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Polycystic ovary syndrome is a risk factor for sarcopenic obesity: a case control study



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Abstract

Background: Polycystic ovary syndrome (PCOS) is the most common endocrine disorder in young women and increases risk of cardiovascular and metabolic disease, and infertility. Women with PCOS share many characteristics commonly associated with aging including chronic inflammation and insulin resistance, which may be associated with "sarcopenic obesity", a term used to describe low appendicular skeletal muscle mass relative to total body mass. The purpose of this work was to determine the prevalence of sarcopenic obesity in women with PCOS. We hypothesized there would be a high prevalence of sarcopenic obesity, and that % appendicular skeletal muscle mass and markers of inflammation and insulin resistance would be inversely correlated in this population.

Methods: Dual energy X-ray absorptiometry was used to assess body composition in 68 women with PCOS aged 18-35y and 60 healthy age-matched women from the same geographic area. Sarcopenic obesity was defined as having % appendicular skeletal muscle mass 2 standard deviations below the mean for the healthy age-matched controls and a % body fat above 35%. Data were analyzed with Mann-Whitney U-tests and Spearman correlations.

Results: 53% of women with PCOS were classified as sarcopenic obese. Women with PCOS had a median (interguartile range) appendicular skeletal muscle mass of 23.8 (22.3-25.8)% which was lower than the control median of 30.4 (28.6–32.4)% (p < 0.0001). Among women with PCOS, there were negative correlations between % appendicular skeletal muscle mass and the homeostasis model assessment insulin resistance index (r = -0.409; p < 0.01), high sensitivity C-reactive protein (r = -0.608; p < 0.0001) and glycosylated hemoglobin (r = -0.430; p < 0.0001). Furthermore, % appendicular skeletal muscle mass correlated positively with vitamin D (r = 0.398; p < 0.0001) in women with PCOS, which is thought to positively affect skeletal muscle mass.

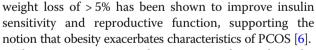
Conclusions: Women with PCOS have a high prevalence of sarcopenic obesity, which is correlated to insulin resistance and inflammation.

Keywords: Muscle, Insulin resistance, Vitamin D, Inflammation

Background

Polycystic Ovary Syndrome (PCOS) is a very common endocrine disorder in young women with a prevalence estimated to be 5-12% of premenopausal women [1]. Clinical features associated with the syndrome include menstrual and ovulatory dysfunction, hyperandrogenemia, hirsutism, polycystic ovaries, insulin resistance, and hyperinsulinism [2]. Women with PCOS are at an increased risk of developing infertility, type 2 diabetes mellitus, obesity and hypertension [1-4]. Approximately 30% of women with PCOS are obese [5], yet a modest

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The term sarcopenic obesity, primarily used to describe many older individuals, is defined as low appendicular skeletal muscle mass (ASM) relative to total body mass [7]. Although the pathogenesis is not fully understood, sarcopenic obesity is thought to be associated with increased visceral adiposity. Adipose tissue releases inflammatory cytokines, which are associated with protein degradation and sarcopenia [8, 9]. Insulin resistance is also thought to be a contributing factor to sarcopenic obesity and may exacerbate skeletal muscle loss [10]. Although sarcopenic obesity is most commonly associated with the



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aging population, women with PCOS share many of the characteristics associated with aging. Women with PCOS have increased visceral adiposity compared to BMI-matched controls [11, 12]. Indicators of impaired glucose tolerance and insulin resistance such as homeostatic model assessment are also higher in women with PCOS [13, 14]. Chronic inflammation is also prevalent in this population with higher levels of inflammatory cytokines compared to controls [15, 16]. Despite a higher prevalence of these risk factors for sarcopenic obesity the prevalence of sarcopenic obesity in women with PCOS is currently unknown.

The primary objective of this study was to determine the prevalence of sarcopenic obesity in a sample of women with PCOS. Furthermore, we wanted to determine whether the metabolic measures associated with sarcopenic obesity in older adults were also associated with sarcopenic obesity in women with PCOS. We hypothesized that women with PCOS would have a high prevalence of sarcopenic obesity and that ASM would correlate negatively with markers of inflammation and insulin resistance (i.e. high sensitivity C-reactive protein [hsCRP], and the homeostasis model assessment insulin resistance index [HOMA]).

Methods

The results in the present report are baseline measures from 68 participants involved in a larger study that determined the effect of a pulse-based diet (e.g., beans, lentils, chick-peas) and exercise intervention in women with PCOS [17]. Sixty healthy age-matched control women from the same city (i.e. Saskatoon SK, Canada) were also recruited to serve as a reference group to determine the prevalence of sarcopenic obesity in women with PCOS. All participants were between 18 and 35 years of age. Control participants were recruited via postings on the University of Saskatchewan website requesting participation from healthy females between 18 and 35 years old with regular menstrual cycles. Controls were excluded if they had any self-identified chronic conditions or irregular menstrual cycles. Women with PCOS were recruited via posters, contacting doctors' offices and postings on the university website. To be included in the control group, women must not have been diagnosed with any chronic medical conditions, must not have been pregnant and must have had regular menstrual cycles. In the PCOS group, a diagnosis of PCOS was made by an obstetrician-gynecologist using criteria specified in the PCOS report of the Androgen Excess-PCOS Society Task Force [2]. A diagnosis of PCOS required 1) either oligoamenorrhea and/or polycystic ovaries defined as > 25 follicles visualized by transvaginal ultrasonography to reflect the newest guidelines for polycystic ovaries (PCO) recommended by the Androgen Excess and Polycystic Ovary Syndrome Society, and 2) hyperandrogenism as defined by a Ferriman and Gallwey score of >6 and/or biochemical hyperandrogenemia [2, 18, 19]. Women with PCOS associated with the following conditions were excluded from the study: Taking anti-seizure or anti-psychotic medications known to induce development of PCO; untreated hyperprolactinemia or thyroid disease; or, excessive adrenal androgen production confirmed by a diagnosis of congenital adrenal hyperplasia, Cushing's disease or syndrome, or an adrenal tumor. Exclusion criteria for the PCOS study group included the use of hormonal birth control methods or fertility medications during the prior 3 months, any uncontrolled medical condition that interfered with ovarian or systemic hormone production, or women who were pregnant or breastfeeding. The screening for the PCOS study group was part of a diet and exercise intervention so exclusion criteria also included residing outside of the local geographic area or women with a medical condition limiting exercise or consumption of a pulse-based diet (allergies or intolerances). The study was approved by the University of Saskatchewan Biomedical Research Ethics Board and all women gave written informed consent before participation in the study.

Body composition was assessed using dual-energy X-ray absorptiometry (Hologic© Discovery Wi; Bedford, MA), performed and analyzed by a certified nuclear medicine technologist. A quality-control phantom scan was performed daily. Coefficients of variation for fat and bone mineral-free lean tissue mass were 3 and 0.5% respectively. Body mass was determined with a calibrated scale and height determined with a standiometer. Body mass index (BMI) was determined as mass (kg) divided by height squared (m²). Waist circumference was measured after removing clothing from the abdomen, by locating the top of the iliac crest and measuring horizon-tally at the tip of the landmark.

Percent ASM was calculated as the total bone mineral-free lean tissue mass of the arms and legs divided by total body mass times 100. Sarcopenic obesity was defined as having a %ASM two standard deviations (SD) below the mean value for the young sex-matched control group [20, 21] as well as a percentage body fat above 35% [22].

Blood was collected for the PCOS group following a 10 to 12 h fast and was centrifuged after a complete clot was formed. All samples were analyzed by the Saskatoon Health Region Laboratory in Saskatoon, Saskatchewan. Glucose and insulin levels were analyzed using the IMMULITE 2000 Systems Analyzer's solid-phase enzyme-labelled chemiluminescent immunometric assay (Siemens, California). HOMA was calculated as fasting plasma insulin (pmol/L) * fasting plasma glucose (mmol/L)/22.5 * 10 (correction factor for SI units). The remaining metabolites were analyzed using Roche Cobas analyzers and kits (Cobas

Roche Diagnostic, Basel, Switzerland) with glycosylated hemoglobin (HbA1C) analyzed using turbidimetric inhibition immunoassay, hsCRP analyzed using a particle enhanced immunoturbidimetric assay, and 25-hydroxyvitamin D analyzed using an electrochemiluminescence immunoassay. The intra-assay % coefficient of variation was < 5% for all assays. The limit of detection was 0.18 mmol/L for HbA1C, 0.15 mg/L for hsCRP, 7.5 nmol/L for vitamin D, and 0.11 mmol/L for glucose. Functional sensitivity was 0.3 mg/L for hsCRP and 4.01 ng/ml for Vitamin D.

The minimal sample size required was determined using the mean and SD of our control group for %ASM ($30.2 \pm 3.0\%$) and a predicted mean for sarcopenic obesity that was 2 SD lower (i.e. 24.1%, as a clinical cut-off for sarcopenic obesity [20–22]), a power of 90% and alpha of 0.05. This resulted in a required sample size of n = 7 per group.

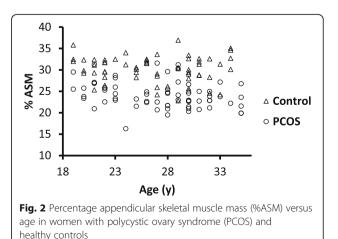
Differences in characteristics between women with PCOS and control women were determined using a Mann-Whitney U-test as data did not follow a normal distribution. Women with PCOS were classified as sarcopenic obese according to the criteria of having a %ASM 2 SD below the mean for our female control group (n = 60), as well a % body fat above 35% [19–21]. All other women were classified as non-sarcopenic obese. Differences between sarcopenic obese and nonsarcopenic obese women with PCOS were determined using a Mann-Whitney U-test as data did not follow a normal distribution. Spearman correlations for anthropometric measures with metabolic markers were determined for all women with PCOS. Spearman correlations were also determined for %ASM and metabolic markers in both non-sarcopenic obese and sarcopenic obese women with PCOS, separately. All data were analyzed using Statistica version 12 (Statsoft, Chicago IL). Group comparisons are presented as median and interquartile range, with descriptive statistics of clinical blood measures and %ASM presented as means ± SD. A P-value < 0.05 was considered significant.

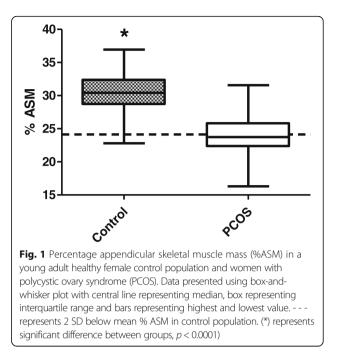
Results

The median BMI and age for the control group were 21.9 (20.5–23.8) kg/m² and 27 (22.0–30.0) years, respectively. The median %ASM for the control group was 30.4 (28.6–32.4) % (Fig. 1). The mean %ASM for the control group was $30.2 \pm 3.0\%$ and the cut-off for sarcopenic obesity (defined as 2 SD below the mean of the control group) was 24.1% (Fig. 1). In women with PCOS the median BMI and age were 31.7 (25.6–37.4) kg/m² and 28 (23.0–30.8) years, respectively. In women with PCOS the median %ASM was 23.8 (22.3–25.8) %; 53% of women with PCOS were defined as sarcopenic obese. Compared to controls, women with PCOS had a higher BMI (p < 0.0001) and lower %ASM (p < 0.0001) (Fig. 1),

with no difference in age between the groups. Figure 2 shows %ASM plotted against age for women with PCOS and controls. The plot shows that across age groups, women with PCOS had consistently lower %ASM compared to controls.

The levels of Vitamin D, hsCRP, and HbA1C in women with PCOS were 61.9 ± 31.6 nmol • L⁻¹, 4.65 ± 4.77 mg • L⁻¹ and $5.32 \pm 0.44\%$, respectively. HOMA in women with PCOS was 3.85 ± 3.88 , which is categorized as insulin resistant [23]. Levels of hsCRP and HbA1C were in the normal clinical range in women with PCOS while levels of vitamin D were insufficient according to our hospital lab (Saskatoon Health region) where normal reference ranges of hsCRP and HbA1C are 0–7 mg • L⁻¹ and 4.5-6.5%, respectively, and normal ranges for





vitamin D are 25–70 and 70–250 nmol \cdot L⁻¹ for relative insufficient and optimal, respectively.

Percent ASM and percent bone-free lean tissue mass (% BFLTM) correlated negatively with hsCRP, HOMA, HbA1C, and positively with Vitamin D in women with PCOS (all p < 0.01) (Table 1). Percent total-body fat mass (% FM), BMI, and waist circumference correlated positively with hsCRP, HOMA and HbA1C, and negatively with Vitamin D in women with PCOS (all p < 0.01) (Table 1).

Among women with PCOS, 36 were classified as sarcopenic obese and 32 were non-sarcopenic obese. Sarcopenic obese women had a higher BMI and waist circumference as well as lower %ASM than non-sarcopenic obese women (p < 0.0001) (Table 2). Among women with PCOS, women with sarcopenic obesity also had higher levels of hsCRP (p < 0.0001), HOMA (p < 0.01), HbA1C (p < 0.01) and lower levels of Vitamin D (p < 0.01) (Table 2). In non-sarcopenic obese women with PCOS, %ASM remained negatively correlated with HOMA (p < 0.05) and hsCRP (p < 0.001), and positively correlated with Vitamin D (p < 0.001) (Fig. 3). In sarcopenic obese women with PCOS, %ASM remained negatively correlated negatively correlated negatively correlated with NCRP (p < 0.05) and HbA1C (p < 0.05) (Fig. 3).

Discussion

This study determined the prevalence of sarcopenic obesity to be 53% in our sample of women with PCOS. Among this population %ASM was positively associated with vitamin D and negatively associated with HOMA, hsCRP and HbA1C, with only hsCRP and HbA1C maintaining significance in sarcopenic obese women with PCOS. The prevalence of sarcopenic obesity was based on the percentage of women who had a %ASM lower than 24.1%, which was 2 SD below the mean for a young age-matched healthy female control group. This reference value is in keeping with other healthy young adult control groups from previous studies, which ranged from 21.1–25.1% [24, 25].

Table 1 Correlations between anthropometric measures andmetabolic markers in women with polycystic ovary syndrome

	%ASM	%BFLTM	%FM	BMI	WC
hsCRP	-0.608**	-0.755**	0.733**	0.571**	0.628**
HOMA	-0.409*	-0.502**	0.520**	0.575**	0.581**
Vitamin D (25-OH)	0.398*	0.430**	-0.430**	-0.374*	-0.379*
HbA1C	-0.430**	-0.538**	0.532**	0.479**	0.410*

Correlation coefficient and *p* values determined using Spearman correlations. (*) represents significant correlation, p < 0.01, (**) represents significant correlation, p < 0.001. % ASM, % appendicular skeletal muscle mass; % BFLTM, % bone-free lean tissue mass; % FM, % fat mass; BMI, body mass index; HbA1C, glycosylated hemoglobin; HOMA, homeostasis model assessment insulin resistance index hsCRP, high sensitive C-reactive protein; WC, waist circumference

The term sarcopenic obesity as a classification [7] is not clearly defined in the literature and currently more than eight definitions can be found, leading to a highly variable prevalence range of 3.6-94% in older women [26]. Using similar criteria of %ASM and %body fat, the prevalence of sarcopenic obesity in a primarily Korean population is 3.3 and 12.5% in women aged 40-59 and \geq 60, respectively [20], which is similar to the prevalence of 10.9% found in a Canadian population of older adults aged 68-82 years [22]. Although the pathogenesis of sarcopenic obesity is not fully understood, physical inactivity, increased visceral adiposity, increased inflammation, hormonal changes and insulin resistance are thought to be the primary factors associated with sarcopenic obesity [27]. From our results, women with PCOS may have a higher prevalence of sacropenic obesity than an older population; this is most likely due to the altered metabolic profile and increased inflammation associated with PCOS.

Both normal-weight and obese women with PCOS have increased visceral adiposity [11, 12], and visceral adiposity is associated with higher levels of inflammatory cytokines [8, 28], as well as the inflammatory marker hsCRP [29]. This body composition profile may in part explain the low grade chronic inflammation [30] and higher levels of hsCRP [15] associated with PCOS. In this study we used waist circumference as an indicator of visceral adiposity, which was positively associated with hsCRP, HOMA and HbA1C in women with PCOS. This is in keeping with another study which found a positive association between hsCRP and waist circumference in women with PCOS [31]. We also found a negative correlation between hsCRP and %ASM in women with PCOS which is consistent with studies in older adults [27] and obese individuals [32]. Inflammatory cytokines are associated with muscle catabolism and sarcopenia [9]. Furthermore, low grade inflammation negatively affects protein synthesis [33]. Thus, chronic inflammation in women with PCOS may be a contributing factor to the high prevalence of sarcopenic obesity in this population. Interestingly, although sarcopenic obesity has primarily been studied in older adults, it has been demonstrated that hsCRP has a stronger association with sarcopenic obesity in females less than 60 years old [34].

Insulin resistance is more prevalent in women with PCOS compared to both lean and overweight BMImatched control groups [35], which may contribute to sarcopenic obesity in this population. In this study, HOMA was used as an indicator of insulin resistance and we demonstrated a negative correlation between HOMA scores and %ASM in women with PCOS. In addition, HbA1C, a marker of glycemic control, was negatively associated with %ASM in women with PCOS. Insulin is an anabolic hormone and insulin resistance is

Table 2 Characteristics and metabolic markers in sarcopenic obese and non-sarcopenic obese women with polycysti	c ovary
syndrome	

	Non-Sarcopenic Obese	Sarcopenic Obese	Normal Clinical Range
BMI	27.1 (22.3–30.9)**	36.0 (30.4–39.7)	
%ASM	26.3 (25.0–28.3)**	22.5 (21.0–23.0)	
Waist Circumference (cm)	88.8 (78.8–100.4)**	108.9 (95.6–119.6)	
hsCRP (mg \cdot L ⁻¹)	0.9 (0.4–4.5)**	5.1 (2.0–9.9)	0-7 ^a
HOMA	2.38 (0.67–3.44)*	3.55 (1.74–5.90)	< 2.73 ^b
Vitamin D (25-OH) (nmol \cdot L ⁻¹)	63.8 (42.9–99.5)*	49.6 (39.1–65.6)	25–70 (relative insufficient); 70–250 (optimal)ª
HbA1C (%)	5.1 (4.9–5.4)*	5.5 (5.2–5.6)	4.5–6.5 ^a

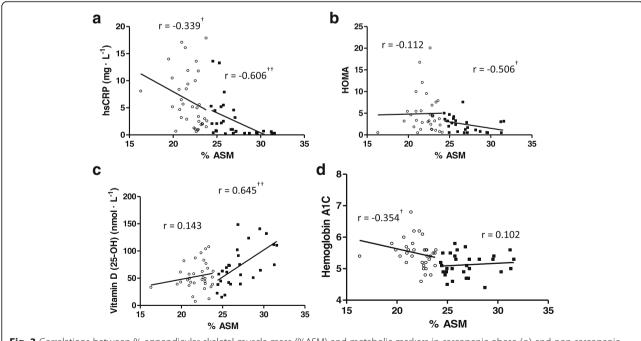
Data are presented as median (interquartile range). (*) represents significant difference between groups, p < 0.01; (**) represents significant difference between groups, p < 0.001. %ASM, % appendicular skeletal muscle mass; BMI, body mass index; HbA1C, glycosylated hemoglobin; HOMA, homeostasis model assessment insulin resistance index hsCRP, high sensitive C-reactive protein

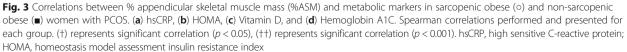
^aSaskatoon Health Region, Saskatoon, SK

^bCut off value for insulin resistance [20]

associated with accelerated muscle protein degradation [36]. Our results are supported by studies demonstrating a positive correlation between fat to lean mass ratio and HOMA in women with PCOS [37] as well as an association between percent lean mass with better insulin sensitivity in obese women [32]. Collectively, these studies and our data support the hypothesis that insulin resistance may contribute to sarcopenia in women with PCOS.

Vitamin D has recently received attention for the role it may play in positively affecting skeletal muscle growth and treatment of sarcopenia [38]. Although the existence of the vitamin D receptor (VDR) in muscle cells is still debated, it has been hypothesized that VDR signaling is important for normal muscle growth and vitamin D may have an anabolic effect by modulating intracellular signaling pathways and contributing to myoblast proliferation [38]. In this study, plasma 25-hydroxyvitamin D correlated positively with %ASM and negatively with waist circumference in women with PCOS. These results are consistent with others who have demonstrated a positive association between vitamin D and percent lean





mass in obese individuals [32] and a negative association between vitamin D and waist circumference in women with PCOS [31]. Furthermore, vitamin D remains positively associated with skeletal muscle mass after adjusting for body fat mass in obese individuals [39]. A large proportion of the women with PCOS had low vitamin D levels (Table 2); however, this is typical of young Canadian women because of the limited sun exposure due to northern latitude. The proportion of women with PCOS with vitamin D level lower than 50 nmol/L in our study (41%) is identical to that of Canadians in the same age group from the general population [40].

In this study women with PCOS and sarcopenic obesity had a higher waist circumference and HOMA, as well as higher levels of hsCRP and HbA1C compared to women with PCOS but without sarcopenic obesity. Low grade chronic inflammation has been proposed as a primary contributor to sarcopenic obesity in older adults, particularly in women [41]. Interestingly, we demonstrated that in sarcopenic obese women with PCOS, only hsCRP and HbA1C maintained a significant negative correlation with %ASM. In non-sarcopenic obese women with PCOS, hsCRP and HOMA were negatively, and 25-hydroxyvitamin D positively correlated with %ASM. These results suggest that in non-sarcopenic obese women with PCOS vitamin D may play a protective role against sarcopenic obesity. Furthermore, sarcopenic obesity may be more associated with inflammation versus insulin resistance in women with PCOS. This is supported by a study demonstrating elevated hsCRP in both obese and non-obese women with PCOS, while only obese women demonstrated insulin resistance [42]. In newly diagnosed untreated diabetics, hsCRP appears to be a reflection of adiposity and is not associated with insulin resistance [43]; thus, the increased visceral adiposity and associated inflammation in women with PCOS may be an independent contributor to sarcopenic obesity in this population.

One of the strengths of this study is the inclusion of > 25 follicles in the definition of PCOS which has been suggested to provide a more accurate definition of the syndrome [19]. In contrast, PCOS was not absolutely ruled out in our control population which is a limitation of this study, as examination for hyperandrogenicity and polycystic ovaries was not completed. Another limitation is the use of correlations between metabolic markers and %ASM; causation cannot be assumed. In addition, in our control group we only determined body composition measures and did not assess metabolic markers or waist circumference; therefore, these measures could not be directly compared to the women with PCO. Although we determined a positive correlation between vitamin D and %ASM in women with PCOS, the interpretation of this result and other correlations is limited given comparison with control correlations cannot be made and results cannot be assumed in this group. Longitudinal studies are required to determine whether changes in metabolic markers over time or with treatment result in increased ASM in women with PCOS. Although our PCOS and control group were recruited from the same population, genetic heterogeneity among the population may have affected our results. A final limitation of the study is that our results are specific to women between the ages of 18 and 35y and therefore not generalizable to ages outside this range.

Conclusion

In conclusion, we report the prevalence of sarcopenic obesity to be 53% in women with PCOS, which is higher than the prevalence in older women reported from the literature. The correlation of %ASM with hsCRP in both sarcopenic obese and non-sarcopenic obese women with PCOS suggests inflammation may play a primary contributing factor to the high prevalence of sarcopenic obesity in this population. Future studies should focus on whether reduction of inflammatory markers can reduce the prevalence of sarcopenic obesity in women with PCOS.

Abbreviations

ASM: Appendicular skeletal muscle mass; BFLTM: Bone-free leant tissue mass; BMI: Body mass index; FM: Fat mass; HbA1C: Glycosylated hemoglobin; HOMA: Homeostasis model assessment insulin resistance index; hsCRP: High sensitivity C-reactive protein; PCOS: Polycystic ovary syndrome; VDR: Vitamin D receptor

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

LEM was involved with data collection, blood analysis, statistical analysis, and writing of the manuscript. PDC was involved with study planning, statistical analysis, and manuscript editing. JJG was involved with data collection and manuscript editing. DRC was involved with study planning, data collection, and manuscript editing. GAZ was involved in study planning and manuscript editing. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the University of Saskatchewan Biomedical Research Ethics Board (Ethics reference numbers Bio 10–98 and Bio-327). Informed consent was obtained from all individual participants included in the study.

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

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References

- Liepa GU, Sengupta A, Karsies D. Polycystic ovary syndrome (PCOS) and other androgen excess-related conditions: can changes in dietary intake make a difference? Nutr Clin Pract. 2008;23:63–71.
- Azziz R, Carmina E, Dewailly D, Diamanti-Kandarakis E, Escobar-Morreale HF, Futterweit W, et al. The androgen excess and PCOS society criteria for the polycystic ovary syndrome: the complete task force report. Fertil Steril. 2009;91:456–88.
- 3. Ehrmann DA. Polycystic ovary syndrome. N Engl J Med. 2005;352:1223-36.
- Lujan ME, Chizen DR, Pierson RA. Diagnostic criteria for polycystic ovary syndrome: pitfalls and controversies. J Obstet Gynaecol Can. 2008;30:671–9.
- Azziz R, Woods KS, Reyna R, Key TJ, Knochenhauer ES, Yildiz BO. The prevalence and features of the polycystic ovary syndrome in an unselected population. J Clin Endocrinol Metab. 2004;89:2745–9.
- Holte J, Bergh T, Berne C, Wide L, Lithell H. (1995) restored insulin sensitivity but persistently increased early insulin secretion after weight loss in obese women with polycystic ovary syndrome. J Clin Endocrinol Metab. 1995;80: 2586–93.
- Baumgartner RN. Body composition in healthy aging. Ann N Y Acad Sci. 2000;904:437–48.
- Chilibeck PD, Pérez-López FR, Bodary P F, Kang ES, Jeon JY. Adipocytokines, metabolic syndrome, and exercise. Int J Endocrinol. 2014;2014:597162.
- Reid MB, Li YP. Tumor necrosis factor-alpha and muscle wasting: a cellular perspective. Respir Res. 2001;2:269–72.
- Lee SW, Youm Y, Lee WJ, Choi W, Chu SH, Park YR, Kim HC. Appendicular skeletal muscle mass and insulin resistance in an elderly korean population: the korean social life, health and aging project-health examination cohort. Diabetes Metab J. 2015;39:37–45.
- Karabulut A, Yaylali GF, Demirlenk S, Sevket O, Acun A. Evaluation of body fat distribution in PCOS and its association with carotid atherosclerosis and insulin resistance. Gynecol Endocrinol. 2012;28:111–4.
- 12. Tripathy P, Sahu A, Sahu M, Nagy A. Ultrasonographic evaluation of intraabdominal fat distribution and study of its influence on subclinical atherosclerosis in women with polycystic ovarian syndrome. Eur J Obstet Gynecol Reprod Biol. 2017;217:18–22.
- Deugarte C, Bartolucci A, Azziz R. Prevalence of insulin resistance in the polycystic ovary syndrome using the homeostasis model assessment. Fertil and Steril. 2005;83:1454–60.
- Kazemi M, Pierson RA, Lujan ME, Chilibeck PD, McBreairty LE, Gordon JJ, Serrao SB, Zello GA, Chizen DR. Comprehensive evaluation of type 2 diabetes and cardiovascular disease risk profiles in reproductive-age women with polycystic ovary syndrome: a large Canadian cohort. J Obstet Gynaecol Can. 2019;(18):30991–5.
- Escobar-Morreale HF, Luque-Ramírez M, González F. Circulating inflammatory markers in polycystic ovary syndrome: a systematic review and metaanalysis. Fertil Steril. 2011;95:1048–58.
- Zafari Zangeneh F, Naghizadeh MM, Masoumi M. Polycystic ovary syndrome and circulating inflammatory markers. Int J Reprod Biomed. 2017;15:375–82.
- 17. Kazemi M, McBreairty LE, Chizen DR, Pierson RA, Chilibeck PD, Zello GA. A comparison of a pulse-based diet and the therapeutic lifestyle changes diet in combination with exercise and health counselling on the cardio-metabolic risk profile in women with polycystic ovary syndrome: a randomized controlled trial. Nutrients. 2018;10:E1387.

- Lujan ME, Jarrett BY, Brooks ED, Reines JK, Peppin AK, Muhn N, et al. Updated ultrasound criteria for polycystic ovary syndrome: reliable thresholds for elevated follicle population and ovarian volume. Hum Reprod. 2013;28:1361–8.
- Dewailly D, Lujan ME, Carmina E, Cedars MI, Laven J, Norman RJ, Escobar-Morreale HF. Definition and significance of polycystic ovarian morphology: a task force report from the androgen excess and polycystic ovary syndrome society. Hum Reprod Update. 2014;20:334–52.
- Kim TN, Yang SJ, Yoo HJ, Lim KI, Kang HJ, Song W, et al. Prevalence of sarcopenia and sarcopenic obesity in Korean adults: the Korean sarcopenic obesity study. Int J Obes. 2009;33:885–92.
- Hwang B, Lim JY, Lee J, Choi NK, Ahn YO, Park BJ. Prevalence rate and associated factors of sarcopenic obesity in Korean elderly population. J Korean Med Sci. 2012;27:748–55.
- Bouchard DR, Dionne IJ, Brochu M. Sarcopenic/obesity and physical capacity in older men and women: data from the nutrition as a determinant of successful aging (NuAge)—the Quebec longitudinal study. Obesity. 2009;17:2082.
- 23. Sumner AE, Cowie CC. Ethnic differences in the ability of triglyceride levels to identify insulin resistance. Atherosclerosis. 2008;196:696–703.
- Moon SS. Low skeletal muscle mass is associated with insulin resistance, diabetes, and metabolic syndrome in the Korean population: the Korea National Health and nutrition examination survey (KNHANES) 2009-2010. Endocr J. 2014;61:61–70.
- Lim S, Kim JH, Yoon JW, Kang SM, Choi SH, Park YJ, et al. Sarcopenic obesity: prevalence and association with metabolic syndrome in the Korean longitudinal study on health and aging (KLoSHA). Diabetes Care. 2010;33: 1652–4.
- Batsis JA, Barre LK, Mackenzie TA, Pratt SI, Lopez-Jimenez F, Bartels SJ. Variation in the prevalence of sarcopenia and Sarcopenic obesity in older adults associated with different research definitions: dual-energy X-ray absorptiometry data from the National Health and nutrition examination survey 1999-2004. J Am Geriatr Soc. 2013;61:974–80.
- Schrager MA, Metter EJ, Simonsick E, Ble A, Bandinelli S, Lauretani F, Ferrucci L. Sarcopenic obesity and inflammation in the InCHIANTI study. J Appl Physiol. 2007;102:919–25.
- Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of Adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. Endocrinology. 2004;145:2273–82.
- Forouhi NG, Sattar N, McKeigue PM. Relation of C-reactive protein to body fat distribution and features of the metabolic syndrome in Europeans and south Asians. Int J Obes Relat Metab Disord. 2001;25:1327–31.
- Tao T, Li S, Zhao A, Zhang Y, Liu W. Expression of the CD11c gene in subcutaneous adipose tissue is associated with cytokine level and insulin resistance in women with polycystic ovary syndrome. Eur J Endocrinol. 2012;167:705–13.
- Maidana P, Fritzler A, Mocarbel Y, Perez Lana MB, González D, Rosales M, González de Chazal F, Sternberg K, Lamas Majek E, Mallea-Gil S, Genovesi E, Pelayo M, Fabre B, Mesch V. Association between vitamin D and adrenal parameters with metabolic and inflammatory markers in polycystic ovary syndrome. Sci Rep. 2019;9:3968.
- Fornari R, Francomano D, Greco EA, Marocco C, Lubrano C, Wannenes F, et al. Lean mass in obese adult subjects correlates with higher levels of vitamin D, insulin sensitivity and lower inflammation. J Endocrinol Investig. 2014;38:367–72.
- Balage M, Averous J, Rémond D, Bos C, Pujos-Guillot E, Papet I, et al. Presence of low-grade inflammation impaired postprandial stimulation of muscle protein synthesis in old rats. J Nutr Biochem. 2010;21:325–31.
- Park CH, Do JG, Lee YT, Yoon KJ. Sarcopenic obesity associated with highsensitivity C-reactive protein in age and sex comparison: a two-center study in South Korea. BMJ Open. 2018;8:e021232.
- Stepto NK, Cassar S, Joham AE, Hutchison SK, Harrison CL, Goldstein RF, et al. Women with polycystic ovary syndrome have intrinsic insulin resistance on euglycaemic-hyperinsulaemic clamp. Hum Reprod. 2013;28:777–84.
- Wang X, Hu Z, Hu J, Du J, Mitch WE. Insulin resistance accelerates muscle protein degradation: activation of the ubiquitin-proteasome pathway by defects in muscle cell signaling. Endocrinology. 2006;147:4160–8.
- Ezeh U, Pall M, Mathur R, Azziz R. Association of fat to lean mass ratio with metabolic dysfunction in women with polycystic ovary syndrome. Hum Reprod. 2014;29:1508–17.

- Wagatsuma A, Sakuma K. Vitamin D signaling in Myogenesis: potential for treatment of sarcopenia. Biomed Res Int. 2014;2014:121254.
- Shantavasinkul PC, Phanachet P, Puchaiwattananon O, Chailurkit LO, Lepananon T, Chanprasertyotin S, et al. Vitamin D status is a determinant of skeletal muscle mass in obesity according to body fat percentage. Nutrition. 2015;31:801–6.
- Vitamin D blood levels of Canadians. Statistics Canada Catalogue no. 82– 624-X. https://www150.statcan.gc.ca/n1/pub/82-624-x/2013001/article/ 11727-eng.htm. Accessed 27 Mar 2019.
- Kim TN, Park MS, Lim KI, Choi HY, Yang SJ, Yoo HJ, et al. Relationships between sarcopenic obesity and insulin resistance, inflammation, and vitamin D status: the Korean Sarcopenic obesity study. Clin Endocrinol. 2013;78:525–32.
- 42. Bannigida DM, Shivananda Nayak B, Vijayaraghavan R. Insulin resistance and oxidative marker in women with PCOS. Arch Physiol Biochem. https://doi.org/10.1080/13813455.2018.1499120 Epub ahead of print.
- Kahn SE, Zinman B, Haffner SM, O'Neill MC, Kravitz BG, Yu D, et al. Obesity is a major determinant of the association of C-reactive protein levels and the metabolic syndrome in type 2 diabetes. Diabetes. 2006;55:2357–64.

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