**Piper sarmentosum as an Antioxidant: A Systematic Review**

**Piper sarmentosum sebagai Antioksidan: Ulasan Sistematik**

**SITI MARJIANA ISMAIL, CHUA KIEN HUI, AMILIA AMINUDDIN & AZIZAH UGUSMAN**

**ABSTRACT**

Piper sarmentosum (PS) is an herb with various medicinal properties. The antioxidant activity of PS contributes to many of its pharmacological effects such as anti-hypertension, anti-cancer and anti-diabetes. This systematic review provides information regarding the antioxidant activity of PS. The review was conducted systematically to identify relevant published articles on the antioxidant activity of PS. The collected data was based on the searched articles through PubMed, Science Direct and Scopus databases between the years 1946 until March 2018. Only articles written in English and related to antioxidant activity of PS were included in this review. Based on the literature searched, 130 potential articles were identified and 19 articles met the inclusion criteria. Ten studies related to chemical assays, five studies combined in vivo animal and chemical assays, three studies combined chemical assays and in vitro studies and a single study combined chemical assay, in vitro and in vivo studies were included in this review. All studies showed positive effects of PS against oxidation, indicating the potential of PS as a source of natural antioxidant.

**Keywords:** Antioxidant; free radicals; free radical scavenging effect; oxidative stress; Piper sarmentosum

**INTRODUCTION**

*Piper sarmentosum* (PS) (Figure 1) is an herbaceous plant that belongs to the family of Piperaceae (Hussain et al. 2009). The vernacular names of PS vary among different countries. It is known as *daun kaduk*, wild pepper, *wild betel* or *sirih duduk* in Malaysia; *karak* or *sirih tanah* in Indonesia; *cha plu* in Thailand; *bo la lot* in Vietnam; *phak i leut* in Laos and *jia ju*, *xi ye ging wei teng* or *qing ju* in China. The plant easily grows in shady areas (Hussain et al. 2012). It is glabrous and creeping with 40-50 cm height of procumbent branches (Rahman et al. 2016). The leaves are alternate, broadly ovate to elliptic with light to dark green colour (Arunrat et al. 2006). The leaves emit a pungent peppery scent when crushed and the size is 4.5-6 cm wide and 7.5-9.5 cm long (Arunrat et al. 2006).

PS has been widely used as food and traditional medicine especially in the South East Asian countries (Abd Jalil et al. 2012). The plant is consumed alone as culinary herb or in combination with other herbs. Traditionally, PS is used for medicinal purposes to treat cough, flu, rheumatism, fever, tooth pain, foot dermatitis, asthma and pleurisy (Abd Jalil et al. 2012; Lee et al. 2011). In Thailand, PS was used to treat minor wounds (Abd Jalil et al. 2012), as an expectorant or anti-diabetic agent (Hutadilok et al. 2006), digestive tonic or anti-malarial agent (Chanwitheesuk et al. 2005).

Oxidative stress happens when there is imbalance between oxidants and antioxidant defense mechanisms in the body (Sies 1997). Oxidative stress causes lipid peroxidation, DNA mutation, membrane protein damage and apoptosis; thereby leading to cancer, cardiovascular diseases and neurodegenerative disorders (Renugadevi & Prabu 2010). Antioxidants are substances that reduce or prevent oxidation by interfering with the transfer of electrons from a substance to an oxidising agent (Velioiglu et al. 1998). The main function of antioxidants is to protect...

The high antioxidant activities of *P. sarmentosum* are related to the amounts of total polyphenols and flavonoids in the extracts (Lee et al. 2014). Other antioxidant compounds found in *P. sarmentosum* are alkaloids, amides, pyrones, sterols and neolignans (Hussain et al. 2009) as well as vitamin C, Vitamin E and carotenes (Chanwitheeusk et al. 2005). The present systematic review is designed to summarize up-to-date relevant citations.

**FIGURE 1. *Piper sarmentosum***

The body against free radicals damage. Endogenous antioxidants include glutathione, uric acid and bilirubin while antioxidant enzymes include superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX). In addition to endogenous antioxidants, optimal antioxidant status can also be achieved by consuming exogenous antioxidants from natural sources such as coffee, honey, tea, fruits and vegetables (Nur Syamsina et al. 2017).

**SEARCH STRATEGY**

A systematic literature searched was conducted to identify relevant research studies on antioxidant activity of *P. sarmentosum*. The method was based on the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) checklist (Moher et al. 2015). The search for the research papers involved three databases; PubMed (http://www.ncbi.nlm.nih.gov/pubmed/), Science Direct (http://www.sciencedirect.com) and Scopus (https://www.scopus.com/) from 1946 to March 2018. The search strategy comprised of combinations of the following two keywords: ‘*Piper sarmentosum*’ and ‘antioxidant’ for the three databases. The references to all retrieved articles were reviewed for relevant citations.

**SELECTION CRITERIA**

Limitation of article type, publication status and language were compulsory. Only the original articles published in English language were included in this review. Review articles, books, chapter in books, conference proceedings, editorial letters, case studies and duplication of publication were excluded from this review. Studies using combined preparation of *P. sarmentosum* with other herbs were also excluded.

**ARTICLES SCREENING**

The articles screening process was done by two reviewers. A discussion was conducted to ensure that all reviewers agreed upon all exclusion and inclusion criteria selected. Data extraction processes was done according to PRISMA guideline (Moher et al. 2015). All the duplicate articles among the databases were removed. The first screening was based on exclusion criteria whereby articles published as a review, book, book chapter, editorial letter, conference proceeding or case study were sorted out based solely on their titles and abstracts. Secondly, published articles with lack of information on antioxidant activities related to *P. sarmentosum* were excluded. Finally, the remaining articles were assessed thoroughly for eligibility by checking all inclusion criteria.

**RESULTS AND DISCUSSION**

**DATA EXTRACTION RESULTS**

Overall, a total of 130 potential articles were found from the three electronic databases. Each article was assessed independently by two reviewers based on the inclusion and exclusion criteria. A total of 26 articles were removed due to duplicate between the three databases, leaving behind 104 articles. Forty articles related with reviews, editorial letters, short communication, conference abstract and book chapters were excluded. Another 45 articles did not contain any parameters related to antioxidant activity of *P. sarmentosum*. Thus, a total of 111 articles were sorted out after the first and second screening. The remaining 19 articles were selected for the final analysis and included in this review. A flowchart of the articles selection process is shown in Figure 2.
STUDY DESIGN CHARACTERISTICS

All the final selected studies were characterised in Table 1. These comprised of 19 studies published between years 2003 and March 2018. Ten of the studies focused on PS only while another nine were focused on multiple herbs or plants including PS. Based on the study design, this review consisted of a single study which combined chemical assay, in vitro and in vivo studies, three studies which combined chemical assays and in vitro studies, five studies which combined in vivo studies and chemical assays and ten studies using chemical assays.

Animal studies used Sprague Dawley rats (Hussain et al. 2010), Wistar rats (Azlina et al. 2011; Mohd et al. 2015), Balb/c albino mice (Lee et al. 2011), Duroc-Landrace-Yorkshire piglets (Wang et al. 2017) and New Zealand white rabbits (Amran et al. 2011) as the research models. There were five different cultured cells used for in vitro studies including human umbilical vein endothelial cells (HUVEC) (Hafizah et al. 2010), human lymphocytes cells (HLC) (Wan Ibrahim et al. 2010), murine monocytic macrophages cell line (RAW 264.7) (Lee et al. 2011), immortalised murine microglial cells (BV-2) and human neuroblastoma cells (SH-SY5Y) (Yeo et al. 2018).

Several antioxidant assays such as 1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging assay, ferric reducing antioxidant power (FRAP) assay, hydroxyl radical scavenging (HRS) assay, 8-carotene linoleate (BCL) assay, iron chelating activity (ICA), superoxide scavenging assay, 2,20-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid diammonium salt (ABTS) assay, erythrocyte hemolysis assay (EHA) and bleomycin-dependent DNA damage (BDD) assay were used in the chemical assay studies. Biochemical parameters such as malondialdehyde (MDA), thiobarbituric acid reactive substances (TBARS), cell viability, superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX) activities were also measured as part of the antioxidant activities.

Besides, the contents of antioxidant compounds in the extracts such as total phenolic content (TPC), total flavonoid content (TFC), total amide content (TAC), total polyphenol content (TPP), total carotenoids content (TCC), total vitamin C content (TVC), total vitamin E content (TVE), total tannins content (TTC) and total xanthophylls content (TXC) were also measured. Overall, all 19 studies showed positive antioxidant activities of P. sarmentosum. The characteristics of all studies are summarised in Table 1.

PIPER SARMENTOSUM AS AN ANTIOXIDANT

Out of 19 studies included in this review, one study tested the antioxidant effect of PS shoot (Sulaiman et al. 2011); one study compared the antioxidant effect between PS fruit and leaf (Hussain et al. 2010); one study compared the antioxidant activities between PS root, stem, leaf and fruit (Hussain et al. 2009); while the other sixteen studies used only PS leaves. The fruit of PS had highest antioxidant activities compared to the root, stem and leaf (Hussain et al. 2009). Hussain et al. (2009) also used different types of solvent for extraction of different parts of PS. Ethanol extracts of different parts of PS showed higher antioxidant activity as well as higher amount of polyphenols, flavonoids and amide compared to aqueous extracts of PS (Hussain et al. 2009). In another study, acetone extract of PS had the highest antioxidant activity followed by ethanol, methanol and aqueous extracts (Sulaiman et al. 2011). Whereas other studies found that methanol extract had the highest antioxidant activity followed by hexane, dichloromethane and ethyl acetate extracts (Yeo et al. 2018). Lee et al. (2014) showed that methanol extract of PS had higher antioxidant activity compared to aqueous, hexane, chloroform, ethyl acetate and butanol extracts. The types of solvent used for extraction influence the efficiency of the extraction of polyphenols and subsequently the

FIGURE 2. The selection process of the articles according to PRISMA guideline
<table>
<thead>
<tr>
<th>Study No</th>
<th>Study Design</th>
<th>Plant source</th>
<th>Plant parts</th>
<th>Type of extracts</th>
<th>Results</th>
<th>Outcomes</th>
<th>Ref.</th>
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<tbody>
<tr>
<td>1</td>
<td>Chemical assay and ten weeks <em>in vivo</em> animal study. Forty-two male New Zealand white rabbits (weight 1.8 - 2 kg) randomly divided into seven groups. Antioxidative activity measured using DPPH assay.</td>
<td>Furley Marketing Sdn Bhd, Malaysia</td>
<td>Leaf</td>
<td>Aqueous</td>
<td>↑ DPPH radical scavenging activity with increasing concentration of PS. IC₅₀ value of PS was 27.12 µg/mL</td>
<td>PS via its antioxidant activity reduced inflammatory markers in hypercholesterolemia and atherosclerosis</td>
<td>Amran et al. 2011</td>
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<tr>
<td>2</td>
<td>Chemical assay study. Antioxidative activity measured using DPPH assay.</td>
<td>Furley Marketing Sdn Bhd, Malaysia</td>
<td>Leaf</td>
<td>Aqueous</td>
<td>PS showed antioxidant activities in different types of encapsulation</td>
<td>PS antioxidant activity was maintained in different types of encapsulation</td>
<td>Chan et al. 2010</td>
</tr>
<tr>
<td>3</td>
<td>Chemical assay and <em>in vitro</em> cell culture study. Antioxidative activity measured using FRAP, MDA, cell viability, SOD, GPX and CAT level in H₂O₂-induced HUVEC. TPC was determined.</td>
<td>Forest Research Institute Malaysia</td>
<td>Leaf</td>
<td>Aqueous, methanol and hexane</td>
<td>FRAP value for PS was 18.90 ± 0.02 µmol Fe(II)/g DM which was close to vitamin C (19.28 ± 0.02 µmol Fe(II)/g DM). TPC value of PS was 90.86 ± 0.37 mg GAE/g DM. ↑ Cell viability of H₂O₂-induced HUVEC treated with aqueous, methanol and hexane extracts of PS. ↓ MDA, SOD, CAT and GPX level in H₂O₂-induced HUVEC treated with aqueous, methanol and hexane extracts of PS</td>
<td>PS had potent antioxidant activity and was able to protect endothelial cells from oxidative stress</td>
<td>Hafizah et al. 2010</td>
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<td>4</td>
<td>Chemical assay study. Antioxidative activity measured using BCL and DPPH assays. TPP, TFC and TAC were determined.</td>
<td>Pulau Pinang, Malaysia</td>
<td>Root, stem, leaf and fruit</td>
<td>Aqueous and ethanol</td>
<td>▲ Antioxidant activity, TPP, TFC and TAC in ethanol extracts of different parts of PS compared to aqueous extracts of different parts of PS. ▲ TPP, TFC and TAC in the fruit ethanol extract compared to leaf, stem and root ethanol extracts. Phyto-constituents: Rutin (in steam and leaf ethanol extracts and all parts of aqueous extracts), flavonone (in root and fruit of ethanol and aqueous extracts) and piperine (in all parts of ethanol extracts)</td>
<td>Ethanol extracts of PS have good antioxidant properties. There is a positive correlation between antioxidant activity and polyphenols, flavonoids and amides content in the extracts</td>
<td>Hussain et al. 2009</td>
</tr>
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<td>5</td>
<td>Fourteen days <em>in vivo</em> animal study. 42 male Sprague Dawley rats (weight 220 ± 20 g) randomly divided into seven groups. Antioxidative activity measured by the levels of TPAA, TBARS, CAT and SOD in the liver homogenates.</td>
<td>Pulau Pinang, Malaysia</td>
<td>Fruit and leaf</td>
<td>Ethanol</td>
<td>↑ TPAA, CAT and SOD levels and ↓ TBARS in PS-treated groups compared to carbon tetrachloride (CCl₄)-induced group</td>
<td>Ethanol extracts of fruit and leaf of PS have promising antioxidant activity against CCl₄-induced oxidative stress</td>
<td>Hussain et al. 2010</td>
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| 6        | Chemical assay study.  
Antioxidative activity measured using DPPH assay.  
TPP and TPC were determined | Bangkok, Thailand | Leaf | Aqueous, methanol, hexane, chloroform, ethyl acetate and butanol | Antioxidant activity exhibited by PS methanol extract with EC₅₀ value of 2.70 ± 0.3 mg/ml.  
▲TPC and TPP in methanol extract compared to aqueous, hexane, chloroform, ethyl acetate and butanol extracts of PS | Methanol extract of PS showed higher antioxidant activity compared to aqueous, hexane, chloroform, ethyl acetate and butanol extracts | Lee et al. 2014 |
| 7        | Chemical assay and 28 days in vivo animal study.  
Rats were divided into five groups (normal Wistar rats, spontaneously hypertensive rats and three groups of hypertensive rats treated with PS)  
Antioxidative activity measured using DPPH assay, SOSc assay and serum MDA | Kuantan, Malaysia | Leaf | Aqueous | Antioxidant activity of PS (Kadukmy™) was 96.21 ± 0.88% from DPPH assay and 95.69 ± 0.18% from SOSc assay.  
PS reduced serum MDA in hypertensive rats | PS reduces blood pressure via its antioxidant property | Mohd et al. 2015 |
| 8        | Chemical assay and in vitro cell culture study.  
Antioxidative activity measured using FRAP and DPPH assays.  
TPC was determined | Selangor, Malaysia | Leaf | Aqueous | PS showed antioxidant activity with FRAP value of 394 ± 20.4 umol/g and DPPH radical scavenging effect of 24.3 ± 2.5 %,  
TPC in PS was 430 ± 3.1 mg GAE/g DM.  
Strong positive correlation between TPC with FRAP value and DPPH radical scavenging effect | PS has high antioxidant effect but higher antioxidant activity is associated with greater genotoxic effect in human lymphocytes | Wan Ibrahim et al. 2010 |
| 9        | Chemical assay study.  
Antioxidative activity measured using DPPH and BCL assays.  
TPC was determined | Chiangrai province, Thailand | Leaf | Aqueous | PS showed antioxidant activity with DPPH radical scavenging effect of 46.84 ± 0.30 % and BCL assay of 1.11 ± 0.00.  
TPC was 10.98 ± 0.25 mg GAE/g DW | PS is a promising antioxidant with antihyperglycaemia potential | Wongsa et al. 2012 |
| 10       | Chemical assay study and in vitro cell culture study.  
Antioxidative activity measured using DPPH assays.  
TPC was determined | Puchong, Malaysia | Leaf | Methanol, hexane, ethyl acetate and dichloromethane | ▲DPPH scavenging activity in in methanol extract followed by hexane, dichloromethane and ethyl acetate extracts.  
▲TPC in in dichloromethane extract followed by hexane, methanol and ethyl acetate extracts.  
There was a weak positive correlation between antioxidant activity and TPC | PS confers neuroprotection on beta-amyloid-induced neuroinflammation in SH-SY5Y cells through its antioxidant and anti-inflammatory actions | Yeo et al. 2018 |
| 11       | Chemical assay study.  
Antioxidative activity measured using BCL assay.  
TVC, TVE, TTC, TXC and TPC were determined | Chiang Mai, Thailand | Leaf | Methanol | High antioxidant activity with antioxidant index of 13.0±0.84.  
The antioxidant compound contents were: TVC (16.6±0.06 mg%), TVE (0.01±0.0006 mg%), TTC (3.8±0.06 mg%), TXC (5.81±0.04 mg%), TTC (17.7±0.06 mg%) and TPC (123±0.12 mg%) | Methanol extract of PS showed high antioxidant activity which is correlated to its antioxidant compound contents | Chanwitthesuk et al. 2005 |
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<td>12</td>
<td>Chemical assay study. Antioxidative activity measured using ICA, DPPH assay, HRS assay, SOD activity, MDA, EHA and BDD. TPC was determined</td>
<td>Thailand</td>
<td>Leaf</td>
<td>Methanol</td>
<td>Positive results in all the antioxidative assays. TPC of 6.41 ± 0.14 μmole of catechin-equivalent/mg of dried extract</td>
<td>PS is a natural source of antioxidant</td>
<td>Hutadilok et al. 2006</td>
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<td>13</td>
<td>Chemical assay, <em>in vivo</em> animal study and <em>in vitro</em> cell culture study. Antioxidative activity measured using DPPH assay. TPC was determined</td>
<td>Universiti Putra Malaysia</td>
<td>Leaf</td>
<td>Methanol</td>
<td>↑ DPPH radical scavenging activity with increasing concentration of PS. IC_{50} value of PS was 129.65 ± 1.04 μg/mL. TPC was 50.01 mg GAE/g DW</td>
<td>PS has antioxidant activity</td>
<td>Lee et al. 2011</td>
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<td>14</td>
<td>Chemical assay study. Antioxidative activity measured using SOSC assay</td>
<td>Forest Research Institute Malaysia</td>
<td>Leaf</td>
<td>Aqueous and methanol</td>
<td>▲Antioxidant activity in methanol compared to aqueous extract of PS. Methanol extract of PS contained naringenin</td>
<td>PS had high antioxidant activity and could be used as antioxidant dietary supplement</td>
<td>Subramaniam et al. 2003</td>
</tr>
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<td>15</td>
<td>Chemical assay study. Antioxidative activity measured using FRAP, DPPH and BCL assays. TVC, TFC and TPC were determined</td>
<td>Serdang, Selangor</td>
<td>Leaf</td>
<td>Aqueous and boiled aqueous</td>
<td>▲FRAP, DPPH and BCL activities were found in PS. ▲FRAP value and TFC in aqueous than boiled aqueous extract of PS. ▲DPPH scavenging activity and TPC in boiled aqueous than aqueous extract of PS</td>
<td>PS exhibits antioxidant activity</td>
<td>Sumazian et al. 2010</td>
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<td>16</td>
<td>Four weeks in vivo animal study. Eighty 21-day-old weaned piglets were randomly divided into four groups. Antioxidative activity measured using serum MDA and GPX activity</td>
<td>Hainan, China</td>
<td>Leaf</td>
<td>Supercritical carbon dioxide</td>
<td>▲GPX activity and ▼MDA in piglets supplemented with PS compared to control</td>
<td>PS supplementation improved antioxidant capability and reduced inflammation</td>
<td>Wang et al. 2017</td>
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<td>Study No</td>
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<td>17</td>
<td>Twenty-eight days <em>in vivo</em> animal study Thirty-two male Wistar rats were randomly divided into four groups. Antioxidative activity measured using plasma TBARS and lung TBARS, GPX and SOD activity</td>
<td>Forest Research Institute Malaysia</td>
<td>Leaf</td>
<td>Methanol</td>
<td>↓ Lung TBARS and GPX activity in rats treated with PS compared to CCl₄-induced group. No change in plasma TBARS and lung SOD activity</td>
<td>PS possesses good antioxidant property and capable to reduce oxidative stress</td>
<td>Azlina et al. 2011</td>
</tr>
<tr>
<td>18</td>
<td>Chemical assay study. Antioxidative activity measured using FRAP, ABTS assay and ICA. TFC and TPC were determined</td>
<td>Mahidol University, Thailand</td>
<td>Leaf</td>
<td>Aqueous</td>
<td>Positive results in FRAP, ABTS assay and ICA. Polyphenolics, especially flavonoids determined the antioxidant capacity.</td>
<td>PS has antioxidant activities</td>
<td>Deetae et al. 2012</td>
</tr>
<tr>
<td>19</td>
<td>Chemical assay study. Antioxidative activity measured using FRAP and DPPH assay. TPC, TFC and TPC were determined</td>
<td>Pulau Pinang, Malaysia</td>
<td>Shoot</td>
<td>Acetone, ethanol, methanol and aqueous</td>
<td>▲FRAP and DPPH scavenging activity in acetone extract followed by ethanol, methanol and aqueous extracts ▲TPC in acetone extract followed by aqueous, ethanol and methanol extracts ▲TFC in ethanol extract followed by acetone, aqueous and methanol extracts</td>
<td>PS antioxidant activities varied depending on the type of solvent used during extraction</td>
<td>Sulaiman et al. 2011</td>
</tr>
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</table>

Footnote: ↑ = Increased, ↓ = Decreased, ▲= Highest, ▼= Lowest. *Piper sarmentosum* (PS), 1-diphenyl-2-picrylhydrazyl (DPPH), Superoxide scavenging (SOSc), Superoxide dismutase (SOD), Ferric-reducing antioxidant power (FRAP), Malondialdehyde (MDA), Thiobarbituric acid reactive substances (TBARS), Total phenolic content (TPC), Catalase (CAT), Glutathione peroxidase (GPX), Total flavonoid content (TFC), Total amide content (TAC), ß-carotene linoleate (BCL), Total polyphenol content (TPP), Total carotenoids content (TCC), Total vitamin C content (TVC), Total vitamin E content (TVE), Total tannins content (TTC), Total xanthophylls content (TXC), Total plasma antioxidant activity (TPAA), Hydroxyl radical scavenging (HRS), Erythrocyte hemolysis assay (EHA), Bloomsyin-dependent DNA damage (BDD), Iron chelation activity (ICA), 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid diammonium salt (ABTS), Dried matter (DM), Gallic acid equivalent (GAE), Inhibition concentration at 50 % (IC₅₀), Hydrogen peroxide (H₂O₂), Carbon tetrachloride (CCl₄), Effective concentration 50% (EC₅₀).
As listed in Table 1, PS samples used for the antioxidant analyses originated from Malaysia, Thailand and China but mostly were from Malaysia. Twelve out of 19 studies used DPPH radical scavenging assay to determine the antioxidant activity of PS. DPPH radical may be neutralised by direct reduction via electron transfer or by radical quenching via hydrogen atom transfer from the antioxidants (Marxen et al. 2007). The next common antioxidant assay used in the studies was FRAP assay. FRAP assay measures the total antioxidant capacity by calculating the reducing power of vitamin C's FRAP value of 18.90 mol Fe(II)/g DM which was comparable with vitamin C’s FRAP value of 19.28 mol Fe(II)/g DM (Hafizah et al. 2010). The results of other antioxidant assays used in the studies such as BCL, ABTS, ICA, EHA, HRS and BDD assays followed the similar trend of DPPH and FRAP assays. There is a positive correlation between antioxidant activity and the content of antioxidant compounds in the extracts. Antioxidant activities as measured by DPPH and BCL assays had positive correlation with polyphenols, flavonoids and amides content of PS (Hussain et al. 2009). Whereas FRAP and DPPH assay results were positively affected by total phenolic content of PS (Wan Ibrahim et al. 2010; Yeo et al. 2010). This is in accordance with a previous study that also reported positive correlation between total polyphenols and total flavonoids content with antioxidant effect measured by DPPH assay (Lee et al. 2014).

TBARS and MDA were commonly analysed as markers of lipid peroxidation and oxidative stress (Del Rio et al. 2005). Five of the studies showed that PS was capable to reduce MDA and/or TBARS (Azlina et al. 2011; Hafizah et al. 2010; Hussain et al. 2010; Mohd et al. 2015; Wang et al. 2017). This shows that PS is protective against oxidative stress. In a study done by Mohd et al. (2015), high antioxidant activity of the extract as measured by DPPH assay contributed to its blood pressure-lowering effect as well as reduction of MDA. The antioxidant enzymes; SOD, CAT and GPX represent a first line of defense against reactive oxygen species (ROS) by metabolising them to innocuous byproducts (Ugusman et al. 2011). There are variations among the effects of PS on antioxidant enzymes. PS was reported to increase SOD, CAT and GPX level (Hussain et al. 2010; Wang et al. 2017). Some studies reported that PS reduced SOD, CAT and GPX levels (Azlina et al. 2011; Hafizah et al. 2010). The increased antioxidant enzymes may be attributed to an instant active role of the enzymes in neutralising the ROS. The decreased antioxidant enzyme could be due to the antioxidant compounds in PS that initially played a role in neutralising the ROS, thus reducing the need for enhanced antioxidant enzyme level (Ashokkumar & Sudhandiran 2008; Hafizah et al. 2010).

Flavonoids are a group of polyphenols that can be found naturally in plants. PS contains flavonoids such as myricetin, quercetin, apigenin (Miean & Mohamed 2001), naringenin (Subramaniam et al. 2003), rutin and vitexin (Ugusman et al. 2012) that have been proven to have good antioxidant activities. Increased intake of dietary flavonoids is associated with a decrease in the risk of coronary artery disease by 65% (Arts & Hollman 2005). Besides, a meta-analysis of prospective cohort studies showed that higher dietary flavonoid intake significantly reduced the risk of stroke (Tang et al. 2016). Rutin and quercetin are more effective than nifedipine in reducing blood pressure and improving oxidative stress markers in high salt diet-induced hypertensive rats (Olalayye et al. 2013). Quercetin has consistent blood pressure-lowering effect in animal and human studies irrespective of the dose, duration or disease status (Clark et al. 2015). Naringenin; a flavonoid present in PS, decreased blood sugar level and oxidative stress markers in diabetic rats (Ren et al. 2016). All 19 studies in this review confirmed the antioxidant effect of PS. However, there was no clinical study carried out to investigate the antioxidant effect of PS in human. Therefore, it is recommended that further studies on the antioxidant activities of PS should be carried out in clinical settings. Apart from this, more studies are also needed to identify the specific phytochemical compound responsible for the antioxidant effect.

CONCLUSION

The review concluded that PS has antioxidant effect and its bioactive compounds such as polyphenols and flavonoids contribute to the antioxidant activities. Thus, PS has the potential to be developed as an alternative treatment for diseases related to oxidative stress such as hypertension, diabetes mellitus and coronary artery disease. A clinical placebo-controlled study may be needed before employing PS as an effective antioxidant.

ACKNOWLEDGEMENTS

This study was supported by the Fundamental Research Grant Scheme, Ministry of Higher Education, Malaysia (FRGS/1/2014/UKM/02/1) and Universiti Kebangsaan Malaysia Medical Centre Fundamental Grant (FF-2017-040). Miss Siti Marianna Ismail receives Ph.D scholarship from the University of Malaya and Ministry of Higher Education, Malaysia.

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Received: 31 March 2018
Accepted: 5 June 2018