#### University of Wollongong

## **Research Online**

Faculty of Science, Medicine and Health - Papers: part A

Faculty of Science, Medicine and Health

1-1-2014

# Phenylpropanoids and furanocoumarins as antibacterial and antimalarial constituents of the Bhutanese medicinal plant Pleurospermum amabile

Phurpa Wangchuk University of Wollongong, pw54@uowmail.edu.au

Stephen G. Pyne University of Wollongong, spyne@uow.edu.au

Paul Keller University of Wollongong, keller@uow.edu.au

Malai Taweechotipatr Srinakharinwirot University

Sumalee Kamchonwongpaisan BIOTEC, Pathum Thani 12120, Thailand

Follow this and additional works at: https://ro.uow.edu.au/smhpapers

Part of the Medicine and Health Sciences Commons, and the Social and Behavioral Sciences Commons

#### **Recommended Citation**

Wangchuk, Phurpa; Pyne, Stephen G.; Keller, Paul; Taweechotipatr, Malai; and Kamchonwongpaisan, Sumalee, "Phenylpropanoids and furanocoumarins as antibacterial and antimalarial constituents of the Bhutanese medicinal plant Pleurospermum amabile" (2014). *Faculty of Science, Medicine and Health - Papers: part A.* 2190.

https://ro.uow.edu.au/smhpapers/2190

Research Online is the open access institutional repository for the University of Wollongong. For further information contact the UOW Library: research-pubs@uow.edu.au

## Phenylpropanoids and furanocoumarins as antibacterial and antimalarial constituents of the Bhutanese medicinal plant Pleurospermum amabile

### Abstract

With the objective of determining safety and verifying the traditional uses of the Bhutanese medicinal plant, Pleurospermum amabile Craib & W. W. Smith, we investigated its crude extracts and the isolated phytochemicals for their biological activities. Four phenylpropanoids [(E)-isomyristicin (1), (E)-isoapiol (2), methyl eugenol (3) and (E)-isoelemicin (4)] and six furanocoumarins [psoralen (5), bergapten (6), isoimperatorin (7), isopimpinellin (8), oxypeucedanin hydrate (9) and oxypeucedanin methanolate (10)] were isolated from this plant. Among the test samples, compound 10 showed weak antibacterial activity against Bacillus subtilis and best antimalarial activity against the Plasmodium falciparum strains, TM4/ 8.2 (chloroquine and antifolate sensitive) and K1CB1 (multidrug resistant). None of the test samples showed cytotoxicity. This study generated scientific data that support the traditional medical uses of the plant.

### Disciplines

Medicine and Health Sciences | Social and Behavioral Sciences

### **Publication Details**

Wangchuk, P., Pyne, S. G., Keller, P. A., Taweechotipatr, M. & Kamchonwongpaisan, S. (2014). Phenylpropanoids and furanocoumarins as antibacterial and antimalarial constituents of the Bhutanese medicinal plant Pleurospermum amabile. Natural Product Communications: an international journal for communications and reviews, 9 (7), 957-960.

## Phenylpropanoids and Furanocoumarins as Antibacterial and Antimalarial Constituents of the Bhutanese Medicinal Plant *Pleurospermum amabile*

Phurpa Wangchuk<sup>a,b,c</sup>\*, Stephen G. Pyne<sup>b</sup>, Paul A. Keller<sup>b</sup>, Malai Taweechotipatr<sup>d</sup> and Sumalee Kamchonwongpaisan<sup>e</sup>

<sup>a</sup>Queensland Tropical Health Alliance Laboratory, Centre for Biodiscovery and Molecular Development of Therapeutics, Faculty of Medicine, Health & Molecular Sciences, James Cook University, Cairns campus, QLD 4878, Australia

<sup>b</sup>School of Chemistry, University of Wollongong, Wollongong, NSW, 2522, Australia

<sup>c</sup>Manjong Sorig Pharmaceuticals, Ministry of Health, Thimphu, Bhutan

<sup>d</sup>Department of Microbiology, Faculty of Medicine, Srinakharinwirot University, Sukhumvit 23, Bangkok, 10110, Thailand

<sup>e</sup>Medical Molecular Biology Research Unit, National Center for Genetic Engineering and Biotechnology, National Science and Technology Development Agency, Thailand Science Park, Pathumthani, 12120, Thailand

phurpaw@yahoo.com

With the objective of determining safety and verifying the traditional uses of the Bhutanese medicinal plant, *Pleurospermum amabile* Craib & W. W. Smith, we investigated its crude extracts and the isolated phytochemicals for their biological activities. Four phenylpropanoids [(*E*)-isomyristicin (1), (*E*)-isoapiol (2), methyl eugenol (3) and (*E*)-isoelemicin (4)] and six furancoumarins [psoralen (5), bergapten (6), isoimperatorin (7), isopimpinellin (8), oxypeucedanin hydrate (9) and oxypeucedanin methanolate (10)] were isolated from this plant. Among the test samples, compound 10 showed weak antibacterial activity against *Bacillus subtilis* and best antimalarial activity against the *Plasmodium falciparum* strains, TM4/8.2 (chloroquine and antifolate sensitive) and K1CB1 (multidrug resistant). None of the test samples showed cytotoxicity. This study generated scientific data that support the traditional medical uses of the plant.

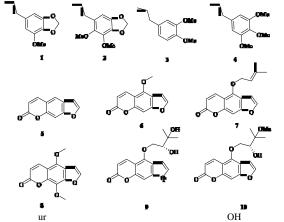
Keywords: Pleurospermum amabile, Bhutanese medicinal plant, Antibacterial, Antimalarial, Furanocoumarins, Phenylpropanoids.

The *Pleurospermum* belong to the family Apiaceae (Umbelliferae) and about 30-50 species are reported from Eastern Europe, north Asia and the Himalayan region [1a-b]. Phytochemicals including essential oils, coumarins, flavonoids, terpenoids, saponins, steroids, fatty acids, esters, glycosides and triterpene saponins were reported

from the few species that were studied [2a-b]. Pharmacologically, the crude extracts, essential oils and the phytochemicals were reported to possess various therapeutic activities including, anticancer, anti-inflammatory, antimalarial, anti-trypanosomal, antibacterial, analgesic, apoptosis inducing, and inhibition of hyperlipidemia and hypercholesterolemia [1a, 2a, 3a-b].

Traditionally, most of these species have been used as vegetables, medicinal plants and herbal perfumes. For example, in Korea, the aerial portion of *P. kamtschaticum* is used as an edible mountain vegetable and for treating colds, arthritis, atherosclerosis and impotence [4]. The root of *P. angelicoides* is used in Himalayan and Chinese folk medicines for treating dysentery and as an antipyretic and a diaphoretic agent [5]. Similarly, *P. densiflorum* is used by the Kumaon Himalayan people (Indian) as one of the best herbal perfumes in the region and is known for its long lasting pleasant smell [6].

In Bhutan, out of the eight reported species of *Pleurospermum* [7], *P. amabile* Craib & W. W. Smith is used in Bhutanese traditional medicine (BTM) for treating dyspepsia, poisoning and fever, and is also often used as a substitute for musk, which is obtained from the Himalayan musk deer (endangered and prohibited) [8]. *P. amabile* 



**Figure 1**: Struct es of compounds (1-10) isolated from the Me extract of *P*. *amabile.* 

grows to a height of 15-50 cm, with a stout root and a solitary stem in open scrub, alpine turf and the semi-stable screes of the high altitude Himalayan mountains (3950 to 4700 masl) of Bhutan [7,8].

We have previously studied the essential oil component of this plant and found interesting bioactivity [1a]. This interesting preliminary finding encouraged us to further investigate the chemical components of the methanol extract of this plant for the first time. The MeOH extract was sequentially fractionated with *n*-hexane, petroleum spirit,  $CH_2Cl_2$ ,  $CHCl_3$ , EtOAc and *n*-butanol to obtain their respective fractionated extracts. These extracts were purified repeatedly using silica gel column chromatography and preparative thin layer chromatography (PTLC). Finally, four phenylpropanoids and six furanocoumarins were identified by comparing the LR-ESI-MS, LR-EI-MS, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of the isolated compounds with those of the relevant literature, as detailed in the experimental section. The compounds are (Figure 1): (*E*)-isoelemicin (1), (*E*)-isoepiol (2), methyl eugenol (3), (*E*)-isoelemicin (4), psoralen (5), bergapten (6) (major compound of a plant), isoimperatorin (7), isopimpinellin (8), oxypeucedanin hydrate (9) and oxypeucedanin methanolate (10).

We investigated these compounds (1-10) and various crude extracts for their antimicrobial, antimalarial and cytotoxicity activities. Some biological activities reported earlier by us [1a, 9] for this plant are reproduced in Table 1 for easy comparisons. The antimicrobial assay was performed using seven bacterial strains and one fungal strain in a modified Agar Well Diffusion method (AWD) and the broth microdilution methods in 96 well plates that determined the minimum inhibitory concentrations (MIC). Unlike the previously reported activity of crude extracts and compounds 1-2 [1a] (Table 1), the crude petroleum spirit, EtOAc and n-BuOH extracts and the pure compounds 3-8 studied here did not show any interesting antimicrobial activities. However, compound 10 showed weak activity against the Gram-positive bacterium, Bacillus subtilis with a minimum inhibition zone (MIZ) of 9 mm and a MIC value of 125  $\mu$ g/mL (Table 1). Compound 6 (bergapten), isolated from Heracleum maximum, was reported to be moderately active against Mycobacterium tuberculosis with a MIC value of 200 µg/mL and an IC\_{50} value of 27  $\mu g/mL$  [10].

Among the three crude extracts (petroleum spirit, EtOAc and *n*-BuOH extracts) and seven pure compounds (**3** and **5-10**) studied here, the petroleum spirit and EtOAc crude extracts, and compounds **7-10** exhibited moderate to significant antiplasmodial activity against a wild type chloroquine and antifolate sensitive strain, TM4/8.2 and the multidrug resistant strain, K1CB1 of *Plasmodium falciparum* (Table 1). The petroleum spirit and EtOAc crude extracts showed moderate activities with  $IC_{50}$  values in the range of 24.3-37.9 µg/mL. The varying activities of the crude extracts and the EO described in Table 1 (including previous data, but inactive fractions not shown) suggested that the more potent chemical components are present in the CH<sub>2</sub>Cl<sub>2</sub> and CHCl<sub>3</sub> extracts when compared with the other extracts.

Out of ten compounds (1-10) isolated from these active extracts, oxypeucedanin methanolate (10) isolated from the CHCl<sub>3</sub> extract exhibited the highest and the most significant antimalarial activity against both *P. falciparum* strains, TM4/8.2 and K1CB1 with IC<sub>50</sub> values of 7.4 µg/mL and 6.7 µg/mL, respectively. Such activities were similar to that of its parent crude CHCl<sub>3</sub> extracts, but threefold better than its closely related compound 9 (oxypeucedanin hydrate) (Table 1). Compounds 7-9 also showed weak to moderate antiplasmodial activity with IC<sub>50</sub> values in a similar range of 15.6-24.9 µg/mL. Compound 5 showed selectivity against the K1CB1 strain with an IC<sub>50</sub> value of 16.4 µg/mL, but it was not active against the TM4/8.2 strain. Interestingly, two major compounds of this plant, (*E*)-isomyristicin (1) and bergapten (6), did not exhibit any biological activity.

While none of the test samples that we studied showed any major cytotoxicity or anticancer activity against human oral carcinoma KB cells, compounds 7 and 9-10 that were isolated from different plant species have been reported to possess antiproliferative activity by inhibiting the growth of nude mouse-transplantable human gastric

 Table 1: Antibacterial, antimalarial and cytotoxicity activities of crude extracts and compounds 1-10 isolated from *P. amabile*.

	Antibacterial Antiplasmodial		Cytotoxicity		
Samples	(MIZ in mm)			<u>(IC<sub>50</sub> in µg/mL)</u>	
•	B. subtilis	TM4/8.2	K1CB1	Vero	KB
CH <sub>2</sub> Cl <sub>2</sub> extract <sup>a</sup>	14	$12.1\pm0.4$	$10.9 \pm 2.3$	>25	>25
CHCl3 extract <sup>a</sup>	6	$7.8 \pm 1.8$	$7.3 \pm 2.6$	>25	>25
Pet. spirit extract*	-	$27.6\pm4.3$	$37.9 \pm 14.1$	>50	>50
Ethyl acetate extract	-	$24.3\pm3.2$	$25.2\pm6.5$	>100	>100
n-Butanol extract	-	>100	>100	>100	>100
Essential oil (EO)b	5	79.0 ±4.6	$72.3 \pm 2.1$	>100	>100
1 <sup>b</sup>	5	>100	>100	>100	>100
2 <sup>b</sup>	6	$52.9\pm2.9$	$69.9\pm2.0$	>100	>100
3	-	>17.8	>17.8	>17.8	>17.8
4 <sup>b</sup>	-	>20	>20	>20	>20
5	-	>18.6	$16.4\pm0.3$	>18.6	>18.6
6	-	>10.8	>10.8	>10.8	>10.8
7	-	$19.1\pm0.9$	$18.5\pm0.7$	>27.0	>27.0
8	-	$19.5\pm2.3$	$15.6\pm2.5$	>24.6	>24.6
9	-	$24.9\pm2.0$	$18.7 \pm 2.3$	>30.4	>30.4
10	9	$7.4 \pm 1.3$	$6.7 \pm 1.1$	>31.8	>31.8
Amoxicillin <sup>c</sup>	8				
Chloroquine <sup>d</sup>		0.01	0.09		
Cycloguanil <sup>d</sup>		0.009	0.81		
Pyrimethamine <sup>d</sup>		0.02	7.70		
Ellipticine <sup>e</sup>				0.09	
Doxorubicine					0.56

<sup>a</sup> Original activity taken from [9]. <sup>b</sup> Original activity taken from [1a]. <sup>c</sup>Positive controls for antibacterial activity. <sup>d</sup> Positive controls for antiplasmodial activity. <sup>e</sup> Positive controls for cytotoxicity activity. \*Petroleum spirit extract. -: Not Active.

adenocarcinoma (MK-1), human uterus carcinoma (HeLa) and murine melanoma (B16F10) cells [11]. Compounds **5** (against HL-60 and K562) and **7** (against L1210, HL-60, K562, B16F10) were also reported to exhibit moderate cytotoxicity against various tumor cell lines [12].

In summary, the petroleum spirit and EtOAc crude extracts exhibited moderate antiplasmodial activity against a wild type chloroquine and antifolate sensitive strain, TM4/8.2 and the multidrug resistant strain, K1CB1 of *Plasmodium falciparum*. These activities were lower than the CHCl<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub> which we reported earlier. Focusing our isolation of phytochemicals on these active fractions (Table 1), ten compounds (1-10) were isolated with isomyristicin (1) and bergapten (6) as the major constituents. Among the ten compounds that were bio-assayed, compound 10 exhibited the highest activity against the same *P. falciparum* strains mentioned above. These biological activities were found to be in conformity with the traditional uses of *P. amabile* in BTM formulations and, therefore, provide scientific insight to this plant.

#### Experimental

**Plant materials:** The aerial components of *P. amabile* Craib & W. W. Smith (voucher number 29) were collected from Lingzhi (4200 m above sea level) in July 2009 and a herbarium specimen was deposited at Manjong Sorig Pharmaceuticals, Ministry of Health, Thimphu, Bhutan. The dried plant material (2 kg) was extracted with methanol (190 g of MeOH obtained).

Isolation and identification of compounds from the MeOH extract: The equipment used, and the isolation and characterization techniques carried out were as described by us previously [13a-b]. The known compounds were identified through MS library matching techniques (NIST and NISTREP mass spectra library) and then confirmed through comparison of their MS and NMR spectra (500 MHz, CDCl<sub>3</sub>) with those reported and discussed as follows. The MeOH extract was dissolved in MeOH/water (1:9) and fractionated first with *n*-hexane and petroleum spirit to obtain their respective extracts. The remaining aqueous portion was acidified with HCl (5%) and fractionated with CH<sub>2</sub>Cl<sub>2</sub> to obtain the acidic extract. The aqueous portion was again basified with NH4OH to pH 9-12 and then fractionated with CHCl<sub>3</sub> to obtain a basic CHCl<sub>3</sub> extract followed by EtOAc and n-butanol giving EtOAc and BuOH extracts, respectively. The remaining aqueous layer was dried to obtain the H<sub>2</sub>O extract. Each of these extracts (50-100 mg) was used for determining their biological activities. The initial purification was focused on the most active CHCl<sub>3</sub> extract (13.3 g) using silica gel CC and a gradient eluent solvent system of MeOH-CH<sub>2</sub>Cl<sub>2</sub> (500 mL, v/v ratios 0:100, 5:95, 10:90, 15:85, 50:50), which yielded 13 fractions, CHCl<sub>3</sub>-F1 to CHCl<sub>3</sub>-F13). Separation of fraction CHCl<sub>3</sub>-5 using PTLC plates with CH<sub>2</sub>Cl<sub>2</sub> (100%) as an isocratic solvent system, yielded compounds 1-4, which were identified as (E)-isomyristicin (1), (E)-isoapiol (2), methyl eugenol (3) and (E)-isoelemicin (4). Fraction CHCl<sub>3</sub>-F7 was separated using the same method and solvent system as above and furnished psoralen (5), bergapten (6) (as major compound) and isoimperatorin (7). Fraction CHCl<sub>3</sub>-F10, upon separation using preparative TLC plates with MeOH-CHCl<sub>3</sub> (1:19) as the solvent system, yielded isopimpinellin (8), oxypeucedanin hydrate (9) and oxypeucedanin methanolate (10). From the petroleum spirit, CH<sub>2</sub>Cl<sub>2</sub> and EtOAc extracts, the same compounds were obtained. The n-hexane, n-BuOH and H<sub>2</sub>O extracts were not studied here.

Identification of phenylpropanoids (1-4): (E)-isomyristicin (1) showed a  $[M+H^+]$  peak at m/z 193, and an ion fragmentation pattern that matched that of isomyristicin in the MS library system (NIST08.LIB, Entry # 15372, CAS: 607-91-0). The  $^{1}$ H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those reported for isomyristicin [14a-b]. (E)-isoapiol (2) was isolated as an oil, which showed a  $[M+H^+]$  peak at m/z 222 in the LR-ESI-MS. The LR-EI-MS exhibited fragmentation ions at m/z 222 (base peak), 207, 191, 177, 163, 149, 134, 121, 106, 91, 77, 65, 53, and 39. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of isoapiol reported in the literature [15]. Methyl eugenol (3) was isolated as colorless oil. Its LR-ESI-MS showed a  $[M+H^+]$  peak and fragmentation ions that matched those of eugenol methyl ether in the MS library system (NIST08s.LIB, Entry # 13416, CAS: 93-15-2). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of methyl eugenol reported in the literature [16]. (E)-isoelemicin (4) was isolated as colorless oil. Its LR-ESI-MS showed a  $[M+H^+]$  peak at m/z 209 and a  $[M^+]$  peak at m/z 208. The <sup>1</sup>H NMR spectroscopic data of this compound agreed with those of (E)-isoelemicin reported in the literature [14b].

*Identification of furanocoumarins* (5-10): Psoralen (5) was isolated and recrystallized as white needles from MeOH/CHCl<sub>3</sub>. Its LR-ESI-MS showed a  $[M+H^+]$  peak at m/z 187, and the LR-EI-MS exhibited an ion fragmentation pattern which matched that of psoralen in the MS library system (NIST08.LIB, Entry # 34463, CAS: 66-97-7). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those reported for psoralen [17]. Bergapten (6) was isolated and recrystallized as clear crystals from MeOH/CHCl<sub>3</sub>. Its LR-ESI-MS displayed a  $[M+H^+]$  peak at m/z 217, and its LR-EI-MS matched that of bergapten in the MS library system (NIST08.LIB, Entry # 53002, CAS: 484-20-8). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those reported for bergapten [10, 18]. Isoimperatorin (7) was isolated and recrystallized from MeOH/CHCl<sub>3</sub>. Its LR-ESI-MS

indicated a [M+H<sup>+</sup>] peak at m/z 271, and its LR-EI-MS fragmentation pattern matched that of isoimperatorin in the MS library system (NIST08.LIB, Entry # 90508, CAS: 482-45-1). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of isoimperatorin reported in the literature [19a-b]. Isopimpinellin (8) was isolated as white solid. The EI-MS, <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of isopimpinellin reported in the literature [20]. Oxypeucedanin hydrate (9) was isolated as light yellowish needles from MeOH/CHCl<sub>3</sub>. Its LR-ESI-MS showed a  $[M+H^+]$  peak at m/z305 and the LR-EI-MS demonstrated an ion fragmentation pattern which matched that of oxypeucedanin hydrate in the MS library system (NIST08.LIB, Entry # 114683, CAS: 24724-52-5). The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of isopimpinellin reported in the literature [11]. Oxypeucedanin methanolate (10) was isolated as a white solid. Its LR-ESI-MS indicated a  $[M+H^+]$  peak at m/z 319, and the LR-EI-MS established the ion fragmentation pattern as m/z 318 [M<sup>+</sup>], 286, 215, 202 (base peak), 185, 174, 157, 145, 118, 99, 85, 73, 59, and 51. The <sup>1</sup>H NMR and <sup>13</sup>C NMR spectroscopic data of this compound agreed well with those of oxypeucedanin methanolate reported in the literature [11].

Bioassay methods: The antimicrobial, antiplasmodial and cytotoxicity tests were carried out in the same ways as we have reported earlier [9, 13a-b]. For the antimicrobial agar well diffusion and broth microdilution bioassay, Escherichia coli (ATCC 25922), Bacillus subtilis (ATCC 6633), Staphylococcus aureus (ATCC 6538), methicillin resistant S. aureus (MRSA), (DMST 20651), S. epidermidis (ATCC 12228), Vibrio cholerae (DMST 2873) and Candida albicans (ATCC 10231) were used as test strains. Amphotericin B (Sigma-Aldrich, USA) was used as a positive control for antifungal testing against Candida albicans (not shown in Table 1 as the samples were found to be inactive). Vancomycin (Edicin, Slovenia) and amoxicillin (GPO, Thailand) were used as the positive controls for antibacterial assays. The K1CB1 (a multidrug resistant) and TM4/8.2 (a wild type chloroquine and antifolate sensitive) strains of Plasmodium falciparum were used for the modified microdilution radioisotope antiplasmodial bioassay. This assay used chloroquine (Sigma-Aldrich), cycloguanil (Sigma-Aldrich) and pyrimethamine (Sigma-Aldrich) as positive controls. For the sulforhodamine B (RSB) cytotoxicity assay, normal vero cells from kidney of African green monkey, Cecopithecus aethiops and the human oral carcinoma KB cells were used. This assay used ellipticine (Sigma-Aldrich, USA) and doxorubicin (Sigma-Aldrich, USA) as reference drugs. All the experiments were performed 3 times in duplicate (3x2) and DMSO (0.1%) and distilled water were used as controls to rule out the solvent effects on the bioassay results of the test samples.

Acknowledgments - We appreciate the support of: a) Ms Norbu Drolma, Ms. Tshring Zam, Mr Samten and Mr Ugyen Dhendup (Head) of Manjong Sorig Pharmaceuticals and Mr Dorji Wangchuk (Director) of the National Institute of Traditional Medicine for providing the materials and necessary administrative services; and b) Roonglawan Rattanajak of BIOTECH, Thailand for antiplasmodial assistance. Mr P. Wangchuk is an Endeavour Award Holder of Australia.

#### References

- (a) Wangchuk P, Keller PA, Pyne SG, Taweechotipatr M, Kamchonwongpaisan S. (2013) GC/GC-MS analysis, isolation and identification of [1] bioactive essential oil components from the Bhutanese medicinal plant, Pleurospermum amabile. Natural Product Communications, 8, 1305-1308; (b) Zehui P, Watson MF. (2005) Pleurospermum Hoffmann, Gen. Pl. Umbell. Viii.1814. In Flora of China, 14, 40-51.
- [2] (a) Lee IK, Choi SU, Lee KR. (2012) Triterpene saponins from Pleurospermum kamtschaticum and their biological activity. Chemical & Pharmaceutical Bulletin, 60, 1011-1018; (b) Luo YG, Li BG, Zhang GL. (2002) Four new glycosides from Pleurospermum franchetianum. Journal of Asian Natural Products Research, 4, 155-163.
- [3] (a) Ming Y. Zhou LY. Miao X. Hui ZX. (2012) Pharmacodynamic study on anti-inflammatory and analgesis effects of Tibetan medicine Pleurospermum hookeri var. thomsonii. Natural Product Research and Development, 24, 972-975; (b) Kim JE, Chung WY, Chun KS, Lee CK, Park HJ, Kim WB, Park KK. (2010). Pleurospermum kamtschaticum extract induces apoptosis via mitochondrial pathway and NAG-1 expression in colon cancer cells. Bioscience, Biotechnology, and Biochemistry, 74, 788-792.
- Jung HJ, Nam JH, Park HJ, Lee KT, Park KK, Kim WB, Choi J. (2007) The MeOH extract of Pleurospermum kamtschaticum and its active [4] component buddlejasaponin (IV) inhibits intrinsic and extrinsic hyperlipidemia and hypercholesterolemia in the rat. Journal of Ethnopharmacology, 112.255-261.
- Shibano M, Naito H, Taniguchi M, Wang NH, Baba K. (2006) Two isocoumarins from Pleurospermum angelicoides. Chemical & Pharmaceutical [5] Bulletin, 54, 717-718.
- Khetwal KS, Rijivi AA, Pande S. (1994) An ester and coumarins from the high altitude herb Pleurospermum densiflorum. Phytochemistry, 35, [6] 1033-1035
- Grierson AJC, Long DG. (1999) Pleurospermum Hoffmann. In Flora of Bhutan: Including a Record of Plants from Sikkim. Royal Botanic Garden, [7] Edinburgh, 2, pp.451-457.
- [8] Wangchuk P, Samten, Ugyen. (2009) High Altitude Medicinal Plants of Bhutan: An Illustrated Guide for Practical Use. Pharmaceutical and Research Unit, Ministry of Health, Thimphu, Bhutan.
- Wangchuk P, Keller PA, Pyne SG, Taweechotipatr M, Tonsomboon A, Rattanajak R, Kamchonwongpaisan S. (2011) Evaluation of an [9] ethnopharmacologically selected Bhutanese medicinal plants for their major classes of phytochemicals and biological activities. Journal of Ethnopharmacology, 137, 730-742.
- [10] O'Neill T, Johnson JA, Webster D, Gray CA. (2013). The Canadian medicinal plant Heracleum maximum contains antimycobacterial diynes and furanocoumarins. Journal of Ethnopharmacology, 147, 232-237.
- Fujioka T, Furumi K, Fujii H, Okabe H, Mihashi K, Nakano Y, Matsunaga H, Katano M, Mori M. (1999) Antiproliferative constituents from [11] Umbelliferae plants. V. A new furanocoumarin and falcarindiol furanocoumarin ethers from the root of Angelica japonica. Chemical & Pharmaceutical Bulletin, 47, 96-100.
- Thanh PN, Jin WY, Song GY, Bae KH, Kang SS. (2004) Cytotoxic coumarins from the root of Angelica dahurica. Archives of Pharmacal [12] Research, 27, 1211-1215.
- (a) Wangchuk P, Keller PA, Pyne SG, Korth J, Samten, Taweechotipatr M, Rattanajak R, Kamchonwongpaisan S. (2013) Antimicrobial, [13] antimalarial and cytotoxic activities of constituents of a Bhutanese variety of Ajania nubigena. Natural Product Communications, 8, 733-736; (b) Wangchuk P, Keller PA, Pyne SG, Sastraruji T, Taweechotipatr M, Rattanajak R, Tonsomboon A, Kamchonwongpaisan S. (2012) Phytochemical and biological activity studies of the Bhutanese medicinal plant *Corydalis crispa*. *Natural Product Communications*, **7**, 575-580. (a) Antunes AM, Palma FMSB, Santos SMBP. (**1995**) Isomyristicin: <sup>1</sup>H and <sup>13</sup>C-NMR assignments. *Fitoterapia*, **66**, 554; (b) Enqiquez RG, Chavez
- [14] MA, Jauregui F. (1980) Propenylbenzenes from Guatteria gaumeri. Phytochemistry, 19, 2024-2025.
- [15] Jakupovic J, Eid F. (1987) Benzoxanthenone derivatives from Polemannia montana. Phytochemistry, 26, 2427-2429.
- Miyazawa M, Kohno G. (2005) Suppression of chemical mutagen-induced SOS response by allylbenzene from Asiasarum heterotropoides in the [16] Salmonella typhimurium TA1535/PSK 1002 umu test. Natural Product Research, 19, 29-36.
- [17] Jiangning G, Xinchu W, Hou W, Qinghua L, Kaishun B. (2005) Antioxidants from a Chinese medicinal herb-Psoralea corylifolia L. Food Chemistry, 91, 287-292.
- [18] Kawaii S, Tomono Y, Katase E, Ogawa K, Yano M. (1999) Isolation of furocoumarins from bergamot fruits as HL-60 differentiation-inducing compounds. Journal of Agricultural and Food Chemistry, 47, 4073-4078.
- (a) Tian YQ, Zhang ZX, Xu HH. (2013) Laboratory and field evaluations on insecticidal activity of Cicuta virosa L. var. latisecta Celak. Industrial [19] Crops and Products 41, 90-93; (b) Jackson GE, Campbell WE, Davidowitz B. (1990) Two dimensional NMR study of some natural coumarins. Spectroscopy Letters: An International Journal for Rapid Communication, 23, 359-367.
- [20] Yoo SW, Kim JS, Kang SS, Son KH, Chang HW, Kim HP, Bae KH, Lee CO. (2002) Constituents of the fruits and leaves of Euodia daniellii. Archives of Pharmacal Research, 25, 824-830.