index of protein body per cell was calculated in each line. All data presented in this paper were obtained from five sections from each of the progeny and parental lines. After double-staining with uranium acetate and lead citrate, observation and image merging of the ultrathin sections were done with a Philips Tecnai 12 transmission electronic microscope.

Characterization of *Sclerotinia sclerotiorum* resistance

Artificial inoculation of plants with *S. sclerotiorum* was performed according to Cruickshank et al. (1983). After 72 h of mycelia inoculation with twenty detached leaves from different plants of each progeny line, the length and width of disease lesions were measured. The level of resistance to *S. sclerotiorum* was judged by the disease lesion area using the formula $S = L \times W$ (*S*: lesion area; *L*: length of lesion; *W*: width of lesion). All trial data were analyzed using the software DPS.

Determination of oil content

The oil content of self-pollinated seed from each of the BCF₄ progeny and parental lines was detected by Near Infrared Reflectance Spectroscopy using the NIRS Model TR-3700 from FOSS (Denmark).

Results

Variation in pod characteristics

Compared with BC_1 plants, pod length and seeds per pod among BCF₄ progeny lines were dramatically improved (Table 1). Three BCF_4 lines (D214, D224, D249) were identified with distinct variation in pod characteristics. D249 had a very high pod density of 1.8/cm, 38.5% higher than the recurrent parent 'Yangyou 6' (1.3/cm). Lines D214 and D224 had considerably longer siliques than the recurrent B. napus parent (Fig. 2e, f). The silique lengths of up to 9.3 and 9.1 cm in length, respectively, correspond to an increase of 69.1 and 65.5%, respectively compared to 'Yangyou 6' (5.5 cm). The number of seeds per pod in these three lines was over 26.8, a increase of 46.5, 55.2 and 60.7% respectively compared to 'Yangyou 6' (18.3 seeds per pod). Furthermore, the pods of the three selected lines had characteristically long beaks (Fig. 2), ranging from 1.4 to 1.8 cm, which resembles the pods of S. alba (2.0 cm beak length) rather than 'Yangyou 6' (1.1 cm beak length).

Table 1 P	od character.	Table 1 Pod characteristics of parents and backcross progenies	nd backcro	ss progenies								
Plant material		Main axisCompared withPodslength'Yangyou 6'main(cm) $(\pm\%)$ racem	Pods per main raceme	per Compared with Pod 'Yangyou 6' density ne (±%) per cn	> с	Compared with 'Yangyou 6' $(\pm\%)$	Pod length (cm)	Compared with 'Yangyou 6' (土%)	Seeds per pod	$ \begin{array}{c cccc} \mbox{Compared with} & \mbox{Pod} & \mbox{Pod} & \mbox{Compared with} & \mbox{Compared with} & \mbox{Compared with} & \mbox{Pod} & \mbox{Compared with} & Compared$	Pod beak length (cm)	Compared with 'Yangyou 6' (土%)
S. alba	30.2	/	84.3	/	2.8	/	1.2	/	3.7	/	2.0	/
Yangyou6 63.8	63.8	/	79.8	/	1.3	/	5.5	/	18.3	/	1.1	/
BC_1	86.3	35.3	119.6	49.9	1.4	T.T	3.1	-43.6	4.6	-74.9	1.3	18.2
D214	73.6	15.4	110.3	38.2	1.5	15.4	9.3	69.1	28.4	55.2	1.8	63.6
D224	72.5	13.6	60.5	-24.2	0.8	-38.5	9.1	65.5	29.4	60.7	1.4	27.3
D249	68.3	7.1	120.4	50.9	1.8	38.5	7.9	43.6	26.8	46.5	1.5	36.4



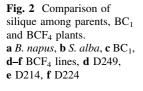


Fig. 3 Seed coat color of backcross progenies and their parents. a *S. alba*,
b 'Yangyou 6', c D244-18,
d D244-52, e D244-6,
f D255-3, g D246-5, h D211

Oil content and anatomical structure of yellow seeded progenies

Several lines with yellow seed color derived from the *S. alba* parent were identified among the backcross progenies. The seed color of these lines ranged from brownish yellow to yellow, while the *B. napus* parents had dark brown seeds (Fig. 3a–h). The oil content of these six yellow-seeded lines was found to be increased to 43.3, 43.4, 47.6, 42.9, 42.8, 44.6%,

respectively, compared to the oil content of 41.6% in 'Yangyou6' and 39.0% in the *S. alba* parental line.

The seed coat pigments were mainly distributed in the palisade layer (Fig. 4), the highest quantity of seed coat pigmentation was observed in 'Yangyou6', followed by the backcross lines D246-5, D211, D255-3 and D244-52. On the other hand, the seeds of the *S. alba* parent and the backcross progenies D244-18 and D244-6 exhibited almost no pigmentation. The thickness of palisade layer was highest in

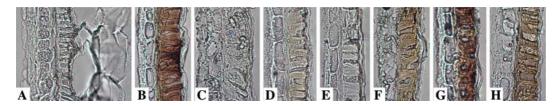


Fig. 4 Pigment distribution in seed coat of backcross progenies and their parents. a S. alba, b 'Yangyou 6', c D244-18, d D244-52, e D244-6, f D255-3, g D246-5, h D211. $\times 200$

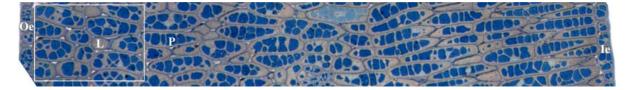


Fig. 5 The semi-thin section of cotyledon in S. alba. $\times 250$. Ie inner epidermical cell, L lipid body, Oe outer epidermical cell, P protein body

B. napus, thinnest in *S. alba* and intermediate in the hybrid progenies (Fig. 4).

When treated by PAS-sudan black B-coomassie blue R, the lipid body and storage proteins on semithin sections of cotyledon embryos were dyed gray and blue, respectively. The first step of PAS staining stains cell walls red, however, this changes to blueblack after sudan black B and coomassie blue R counterstaining (Fig. 5). Statistical analysis of the protein body area index in the first three layers under the outer epidermical of the seed cotyledons (Fig. 5) revealed that the protein body area index in cotyledon cells was highest in *S. alba*, lowest in *B. napus* and intermediate in progenies (Fig. 6).

Characterization of resistance to *Sclerotinia sclerotiorum*

Statistical analysis of disease lesions in the twentyfour progeny lines tested showed a significant variation in disease response among lines after inoculation with mycelia (F = 16.693, v1 = 494, v2 = 25). Compared with line D267 and 'Zhongshuang 9', which were used as susceptible and resistant controls, respectively, different resistance reactions were observed among the tested lines. Through multicomparison of inoculation of detached leaves with mycelia, we founded no significant difference in lines

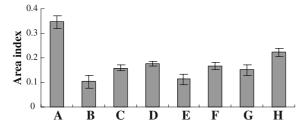


Fig. 6 Area index of protein body of backcross progenies and their parents **a** *S. alba*, **b** 'Yangyou 6', **c** D244-18, **d** D244-52, **e** D244-6, **f** D255-3, **g** D246-5, **h** D211

D216, D222, D225, D226 and D220 compared with susceptible line D267. On the other hand, lines D214, D245 and D219 were shown to have a stronger resistance than D267, but weaker than 'Zhongshuang 9'. Another eleven lines (D218, D223, D246, D212, D265, D211, D273, D241, D230, D237 and D213) were confirmed to have no obvious difference from the resistant control. Lines D244, D243, D229 and D240 exhibited the strongest resistance to *S. sclerotiorum* and showed significant differences to the resistant control 'Zhongshuang 9' (Table 2).

Discussion

Distant hybridization of sexually incompatible plant species via somatic fusion can overcome intergeneric boundaries, enlarge genetic variation, and ultimately result in the creation of new variants and species that do not occur naturally. Both sexual and somatic hybridization have been widely used to introduce traits of interest from related crucifer species to oilseed rape (Snowdon et al. 2006). Roy (1984) and Lu et al. (2006) obtained novel rapeseed with yellow seed color and resistance to blackleg through hybridization between B. juncea and B. napus, respectively. Progenies with extreme pod length variation (20 cm in length) were also created from interspecific hybrids of B. rapa and B. oleracea (Niu et al. 2005). These examples demonstrate that distant hybridization can not only introduce new characteristics from donors into phylogenetically close plants, but also create new variant types beyond the variation present in the parental species. In this study, we obtained lines with many agronomically interesting phenotypes from intergeneric somatic hybrid progenies of B. napus and S. alba through successive backcrosses. These included lines wth high pod density, yellow seed coats, resistance to Sclerotinia sclerotiorum, long siliques, increased numbers of seeds per pod and

rison of ter	Line No.	Mean lesion (cm ²)	Significance at 1%	Significance at 1%
detached	D216	25.12	а	А
PLSD)	D222	24.98	а	А
ILSD)	D267	24.65	ab	AB
	D225	23.16	abc	ABC
	D226	21.66	bcd	ABC
	D220	21.39	bcd	ABC
	D214	20.90	cd	ABCD
	D245	20.29	cd	BCDE
	D219	20.27	cd	BCDE
	D215	18.74	de	CDEF
	D218	16.69	ef	DEFG
	D223	16.53	ef	DEFG
	D246	16.22	efg	EFG
	D212	15.51	efgh	FG
	Zhongshuang9	15.15	fgh	FG
	D265	14.53	fgh	FGH
	D211	14.44	fgh	FGH
	D273	14.23	fgh	FGH
	D241	13.26	fghi	GH
	D230	12.85	ghi	GH
	D237	12.74	ghi	GHI
	D213	12.08	hi	GHI
	D244	10.17	ij	HIJ
	D243	9.88	ij	HIJ
	D229	8.07	j	IJ
	D240	6.85	j	J

Table 2Multi-comparison ofdisease lesion areas afterartificial inoculation of detachedleaves with Sclerotinasclerotiorum mycelia (PLSD)

 $LSD_{0.05} = 3.33,$ $LSD_{0.01} = 4.51$

improved oil content compared to the *B. napus* parent. These variant progenies are of great interest for rapeseed breeding, offering abundant germplasm resources for the creation of new varieties with improved disease resistance, seed quality and yield.

The oil and protein content of the yellow seeds is often higher than that of black ones to the the increased contribution of the cotyledons to the overall seed volume. In addition, the thinner seed coat of yellow-seeded *B. napus* is also associated with a lower antinutritive fiber content, while the reduced pigmentation results in a clear and transparent oil and a higher palatability of the seed meal in animal feeds. The yellow-seeded progenies obtained in the present study were found to have a thinner seed coat and reduced pigmentation compared to the dark-seeded *B. napus* parent. In contrast, the oil bodies of the best progenies resembled oil bodies in *B. napus* seeds rather than the smaller oil bodies of *S. alba* which has a very low oil content. On the other hand, the protein body area was significantly reduced in seeds from yellow-seeded progenies with high oil content, indicating that the reduction in seed coat thickness resulted in an increase in oil rather than an increase in protein proportion in the seeds.

Until now, genetic variation for seed color was only introduced to *B. napus* by interspecific hybridization with the closely related *Brassica* species *B. rapa*, *B. carinata*, *B. juncea* and *B. oleracea*. The seed color of *S. alba* is however, considerably lighter than in most *Brassica* species, raising the possibility that the yellow-seed trait derived from *S. alba* could further improve the meal and oil quality traits associated with light-seeded rapeseed. The progenies identified in the present study combine the yellow seed trait from *S. alba* with other agronomic properties that could improve the oil content, seed yield and seed quality in *B. napus*, and are therefore of considerable interest for rapeseed breeding.

Sinapis alba is phylogenetically close to Brassica A and C genomes (Warwick and Black 1991), and has sufficient genetic homoeology (Gaikward et al. 1996). Nelson et al. (2005) found that 95% of the Brassica DNA fragments could be hybridized strongly to low copy-number sequences in the S. alba genome. Formation of multivalents and chromosome recombination in the progenies derived from B. napus and S. alba hybrid (Wang et al. 2005c) is also indicative of intergenomic homoeology between both S. alba and A/C genomes. The phenotypic variation presented herein with different agronomical characteristics from BCF₄ progenies to BCF₅ exhibited almost stable. One reason is these progenies had a very good fertility and a high seedset. The other reason is these progenies had 38 chromosomes and exhibited a normal meiosis (data not shown). How to identify these introgression lines, especially with yellow seeds, using molecular markers and to use this valuable germplasm for rapeseed breeding is our present task.

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References

- Bodnaryk RP, Lamb RJ (1991) Mechanisms of resistance to the flea beetle, *Phyllotreta cruciferae* (Goeze), in yellow mustard seedlings, *Sinapis alba* L. Can J Plant Sci 71:13– 20
- Brown J, Brown AP, Davis JB, Erickson D (1997) Intergeneric hybridization between *Sinapis alba* and *Brassica napus*. Euphytica 93:163–168
- Cruickshank RH (1983) Distinction between *Sclerotinia* species by their pectic zymograms. Trans Br Mycol Soc 80:117–119
- Downey RK (1987) Rapeseed and mustard. In: Fehr WR (ed) Principles of cultivar development. Macmillan Publishing Company, New York, pp 437–486

- Gaikward K, Kirti PB, Sharma A, Prakash S, Chopra VL (1996) Cytogenetical and molecular investigations on somatic hybrids of *Sinapis alba* and *Brassica juncea* and their backcross progeny. Plant Breed 115:480–483
- Ge YM, Chen LP (2004) Progress of plant cell engineering germplasm enhancement of Cruciferae. Chin J Cell Biol 26:471–474
- Gong ZH, He YK, Wang M (1994) Studies on the resistance of intergeneric hybrids cabbage × white mustard to Alternaria leaf spot. Acta Hort Sin 21:401–403
- Hansen LN, Earle ED (1997) Somatic hybrids between Brassica oleracea L. and Sinapis alba L. with resistance to Alternaria brassicae (Berk.) Sacc. Theor Appl Genet 94:1078–1085
- Hu SY, Xu LY (1990) A cytochemical technique for demonstration of lipids, polysaccharides and protein bodies in thick resin sections. Acta Bot Sin 32:841–846
- Hu Q, Anderson SB, Dixelius C, Hansen LN (2002) Production of fertile intergeneric somatic hybrids between *Brassica napus* and *Sinapis arvensis* for the enrichment of the rapeseed gene pool. Plant Cell Rep 21:147–152
- Lelivelt CLC, Leunissen EHM, Frederiks HJ, Helsper JPFG, Krens FA (1993) Transfer of resistance to the beet cyst nematode (*Heterodera schachtii* Schm.) from *Sinapis alba* L. (white mustard) to the *Brassica napus* L. gene pool by sexual and somatic hybridization. Theor Appl Genet 85:688–696
- Liu HL (1984) Origin and evolution of some Brassicas. Acta Agron Sin 109:9–18
- Liu J, Xu X, Deng X (2005) Intergeneric somatic hybridization and its application to crop genetic improvement. Plant Cell Tiss Org Cult 82:19–44
- Lu KC, Liu SY, Guo JB (2006) Development of the novel yellow-seeded *Brassica napus* germplasm through the interspecific crosses *B. juncea* × *B. napus*. J Hunan Agric Univ 32:116–119
- Nelson MN, Nixon J, Lydiate DJ (2005) Genome-wide analysis of the frequency and distribution of crossovers at male and female meiosis in *Sinapis alba* L. (white mustard). Theor Appl Genet 111:31–34
- Niu YZ, Guo SX, Fu SH (2005) Development of a speciallylong pod variant line from resynthesized *Brassica napus* L. J Plant Genet Res 6:151–155
- Prakash S, Bhat SR, Quiros CF, Kirti PB, Chopra VL (2009) Brassica and its close allies: cytogenetics and evolution. Plant Breed Rev 31:21–187
- Ren JP, Dickson MH, Earle ED (2000) Improved resistance to bacterial soft rot by protoplast fusion between *Brassica rapa* and *B. oleracea*. Theor Appl Genet 100:810–819
- Roy NN (1984) Interspecific transfer of *Brassica juncea*-type high blackleg resistance to *Brassica napus*. Euphytica 33:295–303
- Snowdon RJ, Lühs W, Friedt W (2006) Oilseed rape. In: Kole C (ed) Genome mapping and molecular breeding, vol 2: oilseeds. Springer Verlag, Heidelberg, pp 55–114
- Wang HZ (2005) Problem in the development of oilseed industry and it's countermeasure in China. Chin J Oil Crop Sci 27:100–105
- Wang AY, Li X, Hu DY (2005a) Research advances on distant hybridization breeding in rapeseed. Acta Agriculturae Boreali-occidentalis Sin 14(6):67–71

- Wang YP, Sonntag K, Rudloff E, Chen JM (2005b) Intergeneric somatic hybridization between *Brassica napus* and *Sinapis alba*. J Integr Plant Biol 47:84–91
- Wang YP, Zhao XX, Sonntag K, Wehling P, Snowdon RJ (2005c) Behaviour of *Sinapis alba* addition chromosomes in a *Brassica napus* background revealed by genomic in situ hybridisation. Chromosom Res 13:819–826
- Warwick SI, Black LD (1991) Molecular systematics of *Brassica* and allied genera (Subtribe Brassinae, Brassiceae) chloroplast genome and cytodeme congruence. Theor Appl Genet 82:81–92