Photodegradation of FeDTPA in Nutrient Solutions. I. Effects of Irradiance, Wavelength, and Temperature

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Additional index words. plant nutrition, FeDTPA, FeEDTA, Fe-chelate, photochemistry

Abstract. Irradiation of FeDTPA-containing nutrient solutions by a fluorescent plus incandescent light source resulted in the loss of both Fe-chelate and soluble Fe, the formation of a precipitate that was composed mostly of Fe, and a rise in pH. The rate of Fe-chelate photodegradation in solution increased with irradiance intensity and with solution temperature under irradiation, but irradiance had the greater effect. Fe-chelates absorb in the blue and UV regions of the spectrum. Removal of these wavelengths with a spectral filter eliminated photodegradation. Chemical name used: ferric diethylenetriaminepentaacetic acid (FeDTPA).

Maintaining a sufficient level of micronutrients, particularly Fe, in a soluble, readily available form within a pH range suitable for the production of most plants is difficult without the use of chelates. A metal-chelate complex results when the chelating agent forms multiple bonds with the metal ion. These bonds occur in a ring structure around the metal ion, yielding a configuration in which the ion is nearly surrounded by the chelating agent, maintaining the metal in a soluble form in chemical environments where it would otherwise precipitate (Kolthoff et al., 1969). Chelating agents like diethylenetriaminepentaacetic acid (DTPA), ethylenediaminetetraacetic acid (EDTA), and ethylenediaminedi-o-hydroxy-

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²Professor. Current address: Dept. of Floriculture and Ornamental Horticulture, Cornell Univ., Ithaca, NY 14853. phenylacetic acid (EDDHA) have a high affinity for Fe and will form a stable complex with the metal across a pH range from 4.0 to 7.0, 4.0 to 6.3, and 4.0 to 9.0, respectively (Norvell, 1971).

The vulnerability of FeEDTA in solution to photodegradation was first reported in the mid-1950s when it was found that ultraviolet (UV) and blue radiation could destroy the chelate complex, yielding ferrous Fe (Fe²⁺), glyoxylic acid, formaldehyde, CO₂, and an amine residue (Frisell et al., 1959; Hamaker, 1956). In the early 1990s, Hangarter and Stasinopoulos (1991) showed that FeEDTA incorporated into tissueculture agar was vulnerable to photodegradation. The Fe-chelate FeEDDHA is also vulnerable to photodegradation (Wallace et al., 1967). To our knowledge, there have been no studies on the vulnerability of FeDTPA to photodegradation or how this photochemical event would alter a nutrient solution used in a hydroponic system. Therefore, the objectives of this study were to determine: 1) if FeDTPA photodegrades in nutrient solutions prepared in the laboratory; 2) the effects of light quality, irradiance, and temperature on this photochemical event; and 3) how FeDTPA photodegradation alters the nutrient solution. For comparison, FeEDTA, a Fe-chelate known to be vulnerable to photodegradation, was also included in these studies.

Materials and Methods

Photodegradation of FeDTPA in solution. Solutions of FeDTPA or FeEDTA (89.5 μ mol·L⁻¹ Fe) were contained (400 mL) in translucent 500-mL low-density polyethylene (LDPE) bottles (Nalgene Co., N.Y.). Iron chelate solutions were irradiated for 4 d with 500 μ mol·m⁻²·s⁻¹ of light (330–800 nm), measured at the external container surface. Containers were placed on their sides for irradiation and control containers were covered with aluminum foil (nonirradiated solutions). Irradiance was varied by adjusting lamp-bank distance from the containers. The radiation source was fluorescent plus incandescent lamps. The study was conducted in a controlled-environment growth chamber and solution temperature was maintained at 20 °C by adjusting air temperature. At the end of the study, solutions were centrifuged at 6000 g_n for 1 h in a Sorval SA-600 fixed-angle rotor (DuPont Instruments, Wilmington, Del.) at 22 °C. The supernatant (50 mL) was placed in a 100×15-mmdiameter polystyrene petri dish (Becton Dickinson and Co., Lincoln Park, N.J.) in a controlled-environment growth chamber providing 500 μ mol·m⁻²·s⁻¹ of light (330–800 nm) measured at the surface of the dish from a fluorescent plus incandescent light source. The petri dish was placed on a quantum sensor and the spectral characteristics of the supernatant were determined by spectroradiometric analysis of the radiant flux through the solution (light path = 1 cm) with a spectroradiometer (model LI-1800; LI-COR, Lincoln, Nebr.).

Photodegradation of FeDTPA in nutrient solution. A base nutrient solution, previously described (Albano and Miller, 1996), containing FeDTPA or FeEDTA was prepared as a 5× concentrate stock based on a 14.28 mmol·L⁻¹ N (17.9 µmol·L⁻¹ Fe) 1× concentration. Nutrient solutions (400 mL) were contained in translucent 500 mL LDPE bottles and irradiated for 48 h with 500 µmol·m⁻²·s⁻¹ of light (330-800 nm) as described above except that the radiation source was high intensity discharge (HID), metal halide lamps. Irradiance was varied by adjusting the distance from the lamp to the container surface. Four replications of each treatment were made in a completely randomized design. The pH of the nutrient solution was between 4.8-4.9 and was not modified. At 6-h intervals, a 20-mL aliquot was drawn from each container after agitation and pH was determined. Samples were then centrifuged at $6000 g_n$ for 20 min in a Sorval SA-600 angle-head rotor at 22 °C. The supernatant was analyzed for Fe, Mn, Zn, Cu, K, Ca, and Mg by atomic absorption spectrophotometry (AA), and colorimetrically for P according to Jackson (1958). FeEDTA and FeDTPA were measured spectrophotometrically (model DU-64; Beckman Instruments, Fullerton, Calif.) at 258 nm and 260 nm, respectively, as described by Hill-Cottingham (1957). Absorbance was linear from 1.79 to 89.5 µmol·L⁻¹ for both FeEDTA and FeDTPA standards (data not shown). Iron determined by AA includes all forms of soluble Fe (i.e., Fe-chelates and Fe-salts); FeDTPA and FeEDTA determined spectophotometrically is specific to those forms of Fe only.

Analysis of precipitate. A base nutrient solution, previously described (Albano and Miller, 1996) containing FeDTPA or FeEDTA was prepared as a $4\times$ concentrate stock based on a 14.28 mmol·L⁻¹ N (17.9 µmol·L⁻¹ Fe) 1× concentration. Nutrient solutions (8 L) were

Received for publication 15 Mar. 2000. Accepted for publication 19 July 2000. We thank Dennis R. Decoteau, Thomas M. McInnis, W. Vance Baird, and William C. Bridges, Jr. for consultation in this research. We thank Beth Hardin for technical assistance. This research was supported in part by the Clemson Univ. Ornamental Horticulture Competitive Grants Program and The Fred C. Gloeckner Foundation. Use of trade names does not imply endorsement of the products named nor criticism of similar ones not named. The cost of publishing this paper was defrayed in part by the payment of page charges. Under postal regulations, this paper therefore must be hereby marked *advertisement* solely to indicate this fact.

contained in translucent 10-L LDPE carboys (Nalgene Co.). Nutrient solutions were irradiated for 9 d with 500 µmol·m⁻²·s⁻¹ of light (330-800 nm) as described above. The radiation source was HID, metal halide lamps. Solution temperature was not controlled and was ≈30 °C. The precipitate that formed during irradiation was analyzed by centrifuging 2.4 L of the solution at 12,000 g_n for 1 h in a Sorval GS-3 rotor at 22 °C. The pellet was resuspended in ≈30 mL of distilled-deionized water, vortexed, and centrifuged at $6000 g_n$ for 1 h. This washing step was repeated three times, then the pellet was dissolved in 24 mL of 1 N HCl. Upon centrifugation (6000 g_n), a clear, light yellow supernatant was obtained. The supernatant and pellet-derived solutions were analyzed for Fe, Mn, Zn, Cu, K, Ca, and Mg by AA, and colorimetrically for P according to Jackson (1958).

Kinetic analysis of Fe-chelate photodegradation. A base nutrient solution previously described (Albano and Miller, 1996) was prepared as a $5 \times$ or $10 \times$ concentrate stock based on a 14.28 mmol·L⁻¹ N 1× concentration containing either FeDTPA or FeEDTA. Solutions (400 mL of 5× or 900 mL of 10×) were contained in 500- or 1000-mL LDPE bottles, which were placed on their sides and irradiated for 10 d with 250 µmol·m⁻²·s⁻¹ or 500 μ mol·m⁻²·s⁻¹ of light (330–800 nm), measured at the external container surface. Control containers were covered with aluminum foil. Irradiance was varied by adjusting the distance from the lamp (fluorescent plus incandescent) bank to the container surface. Solution temperature was 20 °C or 40 °C and was maintained by adjusting air temperature. Controlledenvironment growth chambers were programmed to provide the combinations of solution temperature and irradiance indicated in Table 2. Three replications were used per treatment. At 24-h intervals, the containers were agitated and a 20-mL aliquot was drawn from each container and centrifuged at $6000 g_n$ for 20 min in a Sorval SA-600 fixed-angle rotor at 22 °C. The supernatant was analyzed for Fe, Mn, Zn, Cu, K, Ca, and Mg by AA, and colorimetrically for P according to Jackson (1958). Reaction order of FeDTPA and FeEDTA photodegradation was determined by plotting soluble Fe (μ mol·L⁻¹) (zero-order), the log of soluble Fe (first-order), and 1/soluble Fe (second order) vs. time, with the correct reaction order yielding a straight line. Photodegradation of FeDTPA and FeEDTA, based on the disappearance of soluble Fe over time, was determined to be a first-order reaction. The rate constant (k) was calculated from the raw data by linear regression analysis of the logarithm of soluble Fe vs. time (i.e., disappearance of substrate).

Light source spectra. The spectral distribution for the fluorescent plus incandescent and HID light sources and the transmission spectra for the fluorescent plus incandescent light source under spectral filter Acrylic yellow-2208 are presented in Fig 1. Spectra (330–800 nm) were measured at 10-nm intervals with a LI-1800 spectroradiometer (LI-COR) with a quantum sensor.

Statistics. Data were analyzed by analysis of variance (ANOVA) to determine the effects of treatments. Calculations were performed with the general linear model (GLM) procedure of SAS (SAS Institute, Cary, N.C.). Means were separated and planned comparisons were made using LSD or pairwise *t* tests.

Results and Discussion

Fe-chelate photodegradation in solution. Freshly prepared (nonirradiated) solutions of FeDTPA and FeEDTA absorbed strongly in the UV and blue regions of the spectrum (Fig. 2). Upon irradiating Fe-chelate solutions for 4 d with 500 μ mol·m⁻²·s⁻¹ (330–800 nm) from a fluorescent plus incandescent light source, soluble Fe decreased (data not shown) in conjunction with the formation of a precipi-

tate and a decrease of absorbance in the blue and UV regions of the spectrum, indicating the loss of the Fe-chelate complex (Fig. 2). In the irradiated nutrient solutions, the loss of FeDTPA and FeEDTA (Fig. 3A) paralleled the loss of soluble Fe (Fig. 3B), confirming that: 1) FeDTPA photodegrades in lab-prepared nutrient solutions; and 2) the destruction of the chelate results in Fe precipitation in such solutions. The solutions of FeDTPA that were irradiated under a spectral filter with a wavelength cutoff below 454 nm (Acrylic yellow-2208) neither photodegraded, nor lost soluble Fe or formed of a precipitate (Fig. 4A). This indicates that the wave-bands of absorbance (<454 nm) are responsible for the photodegradation of the chelate and subsequent loss of soluble Fe from FeDTPA in solution. These results are similar to those reported for FeEDTA incorporated into



Fig. 1. Spectral photon distribution of a fluorescent plus incandescent light source providing 250 μ mol·m⁻²·s⁻¹ or 500 μ mol·m⁻²·s⁻¹ (\bullet). Transmission spectra of same light source under spectral filter Acrylic yellow-2208 (wavelength cutoff below 454 nm) (O). Spectral energy distribution of a HID light source providing 500 μ mol·m⁻²·s⁻¹(-). Spectra are relative to 580 nm (irradiance = 1) for the fluorescent and incandescent light source, and 590 nm (irradiance = 1) for the HID light source, the wavelengths of maximal irradiance, respectively. Readings were made at 10-nm intervals.



Fig. 2. Absorbance by 89.5 µmol·L⁻¹ solutions of FeDTPA [(**A**) nonirradiated and (**C**) irradiated (supernatant)] and FeEDTA [(**B**) nonirradiated and (**D**) irradiated (supernatant)]. Solutions were irradiated with 500 µmol·m⁻²·s⁻¹ from a fluorescent plus incandescent light source. Soluble Fe concentration in the supernatant of the irradiated solutions was 0. Maximal absorbance by FeDTPA and FeEDTA was 260 and 258 nm, respectively.



Fig. 3. (A) FeDTPA and FeEDTA determined spectrophotometrically at 260 or 258 nm, respectively, and (B) soluble Fe determined by atomic absorption spectrophotometry for a lab-prepared nutrient solution. Nutrient solutions were 5× stocks (14.28 mmol·L⁻¹ N, 17.9 μ mol·L⁻¹ Fe is 1×) irradiated at 30 °C with a HID light source providing 500 μ mol·m⁻²·s⁻¹(330–800 nm) measured at the surface of a 500-mL LDPE container. No absorbance was detected in solutions without Fe-chelate. Vertical bars indicate se (*n* = 4). If none are shown, they fall within the dimensions of the plotting symbol.



Fig. 4. (A) Loss of soluble Fe in linear plot vs. time and (B) linear regression profile of the logarithm of soluble Fe (µmol·L⁻¹) vs. time (h) for 5× or 10× lab-prepared nutrient solutions containing FeDTPA or FeEDTA (14.28 mmol·L⁻¹ N, 17.9 µmol·L⁻¹ Fe = 1×). Nutrient solutions (20 °C or 40 °C) were irradiated with a fluorescent plus incandescent light source providing 250 µmol·m⁻²·s⁻¹ or 500 µmol·m⁻²·s⁻¹ (330–800 nm) measured at the outer surface of the container. Treatments were: 1) FeEDTA, 10×, 20 °C, 250 µmol·m⁻²·s⁻¹ (●); 2) FeDTPA, 10×, 20 °C, 250 µmol·m⁻²·s⁻¹ (○); 3) same as 2) but wrapped in aluminum foil (△); 4) same as 2), but at 40 °C (□); 5) same as 2), but at 500 µmol·m⁻²·s⁻¹ (■); 6) same as 5), but 5× and under spectral filter Acrylic yellow-2208 (▲). All containers were 1 L except for the 5× treatment, which was a 500-mL container. Vertical bars indicate set (*n* = 3). If none are shown, they fall within the dimensions of the plotting symbol.

tissue-culture agar; irradiation from fluorescent lamps degraded the FeEDTA, but this could be prevented using spectral filter Acrylic yellow-2208 to remove the region of the spectrum in which the Fe-chelate maximally absorbed (Hangarter and Stasinopoulos, 1991).

Iron (98% of total) and Mn (5% of total) were lost following irradiation of FeDTPA (Table 1). Analysis of the yellow-tan precipitate that formed upon irradiation indicated that it was primarily composed of Fe, a small amount of Mn (Table 1), and trace amounts of Zn, P, K, Ca, and Mg (data not shown). The precipitate contained 85% of the soluble Fe lost (Table 1). No precipitate formed in irradiated nutrient solutions containing no FeDTPA (data not shown). These data indicate that the photodegradation of nutrient solutions primarily affects only FeDTPA in solution and that the solubility of other nutrients is generally unaffected (remaining \geq 95% soluble) by this photochemical event. Manganese in these studies was supplied as MnEDTA. The chelating-agent EDTA has a higher affinity for, and forms a more stable complex with, Fe than with Mn within a pH range of 4.0-6.3 (Laurie et al., 1991). Therefore, we speculate that Fe released from chelation with DTPA by photodegradation may replace Mn on the MnEDTA chelate complex, causing precipitation of Mn as an oxide and the photodegradation of the EDTA molecule.

The by-products of EDTA, and presumably DTPA, photodegradation are amine residues (Hamaker, 1956). Upon irradiating FeEDTA, carboxyl groups are lost, and the photostability of the amine residue by-product increases (i.e., photostability: ethylenediaminetetraacetic acid < ethylenediaminetriacetic acid < ethylenediaminediacetic acid)(Hamaker, 1956). These by-products are capable of chelating Fe (Hamaker, 1956) and may partially account for the residual levels of Fe-chelate remaining in the irradiated nutrient solutions (Fig. 3A). The residual levels of Fe-chelate remaining in the irradiated nutrient solutions may also be due to Mn and Cu ions and/or chelates that can be a source of interference for the spectrophotometric detection of Fe-chelates (Hill-Cottingham, 1957).

The pH of the FeDTPA- or FeEDTA- containing nutrient solutions rose slightly when irradiated (Fig. 5). Amines are bases and their accumulation during photodegradation of the chelating agent may account for the rise in pH when such solutions are irradiated.

Effects of light intensity and temperature. Lab-prepared nutrient solutions lost >90% of their soluble Fe during exposure to the light (fluorescent plus incandescent source) and temperature environments of 250 µmol·m⁻²·s⁻¹ at 20 °C, 500 µmol·m⁻²·s⁻¹ at 20 °C, or 250 µmol·m⁻²·s⁻¹ at 40 °C (Fig. 4A). Plotting log [soluble Fe] vs. time generated straight lines for all treatments ($r^2 \ge 0.97$), indicating that the reactions, as determined by the disappearance of soluble Fe, were first order (Fig. 4B, Table 2).

Linear regression analysis of the data in Fig. 4 indicated differences in rate constants (*k*) for both temperature and irradiation. At 20 °C, doubling the irradiance from 250 to 500 μ mol·m⁻²·s⁻¹ (330–800 nm) resulted in a



Fig. 5. pH of irradiated or nonirradiated lab-prepared nutrient solution vs. time of irradiation at 30 °C. Solutions containing either (**A**) FeDTPA or (**B**) FeEDTA were prepared as $5 \times \text{stocks}$ (14.28 mmol·L⁻¹ N, 17.9 µmol·L⁻¹ Fe = 1×) and were irradiated with a HID light source providing 500 µmol·m⁻²·s⁻¹ (330–800 nm) measured at the surface of a 500-mL LDPE container. Vertical bars indicate sE (*n* = 4). If none are shown, they fall within the dimensions of the plotting symbol.

Table 1. Iron and Mn composition of supernatant and pellet fractions derived from nonirradiated (NIr) or irradiated (Ir) lab-prepared nutrient solutions. Nutrient solutions were irradiated as 4× stock solutions (14.28 mmol·L⁻¹ N, 17.9 μ mol·L⁻¹ FeDTPA = 1×) for 9 d with 500 μ mol·m⁻²·s⁻¹ from a HID light source.

	Supernatant (µmol·L ⁻¹)		Loss due to	Ir-Pellet	Recovered	
Element	NIr ^z	Ir	irradiation (%)	$(\mu mol \cdot L^{-1})^y$	(%)	
Fe	71.6	1.79	98	59.070	85.0	
Mn	36.4	34.58	5	0.016	0.9	
Fe : Mn ratio	2:1	1:19		3692:1		

^zNo precipitate formed in the nonirradiated solution.

^yThe pellet derived from the irradiated solution was dissolved in 1 N HCl prior to analysis, therefore, data are reported as concentrations.

Table 2. Linear regression (slope, *r*²) and kinetic (*k*) data describing the first order reaction of soluble Fe disappearance from solutions irradiated with a fluorescent plus incandescent light source for 10 d at the temperatures and irradiance levels indicated. Solution temperature was maintained by adjusting air temperature, and irradiance was varied by adjusting the lamp bank distance to containers within a controlled-environment growth chamber. Each solution controlled-environment growth chamber. Each solution (14.28 mmol·L⁻¹ N, 17.9 μmol·L⁻¹ Fe is 1×). There was no loss in soluble Fe in nonirradiated treatments or irradiated treatments under spectral filter Acrylic yellow-2208 (wavelength cut off 454 nm) at either solution temperature or irradiance.

Fe source	Solution temp. (°C)	Irradiance (μ mol·m ⁻² ·s ⁻¹ at 330–800 nm)	Time to ≥90% sol. Fe loss (h)	$\begin{array}{c} Slope^z \\ (\cdot 10^{-4} \cdot h^{-1}) \end{array}$	r ^{2z}	Rate constant ^z $k(\cdot 10^{-5} \cdot \text{min}^{-1})^{y}$
FeEDTA	20	250	96 a ^x	-4.6 c	0.986	1.8 c
FeDTPA	20	250	96 a	-5.0 c	0.970	1.9 c
	20	500	48 c	-10.0 a	0.999	3.8 a
	40	250	72 b	-6.3 b	0.995	2.4 b

^zCalculated to the time point of 90% Fe loss. Rate constant (*k*) was calculated from raw data by linear regression analysis of the logarithm of soluble Fe vs. time (i.e., disappearance of substrate). $y_k = -2.303 \cdot (\text{slope-min}^{-1}).$

^xMeans separation within columns by LSD at $P \le 0.05$. n = 3.

doubling of the rate constant (*k*) of FeDTPA photodegradation (Table 2). At 250 μ mol·m⁻²·s⁻¹ irradiance, a 20 °C increase in temperature from 20 to 40 °C resulted in a 26% increase in the rate constant (*k*) of FeDTPA photodegradation (Table 2). Most thermochemical reactions have a Q₁₀ of 2 to 3 (Petrucci and Wismer, 1993). The low Q₁₀ confirms that the reaction is a photochemical event.

Conclusions. We have demonstrated that FeDTPA incorporated into lab-prepared nutrient solutions is vulnerable to photodegradation, resulting in selective Fe insolubility, but generally not affecting the solubility of other nutrients. The rate of FeDTPA photodegradation increased with temperature or irradiance, but irradiance had a far greater effect. Photodegradation of FeDTPA or FeEDTA resulted in a rise in solution pH. FeDTPA absorbs in the blue and UV regions of the spectrum; removal of these wavelengths with a spectral filter prevented photodegradation. The consequences of using a photodegraded FeDTPA-containing nutrient solution on plant growth and physiology are presented in a second report (Albano and Miller, 2001).

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