



Review

Phytotherapy in the Management of Diabetes: A Review

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Abstract: Phytotherapy has long been a source of medicinal products and over the years there have been many attempts to use herbal medicines for the treatment of diabetes. Several medicinal plants and their preparations have been demonstrated to act at key points of glucidic metabolism. The most common mechanisms of action found include the inhibition of α -glucosidase and of AGE formation, the increase of GLUT-4 and PPARs expression and antioxidant activity. Despite the large amount of literature available, the actual clinical effectiveness of medicinal plants in controlling diabetes-related symptoms remains controversial and there is a crucial need for stronger evidence-based data. In this review, an overview of the medicinal plants, which use in the management of diabetes is supported by authoritative monographs, is provided. References to some species which are currently under increasing clinical investigation are also reported.

Keywords: phytotherapy; hyperglycemia; diabetes; medicinal plants

1. Introduction

In the last decade, the concept of metabolic syndrome (MS) has been extensively debated by the scientific community [1–3]. Despite the difficulty in establishing an exhaustive definition [4], MS is nowadays recognized as a major cardiovascular disease risk factor by the World Health Organization (WHO) and other institutions such as the International Diabetes Federation (IDF) [5–7]. MS can be defined as a concurrence of conditions, including obesity, hypertension, dyslipidemia and altered glycaemia [8]. MS is associated with a higher risk of type 2 diabetes and cardiovascular diseases onset [9] and involves about 25% of the world's adult population [10], with women having a higher risk of developing MS [11].

Phytotherapy has long been a source of medicinal products and there have been many attempts to use herbal medicines for the treatment of diabetes over the years [12,13]. Furthermore, the number of scientific publications regarding herbal medicine and type 2 diabetes is continuously increasing [14].

Among the possible mechanisms of action of natural products in diabetes, such as the inhibition of α -glucosidase and α -amylase, the effects on glucose uptake and glucose transporters, the enhancement of insulin secretion and of pancreatic β -cell proliferation, the inhibition of protein tyrosine phosphatase 1B activity and the antioxidant activity have been studied in depth [15].

Despite the large amount of available literature, the real clinical effectiveness of medicinal plants in the management of diabetes is still controversial and there is a crucial need for stronger evidence-based data [16]. Indeed, despite a long folk medicine history, most of the popular species used suffer from clinical inconsistency, mainly due to the poor quality of the clinical studies [17]. Another aspect to be considered is the variability of the raw herbal materials and preparations used, which may lead to the non-reproducibility of results among different trials [18].

The aim of this review is to provide an overview of the use of medicinal plants in the management of diabetes, focusing on the species that are supported by authoritative documents such as the monographs drafted by the World Health Organization (WHO). Furthermore, an emphasis on some of the most promising species, which are attracting the interest of the scientific community, is also provided.

2. Medicinal Plants Used for the Management of Diabetes

Medicinal plants possessing therapeutic uses in diabetes and supported by clinical data or described in pharmacopoeias and well established documents are covered in the WHO monographs and are reported in Table 1.

Table 1. Species enlisted in WHO monographs with indication of use for diabetes.

Use Supported by Clinical Data	Use Described in Pharmacopoeias and in Traditional Systems of Medicine
<i>Ocimum tenuiflorum</i> L., folium <i>Trigonella foenum-graecum</i> L., semen	<i>Allium cepa</i> L., bulbus <i>Azadirachta indica</i> A. Juss., folium <i>Momordica charantia</i> L., fructus <i>Ocimum tenuiflorum</i> L., folium <i>Panax ginseng</i> C.A. Meyer, radix <i>Panax quinquefolius</i> L., radix <i>Rehmannia glutinosa</i> (Gaertn.) DC., radix

2.1. *Allium cepa* L., Bulbus

Allium cepa L. is a perennial herb belonging to the Amaryllidaceae. The parts of the plant used are the fresh or dried bulbs, commonly known as onion, which are commercially cultivated worldwide [19]. The main chemical constituents are sulfur-containing compounds, such as L-cysteine sulfoxides, and flavonoids, such as quercetin and its glycosides [20]. *A. cepa* seems to exert its antidiabetic activity regardless of the form in which it is administered (i.e., extracts [21–24], juice [25], freeze-dried powder [26], essential oil [27]) [28].

A preliminary study evaluated the hypoglycemic effects of the oral administration of small slices of *A. cepa* (100 g/day) in type 1 and type 2 diabetic patients. Onion exhibited significant antidiabetic effects, reducing fasting blood glucose by about 89 mg/dL in type 1 diabetes patients and by 40 mg/dL in type 2 diabetes patients. A reduction of the induced hyperglycemia by 120 mg/dL in the diabetes 1 group and by 159 mg/dL in the type 2 diabetes was also observed [29].

In 2009, an in vivo study demonstrated that *A. cepa* (7% freeze-dried onion powder added into control diet) may represent an interesting anti-hyperglycaemic dietary adjunct for diabetic therapy, since it decreases serum cholesterol, triacylglycerol and LDL-cholesterol in streptozocin-induced diabetics rats, without alterations in the cholesterol and HDL-cholesterol levels [30]. Hyperglycemia causes glucose autoxidation, impaired mitochondrial bioenergetics and induces reactive oxygen species (ROS) production, leading to an impairment of intracellular pathways (i.e., JAK/STAT, JNK, p38, ERK/MAPK) and to insulin resistance [31]. Onion (400 mg/day) possesses a significant free radical-scavenging property and exerts a regulation on lipid metabolism, decreasing superoxide dismutase activity and lowering lipid hydroperoxide and lipoperoxide concentrations in diabetic rats [32].

A. cepa exerts its antidiabetic activity through multiple pharmacologic actions attributed to the presence of many active constituents: for example, quercetin is responsible for α -glucosidase inhibition [33] and, along with rutin, for the increase of GLUT-4 translocation, glucose uptake and insulin action [34]. Differently, L-cysteine sulfoxides and allyl propyl disulphide can act directly as free radicals scavengers. In fact, they take part in the redox process of glutathione and cysteine, and can also increase the activity of superoxide dismutase and catalase, independently or through the stimulation of insulin secretion [35,36].

2.2. *Azadirachta indica* A. Juss., *Folium*

Azadirachta indica A. Juss., also known as neem, is a deciduous tree belonging to the Meliaceae. The parts of the plant used are the dried leaves [37]. It contains characteristic compounds, such as oxidized tetranotriterpenes, known as azadirachtins [38].

An ethanolic extract (400 mg/kg) obtained from neem leaves demonstrated several effects, such as anti-lipid peroxidation, anti-hyperglycaemic and anti-hypercholesterolaemic activities as well as a reduction in serum triglyceride levels in alloxan-induced diabetic rats [39].

Two water extracts were also tested in high-fat diet-induced diabetic rats (400 mg/kg) and in normal and alloxan-induced diabetic rabbits (500 mg/kg), showing a partial prevention of the rise in blood glucose levels and a normalization of the altered levels of serum insulin, lipid profile and insulin signaling molecules as well as GLUT-4 proteins [40,41].

Chloroform extracts activities were studied in vitro and in vivo in streptozocin-induced diabetic rats (200–300 mg/kg) produced an attenuation of non-enzymatic glycation, inhibiting advanced glycation end product (AGE) formation, and alleviated oxidative stress, increasing the levels of antioxidant enzymes, glucose-6-phosphatase, hepatic glycogen content and insulin plasma levels, and decreasing glucokinase and lipid peroxidation [42,43]. The main mechanism of action of azadirachtins (e.g., azadirachtolide, azadiradione, gedunin and meliacinolin) is the inhibition of α -amylase and α -glucosidase [44–46].

2.3. *Momordica charantia* L., *Fructus*

Momordica charantia L. is a monoecious annual climbing vine belonging to the Cucurbitaceae. The parts of the plant used are the fresh or dried fruits, known as bitter melons [47]. The main chemical constituents are sterols, triterpenes and bioactive proteins [48].

The clinical potential of bitter melon has been examined administering capsules or tablets containing preparations from bitter melon fruits or leaves, in diabetic patients. No statistically significant improvement of blood glucose control, in terms of normalization or reduction of fasting blood glucose, reduction of glycosylated haemoglobin A1c or fructosamine, compared to placebo have been observed [49–51]. More recently, *M. charantia* capsules (500 mg of dried powder of the fruit pulp, containing 0.04–0.05% of charantin), administered in type 2 diabetes patients (2000 mg/day), demonstrated a significant decrease in fructosamine levels after 4 week of treatment and no side effects were observed [52].

The whole plant and/or different plant parts such as fruit pulp, seeds, and leaves have been reported to possess hypoglycemic and anti-hyperglycemic activities in several animal models: in particular, bitter melon reduced blood glucose levels and increased plasma insulin [53].

The activation of the AMP-activated protein kinase (AMPK) system and a role of the α - and γ -peroxisome proliferator-activated receptors (PPAR α and PPAR γ) have been hypothesized as possible mechanisms of action for *M. charantia* in diabetes treatment [54–56].

Cucurbitane triterpenoids from *M. charantia* displayed hypoglycemic effect, reducing blood glucose levels, moderating insulin secretion activity, increasing glucose uptake through stimulation of GLUT-4 translocation and increasing the phosphorylation of AMPK and insulin receptor substrate-1 [57–61].

Among bitter melon triterpenoids, 5 β ,19-epoxy-25-methoxy-cucurbita-6,23-diene-3 β ,19-diol (EMCD) and momordin have been studied in detail; it has been observed that EMCD (20 μ M) can suppress the TNF- α induced expression of iNOS and nuclear translocation of NF- κ B in FL83B mouse liver cells [62], while momordin is responsible for the increase of PPAR δ mRNA expression at nanomolar concentrations in HepG2 human liver cells [63].

Despite the copious data from in vitro and in vivo studies, the available clinical data are often flawed by small sample size, lack of control and poor study designs. Better-designed clinical trials with sufficient sample size and statistical power will be indispensable to further confirm the efficacy of *M. charantia* as a natural treatment for diabetes mellitus [64].

2.4. *Ocimum tenuiflorum* L., *Folium*

Ocimum tenuiflorum L., commonly referred to as tulsi, is a herb or shrub up to 1 meter high belonging to the botanical family Lamiaceae. It is indigenous to India and parts of north and eastern Africa, Hainan Island and Taiwan, China and the herbal substance is represented by the fresh or dried leaves [65]. The main chemical components are tannins and essential oil (mainly composed of eugenol, methyleugenol, and α - and β -caryophyllene) [66].

In 1996, a randomized, placebo-controlled, crossover single blind trial analyzed the effects of *O. tenuiflorum* and *O. album* leaves on fasting and postprandial blood glucose and serum cholesterol levels in patients with noninsulin-dependent diabetes mellitus. A significant decrease in fasting and postprandial blood glucose levels, a similar trend in urine glucose, and a mild reduction of mean total cholesterol levels during treatment period were observed [67].

More recently, a randomized, parallel group, open label pilot study investigated the effect of tulsi extract on metabolic and biochemical parameters in 30 young overweight/obese subjects. The supplementation with *O. tenuiflorum* capsules (250 mg twice daily for 8 weeks) decreased plasma insulin and insulin resistance by 28.49% and 24.79% respectively, caused the normalization of serum lipid profile, and reduced body weight and BMI, compared to the control group (no intervention) [68].

Animal studies showed that the oral administration of *O. tenuiflorum* aqueous extracts (200 mg/kg) could delay the development of insulin resistance. Indeed, tulsi determined a considerable improvement in fasting blood glucose and in glucose tolerance and a correction of the abnormal lipid profile, through the reduction of serum total and LDL cholesterol levels. An increased activity of antioxidant enzymes (i.e., glutathione peroxidase, glutathione S-transferase, superoxide dismutase and catalase), as well as the increase of reduced glutathione levels, have been proposed as a mechanism of action for the decreased lipid peroxidation induced by tulsi [69–71].

Ethanol (80% *w/w*) extracts obtained from *O. tenuiflorum* leaves were effective in lowering blood glucose levels in normal, glucose fed hyperglycemic and streptozocin-induced diabetic rats, also potentiating the action of exogenous insulin in normal rats [72]. The glucose lowering effects was found to be mediated through its insulin secretagogue effects on ex vivo rat pancreas and in BRIN-BD11 rat clonal β -cells [73].

A reduction in the level of hepatic lipids and the reversion of the diminution of lipoprotein lipase, plasma postheparin lipolytic and lecithin cholesterol acyl transferase activities was obtained by administering 500 mg/kg of a different *O. tenuiflorum* ethanolic (95% *w/w*) leaves extract for 15 days in streptozocin-induced diabetic rats [74].

It has been proposed that the fixed oil extracted from *O. tenuiflorum* leaves (46.54 mg/kg/day for three weeks) may be responsible for the free radical scavenging activity, for the decrease in plasma glucose and for the increase in insulin release observed in streptozocin-induced diabetic rats, and this has been related to the content in α -linolenic acid [75].

A tetracyclic triterpenoid, 16-hydroxy-4,4,10,13-tetramethyl-17-(4-methyl-pentyl)-hexadecahydro-cyclopenta[*a*]phenanthren-3-one, isolated from the aerial part of *O. tenuiflorum*, was able to decrease serum glucose levels, total cholesterol, triglycerides and HDL cholesterol and to increase serum LDL cholesterol level in alloxan-induced diabetic rats [76].

2.5. *Panax Ginseng* C.A. Meyer, *Radix* and *Panax quinquefolius* L., *Radix*

Ginseng has been used in traditional Chinese medicine for more than 5000 years [77]. Thanks to its restorative, tonic, nootropic, and anti-aging properties [78], ginseng preparations have been applied to several pathological conditions such as hypodynamia, anorexia, shortness of breath, palpitation, insomnia, impotence, hemorrhage and diabetes [79].

Confirming the clinical strength of many of the traditional uses of ginseng, in 2014 the European Medicines Agency released a monograph which validated the use of ginseng as a traditional medicine for treating asthenia in Western countries too [80].

In medicine, the term ginseng may usually refer to several *Panax* species, which are often used indiscriminately. Nevertheless, the most commonly used are *Panax ginseng* C.A. Meyer and *Panax quinquefolius* L. In this review, the antidiabetic properties of *P. ginseng*, *P. quinquefolius* and their main constituents will be discussed together.

Panax ginseng C.A. Meyer is a perennial herb, native to Korea, China and Japan, with characteristic branched roots belonging to the family of Araliaceae. The herbal substance is represented by the dried roots, whose main constituent are triterpenes saponins, known as ginsenosides [19].

Panax quinquefolius L. is a perennial herb that is native to North America. The parts of the plant used are the dried roots, containing ginsenosides, with a higher concentration of protopanaxadiols compared to *P. ginseng* [81].

Several systematic reviews described the potential of ginseng in the management of diabetes. In 2011, Kim and coworkers analyzed data deriving from four different randomized clinical trials in which ginseng (0.78–6 g/day for a maximum of 12 weeks) demonstrated no significant effects in controlling blood glucose in type 2 diabetes patients [82]. Nevertheless, promising results in improving glucose metabolism were found by Shergis and colleagues in 2013, by analyzing six clinical trials [83]. More recently, Shishtar and colleagues evaluated sixteen trials in which 0.1–20 g/day of different ginseng preparations were administered for 4–24 weeks to patients with and without diabetes. A modest yet significant reduction of fasting blood glucose was observed in both groups [84]. Finally, in 2016, a meta-analysis from Gui and coworkers failed to attribute significant improvement in hemoglobin A1c levels for ginseng treated patients (0.96–13.6 g/day for 4–20 weeks) alone and in combination with conventional therapies, compared to control group. However, improved fasting glucose and post prandial insulin levels were observed when ginseng was administered alone [85].

The effects of ginseng extracts and their main constituents have been extensively studied in vivo. Ginseng total saponins (150–300 mg/kg) significantly reduced hyperglycemia by increasing glucagon-like peptide-1 in high fat and low streptozocin-induced diabetic rats [86]. A fractionated extract containing water soluble ginseng polysaccharides, administered 1 g/kg/day for 2 weeks to streptozocin-induced diabetic rats, caused significant effects on purine, tryptophan, fatty acids and energy metabolism [87]. Protopanaxadiol and protopanaxatriol-type saponins (50–150 mg/day) reduced fasting blood glucose, glucose tolerance and insulin resistance in high fat and low streptozocin-induced diabetic rats. Among the mechanism of action analyzed, a suppression of TNF- α and IL-6 release, an increase in superoxide dismutase and a decrease in malondialdehyde levels, together with the downregulation of PPAR- γ coactivator 1 α , phosphoenolpyruvate carboxykinase and glucose-6-phosphatase were observed [88]. In the same animal model, compound K, a metabolite of panaxadiol, (30–100–300 mg/kg) caused a dose dependent reduction of fasting blood glucose and an enhancement of fasting serum insulin and insulin sensitivity, by promoting the expression of insulin receptor, insulin receptor substrate-1, PI3Kp85, pAkt and GLUT-4 [89]. Ginsenoside-Rg1 was administered 10 mg/kg to streptozocin-induced diabetic mice, improving angiogenesis by increasing eNOS activation, VEGF expression and inhibiting apoptosis [90]. Two in vivo studies described the effects of 20(S)-ginsenoside-Rg3 (20 mg/kg) in streptozocin-induced diabetic rats. The first one reported a decrease in water intake and urine volume, together with a reduction in serum glucose, glycosylated protein and thiobarbituric acid-reactive substances production, leading to an improvement in renal dysfunction compared to control, which was related by the authors to the inhibition of NMDA

receptor-mediated nitrosative stress [91]. The second further describes its positive effects on the metabolism of nucleic acid, energy and gut flora [92]. Finally, ginsenoside-Rh2 (1 mg/kg) lowered plasma glucose in streptozocin-induced diabetic rats, by increasing β -endorphin secretion, which is responsible for opioid μ -receptor activation, resulting in an increase of GLUT-4 expression [93].

It is then worth mentioning that several innovative extraction methods are currently under investigation in order to improve ginseng pharmaceutical applications. A heat-processed Korean ginseng extract was administered 100 mg/kg to streptozocin-induced diabetic rats, causing a reduction of blood glucose levels and an improvement of renal dysfunction without altering the expression of proteins involved in oxidative process, in a more potent manner compared to standard Korean ginseng extract [94]. Black ginseng, obtained by nine cycles of steaming and drying of ginseng, was administered 200 mg/kg to streptozocin-induced diabetic rats in two different studies. A modulation of glucose metabolism [95] as well as a more effective reduction of hyperglycemia, increase in insulin/glucose ratio and improvement of islet architecture and β -cells function was observed compared to standard red ginseng [96]. The inhibition of β -cells apoptosis was presumed by the authors to be related to the suppression of cytokine-induced NF- κ B translocation. In the same animal model, a pectin lyase-modified ginseng extract (20–50–100 mg/kg) decreased the serum levels of AGE and their cross-linking with protein [97]. Finally, a tissue cultured mountain ginseng adventitious root extract enriched with ginsenosides, administered 250–500 mg/kg, was more effective than field cultivated Korean ginseng in lowering blood glucose, total cholesterol and triglycerides in streptozocin-induced diabetic rats [98].

2.6. *Rehmannia glutinosa* (Gaertn.) DC., Radix

Rehmannia glutinosa (Gaertn.) DC. is a perennial herb belonging to the family Plantaginaceae. The parts of the plant used are the dried roots and rhizomas [37]. The main constituents are iridoid glycosides and monoterpenes [99].

Within the various *R. glutinosa* preparations, the aqueous extract is preferred since it contains the largest amount of characteristic constituents [100]. The whole extract, the polysaccharides fraction and some isolated compounds exhibited beneficial activities in improving glucolipid metabolism and redox homeostasis (which is referred to as the balance between oxidant and antioxidant signaling [101]) in both in vitro and in vivo models. In particular, a water extract obtained from fresh rhizome exerted a high free radical scavenging activity and, in addition, was able to reduce ROS production, to suppress NF- κ B activity and to down-regulate the expression of pro-inflammatory genes, such as TNF- α , COX-2, monocyte chemoattractant protein-1 (MCP-1) and inducible protein-10 [102]. Furthermore, two different polysaccharide fractions exerted a significant hypoglycemic effect in normal, glucose- and alloxan-induced diabetic rats at 100 mg/kg (reducing hepatic glucose-6-phosphatase activity, increasing hepatic glycogen content, and raising plasma insulin levels) [103], and in streptozocin-induced diabetic mice at 20, 40 and 80 mg/kg (increasing the mRNA expression of phosphoenolpyruvate and the hepatic glycogen content) [104].

A water extract fraction containing approximately 60.51% of stachyose was administered 200 mg/kg/day for 15 days to normal, glucose-, adrenaline- and alloxan-induced diabetic rats, resulting in a significant hypoglycemic effect, although the mechanism of action was not investigated [105].

Catalpol at micromolar concentrations reduced AGE-induced inflammatory responses, inhibited the formation of intracellular ROS production and had a suppressive effect on NADPH-oxidase activity in THP-1 cells [106]. The protective effects of catalpol (20–120 mg/kg/day) through suppression of AGE-mediated inflammation have been extensively confirmed in animal models [107–110].

2.7. *Trigonella foenum-graecum* L., Semen

Trigonella foenum-graecum L., fenugreek, is an annual aromatic herb belonging to the family Fabaceae. The parts of the plant used are the dried ripe seeds, which contains mucilage and a variety of other secondary metabolites such as trigonelline [37].

A broad range of therapeutic uses, such as to ease childbirth, to increase milk flow, to alleviate menstrual pains and to treat body weakness, have been described in the traditional medicine of eastern Mediterranean areas [111].

Although results from clinical trials are substantially heterogeneous and there is still very limited evidence as to the impact of dietary consumption or supplementation with *T. foenum-graecum* for the management of diabetes, recent systematic reviews and meta-analysis conclude that fenugreek seeds (5–100 g/day) may be a promising complementary option for the clinical management of diabetes. This drug, indeed, contributes to a better glycemic control in type 2 diabetes mellitus patients, reducing fasting blood glucose, 2 h post load blood glucose and glycated haemoglobin [112–115].

In vivo studies agree on fenugreek's hypoglycemic and hypolipemic activities: the seeds powder (5% in the diet for 21 days) decreased the level of lipid peroxidation in alloxan-induced diabetic rats [116], and significantly restored to control values the elevated fasting blood glucose levels in the same animal model (2 g/kg for 7 days) [117].

The mechanisms underlying fenugreek antidiabetic action include the lowering of blood glucose through an insulin signal pathway and the stimulation of glucose uptake in peripheral tissues [118].

The bioactive compounds which has been more deeply studied for the hypoglycemic actions are trigonelline, diosgenin, 4-hydroxyisoleucine and the soluble dietary fiber fraction of fenugreek seeds [119]. Aside from possessing antioxidant activity, trigonelline affects the activity of enzyme related to glucose metabolism, β -cell regeneration and insulin secretion [120]. Diosgenin is implicated in the renewal of pancreatic β -cells and in the stimulation of insulin secretion, has antioxidant effects and promotes adipocyte differentiation and enhancement of insulin-dependent glucose uptake [121–123]. 4-Hydroxyisoleucine stimulates glucose-dependent insulin secretion, reduces insulin resistance and inhibits sucrose α -D-glucosidase and α -amylase [124–127]. The soluble dietary fiber fraction of fenugreek (i.e., galactomannan) enhances glycemic control inhibiting lipid-hydrolyzing and carbohydrate-hydrolyzing enzymes in the digestive system and reducing the rate of glucose uptake [128–130].

3. Other Species with Promising Data for the Management of Diabetes

Several other species have been used in the ethnobotanical traditions of many countries around the world to treat diabetes [131–133], and most of them are under investigation for their potential role in the management of hyperglycemia and related diseases [134–136]. Currently, as retrieved by using the keywords “diabetes” and “phytotherapy” on PubMed, the clinical studies published on this topic are 254, and the review articles are 400 [137]. Nevertheless, the information on medicinal plants are various, with most of the species recording very few articles: many species are rarely used in Western medicine, but very common in other traditional medicine, such as traditional Chinese medicine [138]. Among the most used worldwide, the species reported in Sections 2.1–2.7 are the most studied. However, many other species have been recently considered by the scientific community.

Gymnema sylvestre (Retz.) R.Br. ex Sm., commonly known as gurmar, has been used since ancient times, particularly in Ayurvedic medicine, and its anti-obesity and anti-diabetic efficacy has been clinically demonstrated [139] and confirmed in animal models [140]. The anti-diabetic activity of gurmar has been attributed mainly to gymnemic acids, gymnemasaponins and gurmarin contained in the leaves [141]. A dihydroxygymnemic acetate (20 mg/kg), isolated from *G. sylvestre* leaves, has been administered in streptozocin-induced diabetic rats for 45 days, causing a significative reduction of plasma glucose and glycated hemoglobin level, and an increase of plasma insulin and muscle and liver glycogen [142]. The proposed mechanisms of action include the increase of insulin secretion and the promotion of islet cell regeneration, together with the reduction of intestinal and blood glucose adsorption [13]. Some products derived from *G. sylvestre* have been patented in different European countries. In 2010, a high molecular weight leaves extract (1 g/day for 60 days) was found to significantly increase the circulating insulin and C-peptide levels and reduced fasting and post-prandial blood glucose in a small cohort of patients with type 2 diabetes [143]. In the same year, a different

G. sylvestre leaves extract (500 mg/day for 3 months) was similarly able to reduce fasting and post prandial blood glucose and glycated hemoglobin, causing a favourable shift in lipid profile, in type 2 diabetic patients [144]. Nevertheless, more studies are needed in order to support the use of gurmar in the treatment of diabetic patients [145].

According to a recent systematic review, *Curcuma longa* L. can be considered a promising species for the management of impaired glucose tolerance, as curcumin is able to significantly reduce fasting blood glucose, glycosylated hemoglobin and insulin resistance after 3, 6 and 9 months of treatment [146]. Moreover, when administered together with an absorption enhancer (i.e., piperine 10 mg/day), curcuminoids (1000 mg/day) were able to reduce serum levels of atherogenic lipid indices, which are risk factors for cardiovascular events in type 2 diabetes patients [146].

Morus alba L. leaves demonstrated to possess a wide range of pharmacological activities in both in vitro and in vivo tests, including antidiabetic activity [147]. In a randomized double-blind placebo-controlled trial, the administration of *M. alba* leaf aqueous extract (5 g/day for 4 weeks) improved the postprandial glycemic control in patients with impaired fasting glucose tolerance, by reducing plasma glucose, insulin and C-peptide levels, compared to placebo [148].

The potential of ω -3 fatty acids on glycemic control is widely discussed [149]. *Linum usitatissimum* L. seed oil, also referred to as flaxseed oil, contains over than 50% ω -3 fatty acids, mainly represented by α -linolenic acid [150]. A recent randomized controlled interventional trial demonstrated the ability of *L. usitatissimum* seed oil (25 mg/day) to ameliorate some symptoms of metabolic syndrome, including blood pressure and lipid peroxidation [151]. In type 2 diabetic patients with coronary heart disease, the supplementation with *L. usitatissimum* oil (1000 mg/day for 12 weeks) increased gene expression levels of PPAR- γ and downregulated gene expression of lipoprotein(a), IL-1 and TNF- α [152]. Moreover, *L. usitatissimum* seed powder (10 g/day for 1 month), containing not quantified ω -3 fatty acids and lignans, reduced fasting blood glucose, glycated hemoglobin, triglycerides, total and LDL cholesterol and apolipoprotein B, and increase HDL cholesterol levels in type 2 diabetes patients [153]. Similar results were obtained by the supplementation with *L. usitatissimum* gum (5 g/day for 3 months) [154].

A standardized extract of *Aristotelia chilensis* (Molina) Stuntz berries, containing $\geq 25\%$ delphinidins and $\geq 35\%$ anthocyanins, has been investigated in a double-blind placebo-controlled crossover trial in patients with impaired glucose regulation, showing a significant reduction of postprandial blood glucose levels, which was related to the inhibition of sodium glucose cotransporter (SGLT-1) [155]. However, in a three months clinical trial on prediabetic individuals, the same extract (180 mg/day) caused a reduction in glycated hemoglobin level, but no significant effects were reported for fasting insulin and glucose levels [156].

Insulin-like proteins from *Moringa oleifera* Lam. leaves have been addressed for their potential role in the management of diabetes [157]. In alloxan-induced diabetic mice, a leaf protein isolate from *M. oleifera* (500 mg/kg) reduced blood glucose level and oxidative stress, but did not stimulate insulin secretion [158]. A reduction of blood glucose and a high antioxidant activity was also observed in streptozocin-induced diabetic rats treated with a *M. oleifera* leaves methanol extract (250 mg/kg) [159]. However, clinical validation has to be carried out, since very little information is currently available, with only one preliminary clinical trial been conducted by administering *M. oleifera* leaf powder to healthy subject and evaluating outcome related to diabetes [160].

Zingiber officinale Roscoe has demonstrated beneficial effects on hyperglycemia in several experimental studies, by increasing insulin sensitivity and synthesis, and glucose uptake by tissues, and by reducing oxidative stress, whilst protecting pancreatic β -cells [161]. In a double-blind placebo-controlled randomized clinical trial, the administration of *Z. officinale* powder (3 g/day for months), containing not quantified gingerols, in type 2 diabetic patients reduced serum glucose and glycated hemoglobin levels, decreased insulin resistance and increase total antioxidant capacity [162].

The anti-diabetic potential of several *Morinda citrifolia* L. preparations has been reviewed recently: although there is a wide number of products on the market, there is still the need for well-conducted clinical trials in order to better investigate the role of this herbal product in diabetes [163].

The consumption of tea (*Camellia sinensis* (L.) Kuntze) and coffee (*Coffea Arabica* L.) has been related to a lowered risk of type 2 diabetes onset [164] and these beverage were also found to be effective in impaired glucose tolerance. Some of the effects of coffee involved in the prevention of type 2 diabetes includes: improvement of glucose tolerance, insulin sensitivity and insulin secretion, reduction of glucose intestinal uptake and regulation of glucose metabolism [165]. Moreover, a randomized acute crossover intervention study conducted in healthy volunteers showed the ability of coffee polyphenols to improve postprandial hyperglycemia, increasing glucagon-like peptide 1 secretion and decreasing oxidative stress [166]. Type 2 diabetic patients drinking three cups (600 mL) of black tea per day for 12 weeks reported a significant reduction in glycated hemoglobin level, together with an amelioration of the immune function relevant to prevention and management of type 2 diabetes [167]. The effect of tea seems to be related to its polyphenol contents, and in particular to epigallocatechin-3-gallate, which may act on glucose intestinal and cellular uptake and on oxidative stress [168].

Within the most studied hypoglycemic herbal products, cinnamon is undoubtedly attracting a great interest by the scientific community. For this reason, although there is a lack in authoritative documents reporting specific indications in diabetes, a section of this review is dedicated to a critical analysis of the available literature on cinnamon and its role in diabetes.

Cinnamon

Botanical preparation of cinnamon may results from the dried inner bark of the shoots grown on cut stock of *Cinnamomum verum* J. Presl. as well as from the trunk bark, freed or cork of *Cinnamomum cassia* (L.) J. Presl., both species belonging to the family Lauraceae [19]. The parts of the plant used are the dried bark, free from the outer cork, which contains mainly cinnamaldehyde, eugenol and coumarin in concentrations that can vary abundantly between the two species [169]. Indeed, *C. verum* essential oil contains about 50–63% cinnamaldehyde and only traces of coumarins; *C. cassia* essential oil, instead, contains up to 95% cinnamaldehyde and up to 1% coumarins, together with an higher content of benzaldehyde and methoxycinnamaldehyde, compared to *C. verum* [170].

Many systematic reviews evaluating cinnamon effectiveness on diabetic patients has been published in the last five years. The oral consumption or supplementation with cinnamon, usually in combination with standard hypoglycemic medications or other lifestyle therapies, has been associated to modest effects on fasting plasma glucose and hemoglobin A1c [171], and to a decrease in levels of triglycerides, total and LDL cholesterol and to an increase in HDL cholesterol [172]. Although being judged as promising by Akilen and colleagues [173], the use of cinnamon on glycemic control need to be investigated in better conduced clinical trials [174–176].

In vitro and in vivo evidences indicate that cinnamon may have benefits in improving insulin sensitivity and glycaemic control. Its hypoglycaemic activity may be attributed to multiple mechanisms of action, including the stimulation of insulin release and insulin receptor signalling, the activation and regulation of enzymes involved in carbohydrate metabolism, glycolysis and gluconeogenesis (i.e., inhibition of pancreatic and intestinal amylase and glucosidase and increased of glycogen synthesis in the liver), stimulation of cellular glucose uptake and glycogen content (i.e., increased glucose transporter-4 receptor synthesis) and increased expression of PPARs [177–186].

It has been suggested that cinnamon's effects on blood glucose can be attributed to its active constituent, cinnamaldehyde. The insulinotropic effects of cinnamaldehyde have been preliminarily investigated and are thought indeed to be responsible for promoting insulin release, enhancing insulin sensitivity, increasing insulin disposal, and exerting activity in the regulation of protein-tyrosine phosphatase 1B and insulin receptor kinase [180,182,187]. It is important to consider that the quantity of active cinnamaldehyde may vary among species and formulations [188].

Even if cinnamaldehyde content is higher in *C. cassia*, compared to *C. verum*, it is difficult to state which of the two species is more effective in the management of diabetes. Moreover, the long-term consumption of coumarins have been demonstrated to cause hepatotoxicity in human [189,190] and,

in 2008, the European Food Safety Authority confirmed the previously calculated theoretical added maximum daily intake for coumarins to 0.1 mg/kg bw [191]. Considering the potential toxicity of coumarins in *C. cassia*, it can be speculated that *C. verum* may be safer for clinical application in chronic diseases requiring prolonged treatments, such as type 2 diabetes.

4. Conclusions

Traditional medicine and ethnobotany are an enormous source of information on safety and biological effects of herbal products and many of the medicinal plants currently used to treat hyperglycemia, indeed, derive from traditional use. Medicinal plants possessing anti-diabetes activities, which use has been officially recognized in one or more World regions and is supported by clinical evidence, are considered by WHO and enlisted in WHO monographs on medicinal plants.

Table 2. Pharmacological activities of the main chemical constituents of hypoglycemic medicinal plants.

Herbal Species	Main Chemical Constituents	Pharmacological Activities
<i>Allium cepa</i> L.	Quercetin	Inhibition of α -glucosidase
	Rutin	Increase of GLUT-4 translocation and glucose uptake; stimulation of insulin action
	L-cysteine sulfoxides Allyl-propyl disulphide	Free radical scavenging; increase of SOD and catalase activity
<i>Azadirachta indica</i> A. Juss.	Azadirachtins	Inhibition of α -amylase and α -glucosidase
<i>Momordica charantia</i> L.	Cucurbitane triterpenoids	Reduction of blood glucose levels; modulation of insulin secretion; stimulation of GLUT-4 translocation; upregulation of insulin receptor substrate-1; increase of AMPK phosphorylation
	EMCD	Reduction of TNF- α , iNOS expression and NF- κ B nuclear translocation
	Momordin	Induction of PPAR γ mRNA expression
<i>Ocimum tenuiflorum</i> L.	Essential oil	Reduction of lipid peroxidation; stimulation of antioxidant enzymes; stimulation of insulin secretion; free radical scavenging activity
<i>Panax ginseng</i> C.A. Meyer, and <i>Panax quinquefolius</i> L.	Protopanaxidiols	Increase of glucagone-like peptide-1; reduction of TNF- α and IL-6 release; increase of superoxide dismutase activity; reduction of malondialdehyde activity; down-regulation of PPAR- γ coactivator 1 α , phosphoenolpyruvate carboxykinase and glucose-6-phosphatase; increase of insulin receptor substrate-1, PI3Kp85, pAkt and GLUT-4 mRNA expression
	Ginsenoside-Rg1	Induction of eNOS and VEGF expression; inhibition of apoptosis
	20(S)-ginsenoside-Rg3	Inhibition of NMDA receptor-mediated nitrosative stress; stimulation of nucleic acid and energy metabolism; positive effect on gut flora
	Ginsenoside-Rh2	Increase β -endorphin secretion; up-regulation of GLUT-4 expression
<i>Rehmannia glutinosa</i> (Gaertn.)	Polysaccharides	Improvement of redox homeostasis; reduction of hepatic glucose-6-phosphatase activity; increase of hepatic glycogen level; reduction of ROS production; inhibition of NF- κ B translocation; down-regulation of TNF- α , COX-2, MCP-1 and inducible protein-10
	Catalpol	Inhibition of intracellular ROS production; suppression of NADPH-oxidase activity
	Trigonelline	Anti-oxidant activity; modulation of glucose metabolism; induction of β -cells regeneration
<i>Trigonella foenum-graecum</i> L.	Diosgenin	Increase of insulin secretion; induction of β -cells regeneration; anti-oxidant activity; promotion of adipocyte differentiation; enhancement of insulin-dependent glucose uptake
	4-hydroxyisoleucine	Stimulation of glucose-dependent insulin secretion; reduction of insulin resistance; inhibition of sucrose α -D-glucosidase and α -amylase
	Fiber	Inhibition of lipid- and carbohydrate-hydrolyzing enzymes; reduction of glucose uptake
	<i>Gymnema sylvestre</i> (Retz.) R.Br. ex Sm.	Gymnemic acids; gymnemasaponins gurmarin
<i>Cinnamomum verum</i> J. Presl. and <i>Cinnamomum cassia</i> (L.) J. Presl.	Cinnamaldehyde	Insulino tropic effect; regulation of protein-tyrosine phosphatase 1B; regulation of insulin receptor kinase; modulation of carbohydrate metabolism; inhibition of pancreatic and intestinal amylase and glucosidase; stimulation of cellular glucose uptake; increase of GLUT-4 expression; increase of PPARs expression.

This review highlights the diverse and interesting actions that are attributable to edible plants such as onion, suggesting that simple food preferences could actually help in preventing metabolic diseases.

Among all the medicinal plants reviewed, ginseng and fenugreek possess stronger clinical evidence, and their use is supported not only by WHO monographs, but also by the EMA, that has summarized scientific data on these species, their preparations and chemical constituents in officially published assessment reports [192,193]

Cinnamon anti-diabetic activity lacks of authoritative support, but many clinical trials have been conducted in the last five years, suggesting an increasing interest concerning its application in the management of diabetes.

The most common hypoglycemic mechanisms of action found for the reviewed medicinal plants and their constituents include the inhibition of α -glucosidase and of AGE formation, the increase of GLUT-4 and PPARs expression and the antioxidant activity (Table 2). Moreover, the use of herbal products often relies on the synergistic and multitarget effects of the phytocomplex, which may lead to a clinical effectiveness together with a lower incidence of adverse events [194].

The comprehensive overview of the dataset suggests that dietary natural products and phytotherapy have today an interesting role in controlling the normal to border-line glucidic levels and as an integrative therapy, but they could not be considered as alternative drugs to mono-molecular ones for type II diabetic patients. Well conducted clinical trials using modern standardized extracts are of primordial importance and it is necessary to better investigate correlations between hypoglycemic activity and chemical composition of herbal preparations, with the aim of optimizing extracts to better trigger specific pathways and, finally, to propose correct dosages to enhance safety and effectiveness. Furthermore, as many species which use in the management of diabetes is not enlisted in authoritative documents such as WHO monographs are demonstrating interesting anti-diabetic properties in vitro and in vivo, and clinical trials are being conducted to demonstrate their effectiveness in human patients, it would be of great interest to carry out comparisons between the well documented species, such as those reported in Sections 2.1–2.7, and the new candidate species, such as those reported in Section 3, under similar experimental and clinical conditions.

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References

1. Gale, E.A.M. The myth of the metabolic syndrome. *Diabetologia* **2005**, *48*, 1679–1683. [CrossRef] [PubMed]
2. Kahn, R.; Buse, J.; Ferrannini, E.; Stern, M. The metabolic syndrome: Time for a critical appraisal: Joint statement from the American Diabetes Association and the European Association for the Study of Diabetes. *Diabetes Care* **2005**, *28*, 2289–2304. [CrossRef] [PubMed]
3. Reaven, G.M. The metabolic syndrome: Requiescat in pace. *Clin. Chem.* **2005**, *51*, 931–938. [CrossRef] [PubMed]
4. Parikh, R.M.; Mohan, V. Changing definitions of metabolic syndrome. *Indian J. Endocrinol. Metab.* **2012**, *16*, 7–12. [CrossRef] [PubMed]
5. Alberti, K.G.; Zimmet, P.Z. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: Diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. *Diabet. Med.* **1998**, *15*, 539–553. [CrossRef]
6. Alberti, K.G.M.M.; Zimmet, P.; Shaw, J. The metabolic syndrome—A new worldwide definition. *Lancet* **2005**, *366*, 1059–1062. [CrossRef]
7. International Diabetes Federation. Available online: <https://www.idf.org/e-library/consensus-statements/60-idfconsensus-worldwide-definitionof-the-metabolic-syndrome> (accessed on 29 December 2017).
8. Lam, D.W.; LeRoith, D. Metabolic Syndrome. In *Endotext*; De Groot, L.J., Chrousos, G., Dungan, K., Feingold, K.R., Grossman, A., Hershman, J.M., Koch, C., Korbonits, M., McLachlan, R., New, M., et al., Eds.; MDText.com, Inc.: South Dartmouth, MA, USA, 2000.

9. Stern, M.P.; Williams, K.; Gonzalez-Villalpando, C.; Hunt, K.J.; Haffner, S.M. Does the metabolic syndrome improve identification of individuals at risk of type 2 diabetes and/or cardiovascular disease? *Diabetes Care* **2004**, *27*, 2676–2681. [[CrossRef](#)] [[PubMed](#)]
10. O'Neill, S.; O'Driscoll, L. Metabolic syndrome: A closer look at the growing epidemic and its associated pathologies. *Obes. Rev.* **2015**, *16*, 1–12. [[CrossRef](#)] [[PubMed](#)]
11. Rochlani, Y.; Pothineni, N.V.; Mehta, J.L. Metabolic Syndrome: Does it Differ between Women and Men? *Cardiovasc. Drugs Ther.* **2015**, *29*, 329–338. [[CrossRef](#)] [[PubMed](#)]
12. Kooti, W.; Farokhipour, M.; Asadzadeh, Z.; Ashtary-Larky, D.; Asadi-Samani, M. The role of medicinal plants in the treatment of diabetes: A systematic review. *Electron. Physician* **2016**, *8*, 1832–1842. [[CrossRef](#)] [[PubMed](#)]
13. Ota, A.; Ulrich, N.P. An Overview of Herbal Products and Secondary Metabolites Used for Management of Type Two Diabetes. *Front. Pharmacol.* **2017**, *8*, 1–14. [[CrossRef](#)] [[PubMed](#)]
14. Tabatabaei-malazy, O.; Ramezani, A.; Atlasi, R.; Larijani, B.; Abdollahi, M. Scientometric study of academic publications on antioxidative herbal medicines in type 2 diabetes mellitus. *J. Diabetes Metab. Disord.* **2016**, *15*, 48. [[CrossRef](#)] [[PubMed](#)]
15. Rios, J.L.; Francini, F.; Schinella, G.R. Natural products for the treatment of type 2 Diabetes mellitus. *Planta Med.* **2015**, *81*, 975–994. [[CrossRef](#)] [[PubMed](#)]
16. Yeh, G.Y.; Eisenberg, D.M.; Kaptchuk, T.J.; Phillips, R.S. Systematic review of herbs and dietary supplements for glycemic control in diabetes. *Diabetes Care* **2003**, *26*, 1277–1294. [[CrossRef](#)] [[PubMed](#)]
17. Cefalu, W.T.; Stephens, J.M.; Ribnicky, D.M. Diabetes and Herbal (Botanical) Medicine. In *Herbal Medicine: Biomolecular and Clinical Aspects*, 2nd ed.; CRC Press/Taylor & Francis: Boca Raton, FL, USA, 2011.
18. Chawla, R.; Thakur, P.; Chowdhry, A.; Jaiswal, S.; Sharma, A.; Goel, R.; Sharma, J.; Priyadarshi, S.S.; Kumar, V.; Sharma, R.K.; et al. Evidence based herbal drug standardization approach in coping with challenges of holistic management of diabetes: A dreadful lifestyle disorder of 21st century. *J. Diabetes Metab. Disord.* **2013**, *12*, 35. [[CrossRef](#)] [[PubMed](#)]
19. World Health Organization. *WHO Monographs on Selected Medicinal Plants*; World Health Organization: Geneva, Switzerland, 1999; Volume 1.
20. Farag, M.A.; Ali, S.E.; Hodaya, R.H.; El-Seedi, H.R.; Sultani, H.N.; Laub, A.; Eissa, T.F.; Abou-Zaid, F.O.F.; Wessjohann, L.A. Phytochemical profiles and antimicrobial activities of *Allium cepa* Red cv. and *A. sativum* subjected to different drying methods: A comparative MS-based metabolomics. *Molecules* **2017**, *22*, 761. [[CrossRef](#)] [[PubMed](#)]
21. Augusti, K.T. Studies on the effects of a hypoglycemic principle from *Allium Cepa* Linn. *Indian J. Med. Res.* **1973**, *61*, 1066–1071. [[PubMed](#)]
22. Gupta, R.K.; Gupta, S.; Samuel, K.C. Blood sugar lowering effect of various fractions of onion. *Indian J. Exp. Biol.* **1977**, *15*, 313–314. [[PubMed](#)]
23. Jain, R.C.; Vyas, C.R. Letter: Hypoglycaemia action of onion on rabbits. *Br. Med. J.* **1974**, *2*, 730. [[CrossRef](#)] [[PubMed](#)]
24. Ogunmodede, O.S.; Saalu, L.C.; Ogunlade, B.; Akunna, G.G.; Oyewopo, A.O. An Evaluation of the Hypoglycemic, Antioxidant and Hepatoprotective Potentials of Onion (*Allium cepa* L.) on Alloxan-induced Diabetic Rabbits. *Int. J. Pharmacol.* **2012**, *8*, 21–29.
25. El-Demerdash, F.M.; Yousef, M.I.; El-Naga, N.I.A. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. *Food Chem. Toxicol.* **2005**, *43*, 57–63. [[CrossRef](#)] [[PubMed](#)]
26. Azuma, K.; Minami, Y.; Ippoushi, K.; Terao, J. Lowering effects of onion intake on oxidative stress biomarkers in streptozotocin-induced diabetic rats. *J. Clin. Biochem. Nutr.* **2007**, *40*, 131–140. [[CrossRef](#)] [[PubMed](#)]
27. El-Soud, N.A.; Khalil, M. Antioxidative effects of *Allium Cepa* essential oil in streptozotocin induced diabetic rats. *Maced. J. Med. Sci.* **2010**, *3*, 344–351. [[CrossRef](#)]
28. Akash, M.S.H.; Rehman, K.; Chen, S. Spice plant *Allium cepa*: Dietary supplement for treatment of type 2 diabetes mellitus. *Nutrition* **2014**, *30*, 1128–1137. [[CrossRef](#)] [[PubMed](#)]
29. Taj Eldin, I.M.; Ahmed, E.M.; Elwahab, H.M.A. Preliminary Study of the Clinical Hypoglycemic Effects of *Allium cepa* (Red Onion) in Type 1 and Type 2 Diabetic Patients. *Environ. Health Insights* **2010**, *4*, 71–77. [[PubMed](#)]

30. Bang, M.-A.; Kim, H.-A.; Cho, Y.-J. Alterations in the blood glucose, serum lipids and renal oxidative stress in diabetic rats by supplementation of onion (*Allium cepa*. Linn). *Nutr. Res. Pract.* **2009**, *3*, 242–246. [[CrossRef](#)] [[PubMed](#)]
31. Ahmad, W.; Ijaz, B.; Shabbiri, K.; Ahmed, F.; Rehman, S. Oxidative toxicity in diabetes and Alzheimer's disease: Mechanisms behind ROS/RNS generation. *J. Biomed. Sci.* **2017**, *24*, 76. [[CrossRef](#)] [[PubMed](#)]
32. Campos, K.E.; Diniz, Y.S.; Cataneo, A.C.; Faine, L.A.; Alves, M.J.Q.F.; Novelli, E.L.B. Hypoglycaemic and antioxidant effects of onion, *Allium cepa*: Dietary onion addition, antioxidant activity and hypoglycaemic effects on diabetic rats. *Int. J. Food Sci. Nutr.* **2003**, *54*, 241–246. [[CrossRef](#)] [[PubMed](#)]
33. Kim, S.-H.; Jo, S.-H.; Kwon, Y.-I.; Hwang, J.-K. Effects of Onion (*Allium cepa* L.) Extract Administration on Intestinal α -Glucosidases Activities and Spikes in Postprandial Blood Glucose Levels in SD Rats Model. *Int. J. Mol. Sci.* **2011**, *12*, 3757–3769. [[CrossRef](#)] [[PubMed](#)]
34. Gautam, S.; Pal, S.; Maurya, R.; Srivastava, A.K. Ethanolic extract of *Allium cepa* stimulates glucose transporter typ 4-mediated glucose uptake by the activation of insulin signaling. *Planta Med.* **2015**, *81*, 208–214. [[CrossRef](#)] [[PubMed](#)]
35. Kumari, K.; Augusti, K.T. Antidiabetic and antioxidant effects of S-methyl cysteine sulfoxide isolated from onions (*Allium cepa* Linn) as compared to standard drugs in alloxan diabetic rats. *Indian J. Exp. Biol.* **2002**, *40*, 1005–1009. [[PubMed](#)]
36. Augusti, K.I.; Roy, V.C.; Semple, M. Effect of allyl propyl disulphide isolated from onion (*Allium cepa* L.) on glucose tolerance of alloxan diabetic rabbits. *Experientia* **1974**, *30*, 1119–1120. [[CrossRef](#)] [[PubMed](#)]
37. World Health Organization. *WHO Monographs on Selected Medicinal Plants*; World Health Organization: Geneva, Switzerland, 2007; Volume 3.
38. Alzohairy, M.A. Therapeutics Role of *Azadirachta indica* (Neem) and Their Active Constituents in Diseases Prevention and Treatment. *Evid. Based Complement. Altern. Med.* **2016**, *2016*, 7382506. [[CrossRef](#)] [[PubMed](#)]
39. Ekaidem, I.S.; Akpan, H.D.; Usuh, I.F.; Etim, O.E.; Ebong, P.E. Effects of ethanolic extract of *Azadirachta indica* leaves on lipid peroxidation and serum lipids of diabetic Wistar rats. *Acta Biol. Szeged.* **2007**, *51*, 17–20.
40. Satyanarayana, K.; Sravanthi, K.; Shaker, I.A.; Ponnulakshmi, R. Molecular approach to identify antidiabetic potential of *Azadirachta indica*. *J. Ayurveda Integr. Med.* **2015**, *6*, 165–174. [[CrossRef](#)] [[PubMed](#)]
41. Khosla, P.; Bhanwra, S.; Singh, J.; Seth, S.; Srivastava, R.K. A study of hypoglycaemic effects of *Azadirachta indica* (Neem) in normal and alloxan diabetic rabbits. *Indian J. Physiol. Pharmacol.* **2000**, *44*, 69–74. [[PubMed](#)]
42. Perez Gutierrez, R.M.; de Jesus Martinez Ortiz, M. Beneficial effect of *Azadirachta indica* on advanced glycation end-product in streptozotocin-diabetic rat. *Pharm. Biol.* **2014**, *52*, 1435–1444. [[CrossRef](#)] [[PubMed](#)]
43. Gutierrez, R.M.P.; Gomez, Y.G.Y.; Guzman, M.D. Attenuation of nonenzymatic glycation, hyperglycemia, and hyperlipidemia in streptozotocin-induced diabetic rats by chloroform leaf extract of *Azadirachta indica*. *Pharmacogn. Mag.* **2011**, *7*, 254–259. [[CrossRef](#)] [[PubMed](#)]
44. Kumar, D.B.; Mitra, A.; Manjunatha, M. Azadirachtolide: An anti-diabetic and hypolipidemic effects from *Azadirachta indica* leaves. *Pharmacogn. Commun.* **2011**, *1*, 78–84. [[CrossRef](#)]
45. Ponnusamy, S.; Haldar, S.; Mulani, F.; Zinjarde, S.; Thulasiram, H.; RaviKumar, A. Gedunin and Azadiradione: Human Pancreatic Alpha-Amylase Inhibiting Limonoids from Neem (*Azadirachta indica*) as Anti-Diabetic Agents. *PLoS ONE* **2015**, *10*, e0140113. [[CrossRef](#)] [[PubMed](#)]
46. Perez-Gutierrez, R.M.; Damian-Guzman, M. Meliacinolin: A potent alpha-glucosidase and alpha-amylase inhibitor isolated from *Azadirachta indica* leaves and in vivo antidiabetic property in streptozotocin-nicotinamide-induced type 2 diabetes in mice. *Biol. Pharm. Bull.* **2012**, *35*, 1516–1524. [[CrossRef](#)] [[PubMed](#)]
47. World Health Organization. *WHO Monographs on Selected Medicinal Plants*; World Health Organization: Geneva, Switzerland, 2009; Volume 4.
48. Yuwai, K.E.; Rao, K.S.; Kaluwin, C.; Jones, G.P.; Rivett, D.E. Chemical composition of *Momordica charantia* L. fruits. *J. Agric. Food Chem.* **1991**, *39*, 1762–1763. [[CrossRef](#)]
49. Ooi, C.P.; Yassin, Z.; Hamid, T.-A. *Momordica charantia* for type 2 diabetes mellitus. *Cochrane Database Syst. Rev.* **2012**, *15*, CD007845. [[CrossRef](#)] [[PubMed](#)]

50. Dans, A.M.L.; Villarruz, M.V.C.; Jimeno, C.A.; Javelosa, M.A.U.; Chua, J.; Bautista, R.; Velez, G.G.B. The effect of *Momordica charantia* capsule preparation on glycemic control in type 2 diabetes mellitus needs further studies. *J. Clin. Epidemiol.* **2007**, *60*, 554–559. [[CrossRef](#)] [[PubMed](#)]
51. John, A.J.; Cherian, R.; Subhash, H.S.; Cherian, A.M. Evaluation of the efficacy of bitter gourd (*Momordica charantia*) as an oral hypoglycemic agent—A randomized controlled clinical trial. *Indian J. Physiol. Pharmacol.* **2003**, *47*, 363–365. [[PubMed](#)]
52. Fuangchan, A.; Sonthisombat, P.; Seubnukarn, T.; Chanouan, R.; Chotchaisuwat, P.; Sirigulsatien, V.; Ingkaninan, K.; Plianbangchang, P.; Haines, S.T. Hypoglycemic effect of bitter melon compared with metformin in newly diagnosed type 2 diabetes patients. *J. Ethnopharmacol.* **2011**, *134*, 422–428. [[CrossRef](#)] [[PubMed](#)]
53. Grover, J.K.; Yadav, S.P. Pharmacological actions and potential uses of *Momordica charantia*: A review. *J. Ethnopharmacol.* **2004**, *93*, 123–132. [[CrossRef](#)] [[PubMed](#)]
54. Chuang, C.-Y.; Hsu, C.; Chao, C.-Y.; Wein, Y.-S.; Kuo, Y.-H.; Huang, C. Fractionation and identification of 9c, 11t, 13t-conjugated linolenic acid as an activator of PPARalpha in bitter gourd (*Momordica charantia* L.). *J. Biomed. Sci.* **2006**, *13*, 763–772. [[CrossRef](#)] [[PubMed](#)]
55. Huang, H.-L.; Hong, Y.-W.; Wong, Y.-H.; Chen, Y.-N.; Chyuan, J.-H.; Huang, C.-J.; Chao, P.-M. Bitter melon (*Momordica charantia* L.) inhibits adipocyte hypertrophy and down regulates lipogenic gene expression in adipose tissue of diet-induced obese rats. *Br. J. Nutr.* **2008**, *99*, 230–239. [[CrossRef](#)] [[PubMed](#)]
56. Shih, C.-C.; Lin, C.-H.; Lin, W.-L. Effects of *Momordica charantia* on insulin resistance and visceral obesity in mice on high-fat diet. *Diabetes Res. Clin. Pract.* **2008**, *81*, 134–143. [[CrossRef](#)] [[PubMed](#)]
57. Chang, C.-I.; Tseng, H.-I.; Liao, Y.-W.; Yen, C.-H.; Chen, T.-M.; Lin, C.-C.; Cheng, H.-L. In vivo and in vitro studies to identify the hypoglycaemic constituents of *Momordica charantia* wild variant WB24. *Food Chem.* **2011**, *125*, 521–528. [[CrossRef](#)]
58. Ma, J.; Whittaker, P.; Keller, A.C.; Mazzola, E.P.; Pawar, R.S.; White, K.D.; Callahan, J.H.; Kennelly, E.J.; Krynitsky, A.J.; Rader, J.I. Cucurbitane-type triterpenoids from *Momordica charantia*. *Planta Med.* **2010**, *76*, 1758–1761. [[CrossRef](#)] [[PubMed](#)]
59. Tan, M.-J.; Ye, J.-M.; Turner, N.; Hohnen-Behrens, C.; Ke, C.-Q.; Tang, C.-P.; Chen, T.; Weiss, H.-C.; Gesing, E.-R.; Rowland, A.; et al. Antidiabetic activities of triterpenoids isolated from bitter melon associated with activation of the AMPK pathway. *Chem. Biol.* **2008**, *15*, 263–273. [[CrossRef](#)] [[PubMed](#)]
60. Cheng, H.-L.; Huang, H.-K.; Chang, C.-I.; Tsai, C.-P.; Chou, C.-H. A cell-based screening identifies compounds from the stem of *Momordica charantia* that overcome insulin resistance and activate AMP-activated protein kinase. *J. Agric. Food Chem.* **2008**, *56*, 6835–6843. [[CrossRef](#)] [[PubMed](#)]
61. Harinantenaina, L.; Tanaka, M.; Takaoka, S.; Oda, M.; Mogami, O.; Uchida, M.; Asakawa, Y. *Momordica charantia* constituents and antidiabetic screening of the isolated major compounds. *Chem. Pharm. Bull.* **2006**, *54*, 1017–1021. [[CrossRef](#)] [[PubMed](#)]
62. Cheng, H.-L.; Kuo, C.-Y.; Liao, Y.-W.; Lin, C.-C. EMCD, a hypoglycemic triterpene isolated from *Momordica charantia* wild variant, attenuates TNF-alpha-induced inflammation in FL83B cells in an AMP-activated protein kinase-independent manner. *Eur. J. Pharmacol.* **2012**, *689*, 241–248. [[CrossRef](#)] [[PubMed](#)]
63. Sasa, M.; Inoue, I.; Shinoda, Y.; Takahashi, S.; Seo, M.; Komoda, T.; Awata, T.; Katayama, S. Activating effect of momordin, extract of bitter melon (*Momordica Charantia* L.), on the promoter of human PPARdelta. *J. Atheroscler. Thromb.* **2009**, *16*, 888–892. [[CrossRef](#)] [[PubMed](#)]
64. Efield, J.T.; Choi, Y.M.; Davies, S.W.; Mehra, S.; Anderson, E.J.; Katunga, L.A. Potential for improved glycemic control with dietary *Momordica charantia* in patients with insulin resistance and pre-diabetes. *Int. J. Environ. Res. Public Health* **2014**, *11*, 2328–2345. [[CrossRef](#)] [[PubMed](#)]
65. World Health Organization. *WHO Monographs on Selected Medicinal Plants*; World Health Organization: Geneva, Switzerland, 2002; Volume 2.
66. Pattanayak, P.; Behera, P.; Das, D.; Panda, S.K. *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacogn. Rev.* **2010**, *4*, 95–105. [[CrossRef](#)] [[PubMed](#)]
67. Agrawal, P.; Rai, V.; Singh, R.B. Randomized placebo-controlled, single blind trial of holy basil leaves in patients with noninsulin-dependent diabetes mellitus. *Int. J. Clin. Pharmacol. Ther.* **1996**, *34*, 406–409. [[PubMed](#)]

68. Satapathy, S.; Das, N.; Bandyopadhyay, D.; Mahapatra, S.C.; Sahu, D.S.; Meda, M. Effect of Tulsi (*Ocimum sanctum* Linn.) Supplementation on Metabolic Parameters and Liver Enzymes in Young Overweight and Obese Subjects. *Indian J. Clin. Biochem.* **2017**, *32*, 357–363. [CrossRef] [PubMed]
69. Kapoor, S. *Ocimum sanctum*: A therapeutic role in diabetes and the metabolic syndrome. *Horm. Metab. Res.* **2008**, *40*, 296. [CrossRef] [PubMed]
70. Hussain, E.H.M.A.; Jamil, K.; Rao, M. Hypoglycaemic, hypolipidemic and antioxidant properties of tulsi (*Ocimum sanctum* linn) on streptozotocin induced diabetes in rats. *Indian J. Clin. Biochem.* **2001**, *16*, 190–194. [CrossRef] [PubMed]
71. Reddy, S.S.; Karuna, R.; Baskar, R.; Saralakumari, D. Prevention of insulin resistance by ingesting aqueous extract of *Ocimum sanctum* to fructose-fed rats. *Horm. Metab. Res.* **2008**, *40*, 44–49. [CrossRef] [PubMed]
72. Chattopadhyay, R.R. Hypoglycemic effect of *Ocimum sanctum* leaf extract in normal and streptozotocin diabetic rats. *Indian J. Exp. Biol.* **1993**, *31*, 891–893. [PubMed]
73. Hannan, J.M.A.; Marenah, L.; Ali, L.; Rokeya, B.; Flatt, P.R.; Abdel-Wahab, Y.H.A. *Ocimum sanctum* leaf extracts stimulate insulin secretion from perfused pancreas, isolated islets and clonal pancreatic beta-cells. *J. Endocrinol.* **2006**, *189*, 127–136. [CrossRef] [PubMed]
74. Husain, I.; Chander, R.; Saxena, J.K.; Mahdi, A.A.; Mahdi, F. Antidyslipidemic Effect of *Ocimum sanctum* Leaf Extract in Streptozotocin Induced Diabetic Rats. *Indian J. Clin. Biochem.* **2015**, *30*, 72–77. [CrossRef] [PubMed]
75. Suanarunsawat, T.; Anantasomboon, G.; Piewbang, C. Anti-diabetic and anti-oxidative activity of fixed oil extracted from *Ocimum sanctum* L. leaves in diabetic rats. *Exp. Ther. Med.* **2016**, *11*, 832–840. [CrossRef] [PubMed]
76. Patil, R.; Patil, R.; Ahirwar, B.; Ahirwar, D. Isolation and characterization of anti-diabetic component (bioactivity-guided fractionation) from *Ocimum sanctum* L. (Lamiaceae) aerial part. *Asian Pac. J. Trop. Med.* **2011**, *4*, 278–282. [CrossRef]
77. Mancuso, C.; Santangelo, R. *Panax ginseng* and *Panax quinquefolius*: From pharmacology to toxicology. *Food Chem. Toxicol.* **2017**, *107*, 362–372. [CrossRef] [PubMed]
78. Park, H.J.; Kim, D.H.; Park, S.J.; Kim, J.M.; Ryu, J.H. Ginseng in Traditional Herbal Prescriptions. *J. Ginseng Res.* **2012**, *36*, 225–241. [CrossRef] [PubMed]
79. Xiang, Y.-Z.; Shang, H.-C.; Gao, X.-M.; Zhang, B.-L. A comparison of the ancient use of ginseng in traditional Chinese medicine with modern pharmacological experiments and clinical trials. *Phytother. Res.* **2008**, *22*, 851–858. [CrossRef] [PubMed]
80. European Medicine Agency Community Herbal Monograph on *Panax ginseng* C.A. Meyer, Radix. Available online: http://www.ema.europa.eu/docs/en_GB/document_library/Herbal_-_Community_herbal_monograph/2014/05/WC500167387.pdf (accessed on 9 October 2017).
81. Wang, Y.; Choi, H.-K.; Brinckmann, J.A.; Jiang, X.; Huang, L. Chemical analysis of *Panax quinquefolius* (North American ginseng): A review. *J. Chromatogr. A* **2015**, *1426*, 1–15. [CrossRef] [PubMed]
82. Kim, S.; Shin, B.-C.; Lee, M.S.; Lee, H.; Ernst, E. Red ginseng for type 2 diabetes mellitus: A systematic review of randomized controlled trials. *Chin. J. Integr. Med.* **2011**, *17*, 937–944. [CrossRef] [PubMed]
83. Shergis, J.L.; Zhang, A.L.; Zhou, W.; Xue, C.C. *Panax ginseng* in randomised controlled trials: A systematic review. *Phytother. Res.* **2013**, *27*, 949–965. [CrossRef] [PubMed]
84. Shishtar, E.; Sievenpiper, J.L.; Djedovic, V.; Cozma, A.I.; Ha, V.; Jayalath, V.H.; Jenkins, D.J.A.; Meija, S.B.; de Souza, R.J.; Jovanovski, E.; et al. The effect of ginseng (the genus panax) on glycemic control: A systematic review and meta-analysis of randomized controlled clinical trials. *PLoS ONE* **2014**, *9*, e107391. [CrossRef] [PubMed]
85. Gui, Q.-F.; Xu, Z.-R.; Xu, K.-Y.; Yang, Y.-M. The Efficacy of Ginseng-Related Therapies in Type 2 Diabetes Mellitus: An Updated Systematic Review and Meta-analysis. *Medicine* **2016**, *95*, e2584. [CrossRef] [PubMed]
86. Liu, C.; Zhang, M.; Hu, M.-Y.; Guo, H.-F.; Li, J.; Yu, Y.-L.; Jin, S.; Wang, X.-T.; Liu, L.; Liu, X.-D. Increased glucagon-like peptide-1 secretion may be involved in antidiabetic effects of ginsenosides. *J. Endocrinol.* **2013**, *217*, 185–196. [CrossRef] [PubMed]
87. Niu, J.; Pi, Z.; Yue, H.; Wang, Y.; Yu, Q.; Liu, S. Effect of ginseng polysaccharide on the urinary excretion of type 2 diabetic rats studied by liquid chromatography-mass spectrometry. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **2012**, *907*, 7–12. [CrossRef] [PubMed]

88. Deng, J.; Liu, Y.; Duan, Z.; Zhu, C.; Hui, J.; Mi, Y.; Ma, P.; Ma, X.; Fan, D.; Yang, H. Protopanaxadiol and Protopanaxatriol-Type Saponins Ameliorate Glucose and Lipid Metabolism in Type 2 Diabetes Mellitus in High-Fat Diet/Streptozotocin-Induced Mice. *Front. Pharmacol.* **2017**, *8*, 506. [[CrossRef](#)] [[PubMed](#)]
89. Jiang, S.; Ren, D.; Li, J.; Yuan, G.; Li, H.; Xu, G.; Han, X.; Du, P.; An, L. Effects of compound K on hyperglycemia and insulin resistance in rats with type 2 diabetes mellitus. *Fitoterapia* **2014**, *95*, 58–64. [[CrossRef](#)] [[PubMed](#)]
90. Yang, N.; Chen, P.; Tao, Z.; Zhou, N.; Gong, X.; Xu, Z.; Zhang, M.; Zhang, D.; Chen, B.; Tao, Z.; et al. Beneficial effects of ginsenoside-Rg1 on ischemia-induced angiogenesis in diabetic mice. *Acta Biochim. Biophys. Sin.* **2012**, *44*, 999–1005. [[CrossRef](#)] [[PubMed](#)]
91. Kang, K.S.; Yamabe, N.; Kim, H.Y.; Park, J.H.; Yokozawa, T. Therapeutic potential of 20(S)-ginsenoside Rg(3) against streptozotocin-induced diabetic renal damage in rats. *Eur. J. Pharmacol.* **2008**, *591*, 266–272. [[CrossRef](#)] [[PubMed](#)]
92. Niu, J.; Pi, Z.-F.; Yue, H.; Yang, H.; Wang, Y.; Yu, Q.; Liu, S.-Y. Effect of 20(S)-ginsenoside Rg3 on streptozotocin-induced experimental type 2 diabetic rats: A urinary metabolomics study by rapid-resolution liquid chromatography/mass spectrometry. *Rapid Commun. Mass Spectrom.* **2012**, *26*, 2683–2689. [[CrossRef](#)] [[PubMed](#)]
93. Lai, D.-M.; Tu, Y.-K.; Liu, I.-M.; Chen, P.-F.; Cheng, J.-T. Mediation of beta-endorphin by ginsenoside Rh2 to lower plasma glucose in streptozotocin-induced diabetic rats. *Planta Med.* **2006**, *72*, 9–13. [[CrossRef](#)] [[PubMed](#)]
94. Kim, H.Y.; Kang, K.S.; Yamabe, N.; Yokozawa, T. Comparison of the effects of Korean ginseng and heat-processed Korean ginseng on diabetic oxidative stress. *Am. J. Chin. Med.* **2008**, *36*, 989–1004. [[CrossRef](#)] [[PubMed](#)]
95. Seo, Y.-S.; Shon, M.-Y.; Kong, R.; Kang, O.-H.; Zhou, T.; Kim, D.-Y.; Kwon, D.-Y. Black ginseng extract exerts anti-hyperglycemic effect via modulation of glucose metabolism in liver and muscle. *J. Ethnopharmacol.* **2016**, *190*, 231–240. [[CrossRef](#)] [[PubMed](#)]
96. Kim, J.H.; Pan, J.H.; Cho, H.T.; Kim, Y.J. Black Ginseng Extract Counteracts Streptozotocin-Induced Diabetes in Mice. *PLoS ONE* **2016**, *11*, e0146843. [[CrossRef](#)] [[PubMed](#)]
97. Kim, C.-S.; Jo, K.; Pyo, M.-K.; Kim, J.S.; Kim, J. Pectin lyase-modified red ginseng extract exhibits potent anti-glycation effects in vitro and in vivo. *J. Exerc. Nutr. Biochem.* **2017**, *21*, 56–62. [[CrossRef](#)] [[PubMed](#)]
98. Murthy, H.N.; Dandin, V.S.; Lee, E.J.; Paek, K.Y. Efficacy of ginseng adventitious root extract on hyperglycemia in streptozotocin-induced diabetic rats. *J. Ethnopharmacol.* **2014**, *153*, 917–921. [[CrossRef](#)] [[PubMed](#)]
99. Liu, C.; Ma, R.; Wang, L.; Zhu, R.; Liu, H.; Guo, Y.; Zhao, B.; Zhao, S.; Tang, J.; Li, Y.; et al. *Rehmanniae Radix* in osteoporosis: A review of traditional Chinese medicinal uses, phytochemistry, pharmacokinetics and pharmacology. *J. Ethnopharmacol.* **2017**, *198*, 351–362. [[CrossRef](#)] [[PubMed](#)]
100. Matsumoto, Y.; Sekimizu, K. A hyperglycemic silkworm model for evaluating hypoglycemic activity of *Rehmanniae Radix*, an herbal medicine. *Drug Discov. Ther.* **2016**, *10*, 14–18. [[CrossRef](#)] [[PubMed](#)]
101. Ursini, F.; Maiorino, M.; Forman, H.J. Redox homeostasis: The Golden Mean of healthy living. *Redox Biol.* **2016**, *8*, 205–215. [[CrossRef](#)] [[PubMed](#)]
102. Baek, G.-H.; Jang, Y.-S.; Jeong, S.-I.; Cha, J.; Joo, M.; Shin, S.-W.; Ha, K.-T.; Jeong, H.-S. *Rehmannia glutinosa* suppresses inflammatory responses elicited by advanced glycation end products. *Inflammation* **2012**, *35*, 1232–1241. [[CrossRef](#)] [[PubMed](#)]
103. Zhang, R.; Zhou, J.; Jia, Z.; Zhang, Y.; Gu, G. Hypoglycemic effect of *Rehmannia glutinosa* oligosaccharide in hyperglycemic and alloxan-induced diabetic rats and its mechanism. *J. Ethnopharmacol.* **2004**, *90*, 39–43. [[CrossRef](#)] [[PubMed](#)]
104. Zhou, J.; Xu, G.; Yan, J.; Li, K.; Bai, Z.; Cheng, W.; Huang, K. *Rehmannia glutinosa* (Gaertn.) DC. polysaccharide ameliorates hyperglycemia, hyperlipemia and vascular inflammation in streptozotocin-induced diabetic mice. *J. Ethnopharmacol.* **2015**, *164*, 229–238. [[CrossRef](#)] [[PubMed](#)]
105. Zhang, R.X.; Jia, Z.P.; Kong, L.Y.; Ma, H.P.; Ren, J.; Li, M.X.; Ge, X. Stachyose extract from *Rehmannia glutinosa* Libosch. to lower plasma glucose in normal and diabetic rats by oral administration. *Pharmazie* **2004**, *59*, 552–556. [[PubMed](#)]

106. Choi, H.-J.; Jang, H.-J.; Chung, T.-W.; Jeong, S.-I.; Cha, J.; Choi, J.-Y.; Han, C.W.; Jang, Y.-S.; Joo, M.; Jeong, H.-S.; et al. Catalpol suppresses advanced glycation end-products-induced inflammatory responses through inhibition of reactive oxygen species in human monocytic THP-1 cells. *Fitoterapia* **2013**, *86*, 19–28. [[CrossRef](#)] [[PubMed](#)]
107. Zhu, H.; Wang, Y.; Liu, Z.; Wang, J.; Wan, D.; Feng, S.; Yang, X.; Wang, T. Antidiabetic and antioxidant effects of catalpol extracted from *Rehmannia glutinosa* (Di Huang) on rat diabetes induced by streptozotocin and high-fat, high-sugar feed. *Chin. Med.* **2016**, *11*, 25. [[CrossRef](#)] [[PubMed](#)]
108. Yang, S.; Deng, H.; Zhang, Q.; Xie, J.; Zeng, H.; Jin, X.; Ling, Z.; Shan, Q.; Liu, M.; Ma, Y.; et al. Amelioration of Diabetic Mouse Nephropathy by Catalpol Correlates with Down-Regulation of Grb10 Expression and Activation of Insulin-Like Growth Factor 1 / Insulin-Like Growth Factor 1 Receptor Signaling. *PLoS ONE* **2016**, *11*, e0151857. [[CrossRef](#)] [[PubMed](#)]
109. Zhou, J.; Xu, G.; Ma, S.; Li, F.; Yuan, M.; Xu, H.; Huang, K. Catalpol ameliorates high-fat diet-induced insulin resistance and adipose tissue inflammation by suppressing the JNK and NF-kappaB pathways. *Biochem. Biophys. Res. Commun.* **2015**, *467*, 853–858. [[CrossRef](#)] [[PubMed](#)]
110. Dong, Z.; Chen, C.X. Effect of catalpol on diabetic nephropathy in rats. *Phytomedicine* **2013**, *20*, 1023–1029. [[CrossRef](#)] [[PubMed](#)]
111. Bahmani, M.; Shirzad, H.; Mirhosseini, M.; Mesripour, A.; Rafieian-Kopaei, M. A Review on Ethnobotanical and Therapeutic Uses of Fenugreek (*Trigonella foenum-graecum* L.). *J. Evid.-Based. Complement. Altern. Med.* **2016**, *21*, 53–62. [[CrossRef](#)] [[PubMed](#)]
112. Neelakantan, N.; Narayanan, M.; de Souza, R.J.; van Dam, R.M. Effect of fenugreek (*Trigonella foenum-graecum* L.) intake on glycemia: A meta-analysis of clinical trials. *Nutr. J.* **2014**, *13*, 7. [[CrossRef](#)] [[PubMed](#)]
113. Suksomboon, N.; Poolsup, N.; Boonkaew, S.; Suthisang, C.C. Meta-analysis of the effect of herbal supplement on glycemic control in type 2 diabetes. *J. Ethnopharmacol.* **2011**, *137*, 1328–1333. [[CrossRef](#)] [[PubMed](#)]
114. Shojaii, A.; Dabaghian, F.H.; Goushegir, A.; Fard, M.A. Antidiabetic plants of Iran. *Acta Med. Iran.* **2011**, *49*, 637–642. [[PubMed](#)]
115. Nahas, R.; Moher, M. Complementary and alternative medicine for the treatment of type 2 diabetes. *Can. Fam. Physician* **2009**, *55*, 591–596. [[PubMed](#)]
116. Raju, J.; Gupta, D.; Rao, A.R.; Yadava, P.K.; Baquer, N.Z. Trigonellafoenum graecum (fenugreek) seed powder improves glucose homeostasis in alloxan diabetic rat tissues by reversing the altered glycolytic, gluconeogenic and lipogenic enzymes. *Mol. Cell. Biochem.* **2001**, *224*, 45–51. [[CrossRef](#)] [[PubMed](#)]
117. Ravikumar, P.; Anuradha, C.V. Effect of fenugreek seeds on blood lipid peroxidation and antioxidants in diabetic rats. *Phytother. Res.* **1999**, *13*, 197–201. [[CrossRef](#)]
118. Vijayakumar, M.V.; Singh, S.; Chhipa, R.R.; Bhat, M.K. The hypoglycaemic activity of fenugreek seed extract is mediated through the stimulation of an insulin signalling pathway. *Br. J. Pharmacol.* **2005**, *146*, 41–48. [[CrossRef](#)] [[PubMed](#)]
119. Koupy, D.; Kotolova, H.; Ruda Kucerova, J. Effectiveness of phytotherapy in supportive treatment of type 2 diabetes mellitus II. Fenugreek (*Trigonella foenum-graecum*). *Ceska Slov. Farm.* **2015**, *64*, 67–71. [[PubMed](#)]
120. Zhou, J.; Chan, L.; Zhou, S. Trigonelline: A Plant Alkaloid with Therapeutic Potential for Diabetes and Central Nervous System Disease. *Curr. Med. Chem.* **2012**, *19*, 3523–3531.
121. Kalailingam, P.; Kannaian, B.; Tamilmani, E.; Kaliaperumal, R. Efficacy of natural diosgenin on cardiovascular risk, insulin secretion, and beta cells in streptozotocin (STZ)-induced diabetic rats. *Phytomedicine* **2014**, *21*, 1154–1161. [[CrossRef](#)] [[PubMed](#)]
122. Uemura, T.; Hirai, S.; Mizoguchi, N.; Goto, T.; Lee, J.-Y.; Taketani, K.; Nakano, Y.; Shono, J.; Hoshino, S.; Tsuge, N.; et al. Diosgenin present in fenugreek improves glucose metabolism by promoting adipocyte differentiation and inhibiting inflammation in adipose tissues. *Mol. Nutr. Food Res.* **2010**, *54*, 1596–1608. [[CrossRef](#)] [[PubMed](#)]
123. Son, I.S.; Kim, J.H.; Sohn, H.Y.; Son, K.H.; Kim, J.-S.; Kwon, C.-S. Antioxidative and hypolipidemic effects of diosgenin, a steroidal saponin of yam (*Dioscorea* spp.), on high-cholesterol fed rats. *Biosci. Biotechnol. Biochem.* **2007**, *71*, 3063–3071. [[CrossRef](#)] [[PubMed](#)]
124. Zafar, M.I.; Gao, F. 4-Hydroxyisoleucine: A Potential New Treatment for Type 2 Diabetes Mellitus. *BioDrugs* **2016**, *30*, 255–262. [[CrossRef](#)] [[PubMed](#)]

125. Jette, L.; Harvey, L.; Eugeni, K.; Levens, N. 4-Hydroxyisoleucine: A plant-derived treatment for metabolic syndrome. *Curr. Opin. Investig. Drugs* **2009**, *10*, 353–358. [PubMed]
126. Broca, C.; Manteghetti, M.; Gross, R.; Baissac, Y.; Jacob, M.; Petit, P.; Sauvaire, Y.; Ribes, G. 4-Hydroxyisoleucine: Effects of synthetic and natural analogues on insulin secretion. *Eur. J. Pharmacol.* **2000**, *390*, 339–345. [CrossRef]
127. Sauvaire, Y.; Petit, P.; Broca, C.; Manteghetti, M.; Baissac, Y.; Fernandez-Alvarez, J.; Gross, R.; Roye, M.; Leconte, A.; Gomis, R.; et al. 4-Hydroxyisoleucine: A novel amino acid potentiator of insulin secretion. *Diabetes* **1998**, *47*, 206–210. [CrossRef] [PubMed]
128. Hannan, J.M.A.; Ali, L.; Rokeya, B.; Khaleque, J.; Akhter, M.; Flatt, P.R.; Abdel-Wahab, Y.H.A. Soluble dietary fibre fraction of *Trigonella foenum-graecum* (fenugreek) seed improves glucose homeostasis in animal models of type 1 and type 2 diabetes by delaying carbohydrate digestion and absorption, and enhancing insulin action. *Br. J. Nutr.* **2007**, *97*, 514–521. [CrossRef] [PubMed]
129. Srichamroen, A.; Field, C.J.; Thomson, A.B.R.; Basu, T.K. The Modifying Effects of Galactomannan from Canadian-Grown Fenugreek (*Trigonella foenum-graecum* L.) on the Glycemic and Lipidemic Status in Rats. *J. Clin. Biochem. Nutr.* **2008**, *43*, 167–174. [CrossRef] [PubMed]
130. Hannan, J.M.A.; Rokeya, B.; Faruque, O.; Nahar, N.; Mosihuzzaman, M.; Azad Khan, A.K.; Ali, L. Effect of soluble dietary fibre fraction of *Trigonella foenum graecum* on glycemic, insulinemic, lipidemic and platelet aggregation status of Type 2 diabetic model rats. *J. Ethnopharmacol.* **2003**, *88*, 73–77. [CrossRef]
131. Sathasivampillai, S.V.; Rajamanoharan, P.R.S.; Munday, M.; Heinrich, M. Plants used to treat diabetes in Sri Lankan Siddha Medicine—An ethnopharmacological review of historical and modern sources. *J. Ethnopharmacol.* **2017**, *198*, 531–599. [CrossRef] [PubMed]
132. Afifi-Yazar, F.U.; Kasabri, V.; Abu-Dahab, R. Medicinal plants from Jordan in the treatment of diabetes: Traditional uses vs. in vitro and in vivo evaluations—Part 2. *Planta Med.* **2011**, *77*, 1210–1220. [CrossRef] [PubMed]
133. Ezuruike, U.F.; Prieto, J.M. The use of plants in the traditional management of diabetes in Nigeria: Pharmacological and toxicological considerations. *J. Ethnopharmacol.* **2014**, *155*, 857–924. [CrossRef] [PubMed]
134. Yin, J.; Zhang, H.; Ye, J. Traditional chinese medicine in treatment of metabolic syndrome. *Endocr. Metab. Immune Disord. Drug Targets* **2008**, *8*, 99–111. [CrossRef] [PubMed]
135. Patel, D.K.; Prasad, S.K.; Kumar, R.; Hemalatha, S. An overview on antidiabetic medicinal plants having insulin mimetic property. *Asian Pac. J. Trop. Biomed.* **2012**, *2*, 320–330. [CrossRef]
136. Farzaei, F.; Morovati, M.R.; Farjadmand, F.; Farzaei, M.H. A Mechanistic Review on Medicinal Plants Used for Diabetes Mellitus in Traditional Persian Medicine. *J. Evid.-Based Complement. Altern. Med.* **2017**, in press. [CrossRef] [PubMed]
137. Pubmed. Available online: <http://www.ncbi.nlm.nih.gov/pubmed> (accessed on 29 December 2017).
138. Vasant More, S.; Kim, I.-S.; Choi, D.-K. Recent Update on the Role of Chinese Material Medica and Formulations in Diabetic Retinopathy. *Molecules* **2017**, *22*, 76. [CrossRef] [PubMed]
139. NIH ClinicalTrials.gov. Available online: <https://clinicaltrials.gov/ct2/show/NCT02370121> (accessed on 22 November 2017).
140. Pothuraju, R.; Sharma, R.K.; Chagalamarri, J.; Jangra, S.; Kumar Kavadi, P. A systematic review of *Gymnema sylvestre* in obesity and diabetes management. *J. Sci. Food Agric.* **2014**, *94*, 834–840. [CrossRef] [PubMed]
141. Tiwari, P.; Mishra, B.N.; Sangwan, N.S. Phytochemical and Pharmacological Properties of *Gymnema sylvestre*: An Important Medicinal Plant. *Biomed. Res. Int.* **2014**, *2014*, 830285. [CrossRef] [PubMed]
142. Daisy, P.; Eliza, J.; Mohamed Farook, K.A.M. A novel dihydroxy gymnemic triacetate isolated from *Gymnema sylvestre* possessing normoglycemic and hypolipidemic activity on STZ-induced diabetic rats. *J. Ethnopharmacol.* **2009**, *126*, 339–344. [CrossRef] [PubMed]
143. Al-Romaiyan, A.; Liu, B.; Asare-Anane, H.; Maity, C.R.; Chatterjee, S.K.; Koley, N.; Biswas, T.; Chatterji, A.K.; Huang, G.-C.; Amiel, S.A.; et al. A novel *Gymnema sylvestre* extract stimulates insulin secretion from human islets in vivo and in vitro. *Phytother. Res.* **2010**, *24*, 1370–1376. [CrossRef] [PubMed]
144. Kumar, S.N.; Mani, U.V.; Mani, I. An open label study on the supplementation of *Gymnema sylvestre* in type 2 diabetics. *J. Diet. Suppl.* **2010**, *7*, 273–282. [CrossRef] [PubMed]

145. Martínez-Abundis, E.; Méndez-del Villar, M.; Pérez-Rubio, K.G.; Zuñiga, L.Y.; Cortez-Navarrete, M.; Ramírez-Rodríguez, A.; González-Ortiz, M. Novel nutraceutical therapies for the treatment of metabolic syndrome. *World J. Diabetes* **2016**, *7*, 142–152. [[CrossRef](#)] [[PubMed](#)]
146. Demmers, A.; Korthout, H.; van Etten-Jamaludin, F.S.; Kortekaas, F.; Maaskant, J.M. Effects of medicinal food plants on impaired glucose tolerance: A systematic review of randomized controlled trials. *Diabetes Res. Clin. Pract.* **2017**, *131*, 91–106. [[CrossRef](#)] [[PubMed](#)]
147. Gryn-Rynko, A.; Bazylak, G.; Olszewska-Slonina, D. New potential phytotherapeutics obtained from white mulberry (*Morus alba* L.) leaves. *Biomed. Pharmacother.* **2016**, *84*, 628–636. [[CrossRef](#)] [[PubMed](#)]
148. Kim, J.Y.; Ok, H.M.; Kim, J.; Park, S.W.; Kwon, S.W.; Kwon, O. Mulberry leaf extract improves postprandial glucose response in prediabetic subjects: A randomized, double-blind placebo-controlled trial. *J. Med. Food* **2015**, *18*, 306–313. [[CrossRef](#)] [[PubMed](#)]
149. Jovanovski, E.; Li, D.; Thanh Ho, H.V.; Djedovic, V.; de Ruiz Marques, A.C.; Shishtar, E.; Mejia, S.B.; Sievenpiper, J.L.; de Souza, R.J.; Duvnjak, L.; et al. The effect of alpha-linolenic acid on glycemic control in individuals with type 2 diabetes: A systematic review and meta-analysis of randomized controlled clinical trials. *Medicine* **2017**, *96*, e6531. [[CrossRef](#)] [[PubMed](#)]
150. Singh, K.K.; Mridula, D.; Rehal, J.; Barnwal, P. Flaxseed: A potential source of food, feed and fiber. *Crit. Rev. Food Sci. Nutr.* **2011**, *51*, 210–222. [[CrossRef](#)] [[PubMed](#)]
151. Akrami, A.; Nikaein, F.; Babajafari, S.; Faghih, S.; Yarmohammadi, H. Comparison of the effects of flaxseed oil and sunflower seed oil consumption on serum glucose, lipid profile, blood pressure, and lipid peroxidation in patients with metabolic syndrome. *J. Clin. Lipidol.* **2017**, in press. [[CrossRef](#)] [[PubMed](#)]
152. Hashemzadeh, A.A.; Nasoohi, N.; Raygan, F.; Aghadavod, E.; Akbari, E.; Taghizadeh, M.; Memarzadeh, M.R.; Asemi, Z. Flaxseed Oil Supplementation Improve Gene Expression Levels of PPAR-gamma, LP(a), IL-1 and TNF-alpha in Type 2 Diabetic Patients with Coronary Heart Disease. *Lipids* **2017**, *52*, 907–915. [[CrossRef](#)] [[PubMed](#)]
153. Mani, U.V.; Mani, I.; Biswas, M.; Kumar, S.N. An open-label study on the effect of flax seed powder (*Linum usitatissimum*) supplementation in the management of diabetes mellitus. *J. Diet. Suppl.* **2011**, *8*, 257–265. [[CrossRef](#)] [[PubMed](#)]
154. Thakur, G.; Mitra, A.; Pal, K.; Rousseau, D. Effect of flaxseed gum on reduction of blood glucose and cholesterol in type 2 diabetic patients. *Int. J. Food Sci. Nutr.* **2009**, *60*, 126–136. [[CrossRef](#)] [[PubMed](#)]
155. Hidalgo, J.; Flores, C.; Hidalgo, M.A.; Perez, M.; Yanez, A.; Quinones, L.; Caceres, D.D.; Burgos, R.A. Delphinol(R) standardized maqui berry extract reduces postprandial blood glucose increase in individuals with impaired glucose regulation by novel mechanism of sodium glucose cotransporter inhibition. *Panminerva Med.* **2014**, *56*, 1–7. [[PubMed](#)]
156. Alvarado, J.; Schoenlau, F.; Leschot, A.; Salgad, A.M.; Vigil Portales, P. Delphinol(R) standardized maqui berry extract significantly lowers blood glucose and improves blood lipid profile in prediabetic individuals in three-month clinical trial. *Panminerva Med.* **2016**, *58*, 1–6. [[PubMed](#)]
157. Paula, P.C.; Oliveira, J.T.A.; Sousa, D.O.B.; Alves, B.G.T.; Carvalho, A.F.U.; Franco, O.L.; Vasconcelos, I.M. Insulin-like plant proteins as potential innovative drugs to treat diabetes—The *Moringa oleifera* case study. *New Biotechnol.* **2017**, *39*, 99–109. [[CrossRef](#)] [[PubMed](#)]
158. Paula, P.C.; Sousa, D.O.B.; Oliveira, J.T.A.; Carvalho, A.F.U.; Alves, B.G.T.; Pereira, M.L.; Farias, D.F.; Viana, M.P.; Santos, F.A.; Morais, T.C.; et al. A Protein Isolate from *Moringa oleifera* Leaves Has Hypoglycemic and Antioxidant Effects in Alloxan-Induced Diabetic Mice. *Molecules* **2017**, *22*, 271. [[CrossRef](#)] [[PubMed](#)]
159. Omodanisi, E.I.; Aboua, Y.G.; Oguntibeju, O.O. Assessment of the Anti-Hyperglycaemic, Anti-Inflammatory and Antioxidant Activities of the Methanol Extract of *Moringa Oleifera* in Diabetes-Induced Nephrotoxic Male Wistar Rats. *Molecules* **2017**, *22*, 439. [[CrossRef](#)] [[PubMed](#)]
160. Anthanont, P.; Lumlerdkij, N.; Akarasreenont, P.; Vannasaeng, S.; Sriwijitkamol, A. *Moringa Oleifera* Leaf Increases Insulin Secretion after Single Dose Administration: A Preliminary Study in Healthy Subjects. *J. Med. Assoc. Thail.* **2016**, *99*, 308–313.
161. Akash, M.S.H.; Rehman, K.; Tariq, M.; Chen, S. *Zingiber officinale* and Type 2 Diabetes Mellitus: Evidence from Experimental Studies. *Crit. Rev. Eukaryot. Gene Expr.* **2015**, *25*, 91–112. [[CrossRef](#)] [[PubMed](#)]

162. Shidfar, F.; Rajab, A.; Rahideh, T.; Khandouzi, N.; Hosseini, S.; Shidfar, S. The effect of ginger (*Zingiber officinale*) on glycemic markers in patients with type 2 diabetes. *J. Complement. Integr. Med.* **2015**, *12*, 165–170. [[CrossRef](#)] [[PubMed](#)]
163. Nerurkar, P.V.; Hwang, P.W.; Saksa, E. Anti-Diabetic Potential of Noni: The Yin and the Yang. *Molecules* **2015**, *20*, 17684–17719. [[CrossRef](#)] [[PubMed](#)]
164. Van Dieren, S.; Uiterwaal, C.S.P.M.; van der Schouw, Y.T.; van der A, D.L.; Boer, J.M.A.; Spijkerman, A.; Grobbee, D.E.; Beulens, J.W.J. Coffee and tea consumption and risk of type 2 diabetes. *Diabetologia* **2009**, *52*, 2561–2569. [[CrossRef](#)] [[PubMed](#)]
165. Akash, M.S.H.; Rehman, K.; Chen, S. Effects of coffee on type 2 diabetes mellitus. *Nutrition* **2014**, *30*, 755–763. [[CrossRef](#)] [[PubMed](#)]
166. Jokura, H.; Watanabe, I.; Umeda, M.; Hase, T.; Shimotoyodome, A. Coffee polyphenol consumption improves postprandial hyperglycemia associated with impaired vascular endothelial function in healthy male adults. *Nutr. Res.* **2015**, *35*, 873–881. [[CrossRef](#)] [[PubMed](#)]
167. Mahmoud, F.; Haines, D.; Al-Ozairi, E.; Dashti, A. Effect of Black Tea Consumption on Intracellular Cytokines, Regulatory T Cells and Metabolic Biomarkers in Type 2 Diabetes Patients. *Phytother. Res.* **2016**, *30*, 454–462. [[CrossRef](#)] [[PubMed](#)]
168. Park, J.-H.; Bae, J.-H.; Im, S.-S.; Song, D.-K. Green tea and type 2 diabetes. *Integr. Med. Res.* **2014**, *3*, 4–10. [[CrossRef](#)] [[PubMed](#)]
169. Archer, A.W. Determination of cinnamaldehyde, coumarin and cinnamyl alcohol in cinnamon and cassia by high-performance liquid chromatography. *J. Chromatogr. A* **1988**, *447*, 272–276. [[CrossRef](#)]
170. Ranasinghe, P.; Piger, S.; Premakumara, G.A.S.; Galappaththy, P.; Constantine, G.R.; Katulanda, P. Medicinal properties of “true” cinnamon (*Cinnamomum zeylanicum*): A systematic review. *BMC Complement. Altern. Med.* **2013**, *13*, 275. [[CrossRef](#)] [[PubMed](#)]
171. Costello, R.B.; Dwyer, J.T.; Saldanha, L.; Bailey, R.L.; Merkel, J.; Wambogo, E. Do Cinnamon Supplements Have a Role in Glycemic Control in Type 2 Diabetes? A Narrative Review. *J. Acad. Nutr. Diet.* **2016**, *116*, 1794–1802. [[CrossRef](#)] [[PubMed](#)]
172. Allen, R.W.; Schwartzman, E.; Baker, W.L.; Coleman, C.I.; Phung, O.J. Cinnamon use in type 2 diabetes: An updated systematic review and meta-analysis. *Ann. Fam. Med.* **2013**, *11*, 452–459. [[CrossRef](#)] [[PubMed](#)]
173. Akilen, R.; Tsiami, A.; Devendra, D.; Robinson, N. Cinnamon in glycaemic control: Systematic review and meta analysis. *Clin. Nutr.* **2012**, *31*, 609–615. [[CrossRef](#)] [[PubMed](#)]
174. Bandara, T.; Uluwaduge, I.; Jansz, E.R. Bioactivity of cinnamon with special emphasis on diabetes mellitus: A review. *Int. J. Food Sci. Nutr.* **2012**, *63*, 380–386. [[CrossRef](#)] [[PubMed](#)]
175. Leach, M.J.; Kumar, S. Cinnamon for diabetes mellitus. *Cochrane Database Syst. Rev.* **2012**, *12*, CD007170.
176. Ranasinghe, P.; Jayawardana, R.; Galappaththy, P.; Constantine, G.R.; de Vas Gunawardana, N.; Katulanda, P. Efficacy and safety of “true” cinnamon (*Cinnamomum zeylanicum*) as a pharmaceutical agent in diabetes: A systematic review and meta-analysis. *Diabet. Med.* **2012**, *29*, 1480–1492. [[CrossRef](#)] [[PubMed](#)]
177. Qin, B.; Dawson, H.D.; Schoene, N.W.; Polansky, M.M.; Anderson, R.A. Cinnamon polyphenols regulate multiple metabolic pathways involved in insulin signaling and intestinal lipoprotein metabolism of small intestinal enterocytes. *Nutrition* **2012**, *28*, 1172–1179. [[CrossRef](#)] [[PubMed](#)]
178. Adisakwattana, S.; Lerdsuwankij, O.; Poputtachai, U.; Minipun, A.; Suparpprom, C. Inhibitory activity of cinnamon bark species and their combination effect with acarbose against intestinal alpha-glucosidase and pancreatic alpha-amylase. *Plant Foods Hum. Nutr.* **2011**, *66*, 143–148. [[CrossRef](#)] [[PubMed](#)]
179. Couturier, K.; Batandier, C.; Awada, M.; Hininger-Favier, I.; Canini, F.; Anderson, R.A.; Leverve, X.; Roussel, A.M. Cinnamon improves insulin sensitivity and alters the body composition in an animal model of the metabolic syndrome. *Arch. Biochem. Biophys.* **2010**, *501*, 158–161. [[CrossRef](#)] [[PubMed](#)]
180. Anand, P.; Murali, K.Y.; Tandon, V.; Murthy, P.S.; Chandra, R. Insulinotropic effect of cinnamaldehyde on transcriptional regulation of pyruvate kinase, phosphoenolpyruvate carboxykinase, and GLUT4 translocation in experimental diabetic rats. *Chem. Biol. Interact.* **2010**, *186*, 72–81. [[CrossRef](#)] [[PubMed](#)]
181. Ping, H.; Zhang, G.; Ren, G. Antidiabetic effects of cinnamon oil in diabetic KK-Ay mice. *Food Chem. Toxicol.* **2010**, *48*, 2344–2349. [[CrossRef](#)] [[PubMed](#)]
182. Subash Babu, P.; Prabuseenivasan, S.; Ignacimuthu, S. Cinnamaldehyde—A potential antidiabetic agent. *Phytomedicine* **2007**, *14*, 15–22. [[CrossRef](#)] [[PubMed](#)]

183. Kim, S.H.; Hyun, S.H.; Choung, S.Y. Anti-diabetic effect of cinnamon extract on blood glucose in db/db mice. *J. Ethnopharmacol.* **2006**, *104*, 119–123. [CrossRef] [PubMed]
184. Roffey, B.; Atwal, A.; Kubow, S. Cinnamon water extracts increase glucose uptake but inhibit adiponectin secretion in 3T3-L1 adipose cells. *Mol. Nutr. Food Res.* **2006**, *50*, 739–745. [CrossRef] [PubMed]
185. Qin, B.; Nagasaki, M.; Ren, M.; Bajotto, G.; Oshida, Y.; Sato, Y. Cinnamon extract prevents the insulin resistance induced by a high-fructose diet. *Horm. Metab. Res.* **2004**, *36*, 119–125. [PubMed]
186. Cao, H.; Polansky, M.M.; Anderson, R.A. Cinnamon extract and polyphenols affect the expression of tristetraprolin, insulin receptor, and glucose transporter 4 in mouse 3T3-L1 adipocytes. *Arch. Biochem. Biophys.* **2007**, *459*, 214–222. [CrossRef] [PubMed]
187. Ulbricht, C.; Seamon, E.; Windsor, R.C.; Armbruester, N.; Bryan, J.K.; Costa, D.; Giese, N.; Gruenwald, J.; Iovin, R.; Isaac, R.; et al. An evidence-based systematic review of cinnamon (*Cinnamomum* spp.) by the Natural Standard Research Collaboration. *J. Diet. Suppl.* **2011**, *8*, 378–454. [CrossRef] [PubMed]
188. Corns, C.M. Herbal remedies and clinical biochemistry. *Ann. Clin. Biochem.* **2003**, *40*, 489–507. [CrossRef] [PubMed]
189. Sproll, C.; Ruge, W.; Andlauer, C.; Godelmann, R.; Lachenmeier, D.W. HPLC analysis and safety assessment of coumarin in foods. *Food Chem.* **2008**, *109*, 462–469. [CrossRef] [PubMed]
190. Abraham, K.; Wohrlin, F.; Lindtner, O.; Heinemeyer, G.; Lampen, A. Toxicology and risk assessment of coumarin: Focus on human data. *Mol. Nutr. Food Res.* **2010**, *54*, 228–239. [CrossRef] [PubMed]
191. European Food Safety Authority. Coumarin in flavourings and other food ingredients with flavouring properties—Scientific Opinion of the Panel on Food Additives, Flavourings, Processing Aids and Materials in Contact with Food (AFC). *EFSA J.* **2008**, *6*, 1–15.
192. EMA Assessment Report on *Panax ginseng*, C.A. Meyer, Radix. Available online: http://www.ema.europa.eu/docs/en_GB/document_library/Herbal_-_HMPC_assessment_report/2014/05/WC500167385.pdf (accessed on 29 December 2017).
193. EMA Assessment Report on *Trigonella foenum-graecum* L.; Semen. Available online: http://www.ema.europa.eu/docs/en_GB/document_library/Herbal_-_HMPC_assessment_report/2011/04/WC500105228.pdf (accessed on 29 December 2017).
194. Biagi, M.; Pecorari, R.; Appendino, G.; Miraldi, E.; Magnano, A.R.; Governa, P.; Cettolin, G.; Giachetti, D. Herbal Products in Italy: The Thin Line between Phytotherapy, Nutrition and Parapharmaceuticals; A Normative Overview of the Fastest Growing Market in Europe. *Pharmaceuticals* **2016**, *9*, 65. [CrossRef] [PubMed]



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