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Pleiotropy or linkage? Their relative contributions to the genetic correlation of quantitative traits and detection by multi-trait GWA studies.

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Abstract

Genetic correlations between traits may cause correlated responses to selection. Previous models described the conditions under which genetic correlations are expected to be maintained. Selection, mutation and migration are all proposed to affect genetic correlations, regardless of whether the underlying genetic architecture consists of pleiotropic or tightly-linked loci affecting the traits. Here, we investigate the conditions under which pleiotropy and linkage have differential effects on the genetic correlations between traits by explicitly modeling multiple genetic architectures to look at the effects of selection strength, degree of correlational selection, mutation rate, mutational variance, recombination rate, and migration rate. We show that at mutation-selection(-migration) balance, mutation rates differentially affect the equilibrium levels of genetic correlation when architectures are composed of pairs of physically linked loci compared to architectures of pleiotropic loci. Even when there is perfect linkage (no recombination within pairs of linked loci), a lower genetic correlation is maintained than with pleiotropy, with a

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lower mutation rate leading to a larger decrease. These results imply that the detection of causal loci in multi-trait association studies will be affected by the type of underlying architectures, whereby pleiotropic variants are more likely to be underlying multiple detected associations. We also confirm that tighter linkage between non-pleiotropic causal loci maintains higher genetic correlations at the traits and leads to a greater proportion of false positives in association analyses.

Keywords: Pleiotropy, Linkage, Genetic Architecture, GWAS, Migration, Mutation

1 **Introduction**

2 Both pleiotropy and linkage disequilibrium create genetic correlations be-
3 tween traits so that traits do not vary independently of one another (Wright,
4 1977; Arnold, 1992; Walsh and Blows, 2009). Under natural selection, this
5 process can prevent a combination of traits from reaching their respective op-
6 timum trait values favored by natural selection (Falconer and Mackay, 1996).
7 Likewise, under artificial selection it can constrain breeders from improving
8 one trait due to undesired changes in another, and in medical gene targeted
9 therapy treatments it can cause adverse side-effects (Wright, 1977; Parkes
10 et al., 2013; Visscher et al., 2017; Wei and Nielsen, 2019). Pleiotropy may
11 cause genetic correlation because one gene’s product (e.g., an enzyme or a
12 transcription factor) has more than one target and therefore affects more
13 than one trait or because one gene’s product belongs to a metabolic pathway
14 that has more than one downstream effect (Hodgkin, 1998; Stearns, 2010;
15 Wagner and Zhang, 2011). Linkage disequilibrium (LD) may be the result of
16 a set of loci in close physical proximity on a chromosome that makes a set of
17 alleles at those loci less likely to be split up by recombination and therefore
18 more likely to get passed on together from one generation to the next. But
19 other mechanisms leading to the transmission of one combination of alleles
20 at separate loci over another combination, can also generate LD and cre-

21 ate genetic correlations between traits that those loci affect (e.g., assortative
22 mating, environmental correlations) (Falconer and Mackay, 1996).

23 One of the main objectives of a genome-wide association study (GWAS)
24 is to identify causal genetic variants underlying one or more traits. GWASes
25 leverage the rapid increase in genomic sequencing to find correlations between
26 traits and genotypes, and their success is dependent on the effect sizes of the
27 loci and the distinction between phenotypes. GWASes have had success
28 in associating genetic variants with traits of interest, which have allowed
29 researchers to find the molecular underpinnings of trait change (Visscher
30 et al., 2017). Moving from one trait to two or more trait associations can
31 lead to discovering pleiotropic loci (Saltz et al., 2017). One GWAS using 1094
32 traits and 14,459 genes, found that 44% of genes were “pleiotropic”, but this
33 was determined by assigning genetic variants to the closest gene and even to
34 both flanking genes when the genetic variant was intergenic (Chesmore et al.,
35 2018). This conflates linkage and pleiotropy, and the chain of causality (Platt
36 et al., 2010). Another study, found 90% of genes and 32.4% of SNPs were
37 associated with more than one trait domain, but they could not rule out
38 SNPs associated with traits due to linkage disequilibrium (Watanabe et al.,
39 2018). Unfortunately, determining whether genetic variant associations and
40 trait correlations are actually the result of pleiotropy or linkage is difficult
41 since they often map to large regions of genomes, or are in intergenic regions
42 and don't associate with the closest genes (Flint and Mackay, 2009; Zhu
43 et al., 2016; Peichel and Marques, 2017; Visscher et al., 2017). Distinguishing
44 between the two types of genetic architectures is important for understanding
45 the underlying molecular functions of the traits, and determining how the
46 traits may be differentially affected by selection (Lynch et al., 1998; Barrett
47 and Hoekstra, 2011; Saltz et al., 2017). This is salient at a time when an
48 increasing number of traits of interest (e.g., human diseases) appear to be
49 affected by loci that affect other traits, and especially when targeted gene
50 therapy clinical trials are more widespread than ever (Edelstein et al., 2007;

51 Cai et al., 2016; Pickrell et al., 2016; Visscher and Yang, 2016; Chesmore
52 et al., 2018; Ginn et al., 2018). There are potentially negative implications for
53 gene therapy because fixing a gene underlying one disease might increase risk
54 for another disease. For example, some genetic variants that are associated
55 with greater risk of Ankylosing spondylitis are also associated with less risk
56 of Rheumatoid arthritis, and so “fixing” one gene would have undesired side-
57 effects in this case (Parkes et al., 2013; Gratten and Visscher, 2016).

58 But the evolutionary dynamics of pleiotropic versus linked loci in creat-
59 ing genetic correlations are expected to be different, since pleiotropy requires
60 only one mutation to affect multiple traits and build-up genetic correlations,
61 and linked pairs require two. Mutation rate should be an important factor
62 distinguishing pleiotropy and linked pairs because single mutations affecting
63 more than one trait provides the opportunity for combinations of effects to
64 match patterns of correlational selection better than linked loci that affect
65 one trait at a time. Thus, linked pairs may require high mutation rates to
66 maintain genetic correlations. Recombination can also reduce genetic corre-
67 lations between traits by breaking up associations between alleles at linked
68 loci, but the same cannot occur with a pleiotropic locus (but see Wagner
69 et al. (2007) for other mechanisms to alleviate pleiotropic constraints). Poly-
70 genic analytical models attempting to approximate the level of genetic vari-
71 ance and covariance at mutation-selection balance in a population suggest
72 that tight linkage between pairs of loci affecting separate traits “is nearly
73 equivalent to” pleiotropic loci affecting both traits (Lande, 1984). Therefore,
74 genetic correlations between traits can be approximated using previously
75 elucidated pleiotropic models under certain conditions (Lande, 1980, 1984;
76 Turelli, 1985). On the other hand, more recent extensions of Fisher’s Ge-
77 ometric Model (Fisher, 1930) predict that pleiotropic mutations, compared
78 to mutations that affect only one trait, are less likely to be beneficial over-
79 all since a beneficial effect on one trait may be detrimental to others (Orr,
80 1998; Otto, 2004). The detrimental effect of pleiotropy is exacerbated when

81 increasing the strength of selection or with very strong correlational selection
82 between traits, since both reduce the amount of phenotypic space where mu-
83 tations are beneficial (unless pleiotropic effects are aligned with the fitness
84 surface created by correlational selection). This detriment is not present for
85 linked loci affecting separate traits since their beneficial mutations will not
86 have the collateral effects of pleiotropy. These, therefore suggest that linkage
87 and pleiotropy may have differential effects on genetic variance and covari-
88 ances depending on mutation, recombination and selection regimes, but this
89 comparison was not fully explored in any previous model.

90 Lande (1984) predicted that when loci affecting *different* traits are tightly
91 linked, and there is strong correlational selection between traits, recombina-
92 tion rates between loci affecting different traits can strongly affect genetic
93 correlations between traits, when selection is weak and mutation rates are
94 relatively high. In an extreme case where there is complete linkage between
95 pairs of loci affecting different traits (the recombination rate is 0), and no
96 linkage between sets of these pairs of linked loci (the recombination rate is
97 0.5), then he determined that the maximum genetic correlation due to link-
98 age may be almost as large as the extent of correlational selection, which can
99 be calculated from the (per linkage group) genetic covariance between traits
100 and the genetic variances, respectively, as:

$$genetic\ covariance\ (b) = \frac{\rho\omega^2\mu\alpha^2}{2c}, \quad (1)$$

101

$$genetic\ variance\ (c) = \sqrt{(1 + \sqrt{1 - \rho^2})\omega^2\frac{\mu\alpha^2}{2}}, \quad (2)$$

102 where ρ is the extent of correlational selection acting between the traits, ω^2 is
103 the strength of selection (with lower values representing stronger selection), μ
104 is the per-locus mutation rate, and α^2 is the per-locus mutation variance. If
105 there is equal variances among traits then the genetic correlation is calculated

106 as:

$$107 \quad \text{genetic correlation} = \frac{b}{c} = \frac{\rho}{1 + \sqrt{1 - \rho^2}}. \quad (3)$$

108 From these equations we see that, even in the absence of pleiotropy, genetic
109 covariance may arise from linkage disequilibrium, and depends on both the
110 strength of correlational selection between traits and selection on each trait,
111 as well as on the mutational inputs (mutation rates and mutational variances)
112 of the genes affecting those traits. Yet, from equation (3), the resulting
113 genetic correlation among traits is independent of the genetic architecture of
114 the traits. Lande goes on further to state that the case of complete linkage
115 between pairs of loci affecting different traits is “equivalent to a lesser number
116 of loci with pleiotropic effects”, but this is not quantified nor is the scaling
117 of the two examined. We seek to quantify the equivalence of pleiotropy and
118 linkage in their ability to maintain equilibrium levels of genetic (co)variation
119 under the same conditions. We also wish to extend this to look at a range
120 of linkage distances, selection variances and correlations, and mutation rates
121 and variances, to look at the relative effects of each.

122 The expectations given by Lande are only expected to be accurate under
123 conditions where mutation rates are high compared to the strength of selec-
124 tion on the traits of interest (Turelli, 1984; Turelli and Barton, 1990). When
125 mutation rates are lower ($< 10^{-4}$), predictions for equilibrium levels of genetic
126 variation break down and are better approximated by the “house-of-cards”
127 model (Kingman, 1978; Turelli, 1984). Analytic predictions for equilibrium
128 levels of genetic covariation between traits due to linkage disequilibrium, on
129 the other hand, have not been well explored for the “house-of-cards” model
(Bürger, 2000).

130 Additionally, levels of trait genetic covariation can be influenced by other
131 evolutionary processes that affect allele frequencies, and the covariation of al-
132 lelic values in a population (e.g., migration (Guillaume and Whitlock, 2007),
133 drift (Griswold et al., 2007), inbreeding (Lande, 1984), and phenotypic plas-
134 ticity (Draghi and Whitlock, 2012)). Migration affects genetic covariation

135 because when it is sufficiently high (relative to selection in the focal popula-
136 tion), then combinations of alleles coming from a source population will also
137 be maintained in the focal population. This can lead to higher genetic co-
138 variation between traits in the focal populations, whether the combinations
139 of alleles immigrating are (more likely to be) correlated in their effects on
140 those traits or not (Guillaume and Whitlock, 2007). Migration may also have
141 different effects depending on whether the genetic architecture is pleiotropic
142 or made up of linked loci, but this has not been explored.

143 Here, we are interested in the conditions in which pleiotropic architectures
144 behave similarly or differently to architectures with tight physical linkage
145 between loci affecting different traits, with respect to their effects on genetic
146 correlations between the traits. We use computer simulations to investigate
147 whether the effect of evolutionary forces on the genetic correlation between
148 traits is dependent on the type of genetic architecture, and how. We focus on
149 the relative contributions of selection, mutation and migration to the build
150 up of genetic correlation between traits having different genetic architectures.
151 We show that unless mutation rates are high, genetic architectures with tight
152 linkage between loci maintain much lower equilibrium genetic correlations
153 than pleiotropic architectures. Even when mutation rates are high, other
154 evolutionary forces affecting equilibrium levels of genetic correlation still show
155 a difference between architectures but to a much lesser extent. Additionally,
156 we simulate genomic single-nucleotide polymorphism (SNP) data sets using
157 the different architectures, and show that map distances between causative
158 and non-causative QTL affect false positive proportions in GWA analyses.

159 **Materials and Methods**

160 We modeled four different genetic architectures in a modified version of
161 the individual-based, forward-in-time, population genetics simulation soft-
162 ware NEMO (Guillaume and Rougemont, 2006; Chebib and Guillaume, 2017).
163 NEMO was modified to allow single non-pleiotropic loci to affect different

164 quantitative traits. To compare how pleiotropy and linkage differentially af-
165 fect the genetic correlation between traits, we modeled a set of 120 pairs of
166 linked, non-pleiotropic loci, and a set of 120 pleiotropic loci affecting the two
167 traits. We varied the recombination distance between the two non-pleiotropic
168 loci of each pair with distances 0cM, 0.1cM, or 1cM (Figure 1). Pairs were
169 unlinked to other pairs. The pleiotropic loci were also unlinked to each
170 other. The recombination rates chosen represent no recombination between
171 linked loci, as well as an average and an extreme value of recombination at
172 “hotspots” in the human genome, respectively (Myers et al., 2006). All loci
173 had additive effects on the traits.

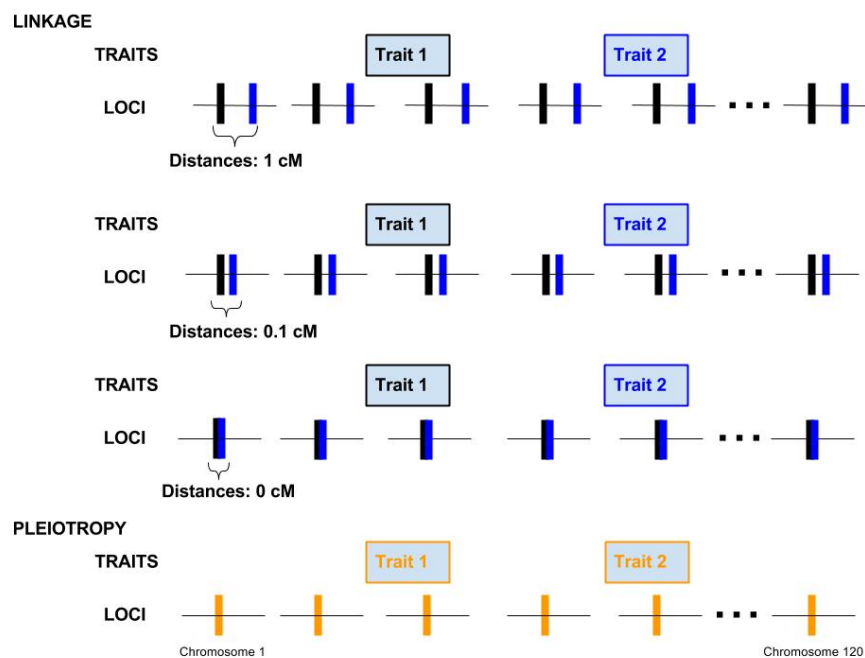


Figure 1: Four genetic architectures showing the distribution of loci on 120 chromosomes. In the case of linkage architectures, pairs of loci affecting the two different traits on each chromosome are either 1, 0.1 or 0 cM apart. In the case of the pleiotropic architecture, each locus on each chromosome affects both traits.

174 Unless otherwise specified, each simulation was run with 5,000 initially
175 monomorphic (variation is gradually introduced through mutations), diploid

176 individuals for 10,000 generations achieving mutation-selection(-migration)
177 balance in order to observe general patterns of genetic correlation in the
178 near-absence of drift. Individuals were hermaphrodites mating at random
179 within a population, with non-overlapping generations. Phenotypes were
180 calculated for each of the two traits modeled by summing the allelic values
181 of all loci affecting one trait. Gaussian stabilizing selection was applied and
182 determined the survival probability of juveniles, whose fitness was calculated
183 as $w = \exp \left[-\frac{1}{2} ((\mathbf{z} - \theta)^T \cdot \mathbf{\Omega}^{-1} \cdot (\mathbf{z} - \theta)) \right]$, where \mathbf{z} is the individual phe-
184 notype vector (initialized to the optimum values), θ is the vector of local
185 optimal trait values (set to 10 for both traits in the focal population), and
186 $\mathbf{\Omega}$ is the selection variance-covariance matrix ($n \times n$, for n traits) describing
187 the multivariate Gaussian selection surface. To examine the effects of the
188 strength of stabilizing selection on each trait and strength of correlational
189 selection between traits, different sets of simulations were run with the di-
190 agonal elements of the $\mathbf{\Omega}$ matrix set as $\omega^2 = 50$, or 100 (selection strength),
191 and off-diagonal set to $\omega^2 \times \rho_\omega$ (where the correlational selection, $\rho_\omega = 0.5$
192 or 0.9). The strength of selection scales inversely with ω^2 where a value of
193 100 corresponds to weak (but non-trivial) selection as opposed to correla-
194 tional selection, ρ_ω , where a value of 0.9 corresponds to strong correlational
195 selection between traits (Lande, 1984; Turelli, 1984).

196 To examine the effects of mutational input on genetic correlation between
197 traits, different sets of simulations were run with mutation rates (μ) of 0.001,
198 0.0001, or 0.00001, and moderate mutational effect sizes (α^2) of 0.1, 0.01,
199 or 0.001 (Turelli, 1984). Mutational effects at each non-pleiotropic locus
200 were drawn from a univariate normal distribution (with a mean of zero) or
201 a bivariate normal distribution (with means of zero and a covariance of 0)
202 for pleiotropic loci. Mutational effects were then added to the existing allelic
203 values (continuum-of-alleles model; Crow and Kimura, 1964). All loci were
204 assumed to have equal mutational variance. No environmental effects on the
205 traits were included.

206 To examine the effects of migration from a source population on genetic
207 correlation between traits, additional sets of simulations were run with uni-
208 directional migration from a second population (as in an island-mainland
209 model with each population consisting of 5000 individuals) with backward
210 migration rates (m) of 0.1, 0.01, and 0.001. The backward migration rate
211 represents the average proportion of new individuals in the focal population
212 whose parent is from the source population. The local optimum values for
213 the two traits in the source population were set at $\theta = [\sqrt{50}, \sqrt{50}]$ (10
214 units distance from the focal population's local optimum). Both focal and
215 source populations had weak stabilizing selection with a strength of $\omega^2 = 100$,
216 the focal population had no correlational selection between the two traits
217 and the source population had a correlational selection of $\rho_\omega = 0$ or 0.9.
218 Fifty replicate simulations were run for each set of parameter values and
219 statistics were averaged over replicates. Averages were also compared against
220 analytical expectations laid out by Lande (1984) and reproduced here in
221 Equations 1–3.

222 *Effects of genetic architecture on false positive/negative proportions in asso-*
223 *ciation studies*

224 In order to elucidate the differential effects of pleiotropy and linkage on the
225 detection of true causal genetic variants in association studies, a genome-wide
226 association (GWA) analysis was performed on data simulated as described
227 above (with only a single population), except that diallelic loci were used in-
228 stead of a continuum-of-alleles model to better represent SNPs. Correlational
229 selection values were chosen that provided equal on-average genetic correla-
230 tions between traits for all genetic architectures of 0.2, 0.3, and 0.4, values
231 frequently observed in both morphological and life-history traits (Roff, 1996).
232 In the association study, a per-locus regression of trait values was performed
233 over genotypes, and the (negative log 10) p-values of regression slopes were
234 plotted with a Benjamini-Hochberg False Discovery Rate (FDR) cutoff to
235 adjust significance levels for multiple tests (Benjamini and Hochberg, 1995).

236 From this, we observed the number (and proportion) of false positives (linked
237 loci that had no effect on a trait but whose regression slope p-values were
238 above the FDR cutoff for that same trait) and false negatives (pleiotropic loci
239 that had an effect on both traits but whose regression slope p-values were
240 below the FDR cutoff for either trait). No correction for population strat-
241 ification was performed during this analysis because each simulation had a
242 single, large, randomly breeding population. Linkage disequilibrium values
243 of D' and R^2 between pairs of linked traits were also calculated using the **R**
244 package genetics (v1.3.8.1) (Warnes et al., 2013). Statistics for number and
245 proportion of false positives and negatives were obtained from the average
246 over 20 replicate simulations of each genetic architecture. We also assessed
247 the false positive rate on an additional set of neutral QTL linked to the causal
248 loci. We simulated a set of 120 independent linkage groups with 200 neutral
249 di-allelic QTL per group, evenly distributed on both sides of the central po-
250 sition occupied by the two causal QTL. Each linkage group was 1 cM long.
251 The minimum recombination rate between two adjacent loci was 10^{-5} . The
252 neutral QTL were set in 10 successive windows of 0.05cM (~ 50 kb) on each
253 side of the causal QTL. The two causal QTL were perfectly linked (0cM)
254 and non-pleiotropic. The simulations were run for 50,000 generations and 10
255 replicates.

256 **Results**

257 *Effects of genetic architecture on genetic correlation at mutation-selection* 258 *balance*

259 By generation 10,000, when mutation-selection balance is reached, simu-
260 lations with the pleiotropic architecture generally maintain a higher average
261 genetic correlation than those with linkage architectures, even when recom-
262 bination is absent (linkage distance of 0cM between pairs of loci) (Figure
263 2). Variation in the mutation rate has the largest effect on the difference of
264 genetic correlation between pleiotropic and fully linked non-pleiotropic loci,

265 with much lower correlations as the mutation rate decreases from 10^{-3} to
266 10^{-5} (Figure 3). This reduction in genetic correlation mostly affected the
267 non-pleiotropic pairs of loci for which a large drop in genetic correlation oc-
268 curred between $\mu = 10^{-3}$ and $\mu = 10^{-4}$ (Figure 3). With lower mutation
269 rates there is also a lower total genetic variance and lower genetic covari-
270 ance. The higher genetic correlation obtained with pleiotropic loci was due
271 to a lower total genetic variance when the mutation rate was high ($\mu = 10^{-3}$),
272 but to a higher genetic covariance when mutation rate was low ($\mu = 10^{-4}$ or
273 10^{-5}).

274 The genetic correlation between the traits decreases with reduction in all
275 four factors tested (μ , ρ_ω , ω^2 , and α^2) and for all genetic architectures, with
276 the coefficient of correlational selection (ρ_ω) having the strongest effect (Fig-
277 ure 4), as expected from equation (3). However, changes in the strength of
278 selection (ω^2) and the mutational variance (α^2) also affect the genetic corre-
279 lation at equilibrium. We find that reducing the strength of selection (Figure
280 5) had a relatively smaller effect than reducing the mutational variance (Fig-
281 ure 6). A decrease in mutational variance leads to a decrease in genetic
282 correlation by a similar amount regardless of genetic architecture (though
283 loose linkage is affected the most). Populations with linkage architectures
284 need both high mutation rates and high mutational variance to maintain
285 strong genetic correlation, whereas the pleiotropic architecture just needs
286 high mutational variance.

287 In contrast to the correlation, the genetic covariance of the two traits
288 was generally equal between pleiotropic and fully linked non-pleiotropic loci,
289 and decreased as recombination increased within pairs of non-pleiotropic loci.
290 The cause of the observed higher trait correlation obtained with pleiotropic
291 loci was the lower genetic variance they maintain under stabilizing selection.

292 *Effects of migration on genetic correlation*

293 A higher migration rate from a source population, whose traits are un-
294 der correlational selection, leads to higher genetic correlations in the focal

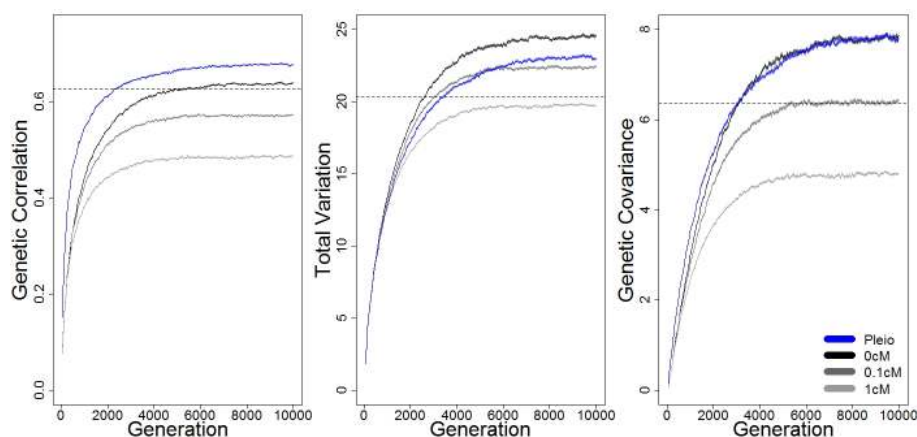


Figure 2: Average genetic correlation, total genetic variation and genetic covariation (and their standard deviations) over 10,000 generations reaching mutation-selection equilibrium for four different genetic architectures: pairs of linked loci affecting two different traits with 0, 0.1 or 1cM between loci, or pleiotropic loci affecting both traits. $N = 5000$, $\omega^2 = 100$, $\rho_\omega = 0.9$, $\alpha^2 = 0.1$, and $\mu = 0.001$. Dashed line represents Lande's 1984 expectations for completely linked loci (0 cm).

295 population regardless of the genetic architecture (Figure 7A). The effect of
296 migration increases with tighter linkage and is highest with pleiotropic archi-
297 tecture. This effect on genetic correlation is still observed when there is no
298 correlational selection on the traits in the source population, but to a largely
299 reduced degree (Figure 7B).

300 *Effects of linkage and pleiotropy on proportion of false positives/negatives*
301 *and linkage disequilibrium in multi-trait GWASes*

302 In simulations where there is linkage between SNPs and equivalent lev-
303 els of genetic correlation between traits, the number and proportion of loci
304 that are false positives (above FDR cutoff but no effect on trait) increase as
305 linkage distance decreases between SNPs affecting different traits (shown in
306 Figure 8 and Supplementary Figure S1). When genetic correlation is higher
307 (due to stronger correlational selection), linkage distance has a greater im-
308 pact on the proportion of false positives. Also, genetic correlation has a larger
309 effect than linkage distance on the number of false positives. In simulations

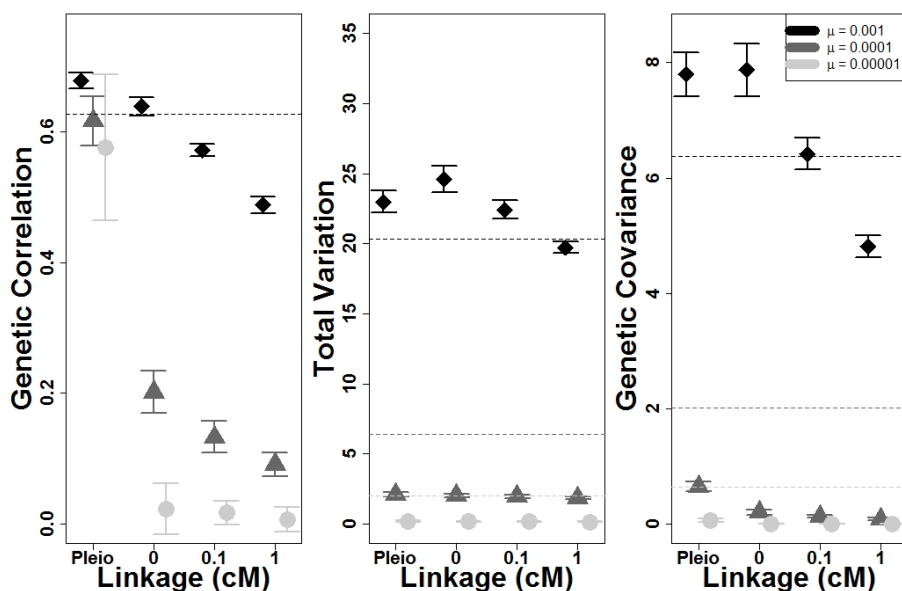


Figure 3: Effect of mutation rate (μ) on average genetic correlation, total variance and genetic covariance (and their standard deviations) after 10,000 generations of correlated, stabilizing selection for four different genetic architectures. $N = 5000$, $\omega^2 = 100$, $\rho_\omega = 0.9$, and $\alpha^2 = 0.1$. Dashed lines represents Lande's 1984 expectations for completely linked loci (0 cM).

310 where SNPs are pleiotropic, genetic correlation due to correlational selection
 311 has little impact on the number and proportion of false negatives (below
 312 FDR cutoff but does affect the traits). Linkage disequilibrium between pairs
 313 of linked SNPs decreases as distance between SNPs increases regardless of
 314 genetic correlation (Figure 9 and Supplemental Table S1). Long-distance
 315 linkage disequilibrium between unlinked SNPs increases with the strength of
 316 correlational selection when the map distance within pairs of linked SNPs
 317 increases (when measured with D' , Supplemental Figure S2). In simulations
 318 where SNPs are pleiotropic, long-distance linkage disequilibrium does not
 319 seem to be affected by a change in genetic correlation. Finally, in simulations
 320 with neutral QTL, the false pleiotropic positive rate is $4.2e-5$ (genetic
 321 correlation: $g_{cor} = 0.2$), $8.3e-5$ ($g_{cor} = 0.3$), and $1.5e-4$ ($g_{cor} = 0.4$) on average
 322 in the first 50kb window (within 0.05 cM of the causal QTL). No false

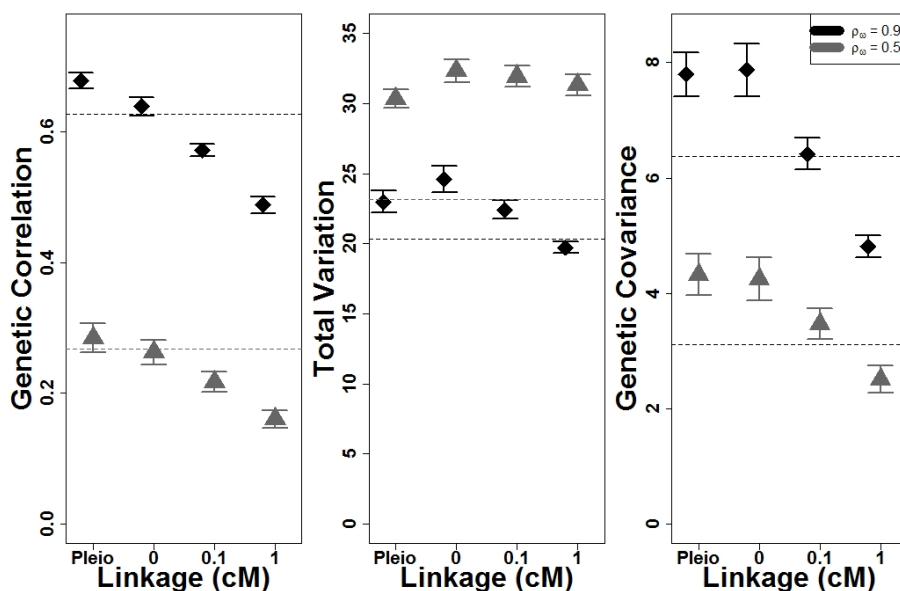


Figure 4: Effect of correlational selection (ρ_ω) on average genetic correlation, total variance and genetic covariance (and their standard deviations) after 10,000 generations of correlated, stabilizing selection for four different genetic architectures. $N = 5000$, $\omega^2 = 100$, $\alpha^2 = 0.1$, and $\mu = 0.001$. Dashed lines represents Lande's 1984 expectations for completely linked loci (0 cM).

323 pleiotropic positives were found at map distance above 0.15 cM for $g_{cor} = 0.2$
 324 and 0.3, and above 0.25 cM for $g_{cor} = 0.4$.

325 Discussion

326 *Pleiotropy and linkage are not the same*

327 The main expectation under an assumption of weak selection and strong
 328 correlational selection is that populations with a genetic architecture con-
 329 sisting of unlinked pairs of two completely linked loci (0cM distance) should
 330 maintain similar equilibrium levels of genetic correlation as with a genetic
 331 architecture consisting of a lesser number of unlinked pleiotropic loci (Lande,
 332 1984). Our results show that this is the case when there are half as many
 333 pleiotropic loci and mutation rates are relatively high. A high rate of muta-
 334 tion (10^{-3}) allows for multiple mutations in both loci in a tightly linked pair

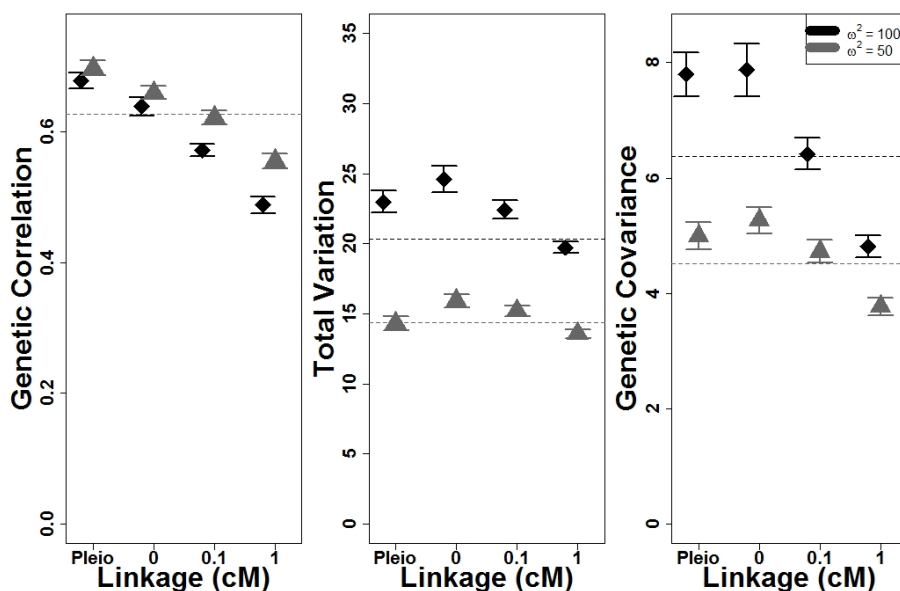


Figure 5: Effect of selection variance (ω^2) on average genetic correlation, total variance and genetic covariance (and their standard deviations) after 10,000 generations of correlated, stabilizing selection for four different genetic architectures. $N = 5000$, $\rho_\omega = 0.9$, $\alpha^2 = 0.1$, and $\mu = 0.001$. Dashed lines represents Lande's 1984 expectations for completely linked loci (0 cM).

335 to accumulate and maintain levels of genetic covariance near to that of muta-
 336 tions in a single pleiotropic locus, but empirical estimations of mutation rates
 337 from varied species like bacteria and humans suggests that *per-nucleotide* mu-
 338 tation rates are in the order of 10^{-8} to 10^{-9} (Nachman and Crowell, 2000;
 339 Ford et al., 2011; Keightley et al., 2015; Lindsay et al., 2019). If a polygenic
 340 locus consists of hundreds or thousands of nucleotides, as in the case of many
 341 quantitative trait loci (QTLs), then per-locus mutation rates may be as high
 342 as 10^{-5} , but the larger the locus the higher the chance of recombination be-
 343 tween within-locus variants that are contributing to genetic correlation. This
 344 leads us to believe that with empirically estimated levels of mutation and re-
 345 combination, strong genetic correlation between traits are more likely to be
 346 maintained if there is an underlying pleiotropic architecture affecting them
 347 than will be maintained due to tight linkage. Consequently, GWASes that

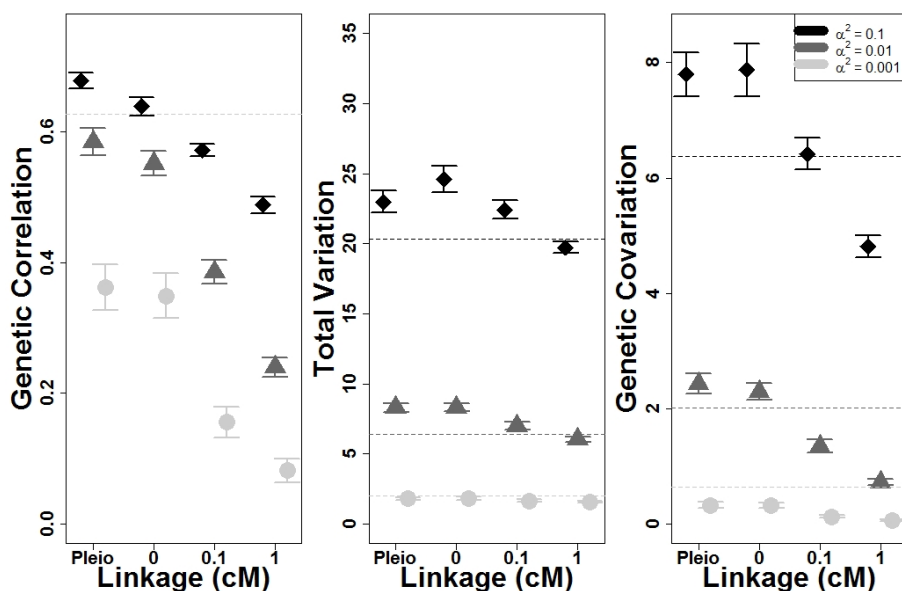


Figure 6: Effect of mutation variance (α^2) on average genetic correlation, total variance and genetic covariance (and their standard deviations) after 10,000 generations of correlated, stabilizing selection for four different genetic architectures. $N = 5000$, $\omega^2 = 100$, $\rho_\omega = 0.9$, and $\mu = 0.001$. Dashed lines represents Lande's 1984 expectations for completely linked loci (0 cM).

348 detect associations between multiple traits and single genetic variants are
 349 more likely to be detecting pleiotropic loci than linked loci. Also, previous
 350 theoretical models suggest that Lande's (1984) equilibrium levels of genetic
 351 variation are not well approximated at low per-locus mutation rates (com-
 352 pared to the strength of selection), which was also true in our simulations
 353 (Supplemental Figure S3) (Turelli, 1984; Bürger, 2000).

354 We find that even under scenarios where pleiotropy and tight linkage
 355 maintain similar levels of genetic covariance, pleiotropic architectures have
 356 higher genetic correlations because they have lower total genetic variance.
 357 This can be explained by understanding the differential fitness effects of loci.
 358 Mutations that affect more than one trait are less likely to be beneficial (Orr,
 359 1998; Otto, 2004). The distribution of fitness effects of pleiotropic mutations
 360 is shifted towards more negative average values as the number of traits af-

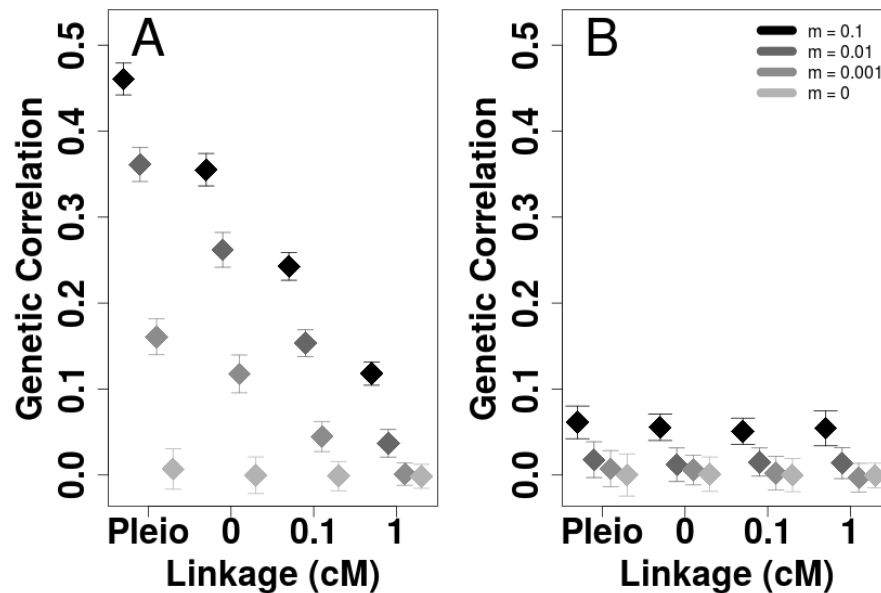


Figure 7: Average genetic correlations in the focal populations (and their standard deviations) after 10,000 generations of migration from a source population with different migration rates (m) for four different genetic architectures. A– Migration from a source population with correlational selection between traits ($\rho_\omega = 0.9$). B– Migration from a source population without correlational selection between traits ($\rho_\omega = 0$).

361 fected increases (Martin and Lenormand, 2006; Chevin et al., 2010). Hence,
 362 pleiotropic architectures that affect more traits have less positive mutational
 363 effects on fitness and maintain a lower equilibrium genetic variation when
 364 compared to linked architectures (Turelli, 1985). It has been suggested that
 365 this might be overcome in more complex organisms with a greater number
 366 of traits by modularization of the effects of different pleiotropic genes to
 367 separate sets of traits and decrease the pleiotropic degree of the mutations
 368 but theoretical models have shown mixed results (Baatz and Wagner, 1997;
 369 Hansen, 2003; Welch et al., 2003; Martin and Lenormand, 2006; Chevin et al.,
 370 2010; Wagner and Zhang, 2011).

371 When correlational selection on the traits is strong in the simulations with
 372 linked architectures, the equilibrium genetic correlation is dependent on the
 373 recombination rates between loci within linkage groups. Tightly linked loci

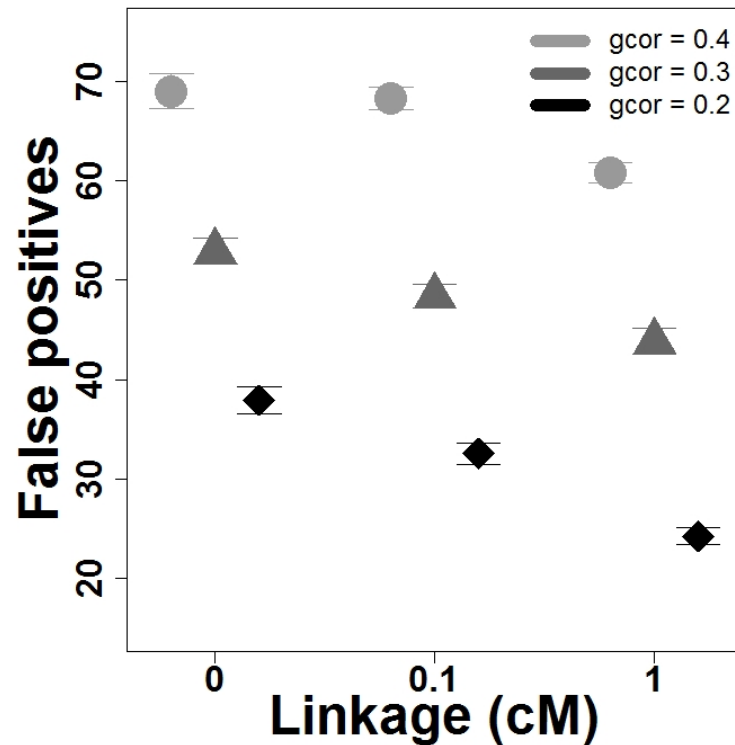


Figure 8: Average number of false positives from GWA analyses (and their standard deviations) for different linkage distances between paired loci and different genetic correlations (gcor). A locus was considered a false positive if associations between the locus' genotypes and trait values, that the locus does not directly affect, are above the Benjamini-Hochberg FDR cutoffs (with a significance level of 0.05).

374 can maintain higher levels of genetic correlation from a build-up of positive
375 linkage disequilibrium than loosely linked loci. This matches the analytical
376 predictions put forth in Lande (1984) under the assumption of weak stabi-
377 lizing selection, strong correlational selection, and loose linkage between loci
378 affecting the same trait.

379 *The impact of pleiotropy and linkage maintaining different genetic correla-*
380 *tions in association studies*

381 When methods like GWA analyses are employed to detect shared ge-
382 netic influences (pleiotropy or linkage) on multiple traits of interest, they

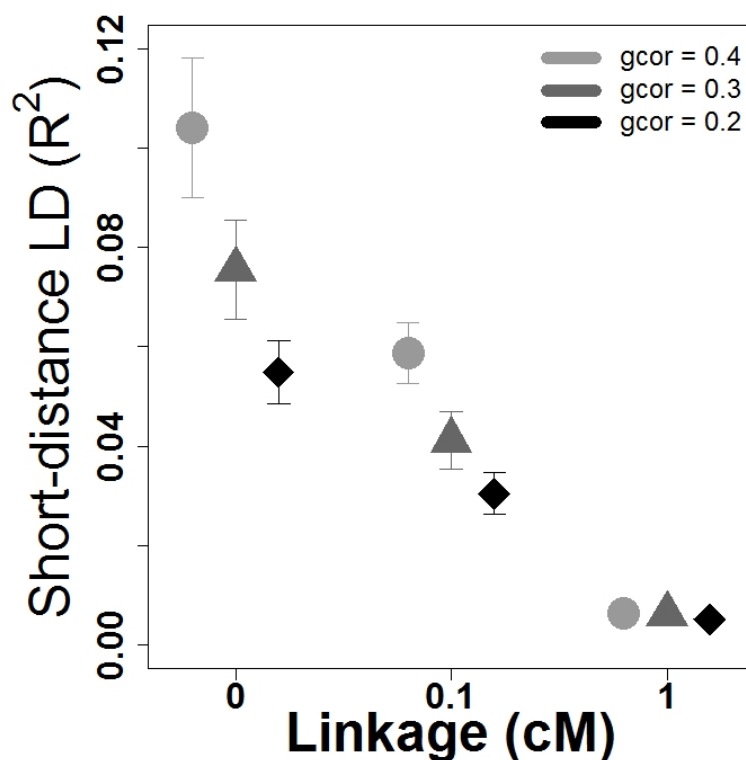


Figure 9: Average linkage disequilibrium (LD) between pairs of linked loci (and their standard deviations) for different linkage distances between paired loci and different genetic correlations (gcor).

383 are dependent upon detecting combinations of effect sizes of genetic variants
384 associated with those traits (Hill and Zhang, 2012b,a; Chung et al., 2014;
385 Visscher and Yang, 2016). The success or failure of this endeavor is directly
386 connected to the ability to detect loci with associations to each trait and
387 the strength of genetic correlation between traits (Wei et al., 2014; Pick-
388 rell et al., 2016; Chesmore et al., 2018; Verbanck et al., 2018). Our results
389 show that (tight) linkage between loci affecting different but correlated traits
390 will lead to “many” false positives. We also show that false positive rates
391 are only marginally affected by linked but non-causative loci. As expected,
392 false positive rates decrease with the map distance to the causative loci and
393 the correlation between the traits (Siegmond and Yakir, 2007). Therefore,

394 GWASes will not be able to empirically distinguish between pleiotropy and
395 linkage when loci affect genetically correlated traits. The proportion of genes
396 associated with two or more phenotypes in the GWAS catalog has increased
397 to around 40% in the last decade (Welter et al., 2013; Pickrell et al., 2016).
398 But it is difficult to determine if this is truly representative of the prevalence
399 of pleiotropy because QTLs are often mapped to loci that can encompass
400 thousands of nucleotides (and more than one gene) and informative SNPs
401 with significant effect sizes are assigned to the closest genes with annotated
402 phenotypes (Chesmore et al., 2018; Liu et al., 2019; Cai et al., 2020). Con-
403 flating inter-genic SNPs with nearby pleiotropic genes (or loci) can distort
404 the prevalence of pleiotropy and reduce the ability to distinguish pleiotropy
405 from physical linkage (Dudley et al., 2005; Gianola et al., 2015). Finding
406 the true false positive rate in GWA studies due to linkage is difficult because
407 it is almost never known whether the source of genetic correlations between
408 traits is linked loci or not, even when fine-scale sequences are available (for the
409 reasons mentioned above and because of the way pleiotropy is erroneously de-
410 fined in GWA studies) (Platt et al., 2010). Watanabe et al. (2018) attempted
411 to break down this issue in a meta-analysis of 558 GWASes by looking at
412 the proportion of genomic loci, genes, and SNPs associated with multiple
413 traits, which may provide a clearer picture of the prevalence of pleiotropic
414 causal variants. They found that 93.3% of loci, 81.0% of genes, and 60.2% of
415 SNPs, were associated with more than one trait. This may seem to provide
416 a better estimate of pleiotropic levels, except that in this study SNPs that
417 were associated with more than one trait could still have been the result of
418 linkage disequilibrium. A point that was brought up by the authors.

419 On the other hand, we observed very few false negatives in pleiotropic loci
420 (regardless of genetic correlation) because we “sampled” the entire popula-
421 tion and therefore had the power to find significant associations with (almost)
422 all causal loci. Had we taken smaller samples of our population to perform
423 the GWA analysis, we would have found a greater number of false nega-

424 tives. The salient consequence is that study design, threshold levels, and
425 genetic correlations between traits will all affect detection of genetic vari-
426 ants, whether the variants are causal themselves or linked to causal variants
427 (Wagner and Zhang, 2011; Hill and Zhang, 2012a). Also, the number of
428 pleiotropic effects a locus has may be under-represented by significance lev-
429 els in association studies (Hill and Zhang, 2012b). Wagner and Zhang 2011
430 go a step further to suggest that number or proportions of traits affected
431 may not be as meaningful as describing the distributions of pleiotropic effect
432 sizes on traits.

433 *There is a difference between pleiotropy and linkage at the nucleotide level*

434 Transgenic experiments may differentiate pleiotropy from linkage at the
435 gene level (Mills et al., 2014), but at the nucleotide level does the distinction
436 between two linked loci and one pleiotropic locus go away? There is evidence
437 that even in the same gene, adjacent polymorphisms affecting different traits
438 in *Drosophila* can be in linkage equilibrium due to fine-scale recombination
439 (Carbone et al., 2006; Flint and Mackay, 2009). But imagine a case where
440 a mutation in a single base-pair has an effect on one trait and a mutation
441 in the base-pair right next to the first base-pair has an effect on a second
442 trait. Now imagine a second case where a mutation in a single base-pair
443 has an effect on two traits. There still seems to be a distinction between
444 these two cases because the probability of a change in both traits in the first
445 case is the mutation rate squared compared to the second case where the
446 probability of a change in both traits is just the mutation rate. Depend-
447 ing on the per-locus mutation rate this difference can be quite large (e.g.
448 10^{-8} versus 10^{-16}). Even in this extreme case, there may indeed still be a
449 gray area in the distinction between pleiotropy and linkage at a mutational
450 level. Mutations may affect the pleiotropic degree (e.g. like enzyme speci-
451 ficity) of a protein-coding gene and the degree to which the gene maintains
452 multi-functionality may itself evolve (Guillaume and Otto, 2012). If there
453 is correlational selection between the catalytic functions of an enzyme, then

454 some pleiotropic mutations that affect more than one catalytic ability will
455 be favoured, and genetic correlations will increase. With this in mind, it
456 makes more sense from a theoretical and functional standpoint to refer to
457 pleiotropy at the nucleotide level (or at the unit of a mutation), than at the
458 gene or larger locus level (but this may depend on the questions of interest
459 (Rockman, 2012; Rausher and Delph, 2015)).

460 *Other factors*

461 Even in the absence of correlational selection it is possible to maintain
462 genetic correlation through continued migration from a source population.
463 High migration brings individuals whose combination of alleles will expand
464 focal population variation in the direction of the source population. This
465 corroborates previous results that showed that slow introgression of allelic
466 combinations into a population can affect the genetic variance-covariance
467 structure of that population (Guillaume and Whitlock, 2007). Whether ge-
468 netic covariance will be maintained in real populations depends on the nature
469 of correlational selection on traits in the population of interest, since migra-
470 tion can reduce local fitness (i.e. migration load) if allele combinations are not
471 favoured by selection or increase it if they are (Nosil et al., 2006; Bolnick and
472 Otto, 2013). Migration into a population will also affect false positive rates
473 since immigrating allele combinations will be in LD from the source popula-
474 tion and will therefore increase the proportion of certain genotypes, even if
475 there is no strong trait correlation in the source population. Although not
476 investigated in this study, a structured population and/or a continual system
477 of inbreeding in a population where there is correlational selection between
478 polygenic traits can result in increased genetic covariation caused by larger
479 LD Lande (1984), which can in turn increase false positive proportions.

480 **Conclusion**

481 Pleiotropic loci maintain stronger genetic correlations between traits than
482 linked loci affecting different traits even when no recombination occurs be-

483 tween the loci, and especially in the magnitude of empirically estimated mu-
484 tation rates. Previous models of the maintenance of genetic covariation at
485 mutation-selection equilibrium describe genetic covariation as a function of
486 the product of mutation rate and variance. These models provide similar
487 expectations for pleiotropic and tight linkage architectures. The discrepancy
488 occurs because of the contingency of mutational covariance input on the oc-
489 currence of mutations (and hence mutation rate). Without high mutation
490 rates, the ability to create genetic covariance between linked loci is highly
491 diminished because the combined likelihood of mutations in each linked loci
492 with both mutational effects in the same direction is low. This result will
493 have implications in the type of underlying architecture we expect to find
494 in multi-trait association studies. On the one hand, tighter linkage between
495 causal loci and detected loci maintains higher genetic correlations, leading to
496 a greater proportion of false positives in pleiotropy tests. More importantly,
497 on the other hand variants are more likely to have pleiotropic effects on
498 traits than linked effects, when they are found to be associated with strongly
499 correlated traits.

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508 **Author Contributions**

509 J.C. performed software modification for model implementation and ac-
510 quisition of data, as well as drafting of manuscript. J.C. and F.G. performed

511 study conception and design, analysis and interpretation of data, and critical
512 revision of manuscript.

513 **Data Archival**

514 The data for this study will be made available online through Zenodo on-
515 line repository at <https://zenodo.org/record/3370185#collapseTwo> and code
516 for simulations can be found [https://sourceforge.net/projects/nemo2/files/Publications-](https://sourceforge.net/projects/nemo2/files/Publications-Code/ChebibGuillaume-PleiotropyOrLinkage-2019/)
517 [Code/ChebibGuillaume-PleiotropyOrLinkage-2019/](https://sourceforge.net/projects/nemo2/files/Publications-Code/ChebibGuillaume-PleiotropyOrLinkage-2019/)

518 **Conflict of interest disclosure**

519 The authors of this article declare that they have no financial conflict of
520 interest with the content of this article. F.G. is a PCI Evolutionary Biology
521 recommender.

522 Supplemental

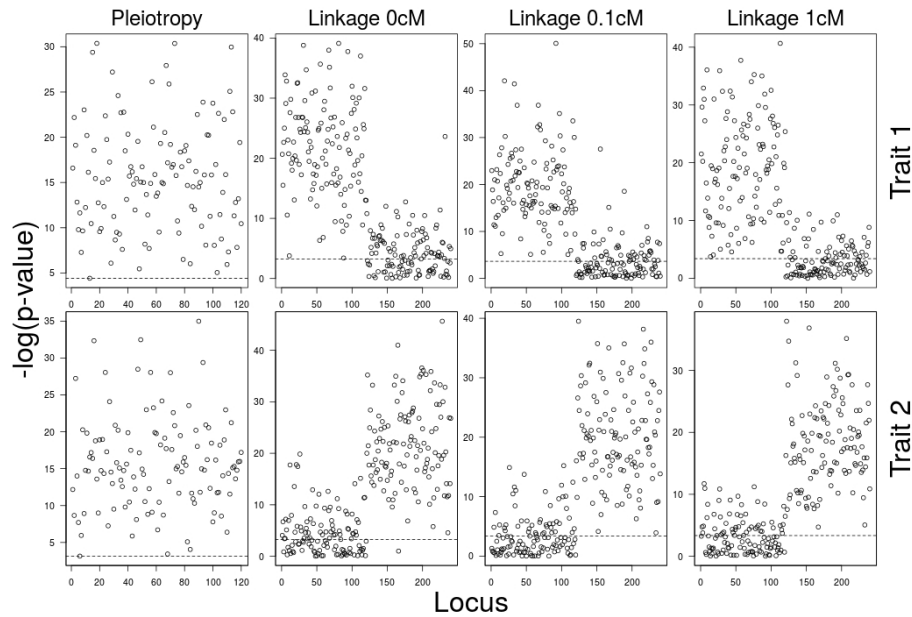


Figure S1: GWA analysis: $-\log(p\text{-values of slope of regression of trait values on genotypes})$ from one set of example simulations. In the case of linkage architectures, the first 120 loci only affected trait 1 and the next 120 loci only affected trait 2. The order of the loci are sorted for visualization purposes whereby linked pairs are separated by the trait they affect (e.g. loci 1 and 121 in the figure are a linked pair). In the case of the pleiotropic architecture, all 120 loci affected both traits. The average genetic correlation of ≈ 0.3 was observed by adjusting the correlational selection levels to 0.88, 0.89, 0.93, and 0.965 for pleiotropy, linkage 0cM, linkage 0.1cM, and linkage 1cM, respectively. Dashed lines represent the Benjamini-Hochberg FDR cutoffs for a significance level of 0.05.

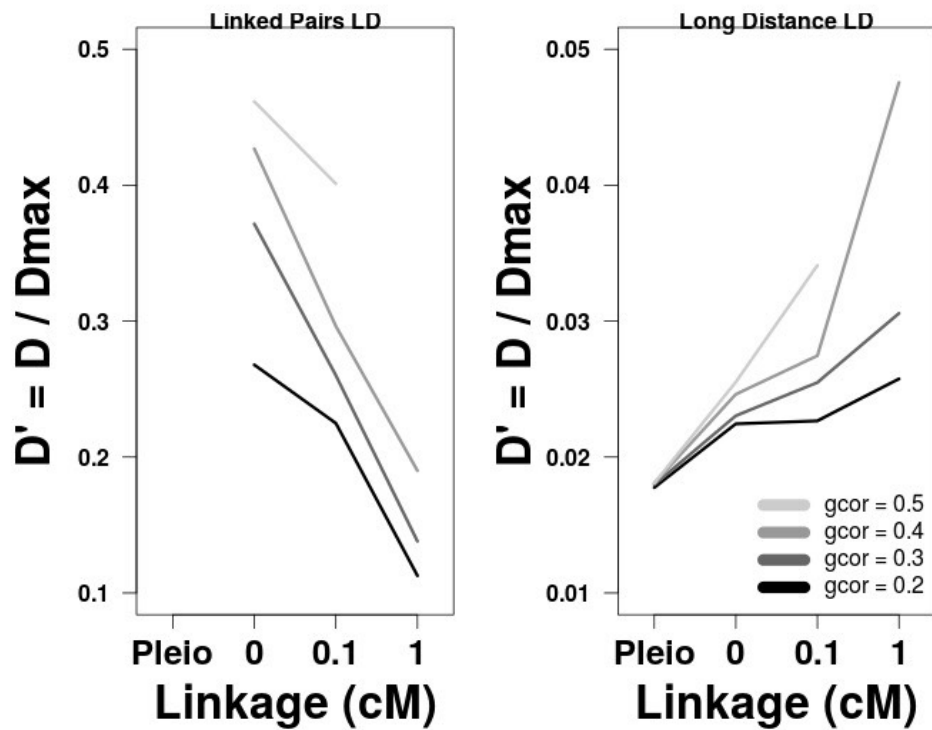


Figure S2: Average linkage disequilibrium (measured by D') between linked pairs (left panel) and between unlinked pairs (right panels) for different genetic correlations and genetic architectures. N.B. No linked pairs existed between pleiotropic loci.

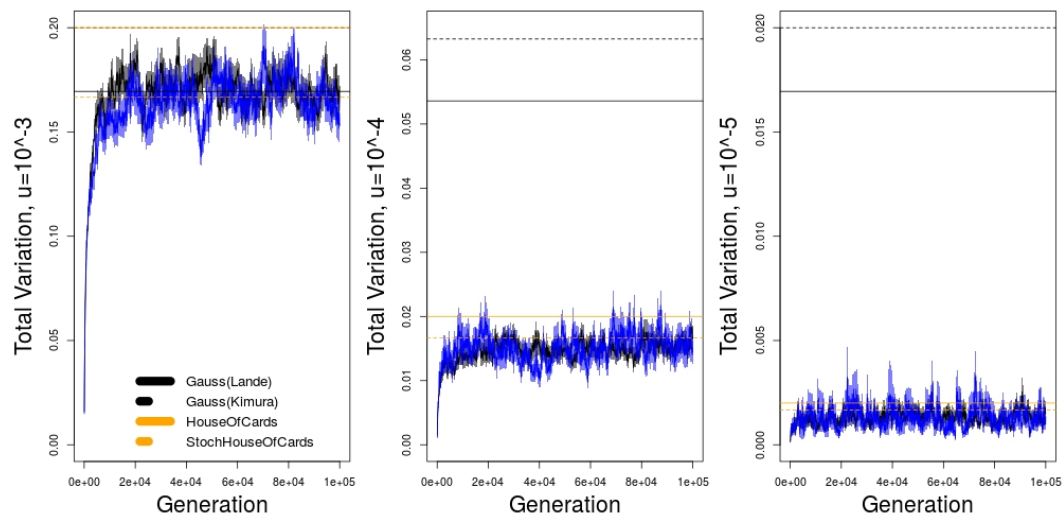


Figure S3: Average genetic variances for different mutation rates and genetic architectures, with either one pleiotropic locus or two completely linked loci, compared against theoretical expectations from several models (Bürger, 2000).

Table S1: Results of GWA analyses for different architectures with average false negatives (Type II errors) for pleiotropic architectures and false positives (Type I errors) for linkage architectures, as well as linkage disequilibrium (LD) measurement averages for short-distance (physically linked loci) and long-distance (unlinked loci) comparisons. The genetic architectures in the bottom half of the table have higher genetic correlations than the top half (created by adjusting correlational selection) to compare the differences at different genetic correlation.

Genetic Architecture	Genetic Cor (SE)	Type I/II Error %	D' short	D' long	R^2 short	R^2 long
Pleiotropy	0.308 (0.0046)	0.35%	NA	0.018	NA	0.00027
Linkage (0cM)	0.300 (0.0055)	22.06%	0.37	0.023	0.089	0.00026
Linkage (0.1cM)	0.300 (0.0045)	20.17%	0.26	0.025	0.047	0.00027
Linkage (1cM)	0.308 (0.0035)	18.28%	0.13	0.030	0.007	0.00027
Pleiotropy	0.407 (0.0048)	0.32%	NA	0.018	NA	0.00027
Linkage (0cM)	0.398 (0.0074)	28.76%	0.43	0.025	0.107	0.00027
Linkage (0.1cM)	0.408 (0.0035)	28.46%	0.30	0.027	0.050	0.00027
Linkage (1cM)	0.404 (0.0029)	25.34%	0.19	0.048	0.006	0.00027

523 **References**

524 **References**

525 Arnold, S. J. (1992). Constraints on phenotypic evolution. *Am. Nat.*, pages
526 S85–S107.

527 Baatz, M. and Wagner, G. P. (1997). Adaptive inertia caused by hidden
528 pleiotropic effects. *Theor. Popul. Biol.*, 51(1):49–66.

529 Barrett, R. D. and Hoekstra, H. E. (2011). Molecular spandrels: tests of
530 adaptation at the genetic level. *Nature Reviews Genetics*, 12(11):767.

531 Benjamini, Y. and Hochberg, Y. (1995). Controlling the false discovery rate:
532 a practical and powerful approach to multiple testing. *Journal of the royal
533 statistical society. Series B (Methodological)*, pages 289–300.

534 Bolnick, D. I. and Otto, S. P. (2013). The magnitude of local adaptation
535 under genotype-dependent dispersal. *Ecology and evolution*, 3(14):4722–
536 4735.

537 Bürger, R. (2000). *The mathematical theory of selection, recombination, and
538 mutation*, volume 228. Wiley Chichester.

539 Cai, L., Fisher, A. L., Huang, H., and Xie, Z. (2016). Crispr-mediated genome
540 editing and human diseases. *Genes & Diseases*, 3(4):244–251.

541 Cai, Z., Dusza, M., Guldbrandtsen, B., Lund, M. S., and Sahana, G. (2020).
542 Distinguishing pleiotropy from linked qtl between milk production traits
543 and mastitis resistance in nordic holstein cattle. *Genetics Selection Evo-
544 lution*, 52:1–15.

545 Carbone, M. A., Jordan, K. W., Lyman, R. F., Harbison, S. T., Leips, J.,
546 Morgan, T. J., DeLuca, M., Awadalla, P., and Mackay, T. F. (2006). Phe-
547 notypic variation and natural selection at catsup, a pleiotropic quantitative
548 trait gene in drosophila. *Current Biology*, 16(9):912–919.

- 549 Chebib, J. and Guillaume, F. (2017). What affects the predictability of
550 evolutionary constraints using a G-matrix? The relative effects of modular
551 pleiotropy and mutational correlation. *Evolution*, 71(10).
- 552 Chesmore, K., Bartlett, J., and Williams, S. M. (2018). The ubiquity of
553 pleiotropy in human disease. *Human genetics*, 137(1):39–44.
- 554 Chevin, L.-M., Martin, G., and Lenormand, T. (2010). Fisher’s model and
555 the genomics of adaptation: restricted pleiotropy, heterogenous mutation,
556 and parallel evolution. *Evolution*, 64(11):3213–3231.
- 557 Chung, D., Yang, C., Li, C., Gelernter, J., and Zhao, H. (2014). Gpa: a
558 statistical approach to prioritizing gwas results by integrating pleiotropy
559 and annotation. *PLoS genetics*, 10(11):e1004787.
- 560 Crow, J. F. and Kimura, M. (1964). The theory of genetic loads. *Proc. XI*
561 *Int. Congr. Genetics*, 2:495–505.
- 562 Draghi, J. A. and Whitlock, M. C. (2012). Phenotypic plasticity facilitates
563 mutational variance, genetic variance, and evolvability along the major
564 axis of environmental variation. *Evolution*, 66(9):2891–2902.
- 565 Dudley, A. M., Janse, D. M., Tanay, A., Shamir, R., and Church, G. M.
566 (2005). A global view of pleiotropy and phenotypically derived gene func-
567 tion in yeast. *Molecular systems biology*, 1(1).
- 568 Edelstein, M. L., Abedi, M. R., and Wixon, J. (2007). Gene therapy clinical
569 trials worldwide to 2007an update. *The Journal of Gene Medicine: A*
570 *cross-disciplinary journal for research on the science of gene transfer and*
571 *its clinical applications*, 9(10):833–842.
- 572 Falconer, D. S. and Mackay, T. F. (1996). *Introduction to quantitative genet-*
573 *ics*. Longman, London, fourth edition.

- 574 Fisher, R. A. (1930). *The genetical theory of natural selection: a complete*
575 *variorum edition*. Oxford University Press.
- 576 Flint, J. and Mackay, T. F. (2009). Genetic architecture of quantitative traits
577 in mice, flies, and humans. *Genome research*, 19(5):723–733.
- 578 Ford, C. B., Lin, P. L., Chase, M. R., Shah, R. R., Iartchouk, O., Galagan,
579 J., Mohaideen, N., Ioerger, T. R., Sacchettini, J. C., Lipsitch, M., et al.
580 (2011). Use of whole genome sequencing to estimate the mutation rate
581 of mycobacterium tuberculosis during latent infection. *Nature genetics*,
582 43(5):482–486.
- 583 Gianola, D., de los Campos, G., Toro, M. A., Naya, H., Schön, C.-C., and
584 Sorensen, D. (2015). Do molecular markers inform about pleiotropy? *Ge-*
585 *netics*, pages genetics–115.
- 586 Ginn, S. L., Amaya, A. K., Alexander, I. E., Edelstein, M., and Abedi, M. R.
587 (2018). Gene therapy clinical trials worldwide to 2017: An update. *The*
588 *journal of gene medicine*, 20(5):e3015.
- 589 Gratten, J. and Visscher, P. M. (2016). Genetic pleiotropy in complex
590 traits and diseases: implications for genomic medicine. *Genome medicine*,
591 8(1):78.
- 592 Griswold, C. K., Logsdon, B., and Gomulkiewicz, R. (2007). Neutral evolu-
593 tion of multiple quantitative characters: a genealogical approach. *Genetics*,
594 176(1):455–466.
- 595 Guillaume, F. and Otto, S. P. (2012). Gene functional trade-offs and the
596 evolution of pleiotropy. *Genetics*, 192:1389–1409.
- 597 Guillaume, F. and Rougemont, J. (2006). Nemo:an evolutionary and popu-
598 lation genetics programming framework. *Bioinformatics*, 22:2556–2557.

- 599 Guillaume, F. and Whitlock, M. C. (2007). Effects of migration on the genetic
600 covariance matrix. *Evolution*, 61(10):2398–2409.
- 601 Hansen, T. F. (2003). Is modularity necessary for evolvability?: Remarks on
602 the relationship between pleiotropy and evolvability. *Biosystems*, 69(2):83–
603 94.
- 604 Hill, W. G. and Zhang, X.-S. (2012a). Assessing pleiotropy and its evolution-
605 ary consequences: pleiotropy is not necessarily limited, nor need it hinder
606 the evolution of complexity. *Nature Reviews Genetics*, 13(4):296.
- 607 Hill, W. G. and Zhang, X.-S. (2012b). On the pleiotropic structure of the
608 genotype-phenotype map and the evolvability of complex organisms. *Ge-
609 netics*, 190(3):1131–1137.
- 610 Hodgkin, J. (1998). Seven types of pleiotropy. *Int. J. Dev. Biol.*, 42(3):501–
611 505.
- 612 Keightley, P. D., Pinharanda, A., Ness, R. W., Simpson, F., Dasmahapatra,
613 K. K., Mallet, J., Davey, J. W., and Jiggins, C. D. (2015). Estimation of
614 the spontaneous mutation rate in *Heliconius melpomene*. *Molecular biology
615 and evolution*, 32(1):239–243.
- 616 Kingman, J. F. (1978). A simple model for the balance between selection
617 and mutation. *Journal of Applied Probability*, 15(1):1–12.
- 618 Lande, R. (1980). The genetic covariance between characters maintained by
619 pleiotropic mutations. *Genetics*, 94(1):203–215.
- 620 Lande, R. (1984). The genetic correlation between characters maintained by
621 selection, linkage and inbreeding. *Genetics Research*, 44(3):309–320.
- 622 Lindsay, S. J., Rahbari, R., Kaplanis, J., Keane, T., and Hurles, M. E. (2019).
623 Similarities and differences in patterns of germline mutation between mice
624 and humans. *Nature communications*, 10(1):1–12.

- 625 Liu, M., Jiang, Y., Wedow, R., Li, Y., Brazel, D., and others, . (2019).
626 Association studies of up to 1.2 million individuals yield new insights into
627 the genetic etiology of tobacco and alcohol use. *Nature Genetics*.
- 628 Lynch, M., Walsh, B., et al. (1998). *Genetics and analysis of quantitative*
629 *traits*, volume 1. Sinauer Sunderland, MA.
- 630 Martin, G. and Lenormand, T. (2006). A general multivariate extension of
631 fisher’s geometrical model and the distribution of mutation fitness effects
632 across species. *Evolution*, 60(5):893–907.
- 633 Mills, M. G., Greenwood, A. K., and Peichel, C. L. (2014). Pleiotropic effects
634 of a single gene on skeletal development and sensory system patterning in
635 sticklebacks. *EvoDevo*, 5(1):5.
- 636 Myers, S., Spencer, C., Auton, A., Bottolo, L., Freeman, C., Donnelly, P., and
637 McVean, G. (2006). The distribution and causes of meiotic recombination
638 in the human genome. *Biochem. Soc. Trans.*, 34(4):536–530.
- 639 Nachman, M. W. and Crowell, S. L. (2000). Estimate of the mutation rate
640 per nucleotide in humans. *Genetics*, 156(1):297–304.
- 641 Nosil, P., Crespi, B., Sandoval, C., and Kirkpatrick, M. (2006). Migration
642 and the genetic covariance between habitat preference and performance.
643 *The American Naturalist*, 167(3):E66–E78.
- 644 Orr, H. A. (1998). The population genetics of adaptation: the distribution
645 of factors fixed during adaptive evolution. *Evolution*, 52(4):935–949.
- 646 Otto, S. P. (2004). Two steps forward, one step back: the pleiotropic effects
647 of favoured alleles. *Proc. R. Soc. Lond (Biol)*, 271(1540):705–714.
- 648 Parkes, M., Cortes, A., Van Heel, D. A., and Brown, M. A. (2013). Genetic
649 insights into common pathways and complex relationships among immune-
650 mediated diseases. *Nature Reviews Genetics*, 14(9):661.

- 651 Peichel, C. L. and Marques, D. A. (2017). The genetic and molecular ar-
652 chitecture of phenotypic diversity in sticklebacks. *Phil. Trans. R. Soc. B*,
653 372(1713):20150486.
- 654 Pickrell, J. K., Berisa, T., Liu, J. Z., Ségurel, L., Tung, J. Y., and Hinds,
655 D. A. (2016). Detection and interpretation of shared genetic influences on
656 42 human traits. *Nature genetics*, 48(7):709.
- 657 Platt, A., Vilhjálmsson, B. J., and Nordborg, M. (2010). Conditions un-
658 der which genome-wide association studies will be positively misleading.
659 *Genetics*.
- 660 Rausher, M. D. and Delph, L. F. (2015). Commentary: When does un-
661 derstanding phenotypic evolution require identification of the underlying
662 genes? *Evolution*, 69(7):1655–1664.
- 663 Rockman, M. V. (2012). The qtn program and the alleles that matter for
664 evolution: all that’s gold does not glitter. *Evolution: International Journal*
665 *of Organic Evolution*, 66(1):1–17.
- 666 Roff, D. A. (1996). The evolution of genetic correlations: an analysis of
667 patterns. *Evolution*, pages 1392–1403.
- 668 Saltz, J. B., Hessel, F. C., and Kelly, M. W. (2017). Trait correlations in the
669 genomics era. *Trends in ecology & evolution*, 32(4):279–290.
- 670 Siegmund, D. and Yakir, B. (2007). *The statistics of gene mapping*. Springer
671 Science & Business Media.
- 672 Stearns, F. W. (2010). One hundred years of pleiotropy: a retrospective.
673 *Genetics*, 186(3):767–773.
- 674 Turelli, M. (1984). Heritable genetic variation via mutation-selection balance:
675 Lerch’s zeta meets the abdominal bristle. *Theoretical population biology*,
676 25(2):138–193.

- 677 Turelli, M. (1985). Effects of pleiotropy on predictions concerning mutation-
678 selection balance for polygenic traits. *Genetics*, 111(1):165–195.
- 679 Turelli, M. and Barton, N. (1990). Dynamics of polygenic characters under
680 selection. *Theoretical Population Biology*, 38(1):1–57.
- 681 Verbanck, M., Chen, C.-Y., Neale, B., and Do, R. (2018). Detection
682 of widespread horizontal pleiotropy in causal relationships inferred from
683 mendelian randomization between complex traits and diseases. *Nature*
684 *genetics*, 50(5):693.
- 685 Visscher, P. M., Wray, N. R., Zhang, Q., Sklar, P., McCarthy, M. I., Brown,
686 M. A., and Yang, J. (2017). 10 years of gwas discovery: biology, function,
687 and translation. *The American Journal of Human Genetics*, 101(1):5–22.
- 688 Visscher, P. M. and Yang, J. (2016). A plethora of pleiotropy across complex
689 traits. *Nature genetics*, 48(7):707.
- 690 Wagner, G. P., Pavlicev, M., and Cheverud, J. M. (2007). The road to
691 modularity. *Nat. Rev. Genet.*, 8(12):921–931.
- 692 Wagner, G. P. and Zhang, J. (2011). The pleiotropic structure of the
693 genotype–phenotype map: the evolvability of complex organisms. *Nat.*
694 *Rev. Genet.*, 12(3):204–213.
- 695 Walsh, B. and Blows, M. W. (2009). Abundant genetic variation + strong se-
696 lection = multivariate genetic constraints: A geometric view of adaptation.
697 *Annu. Rev. Ecol. Evol. Syst.*, 40(1):41–59.
- 698 Warnes, G., with contributions from Gregor Gorjanc, Leisch, F., and Man.,
699 M. (2013). *genetics: Population Genetics*. R package version 1.3.8.1.
- 700 Watanabe, K., Stringer, S., Frei, O., Mirkov, M. U., Polderman, T. J., van der
701 Sluis, S., Andreassen, O. A., Neale, B. M., and Posthuma, D. (2018).

- 702 A global view of pleiotropy and genetic architecture in complex traits.
703 *bioRxiv*, page 500090.
- 704 Wei, W.-H., Hemani, G., and Haley, C. S. (2014). Detecting epistasis in
705 human complex traits. *Nature Reviews Genetics*, 15(11):722.
- 706 Wei, X. and Nielsen, R. (2019). Ccr5- 32 is deleterious in the homozygous
707 state in humans. *Nature medicine*, page 1.
- 708 Welch, J. J., Waxman, D., and Houle, D. (2003). Modularity and the cost
709 of complexity. *Evolution*, 57(8):1723–1734.
- 710 Welter, D., MacArthur, J., Morales, J., Burdett, T., Hall, P., Junkins, H.,
711 Klemm, A., Flicek, P., Manolio, T., Hindorff, L., et al. (2013). The nhgri
712 gwas catalog, a curated resource of snp-trait associations. *Nucleic acids
713 research*, 42(D1):D1001–D1006.
- 714 Wright, S. (1977). Evolution and the genetics of populations. volume 3. exper-
715 imental results and evolutionary deductions. *Evolution and the genetics of
716 populations. Volume 3. Experimental results and evolutionary deductions*.
- 717 Zhu, Z., Zhang, F., Hu, H., Bakshi, A., Robinson, M. R., Powell, J. E.,
718 Montgomery, G. W., Goddard, M. E., Wray, N. R., Visscher, P. M., et al.
719 (2016). Integration of summary data from gwas and eqtl studies predicts
720 complex trait gene targets. *Nature genetics*, 48(5):481.