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Plasma membrane protein OsMCA1 is involved in regulation of hypo-osmotic shock-induced Ca^{2+} influx and modulates generation of reactive oxygen species in cultured rice cells

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Abstract

Background: Mechanosensing and its downstream responses are speculated to involve sensory complexes containing Ca^{2+} -permeable mechanosensitive channels. On recognizing osmotic signals, plant cells initiate activation of a widespread signal transduction network that induces second messengers and triggers inducible defense responses. Characteristic early signaling events include Ca^{2+} influx, protein phosphorylation and generation of reactive oxygen species (ROS). Pharmacological analyses show Ca^{2+} influx mediated by mechanosensitive Ca^{2+} channels to influence induction of osmotic signals, including ROS generation. However, molecular bases and regulatory mechanisms for early osmotic signaling events remain poorly elucidated.

Results: We here identified and investigated OsMCA1, the sole rice homolog of putative Ca^{2+} -permeable mechanosensitive channels in Arabidopsis (MCAs). OsMCA1 was specifically localized at the plasma membrane. A promoter-reporter assay suggested that *OsMCA1* mRNA is widely expressed in seed embryos, proximal and apical regions of shoots, and mesophyll cells of leaves and roots in rice. Ca^{2+} uptake was enhanced in *OsMCA1*-overexpressing suspension-cultured cells, suggesting that OsMCA1 is involved in Ca^{2+} influx across the plasma membrane. Hypo-osmotic shock-induced ROS generation mediated by NADPH oxidases was also enhanced in *OsMCA1*-overexpressing cells. We also generated and characterized *OsMCA1-RNAi* transgenic plants and cultured cells; *OsMCA1*-suppressed plants showed retarded growth and shortened rachises, while *OsMCA1*-suppressed cells carrying Ca^{2+} -sensitive photoprotein aequorin showed partially impaired changes in cytosolic free Ca^{2+} concentration ($[\text{Ca}^{2+}]_{\text{cyt}}$) induced by hypo-osmotic shock and trinitrophenol, an activator of mechanosensitive channels.

Conclusions: We have identified a sole MCA ortholog in the rice genome and developed both overexpression and suppression lines. Analyses of cultured cells with altered levels of this putative Ca^{2+} -permeable mechanosensitive channel indicate that OsMCA1 is involved in regulation of plasma membrane Ca^{2+} influx and ROS generation induced by hypo-osmotic stress in cultured rice cells. These findings shed light on our understanding of mechanical sensing pathways.

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Background

Plants need to sense and respond to mechanical stresses, such as wind, touch, and changes in osmotic pressure [1-3]. Elevation of cytosolic free Ca^{2+} concentration ($[\text{Ca}^{2+}]_{\text{cyt}}$) is induced in response to various stimuli, such as chemical, physical, and mechanical stimuli [2,4-7]. During this process, $[\text{Ca}^{2+}]_{\text{cyt}}$ levels rise through the opening of Ca^{2+} channels located on the plasma membrane and endomembranes. Electrophysiological and bioinformatic studies have revealed the existence of plasma membrane Ca^{2+} -permeable channels activated by mechanical stimuli, although the structural entity involved and their physiological functions remain largely unknown [8-12].

Molecular and electrophysiological studies have shown that *Arabidopsis thaliana* MSL9 and MSL10, homologs of the bacterial mechanosensitive channel MscS, are required for mechanosensitive channel activity in the plasma membrane of root cells, and are more permeable to Cl^- than Ca^{2+} [13,14]. We have recently identified two plasma membrane proteins as putative Ca^{2+} -permeable mechanosensitive channels, MCA1 (At4g35920) and MCA2 (At2g17780), from *Arabidopsis* [15,16], and showed that ectopic overexpression of *MCA1* increases Ca^{2+} uptake in roots, and also enhances $[\text{Ca}^{2+}]_{\text{cyt}}$ elevation upon hypo-osmotic shock. However, the direct effects of MCA proteins on osmotic-induced Ca^{2+} influx through the plasma membrane and the osmotic signaling pathways are little understood.

Upon recognition of osmotic signals, plant cells initiate activation of a widespread signal transduction network that induces second messengers and triggers inducible defense responses. Characteristic early signaling events include Ca^{2+} influx, protein phosphorylation and generation of reactive oxygen species (ROS) [17-20]. These downstream events are often prevented when Ca^{2+} influx is compromised by either Ca^{2+} chelators, such as ethylene glycol-bis-(2-aminoethylether)-*N, N, N', N'*-tetra acetic acid (EGTA), or Ca^{2+} -channel blockers, such as La^{3+} [21]. In tobacco cells, hypo-osmotic shock-induced ROS generation reportedly requires activation of mechanosensitive Ca^{2+} channels [22]. These results suggest that Ca^{2+} influx mediated by mechanosensitive Ca^{2+} channels is involved in the induction of osmotic signals including ROS generation. However, in osmotic responses, molecular bases and regulation mechanisms remain poorly elucidated.

In the present study, we have identified a sole MCA ortholog in the rice genome and developed both overexpression and suppression lines. Studies of these lines with altered levels of this putative mechanosensitive Ca^{2+} channel indicated that *OsMCA1* is involved in regulation of

plasma membrane Ca^{2+} influx and ROS generation induced by hypo-osmotic stress in cultured rice cells.

Results

Identification of *OsMCA1* and its expression patterns

Full-length cDNA of *OsMCA1* was obtained by a rapid amplification of cDNA ends (RACE)-PCR method. It encodes a polypeptide of 418 amino acid residues with a calculated molecular mass of 47,417 (GenBank Accession No. AB601973). The predicted protein showed 66.7% and 57.6% amino acid sequence identity compared with *Arabidopsis* MCA1 and MCA2, respectively; the TopPred program <http://www.sbc.su.se/~erikw/toppred2/> suggests that *OsMCA1* has two potential transmembrane segments (S1 and S2) (Additional file 1), while other transmembrane segment prediction programs suggest different numbers of putative transmembrane segments (data not shown). The PLAC8 motif was found by TMPred prediction http://www.ch.embnet.org/software/TMPRED_form.html in the C-terminal region (Additional file 1). A database search of the whole genome (Rice BLAST; <http://rice-blast.dna.affrc.go.jp/>) indicated that rice has no other homolog of *OsMCA1*.

Quantitative reverse transcriptase (RT)-PCR analysis showed *OsMCA1* mRNA to be expressed in mature leaves, shoots, roots and suspension-cultured cells, suggesting that *OsMCA1* mRNA is expressed throughout the plants in seedlings as well as in cultured cells (Additional file 2). We also consulted the microarray expression database (Rice XPro; <http://ricexpro.dna.affrc.go.jp/GGEP/index.html>, *OsMCA1* locus ID; Os03g0157300), showing expression of *OsMCA1* mRNA throughout the developmental stages, including root, leaf blade, panicle, anther, pistil, and ovary as well as embryo (data not shown). The spatial pattern of *OsMCA1* expression was examined using an *OsMCA1* promoter:: β -glucuronidase (*GUS*) fusion reporter gene construct (*OsMCA1p::GUS*). Figure 1a shows *OsMCA1p::GUS* is expressed in the seed, with relatively high levels in embryo. In the seedling stage, *OsMCA1p::GUS* is highly expressed in proximal and apical regions of shoots (Figure 1b-g). Cross sections of the leaves indicate that *OsMCA1p::GUS* is highly expressed in mesophyll cells, but expressed in vascular tissues at relatively very low levels (Figure 1h-j). *OsMCA1p::GUS* was also expressed in the root, with relatively high levels in the center of primary root as well as the lateral root primordia (Figure 1k-m). These results suggest that *OsMCA1* transcription may be regulated throughout developmental stages. The expression pattern of *OsMCA1* was similar to those of the *Arabidopsis* *MCA2*.

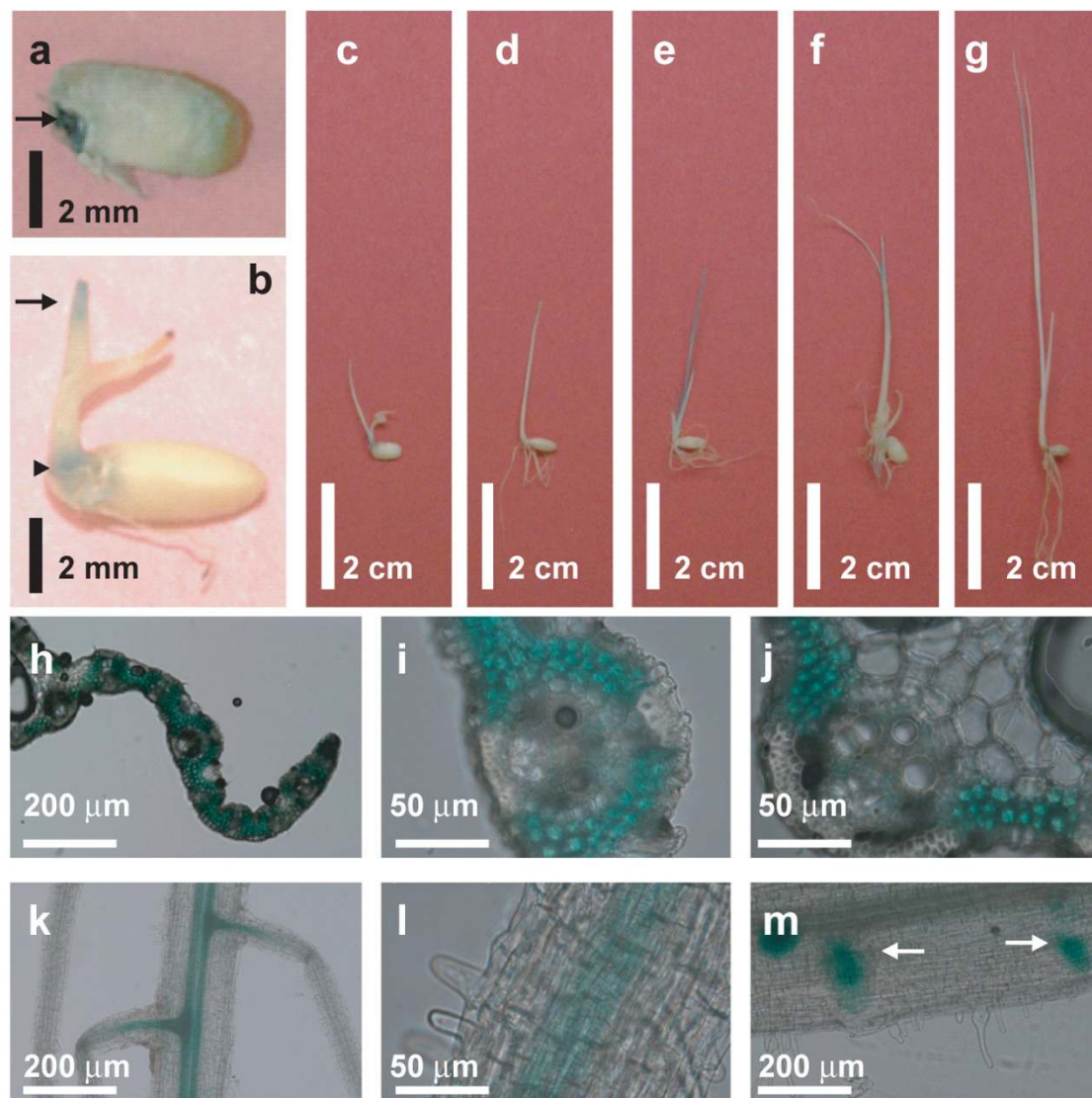
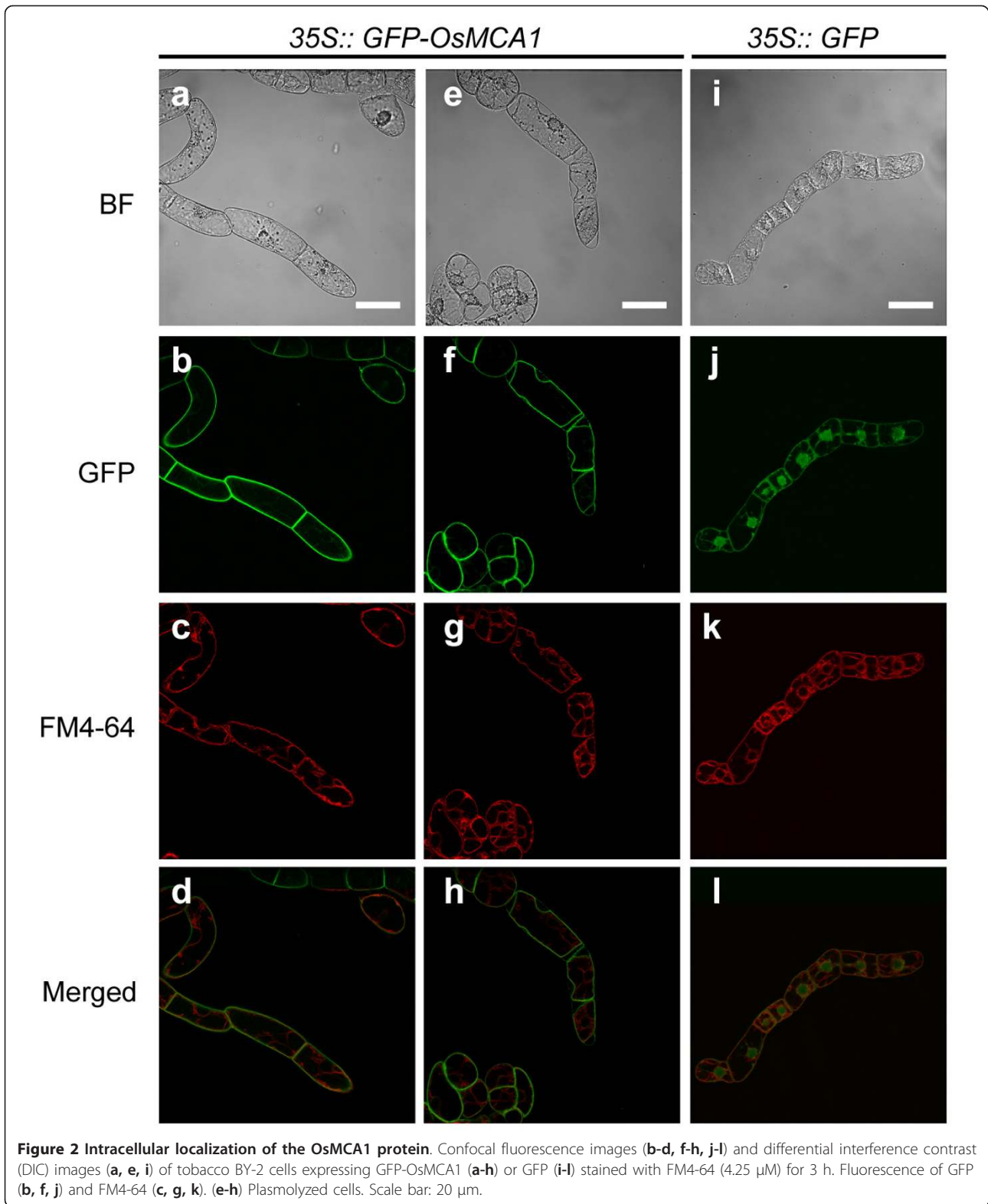


Figure 1 Spatial patterns of *OsMCA1* transcription as revealed by *GUS* staining. Transgenic rice plants harboring *OsMCA1p::GUS* were stained in X-Gluc solution and cleared in methanol. **(a)** Half of a seed. An arrow indicates the seed embryo. **(b-g)** Whole seedlings grown for 7, 7, 7, 8, 13 and 11 days, respectively. Shoot apices and proximal regions are indicated with arrows and arrowheads, respectively. **(h-j)** Cross sections of leaves from plants grown for 14 days. Representative staining images for three transgenic plants are shown. **(k-m)** Roots from plants grown for 7, 14 and 14 days, respectively. Arrows indicate lateral root primordia. Sections are 50 or 200 μm thick.

Intracellular localization of the *OsMCA1* protein

To investigate intracellular localization of the *OsMCA1* protein, we introduced a green fluorescent protein (GFP) construct fused to the coding sequence of the N-terminus of *OsMCA1* into tobacco BY-2 cells and examined its intracellular localization using confocal laser scanning microscopy. When GFP alone was expressed, it localized to the nucleus and the cytoplasm (Figure 2i, j). In contrast, GFP-*OsMCA1* fusion protein

localized specifically to the plasma membrane (Figure 2a, b). This pattern was reinforced by treatment with a high osmotic solution, 1 M mannitol, which induced plasmolysis (Figure 2e, f). In addition, fluorescent images and behavior of GFP-*OsMCA1* before and after plasmolysis were different from those of the intracellular staining marker FM4-64 (Figure 2c, g, k and 2d, h, l), indicating that *OsMCA1* is localized to the plasma membrane.



Effects of *OsMCA1* overexpression on Ca^{2+} uptake in cultured rice cells

To test if *OsMCA1* plays a role in Ca^{2+} transport, we generated cultured cells overexpressing *OsMCA1* and analyzed whether expression levels of *OsMCA1* affected Ca^{2+} uptake activity. As shown in Figure 3A, B, Ca^{2+}

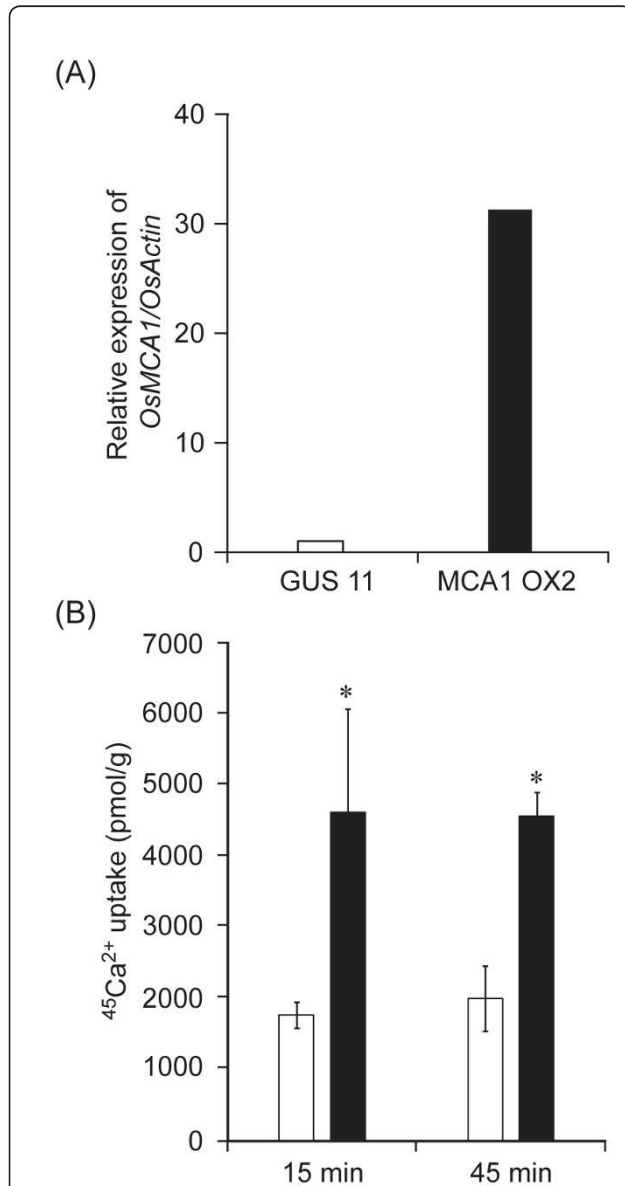


Figure 3 Effect of *OsMCA1*-overexpression on Ca^{2+} uptake in cultured rice cells. Open and closed bars represent *GUS*-expressing control line (*GUS* No.11) and *OsMCA1*-overexpressing line (OX No. 2), respectively. (A) Quantification of *OsMCA1* mRNA by real-time quantitative PCR. The amount of *OsMCA1* mRNA was calculated from the threshold point in the log-linear range of the RT-PCR. The relative level of *OsMCA1* mRNAs in the *GUS*-expressing control line was standardized as 1. (B) $^{45}\text{Ca}^{2+}$ uptake into cultured rice cells. Data are means \pm SD, $n = 3$ independent samples. * $P < 0.05$; significantly different compared with the control.

uptake activity was higher in *OsMCA1*-overexpressing cells than in *GUS*-expressing control cells, suggesting that *OsMCA1* is involved in Ca^{2+} uptake across the plasma membrane in rice. We also generated *OsMCA1*-overexpressing plants, which showed no significant visible phenotypes (data not shown).

Effect of *OsMCA1* suppression on growth and development in plants

To elucidate the physiological roles of *OsMCA1*, transgenic plants were generated in which *OsMCA1* expression was suppressed by RNA interference (*RNAi*) using gene-specific sequences (a 400-bp region of *OsMCA1*). Five independent transgenic plants were generated using *Agrobacterium*-mediated transformation. Non-transgenic plants were investigated simultaneously as controls, whose transduced genes were removed by heterozygous segregation. RT-PCR analyses revealed significant reductions in *OsMCA1* mRNA levels compared with controls (Figure 4A).

The *OsMCA1*-suppressed lines showed slower growth in adult plants (Figure 4B, C). Though germination rates (data not shown) and seedling growth of suppression lines were comparable to controls in the Murashige and Skoog medium (MS medium) (Additional file 3), growth of suppression lines was remarkably retarded after transplantation into soil in a greenhouse, suggesting that *OsMCA1* suppression leads to hypersensitivity to environmental stresses. This phenotype was exhibited in all 5 independent T_2 transgenic *RNAi* lines; severity of the phenotypes correlated well with expression levels of *OsMCA1* transcripts (Figure 4A, C). Furthermore, unlike Arabidopsis *mca* mutants, rachises of the *OsMCA1*-suppressed lines were significantly shorter than those of controls (Figure 4D, E), suggesting that *OsMCA1* plays a different role from Arabidopsis MCAs in some developmental stages.

Effects of *OsMCA1*-suppression on cell growth and Ca^{2+} sensitivity in cultured rice cells

We tested whether *OsMCA1* suppression affects Ca^{2+} sensitivity to growth in rice cells. In regular medium containing 3 mM of Ca^{2+} , growth rates of *OsMCA1*-suppressed lines were comparable to controls (Figure 5). In contrast, when Ca^{2+} concentration of the medium was decreased to 0.1 mM (Figure 5), growth of *OsMCA1*-suppressed lines was significantly restricted compared with controls, suggesting possible involvement of *OsMCA1* in acquisition of Ca^{2+} for cell growth under Ca^{2+} limitation.

Involvement of *OsMCA1* in mechanical stress-induced $[\text{Ca}^{2+}]_{\text{cyt}}$ changes

To test for possible involvement of *OsMCA1* in regulation of Ca^{2+} influx induced by various stimuli, we generated *OsMCA1*-suppressed lines harboring the Ca^{2+}

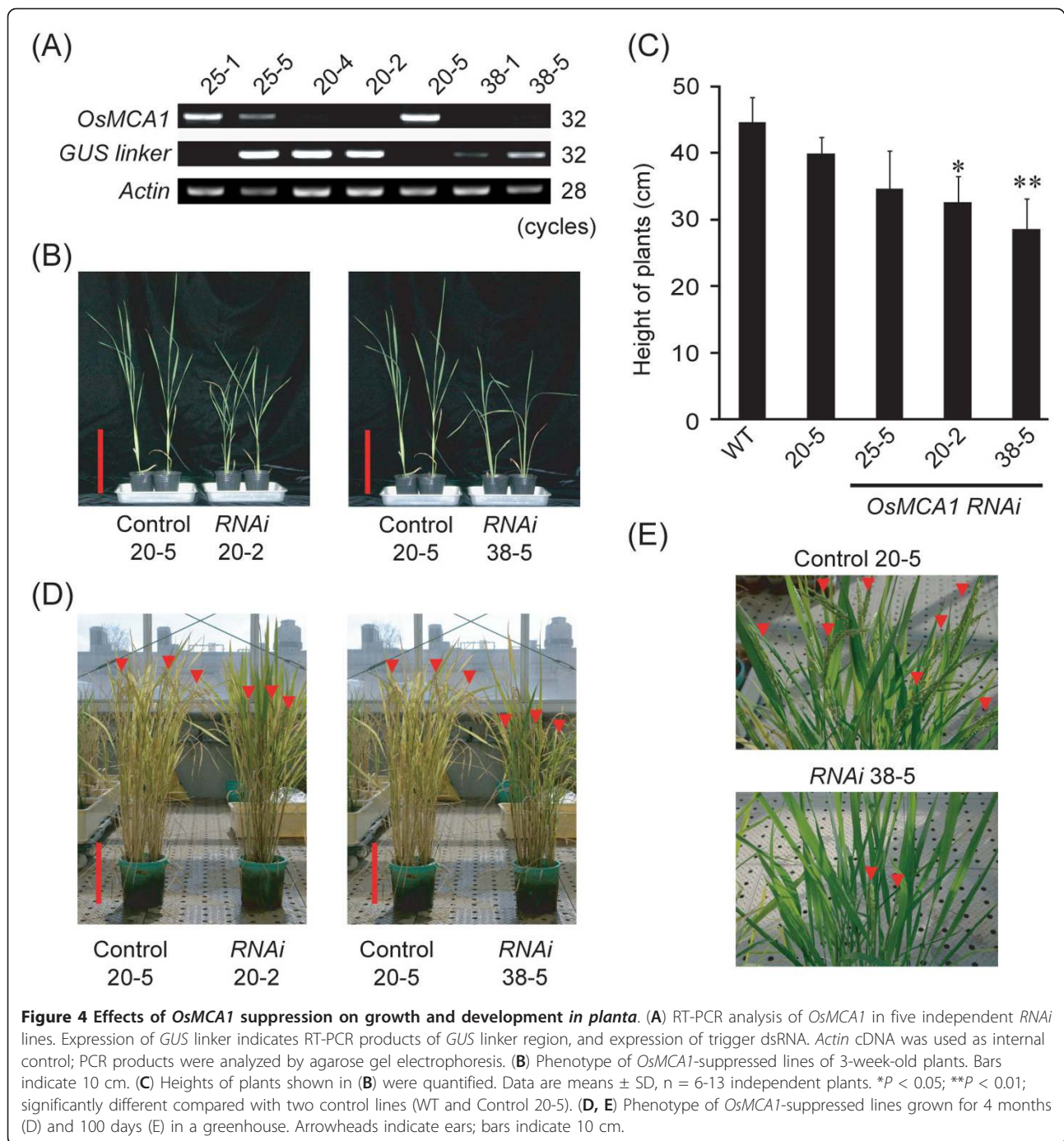
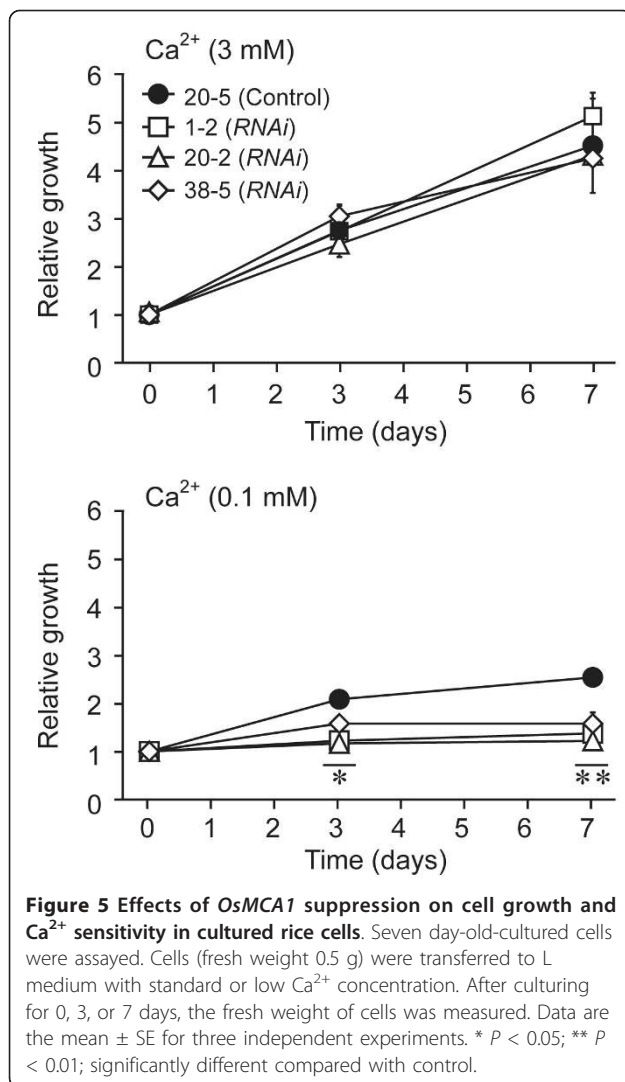


Figure 4 Effects of *OsMCA1* suppression on growth and development in planta. **(A)** RT-PCR analysis of *OsMCA1* in five independent RNAi lines. Expression of *GUS linker* indicates RT-PCR products of *GUS linker* region, and expression of trigger dsRNA. *Actin* cDNA was used as internal control; PCR products were analyzed by agarose gel electrophoresis. **(B)** Phenotype of *OsMCA1*-suppressed lines of 3-week-old plants. Bars indicate 10 cm. **(C)** Heights of plants shown in **(B)** were quantified. Data are means \pm SD, n = 6-13 independent plants. **P* < 0.05; ***P* < 0.01; significantly different compared with two control lines (WT and Control 20-5). **(D, E)** Phenotype of *OsMCA1*-suppressed lines grown for 4 months (D) and 100 days (E) in a greenhouse. Arrowheads indicate ears; bars indicate 10 cm.

⁺-sensitive bioluminescent protein aequorin (Figure 6A). Hypo-osmotic shock-induced transient $[Ca^{2+}]_{cyt}$ change in cultured rice cells (Figure 6B) was inhibited by a Ca^{2+} chelator, 1,2-bis-(2-aminophenoxy)ethane-*N, N, N', N'*-tetra-acetic acid (BAPTA), and Ca^{2+} channel blockers (GdCl₃ and LaCl₃) but not by verapamil, an inhibitor for voltage-dependent Ca^{2+} channels, (Figure 6C), suggesting that plasma membrane Ca^{2+} influx mediated by Gd³⁺ ⁺-sensitive mechanosensitive Ca^{2+} -permeable channel(s)

is induced by hypo-osmotic shock. The hypo-osmotic shock-induced $[Ca^{2+}]_{cyt}$ change was partially impaired in the *OsMCA1*-suppressed cells (Figure 6E), and was proportional to levels of *OsMCA1* expression in various *OsMCA1*-suppressed cells. On the other hand, increased $[Ca^{2+}]_{cyt}$ triggered by Nacetylchito-oligosaccharides, a major microbe-associated molecular pattern (MAMP) recognized by plasma membrane receptors in rice [23,24], was not affected by *OsMCA1* expression levels



(Figure 6D). These results suggest that *OsMCA1* participates in the plasma membrane Ca^{2+} influx triggered by hypo-osmotic shock but not by MAMPs.

Trinitrophenol (TNP) is a potent compound to generate membrane distortion to activate mechanosensitive channels and mimic mechanical stimuli in plants [15] and animal cells. We investigated the effects of TNP on $[\text{Ca}^{2+}]_{\text{cyt}}$ and possible involvement of *OsMCA1* in its regulation in cultured rice cells. TNP induced transient $[\text{Ca}^{2+}]_{\text{cyt}}$ change, which was inhibited by BAPTA, GdCl_3 , and LaCl_3 but not by verapamil (Figure 7A, B). The pharmacology of $[\text{Ca}^{2+}]_{\text{cyt}}$ transients triggered by hypo-osmotic shock and TNP was basically similar (Figure 6C, 7B). TNP-induced $[\text{Ca}^{2+}]_{\text{cyt}}$ change was also impaired in *OsMCA1*-suppressed lines (Figure 7C), suggesting the possible involvement of *OsMCA1* as a putative mechanosensitive Ca^{2+} -permeable channel in the

regulation of mechanical stress-triggered plasma membrane Ca^{2+} influx.

We also tried to examine the effect of overexpression of *OsMCA1* on mechanical stress-triggered Ca^{2+} influx. However, we observed a strong reduction in the total aequorin luminescence in all transgenic cell lines overexpressing *OsMCA1* (data not shown). Thus it was impossible to measure $[\text{Ca}^{2+}]_{\text{cyt}}$ using *OsMCA1*-overexpressing lines. Real-time RT-PCR analysis revealed that the expression level of aequorin gene in the *OsMCA1*-overexpressing lines was comparable to the control (data not shown). Thus constitutive overexpression of *OsMCA1* does not affect the expression but may affect the stability of aequorin protein or inhibit the aequorin chemiluminescence.

Effects of *OsMCA1*-overexpression on sensitivity to hypo-osmotic shock and generation of reactive oxygen species

Hypo-osmotic shock has been shown to trigger ROS generation following $[\text{Ca}^{2+}]_{\text{cyt}}$ increase in cultured tobacco cells [18,22]. As *OsMCA1* has been suggested to affect regulation of hypo-osmotic shock-induced Ca^{2+} influx (Figure 6E), we investigated possible *OsMCA1* involvement in hypo-osmotic shock-induced ROS generation in cultured rice cells, using two distinctive methods sensitive for superoxide anion radical (O_2^-) and hydrogen peroxide (H_2O_2).

Hypo-osmotic shock triggered ROS generation within 5 min (Figure 8A), which was markedly inhibited by Ca^{2+} channel blockers (GdCl_3 and LaCl_3), suggesting an important role of Ca^{2+} influx in hypo-osmotic shock-induced ROS generation (Figure 8B). Diphenylene iodonium (DPI; 10 μM), an NADPH oxidase inhibitor, significantly suppressed ROS generation (Additional file 4). Peroxidase-catalyzed reactions have been also proposed for osmotic shock-induced ROS generation in cultured tobacco and *Arabidopsis* cells [19]. However, a peroxidase inhibitor, salicylhydroxamic acid (SHAM, 3 mM), scarcely affected hypo-osmotic shock-induced ROS generation (Additional file 5), suggesting that a major part of hypo-osmotic shock-induced ROS generation is attributable to ROS-producing NADPH oxidases in cultured rice cells. Hypo-osmotic shock-induced generation of both O_2^- (Figure 8C, D) and H_2O_2 (Additional file 6) was either significantly enhanced or more rapid in *OsMCA1*-overexpressing lines than in the control. Generation of ROS in *OsMCA1*-suppressed lines was comparable to the control (Figure 8A).

Discussion

It has been suggested that Ca^{2+} plays a crucial role in mechanical sensing [7,25]. However, little is known of the molecular mechanisms responsible for Ca^{2+} mobilization. Functional characterization of the *OsMCA1*-

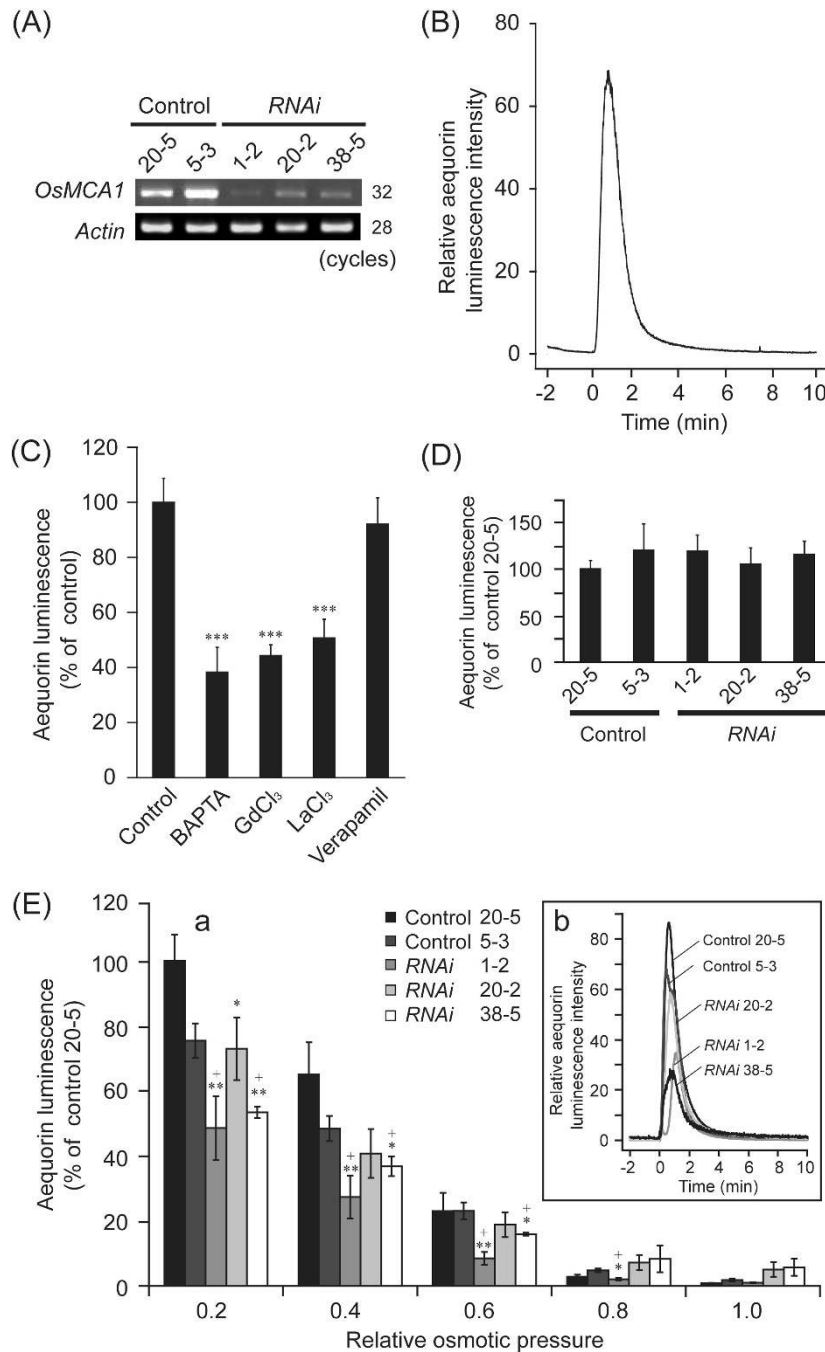


Figure 6 Involvement of *OsMCA1* in the regulation of hypo-osmotic shock-induced $[Ca^{2+}]_{cyt}$ changes. (A) RT-PCR analysis of *OsMCA1* in three independent RNAi lines expressing *apoaequorin* mRNA. (B) Hypo-osmotic shock-induced $[Ca^{2+}]_{cyt}$ changes in cultured rice cells. Cells were diluted four-fold with water at 0 min. A representative result of five experiments is shown. (C) Pharmacological analyses of hypo-osmotic shock-induced $[Ca^{2+}]_{cyt}$ changes. Cells were diluted four-fold with water at 0 min. BAPTA (5 mM), GdCl₃ (5 mM), LaCl₃ (5 mM) and verapamil (1 mM) were added to the cell suspension 15 min before hypo-osmotic shock treatment. No treatment prior to hypo-osmotic shock in the "control." Peak intensities of aequorin chemiluminescence were compared; relative luminescence level in the control was standardized as 100% (C and D). Data are mean \pm SE for five independent experiments. ****P* < 0.005; significantly different compared with control. (D) Effects of *OsMCA1* suppression on *N*-acetylchito-oligosaccharides-induced $[Ca^{2+}]_{cyt}$ changes. Cells were treated with *N*-acetylchitoheptaose (1 μ M). Error bars indicate SE of the mean of five experiments. (E-a) 1 mL of water, medium, or diluted medium was added to *OsMCA1*-suppressed cells harboring *apoaequorin* at 0 min to generate wide-ranging changes in extracellular osmotic pressure. The relative amount of luminescence accumulated for 10 min after hypo-osmotic shock was plotted versus relative extracellular osmotic pressure. Data are means \pm SE, *n* = 3-7. ***P* < 0.05; ****P* < 0.01; significantly different compared with two control lines (No. 20-5 and 5-3). **P* < 0.05; significantly different compared with the control line (No. 20-5). (E-b) Hypo-osmotic shock-induced Ca^{2+} signature in *OsMCA1*-suppressed lines. Cells were diluted four-fold with water at 0 min. A representative result of several experiments is shown.

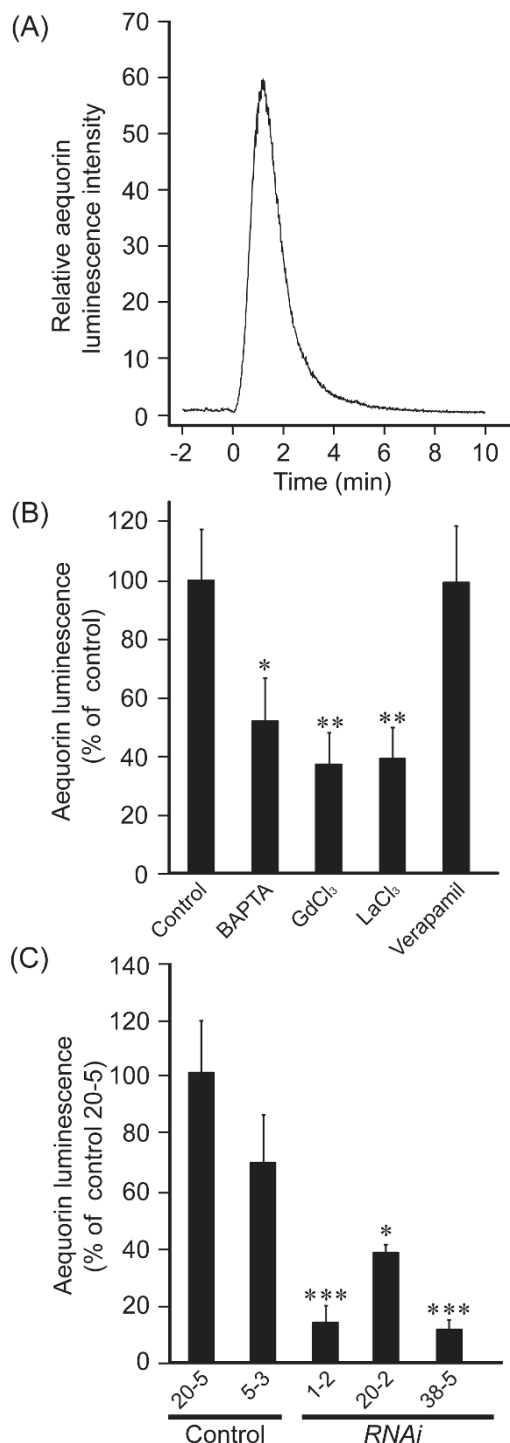


Figure 7 Involvement of OsMCA1 in the regulation of Trinitrophenol (TNP)-induced $[Ca^{2+}]_{cyt}$ changes. (A) TNP-induced $[Ca^{2+}]_{cyt}$ changes in cultured rice cells. Cells were treated with TNP (1 mM) at 0 min. A representative result of five experiments is shown. (B) Pharmacological analyses of TNP-induced $[Ca^{2+}]_{cyt}$ changes. BAPTA (5 mM), GdCl₃ (5 mM), LaCl₃ (5 mM) and verapamil (1 mM) were added to the cell suspension 15 min prior to the TNP treatment; DMSO is used as the control. The peak intensities of

aequorin chemiluminescence were compared; the relative luminescence level in the control was standardized as 100%. Data are the mean \pm SE for three independent experiments. * $P < 0.05$; ** $P < 0.01$; significantly different compared with control. (C) Effects of *OsMCA1*-suppression on TNP-induced $[Ca^{2+}]_{cyt}$ changes. Cells were treated with TNP (1 μ M). The peak intensities of aequorin chemiluminescence were compared; the relative luminescence level in the control line (20-5) was standardized as 100%. Data are the mean \pm SE for three independent experiments. * $P < 0.05$; *** $P < 0.005$; significantly different compared with two control lines (20-5 and 5-3).

RNAi lines as well as overexpressors suggests that *OsMCA1* is involved in hypo-osmotic shock-induced Ca^{2+} influx and ROS generation.

Possible functions of *OsMCA1* in the regulation of growth and development of rice

OsMCA1-suppressed plants displayed stunted growth and shortened rachises (Figure 4D, E). These phenotypes are frequently observed under drought-stress conditions [26]. The suppression of *OsMCA1* might have affected adaptation to drought. Drought stress is known to lead to osmotic stress at a cellular level. Since hypo-osmotic shock-induced $[Ca^{2+}]_{cyt}$ changes were impaired in the *OsMCA1*-suppressed lines, these lines may have defects in osmotic sensing/responses and ability to adapt to drought stress. Future studies to characterize physiological reactions to drought and mechanical signaling in *OsMCA1*-suppressed plants would further elucidate the *in vivo* roles of *OsMCA1* in intact plants.

In *Arabidopsis*, the *mca1 mca2* double mutant shows a growth defect in soil, and reduced accumulation of Ca^{2+} as well as enhanced sensitivity to Mg^{2+} [16]. The balance of Ca^{2+} and Mg^{2+} in soil is an important factor for normal plant growth [27]. Since the growth of the *OsMCA1*-suppressed lines was significantly restricted compared with the control under Ca^{2+} -limitation (Figure 5), growth retardation in the *OsMCA1*-suppressed plants may be attributed to reduced Ca^{2+} uptake, resulting in a low Ca^{2+} - Mg^{2+} ratio.

Possible involvement of *OsMCA1* in osmotic signaling in cultured rice cells

The GFP-*OsMCA1* fusion protein localized specifically to the plasma membrane (Figure 2), suggesting that *OsMCA1* is a plasma membrane protein. In cultured rice cells, both hypo-osmotic shock- and TNP-induced $[Ca^{2+}]_{cyt}$ transients, which were inhibited by La^{3+} and Gd^{3+} , were impaired in *OsMCA1*-suppressed lines (Figures 6 and 7). The temporal pattern of the MAMP-induced $[Ca^{2+}]_{cyt}$ transient was similar (data not shown), but was unaffected by *OsMCA1* suppression (Figure 6D). These results suggest that *OsMCA1* affects regulation of Ca^{2+}

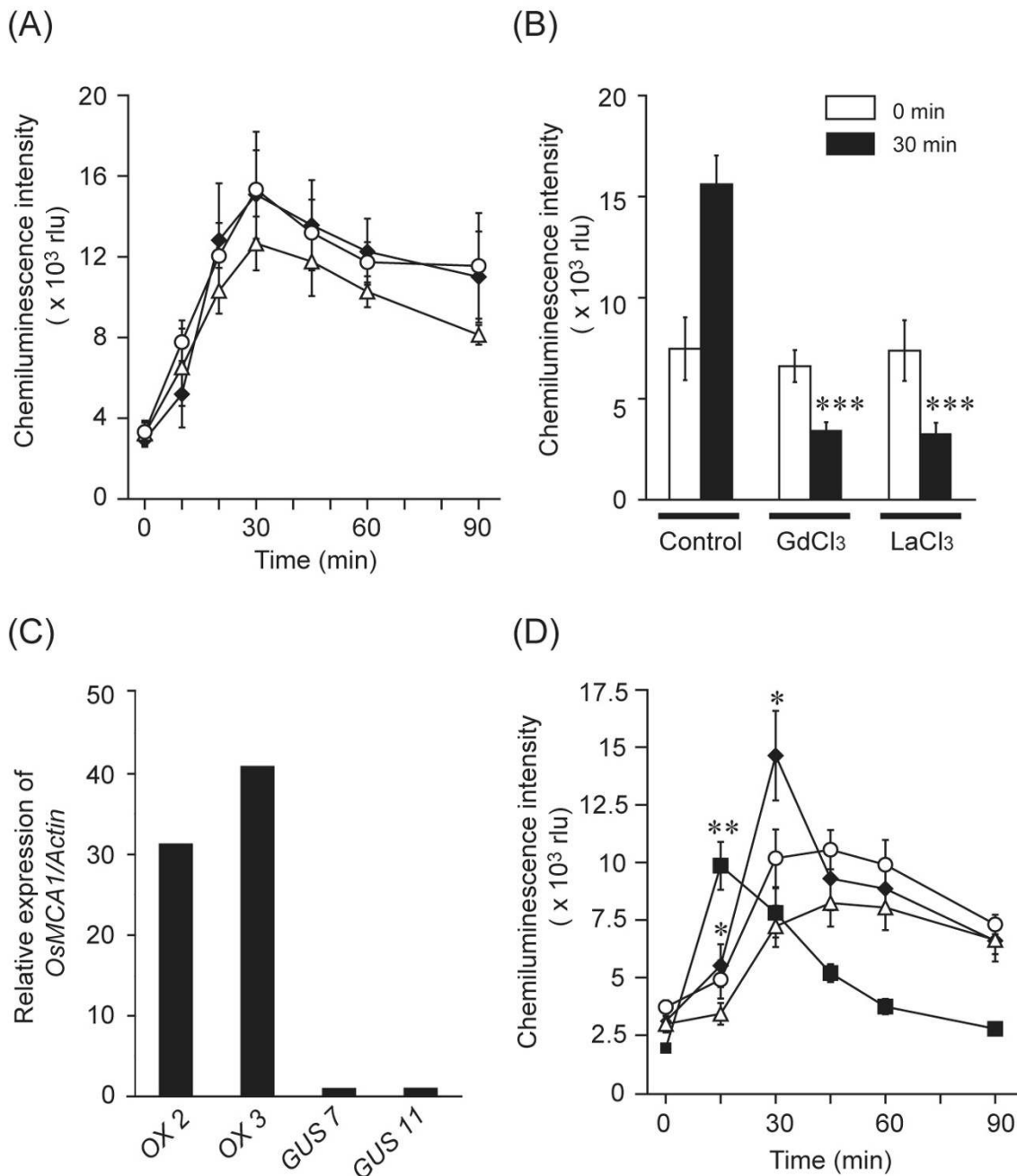


Figure 8 Effects of the expression level of *OsMCA1* on hypo-osmotic shock-induced ROS generation. (A) Effects of *OsMCA1*-suppression on hypo-osmotic shock-induced ROS generation measured by MCLA chemiluminescence. Data are the mean \pm SE for five independent experiments for the control line (closed diamond for control 20-5) and two independent *RNAi* lines (open circle for *RNAi* 20-2; open triangle for *RNAi* 38-5) are shown. As a hypo-osmotic shock, growth medium was replaced by four-fold diluted medium at 0 min (A and B). (B) The effect of several Ca²⁺ channel blockers on hypo-osmotic shock-induced ROS generation in the wild type. GdCl₃ (1 mM) and LaCl₃ (1 mM) were added to the cells 60 min prior to hypo-osmotic shock treatment. Data are the mean \pm SE for three or four independent experiments. ****P* < 0.005; significantly different compared with the control. (C) Quantitative expression levels of *OsMCA1* mRNAs in the *OsMCA1* overexpressor lines by real-time quantitative PCR. The relative level of the *OsMCA1* mRNAs in the control cells (GUS No. 7) was standardized as 1. (D) Effects of *OsMCA1* overexpression on hypo-osmotic shock-induced ROS generation. As a hypo-osmotic shock, growth medium was replaced by three-fold diluted medium at 0 min. Data are the mean \pm SE for five independent experiments for two control lines (open circle for GUS No. 11; open triangle for GUS No. 7) and two overexpressor lines (closed diamond for OX No. 2; closed square for OX No. 3) are shown. **P* < 0.05; ****P* < 0.01; significantly different compared with the control line (GUS No. 7).

influx across the plasma membrane in response to mechanical stimulation in cultured rice cells.

Hypo-osmotic shock triggers ROS generation following a $[Ca^{2+}]_{cyt}$ increase [18,20,22]. Extracellular Ca^{2+} is required for both Ca^{2+} influx and NADPH oxidase-mediated ROS generation induced by hypo-osmotic shock (Figures 6C and 8B, Additional file 4), suggesting that ROS generation requires Ca^{2+} influx across the plasma membrane. Overexpression of *OsMCA1* enhances ROS generation (Figure 8D, Additional file 6). Binding Ca^{2+} to the EF-hand regions of cytosolic regulatory domains of plant NADPH oxidases directly activates them [28-30]. A functional NADPH oxidase *AtrbohC/RHD2* reportedly affects mechanical stress-induced ROS generation in a Ca^{2+} -dependent manner [31]. Overproduction of the plasma membrane Ca^{2+} -permeable channels may induce the mobilization of excess Ca^{2+} in response to mechanical stimuli, which may cause enhanced activation of NADPH oxidases.

In *OsMCA1*-suppressed lines challenged with hypo-osmotic shock, Ca^{2+} influx was partially impaired (Figure 6E), but no significant influence of the impairment on subsequent ROS generation was detected under our assay conditions (Figure 8A). Similar effects of overexpression and a loss-of-function mutation were also observed with another putative Ca^{2+} -permeable channel, *OsTPC1* [32]. A certain level of Ca^{2+} increment may be sufficient for NADPH oxidase-mediated ROS generation. Alternatively, other Ca^{2+} -permeable channels activated by hypo-osmotic shock may redundantly play a role in bypassing *OsMCA1*. It has been suggested that *Arabidopsis* *MSL9* and *MSL10*, homologs of the bacterial mechanosensitive channel *MscS*, are required for mechanosensitive channel activity in root cell plasma membranes, and are able to translocate cations including Ca^{2+} [13,14]. Rice *MSL* homologs may therefore be candidates for such Ca^{2+} -permeable channels.

Conclusions

The present study indicates that *OsMCA1* is involved in regulation of plasma membrane Ca^{2+} influx and NADPH oxidase-mediated ROS generation induced by hypo-osmotic stress in cultured rice cells. These findings shed light on our understanding of mechanical sensing pathways.

Methods

Plant materials and cell culture

Surface-sterilized seeds of rice, *Oryza sativa* L. cv. Nipponbare, were germinated on MS medium [33] containing 0.8% agar and grown for 10 days in a growth chamber under long day conditions (16 h light/8 h darkness, 28°C). Seedlings were transplanted into soil and grown in a greenhouse (16 h light/8 h darkness, 28°C

and 60% humidity). Calli were suspension-cultured at 25°C in a liquid L medium [34] containing 2,4-D (0.5 mg L⁻¹) in the dark and subcultured in fresh medium every week. Cells were filtered through a 20-mesh screen every 2 weeks to make fine aggregates. Cells at 5 days after subculture were used for experiments with osmotic stress and defense responses. *N*-acetylchito-oligosaccharides were kindly provided by Prof. Naoto Shibuya (Meiji University).

Isolation of *OsMCA1* cDNA

The estimated coding region of *OsMCA1* was amplified by PCR using two primers: *OsMCA1* forward, 5'-GAA-GAAGAAGAAGAAGAAGAAGCCGAGTAG-3'; *OsMCA1* reverse, 5'-TATTTATGCTTACCCTG-CATTGTTTTGTGTT-3'. Total RNA was isolated from rice leaves using Trizol (Invitrogen, Carlsbad, CA, USA) in accordance with manufacturer's protocol and quantified spectrophotometrically. First-strand cDNA was synthesized from 3 µg of total RNA with the oligo-dT primer and reverse transcriptase (Invitrogen). To obtain full-length cDNAs for *OsMCA1* and to define the open reading frame, 3'-RACE PCR and 5'-RACE PCR were performed with a 3'-full RACE core kit (Takara, Ohtsu, Japan) and a 5'-RACE system (Invitrogen) in accordance with manufacturers' protocols.

RNA isolation and RT-PCR analyses

Total RNA was isolated using Trizol reagent in accordance with manufacturer's protocol and quantified using a spectrophotometer. First-strand cDNA was synthesized from 3 µg total RNA with an oligo-dT primer and reverse transcriptase. PCR amplification was performed with an initial denaturation at 95°C for 3 min followed by indicated numbers of cycles of incubations at 94°C for 30 s, 55°C for 90 s, and 72°C for 1 min by using specific primers for *OsMCA1*. *Actin* was used as a quantitative control [35]. Aliquots of individual PCR products were resolved by agarose gel electrophoresis and visualized using ethidium bromide staining and exposure to UV light.

Real-time RT-PCR quantification

Real-time RT-PCR assays were performed as described by Kurusu et al. (2010) [36]. First-strand cDNA was synthesized from 3 µg of total RNA using an oligo-dT primer and reverse transcriptase. Real-time PCR was performed using an ABI PRISM 7300 sequence detection system (Applied Biosystems, Foster City, CA, USA) with SYBR Green real-time PCR Master Mix (Toyobo, Osaka, Japan) and *OsMCA1* specific primers *OsMCA1*-RealF, 5'-TGGTCTCAAGCAGAGGATCATACA-3'; *OsMCA1*-RealR, 5'-CTCTGAACAGCAACCAAG-CAAA-3'. Relative mRNA levels were calculated using

the standard curve method and normalized to the corresponding *OsActin1* gene levels. Standard samples of known template amounts were used to quantify PCR products.

Spatial pattern of *OsMCA1* expression using *OsMCA1p::GUS*-expressing plants

A DNA fragment of the *OsMCA1* promoter region was prepared using PCR by synthesizing the 5'-non-coding region spanning -1.5 to 0 kb from the *OsMCA1* initiation codon, using rice (Nipponbare) genomic DNA as a template and the following primers: *OsMCA1pF*, 5'-CACCACAACCCCTAACATGCCTAA-3'; *OsMCA1pR*, 5'-TGCCGTCGTCTACTCGGCTTCTTCT-3' (the CACC sequence used with the Gateway system), subcloned into a pENTR/D-TOPO cloning vector, and then cloned into a Ti-based promoterless *GUS* expression vector, pHGWF57 [37] using the LR clonase reaction; *Agrobacterium*-mediated transformation of rice calli was performed. Transformed calli were screened by hygromycin selection (50 µg mL⁻¹); transgenic plants were then regenerated.

The T₂ transgenic plants were grown at 28°C under a 16 h light/8 h dark cycle for experiments. Histochemical localization of *GUS* activity in situ was performed as follows. Samples were fixed for 1 h with 90% acetone in Eppendorf tubes placed on ice and washed four times with 100 mM sodium phosphate buffer, pH 7.0. Samples were then incubated for 24 h at 37°C in X-Gluc buffer (0.5 mg/mL 5-bromo-4-chloro-3-indolyl glucuronidase (Nacalai Tesque, Osaka, Japan), 50 mM sodium phosphate buffer, pH 7.0, 5% methanol), and then cleaned and fixed by rinsing for 1 h each with 50%, 70%, 90%, and 100% (v/v) ethanol successively. Fixed samples were stored in 100% ethanol before being photographed.

Generation of *OsMCA1*-overexpressing and suppressed lines

To generate RNA-silencing-triggered inverted repeat constructs, a region corresponding to 400 bp of the 3'-UTR of *OsMCA1* was amplified using RNAiFW, 5'-CACC CTCTTATCCAACTTGCCAT-3' and RNAiRV, 5'-AATGTTCCACAGGGGAAAAAGAATGTTCTC-3' as specific primers, subcloned into a pENTR/D-TOPO cloning vector, and cloned into a Ti-based RNAi vector, pANDA [38] using the LR clonase reaction. The construct was introduced into rice calli using *Agrobacterium*-mediated transformation, according to the method of Tanaka et al. 2001 [39]. Transformed calli were screened by hygromycin selection (50 µg mL⁻¹); transgenic plants were then regenerated. Transgenic cell lines derived from T₂ plants were used for various analyses.

To overexpress *OsMCA1* and *GUS* cDNAs, sequences were cloned into a Ti-based vector pPZP2H-lac [40]

downstream of the maize *Ubiquitin* promoter, and *Agrobacterium*-mediated transformation of rice calli was performed. Transformed calli were screened by hygromycin selection (50 µg mL⁻¹), followed by the regeneration of transgenic plants.

To express cytoplasm-targeted *apoaequorin* cDNA [41] in *OsMCA1*-suppressed plants, sequences were cloned into a Ti-based vector pSMAB704 [42] downstream of a *CaMV 35S* promoter, and *Agrobacterium*-mediated transformation of rice calli was performed. Transformed calli were screened using bialaphos (Meiji Seika, Tokyo, Japan) selection (5 µg mL⁻¹), followed by the regeneration of transgenic plants.

Subcellular localization of *OsMCA1* in tobacco BY-2 cells

To generate transgenic BY-2 cells expressing GFP-*OsMCA1*, the coding region was amplified using *OsMCA1(GFP)F*, 5'-CACCATGGCGTCGTGGGA-GAACCT-3' and *OsMCA1(GFP)R*, 5'-TTAGTGTTCCATGTACTGAA-3' as specific primers, subcloned into a pENTR/D-TOPO cloning vector, and then cloned into a pH7WGF2 vector encoding a N-terminal EGFP fusion [37] using the LR clonase reaction.

Transformation of BY-2 cells was carried out in accordance with An (1985) [43] with minor modifications as follows: 4 mL of 3-day-old exponentially growing culture was transferred to 90-mm Petri dishes and incubated at 28°C with 100 µL of fresh overnight-culture of *Agrobacterium tumefaciens* pGV2260 containing the binary vector pH7WGF2. After a 48-h co-cultivation, the tobacco cells were washed and plated on to LS agar medium containing hygromycin (50 µg mL⁻¹) and carbenicillin (250 µg mL⁻¹). Every 3-4 weeks, transformants were selected and transferred onto fresh medium for continued selection.

The fluorescent styryl membrane probe FM4-64 (Molecular Probes, Carlsbad, CA, USA) was kept as a 17 mM stock solution in sterile water, and used at a final concentration of 4.25 µM to label the vacuolar membrane (tonoplast). Five-day-old BY-2 cells were treated with FM4-64 for 3 h and washed twice with culture medium.

Measurement of cytosolic Ca²⁺ concentration

Measurements of Ca²⁺ mobilization were made in accordance with the method described by Kurusu et al. (2011) [44]. Briefly, *apoaequorin*-expressing rice cells (5 day after subculture) were incubated with 1 µM coelenterazine for at least 12 h at 25°C. Cell suspension (250 µL) was transferred to 1.1-cm-diameter culture tubes, and set in a luminometer (Lumicounter 2500, Microtech Niton, Chiba, Japan). In the luminometer, culture tubes rotated 17 revolutions every 3 s clockwise and counter-clockwise in turn, agitating the cells. After a 15-min

incubation to stabilize the cells, Ca^{2+} -dependent aequorin luminescence was measured and expressed as relative luminescence units (rlu).

Ca^{2+} uptake in cultured cells

Rice cells 5 days after subculture were used to measure Ca^{2+} uptake. The rice cells were incubated in Ca^{2+} -free medium for at least 3 h at 25°C. The cell suspension (80 mg mL⁻¹) was transferred to medium containing 0.1 mM CaCl_2 and incubated for 1 h. Ca^{2+} uptake was initiated by adding ⁴⁵ CaCl_2 solution to a final concentration of 33 kBq/g. Cells were then agitated at 25°C; 1 mL of cells was collected at 0, 15 and 45 min after the addition of ⁴⁵ CaCl_2 . Cells were filtered using Whatman filters (GF/C) presoaked with 5 mM CaCl_2 and washed 5 times with an ice-cold solution of 5 mM CaCl_2 , and 2 mM LaCl_3 to remove ⁴⁵ Ca^{2+} from cell walls. Radioactivity retained on each filter was counted as described previously [45].

Measurement of ROS

Rice cells (cv. Nipponbare) 5 days after subculture were used for measurement of $\cdot\text{O}_2^-$ and H_2O_2 in the extracellular medium. $\cdot\text{O}_2^-$ -dependent chemiluminescence was monitored in growth medium supplemented with 20 μM methoxylated cypridina luciferin analog (MCLA (2-methyl-6-[*p*-methoxyphenyl]-3,7-dihydroimidazo [1,2- α]pyrazin-3-one); Invitrogen) using a luminometer (Lumi-counter 2500) under the same conditions as for measuring $[\text{Ca}^{2+}]_{\text{cyt}}$ [46].

To monitor H_2O_2 produced in extracellular medium, cells (80 mg mL⁻¹) were washed and resuspended in 5 mM MES buffer (pH 7.0) containing 0.5 mM CaCl_2 , 0.5 mM K_2SO_4 and with or without 175 mM mannitol (Kurusu et al. 2005). A 25- μL aliquot of medium was mixed in a 96-well microtiter plate with 150 μL 50 mM Tris-HCl (pH 8.0) and 25 μL 0.462 mM luminol in 50 mM Tris-HCl, pH 8.0. Potassium ferricyanide (25 μL , 11.76 mM) was added, and H_2O_2 -dependent chemiluminescence was recorded for 15 s using a luminometer (MicroLumat Plus LB96V, Berthold Technologies, Bad Wildbad, Germany).

Statistical analysis

Statistical significance was determined using an unpaired Student's *t* test; *P* < 0.05 indicated significance.

Additional material

Additional file 1: Multiple amino acid sequence alignment of rice *OsMCA1* and *Arabidopsis MCA1* and *MCA2* using Clustal W. Asterisks indicate identical or conserved residues in the whole sequence in the alignment. Colons indicate conserved substitutions. Dots indicate semi-conserved substitutions. Light and dark gray box indicate the PLAC8

motif and coiled-coil motif, respectively. Two putative transmembrane segments (S1 and S2) are underlined.

Additional file 2: Expression of the *OsMCA1* gene in rice tissues. The expression of *OsMCA1* in rice plants was determined by quantitative RT-PCR analysis. Total RNA was extracted from various tissues of rice plants as well as cultured cells. The amount of *OsMCA1* mRNA was calculated from the threshold point in the log-linear range of the RT-PCR. The relative *OsMCA1* mRNA level in cultured cells was standardized as 1. Data are means \pm SD; *n* = 2-3 independent samples.

Additional file 3: Growth phenotype of *OsMCA1*-suppressed seedlings in MS medium. Length of roots and shoots of the control line (20-5) and the *OsMCA1*-suppressed lines (20-2 and 38-5) of 10-days-old seedlings grown on MS medium plate in a growth chamber under long-day conditions (16 h light/8 h darkness, 28°C). Data are means \pm SD; *n* = 7-10 independent seedlings.

Additional file 4: Effect of NADPH oxidase inhibitor on hypo-osmotic shock-induced ROS generation. H_2O_2 concentration in extracellular medium was determined by ferricyanide-catalyzed oxidation of luminol. Diphenylene iodonium (DPI; 10 μM) was added to the rice cells 30 min before hypo-osmotic shock treatment. Data are the mean \pm SE for five independent experiments for the wild type. ****P* < 0.005; significantly different compared with the control.

Additional file 5: Effect of salicylhydroxamic acid, a peroxidase inhibitor, on hypo-osmotic shock-induced ROS generation. The concentration of $\cdot\text{O}_2^-$ in extracellular medium was measured by MCLA chemiluminescence. Salicylhydroxamic acid (SHAM; 3 mM) was added to the rice cells 30 min before hypo-osmotic shock treatment. Average values and SE of three independent experiments for the wild type.

Additional file 6: Effect of *OsMCA1*-overexpression on hypo-osmotic shock-induced ROS generation. H_2O_2 concentration in the extracellular medium was determined by ferricyanide-catalyzed oxidation of luminol. As a hypo-osmotic shock, growth medium was replaced by three-fold diluted medium at 0 min. Data are the mean \pm SE for four independent experiments for two control lines (open circle for GUS No. 11; open triangle for GUS No. 7) and two overexpressor lines (closed diamond for OX No. 2; closed square for OX No. 3) are shown. **P* < 0.05, significantly different compared with two control lines (GUS No. 7 and 11).

Abbreviations

BAPTA: 1,2-bis-(2-aminophenoxy)ethane-*N, N, N', N'*-tetra acetic acid; $[\text{Ca}^{2+}]_{\text{cyt}}$: cytosolic free Ca^{2+} concentration; DIC: differential interference contrast; DPI: diphenylene iodonium; EGTA: ethylene glycol-bis-(2-aminoethyl ether)-*N, N, N', N'*-tetra acetic acid; GFP: green fluorescent protein; GUS: β -glucuronidase; H_2O_2 : hydrogen peroxide; MAMP: microbe-associated molecular pattern; MCLA: 2-methyl-6-[*p*-methoxyphenyl]-3,7-dihydroimidazo [1,2- α]pyrazin-3-one; $\cdot\text{O}_2^-$: superoxide anion radical; MS medium: Murashige and Skoog medium; RACE: rapid amplification of cDNA ends; rlu: relative luminescence units; *RNAi*: RNA interference; RT: reverse transcriptase; ROS: reactive oxygen species; SHAM: salicylhydroxamic acid; TNP: trinitrophenol.

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Authors' contributions

TK, DN, and YY carried out most of the experiments and data analyses. TK and KK designed the study and wrote the manuscript. DN, YY and MG participated in confocal imaging analyses. MN, TY and KI carried out Ca²⁺ uptake experiments in yeast. HH and HS participated in constructing transgenic lines and PCR analyses. YN, KS, HI participated in the design of the study and critically revised the manuscript. All authors read and approved the final manuscript.

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References

1. Reddy ASN: **Calcium: Silver bullet in signaling.** *Plant Sci* 2001, **160**:381-404.
2. Sanders D, Pelloux J, Brownlee C, Harper JF: **Calcium at the crossroads of signaling.** *Plant Cell* 2002, **14**:S401-S417.
3. Yang T, Poovaiah BW: **Calmodulin-mediated signal network in plants.** *Trends Plant Sci* 2003, **8**:505-512.
4. Fasano JM, Massa GD, Gilroy S: **Ionic signaling in plant responses to gravity and touch.** *J Plant Growth Regul* 2002, **21**:71-88.
5. Braam J: **Genome-wide identification of touch- and darkness-regulated Arabidopsis genes: a focus on calmodulin-like and XTH genes.** *New Phytol* 2005, **165**:373-389.
6. Toyoda M, Furuichi T, Tatsumi H, Sokabe M: **Cytoplasmic calcium increases in response to change in the gravity vector in hypocotyls and petioles of Arabidopsis seedling.** *Plant Physiol* 2008, **146**:505-514.
7. Dodd AN, Kudla J, Sanders D: **The language of calcium signaling.** *Annu Rev Plant Biol* 2010, **2**:593-620.
8. Véry AA, Sentenac H: **Cation channels in the Arabidopsis plasma membrane.** *Trends Plant Sci* 2002, **7**:168-175.
9. White PJ, Bowen HC, Demidchik V, Nichols C, Davies JM: **Genes for calcium-permeable channels in the plasma membrane of plant root cells.** *Biochim Biophys Acta* 2002, **1564**:299-309.
10. Dutta R, Robinson KR: **Identification and characterization of stretch-activated ion channels in pollen protoplasts.** *Plant Physiol* 2004, **135**:1398-1406.
11. Qi Z, Kishigami A, Nakagawa Y, Iida H, Sokabe M: **A mechanosensitive anion channel in Arabidopsis thaliana mesophyll cells.** *Plant Cell Physiol* 2004, **45**:1704-1708.
12. Telewski FW: **A unified hypothesis of mechanoperception in plants.** *Am J Bot* 2006, **93**:1466-1476.
13. Haswell ES, Peyronnet R, Barbier-Brygoo H, Meyerowitz EM, Frachisse JM: **Two MscS homologs provide mechanosensitive channel activities in the Arabidopsis root.** *Curr Biol* 2008, **18**:730-734.
14. Peyronnet R, Haswell ES, Barbier-Brygoo H, Frachisse JM: **AtMSL9 and AtMSL10: Sensors of plasma membrane tension in Arabidopsis roots.** *Plant Signal Behav* 2008, **3**:726-729.
15. Nakagawa Y, Katagiri T, Shinozaki K, Qi Z, Tatsumi H, Furuichi T, Kishigami A, Sokabe M, Kojima I, Sato S, Kato T, Tabata S, Iida K, Terashima A, Nakano M, Ikeda M, Yamanaka T, Iida H: **Arabidopsis plasma membrane protein crucial for Ca²⁺ influx and touch sensing in roots.** *Proc Natl Acad Sci USA* 2007, **104**:3639-3644.
16. Yamanaka T, Nakagawa Y, Mori K, Nakano M, Imamura T, Kataoka H, Terashima A, Iida K, Kojima I, Katagiri T, Shinozaki K, Iida H: **MCA1 and MCA2 that mediate Ca²⁺ uptake have distinct and overlapping roles in Arabidopsis.** *Plant Physiol* 2010, **152**:1284-1296.
17. Zingarelli L, Marré MT, Massardi F, Lado P: **Effects of hyperosmotic stress on K⁺ fluxes, H⁺ extrusion, transmembrane electric potential difference and comparison with the effects of fusaric acid.** *Physiol Plant* 1999, **106**:287-295.
18. Beffagna N, Buffoli B, Busi C: **Modulation of reactive oxygen species production during osmotic stress in Arabidopsis thaliana cultured cells: involvement of the plasma membrane Ca²⁺-ATPase and H⁺-ATPase.** *Plant Cell Physiol* 2005, **46**:1326-1339.
19. Rouet MA, Mathieu Y, Barbier-Brygoo H, Laurière C: **Characterization of active oxygen-producing proteins in response to hypo-osmolarity in tobacco and Arabidopsis cell suspensions: identification of a cell wall peroxidase.** *J Exp Bot* 2006, **57**:1323-1332.
20. Hayashi T, Harada A, Sakai T, Takagi S: **Ca²⁺ transient induced by extracellular changes in osmotic pressure in Arabidopsis leaves: differential involvement of cell wall-plasma membrane adhesion.** *Plant Cell Environ* 2006, **29**:661-672.
21. Moran N: **Osmoregulation of leaf motor cells.** *FEBS Lett* 2007, **581**:2337-2347.
22. Cazalé AC, Rouet-Mayer MA, Barbier-Brygoo H, Mathieu Y, Laurière C: **Oxidative burst and hypoosmotic stress in tobacco cell suspensions.** *Plant Physiol* 1998, **116**:659-669.
23. Kaku H, Nishizawa Y, Ishii-Minami N, Akimoto-Tomiyama C, Dohmae N, Takio K, Minami E, Shibuya N: **Plant cells recognize chitin fragments for defense signaling through a plasma membrane receptor.** *Proc Natl Acad Sci USA* 2006, **103**:11086-11091.
24. Miya A, Albert P, Shinya T, Desaki Y, Ichimura K, Shirasu K, Narusaka Y, Kawakami N, Kaku H, Shibuya N: **CERK1, a LysM receptor kinase, is essential for chitin elicitor signaling in Arabidopsis.** *Proc Natl Acad Sci USA* 2007, **104**:19613-19618.
25. Trewavas A, Knight M: **Mechanical signalling, calcium and plant form.** *Plant Mol Biol* 1994, **26**:1329-1341.
26. Ji XM, Raveendran M, Oane R, Ismail A, Lafitte R, Bruskiwich R, Cheng SH, Bennett J: **Tissue-specific expression and drought responsiveness of cell-wall invertase genes of rice at flowering.** *Plant Mol Biol* 2005, **59**:945-964.
27. Brady KU, Kruckeberg AR, Bradshaw HD Jr: **Evolutionary ecology of plant adaptation to serpentine soils.** *Annu Rev Ecol Syst* 2005, **36**:243-266.
28. Ogasawara Y, Kaya H, Hiraoka G, Yumoto F, Kimura S, Kadota Y, Hishinuma H, Senzaki E, Yamagoe S, Nagata K, Nara M, Suzuki K, Tanokura M, Kuchitsu K: **Synergistic activation of the Arabidopsis NADPH oxidase AtrobohD by Ca²⁺ and phosphorylation.** *J Biol Chem* 2008, **283**:8885-8892.
29. Takeda S, Gapper C, Kaya H, Bell E, Kuchitsu K, Dolan L: **Local positive feedback regulation determines cell shape in root hair cells.** *Science* 2008, **319**:1241-1244.
30. Kimura S, Kaya H, Kawarazaki T, Hiraoka G, Senzaki E, Michikawa M, Kuchitsu K: **Protein phosphorylation is a prerequisite for the Ca²⁺-dependent activation of Arabidopsis NADPH oxidases and may function as a trigger for the positive feedback regulation of Ca²⁺ and reactive oxygen species.** *Biochim Biophys Acta* 2011, **1823**:398-405.
31. Monshausen GB, Bibikova TN, Weisenseel MH, Gilroy S: **Ca²⁺ regulates reactive oxygen species production and pH during mechanosensing in Arabidopsis roots.** *Plant Cell* 2009, **21**:2341-2356.
32. Kurusu T, Yagala T, Miyao A, Hirochika H, Kuchitsu K: **Identification of a putative voltage-gated Ca²⁺ channel as a key regulator of elicitor-induced hypersensitive cell death and mitogen-activated protein kinase activation in rice.** *Plant J* 2005, **42**:798-809.
33. Murashige T, Skoog F: **A revised medium for rapid growth and bioassays with tobacco tissue cultures.** *Physiol Plant* 1962, **15**:473-497.
34. Kuchitsu K, Kikuyama M, Shibuya N: **N-acetylchitooligosaccharides, biotic elicitors for phytoalexin production, induce transient membrane depolarization in suspension-cultured rice cells.** *Protoplasma* 1993, **174**:79-81.
35. Kojima S, Takahashi Y, Kobayashi Y, Monna L, Sasaki T, Araki T, Yano M: **Hd3a, a rice ortholog of the Arabidopsis FT gene, promotes transcription to flowering downstream of Hd1 under short-day conditions.** *Plant Cell Physiol* 2002, **43**:1096-1105.
36. Kurusu T, Hamada J, Nokajima H, Kitagawa Y, Kiyoduka M, Takahashi A, Hanamata S, Ohno R, Hayashi T, Okada K, Koga J, Hirochika H, Yamane H, Kuchitsu K: **Regulation of microbe-associated molecular pattern-induced hypersensitive cell death, phytoalexin production and defense gene expression by calcineurin B-like protein-interacting protein kinases, OsCIPK14/15, in rice cultured cells.** *Plant Physiol* 2010, **153**:678-692.

37. Karimi M, Inzé D, Depicker A: **GATEWAY vectors for Agrobacterium-mediated plant transformation.** *Trends Plant Sci* 2002, **7**:193-195.
38. Miki D, Shimamoto K: **Simple RNAi vectors for stable and transient suppression of gene function in rice.** *Plant Cell Physiol* 2004, **45**:490-495.
39. Tanaka H, Kayano T, Ugaki M, Shiobara F, Onodera H, Ono K, Tagiri A, Nishizawa Y, Shibuya N: **Ultra-fast transformation technique for monocotyledons.** International Patent Application. 2001, No. WO 01/06844 A1.
40. Fuse T, Sasaki T, Yano M: **Ti-plasmid vectors useful for functional analysis of rice genes.** *Plant Biotechnol* 2001, **18**:219-222.
41. Knight MR, Campbell AK, Smith SM, Trewavas AJ: **Transgenic plant aequorin reports the effects of touch and cold-shock and elicitors on cytoplasmic calcium.** *Nature* 1991, **352**:524-526.
42. Igasaki T, Ishida Y, Mohri T, Ichikawa H, Shinohara K: **Transformation of *Populus alba* and direct selection of transformants with the herbicide bialaphos.** *Bulletin of FFPRI* 2002, **1**:235-240.
43. An G: **High efficiency transformation of cultured tobacco cells.** *Plant Physiol* 1985, **79**:568-570.
44. Kurusu T, Hamada H, Sugiyama Y, Yagala T, Kadota Y, Furuichi T, Hayashi T, Umemura K, Komatsu S, Mlyao A, Hirochika H, Kuchitsu K: **Negative feedback regulation of microbe-associated molecular pattern-induced cytosolic Ca²⁺ transients by protein phosphorylation.** *J Plant Res* 2011, **124**:415-424.
45. Iida H, Yagawa Y, Anraku Y: **Essential role for induced Ca²⁺ influx followed by [Ca²⁺]_i rise in maintaining viability of yeast cells late in the mating pheromone response pathway. A study of [Ca²⁺]_i in single *Saccharomyces cerevisiae* cells with imaging of fura-2.** *J Biol Chem* 1990, **265**:13391-13399.
46. Kurusu T, Hamada H, Hanamata S, Kuchitsu K: **Roles of calcineurin B-like protein-interacting protein kinases in plant innate immunity in rice.** *Plant Signal Behav* 2010, **5**:1045-1047.

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