

# Phenotypic variation of *Plasmodium falciparum* merozoite proteins directs receptor targeting for invasion of human erythrocytes

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**The members of the phylum Apicomplexa parasitize a wide range of eukaryotic host cells. *Plasmodium falciparum*, responsible for the most virulent form of malaria, invades human erythrocytes using several specific and high affinity ligand–receptor interactions that define invasion pathways. We find that members of the *P. falciparum* reticulocyte-binding homolog protein family, PfRh2a and PfRh2b, are expressed variantly in different lines. Targeted gene disruption shows that PfRh2b mediates a novel invasion pathway and that it functions independently of other related proteins. Phenotypic variation of the PfRh protein family allows *P. falciparum* to exploit different patterns of receptors on the erythrocyte surface and thereby respond to polymorphisms in erythrocyte receptors and to evade the host immune system.**

**Keywords:** invasion/malaria/merozoite/*P. falciparum*/targeted gene disruption

## Introduction

*Plasmodium falciparum* belongs to the phylum Apicomplexa, a group characterized by a highly specialized apical complex, including pedunculate organelles known as rhoptries that play a central role in the invasion of host cells. *Plasmodium* species infect a very restricted host range in the bloodstream; this specificity is determined by specific receptor–ligand interactions that occur between invasive forms of the parasite and the host erythrocyte (Butcher *et al.*, 1973; Gratzer and Dluzewski, 1993). This is in contrast to some other members of Apicomplexa, such as *Toxoplasma gondii*, that invade a wide range of cell types from many different species. Despite the host cell specificity of *P. falciparum*, it has developed the ability to invade human erythrocytes using multiple parasite ligand–receptor interactions that have become known as alternative invasion pathways (Dolan *et al.*, 1994; Okoyeh *et al.*, 1999).

Erythrocyte invasion is a multistep process involving specific ligand–receptor interactions that mediate apical reorientation and adhesion of the parasite to the host cell (Gratzer and Dluzewski, 1993). A well-defined ligand–receptor interaction is erythrocyte-binding antigen 175 (EBA-175) and glycophorin A on the red blood cell surface (Dolan *et al.*, 1994; Sim *et al.*, 1994). Other invasion pathways have been defined using mutant erythrocytes and enzyme treatments, that use receptors such as glycophorin B and an unknown receptor termed ‘X’ (Dolan *et al.*, 1994). The interactions of parasite ligands with glycophorin A and B depend on sialic acid present on these receptors (Miller *et al.*, 1977; Howard *et al.*, 1982; Pasvol *et al.*, 1982; Pasvol, 1984). Truncation of EBA-175 in a parasite line highly dependent on sialic acid residues was associated with a switch towards invasion via a sialic acid-independent receptor of unknown identity (Reed *et al.*, 2000). Recently, a number of EBA-175 homologs have been identified, and some bind to the erythrocyte surface in a sialic acid-dependent manner (Mayer *et al.*, 2001; Thompson *et al.*, 2001; Narum *et al.*, 2002). The functions of these proteins in merozoite invasion are yet to be determined.

A second family of high molecular weight proteins that bind to erythrocytes with high affinity has been identified in the related human malaria *Plasmodium vivax* (Galinski *et al.*, 1992) and the rodent malaria *Plasmodium yoelii* (Ogun and Holder, 1996). The *P. vivax* proteins bind selectively to reticulocytes and have been termed reticulocyte-binding protein-1 and -2 (RBP-1 and RBP-2) (Galinski *et al.*, 1992). They are located at the apical end of merozoites and have been hypothesized to play a role in the recognition of reticulocytes by merozoites. Merozoite proteins in other species of *Plasmodium*, that are related to the RBPs of *P. vivax*, have been designated as members of a reticulocyte binding-like or RBL family of adhesive invasion proteins (Rayner *et al.*, 2001; Miller *et al.*, 2002). The *P. falciparum* RBL homologs (PfRh) of the *P. vivax* RBPs and the *P. yoelii* Py235 family have been identified and characterized (Rayner *et al.*, 2000, 2001; Taylor *et al.*, 2001; Triglia *et al.*, 2001; Kaneko *et al.*, 2002). There are four members of the PfRh or NBP protein family, encoded by separate genes, and these have been named *PfRh1*, *PfRh2a*, *PfRh2b* and *PfRh4*, or NBP-1, NBP-2a, NBP-2b and NBP-4 (Rayner *et al.*, 2000, 2001; Triglia *et al.*, 2001; Kaneko *et al.*, 2002). A fifth gene, *PfRh3* (Taylor *et al.*, 2001), has been identified, and it has been suggested to be a pseudogene; however, proteomic analysis shows that it may be expressed in sporozoites (Florens *et al.*, 2002). PfRh1/NBP-1 binds to erythrocytes via a neuraminidase-sensitive and trypsin-resistant receptor, and some evidence suggests that PfRh2b may form a complex with PfRh1 (Rayner *et al.*, 2001).

The *P.yoelii* Py235 family is encoded by perhaps up to 14 genes, and individual merozoites originating from a single schizont transcribe distinct members of this gene family (Preiser *et al.*, 1999). This clonal phenotypic variation has been suggested to provide this murine malaria with a survival strategy in the host by expression of a single but different Py235 protein in each merozoite, allowing the parasite to respond efficiently to variations in the host environment. However, the functional consequence of this transcriptional variation remains unclear.

Here we have analyzed the *P.falciparum* Pfrh proteins and have found a remarkable degree of variant expression in different parasite lines. Using gene disruptions, we have identified a novel erythrocyte invasion pathway, and show that variation in expression of these proteins alters the pattern of receptor usage for invasion of human erythrocytes. This provides a unique strategy for parasite invasion in the face of erythrocyte receptor polymorphism and host immune responses.

## Results

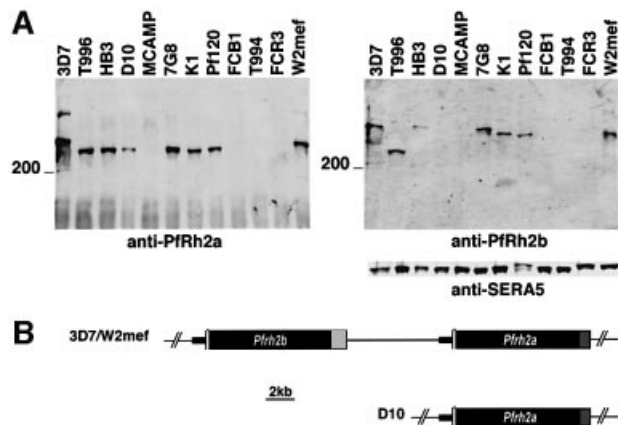
### Variant expression of Pfrh2a and Pfrh2b proteins in multiple *P.falciparum* isolates

The Pfrh family members in *P.falciparum* are large molecular weight proteins, homologous to *P.vivax* PvRBP-1 and -2 (Galinski *et al.*, 2000) and the *P.yoelii* Py235 family (Ogun and Holder, 1996; Preiser *et al.*, 1999). To address the role of Pfrh2a and Pfrh2b in merozoite invasion of *P.falciparum* into erythrocytes, we analyzed expression of these proteins in isolates of *P.falciparum* taken from different geographic areas. Immunoblots of supernatants were probed with antibodies for Pfrh2a or Pfrh2b (Figure 1A) (Rayner *et al.*, 2000). Evidence for variation in expression of these proteins was obtained as Pfrh2a was detectable in 3D7, T996, HB3, D10, 7G8, K1, Pf120 and W2mef, but was not apparent in MCAMP, FCB1, T994 or FCR3. Pfrh2b showed a similar pattern of expression but was absent in the D10 isolate. The SERA5 protein was used as a loading control. These results show that Pfrh2a and Pfrh2b are expressed variably in different isolates, and are in agreement with recent data showing some variation in expression of Pfrh2b in three *P.falciparum* isolates (Taylor *et al.*, 2002).

PCR analysis of genomic DNA from the different isolates 3D7, T996, HB3, 7G8, K1, Pf120, W2mef, MCAMP, FCB1, T994 and FCR3 detected *Pfrh2a* and *Pfrh2b* genes in all isolates, with the exception of D10, which lacks *Pfrh2b* (Triglia *et al.*, 2001). The *P.falciparum* isolates MCAMP, FCB1, T994 and FCR3 do not express Pfrh2a or Pfrh2b protein despite having both intact genes within their genomes (data not shown). To determine whether the variability in expression had any effect on merozoite invasion, we constructed gene disruptions in parasite lines in which they normally are expressed.

### The Pfrh2a and Pfrh2b genes are in a head to tail configuration on chromosome 13

To facilitate gene disruptions of the *Pfrh2a* and *Pfrh2b* genes, we characterized their chromosomal organization. The *Pfrh2a* and *Pfrh2b* genes are identical for their first 7.5 kb, but diverge greatly at their C-termini, sharing only



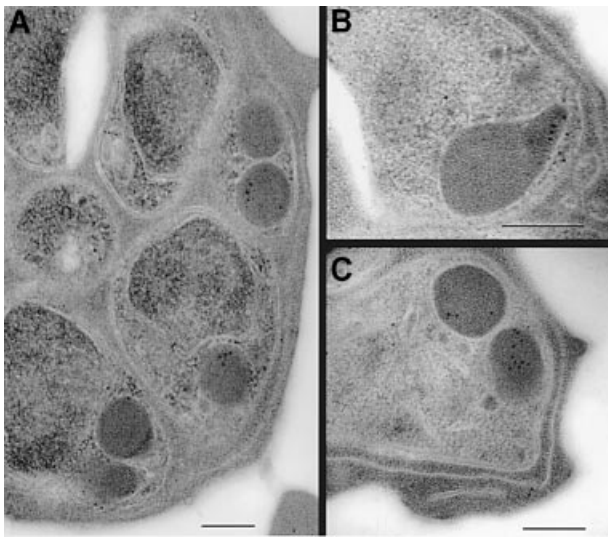
**Fig. 1.** Variant expression, subcellular localization and structure of the Pfrh2a and Pfrh2b genes and proteins in *P.falciparum*. (A) Supernatants from 3D7, T996, HB3, D10, MCAMP, 7G8, K1, Pf120, FCB1, T994, FCR3 and W2mef were probed with anti-Pfrh2a, anti-Pfrh2b or anti-SERA5 antibodies. The position of the 200 kDa molecular weight marker is shown. (B) Structure of the *Pfrh2a* and *Pfrh2b* locus in 3D7, W2mef and D10. The regions in black are identical for the *Pfrh2b* and *Pfrh2a* genes. The light gray (Pfrh2b) and darker gray (Pfrh2a) sections are non-homologous.

a putative membrane-spanning region and a cytoplasmic tail (Figure 1B) (Rayner *et al.*, 2000; Triglia *et al.*, 2001). Using Southern blots and PCR experiments, we showed that *Pfrh2b* and *Pfrh2a* are arranged head to tail in both 3D7 and W2mef, with *Pfrh2b* at the 5' end (Figure 1B). They are separated by ~7.0 kb of sequence, with no intervening coding regions. Southern blots of the D10 *Pfrh2a* gene showed that it is present in a single copy, with the 5'-untranslated region identical to the equivalent region of the 3D7 *Pfrh2b* gene. This suggests that *Pfrh2b* was deleted from the D10 parasite line by an intergenic recombination between the two highly similar 5' ends of *Pfrh2a* and *Pfrh2b*.

### The Pfrh2a and b proteins are located in the neck of the rhoptries of merozoites

To define the subcellular localization of the Pfrh2a/b proteins, we performed immunogold-electron microscopy with the anti-Pfrh2a/b antibodies (Figure 2). First we determined the subcellular localization of Pfrh2a/b proteins in schizont stages of 3D7 parasites. Previous immunofluorescence experiments co-localizing the Pfrh2a/b proteins with rhoptry proteins such as rhoptry-associated protein 1 (RAP1), Pf140 and Pf240/225 suggested that they are located towards the apical end of the merozoite, and co-localization with Pf240/225 but not exactly with RAP-1 or Pf140 suggested perhaps a location in the peduncle of the rhoptries (Rayner *et al.*, 2000, 2001; Triglia *et al.*, 2001).

Immunoelectron microscopy with anti-Pfrh2a/b antibodies showed labeling in the electron-dense rhoptry organelles of the parasite (Figure 2A and C), with label evident in the neck of the rhoptry in some parasites (Figure 2B). The invasion proteins AMA-1 and EBA-175 were found to be present predominantly in the micronemes of parasites from the same preparation (Healer *et al.*, 2002). Thus it appears that Pfrh2b is positioned more apically than micronemal proteins. In 3D7Δ2b parasite

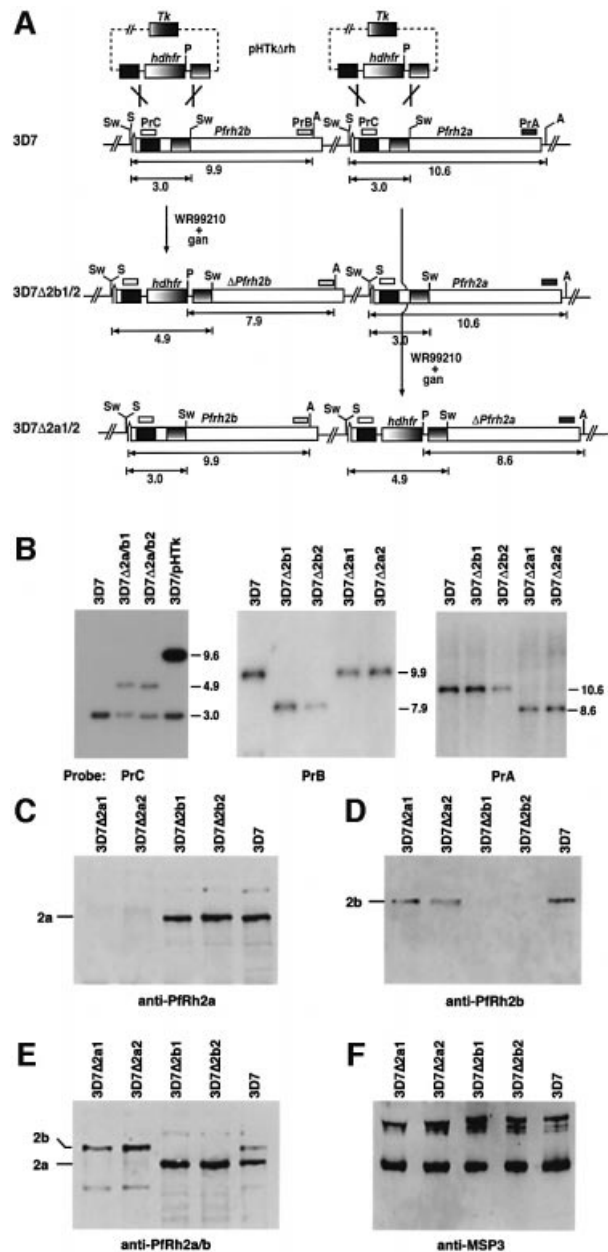


**Fig. 2.** Subcellular localization of Pfrh2a/b in 3D7 parasites by immunoelectron microscopy. (A–C) Cross-sections through merozoites. Scale bars are 200 nm in all panels.

lines lacking Pfrh2b expression (described below), Pfrh2a localization was unaltered (data not shown). Immunofluorescence experiments with anti-Pfrh2a/b antibodies in 3D7Δ2a (described below, lacks Pfrh2a) and 3D7Δ2b (lacks Pfrh2b) parasites showed that localization of Pfrh2a and Pfrh2b in the corresponding Pfrh2a-null or Pfrh2b-null parasites remained unchanged relative to the RAP1 protein (Baldi *et al.*, 2000) (data not shown). These data show definitively that the subcellular localizations of the Pfrh2a and b proteins are not dependent on each other and that they are both localized predominantly in the body and neck of the rhoptries in *P. falciparum* merozoites.

#### **Pfrh2a and Pfrh2b genes can be disrupted**

In order to derive 3D7 parasite lines that lack expression of either Pfrh2a or Pfrh2b proteins, we constructed a plasmid (pHTkΔrh) that would integrate into either gene by double recombination crossover (Figure 3A) (Duraisingh *et al.*, 2002). Integration of pHTkΔrh into 3D7 chromosomes was confirmed by Southern blotting, and four cloned lines were derived in which either *Pfrh2a* or *Pfrh2b* was disrupted (Figure 3B). Hybridization of a PrC probe (Figure 3A) to genomic DNA from 3D7 revealed a hybridizing fragment of 3.0 kb corresponding to the endogenous *Pfrh2a* and *Pfrh2b* genes. In parasites that maintained the pHTkΔrh vector episomally (called 3D7/pHTk parasites), a corresponding 9.6 kb plasmid fragment was evident in addition to the endogenous 3.0 kb fragment. Two clones were derived, 3D7Δ2a/b1 and 3D7Δ2a/b2, that retained the 3.0 kb fragment but also showed a second band of 4.9 kb. This indicated that the transfection plasmid had integrated into either the *Pfrh2a* or the *Pfrh2b* genes. Hybridization with PrB or PrA (which are specific to the *Pfrh2b* or *Pfrh2a* genes, respectively) to genomic DNA from 3D7 and the four cloned lines showed that *Pfrh2b* was disrupted in



**Fig. 3.** Disruption of the *Pfrh2b* and *Pfrh2a* genes in 3D7 parasites. (A) The pHTkΔrh plasmid has the human *dhfr* gene (*dhfr*) flanked by two target sequences for homologous recombination within the *Pfrh2a* and *Pfrh2b* genes. It also includes the thymidine kinase gene (*Tk*) for negative selection with ganciclovir. 3D7Δ2b1/2 and 3D7Δ2a1/2 are the cloned transfected lines showing the structure of the double recombination crossovers. The *Pfrh2a* and *Pfrh2b* genes are shown with relevant restriction enzyme sites: Sw, *Swa*I; S, *Sac*I; A, *Ava*II; P, *Pvu*II. The sizes are in kbp. (B) Southern blotting of genomic DNA from 3D7, 3D7Δ2a/b1 and 3D7Δ2a/b2 (uncloned population transfected with pHTkΔrh), 3D7/pHTk (uncycled parasites transfected with pHTkΔrh), 3D7Δ2b1 and 3D7Δ2b2 (cloned lines), 3D7Δ2a1 and 3D7Δ2a1/a2 (cloned lines). The left panel was digested with *Pvu*I and *Swa*I, whilst the other panels were digested with *Sac*I, *Ava*II and *Pvu*II. (C–F) Pfrh2a and Pfrh2b are not expressed in the parasites with the *Pfrh2a* and *Pfrh2b* genes, respectively, disrupted. Supernatants from 3D7Δ2a1, 3D7Δ2a2, 3D7Δ2b1, 3D7Δ2b2 and 3D7 were probed with the relevant antisera: (C) anti-Pfrh2a sera; (D) anti-Pfrh2b sera; (E) affinity-purified anti-Pfrh2a/2b antibodies; (F) anti-MSP3 antibodies.

**Table I.** Invasion of transfected parasites into enzyme-treated erythrocytes

Erythrocyte treatment <sup>a</sup>	Parasite <sup>b</sup>					
	3D7	3D7Δ2b1	3D7Δ2b2	3D7Δ2a1	3D7Δ2a2	D10
Low trypsin/chymotrypsin	22.8 ± 7.6	70.5 ± 7.7 <sup>c</sup>	65.5 ± 12.9 <sup>c</sup>	18.1 ± 4.8	14.1 ± 2.6	58.0 ± 5.8 <sup>c</sup>
Neuraminidase/low trypsin	23.0 ± 6.9	3.9 ± 0.6 <sup>c</sup>	3.3 ± 0.4 <sup>c</sup>	24.5 ± 6.0	21.9 ± 4.9	10.26 ± 2.3 <sup>c</sup>
Chymotrypsin	67.0 ± 9.1	105.3 ± 9.4 <sup>c</sup>	113.9 ± 14.3 <sup>c</sup>	70.1 ± 10.1	65.3 ± 15.1	99.8 ± 5.9 <sup>c</sup>
Low trypsin	93.5 ± 9.0	63.3 ± 7.8 <sup>c</sup>	61.7 ± 9.7 <sup>c</sup>	80.8 ± 6.7	93.0 ± 14.0	85.9 ± 7.3
Neuraminidase	79.2 ± 8.5	58.2 ± 8.7 <sup>c</sup>	47.5 ± 6.3 <sup>c</sup>	73.9 ± 7.1	82.7 ± 14.9	74.3 ± 11.4

<sup>a</sup>Red blood cells were treated with the enzymes indicated, as described in Materials and methods, before testing in merozoite invasion assays.

<sup>b</sup>Figures are the percentage invasion compared with invasion of 3D7 into untreated blood cells, and errors are 95% confidence limits. The data were from 6–14 independent experiments all carried out in triplicate. In the experiments, the starting parasitemia was 0.5% and the final parasitemia was between 3 and 6% for untreated erythrocytes.

<sup>c</sup>Significantly different invasion when compared with 3D7.

3D7Δ2b1 and 3D7Δ2b2, whereas *PfRh2a* was disrupted in 3D7Δ2a1 and 3D7Δ2a2.

To determine whether disruption of *PfRh2a* and *PfRh2b* resulted in lack of expression of the corresponding protein, we used western blots of supernatants probed with antibodies against each protein (Figure 3C–E). Antibodies to the C-terminus of Pfrh2a (Rayner *et al.*, 2000) revealed two proteins >200 kDa in 3D7, 3D7Δ2b1 and 3D7Δ2b2, whereas these proteins were absent in 3D7Δ2a1 and 3D7Δ2a2 (Figure 3C). This is consistent with disruption of the *PfRh2a* gene in both 3D7Δ2a1 and 2. Similarly, antibodies to the C-terminus of Pfrh2b (Rayner *et al.*, 2000) showed a major band that was also >200 kDa in 3D7, 3D7Δ2a1 and 2, whereas it was absent from 3D7Δ2b1 and 2 (Figure 3D). Using the anti-PfRh2a/b (2A9) antibodies that detect both Pfrh2a and Pfrh2b (Triglia *et al.*, 2001), we confirmed that the corresponding proteins were absent from the recombinant parasite lines (Figure 3E). These results clearly show that Pfrh2a and Pfrh2b proteins are not expressed in 3D7Δ2a1/2 or 3D7Δ2b1/2, respectively. Western blots with schizont pellets of each parasite line gave similar results and confirmed the lack of expression of each protein in the parasite cloned lines (data not shown). The level of expression of the intact *PfRh* gene in each knockout line appeared unchanged when tested in multiple western blot experiments compared with the 3D7 wild-type strain and the control protein MSP3 (Figure 3F). This suggested that integration of the transfection cassette into the neighboring gene had no apparent effect on expression of the adjacent gene.

#### ***PfRh2b* function is required for a novel invasion pathway in *P.falciparum***

Having constructed 3D7 recombinant parasite lines that lack expression of Pfrh2a or Pfrh2b, we initially tested the effect of loss of these proteins on the ability of merozoites to invade untreated human erythrocytes (Dolan *et al.*, 1990, 1994). The growth and merozoite invasion rate of 3D7ΔRh2a1 and 2, and 3D7ΔRh2b1 and 2 parasites was unchanged relative to their 3D7 parent (data not shown). This suggested either that the Pfrh2a and Pfrh2b proteins are not required for merozoite invasion or that the function of Pfrh2a and Pfrh2b is compensated for by other parasite ligands in the knockout parasite lines. This has been described previously for the parasite ligand

EBA-175 which binds to glycoporphin A and mediates an invasion pathway via this receptor (Reed *et al.*, 2000). Loss of function of EBA-175 results in a switch in merozoite invasion to a sialic acid-independent pathway, and this parasite invades and grows as well as parental parasites.

To examine the role of the Pfrh2a and Pfrh2b proteins in erythrocyte invasion further, we determined the effect of different enzyme treatments on the ability of the parental parasites (3D7) to invade protease- or neuraminidase-treated erythrocytes (Table I). Neuraminidase removes sialic acid residues from the erythrocyte surface and blocks invasion pathways dependent on sialic acid present on both glycoporphin A and other receptors (Miller *et al.*, 1977). Trypsin treatment cleaves proteins such as glycoporphin A and C, but does not affect glycoporphin B (Thompson *et al.*, 2001). Chymotrypsin cleaves a non-overlapping set of proteins including glycoporphin B and band 3 on the erythrocyte surface. The different enzyme treatments have the effect of limiting the repertoire of receptors on the erythrocyte, allowing dissection of specific ligand–receptor interactions. This is evident from the effect of different enzyme treatments on erythrocytes and the decreased ability of 3D7 to invade many of these treated cells (Table I).

The 3D7Δ2b1 and 3D7Δ2b2 parasites, lacking expression of Pfrh2b, show a dramatically altered invasion phenotype when compared with 3D7 (Table I). The 3D7Δ2b parasites invade both neuraminidase- and trypsin-treated erythrocytes at a significantly lower efficiency than the 3D7 parent. This reflects a shift in dependency in the 3D7Δ2b parasites from 3D7 parasites using neuraminidase-resistant (sialic acid-independent) and trypsin-resistant receptors, towards using neuraminidase-sensitive (sialic acid-dependent) and trypsin-sensitive receptors in 3D7Δ2b parasites, upon disruption of Pfrh2b. Moreover, when double combinations of enzyme treatments of erythrocytes were used, it was found that the 3D7Δ2b1/2 parasites invade erythrocytes treated with neuraminidase/trypsin at a much lower level compared with 3D7. This treatment appears to limit the erythrocyte receptors available to the parasite, leaving a residual neuraminidase-resistant/trypsin-resistant receptor that can be utilized by a Pfrh2b-dependent invasion pathway in 3D7 (23% invasion) but can no longer be used by 3D7Δ2b1/2 parasites that lack Pfrh2b (3.6% invasion) (Table I;

Figure 6A). Thus, it can be inferred that the receptor for PFRh2b is neuraminidase resistant (sialic acid independent) and trypsin resistant from the PFRh2b-dependent 3D7 invasion of neuraminidase/trypsin-treated cells.

In contrast, the 3D7Δ2b1/2 parasites invade erythrocytes treated with chymotrypsin better than the 3D7 parent, indicating that the loss of the PFRh2b ligand in the null parasites has led to an increased dependency on ligands that utilize chymotrypsin-resistant receptors for invasion. It can therefore be inferred that the cognate receptor for PFRh2b that is used for invasion in wild-type 3D7 is chymotrypsin sensitive (67% invasion compared with 100% for 3D7Δ2b1/2, which cannot use PFRh2b) (Table I; Figure 6A). The reliance of merozoite invasion on this chymotrypsin-sensitive receptor can be increased by limiting the available receptors on the erythrocyte surface as a result of additional treatment with trypsin. 3D7Δ2b parasites invade trypsin/chymotrypsin-treated erythrocytes at least 3-fold better than 3D7 (Table I). To confirm this, an independent 3D7 transfectant with *PfRh2b* disrupted was obtained, and a clone derived from this population gave the same merozoite invasion phenotype as that seen for 3D7Δ2b1/2 (data not shown). Taken together, these data show that PFRh2b functions in a dominant invasion pathway in 3D7 that utilizes an erythrocyte receptor that is chymotrypsin sensitive and neuraminidase/trypsin resistant (Figure 6A). The loss of PFRh2b function leads to an increased reliance on other invasion pathways.

As further evidence that loss of PFRh2b expression in 3D7Δ2b1 and 2 was responsible for the invasion phenotype observed, we tested the ability of D10 merozoites that lack the *PfRh2b* gene (Triglia *et al.*, 2001) to invade enzyme-treated erythrocytes (Table I). Comparison of D10 invasion with 3D7 showed striking similarities to that observed with 3D7Δ2b1 and 2 that also lack PFRh2b. In particular, both the observed ability of D10 merozoites to invade chymotrypsin/trypsin-treated erythrocytes and the impaired invasion of neuraminidase/trypsin-treated cells by D10 lend support to the role of PFRh2b in a merozoite invasion pathway involving a chymotrypsin-sensitive and neuraminidase/trypsin-resistant receptor, in multiple parasite lines from different geographical locations.

Next we tested the effect of loss of expression of PFRh2a on merozoite invasion. A comparison of the ability of 3D7 and 3D7Δ2a merozoites to invade the different enzyme-treated erythrocytes showed no apparent differences (Table I). This was in contrast to the dramatic changes seen with loss of PFRh2b expression. PFRh2a is expressed in 3D7, but its loss of expression was not associated with a measurable shift in reliance to other invasion pathways. Interestingly, attempts to disrupt the *PfRh2a* gene in D10, a parasite that lacks the *PfRh2b* gene, were not successful, suggesting that the *PfRh2a* protein fulfills an important function in this parasite line, which lacks PFRh2b protein (data not shown).

#### **Inhibition of PFRh2b function using specific antibodies impedes merozoite invasion via the PFRh2b invasion pathway**

To dissect the function of PFRh2a/b in merozoite invasion further, we used antibodies specific for the conserved regions of PFRh2a/b to test their ability to inhibit invasion

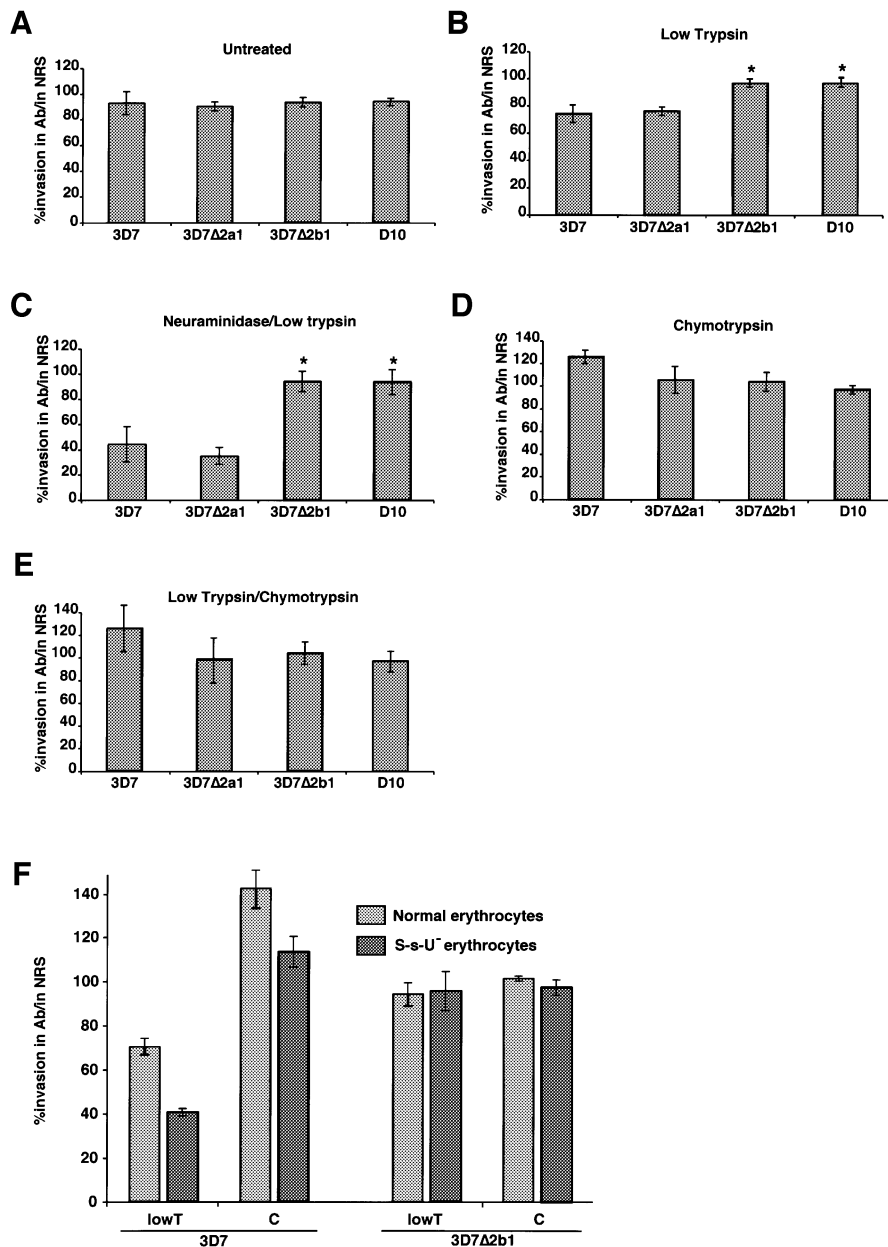
into enzyme-treated erythrocytes in parasites in which the different genes were disrupted (Figure 4). Invasion of 3D7, 3D7Δ2a, 3D7Δ2b and D10 parasites into untreated erythrocytes showed no significant inhibition using these antibodies (Figure 4A). This result was not consistent with previous data suggesting that antibodies to either PFRh2a or PFRh2b can inhibit 3D7 merozoite invasion of untreated erythrocytes (Rayner *et al.*, 2001; Triglia *et al.*, 2001). Our results using parasites with specific *PfRh2a/b* gene disruptions allow specificity controls (Figure 4) that have not been possible previously.

The anti-PFRh2a/b antibodies, however, did show significant inhibition when the potential receptors available on the erythrocyte surface for merozoite invasion were restricted using enzyme-treated erythrocytes. Merozoite invasion of 3D7 into trypsin- and neuraminidase/trypsin-treated erythrocytes was significantly decreased in the presence of anti-PFRh2a/b antibodies (Figure 4B and C). This antibody inhibition was due specifically to blockage of PFRh2b function, as 3D7Δ2b1 parasites, that did not express the PFRh2b protein, showed no antibody-mediated inhibition of invasion into either untreated or enzyme-treated erythrocytes (Figure 4A–C). The level of invasion in the presence of anti-PFRh2b antibodies by 3D7 (43%) versus 3D7Δ2b1 (95%) into neuraminidase/trypsin-treated erythrocytes is consistent with the conclusion that the PFRh2b receptor is neuraminidase and trypsin resistant.

To confirm that PFRh2b functions via a chymotrypsin-sensitive receptor, we tested direct inhibition of this parasite protein using the anti-PFRh2a/b antibodies in a merozoite invasion assay (Figure 4). These antibodies showed no inhibition of merozoite invasion of either 3D7 or 3D7Δ2b1 into chymotrypsin-treated erythrocytes (Figure 4D). Significantly, no inhibition of invasion was seen of either 3D7 or 3D7Δ2b1 into trypsin/chymotrypsin-treated erythrocytes, suggesting that chymotrypsin treatment had removed the PFRh2b receptor, blocking all function of the PFRh2b protein (Figure 4D and E). This also confirms that the antibodies are directly inhibiting the function of the PFRh2b protein rather than acting by non-specific steric hindrance. These results strongly support our data showing that PFRh2b acts in a merozoite invasion pathway using a chymotrypsin-sensitive receptor on the erythrocyte surface.

#### ***PfRh2b* functions in a novel merozoite invasion pathway**

The receptor utilized by PFRh2b is trypsin resistant and chymotrypsin sensitive, and a potential candidate on the erythrocyte was glycophorin B. To test whether PFRh2b utilizes glycophorin B, we used erythrocytes from an S-s-U<sup>-</sup> individual who lacks this protein on their red blood cells (Figure 4F). The PFRh2a/b antibodies significantly inhibited invasion by 3D7 merozoites into trypsin-treated erythrocytes; however, as expected, there was no inhibition of 3D7Δ2b parasites. Inhibition of 3D7 invasion into trypsin-treated erythrocytes by the antibodies was ablated by pre-treatment with chymotrypsin, consistent with removal of the receptor. These results show that PFRh2b function during merozoite invasion is not via glycophorin B as a receptor. The physical properties of the receptor have not been described previously for other defined



**Fig. 4.** Specific antibody inhibition of PfRh2b function in 3D7, 3D7Δ2a1, 3D7Δ2b1 and D10. Assays measure the percentage merozoite invasion in the presence or absence of anti-PfRh2a/b IgG antibodies relative to controls with IgG from normal rabbit sera (NRS). The erythrocytes in each panel are: (A) untreated; (B) trypsin treated; (C) neuraminidase/trypsin treated; (D) chymotrypsin treated; and (E) trypsin/chymotrypsin treated. (F) Assays measure the percentage merozoite invasion into S-s-U<sup>-</sup> erythrocytes that lack glycoprotein B. Error bars in all cases show 95% confidence limits from 4–10 experiments in triplicate.

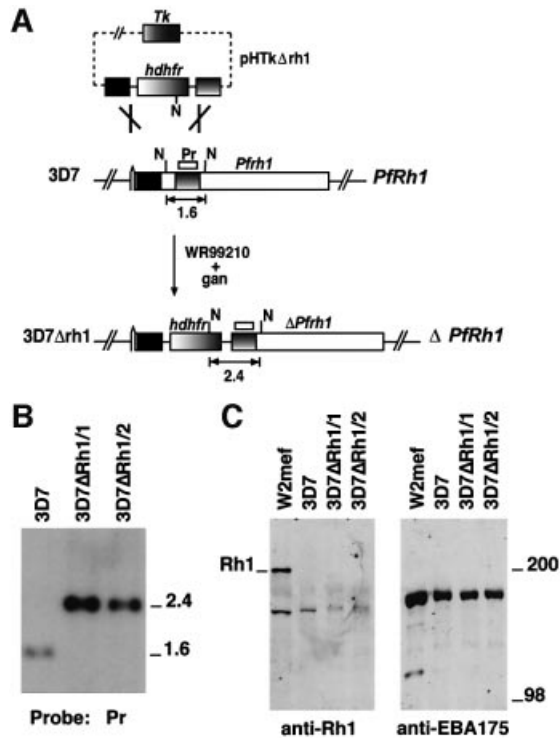
erythrocyte receptors (Dolan *et al.*, 1994; Rayner *et al.*, 2001), and therefore represent a novel merozoite invasion pathway, which we refer to as receptor Z.

#### **The function of PfRh2b is independent of PfRh1**

The PfRh2a and PfRh2b proteins are part of a family that includes PfRh1, and it has been suggested previously that PfRh2b forms a complex with PfRh1 that functions in merozoite invasion (Rayner *et al.*, 2001). To analyze the role of PfRh1, we raised antibodies to different regions of this protein and used them in western blots to detect the protein (Figure 5C). No specific protein bands were observed in 3D7 with antibodies to PfRh1, although a

major protein of ~190 kDa was observed in supernatants from another *P. falciparum* strain W2mef. This contrasted with EBA-175 that was detected in both parasite lines. Identical experiments with schizont pellets gave similar results. These results show that the amount of PfRh1 expressed in 3D7 is below detectable levels, suggesting that the PfRh1 parasite ligand is not functional in merozoite invasion of this parasite line.

To exclude the possibility that PfRh1 was expressed at low levels in 3D7 and still played a role in merozoite invasion of erythrocytes, we made constructs for transfection of 3D7 to disrupt the *PfRh1* gene. The pHTkΔrh1 vector was transfected into 3D7 and, after selection on



**Fig. 5.** Disruption of the *Pfrh1* gene in 3D7. (A) Pr represents the probe used for Southern hybridization in (B). The restriction enzyme shown as N is *NsiI*. pHTk $\Delta$ rh1 has integrated into *Pfrh1* by a double recombination crossover event to yield 3D7 $\Delta$ Rh1. (B) Southern hybridization of genomic DNA digested with *NsiI* from 3D7, 3D7 $\Delta$ Rh1/1 and 3D7 $\Delta$ Rh1/2. Sizes on the right are in kb. (C) Western blot of supernatants from W2mef, 3D7, 3D7 $\Delta$ Rh1/1, 3D7 $\Delta$ Rh1/2 with antibodies to PFRh1 (first panel) and EBA175 (second panel).

WR99210 and ganciclovir, parasites were obtained that had integrated the vector (Duraisingh *et al.*, 2002). Southern blot experiments confirmed that the *Pfrh1* gene had been disrupted by integration of the pHTk $\Delta$ rh1 vector in a double recombination crossover event (Figure 5B) (Duraisingh *et al.*, 2002). Restriction mapping of this locus in two cloned lines 3D7 $\Delta$ Rh1/1 and 2 confirmed the structure of the integration event and disruption of the *Pfrh1* gene.

Western blot analysis of 3D7 $\Delta$ Rh1/1 and 2 with antibodies to the PFRh1 protein showed identical patterns to those seen in the 3D7 parental line, suggesting that this protein was not expressed (Figure 5C). Additionally, we determined whether the PFRh1 protein could be detected in the 3D7 $\Delta$ 2b parasites using western blots. No PFRh1 protein could be detected in these parasites, suggesting that the altered invasion phenotypes of the 3D7 $\Delta$ 2b were not due to increased expression of PFRh1 (data not shown). To confirm further that disruption of PFRh1 had no effect on merozoite invasion, we analyzed the ability of 3D7, 3D7 $\Delta$ Rh1/1 and 2 merozoites to invade untreated and enzyme-treated erythrocytes. The three parasite lines showed identical abilities to invade these cells, suggesting that PFRh1 was not functional in 3D7, 3D7 $\Delta$ Rh1 or 2 (data not shown). Therefore, the PFRh2b protein functions in a merozoite invasion pathway independent of PFRh1 expression and function.

## Discussion

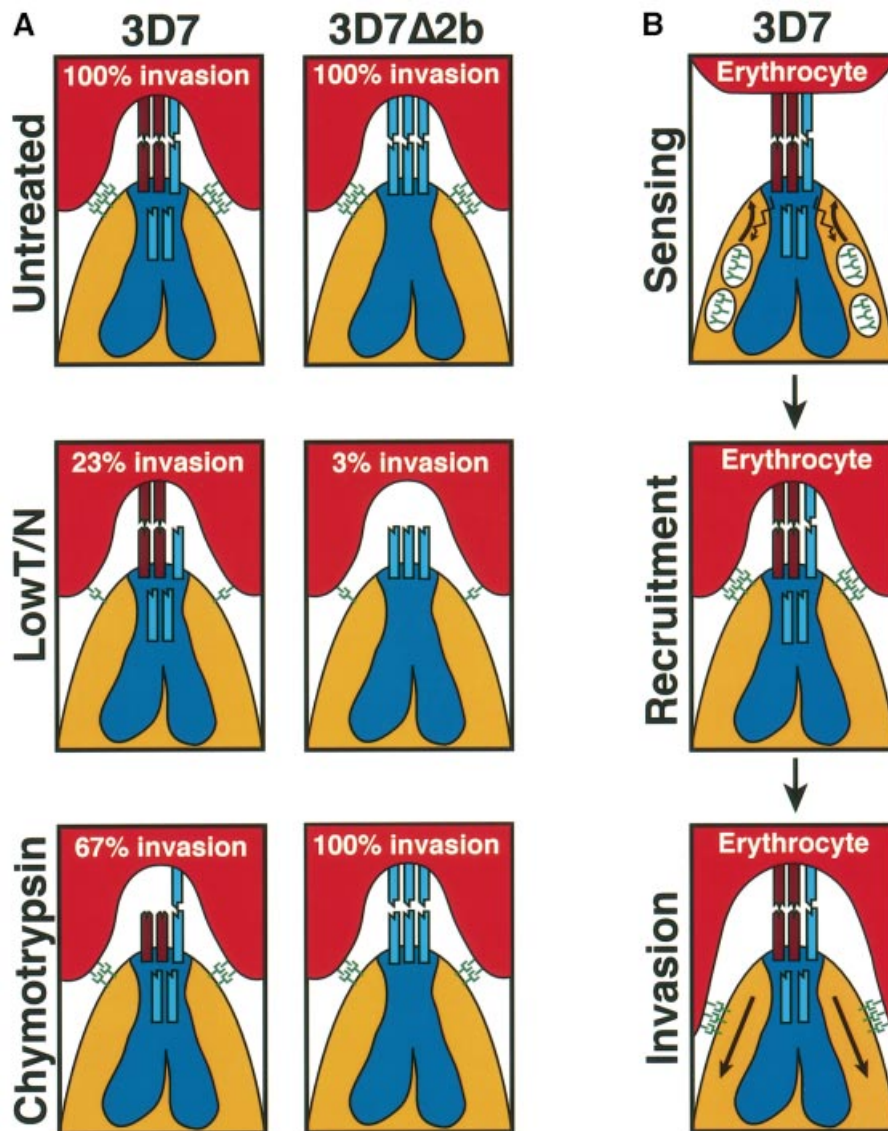
The availability of the *P. falciparum* genome sequence has allowed the identification of two adhesive families of parasite proteins, namely the EBA (Adams *et al.*, 2001) and PFRh proteins (Rayner *et al.*, 2000, 2001; Taylor *et al.*, 2001; Triglia *et al.*, 2001; Kaneko *et al.*, 2002), that may play an important role in determining the pattern of receptor usage on the erythrocyte surface. Here we have used specific gene knockouts to reveal that a member of the PFRh protein family, PFRh2b, functions to define an alternative invasion pathway (Figure 6). Moreover, we find that the PFRh proteins vary in their expression in different parasite lines to provide a mechanism of phenotypic variation utilizing alternative receptors for invasion of merozoites into human erythrocytes. The availability of different parasite ligands for merozoite invasion alters the pattern of receptor utilization, enabling *P. falciparum* parasites to increase the chance of successful invasion in the face of erythrocyte receptor polymorphism and host immune responses.

### Nature of the receptor for the PFRh2b invasion pathway

The PFRh2b ligand mediates invasion through a chymotrypsin-sensitive, trypsin/neuraminidase-resistant receptor, which we refer to as receptor Z (Figure 6A). *Plasmodium falciparum* is known to utilize several receptors on the erythrocyte surface for merozoite invasion, including glycoporphins A, B, C and an unknown receptor X (Dolan *et al.*, 1990, 1994; Mayer *et al.*, 2001; Thompson *et al.*, 2001; Narum *et al.*, 2002). Since receptor Z is trypsin resistant, it cannot correspond to glycoporphins A, C or receptor X, all of which are trypsin sensitive. The putative receptor Y, which binds to the ligand PFRh1, is trypsin resistant but neuraminidase sensitive (Rayner *et al.*, 2001), so is also distinct from receptor Z. Glycophorin B possesses the enzyme sensitivities of receptor Z, but PFRh2b function can be inhibited in merozoite invasion of S-s-U<sup>-</sup> erythrocytes lacking this receptor, demonstrating that it is not utilized by this invasion pathway. This suggests that PFRh2b functions in a merozoite invasion pathway distinct from any described previously.

The PFRh1 protein binds to receptor Y on the erythrocyte surface (Rayner *et al.*, 2001); however, there is no direct evidence that PFRh2b binds directly to receptor Z (Rayner *et al.*, 2000; Triglia *et al.*, 2001). Failure to detect binding in previous assays could be due to either the absence of the binding domain in the processed protein released into the supernatant or low affinity binding that is not detectable by these methods. Since the PvRBP and Py235 homologs of PFRh2b from *P. vivax* (Galinski *et al.*, 1992) and *P. yoelii* (Ogun and Holder, 1996) bind to reticulocytes or normalocytes, respectively, it is therefore likely that PFRh2b also binds directly to receptor Z. However, we cannot rule out the possibility that PFRh2b binds indirectly to receptor Z through an intermediate ligand (Baldi *et al.*, 2000). Importantly, disruption of the *Pfrh2b* gene and demonstration of protein function in merozoite invasion by inhibition with antibody have both provided direct evidence for the role of PFRh2b in a novel invasion pathway.





**Fig. 6.** A simplified model of merozoite invasion into erythrocytes by 3D7 and 3D7Δ2b parasites and the role of the PfRh protein family. **(A)** A model explaining the enzyme sensitivity of receptor Z as trypsin resistant, neuraminidase resistant and chymotrypsin sensitive. The invasion data are from Table I. Top panels: both 3D7 and 3D7Δ2b parasites show 100% invasion of untreated erythrocytes. The PfRh proteins (shown at the apical end of the merozoites and in the neck of the rhoptries) are responsible for sensing the apical end interaction and signaling to the micronemes for release of high affinity ligands (see B). The erythrocyte-binding antigens (EBAs) and their receptors are shown as green symbols. In 3D7, PfRh2b is responsible for the dominant sensing pathway for invasion via the chymotrypsin-sensitive receptor Z (brown). Disruption of the *PfRh2b* gene in 3D7 results in replacement of PfRh2b with an alternative pathway specified by another protein [shown as a blue ligand in the neck of the 3D7 rhoptries and interacting with a receptor (blue) on the erythrocyte in 3D7Δ2b] that binds to a chymotrypsin-resistant receptor. Middle panels: following erythrocyte treatment with trypsin and neuraminidase, 3D7 and 3D7Δ2b give 23 and 3% invasion, respectively. The trypsin and neuraminidase resistance of receptor Z is shown. Bottom panels: following erythrocyte treatment with chymotrypsin, 3D7 and 3D7Δ2b give 67 and 100% invasion, respectively. Some invasion can still be achieved using the alternative parasite ligand shown in the rhoptries that binds to a chymotrypsin-resistant receptor. The chymotrypsin sensitivity of receptor Z is shown. **(B)** Model for the role of the PfRh proteins in sensing the erythrocyte. This would activate release of proteins from micronemes and recruitment at the tight junction, followed by invasion.

The location of the very large PfRh2a/b proteins at the apical end of the merozoite, together with the pivotal role of PfRh2b in an invasion pathway, suggest that the PfRh proteins may be involved in sensing the apical interaction and signaling for subsequent events to form the tight junction (Figure 6) (Galinski *et al.*, 1992; Barnwell and Galinski, 1998). This has also been suggested for the role of the Py235 protein family in *P.yoelii* invasion of erythrocytes (Khan *et al.*, 2001). We speculate that once

apical interaction is established between the merozoite and erythrocyte, the PfRh proteins may recruit high affinity receptors such as EBA-175 by release of micronemal components. Disruption of interactions at the apical end or subsequent recruitment of high affinity receptors by enzyme treatment of erythrocytes and removal of some receptors would have a similar effect on the pattern of merozoite invasion due to blockage of sequential steps in the same process (Figure 6).



### **Compensation for loss of Pfrh2b function by other parasite ligands?**

Binding and entry of *P.falciparum* invasive merozoites into human erythrocytes require specific ligand–receptor interactions. The observed strain variation in the utilization of these invasion pathways is very stable and does not change in the course of routine culture (Dolan *et al.*, 1994; Okoyeh *et al.*, 1999). Merozoites require sufficient affinity of ligand binding to cognate receptors to activate subsequent steps in the invasion process (see Barnwell and Galinski, 1998). It is likely that the area available for interaction between the apical end of the merozoite and the erythrocyte is limited, and therefore the number of ligands occupying that space would also be limited (Figure 6). Even in the absence of Pfrh2b expression, no difference in invasion and growth rates is observed, implying that a sufficient affinity still exists between the remaining ligands and their cognate receptors for efficient invasion of normal erythrocytes. Nevertheless, a large change in receptor usage does result from the loss of Pfrh2b expression, indicating that a significant shift in dependency to other ligands with different erythrocyte receptor specificities has occurred. This shift in receptor usage may either result from the upregulation of ligands, existing or novel, to compensate for the missing Rh2b ligand or, alternatively, be due to the redeployment of existing ligands from a pool containing an excess of ligands. In either case, the apparent shift in the usage of pathways implies that loss of Pfrh2b expression necessitated an increased reliance on other ligands to provide sufficient affinity for efficient invasion (Figure 6).

### **PfRh proteins function independently of each other**

Previous data has suggested that Pfrh2b and Pfrh1 might form a functional complex (Rayner *et al.*, 2001), and this would be consistent with the suggested association of PvRBP1 and PvRBP2 in *P.vivax* (Galinski *et al.*, 1992). However, our data show that Pfrh2b functions in merozoite invasion in the 3D7 parasite line in the absence of Pfrh1 and Pfrh2a expression. Furthermore, immunoprecipitation experiments did not co-precipitate other PfRh proteins using antibodies to Pfrh2b, Pfrh2a and Pfrh1 (data not shown). The functional independence of the PfRh proteins is consistent with the expression of only one *Py235* gene from a large gene family in a single merozoite of *P.yoelii* (Preiser *et al.*, 1999). The role of a fourth member of the PfRh family, Pfrh4 (Kaneko *et al.*, 2002), in merozoite invasion remains to be determined, but it is likely that this protein is involved in specifying a different invasion pathway.

Although Pfrh2b functions in merozoite invasion, we found no invasion role for the closely related protein Pfrh2a in the 3D7 parasite line. Pfrh2a and Pfrh2b are virtually identical for a large part of the N-terminus, and differ at the C-terminus that includes a transmembrane region and a short cytoplasmic tail. Although Pfrh2a does not play an important role in 3D7 parasites, it does not exclude a role in other parasite lines in a different genetic background. Indeed, attempts to disrupt the *PfRh2a* gene in D10 parasites, that lack *PfRh2b*, were unsuccessful, suggesting that the encoded protein plays an important role

in this parasite line. The differences in function between Pfrh2b and Pfrh2a must be specified by the coding divergence in their C-termini. It is possible that the cytoplasmic tail of Pfrh2b is responsible for activating signals for release of micronemes onto the erythrocyte and subsequent steps in tight junction formation (Galinski *et al.*, 1992; Barnwell and Galinski, 1998). Alternatively, the unique region in Pfrh2b N-terminal to the transmembrane domain may be responsible for interacting with a second protein that is involved in receptor binding on the erythrocyte.

### **Variation in expression of PfRh proteins: a selective advantage for P.falciparum**

Different patterns of invasion phenotypes between isolates of *P.falciparum* can be explained in part by variation in expression of the PfRh protein family. This study has identified many parasite isolates that do not express Pfrh2a or Pfrh2b, and, similarly, 3D7 does not express detectable levels of Pfrh1. We have evidence suggesting that there are many parasite isolates that do not express Pfrh1 (data not shown). Loss of expression of the PfRh proteins in any merozoite is compensated by other parasite ligands interacting with available receptors. The presence of non-functional parasite ligands at the apical end of some merozoites, because of blockade by either antibodies or receptor polymorphism, decreases the efficiency of merozoite invasion. It follows that variation in expression of different parasite ligands across parasite populations would overcome this for individual merozoites, since they would lack expression of these non-functional ligands. By concentrating functional ligands at the apical end in place of Pfrh2b, these merozoites could achieve the required affinity for efficient invasion. Obviously, suppression of expression of proteins such as Pfrh2b or Pfrh1 could result in the permanent loss of these proteins within the parasite population. However, genetic recombination during meiosis between frequent mixed infections would constantly generate a pool of parasites that either express or do not express particular PfRh proteins and invasion pathways. Hence, the parasites appear to maintain a varied repertoire of ligands, but probably concentrate on expressing a limited suite at any one time in order to achieve a threshold of binding necessary for invasion.

A number of *P.falciparum* isolates do not express the Pfrh2a or Pfrh2b proteins despite having apparently intact genes within their genomes. The mechanism for lack of expression of these proteins is not yet understood, but there are several possibilities. First, there may be mutations that block transcription or translation. Secondly, it is possible that the genes are silenced, and this would provide a mechanism to switch the expression of these genes on and off, allowing the parasite access to different invasion pathways when required. This would be analogous to the antigenic switching of the *var* gene family of *P.falciparum* (Smith *et al.*, 1995; Su *et al.*, 1995). Expression of the equivalent of the PfRh family in *P.yoelii* (*Py235*) is limited to only one gene per merozoite, suggesting some form of regulated control of this gene family (Preiser *et al.*, 1999).

### Biological significance for *P.falciparum* invasion of human erythrocytes

Multiple parasite ligands and phenotypic variation in expression of the PfRh family in *P.falciparum* have important implications for the parasite. Merozoite invasion requires sufficient ligand–receptor interactions on the erythrocyte surface, and there are circumstances where receptor availability is limited. First, the molecules on the erythrocyte are highly polymorphic in the human population, and this heterogeneity has probably been selected by *P.falciparum* and other infectious agents to protect against severe disease (Miller *et al.*, 2002). The absence or mutation of receptors would limit those available on the erythrocyte for interaction with parasite ligands, and this would decrease invasion efficiency. Examples are populations in East Africa that lack glycophorin B, and Melanesian populations that express a mutant form of glycophorin C on the erythrocyte surface (Patel *et al.*, 2001). Secondly, during development and aging of the human erythrocyte, different receptors such as glycophorin A are not present or vary in their quantity and presentation on the surface of this cell (Issitt and Anstee, 1998). Thirdly, host immune responses to one or more of the parasite ligands involved in merozoite invasion would block utilization of particular alternative invasion pathways, thus limiting the pathways available. Multiple parasite ligands and phenotypic variation of PfRh protein expression provide a mechanism for the parasite to evade these situations by the presence of merozoites within the population that have different dominant invasion pathway specificities, increasing the ability of the parasite to maintain infection and its potential for transmission to subsequent hosts. It will be important to consider the ability of the parasite to vary expression of parasite ligands for merozoite invasion when developing vaccine strategies directed against invasion-related proteins.

## Materials and methods

### Plasmids and parasites

Genomic DNA from the 3D7 parasite line was used as template in the PCR of two ~1 kb fragments corresponding to a region conserved between the *PfRh2a* and *PfRh2b* genes: Flank 1 (5'-GGACCGC-CGCGGAGTCATGAGCATTTTGTGGACAA-3') and Flank 2n (5'-GGACCGGTTAACTTCGTCATGTATATCCAATAGA-3') using the primers 5'-GGACCGGGCGCCCGGGTGATGTATTATGTACAG-CTAAAGC-3' and 5'-GGACCGCCTAGGGAAAAAGCCTCTATGG-ATGAAATG-3'. Similarly, two flanks from the *PfRh1* gene were cloned into a vector derived from pHTk, which had a modified multiple cloning site. Flank 1 was amplified as above using the primers 5'-GATCAC-TAGTTTATGAACCTACCCCTTCAT and 5'-GATCAGATCTATGA-AAGTAGGTTTAATGTA, and Flank 2 was amplified using the primers 5'-GATCGAATTCGATTTACAAGATCTCTTACT and 5'-GATCCC-ATGGTTTGGATATATTTCCCTTA.

*Plasmodium falciparum* asexual stages were maintained in human O+ erythrocytes. 3D7 is a cloned line derived from NF54 and was obtained from David Walliker at Edinburgh University. W2mef is a cloned line derived from the Indochina III/CDC strain. The parasites Tak996 and Tak994 from South-East Asia, and FCB1 of African origin were provided by David Baker from the London School of Hygiene and Tropical Medicine. The FCR3 cloned line is of African origin. HB3 and 7G8 are South American cloned lines, while D10 was cloned from a Papua New Guinean isolate. Malayan Camp, K1 and Pf120 are all South-East Asian lines. Transfection with 80 µg of purified plasmid DNA (Qiagen) and selection for stable transfectants with double recombination crossover were carried out as described previously (Duraisingh *et al.*, 2002).

### SDS-PAGE and immunoblot analysis

Parasite pellets were collected from parasites 40 h after synchronization. Parasite supernatants were obtained from cultures following treatment of the erythrocytes with trypsin (1.0 mg/ml) and neuraminidase (25 mU/ml). Proteins were separated on 6% SDS–polyacrylamide gels. Western blotting onto nitrocellulose (0.45 µm; Schleicher and Schuell, Germany) was performed according to standard protocols.

### Erythrocyte invasion assay

Uninfected erythrocytes were treated with enzymes as described previously (Thompson *et al.*, 2001). The invasion assay was carried out essentially as previously described (Reed *et al.*, 2000). Erythrocytes infected with ring stages were synchronized twice, followed by pre-treatment with trypsin and neuraminidase to prevent reinvasion. Experiments were carried out with the addition of new erythrocytes treated with trypsin (66.7 µg/ml), neuraminidase (66.7 mU/ml) and chymotrypsin (1.0 mg/ml) or a combination of these enzyme treatments (Thompson *et al.*, 2001). Enzyme-treated or normal erythrocytes at 4% hematocrit were inoculated with infected erythrocytes to give a parasitemia of 0.5% and hematocrit of 4% in a volume of 100 µl. Parasites were incubated for 48 h followed by addition of [<sup>3</sup>H]hypoxanthine (Amersham) to 1 µCi/well. After 16 h, the cells were harvested onto glass filters with a cell harvester (Packard). Incorporated [<sup>3</sup>H]hypoxanthine was determined in a scintillation counter. Percentage invasion was calculated in comparison with invasion of the same parasite line into untreated erythrocytes, which was set to 100%.

### Antibody inhibition assay

Antibodies were purified using protein G. Antibody inhibition assays were performed in the same way as the erythrocyte invasion assay except that the hematocrit was kept at 2% with an initial parasitemia of 0.5%. Antibodies were added to a final concentration of 0.5 mg/ml 24 h following the setting up of the assay, prior to reinvasion. Invasion in the presence of specific anti-PfRh2a/b antibodies was compared with invasion in the presence of control antibodies from pre-immune serum.

### Electron microscopy

Immunolabeling for electron microscopy was performed on ultrathin sections of glutaraldehyde-fixed parasites as described previously (Healer *et al.*, 2002). Micrographs were obtained using a Philips CM120 BioTWIN transmission electron microscope at 120 kV.

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