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PHOSPHORUS SPIRALLING IN A WOODLAND STREAM: SEASONAL VARIATIONS¹

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Abstract. Four radiotracer releases were performed over an annual period in 1981–1982 to determine seasonal variation in indices and pathways of phosphorus spiralling in Walker Branch, a small woodland stream in eastern Tennessee, USA. Each release consisted of an addition of ≈ 370 MBq each of carrier-free ${}^{32}\text{PO}_4$ and ${}^{3}\text{H}_2\text{O}$ over a 1-h period during baseflow. Concentrations of ${}^{32}\text{P}$ and ${}^{3}\text{H}$ dissolved in stream water were measured intensively at several stations downstream from the radio-tracer input during and immediately following each release. Activity of ${}^{32}\text{P}$ in coarse particulate organic matter (CPOM), fine particulate organic matter (FPOM), and aufwuchs was measured 2–3 h after each release and at various intervals for 7 wk. Total biomass of CPOM, FPOM, and aufwuchs at the time of each release was also measured.

Uptake of ³²PO₄ from the water was greatest in November and lowest in August. Uptake length (S_w) of phosphorus, defined as the average distance travelled by a PO₄ ion dissolved in water, varied from 22 m in November to 97 m in August. Uptake of ³²PO₄ by CPOM was generally greatest, with $\approx 50\%$ of total uptake, while that by aufwuchs was lowest, with < 15% of the total. CPOM abundance was the major determinant of whole-stream PO₄ uptake rate and S_w . Turnover length (S_p) of phosphorus, defined as the average distance traveled by an atom of P taken up by particulate material, was short compared to S_w , varying from 1 m in November to 3 m in January. Consequently total spiralling length (S) of P varied from 23 m in November, just after peak autumn leaf fall, to 99 m in August, and reflected primarily the travel of P in the dissolved form.

Our results indicate that the greatest increase in S_w (and consequently in S) in Walker Branch occurs in late autumn or winter after storms reduce the abundance of CPOM in the lower portions of the stream bed. Although we calculate that S_p may increase by one to two orders of magnitude for short periods during storms, the greatest effect of storms on P spiralling over the long term is their impact on the quality and quantity of CPOM and FPOM in the stream bed after the return to baseflow. For most of the year, detrital organic carbon probably influences phosphorus spiralling more than phosphorus spiralling influences the processing of organic carbon in Walker Branch. Only during the fall and early winter periods, when CPOM abundance is high and S_w is short, does phosphorus spiralling exert strong control over biotic processes downstream.

Key words: phosphorus; radioisotope; recycling; spiralling length; stream ecosystem; Tennessee; transport.

INTRODUCTION

Webster (1975) and Webster and Patten (1979) suggested that nutrient cycles in streams be considered as spirals because of continuous downstream displacement of nutrients as they cycle. To describe the simultaneous processes of nutrient cycling and downstream transport in streams, Newbold, Elwood, and their coworkers further developed and quantified the concept of spiralling (Newbold et al. 1981, 1982, 1983, Elwood et al. 1983). Newbold et al. (1981) presented an index of spiralling, and reported results from a field measurement of phosphorus spiralling in July 1978 using radiotracers (Newbold et al. 1983). Stream eco-

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² Present address: Stroud Water Research Center, Avondale, Pennsylvania 19311 USA. system models based on spiralling theory have also been developed (Newbold et al. 1982, 1983).

In this paper we present results on the seasonal variability of indices and pathways of phosphorus spiralling in Walker Branch, a small woodland stream in eastern Tennessee, USA. Changes in phosphorus spiralling in streams are expected as a result of seasonal variation in light, temperature, and organic matter storage. For streams with deciduous canopies, the organic matter budget is dominated by large inputs of leaves in autumn (e.g., Fisher and Likens 1973). Gregory (1978) has suggested that phosphorus uptake rates in streams should increase when standing crops of allochthonous organic matter are high. Large amounts of dissolved organic carbon (DOC) are leached and rapidly utilized by stream microbes during autumn (Lock and Hynes 1976, McDowell and Fisher 1976), perhaps increasing microbial phosphorus demand. AlJune 1985

ternatively, autumn leaf inputs contribute substantially to the phosphorus supply of streams (Cowen and Lee 1973, Meyer and Likens 1979) and thus may reduce demand for phosphorus in the water. The pattern of microbial conditioning of decomposing leaves has also been shown to have a large influence on the dynamics of phosphorus (Gregory 1978, Meyer 1980) and nitrogen (Triska and Buckley 1978) in woodland streams. Using data from two previous 32PO₄ releases to Walker Branch, Elwood et al. (1983) estimated that the uptake length of PO_4 (average distance traveled by a PO_4 ion in water before uptake by stream microbes) was only 6 m immediately after peak autumn leaf fall, compared to 165 m in July. In this report we present results of a systematic investigation of seasonal changes in the patterns and controlling mechanisms of phosphorus spiralling in Walker Branch during 1981-1982.

STUDY SITE

Our study was conducted in a 120-m reach of the West Fork of Walker Branch, a first-order woodland stream located in the Ridge and Valley province of eastern Tennessee. Walker Branch flows from a 38.4ha experimental watershed on the Oak Ridge National Environmental Research Park. A detailed description of the system was given by Curlin and Nelson (1968). The stream is fed by springs and seeps that arise in dolomitic limestone. The stream gradient averages 55.9 m/km. Streamflow in the study reach is continuous, with low summer baseflows of 2-4 L/s and occasional brief winter and spring stormflows exceeding 50 L/s. Flow through the reach is augmented 10-25% by springs and groundwater seeps. During baseflow periods the stream averages ≈ 0.05 m in depth and ≈ 2.0 m in width.

Average concentrations of phosphorus and nitrogen, as soluble reactive phosphorus (SRP), NO₃-N, and NH₄-N, were 2, 17, and 72 μ g/L, respectively (from weekly data during 1972–1973, P. J. Mulholland et al., *personal observation*). Alkalinity averages 2.2 mmol/L and pH \approx 7.7 (Elwood and Nelson 1972).

METHODS

We conducted a series of four tracer releases of ${}^{32}PO_4$ to the West Fork of Walker Branch over an annual period. Methods used in the study were similar to those used in a July 1978 experiment reported by Newbold et al. (1983).

About 1 wk prior to each tracer release, standing stocks of coarse particulate organic matter (CPOM, >1 mm) and fine particulate organic matter (FPOM, <1 mm) on the stream bottom were measured. We used stratified random sampling to locate 30 sample sites within relatively homogeneous habitats previously classified as gravel-riffle, bedrock-riffle, moss-riffle, or pool (see Newbold et al. 1983). Samples were taken by

placing a stainless steel cylinder (0.1 m² cross section) into (or on) the stream bottom as deeply as possible (generally 2-10 cm). Water flow into the sampler was suppressed by tightly surrounding the cylinder with cloth towels. Large detritus was removed by hand and transferred to a sample container. Then the bottom material was vigorously agitated with a hand trowel and pumped with a Guzzler hand pump through a 263- μ m mesh net and into a large plastic container, until a volume of 40 L had been transferred. The substrate was sampled to the maximum depth that could be excavated with the trowel, usually <10 cm. The contents of the net were added to the sample of large detritus, which was returned to the laboratory, refrigerated within a few hours, and wet-sieved within a few days into >1 mm (CPOM) and <1 mm (FPOM) fractions. The contents of the large plastic container (FPOM $<263 \ \mu m$) were thoroughly agitated and a 250-mL subsample was taken, returned to the laboratory, and refrigerated. Within a few days the subsamples were filtered on pre-ashed glass-fiber filters (Whatman GF/ C) and processed for ash-free dry mass (AFDM) determination.

A solution containing ≈ 370 MBq of ³²P as carrierfree H₂³²PO₄ and 370 MBq of tritium (³H) as tritiated water was released to Walker Branch over a 1-h period beginning between 1030 and 1200 on 18 November 1981 and on 18 January, 12 April, and 2 August 1982. Weather conditions during the releases ranged from overcast in January to partly cloudy on the other three dates. Tritium was added as a hydrologic tracer to correct for dilution and to determine the average water velocity. Water samples were collected from four or five stations along a 120-m reach at 1- or 2-min intervals during and immediately following the release. At each station, CPOM, benthic and suspended FPOM, and aufwuchs were sampled 5-6 h after the release began (2-4 h after passage of the release slug) and for seven subsequent weeks at intervals increasing from 1 d to 1 wk.

During the release, water samples were collected in polyethylene syringes and filtered (Whatman GF/C) immediately. Then 5 mL of filtrate was added to 10 mL of scintillation cocktail (Aquasol) and counted for ³²P and ³H on a Packard TRI-CARB 460 CD/liquid scintillation spectrometer. Approximately 45 water samples were collected at each station as the ³²P and ³H slug passed. CPOM samples consisted of leaves removed from the stream bottom and cut into small fragments. These were placed on tared planchets, dried at 80°C, ashed at 450° for determination of AFDM, and assayed for ³²P on a Harshaw TASC 12 planchet counter. The AFDM determination and ³²P assay were standard for all particulate samples. Benthic FPOM was sampled by aspirating sediments from depositional areas on the stream bottom. The sediments were collected on a glass-fiber filter (Whatman GF/C) and scraped from the filter onto tared planchets, and AFDM

	Date of release					
Parameter	18 Nov 1981	18 Jan 1982	12 Apr 1982	2 Aug 1982		
Streamflow at station 0 (L/s)*	2.6	3.1	7.5	3.6		
Water temperature at station 0 and 11 (°C)*	14, 16	10, 5	13, 15	15, 16		
Soluble reactive phosphorus (SRP) con- centration (µg/L)	5 (0.7, 5)	4 (0.9, 5)	4 (1.8, 5)	1 (0.7, 5)		
Seston concentration (AFDM, mg/L)	0.50 (0.03, 4)	0.64 (0.03, 4)	0.77 (0.05, 4)	0.69 (0.05, 4)		
Standing stocks (AFDM, g/m ²) Coarse particulate organic matter (CPOM) [†]	536 (340, 30)	102 (110, 30)	185 (240, 30)	75 (90, 30)		
Fine particulate organic matter (FPOM) [†]	340 (350, 30)	210 (200, 30)	400 (305, 30)	370 (380, 30)		
FPOM < 263 μ m (% of total)	82%	79%	73%	78%		
Aufwuchs‡	1.8 (0.6, 5)	0.6 (0.3, 10)	0.4 (0.2, 10)	0.6 (0.3, 10)		

TABLE 1. Selected physical, chemical, and biological measurements of Walker Branch, Tennessee, on each of the indicated ${}^{32}PO_4$ release dates. Standard deviation and number of replicates given in parentheses.

* Station 0 is the upstream and station 11 the downstream boundary of the study reach.

[†] Measured 1 wk prior to date of ${}^{32}PO_4$ release.

‡ January value was measured on 27 January 1982. April value was measured on 22 April 1982.

and ³²P activity were determined. Suspended FPOM (seston) was sampled by filtering 1 L of streamwater in the field through pre-ashed, tared glass-fiber filters (Whatman GF/C). Aufwuchs were scraped from rocks removed from the stream bed immediately after the release in November and January and washed onto tared planchets. The surface area from which the aufwuchs had been removed was traced on paper, cut, and computed by weighing the cuttings. During the April and August experiments, roughened Plexiglas slides (34 cm²) were placed at three stations 6-8 wk prior to the release for aufwuchs colonization. Aufwuchs were sampled by scraping material from the slides onto tared planchets. A study conducted during April indicated that aufwuchs biomass on slides averaged only $\approx 40\%$ of that scraped from a similar area of rock surface. Therefore, to compute biomass of aufwuchs during August, the data from slides was increased by a factor of 2.5.

All ³²P assay values for each tracer release were corrected for decay (to one common time for each release), background activity, and counting efficiency. Results for particulate samples were recorded as either becquerels per milligram AFDM or becquerels per square metre of stream bottom, based on the standing stocks of each component (CPOM, FPOM, aufwuchs) in each habitat type and on the area of each habitat type in each reach.

To compute rates of stable phosphorus uptake from the radiotracer data, specific activity of ${}^{32}PO_4$ in stream water during each release was determined. Total available phosphorus was estimated as soluble phosphorus reacting with molybdate. Samples of stream water for determination of soluble reactive phosphorus (SRP) were collected at each station and immediately filtered through washed glass-fiber filters (Whatman GF/C) into acid-washed 25-mL polyethylene bottles kept chilled, then frozen within 4 h of collection. SRP concentrations were determined using automated methods (Technicon 1974).

Studies of ³²PO₄ equilibration in streamwater between dissolved (passing through a Whatman GF/C filter) and particulate (seston) forms were performed in the laboratory in conjunction with each release. Water from Walker Branch was collected just upstream of ³²PO₄ input within a few days after the January, April, and August ³²PO₄ releases, on 16 December 1981 (4 wk after the November release), and again on 15 November 1982. From each sample, 1-1.5 L of stream water was incubated in continuously stirred, darkenedglass reactors in triplicate at temperatures within 5° of ambient stream water. Approximately 40 kBq of carrier-free ³²PO₄ was added to each reactor, and ³²P activity in dissolved and suspended particulate fractions was monitored for 2-5 d. Periodically during the incubation period, 20 mL of stream water were filtered (Whatman GF/C) and 5 mL of the filtrate was counted by liquid scintillation. Material retained on filters was rinsed with distilled water, dried, and ashed, and the ³²P was assayed on the planchet counter. After correcting for differences in counting efficiency, background, and decay for samples counted with different techniques, we computed the relative activity of ³²P associated with seston and dissolved in water over time in each reactor. Steady-state distribution of ³²P between water and seston was generally achieved within 2-4 h after ³²PO₄ addition. Although sorption of ³²PO₄ to walls of the glass reactors did occur, it did not effect our computation of ³²P distribution between water and seston, since these were measured directly.

Water samples for seston were collected in 250-mL plastic bottles at 10-min intervals during a number of

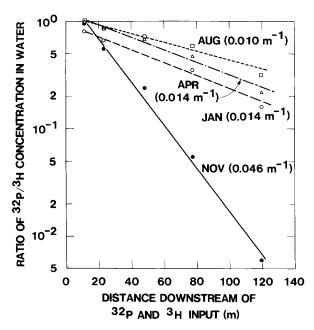


FIG. 1. Semi-logarithmic plot of means (n = 24-28) of ${}^{32}P/{}^{3}H$ concentration ratios during each release at various distances downstream from the input. (Standard errors are smaller than symbols [$\leq 3\%$ of mean for all points] and thus are not shown in figure.) The inverse of the slope of each line (given in parentheses) is the uptake length of PO₄ (S_w) during that release (Newbold et al. 1981).

storms by an automated fraction-collector activated when streamflow began to increase. The samples were retrieved within 2 d, filtered on ashed, tared glass-fiber filters (Whatman GF/C), and processed for AFDM and ³²P activity. Streamflow at a station ≈ 25 m downstream was continuously measured and recorded during each storm as well as during baseflow periods.

RESULTS

Environmental conditions

Physical, chemical, and biological factors varied seasonally in Walker Branch (Table 1). Although all releases were performed during baseflow periods, streamflow at the upstream reach boundary (station 0) during the releases ranged from 2.6 to 7.5 L/s. Streamflow increased by nearly 20% over the 120 m experimental reach on 18 November 1981 and 18 January 1982, by \approx 10% on 12 April 1982, and by 3% on 2 August 1982. Water temperature was lowest during the January release, decreasing from 10° at station 0 to 5° at the downstream boundary (station 11). During the other three releases, water temperature was higher and increased slightly from upstream to downstream. Water temperature at station 0 varied less than that at the other stations as a result of a large spring ≈ 20 m upstream that supplied about one-third of the total flow at a temperature ranging from 10° to 13° throughout the year.

Over the annual period beginning about the time of our first release in November, SRP concentration ranged from $<1 \ \mu g/L$ on a few occasions to 23 $\mu g/L$ on one date in April, and averaged 2.9 ± 2.0 (sD), 3.4 ± 3.1 , and $3.2 \pm 3.8 \ \mu g/L$ at the spring, station 0, and station 11, respectively (n = 36 samples for each station). There was no apparent seasonal trend, and the longitudinal variation in mean SRP concentration was not statistically significant (P > .05).

Standing stock of CPOM was greatest during the November release, immediately after peak autumn leaf fall. On 4 January 1982 a large storm produced a peak streamflow of 114 L/s (recurrence interval of 1-2 yr) and greatly reduced CPOM standing stock within the portion of the stream bed wetted during baseflow. Abundance of FPOM also declined from the November value. Minimum CPOM abundance occurred during August, while FPOM abundance increased from January to April and remained high in August. At all times, aufwuchs standing stock was low; however, it was about three times as great in November as in the other seasons. The relatively high November aufwuchs value may be the result of increased heterotrophic biomass supported by the large inputs of dissolved organic matter (DOM) leached from newly shed leaves. A number of researchers have shown that the autumn DOM influx to streams rapidly disappears from the water, via either uptake by heterotrophic microbes coating particulate surfaces (e.g., leaves, FPOM, clay, rocks) or physical adsorption to inorganic surfaces (Wetzel and Manny 1972, Lock and Hynes 1976, McDowell and Fisher 1976, Dahm 1981).

Phosphorus uptake

As the 1-h release slug passed downstream during each release, uptake of ${}^{32}PO_4$ conformed to an exponential decay model (Fig. 1); i.e., dissolved ${}^{32}PO_4$ concentration corrected for dilution decreased exponentially with distance downstream. The distance rate constants describing ${}^{32}PO_4$ uptake varied from 0.010 m⁻¹ in August to 0.046 m⁻¹ in November (Table 2). Time-specific and area-specific rate constants for total PO₄ uptake were computed using SRP data, average water velocities, and the distance rate constants for ${}^{32}PO_4$ uptake (Table 2).

Of the three principal biotic stream bed components that take up PO_4 directly from the water, aufwuchs had the highest mass-specific PO_4 uptake rates and FPOM the lowest, for all releases (Table 2). The mass-specific PO_4 uptake rate by aufwuchs was greatest during April, perhaps as a result of high light levels in early spring prior to development of the leaf canopy, while uptake rates by CPOM and FPOM were greatest in November, probably because of recent inputs of fresh leaves. The differences in uptake rates among aufwuchs, CPOM, and FPOM reflect the relative amount of active microbial biomass in each substrate. While much of the organic material scraped from rocks or glass slides may

TABLE 2.	Parameters describing	uptake of PO ₄ on 1	the basis of lo	ongitudinal dis	stance, time, ar	ea of stream b	ed, and mass o	эf
stream s	substrates. Standard dev	viation given in par	rentheses who	en it could be	calculated.			

	Date of release				
	18 Nov 1981	18 Jan 1982	12 Apr 1982	2 Aug 1982	
Average water velocity (cm/s)	2.3	3.0	5.5	4.0	
Distance rate constant for ${}^{32}PO_4$ up-take (m ⁻¹)	0.0464 (0.0021)	0.0143 (0.0011)	0.0137 (0.0012)	0.0103 (0.0010)	
Time-specific uptake rate of PO ₄ (min ⁻¹)*	0.062	0.027	0.046	0.026	
Area-specific P uptake rate (as PO ₄) (μ g m ⁻² min ⁻¹) [†]	15.5	5.4	11.1	1.3	
Mass-specific P uptake rates (ng mg ⁻¹ h	n-1)‡:				
CPOM FPOM Aufwuchs	1.7 (1.2) 1.2 (0.5) 5.2 (0.7)	0.9 (0.5) 0.3 (0.2) §	1.4 (0.5) 0.3 (0.1) 118 (11)	0.5 (0.2) 0.1 (0.02) 6.8 (3.0)	
Mass balance of ³² P uptake (% of total)) :				
CPOM FPOM Aufwuchs	89 48 5	26 23 §	44 23 8	48 57 9	
Rapid release to water¶	1	5	4	8	

* Fraction of available phosphorus pool in water taken up per unit time. Calculated by multiplying the distance rate constant of ${}^{32}PO_4$ uptake by the average water velocity.

† Calculated by multiplying the time-specific P uptake rate by the concentration of soluble reactive phosphorus [SRP] and the average water depth (0.05 m in November, January, and August, 0.06 m in April).

[‡] Computed by multiplying specific activity of ³²P in substrates (³²P/mg AFDM) sampled 4 h after beginning of release at each station by specific activity of ³²PO₄ in water (³²PO₄/SRP) during passage of the release slug. § Not measured.

Computed by multiplying mass-specific ${}^{32}P$ uptake rates by average standing stocks and presented as % of total uptake as measured by disappearance of ${}^{32}P$ from water.

 1^{32} P taken up by sediments but released back to water after passage of 32 PO₄ slug but before initial sampling of sediments. Computed by comparing ratios of 3 H and 32 P concentrations in each water sample after passage of the 32 PO₄ slug and 3 H and 32 P concentrations during passage of the 32 PO₄ slug.

be living, CPOM and FPOM are composed primarily of nonliving organic material; a surface coating of living microbes makes up only a small fraction of the total mass. Despite a greater surface area per unit mass of FPOM, the lower mass-specific uptake rate of FPOM as compared with CPOM may indicate that microbes on FPOM are less active metabolically. Perhaps FPOM is of poorer quality as an organic substrate for microbial growth. Alternatively, a portion of FPOM sampled may not have been on the sediment surface and thus was more isolated from ³²PO₄ in the overlying water than was CPOM.

A mass balance of ³²P taken up by stream substrates during each release was calculated (Table 2). Only for the April and August releases was the sum of ³²PO₄ uptake accounted for by aufwuchs, CPOM, and FPOM (calculated using mass-specific uptake rates and standing stocks) within 25% of the total uptake computed from disappearance of ³²PO₄ in the water over the experimental reach. In November, ³²PO₄ uptake accounted for by CPOM, FPOM, and aufwuchs was considerably greater (143%) than that computed as loss from the water. This may be the result of a sampling bias favoring surface CPOM and FPOM, which may have been more readily labeled than deeper substrates, particularly when standing stocks were large, as they were in November. The low fraction of total ³²PO₄ uptake accounted for by CPOM and FPOM in January (54% of total loss of ${}^{32}\text{PO}_4$ from water) is more puzzling. Certainly aufwuchs, which was not sampled after the release in January, was responsible for some of the uptake unaccounted for. Assuming a mass-specific ${}^{32}\text{PO}_4$ uptake rate by aufwuchs equal to that in April (when SRP concentrations were similar), aufwuchs would account for 12% of total ${}^{32}\text{PO}_4$ uptake in January. This leaves \approx 35% of the total ${}^{32}\text{PO}_4$ loss from water still unaccounted for. Perhaps standing stocks of CPOM and/or FPOM were underestimated in January.

Despite the limited success of the mass balance of ^{32}P , two observations can be made concerning partitioning of PO₄ uptake in Walker Branch. First, although aufwuchs had high mass-specific PO₄ uptake rates, the portion of whole-stream PO₄ uptake by aufwuchs was relatively low (probably <15%) owing to very low biomass. Second, during most releases, CPOM was the most important stream substrate for removal of PO₄ from water, although at times of relatively low CPOM abundance (e.g., January and August) PO₄ uptake by FPOM was similar to that by CPOM.

Phosphorus turnover

Activity of ³²P in CPOM, FPOM, and aufwuchs over time followed multi-first-order kinetics, which indicates the existence of at least two phosphorus pools

	Date of release			
	18 Nov 1981	18 Jan 1982	12 Apr 1982	2 Aug 1982
Temperature range (°C)	10-16	5-12	11–16	14-16
Turnover rates (d ⁻¹)*				
CPOM: rapid	0.19 (0.19, 2)	0.61 (0.10, 3)	0.57 (0.16, 3)	0.69 (0.05, 3)
slow	0.09 (0.03, 5)	0.08 (0.02, 5)	0.02 (0.01, 5)	0.08 (0.03, 4)
FPOM: rapid slow	0.17 (0.15, 2) 0.04 (0.01, 5)	0.26 (0.19, 2) 0.02 (0.01, 5)	0.11 (0.11, 2) 0.03 (0.01, 5)	0.13 (0.05, 2) 0.03 (0.01, 5)
Aufwuchs: rapid slow	···+ ···+	· · · † · · <i>·</i> †	0.32 (0.09, 3) 0.11 (0.02, 6)	0.22 (0.01, 4) 0.03 (0.01, 3)

TABLE 3. Range of water temperature during the 7 wk immediately following each ³²P release and turnover rates of ³²P associated with CPOM, FPOM, and aufwuchs (standard error and df in parentheses).

* Turnover rates were calculated by fitting a two-compartment first-order model to data on ^{32}P content in each substrate over time at stations near the upper end of the experimental reach.

† Insufficient data to compute turnover rate.

associated with each substrate. Turnover of ³²P from the rapidly cycling pools dominated ³²P kinetics from 4 h after the ³²P release to 4–7 d later, and release from the slow pool dominated ³²P kinetics thereafter (to 49 d). Turnover rates of ³²P in both pools associated with CPOM were generally greatest, and turnover rates of ³²P in the slow FPOM and aufwuchs pools were generally lowest (Table 3). The lower ³²P turnover rates in FPOM as compared to CPOM are consistent with the lower mass-specific uptake rates by FPOM (Table 2). However, the relatively low ³²P turnover rates in aufwuchs are in contrast to the relatively high massspecific ³²P uptake rates by aufwuchs (Table 2), perhaps indicating relatively high net storage of phosphorus by aufwuchs compared to microbes associated with CPOM and FPOM. Further, the relatively low ³²P turnover rate in CPOM in November compared to other seasons (Table 3) is in contrast to the relatively high massspecific ³²P uptake rate by CPOM in November (Table 2), perhaps again indicating greater net storage of phosphorus by CPOM at this time.

Indices of phosphorus spiralling

The uptake length of PO₄ in Walker Branch, designated S_{w} , was defined as the average distance travelled by a PO₄ ion in water (Newbold et al. 1981, 1983) and was computed as the inverse of the distance rate constant for ³²PO₄ uptake (Table 2). This distance was shortest in November (22 m) and longest in August (97 m) (Table 4). In earlier studies S_w values of 6 m in November and 165 m in July were measured (Elwood et al. 1983). The present results concur with the pattern of a short S_w in autumn after leaf fall and a long S_w in summer; however, the range is not as great.

The importance of CPOM in PO_4 uptake in Walker Branch is illustrated by the inverse relationship between S_w and CPOM abundance (Fig. 2). When CPOM was abundant, as in autumn after leaf fall, uptake length of PO_4 was shortest, which indicates that PO_4 supplies were utilized most efficiently.

Phosphorus turnover length (S_p) , defined as the av-

erage distance traveled by an atom of P taken up by particulate material, including microbes and algae (Newbold et al. 1981, 1983), can be computed from the total downstream flux of P in dissolved and particulate phases. Newbold et al. (1981) showed that, for a stream at or near steady state, S_p is related to the ratio of the particulate (seston) P flux (F_p) and dissolved P flux (F_w) by the following equation: $S_p = (F_p/F_w)S_w$. Although calculation of F_w is straightforward, calculation of F_p is complicated by the fact that not all of the seston P is actively cycling and thus potentially exchangeable with the dissolved P pool.

The ratio of exchangeable-P flux in secton (F_v) to that dissolved in water (F_w) , was computed using data on the equilibrium distribution of ³²P between water and seston for stream water collected a few days after each release. The equilibration experiment following the 18 November 1981 release was not performed until 16 December 1981. An additional equilibration experiment was performed on 15 November 1982 to approximate conditions during the previous year more closely. Although steady-state distribution of ³²P between water and seston was usually achieved within the first 2-4 h and was usually maintained for 1-2 d, during the August 1982 experiment an initial steady state was achieved after only a few minutes but was maintained for only 2-4 h. Other investigators (Lean 1973, Berman and Skyring 1979, Tarapchak et al. 1981) have reported rapid (30 min to a few hours) biotic equilibration of 32P between bacteria or algae and lake water. In our study, stream-water incubations carried out for >24-48 h resulted in increases in particle concentration and a second phase of net ³²P uptake by particles in all experiments, probably the result of artifacts of incubating samples in containers (e.g., wall growth and subsequent sloughing). Therefore, the ³²P distribution during the steady state achieved in the first 2-4 h was used to compute F_p/F_w for all experiments.

Steady-state distributions of ${}^{32}P$ on seston were $\leq 5\%$ of the total ${}^{32}P$ dissolved in water at all times (Table 5). These distribution values are much lower than val-

ues reported for lake seston (Lean 1973, Tarapchak et al. 1981), probably reflecting lower seston concentrations and/or lower microbial activity associated with stream seston compared with lake seston. Values of S_p , computed from the steady-state ³²P distribution between seston and water, were ≤ 3 m for all releases, much lower than the respective values of S_w (Table 4). Thus, total spiralling length of phosphorus (*S*, equal to $S_w + S_p$) was dominated by S_w and its seasonal variation is due to variation in S_w (Table 4).

Storm effects

The large storm of 4 January 1982 (peak flow of 114 L/s) greatly reduced CPOM abundance. Average standing stock of CPOM decreased by 434 g/m² over a period of 63 d: from 536 g/m² measured on 9 and 10 November 1981 to 102 g/m² measured on 12 January 1982, a decline of 81% (Table 1). Decomposition processes alone were calculated to account for only $\approx 60\%$ of the decline in CPOM standing stock (≈ 270 g/m²) during this period (Elwood et al. 1981*a*). We attribute the additional CPOM reduction of 164 g/m² ($\approx 60\%$ of the CPOM available just prior to the 4 January storm) to the scouring effects of the storm.

To determine the fate of the CPOM removed from the stream bed during this storm, on 12 January 1982 we attempted to compute a mass balance of ³²P-labeled CPOM in portions of the stream channel and bank flooded during large storms. After correcting for radioisotope decay and ³²P release from CPOM since the November release, we could account for only $\approx 36\%$ of the expected CPOM ³²P activity on the stream bed, and another 18% on stream banks. This leaves $\approx 46\%$ of the expected CPOM ³²P activity unaccounted for; it was washed downstream. Although the movement of CPOM out of the reach by way of fluvial processes could have occurred any time after the November 1981 release of ³²PO₄, the storm of 4 January 1982 was the first sizeable storm since the release. We observed, visually, little decline in CPOM standing stock in the study reach prior to that event. Thus, we estimate that the January storm was responsible for removing $\approx 64\%$ of the CPOM standing stock from the stream bed of the experimental reach. However, $\approx 30\%$ of the CPOM removed (18% of the CPOM standing stock) was not transported any great distance downstream, but was

TABLE 4. Uptake length (S_w) , turnover length (S_p) , and total spiralling length (S) of phosphorus in the experimental reach of Walker Branch.

Date of release	<i>S</i> _w (m)	$S_p(\mathbf{m})$	<i>S</i> (m)
18 Nov 1981	22	1	23
18 Jan 1982	70	3	73
12 Apr 1982	73	2	75
2 Aug 1982	97	2	99

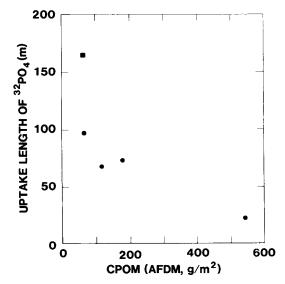


FIG. 2. Plot of uptake length of PO₄ vs. coarse particulate organic matter (CPOM) standing stock on dates when both were measured. Data points presented are for the four ${}^{32}PO_{4}$ releases in 1981–1982 (•) and for the ${}^{32}PO_{4}$ release in July 1978 (•).

deposited in unwetted regions of the stream channel (islands and debris accumulations above-water during baseflow) or on stream banks within the reach.

We also intensively sampled seston during a number of storms following ³²PO₄ releases. Seston mass concentration increased rapidly at the onset of rainfall and peaked prior to the peak in streamflow, as has been observed in other studies (e.g., Bilby and Likens 1979, Webster 1983). Concentrations of ³²P in the seston (measured as becquerels per milligram AFDM) during storms were substantially lower than ³²P concentrations found in seston just prior to storms. Results from the largest storm sampled (peak streamflow of 82 L/s), which began on 20 January 1982 \approx 40 h after release of ³²PO₄, illustrate the general trends in mass and ³²P concentration for seston found during storms (Fig. 3). During the 2-h period of peak seston mass concentration (580-700 min; Fig. 3), mean seston ³²P concentration was 1.4 Bq/mg AFDM, one-sixth of the seston ³²P concentration prior to the storm, indicating dilution of ³²P-labeled seston with unlabeled seston. However, this decline in seston ³²P content was not commensurate with the even larger increase in seston mass concentration, which increased by a factor of 24, from a prestorm value of 0.6 mg/L to a mean of 14.3 mg/L during this 2-h period. Thus, $\approx 75\%$ of the increase in seston concentration during the rising limb of the storm hydrograph was generated from within the ³²P-labeled study reach, i.e., within the 120 m of stream wetted during baseflow immediately upstream from our sampling location. This implies that during storms in Walker Branch, average travel distance of seston particles is

TABLE 5. Results of studies of ${}^{32}P$ equilibration between seston and water in Walker Branch conducted in conjunction with ${}^{32}PO_4$ releases. Standard deviation and number of replicates in parentheses.

Date	Seston concen- tration (AFDM, mg/L)	Tempera- ture† (°C)	D^* (F_p/F_w)
12 Dec 1981	0.30	9	0.05 (0.01, 3)
20 Jan 1982	0.42	7	0.04 (0.01, 3)
19 Apr 1982	0.70	17	0.02 (0.004, 3)
3 Aug 1982	0.73	15	0.02(0.002, 3)
15 Nov 1982§	0.37	15	0.03 (0.02, 3)

* Steady-state ratio of ³²P in secton (F_p) and ³²P in water (F_w) is presented as the ³²P fraction associated with secton $(D = F_p/F_w)$; see Results: Indices of Phosphorus Spiralling).

† Incubation temperatures were chosen to approximate field conditions.

[‡] Seston ³²P distribution did not reach steady state during first 48 h (see Results: Indices of Phosphorus Spiralling).

Additional study conducted because 16 December 1981 study was 4 wk after ³²PO₄ release (18 November 1981).

probably < 120 m and perhaps not greatly longer than at baseflow.

For the storm that began on 20 January 1982, seston ³²P flux at station 11 during the 24-h period of greatest transport was 13 times as great as seston ³²P flux during a 24-h baseflow period 1 d earlier. If, as assumed, the rate of P remineralization from seston did not increase during the storm, then the average S_p increased by more than an order of magnitude over that at baseflow during this period (Table 4).

DISCUSSION

Spiralling theory assumes a strong biological control of nutrient uptake and release in streams (Newbold et al. 1982, Elwood et al. 1983). Although Meyer (1979) reported that abiotic sorption by stream bed sediments controlled phosphorus concentrations in Bear Brook, New Hampshire, studies of other streams in Oregon (Gregory 1978) and Michigan (Ball and Hooper 1961), USA, and in New Zealand (Vincent and Downes 1980) indicated strong biotic control of phosphorus uptake. In hard-water streams such as Walker Branch, precipitation of calcium carbonate could remove PO₄ from solution. Total alkalinity and dissolved Ca concentrations in Walker Branch exhibit a seasonal pattern, with summer and early autumn maxima of 2.8 mmol/L and 30 mg/L, respectively, at pH of 8.1 (J. W. Elwood, personal observation). However, data on ³²P content of seston during our experimental releases indicated that uptake onto suspended particles was very low compared to total loss from the water, even during the summer and autumn studies. Total amount of ³²P retained on filters during passage of the ³²PO₄ slug at each stream station during each release was <1% of the ³²P loss from water between each set of stations, distances ranging from 10 to 43 m. Even if the mean travel distance of precipitated CaCO₃ particles was only a few metres prior to deposition on the stream bottom, co-precipitation of ${}^{32}PO_4$ would still be <5% of the measured ${}^{32}P$ loss from water during each release. Past studies of phosphorus spiralling in Walker Branch have also indicated that >90% of phosphorus uptake by sediments on the stream bottom is due to biotic processes (Elwood et al. 1981*a*).

It is also possible that we overestimated dissolved ³²PO₄ concentrations at downstream stations in Walker Branch during some releases if ³²PO₄ adsorbed to very small ($< 1.2 \mu m$) CaCO₃ particles that passed through the GF/C filters used to separate dissolved from particulate fractions. This potential overestimation of biologically available ³²PO₄ dissolved in water would result in similar overestimation of the biologically mediated PO_4 uptake length (S_w). To determine whether ³²PO₄ adsorption to small particles occurs, water samples (1 L) were collected at two stations in Walker Branch on 15 May 1984, spiked with ³²PO₄, and incubated for 1 h, and each of four replicate subsamples was filtered through GF/C, 0.4 µm Nuclepore, and 0.2 μ m Nuclepore filters. Total alkalinity was 1.8 mmol/L and pH was 7.9 at this time. No differences between filter types in the concentrations of ³²P in the filtrate were observed which indicates that no measurable ³²PO₄ adsorption to particles in the size range 0.2–1.2 μ m occurred. Although it is still possible that some adsorption may occur in summer and autumn when Ca concentrations and pH are at maximum values, the large decrease in S_w between these two seasons was

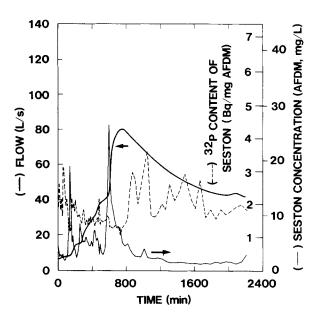


FIG. 3. Seston concentrations and ³²P concentrations in seston collected during a storm in Walker Branch beginning 20 January 1982, ≈ 40 h after release of ³²PO₄. Arrows on curves point to the associated ordinate scale.

probably not the result of differences in ${}^{32}PO_4$ adsorption to fine CaCO₃ particles.

Our data indicate an inverse relationship between PO₄ uptake rate and standing stock of CPOM in Walker Branch (Fig. 2). Our data also indicate that water velocity may positively influence PO4 uptake rate. In April, when water velocity was greatest, time- and area-specific uptake rates were somewhat higher (Table 2) than might have been expected from CPOM abundance data alone (Table 1). Various researchers (Lock and John 1979, Horner and Welch 1981) have reported a stimulation of phosphorus uptake by periphyton with increasing water velocities. Although increases in water velocity may stimulate PO4 uptake rates by stream bed substrates, increases in water velocity also increase the rate of downstream transport of dissolved P. Our results from April indicate that the net result of these counteracting effects was to reduce the stream distance rate constant for PO4 uptake, consequently increasing S_w . We would have expected S_w to be considerably shorter in April than in January because of higher CPOM abundance and greater area-specific PO₄ uptake rates in April (Tables 1 and 2). However, S_w was slightly longer in April than in January (Table 4) because increases in transport velocity compensated for increases in uptake of PO₄. Our results emphasize the dependence of S_w on factors controlling both uptake rates and transport rates of PO4, and indicate its usefulness as an index describing the simultaneous processes of PO₄ utilization and downstream transport, which define stream ecosystem efficiency.

Our calculations of area-specific and mass-specific PO₄ uptake rates were dependent upon the assumption that SRP represents total available phosphorus (i.e., that all forms are taken up as readily as PO₄). Although there is considerable evidence indicating that the SRP measurement overestimates orthophosphate (Rigler 1968, Stainton 1980, Tarapchak et al. 1982), a recent study indicates that SRP is a reasonable estimate of biologically available phosphorus in most natural waters (Nurnberg and Peters 1984). If SRP measurements in Walker Branch overestimated total available phosphorus, our values for area-specific and mass-specific PO4 uptake were correspondingly overestimated (Table 2). However, our calculations of the distance rate constants for PO_4 uptake and spiralling indices (S_w and S_{ν}) were dependent only on ³²P dynamics during our tracer additions in the field and in the laboratory equilibration studies, and are not affected by uncertainties about the SRP measurement.

We should also note that our computations of PO_4 uptake using tracer additions of ${}^{32}PO_4$ over a 1-h period represent gross uptake rates. We presume that release of ${}^{32}PO_4$ taken up by stream substrates was minimal during the 1-h releases and that the depletion of ${}^{32}P$ dissolved in stream water as the slug passed stations at various distances downstream from the input represents gross ${}^{32}PO_4$ uptake. Actually, we observed slight

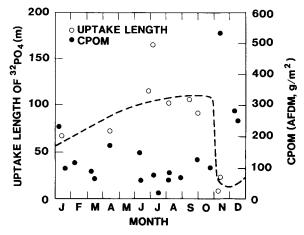


FIG. 4. Seasonal patterns of uptake length of PO_4 and coarse particulate organic matter (CPOM) abundance in Walker Branch (data from 1968 to present).

increases in ³²P (relative to ³H) during the latter portion of the slug passage at the most downstream station, and these data were deleted when computing average ³²P concentrations at that station. Thus, S_{ψ} calculated from the tracer data reflects the dynamic behavior of PO₄ despite the apparent steady-state concentrations (SRP concentrations) with distance and time.

From data of past releases of ³²PO₄ to Walker Branch as well as those of this study, a seasonal pattern in uptake length of PO₄ emerges (Fig. 4). After autumn leaf fall, when CPOM standing stock in Walker Branch is at its maximum, S_w is at its minimum. Usually a storm occurs in late fall or early winter that is large enough to reduce the standing stock of CPOM within portions of the stream bed inundated during baseflow. This reduction in CPOM results in a similar reduction in PO₄ uptake from stream waters, and consequently an increase in S_w . During spring and summer, S_w increases further in response to a gradual decline in CPOM standing stock. However, occasional new inputs of CPOM or reintroduction of stream-bank CPOM probably results in small decreases in S_w particularly during spring. The S_w measured during July 1978 (165 m) and reported in a previous paper (Newbold et al. 1983), is considerably longer than the other summer and early autumn values of S_w (Fig. 4), and this may indicate that S_w can exhibit considerable variability at this time of year. Although CPOM abundance (85 g/m²), water velocity (3.7 cm/s) and water temperature (15°-16°) in July 1978 were similar to values measured in August 1982 when S_w was only 97 m (Tables 1 and 4), FPOM abundance was considerably lower (150 g/m²) in July 1978 (Newbold et al. 1983). Relatively low uptake of PO₄ by FPOM in July 1978 may at least partially account for the longer S_w at that time compared to August 1982.

While our data implicate the quantity of CPOM as

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a major determinant of the efficiency of PO₄ utilization in Walker Branch, they do not reveal the importance of CPOM quality. In a separate study examining the role of microbial conditioning of CPOM on PO₄ uptake in laboratory streams, we found the PO₄ uptake by CPOM increased sharply, then declined during the first few weeks after leaves were added to the stream (Mulholland et al. 1984). However, the rate of PO₄ uptake by CPOM and S_w appeared to stabilize after CPOM had been in the streams $\approx 4-5$ wk. Therefore, during the first month after autumn leaf fall, S_w may undergo an additional temporal pattern of shortening and lengthening due to changes in CPOM quality alone.

Although the spiralling of phosphorus is clearly essential to the function of the stream ecosystem, its immediate influence on other aspects of ecosystem dynamics depends on (1) the degree to which concentrations of available phosphorus are controlled within the stream ecosystem, and (2) the level of response of other ecosystem processes to variations in available phosphorus concentrations. In a theoretical analysis of the spiralling concept, Newbold et al. (1982) argued that these two conditions should vary together, i.e., that the ecosystem's influence on concentrations of dissolved, biologically available phosphorus increases as the degree of nutrient limitation increases. When both are high, spiralling mechanisms such as uptake, regeneration, and downstream transport of nutrients should strongly influence other ecosystem dynamics (such as primary production, microbial decomposition, secondary production), and such influences can be transmitted downstream. Newbold et al. (1982) predicted that strong nutrient limitation in Walker Branch should be reflected in relatively low S_{ν}/S_{ρ} ratios (values approaching unity).

The lack of significant differences between SRP concentrations in spring water and in stream water downstream throughout the year in Walker Branch would appear to indicate a lack of ecosystem control of available phosphorus concentrations and hence a lack of strong phosphorus limitation. However, depletion of up to $1-2 \mu g/L$ in SRP concentration might not have been evident from our data due to a lack of sensitivity of our method of measurement. Measurements of SRP repeated on the same samples using our automated technique (Technicon 1974) indicated an average deviation of 2 μ g/L from the initial measurement. More recent measurements of SRP in Walker Branch using a more sensitive and precise nonautomated technique (Murphy and Riley 1962) indicated a consistent depletion of SRP of 1–2 μ g/L between spring water and stream water. The relatively large S_{u}/S_{n} ratios measured in Walker Branch (22-48; Table 4) also appear to indicate a lack of strong phosphorus limitation. Nonetheless, Elwood et al. (1981b) reported a phosphorus limitation of leaf decomposition during winter and spring in Walker Branch, although conditions of their study may have more nearly represented those of

autumn, because leaf packs of autumn-shed leaves were placed in the stream at the beginning of the study. Results of our present study indicating relatively low ³²P turnover rates in CPOM in November (Table 3) despite relatively high mass-specific ³²P uptake rates in November (Table 2) may also indicate relatively greater net phosphorus uptake by CPOM at that time. Therefore, our results in total indicate that phosphorus is most likely to be limiting in Walker Branch in late autumn and winter, when CPOM is maximum and S_{w} is minimum. At this time of year, biota in downstream reaches of Walker Branch are most strongly influenced by phosphorus spiralling mechanisms in upstream reaches. By contrast, in summer, when CPOM is minimum and consequently phosphorus demand is lowest, S_w is maximum and phosphorus spiralling has its least impact on ecosystem dynamics.

During storms, the flux of particulate phosphorus and S_p in Walker Branch increases sharply, potentially removing phosphorus accumulated on the stream bed during interstorm periods. Malfunctions in our automated sampler prevented the determination of a complete budget on particulate transport during storms. However, it is possible to obtain an estimate for average annual storm transport. Huff et al. (1977) found that storms accounted for 17% of the flow in Walker Branch between 1971 and 1975. The flow-weighted annual average concentration of transported FPOM during all storms in 1974–1975 was 8 mg/L, or ≈ 15 times the baseflow concentration (Comiskey 1978). Although we calculate that average CPOM concentration could be on the order of 10-20 mg/L during the first large storm following autumn leaf fall, over the entire year CPOM transport is small compared to FPOM transport (Sedell et al. 1978, Gurtz et al. 1980, Webster 1984). Thus, in an average year in which 17% of the flow is due to storms, $\approx 75\%$ of the particulate organic matter transport may occur during storms. As we have shown in this study, most but not all of the extra particulate transport is derived from the stream bottom. Therefore, ignoring the extra dissolved transport that occurs during storms, the average annual proportion of the downstream flux of phosphorus that is in exchangeable particulate form is probably no more than 8-20%, or four times that measured in baseflow.

In conclusion, this study further demonstrates the usefulness of the spiralling concept in analyzing nutrient dynamics in stream ecosystems. Seasonal variation in phosphorus spiralling length in Walker Branch is substantial and indicates a seasonal pattern in the efficiency with which supplies of PO_4 are utilized, despite lack of temporal and longitudinal variation in such measures of phosphorus concentration as SRP. Additional studies are needed to determine if the observed annual patterns in phosphorus spiralling indices continue in Walker Branch and whether they exist in other streams with different physical and biological characteristics. Additional field measurements are also

needed to determine how phosphorus spiralling indices vary between streams of similar size and longitudinally within stream systems.

Our study of annual variation in phosphorus spiralling has also shown that for most of the year, detrital organic matter influences phosphorus spiralling in Walker Branch probably more than phosphorus spiralling influences the processing of organic matter. Uptake of phosphorus and, by inference, ecosystem microbial production during much of the year appear to be limited by the abundance of CPOM and perhaps also by its quality (i.e., by availability of the biologically more labile carbon substances). Phosphorus spiralling is most likely to be important to metabolic processes in Walker Branch during late autumn and early winter, when CPOM abundance is large. Because of the relative unimportance of particulate transport in Walker Branch at baseflow, downstream influences of phosphorus spiralling, if they exist, are probably mediated temporally through storage and release (buffering), or possibly through downstream transport of dissolved organic phosphorus.

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