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## **Plant budding speciation predominant by ecological and geographical differentiation: an ‘evolutionary snapshot’ in Iberodes (Boraginaceae) — Source link**

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1 **A snapshot of progenitor-derivative speciation in action in *Iberodes* (Boraginaceae)**

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## 15 SUMMARY

- 16 • Traditional classification of speciation modes has focused on physical barriers to  
17 gene flow. While allopatry has been viewed as the most common mechanism of  
18 speciation, parapatry and sympatry, both entail speciation in the face of ongoing  
19 gene flow and thus both are far more difficult to detect and demonstrate.  
20 *Iberodes* (Boraginaceae, NW Europe) with a small number of recently derived  
21 species (five) and contrasting morphological traits, habitats and distribution  
22 patterns constitutes an ideal system in which to study drivers of lineage  
23 divergence and differentiation.
- 24 • To reconstruct the evolutionary history of the genus, we undertook an  
25 integrative study entailing: (i) phylogenomics based on restriction-site  
26 associated DNA sequencing (RAD-seq), (ii) morphometrics, and (iii) climatic  
27 niche modelling.
- 28 • Key results revealed a history of repeated progenitor-derivative speciation,  
29 manifesting in paraphyletic pattern within *Iberodes*. Climatic niche analyses,  
30 together with the morphometric data and species distributions, suggest that  
31 ecological and geographical differentiation have interacted to shape the diversity  
32 of allopatric and parapatric distributions observed in *Iberodes*.
- 33 • Our integrative study has enabled to overcome previous barriers to  
34 understanding parapatric speciation by demonstrating the recurrence of  
35 progenitor-derivative speciation in plants with gene flow and ecological  
36 differentiation, explaining observed parapatry and paraphyly.

37

38 **Keywords:** Budding speciation, ecological speciation, restriction-site associated DNA  
39 sequencing (RAD-seq), molecular dating, species distribution modelling, niche overlap,  
40 paraphyly, parapatry.

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## 47 INTRODUCTION

48 Understanding what modes of speciation dominate diverse clades across the tree of life  
49 has been a contentious domain of evolutionary biology for decades (e.g. Templeton,  
50 1981; Rieseberg & Brouillet, 1994; Butlin *et al.*, 2008; Horandl & Stuessy, 2010).  
51 Reproductive barriers are the most frequently used criteria for distinguishing among  
52 three overarching modes of speciation (Futuyma, 2009). Speciation of adjacent  
53 populations in the absence of clear physical or reproductive barriers is frequently  
54 explained under the process of (1) parapatric speciation, which entails the evolution of  
55 reproductive isolation in the face of limited but ongoing gene flow (Coyne & Orr, 2004,  
56 p. 112). Parapatric speciation can be considered intermediate between allopatric and  
57 sympatric speciation. In (2) allopatric speciation, gene flow is prevented by geographic  
58 barriers between populations. In (3) sympatric speciation, partial or complete  
59 reproductive isolation arises between subsets of a single population with overlapping  
60 ranges, without spatial segregation. Sympatric speciation often entails significant levels  
61 of gene flow at the early stages of differentiation (Gavrilets, 2003; Futuyma, 2009).  
62 Whereas allopatry is widely viewed to be the most common mode of speciation (Coyne  
63 & Orr, 2004; Vargas *et al.*, 2018), the relative importance of parapatry and sympatry has  
64 been the subject of considerable debate (Fitzpatrick *et al.*, 2008; Fitzpatrick *et al.*, 2009;  
65 Arnold, 2015) in large part due to the difficulty of (1) resolving phylogenetic  
66 relationships among very closely related populations that may still be exchanging genes  
67 at a low level, and (2) quantifying the amount of gene flow between populations during  
68 population divergence (Barluenga *et al.*, 2006; Fontdevilla, 2014). In the last decade,  
69 new genomic, phylogenetic, and ecological tools have made non-allopatric modes of  
70 speciation increasingly amenable to study, allowing us to explore with statistical rigor a  
71 wider range of speciation scenarios (e.g. Savolainen *et al.*, 2006; Chozas *et al.*, 2017;  
72 Zheng *et al.*, 2017). In particular, there are some critical points of evidence that together  
73 provide support for a parapatric speciation scenario: (1) phylogenetic relationships  
74 usually reflect a progenitor-derivative (budding) pattern in which the more widely  
75 distributed species is the living progenitor of the more restricted one; (2) the absence of  
76 any physical barrier to gene-flow in both present and past times, i.e. distribution ranges  
77 are adjacent; (3) interspecific genetic differentiation that putatively discards processes  
78 of secondary contact; (4) alternative forces of reproductive isolation factors—e.g.,  
79 differential climatic niches, reproductive exclusion, ploidy variation, among others—are  
80 present.

81 Ecological differentiation is often associated with non-allopatric speciation (Sobel  
82 *et al.*, 2010; Nosil, 2012; Stankowsky *et al.*, 2015), particularly in plants, because a  
83 sessile habit makes plants more sensitive to fine-scale environmental heterogeneity  
84 (Anacker & Strauss, 2014). Both the influence of niche conservatism (i.e. retention of  
85 ancestral ecological characteristics of species over time, Peterson *et al.*, 1999) and niche  
86 evolution (i.e. adaptation of lineages to changes in the environment, Donoghue &  
87 Edwards, 2014) have been argued to play a role in speciation and lineage diversification  
88 (Wiens & Graham, 2005; Cavender-Bares, 2019). Nevertheless, the contribution of  
89 ecological factors as primary drivers of speciation remains more elusive (Mayr, 1954;  
90 Pyron & Burbrink, 2010; Yin *et al.*, 2016).

91 The advent of inexpensive phylogenomic approaches in the last decade have made  
92 the genetic side of testing recent speciation scenarios tractable (McCormack *et al.*,  
93 2013; McVay *et al.*, 2017a). High Throughput Sequencing (HTS) approaches make it  
94 possible to analyze loci sampled from the entire genome in reconstructing species trees  
95 (Fernández-Mazuecos *et al.*, 2018) and the history of speciation in the face of gene flow  
96 (Leroy *et al.*, 2017; Folk *et al.*, 2018; Crowl *et al.*, 2019). Restriction-site associated  
97 DNA sequencing (RAD-seq) has contributed particularly strongly to our understanding  
98 of recent evolutionary processes, especially in non-model organisms, since a reference  
99 genome is not needed for accurate phylogenetic inference (e.g. Fitz-Gibbon *et al.*,  
100 2017). RAD-seq has as a consequence been useful in inferring complex speciation  
101 histories: repeated cycles of island connectivity and isolation (the pump hypothesis;  
102 Papadopoulou & Knowles, 2015), incipient sympatric speciation (Kautt *et al.*, 2016),  
103 allopatric speciation despite historical gene flow (Maguilla *et al.*, 2017), ancient  
104 introgression among now-extinct species (McVay *et al.*, 2017b).

105 The Mediterranean subendemic genus *Iberodes* M.Serrano, R.Carbajal & S.Ortiz  
106 (Boraginaceae, Cynoglossoidae; Serrano *et al.*, 2016) provides an excellent system for  
107 studying recent speciation, morphological and ecological differentiation because it is  
108 clearly demonstrated to be a recently-derived genus (Chacón *et al.*, 2017; Otero *et al.*,  
109 2019a). *Iberodes* comprises five species and one subspecies that differ in geographic  
110 ranges (including allopatric and parapatric species) and habitat (ranging from forest  
111 understories to scrublands and coastal dunes). Moreover, all species are endangered and  
112 narrowly endemic, except for the widely-distributed *I. linifolia* of the Iberian Peninsula  
113 and southeastern France (Talavera *et al.*, 2012). Sanger sequencing has failed to resolve  
114 phylogenetic relationships among *Iberodes* species because the markers used to date

115 provide minimal variability (Otero *et al.*, 2014; Holstein *et al.*, 2016a; Holstein *et al.*,  
116 2016b; Otero *et al.*, 2019a). The question consequently remains as to what is the relative  
117 importance of extrinsic geographical and ecological barriers to the complex speciation  
118 patterns observed in *Iberodes*. Given the narrow endemism in most *Iberodes* species,  
119 we hypothesize that geographic barriers rather than ecological factors have been  
120 predominant in the speciation history of *Iberodes*. To test this hypothesis, we aim to: (1)  
121 reconstruct phylogenetic relationships of the five species; (2) infer speciation patterns  
122 and lineage diversification during the last geological epochs comparing diverse  
123 morphologies and ecologies; and (3) evaluate the relative contribution of geographical  
124 and ecological processes in speciation.

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## 126 **MATERIAL AND METHODS**

### 127 **Taxon sampling**

128 Based on species distributions, between one and four individuals from one to nine  
129 populations were sampled for each of the five species of *Iberodes* including ssp.  
130 *gallaecica* and ssp. *littoralis* of *I. littoralis*) (Fig. 1, Table S1, in Supporting  
131 Information). Samples of *I. littoralis*, *I. kuzinskyana* and *I. brassicifolia* were collected  
132 in the field during 2015 and stored in silica gel until extraction. Since these species are  
133 considered threatened according to IUCN criteria (Lopes & Carvalho, 1990; Serrano &  
134 Carbajal, 2003; ICNB, 2007; Moreno, 2008; IUCN, 2019; INPN, 2019), permits were  
135 obtained prior to collecting. Mature seeds of *I. commutata* were collected in 2015 and  
136 germinated in a glasshouse to obtain green leaves for DNA extraction. Both field and  
137 herbarium materials were used to represent the large distribution of *I. linifolia* (Table  
138 S1). Three species (17 individuals) of the tribe Omphalodeae were included as the  
139 outgroup (Otero *et al.*, 2019b): *Omphalodes nitida*, *Myosotidium hortensia*, and  
140 *Gyrocaryum oppositifolium* (Table S1).

### 141 **DNA extraction and RAD library preparation**

142 DNA was extracted from leaf tissue using a modified CTAB protocol from Doyle and  
143 Doyle (1987) and Shepherd and Mc Lay (2011), including a precipitation step in  
144 isopropanol and ammonium acetate 7.5M at -20°C overnight to improve yield. DNA  
145 extractions were visualized and quantified with NanoDrop 2000/2000c (Thermo-Fisher,  
146 Waltham, Massachusetts) and Qubit Fluorometric Quantification (Thermo-Fisher,  
147 Waltham, Massachusetts) in the *Instituto de Investigaciones Biomédicas* (IIBm, CSIC-  
148 UAM). Final DNA sample concentrations were standardized to 10 ng/ul. Preparation of

149 single-end RAD-seq libraries using restriction enzyme *Pst*I from genomic DNA was  
150 conducted at Floragenex Inc. (Eugene, Oregon) following Baird *et al.* (2008) and  
151 sequenced as single-end, 100-bp reactions on an Illumina HiSeq 2500 at the University  
152 of Oregon Genomics & Cells Characterization Core Facility (library C606). Processed  
153 data were returned in the Illumina 1.3 variant of the FASTQ format, with Phred quality  
154 scores for all bases.

### 155 **Data clustering**

156 Sequences from Illumina were analyzed in *PyRAD* v. 3.0.6, a *de novo* clustering  
157 pipeline with seven main steps that filter and cluster putatively orthologous loci from  
158 RAD sequencing reads (Eaton, 2014). This pipeline is able to cluster highly divergent  
159 sequences, taking into account indel variation and nucleotide (SNP) variation. It is thus  
160 well-suited to the inter- and intraspecific scales addressed in our study. As no evidence  
161 of polyploidy was found from flow cytometry, we did not assess sensitivity of paralog  
162 detection to sequencing depth or number of heterozygotic positions allowed per loci or  
163 shared among individuals. Base calls with a Phred Quality score <20 were replaced by  
164 the ambiguous base code (N). Three similarity percentages were set for the clustering  
165 (c) threshold of reads: 85%, 90% and 95%. Four levels of minimum coverage (m) of a  
166 sample in a final locus (4, 12, 20, 28) were tested for each of the three percentages of  
167 clustering similarity, resulting in a fully factorial set of 12 matrices from the  
168 combination of filtering parameters, so we could assess sensitivity of our phylogenetic  
169 inferences to clustering assumptions. Consensus base calls were made for clusters with  
170 a minimum depth of coverage >5. From each parameter combination we obtained three  
171 kinds of output: (1) a matrix with all loci retrieved for each individual (concatenated  
172 matrix), (2) a matrix with single nucleotide polymorphisms (SNP matrix), and (3) a  
173 matrix with one SNP per locus (unlinked SNP matrix). The loci matrix was explored  
174 with the RADami (Hipp, 2014) and vegan (Oksanen *et al.*, 2013) packages of R version  
175 3.3.2 (R Core Team, 2013).

### 176 **Phylogenetic analysis**

177 Maximum likelihood analyses of concatenated matrices obtained in *PyRAD* were  
178 performed in RAxML v8.2.10 (Stamatakis, 2014) through the Cipres Portal (Miller *et al.*,  
179 2010). We used rapid bootstrapping, the GTRGAMMA model, and automatic-stop  
180 bootstrapping with the majority rule criterion. We conducted a maximum likelihood  
181 analysis for each of the 12 *PyRAD* concatenated matrices. Finally, we compared the  
182 topologies based on average bootstrap support (BS). Between the two topologies with

183 average BS>98%, we chose the one maximizing the number of loci (HBS tree) for the  
184 remaining phylogenetic-based analyses. Moreover, we used the unlinked SNP matrix  
185 from the HBS tree (HBS matrix) to perform a coalescent-based phylogeny using  
186 SVDquartets (Chifman & Kubatko, 2014) in PAUP (Swofford, 2001) through the  
187 Cipres portal (Miller *et al.*, 2010). For SVDquartets we grouped individuals based on  
188 main clades obtained from concatenated analysis in RAxML and evaluated all possible  
189 quartets with 100 bootstrap replicates.

#### 190 **Estimates of divergence times**

191 We used penalized likelihood as implemented in TreePL (Smith & O'Meara, 2012) to  
192 estimate a time-calibrated tree for all bootstrap trees (with branch lengths) obtained in  
193 RAxML from the HBS tree. TreePL is suitable for divergence time estimation when  
194 dealing with large amounts of data, such as those yielded by high throughput  
195 sequencing RAD-sequencing (Zheng & Wiens, 2015). Two calibration points were  
196 used: (1) tribe Omphalodeae (root) (minimum age (minAge) = 9.062; maximum age  
197 (maxAge) = 24.4097) and (2) clade *Iberodes* (ingroup) (minAge = 0.6591; maxAge =  
198 3.7132) based on the averaged ages and standard deviation inferred in Otero *et al.*  
199 (2019a). While secondary calibrations tend to underestimate the uncertainty of the age  
200 estimates in the trees from which they are derived, use of multiple secondary  
201 calibrations with the full range of uncertainty found in the original study have the  
202 potential to recover more accurate clade ages (Schenk, 2016). Average node ages over  
203 all posterior trees from the latter study were calculated using *treeStat* from the BEAST  
204 v 1.8.2 package (Drummond *et al.*, 2012). We first conducted an analysis under the  
205 “prime” option to select the optimal set of parameter values, using random subsample  
206 and replicate cross-validation (RSRCV) to identify the best value for the smoothing  
207 parameter, lambda. For this first analysis we used the thorough analysis option and set  
208 200,000 iterations for penalized likelihood and 5,000 iterations for cross validation. We  
209 then repeated the same thorough analysis and iterations but setting the smoothing value  
210 of lambda to 10, gradient based (opt) and auto-differentiation based optimizers (optad)  
211 to five, and auto-differentiation cross-validation-based optimizers to two (optcvad).  
212 Although TreePL does not draw a prior distribution for the phylogenetic tree, we took  
213 into account the bootstrapped variance in topology and branch length by running the  
214 same TreePL analysis for each bootstrap tree and then a maximum clade credibility tree  
215 using the mean heights was reconstructed in *TreeAnnotator* from the BEAST v. 1.8.2  
216 package (Drummond *et al.*, 2012). Our dating analysis thus accounts for the uncertainty



217 in branch length that is due to variance in molecular substitution process across the  
218 RAD-seq loci used.

### 219 **Species-level analyses**

220 Phylogenetic inferences showed two cases of paraphyly within the lineage of *I. linifolia*  
221 that involved three other taxa (*I. kuzinskyana*, *I. littoralis* ssp. *littoralis* and ssp.  
222 *gallaecica*; see results). To evaluate these cases of paraphyly, a set of analyses were  
223 done to analyze the degree of genetic, morphological and ecological differentiation  
224 among the four taxa involved. Therefore, the following analyses were performed on a  
225 subset of data including only these four taxa.

### 226 **Genetic structure—**

227 The HBS matrix was used to perform a Discriminant Analysis of Principal Components  
228 (DAPC) to identify and describe clusters of genetically related individuals (Jombart *et*  
229 *al.*, 2010). The format of the HBS matrix was converted from *vcf* to *genlight* with *vcfR*  
230 (Knaus & Grünwald, 2017). DAPC analysis was performed in *adegenet* (Jombart,  
231 2008). We fixed the number of clusters to four, based on the natural groups of four  
232 taxonomic species and subspecies (the Linifolia clade, see results). We used the  
233 function ‘optim.a.score’ to estimate the optimal number of principal components for  
234 discriminant analysis. We represented genetic differentiation using scatterplots of the  
235 principal components and a barplot representing assignment probabilities of individuals  
236 to groups (*compoplot*). In addition, analysis of molecular variance (AMOVA) was  
237 performed in *Arlequin* v. 3.5.2.1 (Excoffier & Lischer, 2010). AMOVA was run to  
238 evaluate the level of genetic differentiation of the paraphyletic *I. linifolia* as a whole  
239 with respect to the other two species (*I. kuzinskyana* and *I. littoralis*) of the Linifolia  
240 clade. Therefore, we considered only two groups, *I. linifolia* on one hand, and the two  
241 coastal species together (*I. kuzinskyana* and *I. littoralis*) on the other. Finally, to  
242 evaluate a potential role of historical introgression in determining paraphyly, we  
243 conducted D-statistic tests in *PyRAD* (see Methods S1 for details).

### 244 **Morphometric evaluation—**

245 We performed a morphometric exploration using principal component analysis (PCA).  
246 We consulted the taxonomic literature to identify diagnostic characters in *Iberodes*  
247 (Talavera *et al.*, 2012). We measured 12 diagnostic macromorphological vegetative and  
248 reproductive characters for 111 individuals from 58 vouchers (two different individuals  
249 per voucher except for 5 vouchers with only one individual). We averaged measures  
250 from each two individuals of the same voucher. As a result, we obtained a final matrix

251 of 58 accessions from 47 locations (74 individuals from 35 locations of *I. linifolia*, 14  
252 from five locations of *I. kuzinskyana*, and 23 from seven locations of *I. littoralis*  
253 (including the two subspecies) (Fig. 1, Table S2). Ten morphological characters were  
254 quantitatively continuous: four vegetative characters (length of stem, leaves and  
255 pedicels, and width of leaves) and six reproductive characters (length of fruiting calix,  
256 width and length of nutlet, length of nutlet margin, length of the nutlet margin teeth, and  
257 hair length). In addition, one binomial character (presence or absence of bracts) and one  
258 quantitative, discrete character (hairs per mm<sup>2</sup> on the abaxial surface of the nutlet) were  
259 also considered.

260 PCA was run using the function ‘pcomp’ from Stats R package (R Core Team,  
261 2013). Results were visualized with the function ‘ggbiplot’ in the ggbiplot R package  
262 (Vu, 2011).

### 263 ***Climatic niche differentiation and distribution modelling—***

264 Occurrences of the four taxa were collected from GBIF (<https://www.gbif.org/>). We  
265 filtered the resulting dataset by removing points suspected to be incorrect, such as those  
266 placed clearly outside of the known species distribution range (based on taxonomic  
267 literature and databases; Fernández & Talavera, 2012; Muséum National d’Histoire  
268 Naturelle, 2003-2019). Additional geographic coordinates were obtained from  
269 herbarium material and our own fieldwork data. We also estimated the geographic  
270 coordinates of non-georeferenced herbarium specimens, whenever the locality provided  
271 could be pinpointed with an accuracy of  $\pm 5$  km (Table S3, Fig. 1). To reduce sampling  
272 bias, we filtered the dataset by randomly removing points that were within 0.2° latitude  
273 or longitude of each other for *I. linifolia* and 0.1° for *I. littoralis* ssp. *littoralis* and ssp.  
274 *gallaecica*. Filtering was not applied for *I. kuzinskyana* because of the low number of  
275 occurrences. The final dataset after filtering included 115 occurrences of *I. linifolia*, 11  
276 of *I. kuzinskyana*, 16 of *I. littoralis* ssp. *littoralis* and 11 of ssp. *gallaecica*.

277 We obtained 19 bioclimatic variables (resolution of 30”) from WorldClim 1.4  
278 (<http://www.worldclim.org/>; Hijmans *et al.*, 2006) for the study region (34° to 50° N;  
279 11° W to 7° E). To avoid collinearity among bioclimatic variables, we first excluded  
280 variables displaying a high correlation coefficient with other variables ( $|r| > 0.7$ ) in the  
281 study area (Dormann *et al.*, 2013). Then, we calculated the variance inflation factor  
282 (VIF) for each remaining variable using the HH package in R (Heiberger, 2017) and  
283 selected variables with VIF <5 following Benítez-Benítez *et al.* (2018). As a result, the  
284 following six variables were selected: bio1, bio3, bio4, bio8, bio12 and bio14. We

285 assessed climatic niche overlap among the four taxa by pairwise comparisons. The E-  
286 space (i.e. environmental space resulting from the six climatic variables) of each taxon  
287 was analyzed in a Principal Component Analysis (PCA) as implemented in the R  
288 package *ecospat* (Di Cola *et al.*, 2017). To visualize niche differences along each of the  
289 first two principal components, the result was plotted using the *niceOverPlot* function  
290 (Fernández-López & Villa-Machío, 2017), which relies on the *ggplot2* package  
291 (Wickham, 2016). We calculated values of Schoener's index (D) as a measure of niche  
292 overlap (Schoener, 1968; Warren *et al.*, 2008). Then, we conducted tests of niche  
293 equivalency and niche similarity using *ecospat* (Di Cola *et al.*, 2017). The niche  
294 equivalency test evaluates whether the observed niche overlap is significantly different  
295 from a null simulated by randomly reallocating the occurrences of both entities between  
296 their ranges (Broennimann *et al.*, 2012; Warren *et al.*, 2008). The niche similarity test  
297 checks whether the overlap between two niches is significantly different from the  
298 overlap obtained if random shifts within each environmental space are allowed  
299 (Schoener, 1968; Warren *et al.*, 2008). In both cases, we tested for niche divergence by  
300 using the `alternative="lower"` option. All tests were based on 100 iterations. In  
301 similarity tests, both ranges were randomly shifted (`rand.type=1`).

302 In addition, we modeled the potential range of the four taxa of the Linifolia clade  
303 under present conditions, which was then projected to last interglacial (LIG, c. 120–140  
304 kya) and last glacial maximum (LGM, c. 21 kya) conditions. To this end, we used the  
305 same six WorldClim bioclimatic variables employed for niche differentiation analyses,  
306 including present layers, LIG layers from Otto-Bliesner *et al.* (2006) and LGM layers  
307 based on the CMIP5 project (three global climate models (GCMs): CCSM4, MIROC-  
308 ESM and MPI-ESM-P). Resolution was 30" for current and LIG, and 2.5' for LGM  
309 layers. We performed species distribution modeling (SDM) using the maximum entropy  
310 algorithm, as implemented in Maxent v3.4 (Phillips *et al.*, 2006). We used the same  
311 occurrences for each taxon employed for niche differentiation analyses (see above).  
312 80% of occurrences for each taxon were used for model training and 20% for model  
313 evaluation. For each taxon, ten subsample replicates were run by randomly partitioning  
314 the data into a training set and evaluation set, and a mean model was calculated. For the  
315 LGM, an average of the three GCMs was calculated. Logistic outputs were converted  
316 into presence/absence using the maximum training sensitivity plus specificity logistic  
317 threshold.

318 ***Ploidy level estimation—***

319 Flow cytometry was used to estimate genome size, explore the possible role of  
320 polyploidization in speciation, and assess the risk of clustering paralogs in RAD-seq *de*  
321 *novo* clustering. Earlier cytogenetic studies for three *Iberodes* species showed them to  
322 be diploids ( $2n=28$  for *I. linifolia*, *I. commutata* and *I. kuzinskyana* (Franco, 1984; Saly,  
323 1997; Talavera *et al.*, 2012), and aneuploidy was inferred for *I. littoralis* based on its  
324 chromosome count of  $2n=24$  (Fernández-Casas, 1975). Nevertheless, some of these  
325 references do not cite the plant sources, making it difficult to assess the connection of  
326 cytotypes to populations. Individuals of the five *Iberodes* species were cultivated at the  
327 glasshouse and five individuals per species were sampled. In addition, samples of the  
328 diploid *Solanum lycopersicum* of known genome size ( $2n=24$ ,  $2C=1.88-2.07$  pg,  
329 Grandillo *et al.*, 2011) were also cultivated to compare standardized results with our  
330 study species and estimate genome sizes (Doležel & Bartoš, 2005). We performed a  
331 two-step nuclear DNA Content Analysis using Partec Buffer (de Laat *et al.*, 1987) and  
332 DAPI fluorochrome through a Cell Lab Quanta SC flow cytometer (Beckman Coulter,  
333 Fullerton, CA, USA) equipped with a mercury lamp following the protocol of Doležel  
334 *et al.* (2007). Samples were visualized through the Cell Lab Quanta SC Software  
335 package. FL1 detector (530 nm / 28 nm; DAPI emission maximum=461) and FL1-Area  
336 was used as directly correlated to DNA content. A minimum number of 1000 nuclei for  
337 G1 peak were analyzed. Only histograms with a coefficient of variation (CV) lower than  
338 10% for the G1 peak were accepted.

339

## 340 **RESULTS**

### 341 **Phylogenetic analysis**

342 After filtering and processing all samples under the different parameter combinations in  
343 *PyRAD*, 12 matrices of 54 taxa each but varying in numbers of loci and SNPs were  
344 analyzed (see Table S4). The same topology was obtained in RAxML from each of the  
345 12 matrices, differing only in bootstrap support (BS) values for the nodes (Fig. S1). The  
346 highest BS supports and highest number of loci were obtained for the c95m20 matrix  
347 (95% similarity within clusters and minimum coverage of 20 samples for a final locus).  
348 Consequently, this matrix was used for the remaining phylogenetic analyses (HBS  
349 matrix). *Iberodes brassicifolia* was inferred to be the earliest-diverging lineage, sister to  
350 a clade that contains *I. commutata* sister to another subclade with the remaining species  
351 (Fig. 2). Paraphyly was inferred for *I. linifolia* since a clade including *I. kuzinskyana*

352 and *I. littoralis* is nested within. Likewise, paraphyly was inferred for *I. littoralis* ssp.  
353 *gallaecica* since ssp. *littoralis* is nested within populations of subsp. *gallaecica* (Fig. 2).

354 Taxon groups for coalescent-based phylogenetic inference (SVDquartets) were  
355 based on the eight main lineages recovered using RAxML (*Iberodes brassicifolia*, *I.*  
356 *commutata*, populations of *I. linifolia* from Seville, core clade of *I. linifolia*,  
357 populations of *I. linifolia* from southern Spain, populations of *I. linifolia* from central  
358 Portugal, *I. kusinskyana*, and *I. littoralis*, Fig. S2). The species tree topology was mostly  
359 congruent with the concatenated phylogenetic topology. The only difference was the  
360 placement of *I. linifolia* populations from Seville and southern Spain, which were  
361 nested within the core of *I. linifolia* (Fig. S2). In any case, paraphyly of *I. linifolia* is  
362 still inferred because of the nested position of *I. kusinskyana* and *I. littoralis* sister to  
363 populations of *I. linifolia* from Portugal (Fig. S2).

#### 364 **Divergence time estimates**

365 The stem age estimated for *Iberodes* was 17.56 myr (17.26-17.86 Bootstrapped  
366 Variance, BV) with a crown age of 5.32 myr (5.32-5.32 BV) (Fig. 2). Crown age  
367 estimates for *I. brassicifolia* and *I. commutata* were 1.29 (1.21-1.33 BV) and 2.22 myr  
368 (2.17-2.28 BV), respectively. The crown age of the Linifolia clade (*I. linifolia* s.l., *I.*  
369 *kusinskyana* and *I. littoralis*) was inferred to be 4.33 myr (4.25-4.59 BV). Divergence  
370 time between Portuguese populations of *I. linifolia* and ancestor of the coastal species  
371 (*I. kusinskyana* and *I. littoralis*) was estimated at 3.67 myr (3.60-3.85 HPD). The origin  
372 of *I. littoralis* ssp. *littoralis*, possibly from an *I. littoralis* ssp. *gallaecica* ancestor, was  
373 inferred to have taken place ca. 10,000 years ago (Fig. 2).

#### 374 **Genetic structure**

375 We set four as the *a priori* number of clusters (K=4) based on the four taxa  
376 circumscribed within the Linifolia clade. DAPC analysis identified only one optimal  
377 principal component for the scatterplot which revealed two well-differentiated clusters:  
378 *I. linifolia* (LIN) on the one side, and the coastal *I. kusinskiana* (KUZ) and *I. littoralis*  
379 (LIT\_LIT, LIT\_GAL) on the other (Fig. 3c). Within the coastal species cluster, *I.*  
380 *kusinskyana* occupied a position closer to *I. linifolia*, while the two subspecies of *I.*  
381 *littoralis* were intermingled (Fig. 3c). Compoplot assigned all individuals of *I. linifolia*  
382 to a separate cluster, without admixture (Fig. 3c). The other three clusters were shared  
383 among coastal species with different membership probability (Fig. 3c). One of the two  
384 individuals of *I. kusinskyana* was uniquely assigned to one of the clusters, whereas the  
385 other individual showed a mixed assignment, with admixture of the three genetic groups

386 (Fig. 3c). Most individuals (nine) of both subspecies of *I. littoralis* were placed in the  
387 same two clusters, and two of them showed admixture with the same cluster shared by  
388 the two samples of *I. kuzinskyana* (Fig. 3c). Given the interdigitated pattern of the two  
389 subspecies of *I. littoralis*, we repeated the analysis by considering the two subspecies as  
390 part of a single group (K=3) to evaluate the genetic clustering of *I. littoralis* as a genetic  
391 group. Compoplot for K=3 assigned most individuals (nine) of *I. littoralis* to the same  
392 cluster, while two samples had mixture assignment probability to the cluster of *I.*  
393 *kuzinskyana* (Fig. S3). Likewise, AMOVA of the two groups (*I. linifolia* vs coastal  
394 species) showed significant among-group variance (58.88%) higher than within-  
395 population variance (38.09%) or variance within populations among groups (3.03%),  
396 with a fixation index of  $F_{ST}= 0.619$  (Table 1). D-statistic tests did not conclusively  
397 support a role of historical introgression in determining paraphyletic pattern since  
398 ancient introgression was inferred indiscriminately among lineages of *I. linifolia* and  
399 both coastland species (see Methods S1).

#### 400 **Morphometric analysis**

401 Principal Component Analysis (PCA) revealed morphological differentiation between  
402 the three taxa. *Iberodes linifolia* exhibited the greatest morphological variation, ranging  
403 from the two extremes of both PC1 and PC2 compared to the more restricted variation  
404 of coastal species that is concentrated in higher scores of PC1 and lower of PC2. Three  
405 species of the Linifolia clade were almost completely differentiated, which supports the  
406 previous taxonomic observations. *Iberodes linifolia* and *I. littoralis* occupied the two  
407 extremes of variation ranging from higher length of vegetative characters, higher nutlet  
408 and nutlet teeth sizes for *I. linifolia* to higher length and density of nutlet trichomes and  
409 presence of bracts for *I. littoralis* (Fig. 3a). *Iberodes kuzinskyana* occupied an  
410 intermediate position between *I. littoralis* and *I. linifolia*, overlapping very slightly with  
411 the latter. The two subspecies of *I. littoralis* were intermingled at the highest scores of  
412 PC1 and the lowest of PC2 (Fig. 3a).

#### 413 **Climatic niche differentiation and distribution modelling**

414 The two main principal components of the 6 climatic variables explained 73.59%  
415 (PC1=48.86% and PC2=24.73%) of the climatic variability in the study region. The  
416 variables that contributed most to PC1 were: bio1 (annual mean temperature), bio8  
417 (mean temperature of wettest quarter), bio12 (annual precipitation), and bio14  
418 (precipitation of the driest month). Bio3 (isothermality) and bio4 (temperature  
419 seasonality) were the two variables most related to PC2. The visualization of the E-

420 space showed little overlap among taxa (Fig. 3b). The geographically widespread  
421 *Iberodes linifolia* showed the widest niche, whereas the more narrowly distributed  
422 coastal species are more restricted in climatic niche space (Fig. 3b). Among coastal  
423 species, *I. kuzinskyana* differs from *I. littoralis* in PC1 (less precipitation and higher  
424 temperatures for *I. kuzinskyana*) and overlaps with *I. littoralis* ssp. *gallaecica* but not  
425 with *I. littoralis* ssp. *littoralis* in PC2 (higher isothermality in *I. kuzinskyana* and *I.*  
426 *littoralis* ssp. *gallaecica*). The two subspecies of *I. littoralis* have well-differentiated  
427 niches particularly along PC2. *Iberodes kuzinskyana* occupies an intermediate position  
428 between *I. linifolia* and *I. littoralis* subsp. *gallaecica*. Thus, *I. kuzinskyana* marginally  
429 overlaps *I. linifolia* in those parts of the niche of *I. linifolia* with higher isothermality,  
430 lower temperature and higher precipitation. Indeed, minimum niche overlap was  
431 inferred between both subspecies of *I. littoralis* and *I. linifolia* due to lower temperature  
432 and higher precipitation for coastal taxa. Consequently, although tests of similarity  
433 indicated that observed values of niche overlap are not significantly lower than random  
434 overlap, all pairwise equivalency tests significantly rejected the equivalency of niches  
435 and this is supported by the low values of D obtained (Table 2).

436 The potential ranges for the species of the Linifolia clade in the present are  
437 broadly congruent with known geographic distributions, with overlapping distributions  
438 of *I. linifolia* and *I. kuzinskyana* (Fig. 4). In contrast, a contraction of distribution ranges  
439 was inferred for the LIG, including the disappearance of the potential range of *I.*  
440 *littoralis* ssp. *gallaecica* (Fig. 4), but maintaining the overlap and absence of physical  
441 barriers between *I. linifolia* and *I. kuzinskyana*. Projections to the LGM showed a range  
442 similar to the present one for *I. linifolia* and expanded ranges for both *I. kuzinskyana*  
443 and *I. littoralis* ssp. *gallaecica*. The range of *I. littoralis* ssp. *gallaecica* is inferred to  
444 have occupied most of the current range of both subspecies in the LGM, before subsp.  
445 *littoralis* differentiated from subsp. *gallaecica*.

#### 446 **Ploidy level estimation**

447 Mean peaks of fluorescence, CV values, and genome sizes obtained for each species are  
448 summarized in Fig. S4 and Table S5. Fluorescence peaks were similar among all  
449 *Iberodes* species, ranging from 45.2 to 62.6 nm for G1. No evidence of duplicated DNA  
450 content was observed, suggesting no ploidy differences among taxa or populations.  
451 Likewise, similar genome sizes were obtained for the five species ranging from 1.788 to  
452 1.980 pg. *Iberodes kuzinskyana* and *I. littoralis* ssp. *littoralis* exhibited the largest and

453 the smallest genome sizes in the genus, respectively. The five species are similar in  
454 genome size to the *S. lycopersicum* standard (1.88-2.07 pg, Grandillo *et al.*, 2011).

455

## 456 **DISCUSSION**

457 Phylogenetic relationships among *Iberodes* species are robustly inferred by RAD-  
458 sequencing, which demonstrated the monophyly of populations except for *I. linifolia*.  
459 Two early-diverging monophyletic species (*I. brassicifolia* and *I. commutata*) contrast  
460 with paraphyly of three species in the Linifolia clade (Fig. 1). The paraphyly of *I.*  
461 *linifolia* with respect to *I. kuzinskyana* and *I. littoralis* suggests a progenitor-derivative  
462 relationship between the species, with the widespread species giving rise to the coastal  
463 endemics in the course of a northward colonization from the coast of Portugal to the  
464 west coast of France. Our results provide multiple lines of evidence that support  
465 parapatric divergence of the coastland lineage (*I. kuzinskyana*-*I. littoralis* clade): (1)  
466 lack of any physical barrier to gene flow between inland and coastland in the present or  
467 the past; (2) low level of contemporary gene flow combined with evidence of historical  
468 introgression, supporting budding differentiation rather secondary contact; (3) recent  
469 niche differentiation from inland habitats with higher temperature seasonality to  
470 coastland ones with higher precipitation; (4) paraphyly of *I. linifolia*, which can be  
471 interpreted as the extant inland ancestor of the coastland lineage; and (5) incipient  
472 morphological differentiation between *I. linifolia* and the coastal lineage. Subsequently,  
473 differentiation in allopatry of the two coastal species during the mid-Pleistocene led to  
474 the modern-day species *I. kuzinskyana* and *I. littoralis*, most likely mediated by  
475 geographic isolation and niche differentiation. Thus ecological and geographic  
476 differentiation contributed to both parapatric and allopatric speciation in the clade.

### 477 **A predominant paraphyletic pattern during *Iberodes* evolution**

478 Our RAD-seq analyses reveal a pattern of predominant paraphyly within the recently  
479 diverged Linifolia clade (Fig. 2). While the populations of *I. brassicifolia* and those of *I.*  
480 *commutata* form monophyletic groups, paraphyly is inferred for *I. linifolia* populations  
481 with respect to the two coastal species. While apparent paraphyly of species may be a  
482 consequence of introgression dragging populations toward the species to which they  
483 have introgressed (e.g., Eaton *et al.*, 2015), our analyses of historical introgression using  
484 *D*-statistics do not conclusively support a role of introgressive hybridization in shaping  
485 the paraphyletic topology (Methods S1), and genetic structure shows no evidence of  
486 recent admixture involving *I. linifolia* (Fig. 3c). This suggests that *Iberodes* contains



487 some truly paraphyletic species, supporting a progenitor-derivative or “budding”  
488 speciation history. Despite the tendency in taxonomy to only consider monophyletic  
489 species, paraphyletic ones have been reported as particularly common in groups  
490 showing recent local speciation, i.e. groups composed of a ‘mother’ species that  
491 coexists in time and space with more restricted derivative species (Rajakaruna, 2018).

#### 492 **Recent ecological speciation in parapatric *Iberodes***

493 Although parapatry has been argued to be common in plants (Rieseberg & Brouillet,  
494 1994), few empirical examples are found on the literature (Table 3). Our study provides  
495 strong evidence for parapatry through a process of progenitor-derivative (or budding)  
496 speciation between *I. linifolia* and the ancestor of *I. kuzinskyana* and *I. littoralis*. Indeed,  
497 the projection of distribution models to past climatic periods during the Quaternary  
498 revealed a similar or greater degree of geographic overlap between the potential ranges  
499 of *I. linifolia* and *I. kuzinskyana* (Fig. 4), which makes allopatry unlikely (Zheng *et al.*,  
500 2017). Peripheral populations of the widespread *I. linifolia* close to the coast of Portugal  
501 are the most closely related potential living progenitor of the coastal species (*I.*  
502 *kuzinskyana* and *I. littoralis*; Fig. 2), which seem to have differentiated in an adjacent  
503 area with no apparent physical barrier. Paraphyly similarly supports the hypothesis of  
504 budding (progenitor-derivative) speciation (Stuessy & Hörandl, 2013). In addition, the  
505 historical introgression inferred among *I. linifolia* and coastland species (Methods S1),  
506 does not conclusively underlie the paraphyletic topology. Moreover, our results of  
507 AMOVA and DAPC pointed to lack of recent gene flow between the nearby species (*I.*  
508 *kuzinskyana* and *I. linifolia*). This pattern supports strong isolation rather than secondary  
509 contacts as suggested in other cases of ecological differentiation (e.g. *Mimulus*,  
510 Stankowski *et al.*, 2015; *Stauracanthus*, Chozas *et al.*, 2017; Table 3). Nevertheless, a  
511 deeper population genetics study focused on the adjacent populations of Portugal is  
512 necessary to obtain more detailed estimates of gene flow.

513 In the earliest stages of parapatric speciation mediated by ecological differentiation,  
514 reproductive barriers are expected to evolve in response to selection towards locally  
515 adapted, divergent ecotypes (Sakaguchi *et al.*, 2019). The niche identity tests point to a  
516 role of ecologically-based selection in the speciation of the coastal lineage from *I.*  
517 *linifolia*. The coastal lineage is distributed in niches with less temperature seasonality  
518 and higher precipitation means than *I. linifolia* (Fig. 3b). Interestingly, although niche  
519 overlap is close to zero, similarity tests indicated a certain degree of similarity within  
520 the E-space, which points to recent divergence and suggests ongoing differentiation or

521 some degree of niche conservatism. Besides, edaphic factors such as the specialization  
522 in coastal dune substrates may also have contributed to ecological differentiation,  
523 although this is a fine-scale soil variable that has not been mapped at high enough  
524 resolution to include meaningfully in niche models. The differentiation of coastal  
525 lineages (*I. kuzinskyana* and *I. littoralis*) from inland populations of *I. linifolia* seems to  
526 have started in western Iberian Peninsula during the Mid Pliocene (3.85-3.60 myr, Fig.  
527 2), when increasing seasonality led to the origin of Mediterranean climate with its  
528 summer droughts (Suc, 1984). The onset of Mediterranean climate favored the  
529 differentiation and configuration of modern Mediterranean flora (Vargas *et al.*, 2018),  
530 which agrees with recent *Iberodes* speciation (Fig. 2). Posterior glacial/interglacial  
531 periods during the Late Pliocene promoted the expansion and contraction of geographic  
532 ranges and the formation of sources and sinks of genetic diversity for most  
533 Mediterranean species (Médail & Diadema, 2009). Interestingly, the overlapping area  
534 inferred for *I. linifolia* and *I. kuzinskyana* in the past is congruent with the location of  
535 glacial refugia (Estremadura and Beira litoral) proposed by Médail and Diadema (2009)  
536 and plant species previously studied as glacial refugial species (e.g. *Drosophyllum*  
537 *lusitanicum*: Müller & Deil, 2001; *Cistus monspeliensis*: Fernández-Mazuecos &  
538 Vargas, 2010; Coello *et al.*, unpublished; *Linaria elegans*: Fernández-Mazuecos &  
539 Vargas, 2013).

540 In addition, ecological differentiation in parapatric speciation involves divergent  
541 natural selection towards contrasting environments that promotes adaptation to different  
542 environments and subsequent barriers to gene flow (Schluter, 2001; Rundle & Nosil,  
543 2005). The strength of divergent selection can modulate the degree of completeness of a  
544 speciation process (Nosil *et al.*, 2009). In our case, both the high interspecific genetic  
545 differentiation and clear morphological and ecological divergence (Fig. 3) between  
546 inland and coastal species may reflect strong divergent selection promoting rapid  
547 speciation (Pliocene-Pleistocene, Fig. 2). Indeed, our study reinforces the common trend  
548 for Mediterranean plants in which narrow endemics arise from ecological differentiation  
549 of peripheral populations of more widespread progenitor species (Papuga *et al.*, 2018).  
550 Moreover, this trend highlights the role of peripheral populations in setting the scene for  
551 diversification, thus revealing their conservation value (Debussche & Thompson, 2003).

552

553 **Allopatric differentiation along the Atlantic coast**

554 An allopatric pattern is also found in the Linifolia clade. Given the current separation  
555 between ranges of the three coastal taxa (Fig. 1), allopatry seems to have triggered  
556 speciation. On the one hand, the widely separated ranges of *I. kuzinskyana* and *I.*  
557 *littoralis* subsp. *gallaecica* both in current times and in projections to the LIG (despite  
558 probable intermittent contact during the LGM) suggest either a colonization mediated  
559 by LDD followed by geographic isolation and niche shift (Fig. 3, 4) or a vicariant  
560 process mediated by an early expansion and contraction of distribution ranges. We find  
561 similar examples of recent allopatric speciation for both hypotheses: by LDD among  
562 coastal Mediterranean flora in the same period (e.g. Jakob *et al.*, 2007; Carnicero *et al.*,  
563 2017; Herrando-Moraira *et al.*, 2017); and expansion/contraction of distribution ranges  
564 (e.g. Ortiz *et al.*, 2007; Lo Presti & Oberprieler, 2011; Fernández-Mazuecos & Vargas,  
565 2011). On the other hand, the allopatric differentiation of the two subspecies of *I.*  
566 *littoralis* was likely preceded by an expansion of *I. littoralis* during the LGM, followed  
567 by geographic isolation as its range contracted and niche shifted, leading to the  
568 differentiation of *I. littoralis* subsp. *littoralis* in more recent times (Fig. 3, 4). In this  
569 case, the probable role of long distance dispersal (LDD) is further supported by the fact  
570 that the expanded area during the LGM does not seem to have formed a continuous  
571 range (Fig. 4). Indeed, unlike inland species of *Iberodes*, the three coastal taxa seem to  
572 have specializations for LDD in the form of uncinated hooks (Fig. 1) in the nutlet,  
573 related to the attachment to feathers and fur in other closely related Boraginaceae genera  
574 (Selvi *et al.*, 2011).

575

## 576 CONCLUSIONS

577 Our integrative approach provides a comprehensive history of parapatric speciation,  
578 which has long posed a challenge to our understanding of ecological speciation. The  
579 integration of RAD-sequencing, morphometrics and climatic niche modelling provide  
580 strong evidence that progenitor-derivative speciation with gene flow, associated with  
581 ecological divergence, shapes the parapatry we observe today in *Iberodes*. Our work  
582 thus serves as a model for how integrative studies may serve as a powerful resource for  
583 investigating scenarios of non-allopatric processes in other groups of plants. Processes  
584 such as budding speciation bring to light the need to extend species concepts to  
585 encompass paraphyletic species as natural units of both taxonomic classification and  
586 evolutionary history: species in every sense of the word.

587

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612 AO, PJM, VV and PV contributed to developing the question and experimental design.  
613 AO led the material collection. AO, MFM and ALH conducted the data analysis. AO  
614 wrote the manuscript with the contribution of all authors in both interpretation and  
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## 951 **FIGURE LEGENDS**

952 **Fig. 1** Locations of the *Iberodes* species sampled for the different analyses performed.  
953 The star symbols indicate locations for RAD-sequencing, squares represent the  
954 locations for the morphometric analysis, and circles indicate locations for the ecological  
955 niche modelling. Color legend for each taxa of *Iberodes* is shown in the figure. Images  
956 of the six taxa are framed following each taxa color in the map.

957 **Fig. 2** Time-calibrated phylogeny obtained from TreePL by using maximum clade  
958 credibility from 1000 bootstrap trees of the maximum likelihood analysis of c95m20  
959 matrix. Branch thickness indicates the ML bootstrap supports above 98. Node bars in  
960 blue indicate the node age ranges taking into account the branch length variance along  
961 the 1000 bootstrap trees. No branch length variation was inferred for nodes without blue  
962 bar. The international stratigraphic scale is included from 26 myr until present. Text  
963 codes for each individual location are detailed in the Table S1. The color for each  
964 lineage of the Linifolia clade is geographically represented through the colored ellipses  
965 of the map. Grey ellipses group the different lineages of the four taxa within the  
966 Linifolia clade.

967 **Fig. 3** Morphometric, niche overlap and genetic clustering analyses of the Linifolia  
968 clade. Codes for each taxa are: LIN for *I. linifolia*, KUZ for *I. kuzinskyana*, LIT-GAL  
969 for *I. littoralis* ssp. *gallaecica*, and LIT-LIT for *I. littoralis* ssp. *littoralis*. (a)  
970 Morphometric analysis. Principal Component Analysis (PCA) of the 12 morphologic  
971 characters: LS, length of the stem; LL, length of the leaf; WL, wide of the leaf; PINFL,  
972 pedicel of inflorescence; LCFR, length of fruit calix; DN, diameter of the nutlet; LN,  
973 length of the nutlet; LM, length of the margin nutlet; LT, length of the margin teeth;  
974 DH, density of trichomes; LH, length of trichome; BR: absence=0, presence=1 of  
975 flowering bracts). The contribution of each trait to the two principal components (PC1,  
976 PC2) as well as the positive or negative sense of the relationship is represented through  
977 the length of each arrow and the directionality of the arrow, respectively. The  
978 percentages of the variability explained by the two PCs are indicated close to the axes.  
979 (b) Niche overlap analysis. PCA of the six climatic variables obtained from WorldClim.  
980 BIO1, Annual Mean Temperature; BIO3, Isothermality; BIO4, Temperature  
981 Seasonality; BIO8, Mean Temperature of Wettest Quarter; BIO12, Annual  
982 Precipitation; BIO14, Precipitation of Driest Month. The strength of contribution of  
983 each variable to PCs and the sense of the relationship is represented, as explained above,  
984 through the length and the directionality of the arrows. The percentages of the

985 variability explained by the two PCs are indicated close to the axes. Values of the  
 986 variables are also represented for each PC separately following Fernández-López and  
 987 Villa-Machío (2017). (c) Genetic clustering. Scatterplot from the DAPC analysis  
 988 showing the genetic differentiation retained in one PC as the optimal number of PC for  
 989 the discriminant function. Compoplot for K=4 is also represented indicating  
 990 membership probability to each genetic cluster for each taxa.

991 **Fig. 4** Species distribution modelling obtained through the entropy algorithm, as  
 992 implemented in Maxent for the four taxa of the Linifolia clade. Codes for each taxa are:  
 993 LIN for *I. linifolia*, KUZ for *I. kuzinskyana*, and LIT for *I. littoralis* (LIT-GAL for *I.*  
 994 *littoralis* ssp. *gallaecica*, LIT-LIT for *I. littoralis* ssp. *littoralis*). Analyses are based on  
 995 WorldClim and projected to past conditions: last interglacial (LIG, c. 120–140 kya) and  
 996 last glacial maximum (LGM, c. 21 kya). The presence/absence was estimated using the  
 997 maximum training sensitivity plus specificity logistic threshold. The projection to the  
 998 past is only presented according to the inferred times of divergence of each taxa.

999

## 1000 TABLES

1001 **Table 1** Analysis of molecular variance (AMOVA) results for individuals from two  
 1002 contrasted groups: (1) mainland, *I. linifolia* and (2) coastal, *I. kuzinskyana* and *I.*  
 1003 *littoralis*.

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Source of Variation	d.f.	Sum of squares	Variance components	Percentage of variation
Among groups	1	1602.666	106.33509 Va	58.88
Among populations within groups	2	181.372	5.47321 Vb	3.03
Within populations	24	1651.033	68.79306 Vc	38.09
Total	27	3435.071	180.60135	

1005 Note: Variance components are as in Excoffier *et al.* (1992). Fst = 0.61909; Fsc = 0.07370; Fct  
 1006 = 0.58878.

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1010 **Table 2** Pairwise statistical test for comparison of ecological niche overlap between the  
 1011 different taxa within the Linifolia clade. The statistical significance is represented by p-  
 1012 values (Warren *et al.*, 2008). Asterisk (\*) indicate significant values for  $p$ -values < 0.01.

Taxa comparison	Schoener's D	Niche equivalency p-value	Niche similarity p-value
<i>I. linifolia</i> vs <i>I. kuzinskyana</i>	0.026	0.0099*	0.94059
<i>I. linifolia</i> vs <i>I. littoralis</i> ssp. <i>gallaecica</i>	0.003	0.003	0.60396
<i>I. linifolia</i> vs <i>I. littoralis</i> ssp. <i>littoralis</i>	0.004	0.0099*	0.69307
<i>I. kuzinskyana</i> vs <i>I. littoralis</i> ssp. <i>gallaecica</i>	0.000	0.0099*	0.64356
<i>I. kuzinskyana</i> vs <i>I. littoralis</i> ssp. <i>littoralis</i>	0.000	0.0099*	0.71287
<i>I. littoralis</i> ssp. <i>gallaecica</i> vs <i>I. littoralis</i> ssp. <i>littoralis</i>	0.000	0.0099*	0.67327

1013

1014 **Table 3** Examples of parapatric speciation in plants.

Genus	Family	Event suggested	Reference
<i>Ephedra</i>	Ephedraceae	Ecological divergence mediated by bioclimatic niche	Yin <i>et al.</i> (2016)
<i>Fosterella</i>	Bromeliaceae	Ecological divergence mediated by polyploidization and	Paule <i>et al.</i> (2017)
<i>Helianthus</i>	Asteraceae	Ecological divergence mediate by bioclimatic niche	Andrew & Rieseberg (2013)
<i>Mimulus</i>	Phrymaceae	Ecological divergence mediated by pollinator shift	Sobel & Streisfeld (2015), Stankowski <i>et al.</i> (2015)
<i>Miscanthus</i>	Poaceae	Ecological divergence mediated by bioclimatic	Huang <i>et al.</i> (2014)

niche			
<i>Oryza</i>	Poaceae	Ecological divergence mediated by bioclimatic niche	Zheng & Ge (2010)
<i>Populus</i>	Salicaceae	Ecological divergence mediated by bioclimatic niche	Zheng <i>et al.</i> (2017)
<i>Roscoea</i>	Zingiberaceae	Ecological divergence mediated by bioclimatic niche	Zhao <i>et al.</i> (2016)
<i>Senecio</i>	Asteraceae	Ecological divergence mediated by ecological niche	Richards & Ortiz-Barrientos (2016)
<i>Stauracanthus</i>	Fabaceae	Ecological divergence mediated by bioclimatic niche	Chozas <i>et al.</i> (2017)
<i>Columnnea</i>	Gesneriaceae	Ecological divergence mediated by bioclimatic niche and pollinator shift	Schulte <i>et al.</i> (2015)

1015

1016

## 1017 SUPPORTING INFORMATION

1018 Additional Supporting Information may be found online in the Supporting Information  
1019 section at the end of the article.

1020 **Fig. S1** RAxML trees of *Iberodes* using three different similarity percentages (85%,  
1021 90%, 95%) and four levels of minimum coverage (m4, m12, m20, m28). Bootstrap  
1022 support are indicated when is lower than 100.

1023 **Fig. S2.** SVDquartets species tree of *Iberodes* using the eight main lineages obtained in  
1024 the RAxML analyses.

1025 **Fig. S3** Compplot showing individual assignment probability to different species-  
1026 groups of the Linifolia clade of *Iberodes*, considering the two subspecies of *I. littoralis*  
1027 as one group (k=3). Codes for each taxa are: LIN for *I. linifolia*, KUZ for *I.*  
1028 *kuzinskyana*, LIT-GAL for *I. littoralis* ssp. *gallaecica*, and LIT-LIT for *I. littoralis* ssp.  
1029 *littoralis*.

1030 **Fig. S4** Flow cytometry peaks for each of five samples used for each of the five species  
1031 of *Iberodes* (including the two subspecies of *I. littoralis*. Peak of the standard *Solanum*  
1032 *lycopersicum* is included. FL1 in the X axis indicates the signal intensity (530 nm / 28  
1033 nm; DAPI emission maximum=461). Y axis indicates the number of events found.  
1034 Mean, mode, and median values of FL and percentage of variation coefficient is shown  
1035 for each colored peak.

1036 **Table S1** Data information and NCBI SRA accession numbers of all individuals  
1037 sampled for RADseq.

1038 **Table S2** Morphological characters analyzed. Values represent the mean of two  
1039 individuals measured per herbarium sheet. LS: length of the stem, LL: length of the leaf,  
1040 WL: wide of the leaf, PINFL: pedicel of inflorescence, LCFR: length of fruit calix, DN:  
1041 diameter of the nutlet, LN: length of the nutlet, LM: length of the margin nutlet, LT:  
1042 length of the margin teeth, DH: density of the hair, LH: length of trichome.

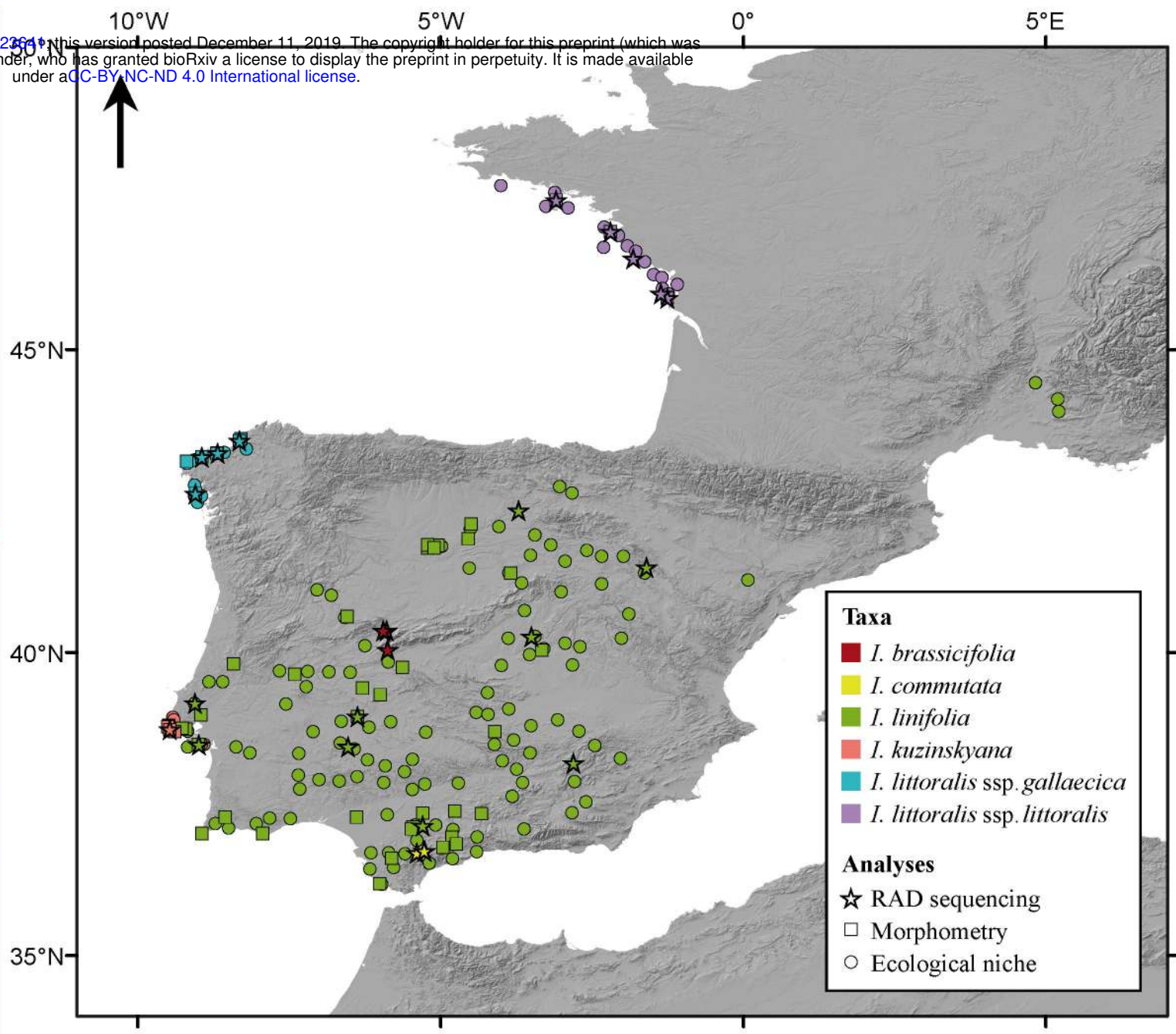
1043 **Table S3** Data points for environmental analysis

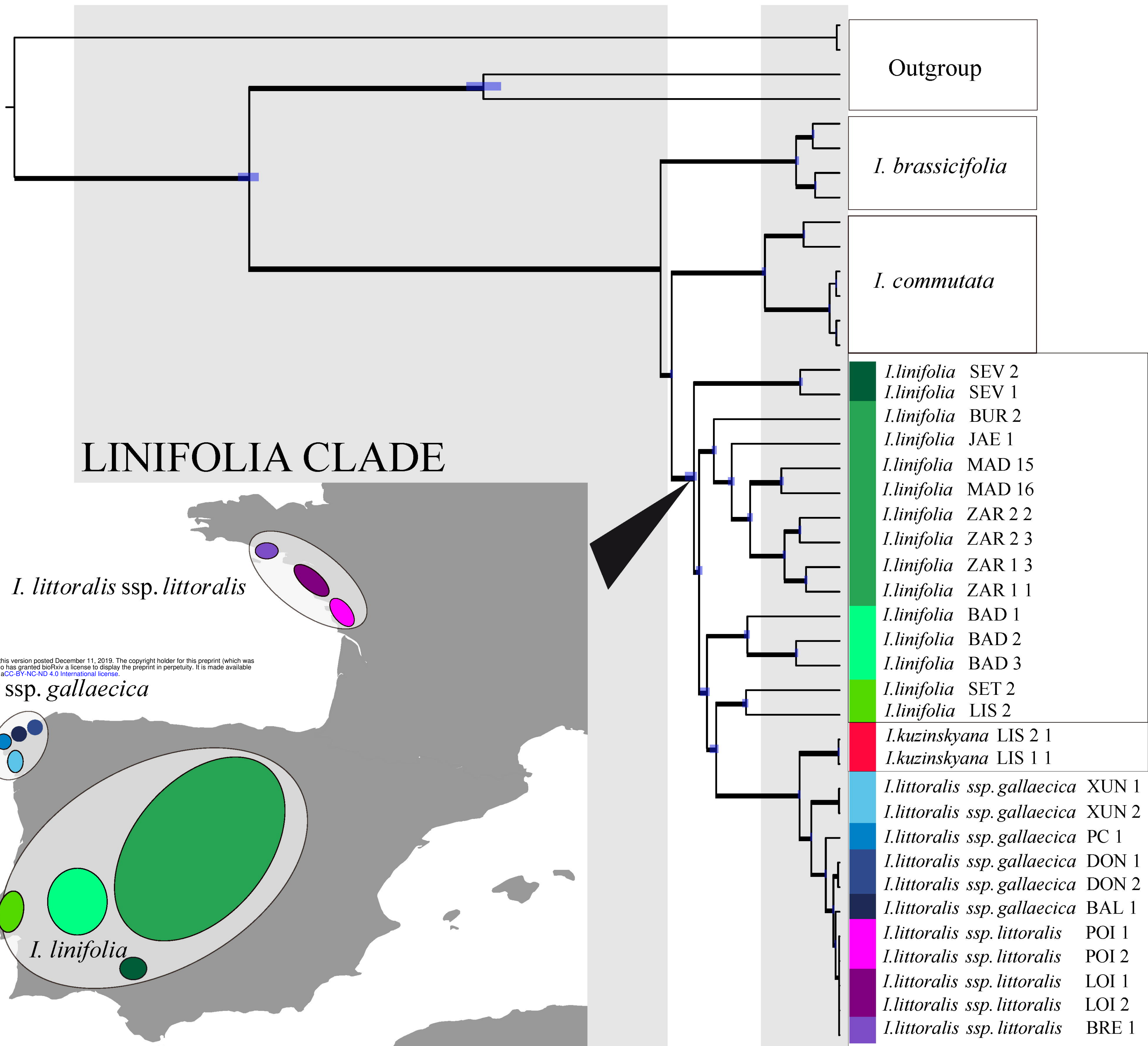
1044 **Table S4** Number of loci, SNPs retrieved for each of the 12 parameter combinations.  
1045 Parameter abbreviations indicate: c, clustering threshold; m, minimum taxon coverage to  
1046 consider a locus.

1047 **Table S5** Genome size estimation for the five species of *Iberodes* (including the two  
1048 subspecies of *I. littoralis*.

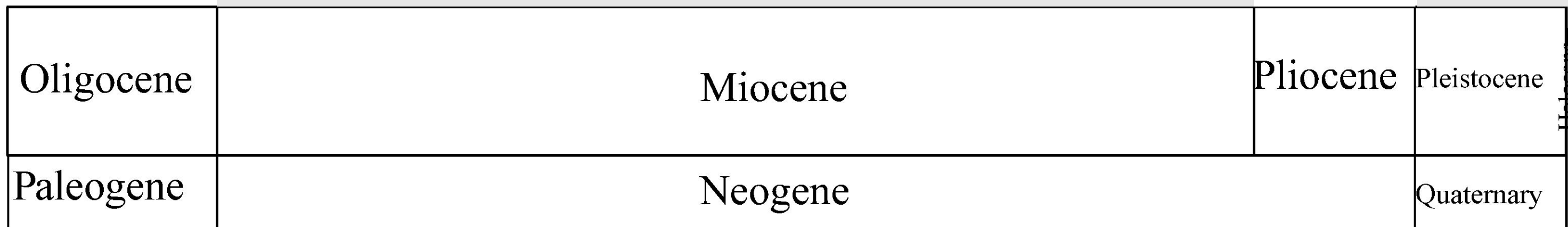
1049 **Methods S1** Introgression analysis.

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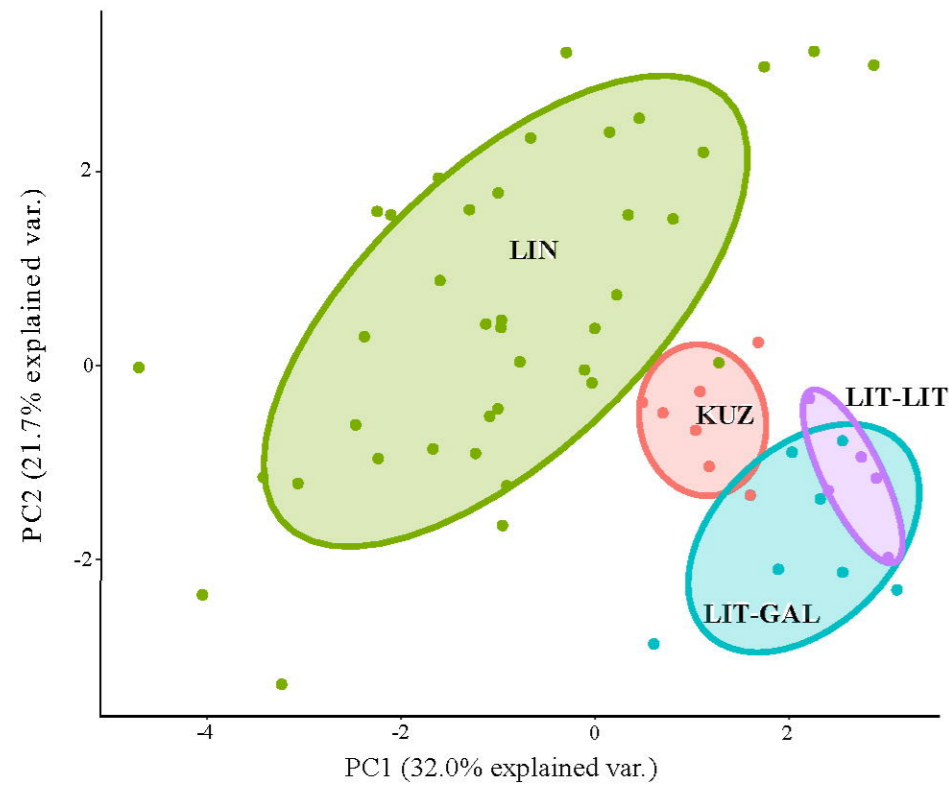
20

10

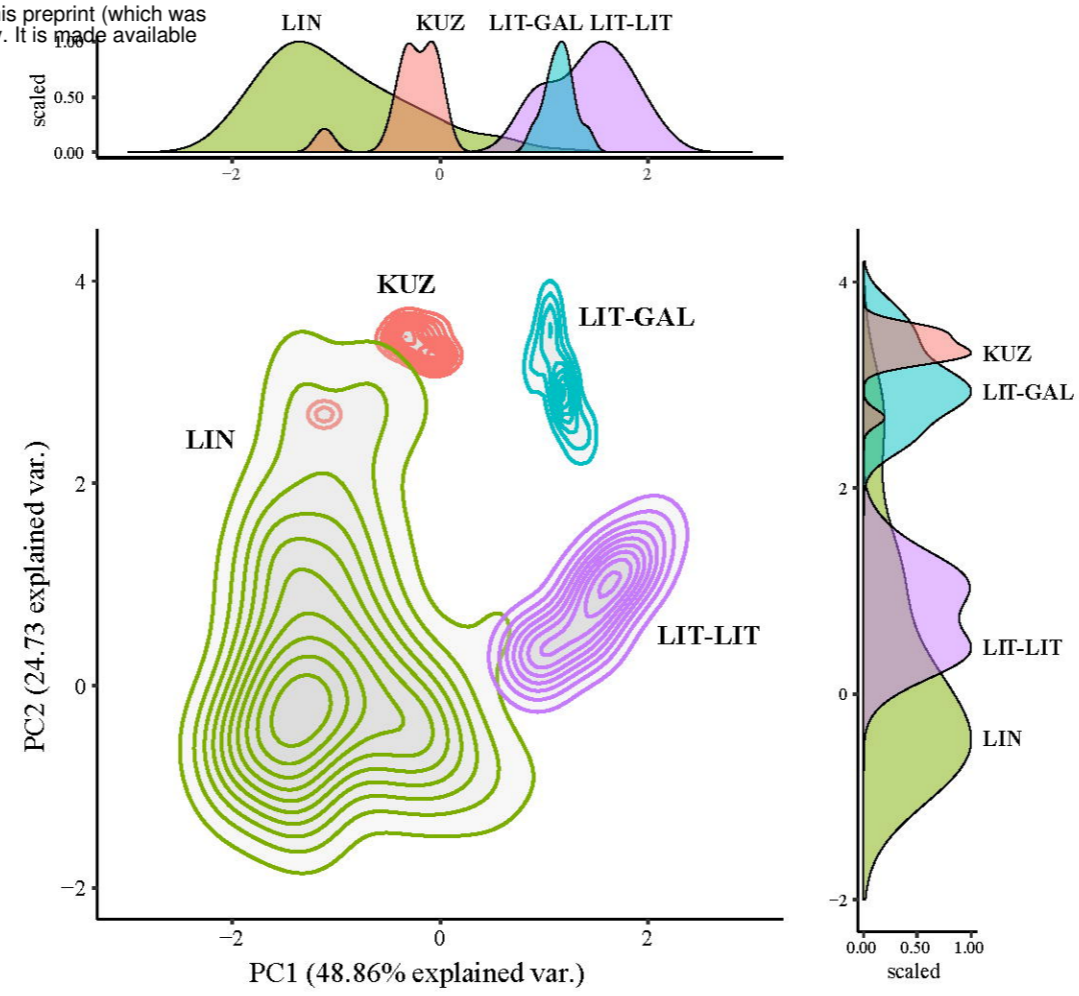
0 (myr)

### (a) Morphometric analysis

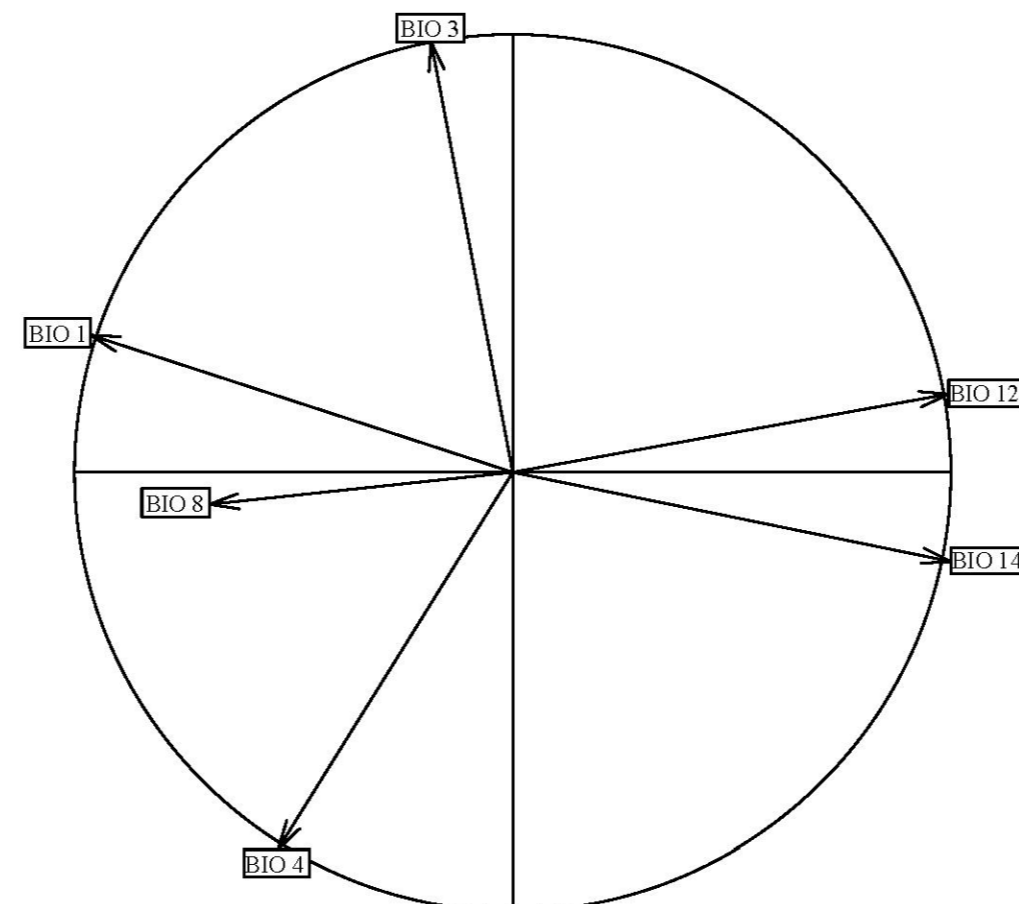
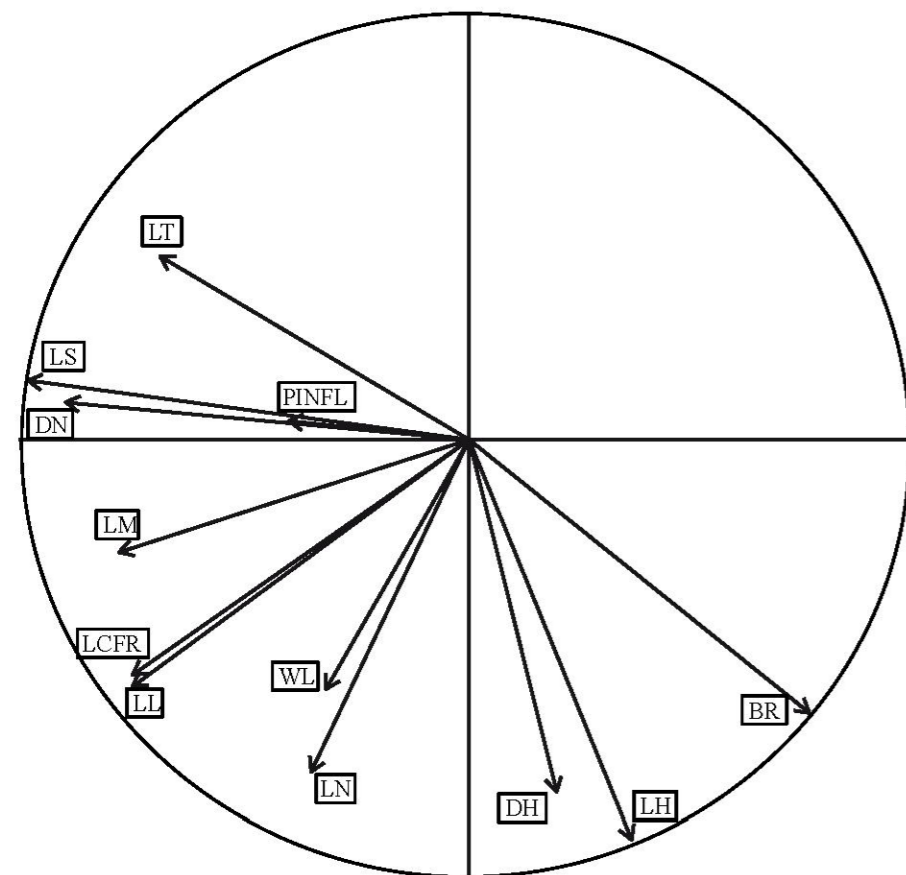
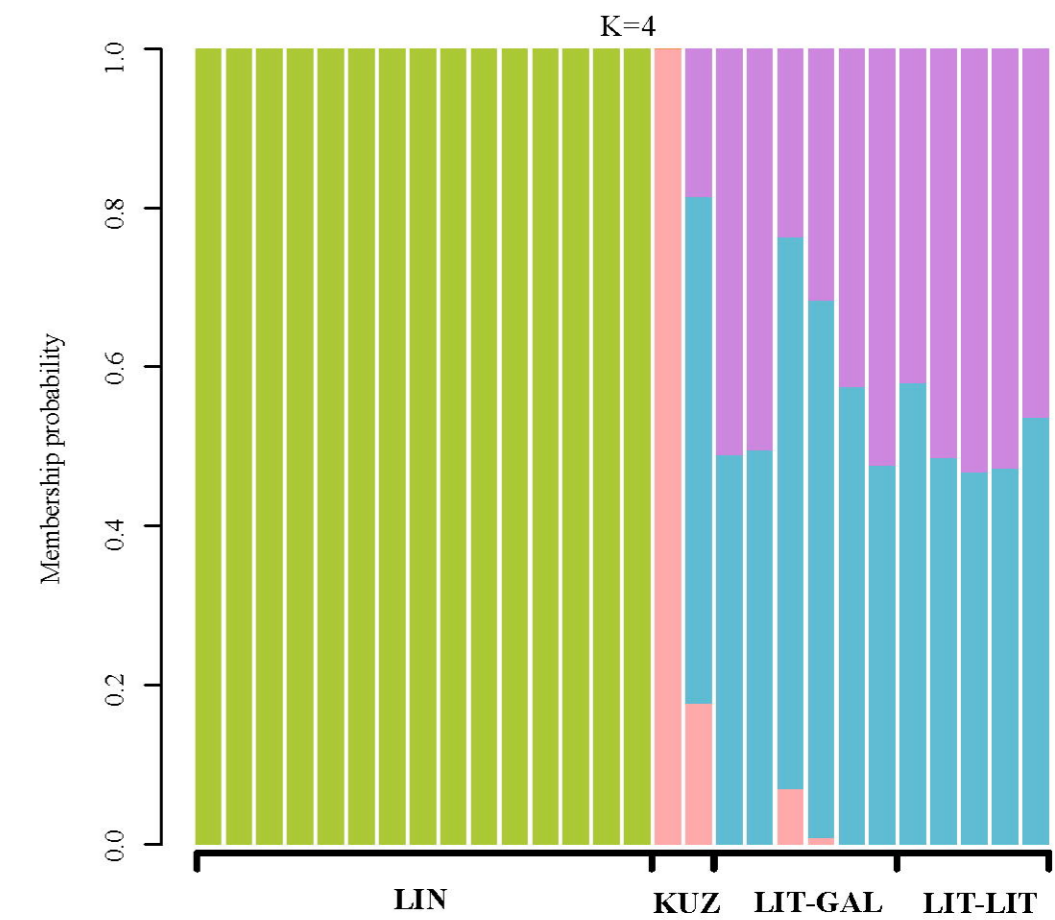
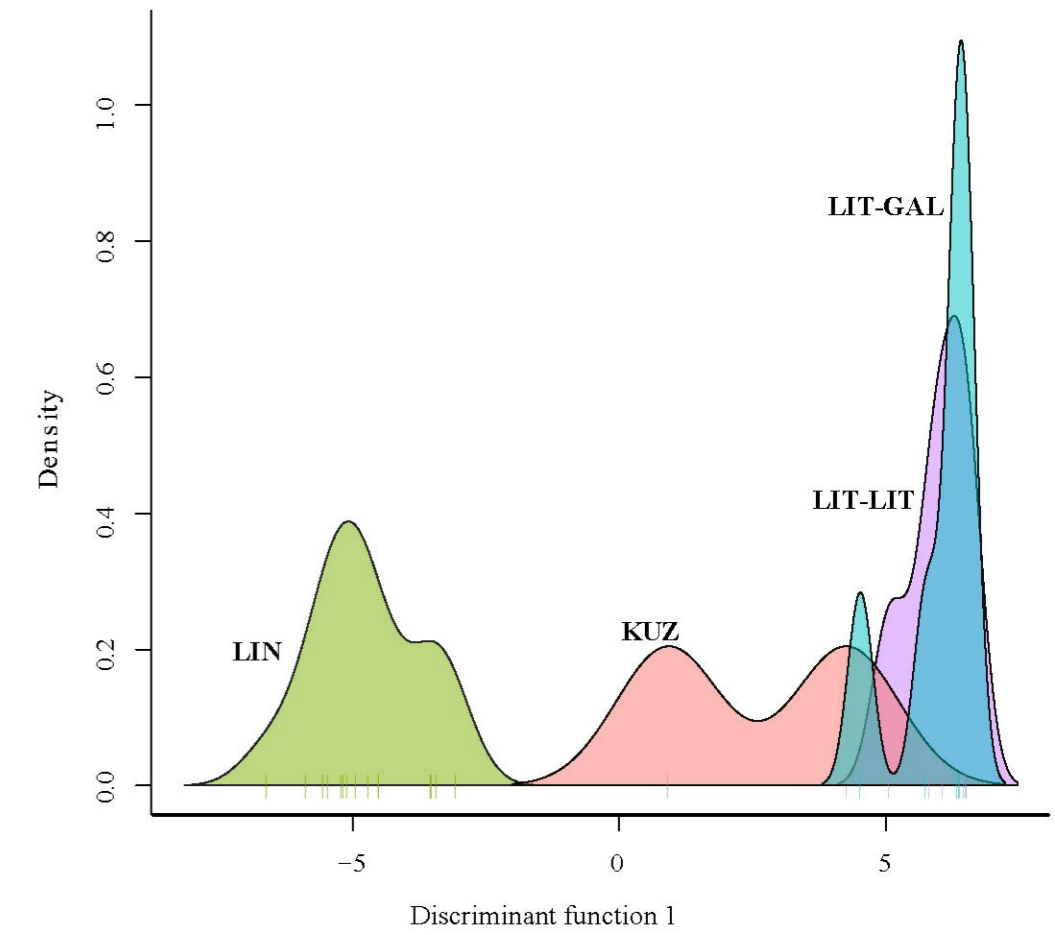
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### (b) Niche overlap



### (c) Genetic clustering



**Last interglacial**  
(120,000 - 140,000  
yr BP)

**Last Glacial  
Maximum**  
(21,000 yr BP)

**Present**

**LIN**



**KUZ**



**LIT-GAL**



**LIT-LIT**

