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REVIEW ARTICLE

Phytotoxicity of Nanomaterials in Agriculture

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Abstract:

Science and technology have advanced rapidly in every aspect; thus, nanotechnology is one of the highly promising interdisciplinary approaches which has swiftly emerged in the world. The inherent properties of nanomaterials (NMs) made them widely accepted to use in many fields, including agriculture. Because of this, NMs have attracted novel agrochemical formulations to enhance crop productivity. However, deliberate and accidental release of nanoparticulate based agrochemical formulations and engineered NMs have raised concerns on the possible effects on agricultural crops. Therefore, the interaction of NMs leading to phytotoxicity is the biggest concern that is required to be assessed prior to their applications. Hence, this review discusses whether NMs can be used as a feasible stand-in candidate for agriculture.

Keywords: Carbon based nanomaterials, Metal based nanomaterials, Nanomaterials, Phytotoxicity, Silica nanoparticles, Rare earth oxide nanoparticles.

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1. INTRODUCTION

The development of nanotechnology has touched the quality of life in almost all stakeholders providing several products and benefits to the society, environment and economy [1]. Nanotechnology utilizes particles with at least a single dimension between 1-100 nm for these applications, where it is simply known as nanoparticles (NPs). Therefore, unique and novel properties of NPs enable to use them in various applications in medicine, energy, food industry, environmental remediation, electronics *etc* [2 - 7]. Also, agricultural sector possesses a great interest in using nanomaterials (NMs) to overcome the issues causing loss of soil fertility [8]. Therefore, as an interdisciplinary approach, nanotechnology has enabled to expand its applications over a range of fields.

Although the use of nanotechnology has gained a significant interest, the nonregulated use of NPs has caused several problems, challenges and consequences to living organisms and ecosystem balance [6]. The release of NMs derived from natural and anthropogenic sources raised several concerns on their potential toxicity. The NPs are released to the environment through the processes such as photochemical reactions, volcanic eruptions, forest fires, simple erosions as well as from plants and animals [9]. Furthermore, agricultural applications, atmospheric depositions, rain erosion, surface

runoffs lead to the entry of NPs into the soil. Reports have shown that weak migration ability of NPs in the soil leads to more accumulation in high concentrations than in water or air. Therefore, soil can be regarded as the major compact where NMs will end up [10, 11]. Hence, the deliberate or accidental release of NMs to the environment has intensified the questions regarding their use and effects on the ecosystem.

Plants as primary producers play a critical role in any community by harnessing solar energy, especially to carbohydrates and providing energy for other trophic levels [11]. Plants were evolved in the presence of natural NPs, but the present use of engineered NMs has enhanced their probability of exposure. Hence, direct and accidental release of NMs from contaminated soils and sediments or atmospheric fallouts results in the accumulation of NMs in plants [12].

The interaction of NMs with plants may show positive as well as negative impacts on plants. The capability of NMs to bioaccumulate, biomagnify and transfer *via* trophic levels led to make more concerns on their potential toxicity to plants. Because plants are the pioneers of fulfilling the food requirement of organisms either *via* direct or indirect modes, any harm to this trophic level will adversely affect other trophic groups [11 - 13]. Therefore, assessing the NMs as potential candidates for agricultural applications is imperative.

2. FACTORS AFFECTING THE TOXICITY OF NMs

The simplest explanation to understand the fate of NMs

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against biological systems is to study their physicochemical properties and the interactions with the environment [14]. Physicochemical properties include size, shape, composition and surface chemistry of NPs and these properties will determine their effects on living components. Primarily, NPs can be grouped according to their dimensionality, morphology, composition, uniformity and agglomeration properties. The dimensionality of NPs plays a critical role in its function, especially in their potential effects. Based on its dimensionality, NPs can be categorized as one-dimensional (1D), two-dimensional (2D) or three-dimensional (3D). Moreover, morphological characteristics, including flatness, sphericity, and aspect ratio can enhance their properties. Also, chemistry and electromagnetic properties allow the NPs to exist as dispersed aerosols, suspensions/colloids or in agglomerated states [14, 15]. Despite different classifications, NPs are generally classified based on their composition [9, 16, 17] as carbon-based nanomaterials (CNMs), metal based NMs, dendrimers and polymers, bio-inorganic complexes and quantum dots, etc.

Furthermore, interaction with the environment can alter their chemical properties and reactivity. Hence, this can lead to aggregate NMs forming complexes with organic matter and colloids in soil and water thereby, severely influencing bioavailability by abiotic factors [9, 14]. Moreover, same NM has different effects on different model plants. For instance, TiO₂ NPs on *Spinacia oleraces* show a 60% increment in plant fresh and dry weight. In contrast, the same NM at the same concentration tested on *Zea mays* shows a growth inhibition [18]. Therefore, the model plant selected for a study plays an important role in the phytotoxicity of NPs as well.

3. PLANT UPTAKE OF NPs

The NPs can be dispersed in different environmental matrices, in which their interaction can be extended from root to leaves [14]. Therefore, NPs accumulated in the atmosphere can directly interact with aerial parts of the plants where they can enter into plants through stomatal pathways, through plant stems and through other plant cellular channels [19]. In contrast to accumulation in the atmosphere, NPs can be frequently found in other environmental matrices such as water and soil. Hence, uptake of different NMs into plants occurs through its root systems *via* selective uptake and NPs even undergo biotransformation within the roots. Eventually, NPs may translocate to different parts of the plant [12, 20, 21].

In comparison to bulk counterparts of NMs, the physicochemical properties of NPs enhance their reactivity to their environs, thus leading to interaction with plant root exudates and peculiar membrane transporter. Studies have exploited three different means of NP penetrations into plant cells based on the size, shape, charge, hydrophobicity, chemical composition and stability as mentioned elsewhere in the text [22]. Direct diffusion of NPs through the phospholipid bilayer is regarded as the first mechanism. Secondly, penetration *via* endocytosis in which plasma membrane develops deformed inwards around the NPs, where this will invaginate to surrounding NMs followed by internalized

vesicles in the cells. The third mechanism involves ion channels and aquaporin. This mechanism is limited by the selectivity and small pore-sizes. Also, the establishment can be induced by the mechanical action of NPs and it is observed in the cell membranes [23].

Once NPs are internalized, they can be transported to different parts of the plant *via* apoplastic or symplastic pathways, *via* xylem vessels and crossing the plasmoderms. However, the efficiency of uptake and translocation is species-dependent where it may be based on the physiology of the plant species [14, 24]. In addition, many studies have exploited the characteristics of NPs, such as size, surface charge, mobility and dissolution within the biological system play a critical role in internalization within plants [25]. However, more studies are in need to better understand the behavior of NMs under realistic conditions.

4. PHYTOTOXICITY OF MOST WIDESPREAD NPs

Many reports available on the interactions between plants and NPs include the metallic NPs, CNMs and Silica NPs. Most of the studies have reported on metallic NPs because of their ease of absorption by roots and the ability to provide essential micronutrients in easily accessible form to plants [8, 14]. It is noteworthy to point out that the interactions of NMs with plants may show positive as well as negative impacts depending on the physicochemical properties. Hence, Fig. (1) has summarized the effects and interactions of NMs with plants. Therefore, some of the most utilized NPs as nanofertilizers, and their phytotoxic effects on different plant systems as well as the bioassays to assess the phytotoxic effects are reported here.

4.1. Plant Bioassays

Phytotoxicity studies are usually performed using well-established plant bioassays, which will allow screening and monitoring the environmental contaminants by analyzing the relation between the dose applied and the extent of induced damage in plants [26]. Ideal plant bioassays include easiness of handling, low cost, high sensitivity, and good correlation with the results of animal assays. Model plants include dicotyledons, such as *Vicia faba* L. and *Allium cepa* L. and monocotyledons; *Hordeum vulgare* L. and *Zea mays* L. Furthermore, edible plants such as ryegrass, zucchini, lettuce, radish crop plants and wheat have also been used in plant assays, especially on NMs [27]. The tests include a range of endpoints as physiological and morphological parameters and may evaluate the effects on mitotic activity, cell cycle (cytotoxicity), and DNA damage caused by clastogenic and mutagenic activity (genotoxicity) after an exposure period to assess acute or chronic damage [14].

4.2. Effects of Metal and Metal Oxide- Based NMs

A number of studies has been carried out on the phytotoxicity of metallic NPs. Metallic NMs such as Zn, Cu, Fe, Mn, Ag and their oxides, including TiO₂, Al₂O₃, CuO, Fe₂O₃ is widely exploited for their phytotoxicity in different model plants [8, 28 - 35].

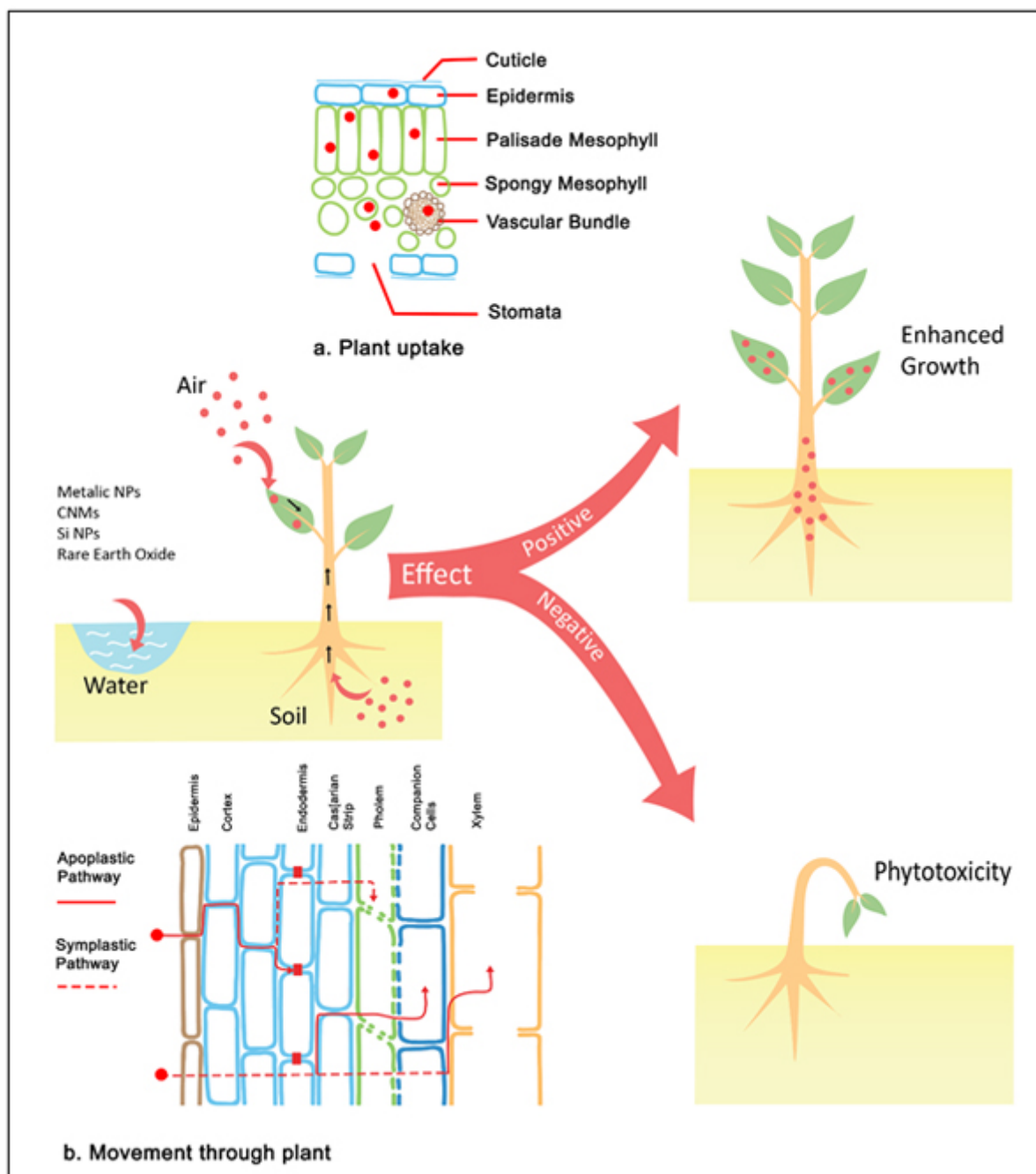


Fig. (1). Distribution, interactions and the effects of nanomaterials on plants.

4.2.1. Effects of Copper Oxide NPs

Rajput *et al.* [36] observed that CuO NPs inhibit the growth by affecting the shoot and root elongations, maximal quantum yield of photosystem II, and transpiration rate of *Hordeum sativum*. It was speculated that the suppression of photosynthesis might be due to decreased electron transport, the number of thylakoids per granum, photosynthetic rate, transpiration rate and stomatal conductance. Phytotoxicity of CuO NPs of size 43 nm in the presence of humic acid was reported by Peng *et al.* [37]. The authors reported that CuO NPs had induced the inhibition of root elongation, abnormality

in root morphology, and its ultrastructure. Further, cell viability and membrane integrity were also reported. These adverse effects had resulted from the generation of reactive oxygen species caused by CuO NPs, which led to mitochondrial dysfunction, lipid peroxidation, and programmed cell death in rice seedlings.

4.2.2. Effects of Iron and Zinc Oxide NPs

Treatment of Fe₂O₃ NPs at 50 mg/L concentration has decreased root elongation of *Lactuca sativa* by 20%, compared to control plants [38]. More recently, Sun *et al.* [39] reported

that *Allium cepa* plants treated with 5 and 50 µg/mL ZnO NPs showed cytotoxic and genotoxic effects in the root meristems by affecting the cell membrane integrity, metabolic activity, reactive oxygen species accumulation, DNA damage, chromosome aberrations and cell cycle progression. It was found that ZnO NPs treatments accounted for 24.2 or 36.1% root inhibitions when the plants were exposed to 36 h period.

4.2.3. Effects of Titanium dioxide NPs

Hu *et al.* [40] depicted that TiO₂ NPs with 400 mg/L concentration show a negative impact on the nutritional quality of *Lactuca sativa*. Exposure of *Oryza sativa* to higher concentrations of TiO₂ NPs (1000 ppm) led to accumulating higher amounts of Ti in roots than in shoots. Significant reduction in CO₂ fixation, transpiration rate and stomatal conductance was reported on tests. In addition, insignificant effects on photosynthetic pigments, quantum efficiency of PSII (Fv/Fm ratio) and photochemical quenching were also exploited by DaCosta and Sharma [41]. Frazier *et al.* [42] investigated the effect of TiO₂ NPs on tobacco and found that higher concentrations of NPs could significantly inhibit the germination rates, root lengths, and biomasses of tobacco seedlings. Similarly, TiO₂ NPs significantly influenced the expression profiles of microRNAs (miRNAs) that have been shown to play an imperative role in plant development as well as plant tolerance to abiotic stresses such as drought, salinity, cold, and heavy metal was also highlighted by authors.

4.2.4. Effects of Silver NPs

The Ag is another group of metallic NPs with wide use in different fields; however, Rui *et al.* [43] reported that soil amendment of Ag NPs on peanut cultivars (50, 500, and 2000 mg/kg) for 98 days had shown a significant reduction in plant growth and crop yield at the concentrations tested. Furthermore, the fatty acids in the grains of peanuts indicated the presence of Ag NPs could significantly alter the crop quality. In another study, Li *et al.* [44] point out that the possibility of contaminating fruits, seeds, and other edible parts by translocation processes in plants exposed to AgNPs.

In contrast, the same NMs show promising results in agricultural applications as potential candidates as nanofertilizers. Ghafariyan *et al.* [24] reported that low concentrations of superparamagnetic Fe-NPs (0-2 mg/L) significantly increased the chlorophyll contents in sub-apical leaves of soybeans under hydroponic conditions, proving that soybean could use this type of Fe-NPs as a source of Fe. The cucumber plants grown in soil treated with ZnO NPs at 400 and 800 mg/kg showed that at 400 mg/kg ZnO NPs had significantly increased the starch content was reported by Zhao *et al.* [45]. Green synthesized AuNPs using onion extracts on onion seeds were evaluated by Acharya *et al.* [46]. It was exhibited that applying AuNPs as priming agent at low concentrations (5.4 ppm) resulted in enhancement of germination, plant height, leaf length, leaf diameter, neck diameter, and leaf surface area. Moreover, Sathiyabama and Manikandan [47], speculated, both foliar spray or as a combined application (involving seed coat and foliar spray) of copper-chitosan NPs had enhanced the growth profile and crop

productivity of finger millet plants. In addition, suppression of blast disease development *via* increased defense enzymes of finger millet plants by copper-chitosan NPs portrays the capability of NPs to enhance the disease resistance as extra merit to use in agricultural applications.

4.3. Effects of Carbon Based NMs

The CNMs are recognized as one of the most promising NMs in nanotechnology because of its intense applications. Thus, many phytotoxic studies have been performed to evaluate its potential toxicity.

4.3.1. Effects Of Multi Walled Carbon Nanotubes And Single Walled Carbon Nanotubes

Studies on red spinach, cucumber and lettuce showed that exposure of 1000 and 2000 mg/L concentrations of multi walled carbon nanotubes (MWCNTs) on hydroponic systems had significantly reduced the shoot and root elongation [48]. Zaytseva *et al.* [49] confirmed that phytotoxicity of MWCNTs coupled with the oxidative stress in relation to disturbance of micronutrient homeostasis. Shen *et al.* [50] experimented with single walled carbon nanotubes (SWCNTs) on protoplasts of *Arabidopsis* and *Oryza* and reported that SWCNTs have a dose-dependent programmed cell death through oxidative stress. Moreover, test plants had shown adverse cellular responses such as cell aggregation, chromatin condensation, and plasma membrane deposition.

4.3.2. Effects of Fullerene and Graphene NPs

Fullerene (C₆₀) is another CNMs tested for phytotoxic effect in a study conducted by Santos *et al.* [51] on the aquatic plant *Lemna gibba*. They reported that the contents of chlorophylls a and b and the oxygen generation by chloroplasts were considerably decreased by the treatment of C₆₀. Furthermore, graphene was evaluated for its phytotoxic effects on cabbage, tomato and red spinach at a concentration range of 500 to 2000 mg/L. Graphene caused an inhibitory effect on plant growth and biomass on test plants compared to their controls. In addition, the leaf number and the size of the leaves were considerably reduced in a dose-dependent manner. Moreover, concentration dependent intensification in reactive oxygen species generation, as well as cell death, was also observed along with necrotic lesions on test plants [52]. Hao *et al.* [53] suggested that exposure to 50 and 150 mg/L mesoporous CNMs negatively affect rice growth *via* altering the levels of vital phytohormones and also, toxicity was particle size-dependent. In addition, exposure to 150 mg/L mesoporous CNMs (150 nm) reduced root length and shoot lengths by 21% and 29%, respectively. Whereas 80 nm sized CNMs significantly reduced the root and shoot lengths by 70% and 57% at the concentration of 150 mg/L.

Despite their phytotoxic effects, some studies have emphasized the merits using CNMs on enhanced agricultural production. Morphologically different CNMs such as helical-MWCNTs, few-layered graphene, long MWCNTs, and short MWCNTs can activate cell growth, germination, and plant growth, in which tests performed on tobacco cell cultures confirms the growth was found to increase by 22%–46% when

CNMs were introduced to the growth medium at a concentration of 50 µg/mL [54]. Application of graphene and MWCNTs on bioenergy crops (sorghum and switchgrass) increased the germination rate of switchgrass seeds and led to early germination of sorghum seeds. Furthermore, exposure of switchgrass to graphene (200 mg/l) enhanced the total biomass by 28% produced compared to untreated plants was reported by Pandey *et al.* [55]. Moreover, Pandey *et al.* [56] exposed MWCNTs and graphene had activated the early seed germination in *Catharanthus* and higher germination rate in cotton and *Catharanthus* seeds. Also, the soil growth of *Catharanthus* plants stimulated the reproductive system by inducing early flower development and accelerated total flower production by 37 and 58%, respectively.

Specifically, it was dictated that CNMs are multifaceted candidates to enhance plant growth as well as to eliminate the detrimental effects of environmental stresses from salt and drought with no symptoms on economically important crops [55, 56].

4.3.3. Effects of Carbon Dot NPs

Recently, Carbon dots (CDs) have been exploited as a novel form of CNMs showing a greater interest as markers for plant bioimaging. Because of their high aqueous solubility and flexibility in surface modification, the use of CDs has been spotlighted to be a promising substitute to conventional semiconductor quantum dots (QDs) and organic dyes [57 - 59]. When maize seedlings were exposed to 1000 and 2000 mg/L of CDs for 4 weeks, it was found that fresh weight of roots (57% and 68%) and shoot fresh weight (38% and 72%) were significantly reduced by CDs [60]. In contrast to other CNMs, the non-toxic carbon backbone of CDs shows much lower environmental toxicity and higher biocompatibility [58]. Li *et al.* [57] studied the phytotoxicity of fluorescent CDs on mung beans pointed out that, up to 0.4 mg/mL, there is a positive impact on the seed germination, root and stem elongation, fresh biomass, and moisture level of mung beans. Wang *et al.* [61] reported that CDs showed a dose-response effect on the growth of mung bean sprouts by promoting root elongation, stem elongation and biomass. In addition, mung bean sprouts treated with CDs had an elevated level in carbohydrate content by 21.9% compared to the control proving it a good candidate for fertilizers in agriculture.

4.4. Effects of Silica NPs

At an industrial scale, silica NPs rank top as a globally demanding product [14]. Silica NPs are widely utilized in fields like medicine, cosmetics, food industry, in biomedical and biotechnological applications as well as a nanofertilizer in agricultural applications [62]. Because of its extensive use, many studies have examined the potential toxicity of silica NPs on living beings [63]. Silicon (Si) is identified as a favorable element for the growth and development of plant systems where its contents are comparable to that of macronutrients essential for plant growth. Therefore, it has been widely used as a fertilizer to increase the yield of many crops [64]. Despite that, inherent physicochemical properties and the high reactivity of NPs led to assess the effects of silica NPs on seed germination and plant growth [14].

Silica NPs show phytotoxic effects principally at higher concentrations and sometimes the effects might depend on size, concentration and charge [65]. For example, at very high concentrations (540- 1820 mg/L), silica NPs had shown phytotoxic effects on *Allium cepa* with three different size ranges (7, 12, 22 nm). It was observed that silica NPs affected plant growth parameters such as germination and root elongation in seedlings. Moreover, cytogenetic analysis on root meristems has exhibited chromosomal abnormalities indicating its genotoxic effects [66]. The Bt- transgenic cotton plants tested with SiO₂ NPs at 2000 mg/L showed a significant decrease in the plant height, shoot and root biomasses as well as affected the micronutrient contents such as Cu, Mg in roots and Na content in roots of transgenic cotton. In addition, the activity of superoxide dismutase and IAA content were considerably impacted by SiO₂ NPs [67]. Therefore, it is important to improve the risk assessment of NPs under environmental exposures because the effects of NMs are concentration dependent as well as with the other physiochemical parameters.

Distinctive properties such as a large volume of tunable pores, high surface area, ease of surface functionalization, physical and chemical stability, high biocompatibility and low degradability under physiological conditions of mesoporous Si NPs, that make them ideal reservoirs for smart delivery systems [68 - 70]. Therefore, the applications of mesoporous Si NPs as a smart delivery system to increase the crop productivity has gained more attention in modern agriculture. Sun *et al.* [71] investigated the uptake of mesoporous SiNPs at specific concentrations (500 and 1000 mg/L) on wheat and lupin enhanced seed germination, plant biomass, leaf total protein and chlorophyll pigments. Also, the interaction of mesoporous SiNPs with the chloroplasts promotes plant growth by uplifting the photosynthesis of test plants.

4.5. Effects of Rare Earth Oxide NPs

Ma *et al.* [72] studied the effect of La₂O₃ NPs on the root growth of cucumber plants at concentrations over 200 mg/L. It was speculated that La₂O₃ NPs at the highest concentration (2000 mg/L) tested had shown a decrease in root elongation by 66% as a dose dependent response on cucumber. The release of Ce³⁺ ions plays a critical role in the phytotoxicity of CeO₂ NPs on romaine lettuce was highlighted by Zhang *et al.* [73]. Romaine lettuce grown under sand containing CeO₂ NPs had diminished the chlorophyll content by 16.5% and 25.8% at the highest concentrations tested (1000 and 2000 mg/kg). Moreover, there is a significant inhibition in the biomass production of lettuce as well as the concentrations assessed. Phytotoxicity of Y₂O₃ NPs on rice seedlings under hydroponic cultures was assessed by Zhao *et al.* [74]. Results indicated that high concentrations of Y₂O₃ NPs at 50 and 100 mg/L delayed seed germination. In contrast, lower concentrations including 1, 5, and 10 mg/L have shown positive effects on the root elongation of rice seedlings. Notably, Y₂O₃ NPs ranging from 20 to 100 mg/L significantly diminished the root activity and chlorophyll contents of rice. Furthermore, CeO₂ NPs at 250 mg/L significantly reduced the disease severity to fusarium wilt while improving the chlorophyll content and the nutritional

value of the tomato [75]. Therefore, all most all the studies available in the literature manifest the effect of different NMs have shown dose dependent responses to agricultural crops.

4.6. Effects of Polymeric and Other NPs

Chitosan/tripolyphosphate NPs had inhibited the germination *Zea mays*, *Brassica rapa*, and *Pisum sativum*. It was exploited that size, composition and the charge of the polymeric NPs had a significant effect with variable phytotoxicities [76]. Xin *et al.* [77] experimented on the effect of polysuccinimide polymeric NPs using *Zea mays* for

germination and seedling growth with response to Cu stress. These NPs had improved the germination index of corn by mitigating the Cu stress. Also, it was reported as a novel opportunity to promote plant growth under heavy metal stresses. However, only a handful of studies have been performed on the effects of polymeric NPs on different model plants [76]. Tables 1 and 2 summarized the positive and negative effects of different NMs tested for phytotoxic effects. Madanayake *et al.* [78] and Kottegoda *et al.* [79] suggested hydroxyapatite NPs and their modified form with urea can be better macronutrient supplier enhancing crop growth.

Table 1. Positive effects of NMs on different model plants.

Type of NMs	Size	Concentration	Test Plant	Effects	References
Superparamagnetic Fe-NPs	9 nm	0-2 mg/L	Soybeans	Increased the chlorophyll contents in sub-apical leaves	[24]
ZnO NPs	10 nm	400 and 800 mg/kg	Cucumber	At 400 mg/kg ZnO NPs had significantly increased the starch content	[45]
Green synthesized AuNPs	30–113 nm	5.4 ppm	Onion	Enhanced germination, plant height, leaf length, leaf diameter, neck diameter, and leaf surface area	[46]
Helical-MWCNTs, layered graphene, long MWCNTs, and short MWCNTs	13-18 nm	50 µg/mL	Tobacco cell cultures	Activate cell growth, germination, and plant growth Cell growth was increased by 22%–46% when CNMs were introduced to the growth medium	[54]
Graphene and MWCNTs	13–18 nm	200 mg/l	Sorghum and switchgrass	Switchgrass to graphene enhanced the total biomass by 28%	[55]
MWCNTs and graphene	13-18 nm		Catharanthus and cotton	Activated the early seed germination in Catharanthus and higher germination rate in cotton and Catharanthus seeds Stimulated the reproductive system by inducing early flower development and accelerated total flower production by 37 and 58%	[56]
CDs	4 - 6 nm	0.02-0.12 mg/mL	Mung bean	Promote root elongation, stem elongation and biomass Elevated levels in carbohydrate content by 21.9%	[61]
Mesoporous SiNPs	20 nm	500 and 1000 mg/L	Wheat and Lupin	Enhanced seed germination, plant biomass, leaf total protein and chlorophyll pigments	[71]
CeO ₂ NPs	8 ± 1 nm	250 mg/L	Tomato	Reduced disease severity to fusarium wilt Improved chlorophyll content and the nutritional value	[75]

Table 2. Negative effects of NMs on different model plants.

Type of NMs	Size	Concentration	Test Plant	Effects	References
CuO NPs	30–50 nm	10 g/L	<i>Hordeum sativum</i>	Inhibited growth by affecting the shoot and root elongations, maximal quantum yield of photosystem II, and transpiration rate	[36]
CuO NPs in the presence of humic acid	43 nm	0, 2, 5, 10, 20, 50, 100 mg /L	Rice	Induced the inhibition of root elongation, abnormality in root morphology and its ultrastructure	[37]
Fe ₂ O ₃ NPs at concentration	60 ± 27 × 30 ± 12 nm	50 mg/L	<i>Lactuca sativa</i>	Decreased the root elongation by 20%, compared to control	[38]
ZnO NPs	< 50 nm	5 and 50 µg/mL	<i>Allium cepa</i>	Cytotoxic and genotoxic effects in the root meristems	[39]
TiO ₂ NPs		400 mg/L	<i>Lactuca sativa</i>	Negative impact on the nutritional quality	[40]

(Table 4) contd....

Type of NMs	Size	Concentration	Test Plant	Effects	References
TiO ₂ NPs		1000 ppm	<i>Oryza sativa</i>	Reduction in CO ₂ fixation, transpiration rate and stomatal conductance Effects on photosynthetic pigments, quantum efficiency of PSII (Fv/Fm ratio) and photochemical quenching	[41]
TiO ₂ NPs	<25 nm	0, 0.1, 1, 2.5, or 5%	Tobacco	Inhibit the germination rates, root lengths, and biomasses of tobacco seedlings	[42]
Ag NPs	20 nm	50, 500, and 2000 mg/kg	Peanut	Reduction in plant growth and crop yield	[43]
MWCNTs	Outer mean diameter ~13 nm, Inner mean diameter ~4 nm	1000 and 2000 mg/L	Red spinach, cucumber and lettuce	Significantly reduced the shoot and root elongation	[48]
SWCNTs	Diameter of 1 – 2 nm	5–250 µ g/mL	<i>Arabidopsis</i> and <i>Oryza</i>	Dose-dependent programmed cell death through oxidative stress	[50]
C ₆₀	29-38 nm		<i>Lemna gibba</i>	Contents of chlorophylls a and b and the oxygen generation by chloroplasts were considerably decreased	[51]
Graphene	1 nm	500 to 2000 mg/L	Cabbage, tomato and red spinach	Inhibitory effect on plant growth and biomass Leaf number and the size of the leaves were considerably reduced	[52]
Mesoporous CNMs	80 nm and 150 nm	50 and 150 mg/L	Rice	Exposure to 150 mg/L (150 nm) reduced root length and shoot lengths by 21% and 29%. Significantly reduced the root and shoot lengths by 70% and 57% at the concentration 150 mg/L (80 nm)	[53]
Silica NPs	7, 12, 22 nm	540- 1820 mg/L	<i>Allium cepa</i>	Affected on germination and root elongation in seedlings Cytogenetic analysis on root meristems showed chromosomal abnormalities	[66]
SiO ₂ NPs	35 nm	2000 mg/L	Bt- transgenic cotton	Decrease in the plant height, shoot and root biomasses Affected micronutrient contents such as Cu, Mg in roots and Na content in roots	[67]
La ₂ O ₃ NPs	65.8 ± 10.5 nm	200 mg/L. - 2000 mg/L	Cucumber	Decrease in root elongation by 66% at the highest concentration	[72]
CeO ₂ NPs	16.5 ± 6.8 nm	1000 and 2000 mg/kg	Lettuce	Diminished chlorophyll content by 16.5% and 25.8% at highest concentrations tested	[73]
Y ₂ O ₃ NPs	20-30 nm	50 and 100 mg/L	Rice seedlings	Delayed seed germination	[74]
Hydroxyapatite NPs	diameter of 20 nm and an average length 150 nm	0-10000 mg/L	<i>Raphanus sativus</i>	Hydroxyapatite NPs were biotransformed within the roots of the test plant. Shoot, root lengths and dry biomass were significantly enhanced with no effect on soluble protein and indole acetic acid content at the highest concentration tested.	[78]

CONCLUSION AND FUTURE PROSPECTUS

Nanotechnology as an interdisciplinary approach has been linked with almost each and every field in science. Therefore, the utilization of NMs leads to release them deliberately or accidentally to the environment. Currently, agriculture has focused on using different nanoformulations on agricultural different applications, including fertilizers. However, NMs show differential effects of various plants based on their concentrations, environmental factors, and the tested species where it might either have positive or negative impacts. Despite their phytotoxic effects, NMs exhibits a range of beneficial effects with respect to the growth and development of plants that are particularly exposed to environmental stresses. Thus, it is important to rationally and safely apply NPs in agriculture as well as to establish a comprehensive system to effectively evaluate their impacts on crop plants. Therefore, in

the future it important to exploit effective and safe strategies of application of different NPs incorporated products in agriculture. Also, more comprehensive studies should be carried out to evaluate the most effective concentrations of NPs on different crops as recommendations for agricultural applications. Finally, how to control their potential impact on food safety and food quality should draw more attention as NPs themselves could be taken up by crops and humans exposed to them through food consumption.

CONSENT FOR PUBLICATION

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CONFLICT OF INTEREST

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REFERENCES

- [1] Tolaymat T, El Badawy A, Sequeira R, Genaidy A. A system-of-systems approach as a broad and integrated paradigm for sustainable engineered nanomaterials. *Sci Total Environ* 2015; 511: 595-607. [http://dx.doi.org/10.1016/j.scitotenv.2014.09.029] [PMID: 25590540]
- [2] Boisseau P, Loubaton B. Nanomedicine, nanotechnology in medicine. *C R Phys* 2011; 12(7): 620-36. [http://dx.doi.org/10.1016/j.crh.2011.06.001]
- [3] Joo SH, Cheng F. Nanotechnology for environmental remediation. Springer Science & Business Media 2006. [http://dx.doi.org/10.1007/b137366]
- [4] Rae A. Real life applications of nanotechnology in electronics. *OnBoard Technol* 2006; 2006: 28.
- [5] Rashidi L, Khosravi-Darani K. The applications of nanotechnology in food industry. *Crit Rev Food Sci Nutr* 2011; 51(8): 723-30. [http://dx.doi.org/10.1080/10408391003785417] [PMID: 21838555]
- [6] Yang J, Cao W, Rui Y. Interactions between nanoparticles and plants: phytotoxicity and defense mechanisms. *J Plant Interact* 2017; 12(1): 158-69. [http://dx.doi.org/10.1080/17429145.2017.1310944]
- [7] Zang L, Ed. Energy efficiency and renewable energy through nanotechnology. Berlin: Springer 2011. [http://dx.doi.org/10.1007/978-0-85729-638-2]
- [8] Ruttkey-Nedecky B, Krystofova O, Nejd L, Adam V. Nanoparticles based on essential metals and their phytotoxicity. *J Nanobiotechnology* 2017; 15(1): 33. [http://dx.doi.org/10.1186/s12951-017-0268-3] [PMID: 28446250]
- [9] Remédios C, Rosário F, Bastos V. Environmental nanoparticles interactions with plants: Morphological, physiological, and genotoxic aspects. *J Bot* 2012. [http://dx.doi.org/10.1155/2012/751686]
- [10] Gottschalk F, Sonderer T, Scholz RW, Nowack B. Modeled environmental concentrations of engineered nanomaterials (TiO₂, ZnO, Ag, CNT, Fullerenes) for different regions. *Environ Sci Technol* 2009; 43(24): 9216-22. [http://dx.doi.org/10.1021/es9015553] [PMID: 20000512]
- [11] McKee MS, Filsler J. Impacts of metal-based engineered nanomaterials on soil communities. *Environ Sci Nano* 2016; 3(3): 506-33. [http://dx.doi.org/10.1039/C6EN00007J]
- [12] Rico CM, Majumdar S, Duarte-Gardea M, Peralta-Videa JR, Gardea-Torresdey JL. Interaction of nanoparticles with edible plants and their possible implications in the food chain. *J Agric Food Chem* 2011; 59(8): 3485-98. [http://dx.doi.org/10.1021/jf104517j] [PMID: 21405020]
- [13] Rienzle R, Adassooriya NM. Toxicity of nanomaterials in agriculture and food. In: Rai M, Biswas J, Eds. *Nanomaterials: Ecotoxicity, safety, and public perception*. Cham: Springer 2018; pp. 207-34. [http://dx.doi.org/10.1007/978-3-030-05144-0_11]
- [14] Giorgetti L. Effects of nanoparticles in plants: Phytotoxicity and genotoxicity assessment. In: Durgesh KT, Parvaiz A, Shivesh S, Devendra KC, Nawal KD, Eds. *Nanomaterials in Plants, Algae and Microorganisms*. Academic Press 2019; pp. 65-87.
- [15] Buzea C, Pacheco II, Robbie K. Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases* 2007; 2(4): MR17-71. [http://dx.doi.org/10.1116/1.2815690] [PMID: 20419892]
- [16] Zhang D, Hua T, Xiao F, et al. Phytotoxicity and bioaccumulation of ZnO nanoparticles in *Schoenoplectus tabernaemontani*. *Chemosphere* 2015; 120: 211-9. [http://dx.doi.org/10.1016/j.chemosphere.2014.06.041] [PMID: 25063888]
- [17] Zhang P, Ma Y, Zhang Z. Interactions between engineered nanomaterials and plants: phytotoxicity, uptake, translocation, and biotransformation. In: Siddiqui M, Al-Wahaibi M, Mohammad F, Eds. *Nanotechnology and plant sciences*. Cham: Springer 2015; pp. 77-99.
- [18] Rastogi A, Zivcak M, Sytar O, et al. Impact of metal and metal oxide nanoparticles on plant: a critical review. *Front Chem* 2017; 5: 78. [http://dx.doi.org/10.3389/fchem.2017.00078] [PMID: 29075626]
- [19] Wang WN, Tarafdar JC, Biswas P. Nanoparticle synthesis and delivery by an aerosol route for watermelon plant foliar uptake. *J Nanopart Res* 2013; 15(1): 1417. [http://dx.doi.org/10.1007/s11051-013-1417-8]
- [20] Tripathi A, Liu S, Singh PK, et al. Differential phytotoxic responses of silver nitrate (AgNO₃) and silver nanoparticle (AgNps) in *Cucumis sativus* L. *Plant Gene* 2017; 11: 255-64. [http://dx.doi.org/10.1016/j.plgene.2017.07.005]
- [21] Tripathi DK, Singh S, Singh S, et al. Nitric oxide alleviates silver nanoparticles (AgNps)-induced phytotoxicity in *Pisum sativum* seedlings. *Plant Physiol Biochem* 2017; 110: 167-77. [http://dx.doi.org/10.1016/j.plaphy.2016.06.015] [PMID: 27449300]
- [22] Mehrian SK, De Lima R. Nanoparticles cyto and genotoxicity in plants: Mechanisms and abnormalities. *Environ Nanotechnol Monit Manag* 2016; 6: 184-93. [http://dx.doi.org/10.1016/j.enmm.2016.08.003]
- [23] Schmidt J. Nanoparticle-Induced Membrane Pore Formation Studied with Lipid Bilayer Arrays. *Biophys J* 2015; 108(2): 344a-5a. [http://dx.doi.org/10.1016/j.bpj.2014.11.1888]
- [24] Ghafariyan MH, Malakouti MJ, Dadpour MR, Stroeve P, Mahmoudi M. Effects of magnetite nanoparticles on soybean chlorophyll. *Environ Sci Technol* 2013; 47(18): 10645-52. [http://dx.doi.org/10.1021/es402249b] [PMID: 23951999]
- [25] Singh D, Kumar A. Impact of irrigation using water containing CuO and ZnO nanoparticles on Spinach oleracea grown in soil media. *Bull Environ Contam Toxicol* 2016; 97(4): 548-53. [http://dx.doi.org/10.1007/s00128-016-1872-x] [PMID: 27370820]
- [26] Grant WF. The present status of higher plant bioassays for the detection of environmental mutagens. *Mutat Res* 1994; 310(2): 175-85. [http://dx.doi.org/10.1016/0027-5107(94)90112-0] [PMID: 7523890]
- [27] Gui X, Rui M, Song Y, et al. Phytotoxicity of CeO₂ nanoparticles on radish plant (*Raphanus sativus*). *Environ Sci Pollut Res Int* 2017; 24(15): 13775-81. [http://dx.doi.org/10.1007/s11356-017-8880-1] [PMID: 28401392]
- [28] Mazumdar H, Ahmed GU. Phytotoxicity effect of silver nanoparticles on *Oryza sativa*. *Int J Chemtech Res* 2011; 3(3): 1494-500.
- [29] Atha DH, Wang H, Petersen EJ, et al. Copper oxide nanoparticle mediated DNA damage in terrestrial plant models. *Environ Sci Technol* 2012; 46(3): 1819-27. [http://dx.doi.org/10.1021/es202660k] [PMID: 22201446]
- [30] Burklew CE, Ashlock J, Winfrey WB, Zhang B. Effects of aluminum oxide nanoparticles on the growth, development, and microRNA expression of tobacco (*Nicotiana tabacum*). *PLoS One* 2012; 7(5): e34783. [http://dx.doi.org/10.1371/journal.pone.0034783] [PMID: 22606225]
- [31] Faisal M, Saquib Q, Alatar AA, Al-Khedhairi AA, Hegazy AK, Musarrat J. Phytotoxic hazards of NiO-nanoparticles in tomato: a study on mechanism of cell death. *J Hazard Mater* 2013; 250-251: 318-32. [http://dx.doi.org/10.1016/j.jhazmat.2013.01.063] [PMID: 23474406]
- [32] Gui X, Deng Y, Rui Y, et al. Response difference of transgenic and conventional rice (*Oryza sativa*) to nanoparticles (γ-Fe₂O₃). *Environ Sci Pollut Res Int* 2015; 22(22): 17716-23. [http://dx.doi.org/10.1007/s11356-015-4976-7] [PMID: 26154040]
- [33] Chichiricò G, Poma A. Penetration and toxicity of nanomaterials in higher plants. *Nanomaterials (Basel)* 2015; 5(2): 851-73. [http://dx.doi.org/10.3390/nano5020851] [PMID: 28347040]
- [34] Hossain Z, Mustafa G, Sakata K, Komatsu S. Insights into the proteomic response of soybean towards Al₂O₃, ZnO, and Ag nanoparticles stress. *J Hazard Mater* 2016; 304: 291-305. [http://dx.doi.org/10.1016/j.jhazmat.2015.10.071] [PMID: 26561753]
- [35] Zhao L, Huang Y, Hu J, Zhou H, Adeleye AS, Keller AA. 1H NMR and GC-MS based metabolomics reveal defense and detoxification mechanism of cucumber plant under nano-Cu stress. *Environ Sci Technol* 2016; 50(4): 2000-10. [http://dx.doi.org/10.1021/acs.est.5b05011] [PMID: 26751164]
- [36] Rajput V, Minkina T, Fedorenko A, et al. Toxicity of copper oxide nanoparticles on spring barley (*Hordeum sativum distichum*). *Sci Total Environ* 2018; 645: 1103-13. [http://dx.doi.org/10.1016/j.scitotenv.2018.07.211] [PMID: 30248835]
- [37] Peng C, Zhang H, Fang H, et al. Natural organic matter-induced alleviation of the phytotoxicity to rice (*Oryza sativa* L.) caused by copper oxide nanoparticles. *Environ Toxicol Chem* 2015; 34(9):

- 1996-2003.
[http://dx.doi.org/10.1002/etc.3016] [PMID: 25868010]
- [38] Liu R, Zhang H, Lal R. Effects of stabilized nanoparticles of copper, zinc, manganese, and iron oxides in low concentrations on lettuce (*Lactuca sativa*) seed germination: nanotoxicants or nonnutrients? *Water Air Soil Pollut* 2016; 227(1): 42.
[http://dx.doi.org/10.1007/s11270-015-2738-2]
- [39] Sun Z, Xiong T, Zhang T, Wang N, Chen D, Li S. Influences of zinc oxide nanoparticles on *Allium cepa* root cells and the primary cause of phytotoxicity. *Ecotoxicology* 2019; 28(2): 175-88.
[http://dx.doi.org/10.1007/s10646-018-2010-9] [PMID: 30612257]
- [40] Hu J, Wu X, Wu F, *et al.* TiO₂ nanoparticle exposure on lettuce (*Lactuca sativa* L.): dose-dependent deterioration of nutritional quality. *Environ Sci Nano* 2020; 7(2): 501-13.
[http://dx.doi.org/10.1039/C9EN01215J]
- [41] DaCosta MV, Sharma PK. Influence of titanium dioxide nanoparticles on the photosynthetic and biochemical processes in *Oryza sativa*. *Int J Rec Sci Res* 2015; 6(1): 2445-51.
- [42] Frazier TP, Burklew CE, Zhang B. Titanium dioxide nanoparticles affect the growth and microRNA expression of tobacco (*Nicotiana tabacum*). *Funct Integr Genomics* 2014; 14(1): 75-83.
[http://dx.doi.org/10.1007/s10142-013-0341-4] [PMID: 24132512]
- [43] Rui M, Ma C, Tang X, *et al.* Phytotoxicity of silver nanoparticles to peanut (*Arachis hypogaea* L.): physiological responses and food safety. *ACS Sustain Chem& Eng* 2017; 5(8): 6557-67.
[http://dx.doi.org/10.1021/acsschemeng.7b00736]
- [44] Li CC, Dang F, Li M, *et al.* Effects of exposure pathways on the accumulation and phytotoxicity of silver nanoparticles in soybean and rice. *Nanotoxicology* 2017; 11(5): 699-709.
[http://dx.doi.org/10.1080/17435390.2017.1344740] [PMID: 28627335]
- [45] Zhao L, Peralta-Videa JR, Rico CM, *et al.* CeO₂ and ZnO nanoparticles change the nutritional qualities of cucumber (*Cucumis sativus*). *J Agric Food Chem* 2014; 62(13): 2752-9.
[http://dx.doi.org/10.1021/jf405476u] [PMID: 24611936]
- [46] Acharya P, Jayaprakasha GK, Crosby KM, Jifon JL, Patil BS. Green-synthesized nanoparticles enhanced seedling growth, yield, and quality of onion (*Allium cepa* L.). *ACS Sustain Chem& Eng* 2019; 7(17): 14580-90.
[http://dx.doi.org/10.1021/acsschemeng.9b02180]
- [47] Sathiyabama M, Manikandan A. Application of copper-chitosan nanoparticles stimulate growth and induce resistance in finger millet (*Eleusine coracana Gaertn.*) plants against blast disease. *J Agric Food Chem* 2018; 66(8): 1784-90.
[http://dx.doi.org/10.1021/acs.jafc.7b05921] [PMID: 29443531]
- [48] Begum P, Ikhtiar R, Fugetsu B, Matsuoka M, Akasaka T, Watari F. Phytotoxicity of multi-walled carbon nanotubes assessed by selected plant species in the seedling stage. *Appl Surf Sci* 2012; 262: 120-4.
[http://dx.doi.org/10.1016/j.apsusc.2012.03.028]
- [49] Zaytseva O, Wang Z, Neumann G. Phytotoxicity of carbon nanotubes in soybean as determined by interactions with micronutrients. *J Nanopart Res* 2017; 19(2): 29.
[http://dx.doi.org/10.1007/s11051-016-3722-5]
- [50] Shen CX, Zhang QF, Li J, Bi FC, Yao N. Induction of programmed cell death in *Arabidopsis* and rice by single-wall carbon nanotubes. *Am J Bot* 2010; 97(10): 1602-9.
[http://dx.doi.org/10.3732/ajb.1000073] [PMID: 21616795]
- [51] Santos SM, Dinis AM, Rodrigues DM, Peixoto F, Videira RA, Jurado AS. Studies on the toxicity of an aqueous suspension of C60 nanoparticles using a bacterium (gen. *Bacillus*) and an aquatic plant (*Lemna gibba*) as in vitro model systems. *Aquat Toxicol* 2013; 142-143: 347-54.
[http://dx.doi.org/10.1016/j.aquatox.2013.09.001] [PMID: 24084257]
- [52] Begum P, Ikhtiar R, Fugetsu B. Graphene phytotoxicity in the seedling stage of cabbage, tomato, red spinach, and lettuce. *Carbon* 2011; 49(12): 3907-19.
[http://dx.doi.org/10.1016/j.carbon.2011.05.029]
- [53] Hao Y, Xu B, Ma C, *et al.* Synthesis of novel mesoporous carbon nanoparticles and their phytotoxicity to rice (*Oryza sativa* L.). *J Saudi Chem Soc* 2019; 23(1): 75-82.
[http://dx.doi.org/10.1016/j.jscs.2018.05.003]
- [54] Lahiani MH, Dervishi E, Ivanov I, Chen J, Khodakovskaya M. Comparative study of plant responses to carbon-based nanomaterials with different morphologies. *Nanotechnology* 2016; 27(26):265102
[http://dx.doi.org/10.1088/0957-4484/27/26/265102] [PMID: 27195934]
- [55] Pandey K, Lahiani MH, Hicks VK, Hudson MK, Green MJ, Khodakovskaya M. Effects of carbon-based nanomaterials on seed germination, biomass accumulation and salt stress response of bioenergy crops. *PLoS One* 2018; 13(8): e0202274.
[http://dx.doi.org/10.1371/journal.pone.0202274] [PMID: 30153261]
- [56] Pandey K, Anas M, Hicks VK, Green MJ, Khodakovskaya MV. Improvement of commercially valuable traits of industrial crops by application of carbon-based nanomaterials. *Sci Rep* 2019; 9(1): 19358.
[http://dx.doi.org/10.1038/s41598-019-55903-3] [PMID: 31852946]
- [57] Li W, Zheng Y, Zhang H, *et al.* Phytotoxicity, uptake, and translocation of fluorescent carbon dots in mung bean plants. *ACS Appl Mater Interfaces* 2016; 8(31): 19939-45.
[http://dx.doi.org/10.1021/acsmi.6b07268] [PMID: 27425200]
- [58] Li Y, Xu X, Wu Y, *et al.* A review on the effects of carbon dots in plant systems. *Mater Chem Front* 2020; 4(2): 437-48.
[http://dx.doi.org/10.1039/C9QM00614A]
- [59] Xu X, Mao X, Zhuang J, *et al.* PVA-Coated Fluorescent Carbon Dot Nanocapsules as an Optical Amplifier for Enhanced Photosynthesis of Lettuce. *ACS Sustain Chem& Eng* 2020; 8(9): 3938-49.
[http://dx.doi.org/10.1021/acsschemeng.9b07706]
- [60] Chen J, Dou R, Yang Z, *et al.* The effect and fate of water-soluble carbon nanodots in maize (*Zea mays* L.). *Nanotoxicology* 2016; 10(6): 818-28.
[http://dx.doi.org/10.3109/17435390.2015.1133864] [PMID: 26694806]
- [61] Wang H, Zhang M, Song Y, *et al.* Carbon dots promote the growth and photosynthesis of mung bean sprouts. *Carbon* 2018; 136: 94-102.
[http://dx.doi.org/10.1016/j.carbon.2018.04.051]
- [62] Napierska D, Thomassen LC, Lison D, Martens JA, Hoet PH. The nanosilica hazard: another variable entity. *Part Fibre Toxicol* 2010; 7(1): 39.
[http://dx.doi.org/10.1186/1743-8977-7-39] [PMID: 21126379]
- [63] Fubini B, Hubbard A. Reactive oxygen species (ROS) and reactive nitrogen species (RNS) generation by silica in inflammation and fibrosis. *Free Radic Biol Med* 2003; 34(12): 1507-16.
[http://dx.doi.org/10.1016/S0891-5849(03)00149-7] [PMID: 12788471]
- [64] Takahashi E, Ma JF, Miyake Y. The possibility of silicon as an essential element for higher plants. *Comments on Agricultural and Food Chemistry* 1990; 2(2): 99-102.
- [65] Slomberg DL, Schoenfisch MH. Silica nanoparticle phytotoxicity to *Arabidopsis thaliana*. *Environ Sci Technol* 2012; 46(18): 10247-54.
[PMID: 22889047]
- [66] Da Silva GH, Monteiro RT. Toxicity assessment of silica nanoparticles on *Allium cepa*. *Ecotoxicol Environ Contam* 2017; 12(1): 25-31.
[http://dx.doi.org/10.5132/eec.2017.01.04]
- [67] Le VN, Rui Y, Gui X, Li X, Liu S, Han Y. Uptake, transport, distribution and Bio-effects of SiO₂ nanoparticles in Bt-transgenic cotton. *J Nanobiotechnology* 2014; 12(1): 50.
[http://dx.doi.org/10.1186/s12951-014-0050-8] [PMID: 25477033]
- [68] Li X, Xie QR, Zhang J, Xia W, Gu H. The packaging of siRNA within the mesoporous structure of silica nanoparticles. *Biomaterials* 2011; 32(35): 9546-56.
[http://dx.doi.org/10.1016/j.biomaterials.2011.08.068] [PMID: 21906804]
- [69] Nandiyanto AB, Kim SG, Iskandar F, Okuyama K. Synthesis of spherical mesoporous silica nanoparticles with nanometer-size controllable pores and outer diameters. *Microporous Mesoporous Mater* 2009; 120(3): 447-53.
[http://dx.doi.org/10.1016/j.micromeso.2008.12.019]
- [70] Sun D, Hussain HI, Yi Z, *et al.* Uptake and cellular distribution, in four plant species, of fluorescently labeled mesoporous silica nanoparticles. *Plant Cell Rep* 2014; 33(8): 1389-402.
[http://dx.doi.org/10.1007/s00299-014-1624-5] [PMID: 24820127]
- [71] Sun D, Hussain HI, Yi Z, Rookes JE, Kong L, Cahill DM. Mesoporous silica nanoparticles enhance seedling growth and photosynthesis in wheat and lupin. *Chemosphere* 2016; 152: 81-91.
[http://dx.doi.org/10.1016/j.chemosphere.2016.02.096] [PMID: 26963239]
- [72] Ma Y, He X, Zhang P, *et al.* Phytotoxicity and biotransformation of La₂O₃ nanoparticles in a terrestrial plant cucumber (*Cucumis sativus*). *Nanotoxicology* 2011; 5(4): 743-53.
[http://dx.doi.org/10.3109/17435390.2010.545487] [PMID: 21261455]
- [73] Zhang P, Ma Y, Liu S, *et al.* Phytotoxicity, uptake and transformation of nano-CeO₂ in sand cultured romaine lettuce. *Environ Pollut* 2017; 220(Pt B): 1400-8.

- [74] [http://dx.doi.org/10.1016/j.envpol.2016.10.094] [PMID: 27843018] Zhao X, Zhang W, He Y, *et al.* Phytotoxicity of Y_2O_3 nanoparticles and Y^{3+} ions on rice seedlings under hydroponic culture. *Chemosphere* 2021; 263(1): 127943. [http://dx.doi.org/10.1016/j.chemosphere.2020.127943] [PMID: 32822939]
- [75] Adisa IO, Reddy Pullagurala VL, Rawat S, *et al.* Role of cerium compounds in Fusarium wilt suppression and growth enhancement in tomato (*Solanum lycopersicum*). *J Agric Food Chem* 2018; 66(24): 5959-70. [http://dx.doi.org/10.1021/acs.jafc.8b01345] [PMID: 29856619]
- [76] Nakasato DY, Pereira AES, Oliveira JL, Oliveira HC, Fraceto LF. Evaluation of the effects of polymeric chitosan/tripolyphosphate and solid lipid nanoparticles on germination of *Zea mays*, *Brassica rapa* and *Pisum sativum*. *Ecotoxicol Environ Saf* 2017; 142: 369-74. [http://dx.doi.org/10.1016/j.ecoenv.2017.04.033] [PMID: 28437729]
- [77] Xin X, Zhao F, Rho JY, Goodrich SL, Sumerlin BS, He Z. Use of polymeric nanoparticles to improve seed germination and plant growth under copper stress. *Sci Total Environ* 2020; 745(1): 141055. [http://dx.doi.org/10.1016/j.scitotenv.2020.141055] [PMID: 32736110]
- [78] Madanayake NH, Adassooriya NM, Salim N. The effect of hydroxyapatite nanoparticles on *Raphanus sativus* with respect to seedling growth and two plant metabolites. *Environ Nanotechnol Monit Manag* 15100404 [http://dx.doi.org/10.1016/j.enmm.2020.100404]
- [79] Kottegoda N, Munaweera I, Madusanka N, Karunaratne V. A green slow-release fertilizer composition based on urea-modified hydroxyapatite nanoparticles encapsulated wood. *Curr Sci* 2011; 73-8.

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