





Review

# Polyester Polymeric Nanoparticles as Platforms in the Development of Novel Nanomedicines for Cancer Treatment

Enrique Niza <sup>1,2</sup> , Alberto Ocaña <sup>3</sup>, José Antonio Castro-Osma <sup>1,2</sup> , Iván Bravo <sup>1,2,\*</sup>   
and Carlos Alonso-Moreno <sup>1,2,\*</sup> 

<sup>1</sup> Centro Regional de Investigaciones Biomédicas, Unidad NanoCRIB, 02008 Albacete, Spain; enrique.niza@uclm.es (E.N.); joseantonio.castro@uclm.es (J.A.C.-O.)

<sup>2</sup> School of Pharmacy, University of Castilla-La Mancha, 02008 Albacete, Spain

<sup>3</sup> Experimental Therapeutics Unit, Hospital Clínico San Carlos, IdISSC and CIBERONC, 28040 Madrid, Spain; albertoo@sescam.jccm.es

\* Correspondence: ivan.bravo@uclm.es (I.B.); Carlos.amoreno@uclm.es (C.A.-M.); Tel.: +34-967-599200 (I.B. & C.A.-M.)

**Simple Summary:** Despite the existence of powerful therapeutic agents, cancer is still an incurable disease in many clinical scenarios. In this regard, nanomedicine and particularly polymeric nanoparticles have raised attention as a manner to improved drug delivery. Polymeric nanoparticles can optimize existent compounds or be used to improve the formulation for novel therapeutics. In this article the advantages and disadvantages of polymeric nanoparticles will be discussed, and current nanodevices, raw materials for their formulation, methods of formulation, and polymeric nanoparticles in clinical investigations will be reviewed. Finally, options for improvement and clinical applications will be suggested.



**Citation:** Niza, E.; Ocaña, A.; Castro-Osma, J.A.; Bravo, I.; Alonso-Moreno, C. Polyester Polymeric Nanoparticles as Platforms in the Development of Novel Nanomedicines for Cancer Treatment. *Cancers* **2021**, *13*, 3387. <https://doi.org/10.3390/cancers13143387>

Academic Editor: Clare Hoskins

Received: 28 May 2021

Accepted: 29 June 2021

Published: 6 July 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Abstract:** Many therapeutic agents have failed in their clinical development, due to the toxic effects associated with non-transformed tissues. In this context, nanotechnology has been exploited to overcome such limitations, and also improve navigation across biological barriers. Amongst the many materials used in nanomedicine, with promising properties as therapeutic carriers, the following one stands out: biodegradable and biocompatible polymers. Polymeric nanoparticles are ideal candidates for drug delivery, given the versatility of raw materials and their feasibility in large-scale production. Furthermore, polymeric nanoparticles show great potential for easy surface modifications to optimize pharmacokinetics, including the half-life in circulation and targeted tissue delivery. Herein, we provide an overview of the current applications of polymeric nanoparticles as platforms in the development of novel nanomedicines for cancer treatment. In particular, we will focus on the raw materials that are widely used for polymeric nanoparticle generation, current methods for formulation, mechanism of action, and clinical investigations.

**Keywords:** nanomedicine; polymeric nanoparticles; drug delivery system; cancer treatment



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

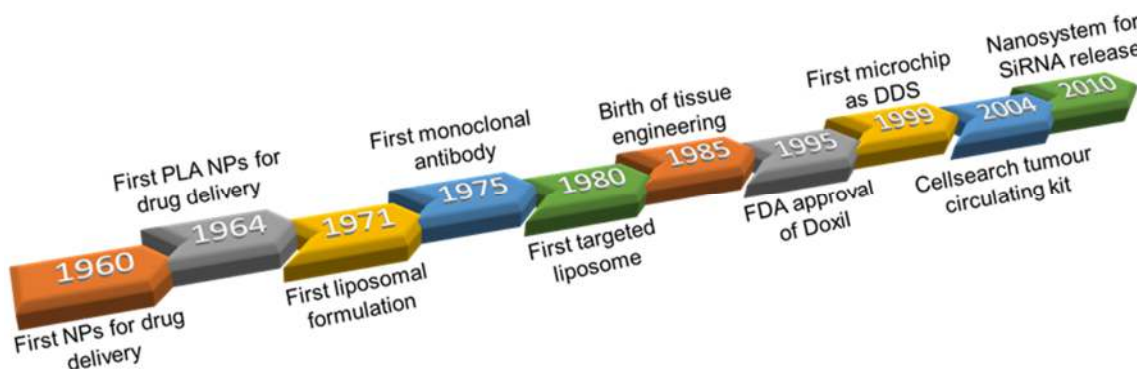
## 1. Introduction

At the end of the nineties, nanomedicine arose as a panacea for the diagnosis and treatment of diseases. However, today, nanomedicine is still there, waiting for its potential to be fully tapped.

The history of nanomedicine is linked to the evolution of high-resolution microscopy. The first visualization of <4 nm structures took place in 1902, and was performed by Richard Zsigmondy and Henry Siedentopf. Fifty years later, the disposition of the atoms over surfaces was reported, for the first time, by Erwin Müller in 1951 using ion field microscopy. In this regard, the development of the first atomic force microscope in 1986 allowed us to finally see nanostructures in high resolution [1]. These findings attracted great interest in the field of medicine, and a surge of scientific studies and research allowed

us to come up with the term “nanomedicine” [2]. Nevertheless, the first time we saw this concept on paper was in the book “What is Nanomedicine” in 1999, by Robert A. Freiras [3].

Figure 1 illustrates the most important events in the development of nanomedicine as a branch of science. Looking back, we saw the first nanoparticles (NPs) as drug delivery systems (DDS), reported in the late 1960s by Peter Paul Speiser [4]. In 1964, Kulkarni et al. used biodegradable and biocompatible polymers to make NPs for the first time [5]. In the early seventies, Georges Jean Franz Köhler and César Milstein produced monoclonal antibodies (mAbs), which Leserman et al. used to make immunoliposomes years later [6]. This event is considered one of the greatest moments in the history of nanomedicine, because it represented the first “Targeted Nanotherapy”. There were other important findings aimed to implement nanotechnology in the field of medicine, such as the use of dendrimers [7] or chips [8], but, undoubtedly, the use of nanomaterials in tissue engineering paved the way for considering nanomedicine as an area of expertise in science [9]. Aside from this, of particular interest to cancer treatment researchers, is the approval of Doxil® [10] by the Food and Therapeutic Administration (FDA) as the first encapsulated therapeutic, which meant a step forward in the development of new treatments. In fact, it accelerated the registration and marketing approvals of key pharmaceuticals in developed countries [11].



**Figure 1.** Major discoveries made in the field of nanomedicine, which are linked to the evolution of high-resolution microscopy.

Nanomedicine provides hope to improve current cancer treatment. In this sense, nanoparticles can offer several advantages in comparison to conventional chemotherapeutics based on the enhanced permeability and retention (EPR) effect. The drug can be delivered in high concentrations to the site of interest, reducing the effects to the surrounding tissues (Table 1 collects the advantages of nanomedicines compared to conventional chemotherapeutics). Apart from all the advantages of the use of DDS for cancer treatment, polymeric DDS are notable for clinical translation, due to the biocompatibility of the raw materials and the easy modulation to improve efficacy.

**Table 1.** Advantages of nanomedicine compared to conventional chemotherapeutics.

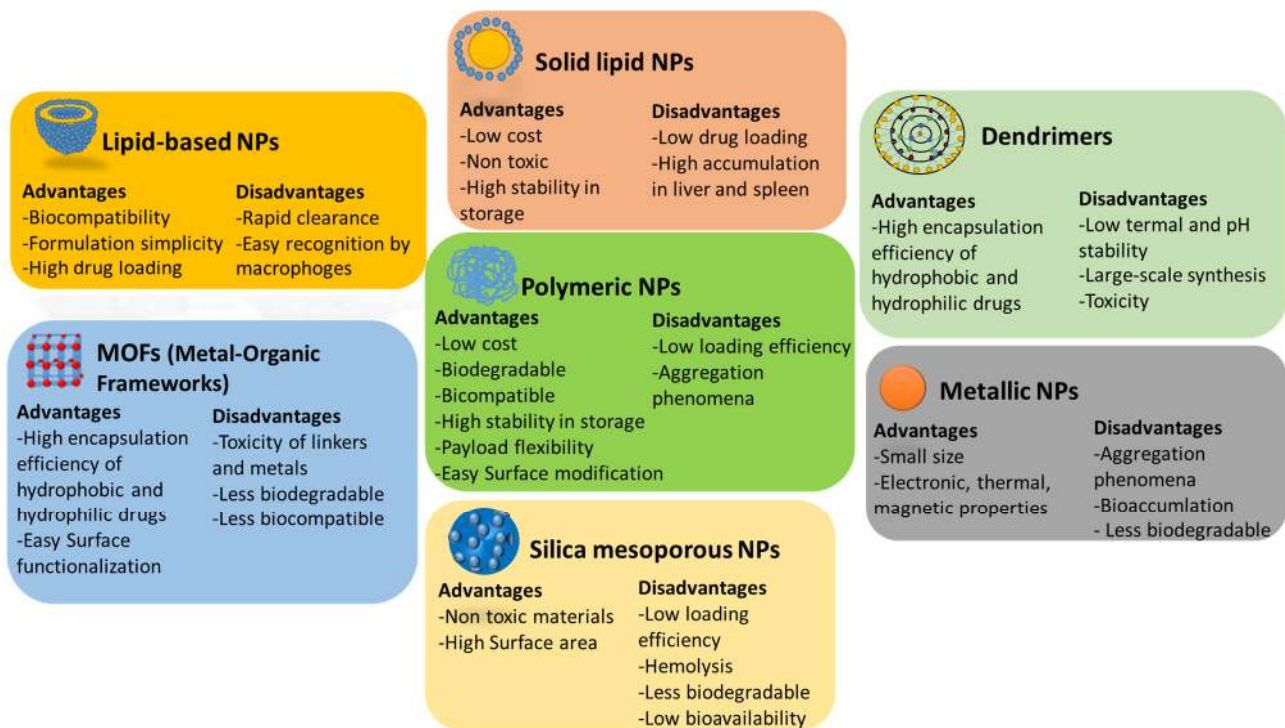
Advantages of Nanomedicine
Improved bioavailability
Greater dose response
Enhanced solubility
Scope for improvement efficacy
Assess therapeutic efficacy in real time: simultaneous monitoring of drug release and imaging distribution
Reduced toxicity
Allow to design specifically targeted therapies
Possibility of designing smart nanosystems capable of responding to external stimuli

Nanomedicine can be divided into the following three main areas depending on its application: nanodiagnosis, regenerative medicine, and nanotherapy [12]. The main aim of nanodiagnosis is the early detection of diseases by the use of nanomaterials [13–15]. The *in vivo* diagnosis consists of the administration of different nanodevices for the quantification of several parameters, compounds or metabolites in the organism, while *in vitro* diagnosis achieves disease detection through samples obtained from patients. One of the principal nanomaterials for nanodiagnosis is the nanobiosensor, which can detect a number of compounds in real time [16].

On the other hand, regenerative medicine consists of the repair or substitution of damaged tissues and organs by nanomaterials [12]. The most commonly used nanomaterials employed for this purpose are based on carbon nanotubes, hydroxyapatite nanodevices, nanocomposites, and biodegradable polymers [17].

Unfortunately, therapeutic agents are not free of side effects and contraindications. In fact, there is a significant proportion of patients who experience adverse effects with the current therapies. Some therapeutics are very quickly metabolized and require high doses to be effective. On the other hand, minor, but frequent, side effects produced mean that many patients report low levels of adherence to treatments [18]. Chemical modifications of approved therapeutics, to improve their pharmacokinetics and safety profile, are costly. In this context, nanotherapy seems to provide solutions by therapeutic encapsulation in controlled-release systems. NPs of 100–400 nm diameter can accumulate within the tumor, through the EPR effect [19]. They can deliver high concentrations of the therapeutic to the target site by convection and diffusion processes, which can also reduce the effects on the surrounding tissues [20].

Figure 2 displays the advantages and disadvantages of the most important DDS (structure, benefits and intended use of the different NPs are described in Table 2). Lipid-based NPs are simply formulated and are able to carry large payloads [21], but they are rapidly retained by the reticuloendothelial system, and modifications to extend their half-time circulation are requested for clinical use. Solid lipid NPs are particularly important in genetic therapy, due to their efficacy for nucleic acid delivery [22]. However, the low therapeutic loading, and accumulation in the liver and spleen limit their options for clinical development [21]. Inorganic NPs possess additional magnetic, electrical, and optical properties for application, such as diagnostics, imaging and photothermal therapies [23]. Nevertheless, their clinical application is limited by their low solubility and toxicity [24]. Amongst the many materials used in nanomedicine, with promising properties as therapeutic carriers, the following one stands out: biodegradable and biocompatible polymers [25,26]. Polymeric NPs are ideal candidates for drug delivery, given the versatility of the raw materials used for their production. These NPs are also stable during storage, and large-scale production is feasible. Furthermore, they show great potential for easy surface modifications to optimize pharmacokinetics, including the half-life in circulation and tissue delivery, and, by modulating the polymer structure, loading and release kinetics can be controlled [27–29]. It is worth noting other advantages of polymeric NPs, such as the ease of customized surfaces to be constructed to recognize target proteins and cells [30], along with the generation of stimuli-responsive nanodevices [31,32]. To date, many polymeric NPs are in clinical trials (Table 3) [33].



**Figure 2.** Advantages and disadvantages of different DDS for cancer treatment. Lipid-based NPs reach high drug loading, but their half-life in blood limits their clinical translation; solid lipid NPs fails due to low drug loading and high accumulation; dendrimers are mainly toxic and difficult to scale their synthesis; metallic NPs are easily accumulated in the body and cause aggregation phenomena; MOFs and silica mesoporous NPs are not biodegradable and toxic; polymeric NPs are the most potential candidates for clinical translation due to their high biocompatibility and payload and surface modification flexibility.

**Table 2.** Structure, benefits and intended use of the most significant DDS in medicine.

Type of DDS	Structure	Benefits	Indented Use
Lipid-based NPs	Spherical vesicle having a lipid bilayer	Formulation simplicity and high drug loading	First vehicle for administration of pharmaceutical drugs
Solid lipid NPs	Spherical solid lipid core stabilized by surfactants	Temporal and in vivo stability	Delivery vehicle for nucleic acids
Dendrimers	Branched polymeric molecules	Encapsulation of hydrophobic drugs	Easy chemical modification to increase in vivo suitability
MOFs	Clusters of metal ions coordinated to organic ligands	High encapsulation and loading efficiency	pH-, magnetic-, ion-, temperature- and pressure-response carriers
Metallic NPs	Mainly iron oxide, gold, or silver core	Small size and easy surface functionalization	Development of diagnostic and therapeutic agents
Silica mesoporous NPs	Mesoporous silica core	Extensive multi-functionality based on its high specific surface	Stimuli-reactive guided liberation of drugs through chemical coatings
Polymeric NPs	Biodegradable and biocompatible polymers	Low cost, large-scale synthesis, payload flexibility, biocompatibility and easy surface modification	Clinical translation of nanomedicines

**Table 3.** Polymeric NPs in clinical trials.

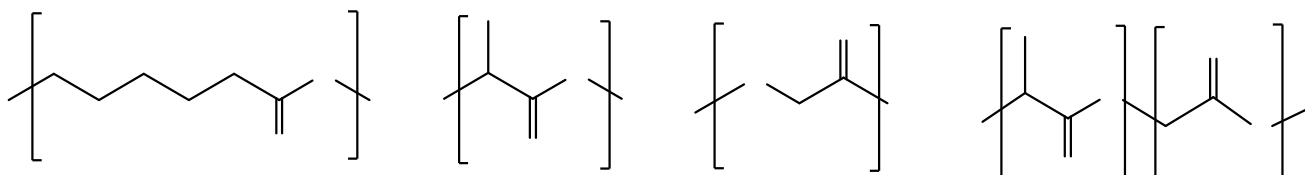
Nanomedicine	Drug	Polymer	Conditions	Reference Clinical Trials
Genexol-PM®	Paclitaxel	polymeric micelle formulation	Metastatic adenocarcinoma of the pancreas	NCT02739633
			Hepatocellular carcinoma after failure of sorafenib	NCT03008512
			Advanced urothelial cancer	NCT01426126
			Advanced non-small-cell lung cancer	NCT01770795
			Advanced, metastatic and recurrent breast cancer	NCT01784120 NCT00876486 NCT01169870 NCT00912639 NCT02263495 NCT02064829
			Gynecologic cancer	NCT02739529
			Advanced ovarian cancer	NCT00877253 NCT01276548 NCT00886717
			Advanced and metastatic pancreatic cancer	NCT00882973 NCT00111904
			Advanced non-small-cell lung cancer	NCT01023347
			Advanced head and neck cancer	NCT01689194
			Advanced esophageal squamous cell carcinoma	NCT01474642 NCT00816634
PICN®	Paclitaxel	copolymer poly lactide-polyethylene glycol	Metastatic breast cancer	CTRI/2010/091/001116
BIND-014®	Docetaxel	copolymer poly lactide-polyethylene glycol	Metastatic castration-resistant prostate cancer	NCT01812746
			Non-small-cell lung cancer	NCT01792479
			Advanced or metastatic cancer	NCT01300533
			KRAS positive or squamous cell non-small-cell lung cancer	NCT02283320
			Urothelial carcinoma, cholangiocarcinoma, cervical cancer and squamous cell carcinoma of the head and neck	NCT02479178
Livatag®	Doxorubicin	Polyalkylcyanoacrylate	Advanced hepatocarcinoma	EudraCT-2006-004088-77
CALAA-01®	siRNA	Adamantane polyethylene glycol containing cyclodextrin	Solid tumor cancers	NCT00689065

## 2. From Raw Materials to Polymeric NPs

Polyesters are the most used raw materials for polymeric DDS generation. Ideally, the polymers selected must be biocompatible and biodegradable, and therefore the existence of ester bonds in the macrostructure make these devices easily broken in biological environments. Non-synthetic biodegradable polymers, such as alginate, chitosan, and albumin, have been used to prepare polymeric NPs. Despite their biocompatibility, biodegradability and non-toxic properties, these raw materials present limitations to clinical translation, due to the high variability in batch productions and high immunogenicity of some natural polymers. In addition, their degradation strictly depends on intrinsic properties. On the other hand, synthetic polymers can be designed to modulate delivery parameters such as loading efficiencies, therapeutic release kinetics, surface charge, stability, responsivity, and size and polydispersities of the polymeric NPs. In this sense, polylactide (PLA), polyglycol-



ide (PGA), polycaprolactone (PCL) and polylactide-co-glycolide (PLGA) are commercial FDA-approved polymers for DDS generation [34] (see chemical structures in Figure 3).



**Figure 3.** Chemical structures of the most widely used polymers for DDS generation. PCL is used for DDS generation in the form of copolymers with other low degradation rate polymers; PLA is an ideal environment for the generation of polymeric NPs capable of encapsulating hydrophobic therapeutics; PGA is mainly used in tissue engineering; PLGA is by far the most used for DDS generation due to the easy hydrophobic/hydrophilic ratio modulation.

### 2.1. PLA

PLA is an FDA-approved polymer, due to its biodegradability, low immunogenicity, low toxicity and high biocompatibility. PLA is degraded to lactic acid, which, in turn, is used in other metabolic routes [35]. Some studies with PLA NPs showed that lactic acid was metabolized fast, to H<sub>2</sub>O and CO<sub>2</sub>, and, therefore, was easily eliminated by the body [36]. PLA is produced by a polycondensation reaction of lactic acid, or by ring-opening polymerization (ROP) of the cyclic ester. Both raw materials are obtained by the anaerobic fermentation of organic substrates such as sugars, beet, or wheat starch [37,38]. PLA has high hydrophobicity, which is an ideal environment for the encapsulation of hydrophobic therapeutics [39]. Representative examples for the development of new cancer treatments using PLA NPs are the work carried out by Coolen et al., where PLA NPs were used for cell transfection [40], or the work reported by Feng et al., to encapsulate fisetin for breast cancer therapy [41].

### 2.2. PGA

PGA is an FDA-approved polymer, obtained by ROP of glycolide. PGA was used for the generation of the first bioresorbable suture in the seventies [42]. PGA is a biodegradable thermoplastic that produces glycolic acid after degradation, and is then excreted in urine. The low solubility in organic solvent, low stability in water, and quick enzymatic degradability limited the use of PGA for NPs formulation. Indeed, the use of PGA is focused on tissue engineering for bone, tendons, cartilage, teeth, and spinal regeneration [43].

### 2.3. PLGA

PLGA is an FDA-approved biodegradable and biocompatible copolymer, obtained by ROP of lactide and glycolide. The degradation products of PLGA are lactic acid and glycolic acid, which are innocuous for humans [44]. The physicochemical properties of the polymer depend on the PLA:PGA ratio. Due to the high degradability of PGA, the degradation of PLGA is marked by the amount of PLA in the copolymer structure [34,44]. PLGA is by far the most used for DDS generation [45]. However, as has happened for all the polymers mentioned, PLGA needs to be pegylated for enhancing its *in vivo* efficiency [46,47]. The incorporation of polyethylene glycol into the macromolecular structure allows the circulation time of the NPs to increase [48], and the bio-adhesion to different immune cell lines or different plasmatic components to decrease [49,50]. Also, PLGA is used as a raw material for the generation of molecule-targeted therapy. As an original strategy, Pan et al. reported hyaluronic acid-decorated hybrid PLGA nanoparticles as 17-allylaminogeldanamycin delivery carriers for targeted colon cancer therapy. *In vivo* studies showed much better therapeutic efficiency than the free therapeutic [51].

## 2.4. PCL

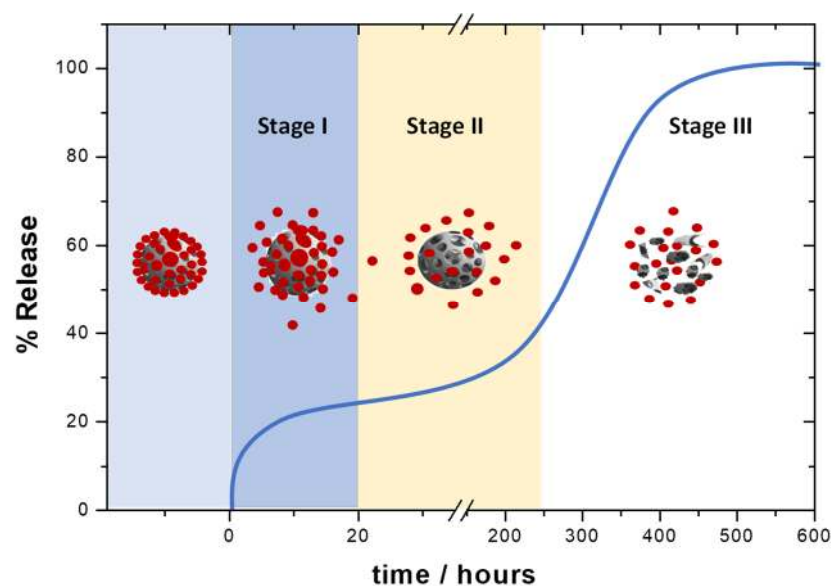
PCL is a biodegradable and biocompatible FDA-approved polymer, obtained from fossil resources [52]. This polyester is obtained by ROP of  $\epsilon$ -caprolactone. It is soluble in a wide range of organic solvents and presents slow degradation rates (2–3 years). Once again, its use is focused on tissue engineering [53]. However, it has also been used as a raw material for DDS generation, in the form of copolymers with other low degradation rate polymers, such as PLGA or PEG [54,55]. There are some successful examples of PCL formulations for the treatment of cancer [56–58].

## 2.5. Other Synthetic Polymers

Poly(anhydrides) [59], poly(orthoesters) [60], poly(amides) [61], poly(phosphoesters) [62], and poly(alkylcyanoacrylates) [63] are examples of other polymers used for the generation of DDS. In many cases, successful devices have been formulated, such as the one reported by Fusser et al., using poly(2-ethylbutyl cyanoacrylate) to encapsulate cabazitaxel for breast cancer treatment [64], or the poly(ester amides) NPs reported by Villamagna et al. to encapsulate Celecoxib [65].

## 3. Mechanism of Action of Polymeric NPs

Three mechanisms govern the release of the therapeutic load from polymeric NPs. Burst release, diffusion through the polymer matrix, and erosion are the mechanisms proposed for the release of therapeutics. The extension of each one will depend on the therapeutic structure, raw material employed, NP morphology, and release media [66]. A triphasic profile is expected for the release of the therapeutic from polymeric NPs (see example of profile in Figure 4). Note that mechanisms can act simultaneously.



**Figure 4.** Representative release profile for polymeric NPs. In stage I the drug is quickly released through desorption–diffusion processes. The release of the drug in stage II is governed by diffusion through the polymer matrix or pores. Stage III is governed by the erosion of the polymeric matrix.

### 3.1. Stage I

This stage is governed by the burst release mechanism. The therapeutics physically absorbed over the surface of the NPs are released through desorption–diffusion processes. This stage is characterized by very short times.

### 3.2. Stage II

In this case is the diffusion of the therapeutic through the polymer matrix or pores, which explains the release phenomena. This stage is much slower and can coexist with

polymer erosion [34,66]. The diffusion relies on the intermolecular therapeutic/polymer forces, as well as the permeability and thickness of the nanodevices [67]. Stage I and II follow the Higuchi pattern, a mathematical model to describe the release of water-soluble and low-soluble drugs incorporated into semisolid and solid matrices. Thus, the simplified Higuchi model describes the release of therapeutics as a square root of a time-dependent process ( $t^{1/2}$ ), based on a pseudo-steady state approach that matches the Fick's 1st law (Fickian release) for an ideal steady-state in stage II [68].

### 3.3. Stage III

The erosion mechanism governs this stage and will depend on the physico-chemical properties of the raw materials [66]. Usually, the release is very slow. Nevertheless, those polymers easily hydrolyzed show this phase in shorter times. In this case, the release kinetic is unpredictable [69]. The Korsmeyer and Peppas model is based on an empirical equation, to include both the Fickian and non-Fickian release of the drug from the polymeric matrix. This describes the release as a time-dependent  $t^n$  process, where  $n$  is the release exponent, indicative of the mechanism of drug transport through the polymeric matrix, allowing the erosion process to be included in the release mechanism in stage III [68].

Biphasic profiles and constant release profiles have been also reported in a few cases [34].

### 3.4. The Release of the Therapeutic Depends on Many Factors

Cellular and intracellular barriers, crystallinity and degradability of the polymer, the hydrophilic/hydrophobic ratio between the therapeutic and polymer, and the type of surfactants are key variables for the delivery.

#### 3.4.1. Cellular and Intracellular Barriers

Cells and intracellular membranes vary widely, as well as lipid rafts and transmembrane proteins. Therefore, membranes are heterogeneous within a patient or across a patient population, which creates diverse barriers to NPs delivery [70]. In addition, polymeric NPs can contact biomolecules such as serum proteins and lipids, to form a corona over their surface. This fact can dictate the uptake of the polymeric NPs, even their stability and cargo [71].

#### 3.4.2. Crystallinity

Polymer crystallinity is defined as the rate of ordered regions inside the polymer matrix in relation with the disordered regions. The orientation of the polymer chains into the NPs can influence the physicochemical properties of the nanodevices. Thus, ordered macrostructures will hamper the water entrance, slowing down the erosion. It is mandatory to calculate the glass transition temperature to know about the crystallinity rate of a polymer ( $T_g$ ). The  $T_g$  is the temperature in which the polymer changes from a rigid state to a more flexible state, and it is calculated by dynamic scanning calorimetry (DSC) [39]. In general terms, the therapeutic release decreases with the increasing  $T_g$ , and the crystallinity is directly related to the molecular weight of the polymer [44]. Polymers with low molecular weights are less crystalline and degrade easily, which causes faster therapeutic release rates [34].

#### 3.4.3. Hydrophilic/Hydrophobic Ratio

This ratio is a limiting factor to achieving high encapsulation efficiencies. Polymers are hydrophobic in nature and, therefore, hydrophobic therapeutics are suited to being entrapped in polymeric NPs. Those polymers with high solubility in water will give rise to DDS with fast therapeutic release profiles [72].

#### 3.4.4. Degradability

Polymer degradation mainly depends on the lability of its chemical bonds, and the crystallinity and hydrophobicity. Polymers with high crystallinity and high hydrophobicity



present low degradation rates and low susceptibility to hydrolysis. In the same way, ester and amide bonds facilitate the DDS degradation for enzyme action [66].

#### 3.4.5. Type of Surfactants

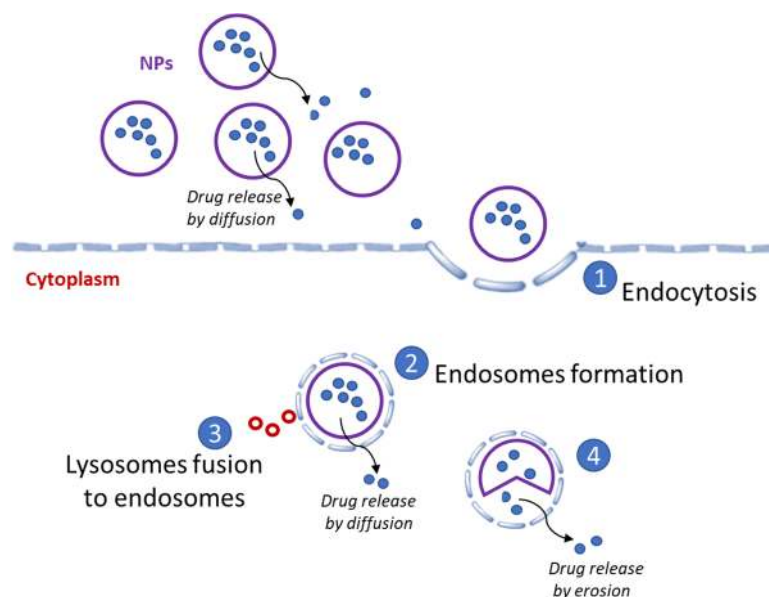
Surfactants are humectant, and are capable of reducing the superficial tension of liquids and the interfacial tension between liquids. Each surfactant has a hydrophilic grade, which is a determinant for the generation of NPs [73]. Surfactants can be divided into anionic, cationic and zwitterionic, and non-ionic surfactants. Tweens<sup>®</sup>, spans<sup>®</sup>, pluronic<sup>®</sup> and poly(vinyl alcohol) are the most common surfactants employed in formulation. The surfactants are determined to modulate the size, shape, and surface of polymeric NPs. In general terms, non-ionic surfactants are less toxic. Aggregation phenomena can be observed if the surfactant is not appropriate, impacting the size and polydispersities of the formulation [74]. Also, the release of the therapeutic can be modified [75]. Gagliardi et al. recently discussed the role of surfactants for the construction of polymeric NPs [25].

#### 3.5. Systemic Administration of Polymeric NPs Is Common for Cancer Treatment

In terms of size, NPs of less than 10 nm are rapidly eliminated by the kidneys, and those larger than 20 nm stimulate recognition by macrophages or other cells of the immune system [76]. Thus, PEG is usually incorporated into the polymeric NPs to improve their circulation time. Another option is the surface modification by conjugation techniques. Many polymeric NPs have successfully added moieties, such as antibodies, glucose, or proteins, to direct their delivery [77,78].

In terms of surface charge, neutral and slightly negative polymeric NPs have a longer circulation time. Anionic polymeric NPs have to overcome repulsive forces to make contact with the cell surface, and, on the contrary, the attractive forces suffered by cationic polymeric NPs can damage the cells and cause toxicity [79].

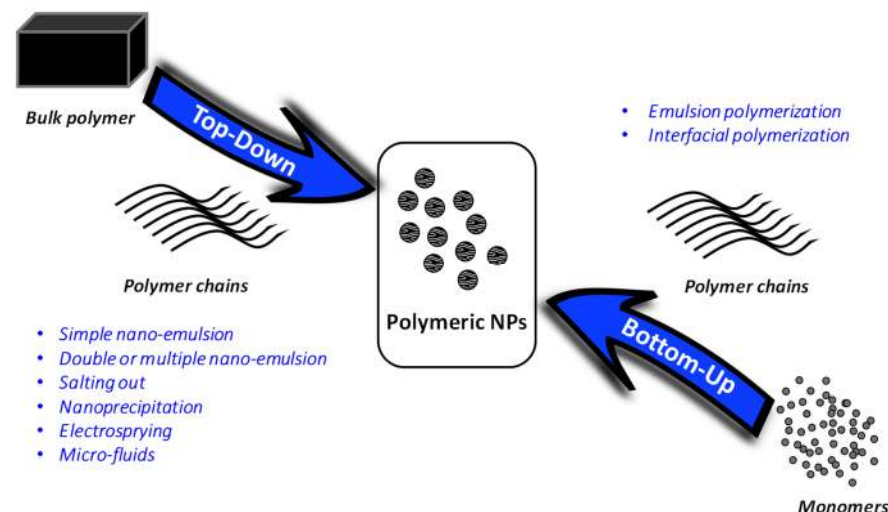
The mechanism of action of polymeric NPs is based on the EPR effect. NPs in the range of 100–400 nm have been widely reported to accumulate at the tumor site via the EPR effect [80]. This favors high accumulation of the therapeutic, facilitating its delivery to the site of interest by convection and diffusion processes [81]. However, works suggest potential limitations, because the EPR effect can differ among patients and types of tumors [82,83]. The internalization of the NPs into the cell is produced through endocytosis [82], and results in early endosome formation. The type of endocytosis is determined by cell type and size of the NPs. It should be noted that the endocytosis process can cause changes in the stability of the NPs and their cargo. Figure 5 illustrates the mechanism of action. Endosomes couple to lysosomes that cleave the NPs, which subsequently release the free cytotoxic therapeutics into the cytoplasm, interfering with the cellular mechanisms, and ultimately promoting cell death [84].



**Figure 5.** The mechanism of action of polymeric NPs consists of the internalization of the NPs into the cell that is produced through endocytosis, producing changes in the stability and cargo of the NPs. The following step is the cleavage of the NPs by endosome formation and lysosome fusion to release the drug by erosion of the NPs.

**4. Methods to Formulate Polymeric NPs**

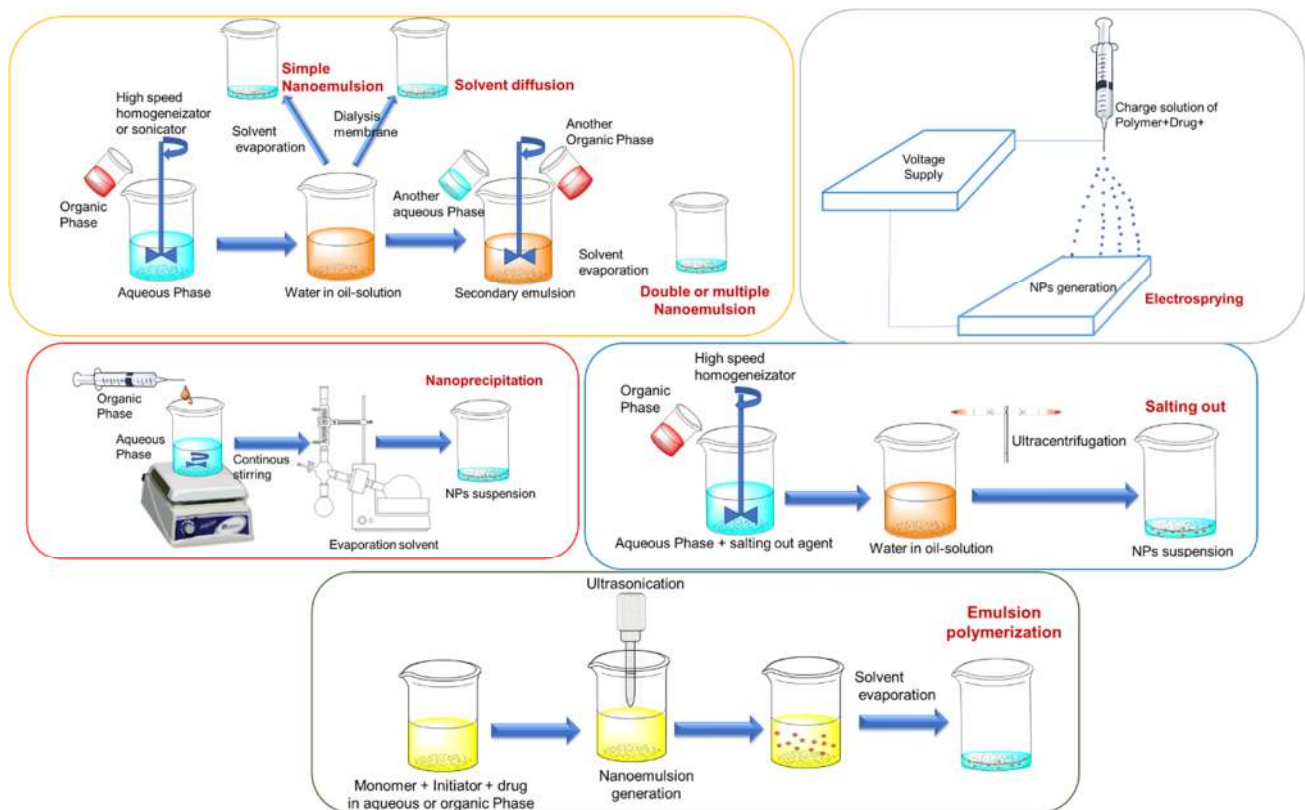
There are several methods to formulate polymeric NPs. The methods can be broken down in two main strategies, top-down and bottom-up methodologies (Figure 6).



**Figure 6.** The methods to formulate polymeric NPs are divided into top-down and bottom-up methodologies. Top-down methodologies generate the NPs from preformed polymers, whereas in bottom-up methodologies the polymerization of the monomers is achieved during formulation.

In top-down methodologies, the NPs are obtained from preformed polymers; meanwhile, in bottom-up methodologies, the polymerization of the monomers is achieved during formulation [85,86]. The nanoprecipitation and displacement solvent method, several techniques of emulsification and evaporation, solvent diffusion, dialysis methods, salting-out, electro-static spraying and micro-fluids are the most important ones in the case of top-down methodologies. Bottom-up strategies have not been widely explored, but, among them, emulsion polymerization, interfacial polymerization, interfacial polycondensation and the coacervation approach are the most used [86]. The following is a more

detailed explanation of the most widely used methods for the generation of polymeric NPs (see illustrations in Figure 7).



**Figure 7.** Illustration of the formulation of polymeric NPs by the most significant methods. Emulsion polymerization is a bottom-up methodology, whereas nanoprecipitation, simple nano-emulsion, solvent diffusion, double or multiple nano-emulsion, and salting out are top-down methodologies.

#### 4.1. Simple Nano-Emulsion (Top-Down)

In this approach, the therapeutic and polymer are solubilized in immiscible organic solvents, such as ethyl acetate or dichloromethane, within the aqueous phase containing surfactants [87]. The phases are emulsified with the help of a high-speed homogenizer or sonicator. Once the nano-emulsion is stabilized, the solvent is removed. This methodology is characterized to give rise to large particle sizes [88].

#### 4.2. Double or Multiple Nano-Emulsion (Top-Down)

It was designed to encapsulate hydrophilic therapeutics and proteins. This approach consists of the formulation of two nano-emulsions, once a simple nano-emulsion preparation is added to an external aqueous phase, and again emulsified to obtain the double nano-emulsion. NPs are formed when the organic solvent is removed. This approach was designed in order to attain higher encapsulation efficiencies for hydrophilic therapeutics [89].

#### 4.3. Salting Out (Top-Down)

This approach is a modified formulation of nano-emulsion in which the mixture to be emulsified contains a polymer, a therapeutic, surfactants, and salting-out agents. The common choices of salting-out agents are magnesium chloride, calcium chloride or sucrose. Fast mechanical stirring is used to emulsify, and the solvent is removed via reduced pressure. The mixture needs ultracentrifugation and repeated washing to eliminate the

salting-out agents and surfactants [90]. The main disadvantage of this methodology is that the salting out agents are, in many cases, incompatible with therapeutics [91].

#### 4.4. Nanoprecipitation and Displacement Solvent Method (Top-Down)

The polymer and therapeutic are solubilized in miscible organic solvents, and then the mixture is added in a controlled manner over an aqueous solution during continuous stirring [92]. During nanoprecipitation, NPs are formed instantly and the therapeutic is entrapped in the polymer matrix. In this case, the solvent is removed by reduced pressure [28]. The formation of NPs is governed by the Gibbs-Marangoni effect, which describes a mass transfer in an interphase between two fluids, due to a gradient of superficial tension [93]. The nanoprecipitation method prevents the loss of therapeutics during the emulsification process.

#### 4.5. Electrospraying (Top-Down)

The basic principles of this approach are based on the application of electrostatic charges to manufacture the NPs. For this approach, a charged solution where the therapeutic and polymer are dissolved is used, and the concentration, caudal, voltage and other parameters are adjusted to generate little drops with different defined shapes and sizes in the matrix solution. This technique achieves very high therapeutic loading efficiency with a low polydispersity index [94].

#### 4.6. Microfluids (Top-Down)

The microfluid devices are designed to manipulate fluids in microscale channels. Obtaining NPs in microfluid systems is carried out by microdevices with internal dimensions of less than 1 mm [95,96].

#### 4.7. Emulsion Polymerization (Bottom-Up)

Emulsion polymerization is the fastest scale-up method to manufacture polymeric NPs. There are two types [97], emulsion polymerization with a continuous organic phase, which consists of the dispersion of the monomer into an emulsion, and emulsion polymerization, with a continuous aqueous phase in which the monomer is dissolved in an aqueous solution without surfactants. The former is less used because of the use of toxic solvents, surfactants, and initiators, which are difficult to be removed [97].

#### 4.8. Interfacial Polymerization (Bottom-Up)

In this case, the mixture of the therapeutic, monomers and initiator are extruded through a needle over an aqueous solution within a surfactant. During the process, NPs are spontaneously formed by monomer polymerization. Later, the solvent is removed, and the NPs are obtained. The advantage of this approach is the high encapsulation efficiency in the one-step formulation. However, the organic solvent is very difficult to remove [97].

### 5. Polymeric NPs in Clinical Investigations

There are more than 15 nanomedicines on the market for cancer treatment [98]. Concerning the polymeric NPs (see Table 3), PICN<sup>®</sup> is a polymeric formulation of paclitaxel that is approved in India for metastatic breast cancer [99]. The non-targeted PICN<sup>®</sup> is currently in clinical trials in the USA [98]; Genexol<sup>®</sup>, produced by Samyang Biopharm, is a polymeric micelle formulation of paclitaxel that is clinically approved to treat breast cancer in South Korea [100–102]. BIND-014<sup>®</sup> is composed of a copolymer PLA–PEG and is used for the controlled release of docetaxel against prostate cancer [103,104]. In this particular case, the small-molecule S,S-2-[3-[5-amino-1-carboxypentyl]-ureido]-pentanedioic acid was used to guide the NPs. Preclinical studies showed different pharmacokinetic properties than those reported with sb-docetaxel. Negative phase II clinical trials were reported, due to their little activity [105]. Livatag<sup>®</sup> is a polymeric NPs formulation of doxorubicin for the treatment of primary liver cancer. Poly (alkyl cyanoacrylate) and cyclodextrin are

the raw materials for the generation of this formulation. Initial studies reported higher activities and a very low toxicity in the heart compared to free doxorubicin. However, several adverse pulmonary events led to the termination of the clinical development of Livatag<sup>®</sup> [106]. CALAA-01<sup>®</sup> are targeted polymeric NPs generated for siRNA-mediated treatment of solid tumors [107]. They were constructed by the self-assembly of a mixture of cyclodextrin-containing polymer backbone, adamantane-conjugated PEG, and transferrin-conjugated PEG and siRNA. Increased inhibition of tumor growth in mice by CALAA-01<sup>®</sup> was demonstrated in a mouse model of metastatic Ewing's sarcoma. It was the first experimental interfering RNA therapeutic agent to be administered in cancer patients [108]. Early results in patients with solid tumors showed dose-dependent intracellular localization in tumor cells. Phase II clinical studies are completed, but no data has been released yet.

The identification of genomic alterations, such as gene amplifications or mutations, in cancers has permitted the design of chemical entities against those alterations. This approach has been shown to be effective, with a wide range of compounds reaching the clinical setting. However, many of those have failed in their clinical development due to an inadequate toxicity profile [109]. The toxic effect of the therapeutic agent against a non-transformed tissue that also expressed the target protein has clearly had a constant limitation: this concept is called on-target off-tumor toxicity [110]. A classic example has been the cardiotoxicity observed with anthracyclines, which is a type of chemotherapy that is widely used for the treatment of many solid tumors. Similar findings can be described for targeted agents, such as the kinase inhibitor against HER2 neratinib that shows an inadequate toxicity profile in relation with diarrhea. In both examples, long-term exposure requires reductions, or even limitations, in their use. Another example is the mucositis and glucose deregulation observed with everolimus, which produces treatment discontinuations [111]. In this context, it is expected that strategies targeting pan-essential genes will be toxic, having an inverse therapeutic index [112,113]. Nano-vectorization is a potential approach to improve their delivery and reduce their toxicity. Another approach will be the identification of smart drug combinations, but, again, both agents could be loaded in the same nanoparticle [114].

In addition, the pharmacokinetic (PK) profile can influence the toxicity and particularly when the toxicity itself is not reversible [109]. To resolve this problem, encapsulation of compounds to improve their PK profile, limiting their exposure to non-transformed tissue, is a main area of research. The encapsulation of PROTACs is an example of success [115], but there is still a long way to go, which requires safety and efficacy experiments in different animal models. Novel methods for the encapsulation of targeted agents, such as small chemical entities, are under evaluation.

## 6. Guided and Smart NPs

Polymeric NPs made of polymers that are naturally pharmaceutically active, or polymers conjugated with therapeutics, or with biomacromolecules such as proteins or antibodies, are being designed to enhance delivery and optimize precision medicine therapies [116]. Therefore, polymeric NPs with targeting moieties, such as antibodies, transferrin or folate, use ligand–receptor, enzyme–substrate or antibody–antigen interactions to improve NP uptake and distribution. Such is the case of the transferrin-targeted polymeric NPs formulated for the codelivery of tarquidar and paclitaxel. These polymeric NPs enhanced cytotoxicity in ovarian cancer cell lines and cancer spheroid cultures [117]. Another example is the conjugation of doxorubicin to PLGA, to form polymeric NPs. The surface of the polymeric NPs was decorated with a hepatocellular carcinoma-specific peptide moiety, to mitigate toxicity in animals and enhance the efficacy of doxorubicin against hepatocellular carcinoma [118]. Another representative example is the nutlin-3-loaded PLGA NPs conjugated with rituximab to successfully target CD20-positive leukemic cancer cells [119], or the paclitaxel-loaded PLGA/montmorillonite NPs conjugated with trastuzumab antibodies, formulated to reduce the side effects of paclitaxel [120]. To exploit this approach, the



identification of biomarkers on the surface of tumoral cells is mandatory to avoid on-target non-tumoral toxicity.

On the other hand, smart polymeric NPs can activate depending on the tumor microenvironment or a specific biological stimuli [121]. Some pH-sensitive, visible-light- and temperature-responsive polymeric NPs have been reported to provide successful targeted delivery. A representative example of pH-responsive drug delivery is the ultra-pH-responsive polymers, which are designed for the generation of polymeric NPs for the in vivo delivery of RNAi therapeutics [122]. Interestingly, nanomedicines loaded with paclitaxel and siRNA's targeting antiapoptotic genes were reported for hepatocellular carcinoma treatment. By the use of low-frequency ultrasound on the tumor site, an enhanced antitumoral activity was observed in vivo [123]. It is worth highlighting that recent technological advances tend to archive multifunctional polymeric NPs that allow the simultaneous accommodation of therapeutic and imaging agents, which, together with the correct choice of the appropriate functionalized smart polymer, facilitate the construction of hybrid multi-modal NPs for theranostic, simultaneous monitoring of drug release and imaging distribution, and to assess therapeutic efficacy in real time [124]. This is the case of the semiconducting polymer nano-PROTACS, which was recently reported for activable photo-immunometabolic cancer therapy [125].

## 7. Conclusions

Over the last decade, polymeric NPs have been designed to overcome the limitations of free therapeutics for the treatment of cancer. Polymeric NPs have shown a more favorable pharmacokinetic profile than the free chemotherapeutics, but optimization of the formulation, in terms of the polydispersity and size of the NPs, is still needed to improve efficacy. In the same way, drug release from polymeric NPs can be more precisely controlled, with a range of polymers designed specifically for that purpose. In this sense, therapeutic polymers emerge as polymers with pharmaceutical and biomedical applications, and with a promising future in cancer research. On the other hand, selective-targeting tumoral cells can augment the permeability and penetration of the polymeric NPs within the tumor, and, therefore, diminish toxicity and avoid the adverse effects of prolonged treatment. For this purpose, guided polymeric NPs may provide benefit over NPs when targeting receptors that also have a biological effect.

In conclusion, although great advances have been performed, there is still a long way to go, and combined efforts from the scientific and biotechnology community are needed in order to speed up the development of this technology.

**Author Contributions:** Conceptualization, C.A.-M. and I.B.; investigation, E.N.; writing—original draft preparation, E.N. and J.A.C.-O.; writing—review and editing, C.A.-M., A.O. and I.B.; supervision, C.A.-M. and I.B.; funding acquisition, A.O. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Ministerio de Ciencia e Innovación, Spain, grant number CTQ2017-84131-R and CTQ2016-81797-REDC Programa Redes Consolider, Instituto de Salud Carlos III, grant number PI16/01121, and under the agreement UCLM-AFANION and UCLM-ACEPAIN to support research activity on the campus of the UCLM Albacete.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Binnig, G.; Quate, C.F.; Gerber, C. Atomic Force Microscope. *Phys. Rev. Lett.* **1986**, *56*, 930–933. [[CrossRef](#)] [[PubMed](#)]
2. Drexler, K.; Peterson, C.L.; Pergamit, G. *Unbounding the Future: The Nanotechnology Revolution*; William Morrow and Company, Inc.: New York, NY, USA, 1991.
3. Freitas, R.A. What Is Nanomedicine? *Nanomed. Nanotechnol. Biol. Med.* **2005**, *1*, 2–9. [[CrossRef](#)] [[PubMed](#)]
4. Kreuter, J. Nanoparticles—A historical perspective. *Int. J. Pharm.* **2007**, *331*, 1–10. [[CrossRef](#)]
5. Kulkarni, R.K.; Moore, E.G.; Hegyeli, A.F.; Leonard, F. Biodegradable Poly(Lactic Acid) Polymers. *J. Biomed. Mater. Res.* **1971**, *5*, 169–181. [[CrossRef](#)] [[PubMed](#)]



6. Köhler, G.; Milstein, C. Continuous Cultures of Fused Cells Secreting Antibody of Predefined Specificity. *Nature* **1975**, *256*, 495–497. [[CrossRef](#)]
7. Abbasi, E.; Aval, S.F.; Akbarzadeh, A.; Milani, M.; Nasrabadi, H.T.; Joo, S.W.; Hanifehpour, Y.; Nejati-Koshki, K.; Pashaei-Asl, R. Dendrimers: Synthesis, Applications, and Properties. *Nanoscale Res. Lett.* **2014**, *9*, 247. [[CrossRef](#)] [[PubMed](#)]
8. Santini, J.T.; Cima, M.J.; Langer, R. A Controlled-Release Microchip. *Nature* **1999**, *397*, 335–338. [[CrossRef](#)] [[PubMed](#)]
9. Langer, R.; Vacanti, J.P. Tissue Engineering. *Science* **1993**, *260*, 920–926. [[CrossRef](#)]
10. Barenholz, Y. Doxil<sup>®</sup>—The First FDA-Approved Nano-Drug: Lessons Learned. *J. Control. Release* **2012**, *160*, 117–134. [[CrossRef](#)] [[PubMed](#)]
11. Zingg, R.; Fischer, M. The Consolidation of Nanomedicine. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* **2019**, *11*, e1569. [[CrossRef](#)]
12. Shi, J.; Votruba, A.R.; Farokhzad, O.C.; Langer, R. Nanotechnology in Drug Delivery and Tissue Engineering: From Discovery to Applications. *Nano Lett.* **2010**, *10*, 3223–3230. [[CrossRef](#)]
13. Jain, K.K. Nanodiagnosics: Application of Nanotechnology in Molecular Diagnostics. *Expert Rev. Mol. Diagn.* **2003**, *3*, 153–161. [[CrossRef](#)]
14. Bonam, S.R.; Kotla, N.G.; Bohara, R.A.; Rochev, Y.; Webster, T.J.; Bayry, J. Potential Immuno-Nanomedicine Strategies to Fight COVID-19 like Pulmonary Infections. *Nano Today* **2021**, *36*, 101051. [[CrossRef](#)]
15. Rana, M.M. Polymer-Based Nano-Therapies to Combat COVID-19 Related Respiratory Injury: Progress, Prospects, and Challenges. *J. Biomater. Sci. Polym. Ed.* **2021**, *32*, 1219–1249. [[CrossRef](#)]
16. Windmiller, J.R.; Wang, J. Wearable Electrochemical Sensors and Biosensors: A Review. *Electroanalysis* **2013**, *25*, 29–46. [[CrossRef](#)]
17. Madurantakam, P.A.; Cost, C.P.; Simpson, D.G.; Bowlin, G.L. Science of Nanofibrous Scaffold Fabrication: Strategies for next Generation Tissue-Engineering Scaffolds. *Future Med.* **2009**, *4*, 193–206. [[CrossRef](#)] [[PubMed](#)]
18. Seruga, B.; Ocana, A.; Tannock, I.F. *Drug Resistance in Metastatic Castration-Resistant Prostate Cancer*; Nature Publishing Group: Berlin, Germany, 2011; Volume 8.
19. Scheetz, L.; Park, K.S.; Li, Q.; Lowenstein, P.R.; Castro, M.G.; Schwendeman, A.; Moon, J.J. Engineering Patient-Specific Cancer Immunotherapies. *Nat. Biomed. Eng.* **2019**, *3*, 768–782. [[CrossRef](#)]
20. Brannon-Peppas, L.; Blanchette, J.O. Nanoparticle and Targeted Systems for Cancer Therapy. *Adv. Drug Deliv. Rev.* **2012**, *64*, 206–212. [[CrossRef](#)]
21. Fenton, O.S.; Olafson, K.N.; Pillai, P.S.; Mitchell, M.J.; Langer, R. Advances in Biomaterials for Drug Delivery. *Adv. Mater.* **2018**, *30*, 1705328. [[CrossRef](#)]
22. Cheng, Q.; Wei, T.; Farbiak, L.; Johnson, L.T.; Dilliard, S.A.; Siegwart, D.J. Selective Organ Targeting (SORT) Nanoparticles for Tissue-Specific mRNA Delivery and CRISPR–Cas Gene Editing. *Nat. Nanotechnol.* **2020**, *15*, 313–320. [[CrossRef](#)] [[PubMed](#)]
23. Huang, H.-C.; Barua, S.; Sharma, G.; Dey, S.K.; Rege, K. Inorganic Nanoparticles for Cancer Imaging and Therapy. *J. Control. Release* **2011**, *155*, 344–357. [[CrossRef](#)]
24. Aguado, B.A.; Grim, J.C.; Rosales, A.M.; Watson-Capps, J.J.; Anseth, K.S. Engineering Precision Biomaterials for Personalized Medicine. *Sci. Transl. Med.* **2018**, *10*. [[CrossRef](#)]
25. Gagliardi, A.; Giuliano, E.; Venkateswararao, E.; Fresta, M.; Bulotta, S.; Awasthi, V.; Cosco, D. Biodegradable Polymeric Nanoparticles for Drug Delivery to Solid Tumors. *Front. Pharm.* **2021**, *12*. [[CrossRef](#)] [[PubMed](#)]
26. Sun, L.; Wu, Q.; Peng, F.; Liu, L.; Gong, C. Strategies of Polymeric Nanoparticles for Enhanced Internalization in Cancer Therapy. *Colloids Surf. B Biointerfaces* **2015**, *135*, 56–72. [[CrossRef](#)]
27. Liu, Y.; Chen, Z.; Liu, C.; Yu, D.; Lu, Z.; Zhang, N. Gadolinium-Loaded Polymeric Nanoparticles Modified with Anti-VEGF as Multifunctional MRI Contrast Agents for the Diagnosis of Liver Cancer. *Biomaterials* **2011**, *32*, 5167–5176. [[CrossRef](#)] [[PubMed](#)]
28. Niza, E.; Nieto-Jiménez, C.; Noblejas-López, M.d.M.; Bravo, I.; Castro-Osma, J.A.; De La Cruz-Martínez, F.; Martínez de Sarasa Buchaca, M.; Posadas, I.; Canales-Vázquez, J.; Lara-Sanchez, A. Poly (Cyclohexene Phthalate) Nanoparticles for Controlled Dasatinib Delivery in Breast Cancer Therapy. *Nanomaterials* **2019**, *9*, 1208. [[CrossRef](#)]
29. Gu, F.X.; Karnik, R.; Wang, A.Z.; Alexis, F.; Levy-Nissenbaum, E.; Hong, S.; Langer, R.S.; Farokhzad, O.C. Targeted Nanoparticles for Cancer Therapy. *Nano Today* **2007**, *2*, 14–21. [[CrossRef](#)]
30. Valcourt, D.M.; Dang, M.N.; Scully, M.A.; Day, E.S. Nanoparticle-Mediated Co-Delivery of Notch-1 Antibodies and ABT-737 as a Potent Treatment Strategy for Triple-Negative Breast Cancer. *ACS Nano* **2020**, *14*, 3378–3388. [[CrossRef](#)]
31. Jia, R.; Teng, L.; Gao, L.; Su, T.; Fu, L.; Qiu, Z.; Bi, Y. Advances in Multiple Stimuli-Responsive Drug-Delivery Systems for Cancer Therapy. *Int. J. Nanomed.* **2021**, *16*, 1525–1551. [[CrossRef](#)]
32. Zhao, X.; Bai, J.; Yang, W. Stimuli-Responsive Nanocarriers for Therapeutic Applications in Cancer. *Cancer Biol. Med.* **2021**, *18*, 319–335. [[CrossRef](#)]
33. Anselmo, A.C.; Mitragotri, S. Nanoparticles in the Clinic: An Update. *Bioeng. Transl. Med.* **2019**, *4*, e10143. [[CrossRef](#)] [[PubMed](#)]
34. Kamaly, N.; Yameen, B.; Wu, J.; Farokhzad, O.C. Degradable Controlled-Release Polymers and Polymeric Nanoparticles: Mechanisms of Controlling Drug Release. *Chem. Rev.* **2016**, *16*, 2602–2663. [[CrossRef](#)] [[PubMed](#)]
35. James, R.; Manoukian, O.S.; Kumbar, S.G. Poly(Lactic Acid) for Delivery of Bioactive Macromolecules. *Adv. Drug Deliv. Rev.* **2016**, *107*, 277–288. [[CrossRef](#)] [[PubMed](#)]

36. Bazile, D.V.; Ropert, C.; Huve, P.; Verrecchia, T.; Marlard, M.; Frydman, A.; Veillard, M.; Spenlehauer, G. Body Distribution of Fully Biodegradable [<sup>14</sup>C]-Poly(Lactic Acid) Nanoparticles Coated with Albumin after Parenteral Administration to Rats. *Biomaterials* **1992**, *13*, 1093–1102. [[CrossRef](#)]
37. Nagarajan, V.; Mohanty, A.K.; Misra, M. Perspective on Polylactic Acid (PLA) Based Sustainable Materials for Durable Applications: Focus on Toughness and Heat Resistance. *ACS Sustain. Chem. Eng.* **2016**, *4*, 2899–2916. [[CrossRef](#)]
38. Avérous, L. Chapter 21—Polylactic Acid: Synthesis, Properties and Applications. In *Monomers, Polymers and Composites from Renewable Resources*; Belgacem, M.N., Gandini, A., Eds.; Elsevier: Amsterdam, The Netherlands, 2008; pp. 433–450. ISBN 978-0-08-045316-3.
39. Jenkins, A.D. *Contemporary Polymer Chemistry* (3rd Edition). HR Allcock, FW Lampe and JE Mark. Pearson Education, Inc. (Pearson/Prentice Hall): Upper Saddle River, NJ, USA, 2003, ISBN 0-13-065056-0 Pp Xviii + 814. *Polym. Int.* **2004**, *53*, 1395. [[CrossRef](#)]
40. Coolen, A.-L.; Lacroix, C.; Mercier-Gouy, P.; Delaune, E.; Monge, C.; Exposito, J.-Y.; Verrier, B. Poly(Lactic Acid) Nanoparticles and Cell-Penetrating Peptide Potentiate mRNA-Based Vaccine Expression in Dendritic Cells Triggering Their Activation. *Biomaterials* **2019**, *195*, 23–37. [[CrossRef](#)]
41. Feng, C.; Yuan, X.; Chu, K.; Zhang, H.; Ji, W.; Rui, M. Preparation and Optimization of Poly (Lactic Acid) Nanoparticles Loaded with Fisetin to Improve Anti-Cancer Therapy. *Int. J. Biol. Macromol.* **2019**, *125*, 700–710. [[CrossRef](#)]
42. A New Absorbable Suture—Frazza—1971—Journal of Biomedical Materials Research—Wiley Online Library. Available online: <https://onlinelibrary.wiley.com/doi/abs/10.1002/jbm.820050207> (accessed on 28 May 2021).
43. Tang, X.; Thankappan, S.K.; Lee, P.; Fard, S.E.; Harmon, M.D.; Tran, K.; Yu, X. Chapter 21—Polymeric Biomaterials in Tissue Engineering and Regenerative Medicine. In *Natural and Synthetic Biomedical Polymers*; Kumbar, S.G., Laurencin, C.T., Deng, M., Eds.; Elsevier: Oxford, UK, 2014; pp. 351–371. ISBN 978-0-12-396983-5.
44. Xu, Y.; Kim, C.-S.; Saylor, D.M.; Koo, D. Polymer Degradation and Drug Delivery in PLGA-Based Drug-Polymer Applications: A Review of Experiments and Theories. *J. Biomed. Mater. Res. B Appl. Biomater.* **2017**, *105*, 1692–1716. [[CrossRef](#)]
45. Kapoor, D.N.; Bhatia, A.; Kaur, R.; Sharma, R.; Kaur, G.; Dhawan, S. PLGA: A Unique Polymer for Drug Delivery. *Ther. Deliv.* **2015**, *6*, 41–58. [[CrossRef](#)]
46. Song, Z.; Feng, R.; Sun, M.; Guo, C.; Gao, Y.; Li, L.; Zhai, G. Curcumin-Loaded PLGA-PEG-PLGA Triblock Copolymeric Micelles: Preparation, Pharmacokinetics and Distribution in Vivo. *J. Colloid Interface Sci.* **2011**, *354*, 116–123. [[CrossRef](#)] [[PubMed](#)]
47. Mittal, G.; Sahana, D.K.; Bhardwaj, V.; Ravi Kumar, M.N.V. Estradiol Loaded PLGA Nanoparticles for Oral Administration: Effect of Polymer Molecular Weight and Copolymer Composition on Release Behavior in Vitro and in Vivo. *J. Control. Release* **2007**, *119*, 77–85. [[CrossRef](#)] [[PubMed](#)]
48. Torchilin, V.P. Polymer-Coated Long-Circulating Microparticulate Pharmaceuticals. *J. Microencapsul.* **1998**, *15*, 1–19. [[CrossRef](#)]
49. Knop, K.; Hoogenboom, R.; Fischer, D.; Schubert, U.S. Poly(Ethylene Glycol) in Drug Delivery: Pros and Cons as Well as Potential Alternatives. *Angew Chem. Int. Ed. Engl.* **2010**, *49*, 6288–6308. [[CrossRef](#)] [[PubMed](#)]
50. Torchilin, V.P.; Omelyanenko, V.G.; Papisov, M.I.; Bogdanov, A.A.; Trubetsky, V.S.; Herron, J.N.; Gentry, C.A. Poly(Ethylene Glycol) on the Liposome Surface: On the Mechanism of Polymer-Coated Liposome Longevity. *Biochim. Biophys. Acta* **1994**, *1195*, 11–20. [[CrossRef](#)]
51. Pan, C.; Zhang, T.; Li, S.; Xu, Z.; Pan, B.; Xu, S.; Jin, S.; Lu, G.; Yang, S.; Xue, Z.; et al. Hybrid Nanoparticles Modified by Hyaluronic Acid Loading an HSP90 Inhibitor as a Novel Delivery System for Subcutaneous and Orthotopic Colon Cancer Therapy. *Int. J. Nanomed.* **2021**, *16*, 1743–1755. [[CrossRef](#)] [[PubMed](#)]
52. Rudnik, E. *Compostable Polymer Materials*; Elsevier: Amsterdam, The Netherlands, 2019; ISBN 978-0-08-099442-0.
53. Dash, T.K.; Konkimalla, V.B. Poly-ε-Caprolactone Based Formulations for Drug Delivery and Tissue Engineering: A Review. *J. Control. Release* **2012**, *158*, 15–33. [[CrossRef](#)]
54. Dash, T.K.; Konkimalla, V.B. Polymeric Modification and Its Implication in Drug Delivery: Poly-ε-Caprolactone (PCL) as a Model Polymer. *Mol. Pharm* **2012**, *9*, 2365–2379. [[CrossRef](#)] [[PubMed](#)]
55. Lu, X.L.; Sun, Z.J.; Cai, W.; Gao, Z.Y. Study on the Shape Memory Effects of Poly(L-Lactide-Co-Epsilon-Caprolactone) Biodegradable Polymers. *J. Mater. Sci. Mater. Med.* **2008**, *19*, 395–399. [[CrossRef](#)]
56. Jeong, J.-C.; Lee, J.; Cho, K. Effects of Crystalline Microstructure on Drug Release Behavior of Poly(Epsilon-Caprolactone) Microspheres. *J. Control. Release* **2003**, *92*, 249–258. [[CrossRef](#)]
57. Chawla, J.S.; Amiji, M.M. Biodegradable Poly(Epsilon-Caprolactone) Nanoparticles for Tumor-Targeted Delivery of Tamoxifen. *Int. J. Pharm.* **2002**, *249*, 127–138. [[CrossRef](#)]
58. Niza, E.; Castro-Osma, J.A.; Posadas, I.; Alonso-Moreno, C.; Bravo, I.; Garzón, A.; Canales-Vázquez, J.; Ceña, V.; Lara-Sánchez, A.; Albaladejo, J. Assessment of Doxorubicin Delivery Devices Based on Tailored Bare Polycaprolactone against Glioblastoma. *Int. J. Pharm.* **2019**, *558*, 110–119. [[CrossRef](#)]
59. Pachence, J.M.; Bohrer, M.P.; Kohn, J. Biodegradable Polymers. *Princ. Tissue Eng.* **2007**, 323–339. [[CrossRef](#)]
60. Pulapura, S.; Kohn, J. Trends in the Development of Bioresorbable Polymers for Medical Applications. *J. Biomater. Appl.* **1992**, *6*, 216–250. [[CrossRef](#)] [[PubMed](#)]
61. Poly (γ-Glutamic Acid) Based Combination of Water-Insoluble Paclitaxel and TLR7 Agonist for Chemo-Immunotherapy—PubMed. Available online: <https://pubmed.ncbi.nlm.nih.gov/24954733/> (accessed on 28 May 2021).

62. Polyphosphoesters in Drug and Gene Delivery—PubMed. Available online: <https://pubmed.ncbi.nlm.nih.gov/12706047/> (accessed on 28 May 2021).
63. Andrieux, K.; Couvreur, P. Polyalkylcyanoacrylate Nanoparticles for Delivery of Drugs across the Blood-Brain Barrier. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* **2009**, *1*, 463–474. [[CrossRef](#)]
64. Fusser, M.; Øverbye, A.; Pandya, A.D.; Mørch, Y.; Borgos, S.E.; Kildal, W.; Snipstad, S.; Sulheim, E.; Fleten, K.G.; Askautrud, H.A.; et al. Cabazitaxel-Loaded Poly(2-Ethylbutyl Cyanoacrylate) Nanoparticles Improve Treatment Efficacy in a Patient Derived Breast Cancer Xenograft. *J. Control. Release* **2019**, *293*, 183–192. [[CrossRef](#)] [[PubMed](#)]
65. Villamagna, I.J.; Gordon, T.N.; Hurtig, M.B.; Beier, F.; Gillies, E.R. Poly(Ester Amide) Particles for Controlled Delivery of Celecoxib. *J. Biomed. Mater. Res. A* **2019**, *107*, 1235–1243. [[CrossRef](#)]
66. The Mechanisms of Drug Release in Poly(Lactic-Co-Glycolic Acid)-Based Drug Delivery Systems—A Review—PubMed. Available online: <https://pubmed.ncbi.nlm.nih.gov/21640806/> (accessed on 28 May 2021).
67. Fu, Y.; Kao, W.J. Drug Release Kinetics and Transport Mechanisms of Non-Degradable and Degradable Polymeric Delivery Systems. *Expert Opin. Drug Deliv.* **2010**, *7*, 429–444. [[CrossRef](#)] [[PubMed](#)]
68. Paarakh, M.P.; Jose, P.A.; Setty, C.; Christopher, G.P. Release kinetics—Concepts and applications. *Int. J. Pharm. Res. Technol.* **2018**, *8*, 12–20.
69. Uhrich, K.E.; Cannizzaro, S.M.; Langer, R.S.; Shakesheff, K.M. Polymeric Systems for Controlled Drug Release. *Chem. Rev.* **1999**, *99*, 3181–3198. [[CrossRef](#)] [[PubMed](#)]
70. Wilhelm, S.; Tavares, A.J.; Dai, Q.; Ohta, S.; Audet, J.; Dvorak, H.F.; Chan, W.C.W. Analysis of Nanoparticle Delivery to Tumours. *Nat. Rev. Mater.* **2016**, *1*, 1–12. [[CrossRef](#)]
71. Tenzer, S.; Docter, D.; Kuharev, J.; Musyanovych, A.; Fetz, V.; Hecht, R.; Schlenk, F.; Fischer, D.; Kiouptsi, K.; Reinhardt, C.; et al. Rapid Formation of Plasma Protein Corona Critically Affects Nanoparticle Pathophysiology. *Nat. Nanotechnol.* **2013**, *8*, 772–781. [[CrossRef](#)] [[PubMed](#)]
72. Liechty, W.B.; Kryscio, D.R.; Slaughter, B.V.; Peppas, N.A. Polymers for Drug Delivery Systems. *Annu. Rev. Chem. Biomol. Eng.* **2010**, *1*, 149–173. [[CrossRef](#)] [[PubMed](#)]
73. Zheng, Y.; Zheng, M.; Ma, Z.; Xin, B.; Guo, R.; Xu, X. 8—Sugar Fatty Acid Esters. In *Polar Lipids*; Ahmad, M.U., Xu, X., Eds.; Elsevier: Amsterdam, The Netherlands, 2015; pp. 215–243. ISBN 978-1-63067-044-3.
74. Sharipova, A.A.; Aidarova, S.B.; Mutaliyeva, B.Z.; Babayev, A.A.; Issakhov, M.; Issayeva, A.B.; Madybekova, G.M.; Grigoriev, D.O.; Miller, R. The Use of Polymer and Surfactants for the Microencapsulation and Emulsion Stabilization. *Colloids Interfaces* **2017**, *1*, 3. [[CrossRef](#)]
75. Mu, L.; Seow, P.-H.; Ang, S.-N.; Feng, S.-S. Study on Surfactant Coating of Polymeric Nanoparticles for Controlled Delivery of Anticancer Drug. *Colloid Polym Sci.* **2004**, *283*, 58–65. [[CrossRef](#)]
76. Hoshyar, N.; Gray, S.; Han, H.; Bao, G. The Effect of Nanoparticle Size on in Vivo Pharmacokinetics and Cellular Interaction. *Nanomedicine* **2016**, *11*, 673–692. [[CrossRef](#)]
77. Juan, A.; Cimas, F.J.; Bravo, I.; Pandiella, A.; Ocaña, A.; Alonso-Moreno, C. An Overview of Antibody Conjugated Polymeric Nanoparticles for Breast Cancer Therapy. *Pharmaceutics* **2020**, *12*, 802. [[CrossRef](#)] [[PubMed](#)]
78. Juan, A.; Cimas, F.J.; Bravo, I.; Pandiella, A.; Ocaña, A.; Alonso-Moreno, C. Antibody Conjugation of Nanoparticles as Therapeutics for Breast Cancer Treatment. *Int. J. Mol. Sci.* **2020**, *21*, 6018. [[CrossRef](#)] [[PubMed](#)]
79. Xiao, K.; Li, Y.; Luo, J.; Lee, J.S.; Xiao, W.; Gonik, A.M.; Agarwal, R.G.; Lam, K.S. The Effect of Surface Charge on in Vivo Biodistribution of PEG-Oligocholic Acid Based Micellar Nanoparticles. *Biomaterials* **2011**, *32*, 3435–3446. [[CrossRef](#)]
80. Nakamura, Y.; Mochida, A.; Choyke, P.L.; Kobayashi, H. Nanodrug Delivery: Is the Enhanced Permeability and Retention Effect Sufficient for Curing Cancer? *Bioconjugate Chem.* **2016**, *27*, 2225–2238. [[CrossRef](#)]
81. Wicki, A.; Witzigmann, D.; Balasubramanian, V.; Huwyler, J. *Nanomedicine in Cancer Therapy: Challenges, Opportunities, and Clinical Applications*; Elsevier: Amsterdam, The Netherlands, 2015; Volume 200.
82. Bareford, L.M.; Swaan, P.W. Endocytic Mechanisms for Targeted Drug Delivery. *Adv. Drug Deliv. Rev.* **2007**, *59*, 748–758. [[CrossRef](#)] [[PubMed](#)]
83. Bregoli, L.; Movia, D.; Gavigan-Imedio, J.D.; Lysaght, J.; Reynolds, J.; Prina-Mello, A. Nanomedicine Applied to Translational Oncology: A Future Perspective on Cancer Treatment. *Nanomed. Nanotechnol. Biol. Med.* **2016**, *12*, 81–103. [[CrossRef](#)] [[PubMed](#)]
84. Battistella, C.; Klok, H.-A. Controlling and Monitoring Intracellular Delivery of Anticancer Polymer Nanomedicines. *Macromol. Biosci.* **2017**, *17*, 1700022. [[CrossRef](#)]
85. Han, J.; Zhao, D.; Li, D.; Wang, X.; Jin, Z.; Zhao, K. Polymer-Based Nanomaterials and Applications for Vaccines and Drugs. *Polymers* **2018**, *10*, 31. [[CrossRef](#)] [[PubMed](#)]
86. Krishnaswamy, K.; Orsat, V. Chapter 2—Sustainable Delivery Systems Through Green Nanotechnology. In *Nano- and Microscale Drug Delivery Systems*; Grumezescu, A.M., Ed.; Elsevier: Amsterdam, The Netherlands, 2017; pp. 17–32. ISBN 978-0-323-52727-9.
87. Avgoustakis, K. Pegylated Poly(Lactide) and Poly(Lactide-Co-Glycolide) Nanoparticles: Preparation, Properties and Possible Applications in Drug Delivery. *Curr. Drug Deliv.* **2004**, *1*, 321–333. [[CrossRef](#)]
88. Wang, Q.; Wu, P.; Ren, W.; Xin, K.; Yang, Y.; Xie, C.; Yang, C.; Liu, Q.; Yu, L.; Jiang, X.; et al. Comparative Studies of Salinomycin-Loaded Nanoparticles Prepared by Nanoprecipitation and Single Emulsion Method. *Nanoscale Res. Lett.* **2014**, *9*, 351. [[CrossRef](#)] [[PubMed](#)]



89. Cohen-Sela, E.; Teitlboim, S.; Chorny, M.; Koroukhov, N.; Danenberg, H.D.; Gao, J.; Golomb, G. Single and Double Emulsion Manufacturing Techniques of an Amphiphilic Drug in PLGA Nanoparticles: Formulations of Mithramycin and Bioactivity. *J. Pharm. Sci.* **2009**, *98*, 1452–1462. [[CrossRef](#)]
90. Cegnar, M.; Kos, J.; Kristl, J. Cystatin Incorporated in Poly(Lactide-Co-Glycolide) Nanoparticles: Development and Fundamental Studies on Preservation of Its Activity. *Eur. J. Pharm. Sci.* **2004**, *22*, 357–364. [[CrossRef](#)]
91. Allémann, E.; Gurny, R.; Doelker, E. Preparation of Aqueous Polymeric Nanodispersions by a Reversible Salting-out Process: Influence of Process Parameters on Particle Size. *Int. J. Pharm.* **1992**, *87*, 247–253. [[CrossRef](#)]
92. Das, S.; Suresh, P.K.; Desmukh, R. Design of Eudragit RL 100 Nanoparticles by Nanoprecipitation Method for Ocular Drug Delivery. *Nanomedicine* **2010**, *6*, 318–323. [[CrossRef](#)]
93. Draheim, C.; de Crécy, F.; Hansen, S.; Collnot, E.-M.; Lehr, C.-M. A Design of Experiment Study of Nanoprecipitation and Nano Spray Drying as Processes to Prepare PLGA Nano- and Microparticles with Defined Sizes and Size Distributions. *Pharm. Res.* **2015**, *32*, 2609–2624. [[CrossRef](#)]
94. Jaworek, A. Micro- and Nanoparticle Production by Electrospraying. *Powder Technol.* **2007**, *176*, 18–35. [[CrossRef](#)]
95. Xu, S.; Nie, Z.; Seo, M.; Lewis, P.; Kumacheva, E.; Stone, H.A.; Garstecki, P.; Weibel, D.B.; Gitlin, I.; Whitesides, G.M. Generation of Monodisperse Particles by Using Microfluidics: Control over Size, Shape, and Composition. *Angew Chem. Int. Ed. Engl.* **2005**, *44*, 724–728. [[CrossRef](#)] [[PubMed](#)]
96. DeMello, A.J. Control and Detection of Chemical Reactions in Microfluidic Systems. *Nature* **2006**, *442*, 394–402. [[CrossRef](#)]
97. Reis, C.P.; Neufeld, R.J.; Ribeiro, A.J.; Veiga, F. Nanoencapsulation I. Methods for Preparation of Drug-Loaded Polymeric Nanoparticles. *Nanomedicine* **2006**, *2*, 8–21. [[CrossRef](#)] [[PubMed](#)]
98. He, H.; Liu, L.; Morin, E.E.; Liu, M.; Schwendeman, A. Survey of Clinical Translation of Cancer Nanomedicines—Lessons Learned from Successes and Failures Published as Part of the Accounts of Chemical Research Special Issue “Nanomedicine and Beyond. *Acc. Chem. Res.* **2019**, *52*, 2445–2461. [[CrossRef](#)] [[PubMed](#)]
99. Jain, M.M.; Gupte, S.U.; Patil, S.G.; Pathak, A.B.; Deshmukh, C.D.; Bhatt, N.; Haritha, C.; Babu, K.G.; Bondarde, S.A.; Digumarti, R.; et al. Paclitaxel Injection Concentrate for Nanodispersion versus Nab-Paclitaxel in Women with Metastatic Breast Cancer: A Multicenter, Randomized, Comparative Phase II/III Study. *Breast Cancer Res. Treat.* **2016**, *156*, 125–134. [[CrossRef](#)] [[PubMed](#)]
100. Saif, M.W.; Podoltsev, N.A.; Rubin, M.S.; Figueroa, J.A.; Lee, M.Y.; Kwon, J.; Rowen, E.; Yu, J.; Kerr, R.O. Phase II Clinical Trial of Paclitaxel Loaded Polymeric Micelle in Patients with Advanced Pancreatic Cancer. *Cancer Invest.* **2010**, *28*, 186–194. [[CrossRef](#)]
101. Kim, T.Y.; Kim, D.W.; Chung, J.Y.; Shin, S.G.; Kim, S.C.; Heo, D.S.; Kim, N.K.; Bang, Y.J. Phase I and Pharmacokinetic Study of Genexol-PM, a Cremophor-Free, Polymeric Micelle-Formulated Paclitaxel, in Patients with Advanced Malignancies. *Clin. Cancer Res.* **2004**, *10*, 3708–3716. [[CrossRef](#)] [[PubMed](#)]
102. Lee, K.S.; Chung, H.C.; Im, S.A.; Park, Y.H.; Kim, C.S.; Kim, S.B.; Rha, S.Y.; Lee, M.Y.; Ro, J. Multicenter Phase II Trial of Genexol-PM, a Cremophor-Free, Polymeric Micelle Formulation of Paclitaxel, in Patients with Metastatic Breast Cancer. *Breast Cancer Res. Treat.* **2008**, *108*, 241–250. [[CrossRef](#)]
103. Natale, R.; Socinski, M.; Hart, L.; Lipatov, O.; Spigel, D.; Gershengorn, B.; Weiss, G.; Kazmi, S.; Karaseva, N.; Gladkov, O.; et al. 41 Clinical Activity of BIND-014 (Docetaxel Nanoparticles for Injectable Suspension) as Second-Line Therapy in Patients (Pts) with Stage III/IV Non-Small Cell Lung Cancer. *Eur. J. Cancer* **2014**, *50*, 19. [[CrossRef](#)]
104. Hoff, D.D.V.; Mita, M.M.; Ramanathan, R.K.; Weiss, G.J.; Mita, A.C.; Lorusso, P.M.; Burris, H.A.; Hart, L.L.; Low, S.C.; Parsons, D.M.; et al. Phase I Study of PSMA-Targeted Docetaxel-Containing Nanoparticle BIND-014 in Patients with Advanced Solid Tumors. *Clin. Cancer Res.* **2016**, *22*, 3157–3163. [[CrossRef](#)]
105. Autio, K.A.; Garcia, J.A.; Alva, A.S.; Hart, L.L.; Milowsky, M.I.; Posadas, E.M.; Ryan, C.J.; Summa, J.M.; Youssoufian, H.; Scher, H.I.; et al. A Phase 2 Study of BIND-014 (PSMA-Targeted Docetaxel Nanoparticle) Administered to Patients with Chemotherapy-Naive Metastatic Castration-Resistant Prostate Cancer (mCRPC). *J. Clin. Oncol.* **2016**, *34*, 233.
106. Merle, P.; Camus, P.; Abergel, A.; Pageaux, G.P.; Masliah, C.; Bronowicki, J.P.; Zarski, J.P.; Pelletier, G.; Bouattour, M.; Farloux, L.; et al. Safety and Efficacy of Intra-Arterial Hepatic Chemotherapy with Doxorubicin-Loaded Nanoparticles in Hepatocellular Carcinoma. *ESMO Open* **2017**, *2*. [[CrossRef](#)] [[PubMed](#)]
107. Ribas, A.; Kalinoski, L.; Heidel, J.D.; Peterkin, J.; Seligson, D.B.; Zuckerman, J.E.; Choi, C.; Yen, Y.; Davis, M.E.; Tolcher, A.W. Systemic Delivery of siRNA via Targeted Nanoparticles in Patients with Cancer: Results from a First-in-Class Phase I Clinical Trial. *J. Clin. Oncol.* **2010**, *28*, 3022. [[CrossRef](#)]
108. Zuckerman, J.E.; Gritli, I.; Tolcher, A.; Heidel, J.D.; Lim, D.; Morgan, R.; Chmielowski, B.; Ribas, A.; Davis, M.E.; Yen, Y. Correlating Animal and Human Phase Ia/Ib Clinical Data with CALAA-01, a Targeted, Polymer-Based Nanoparticle Containing siRNA. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 11449–11454. [[CrossRef](#)] [[PubMed](#)]
109. Ocana, A.; Pandiella, A.; Siu, L.L.; Tannock, I.F. Preclinical Development of Molecular-Targeted Agents for Cancer. *Nat. Rev. Clin. Oncol.* **2010**, *8*, 200–209. [[CrossRef](#)]
110. Seruga, B.; Templeton, A.J.; Badillo, F.E.V.; Ocana, A.; Amir, E.; Tannock, I.F. Under-Reporting of Harm in Clinical Trials. *Lancet Oncol.* **2016**, *17*, e209–e219. [[CrossRef](#)]
111. Saleh, R.R.; Meti, N.; Ribnikar, D.; Goldvaser, H.; Ocana, A.; Templeton, A.J.; Seruga, B.; Amir, E. Associations between Safety, Tolerability, and Toxicity and the Reporting of Health-Related Quality of Life in Phase III Randomized Trials in Common Solid Tumors. *Cancer Med.* **2020**, *9*, 7888–7895. [[CrossRef](#)]

112. Fernandes Neto, J.M.; Nadal, E.; Bosdriesz, E.; Ooft, S.N.; Farre, L.; McLean, C.; Klarenbeek, S.; Jurgens, A.; Hagen, H.; Wang, L.; et al. Multiple Low Dose Therapy as an Effective Strategy to Treat EGFR Inhibitor-Resistant NSCLC Tumours. *Nat. Commun.* **2020**, *11*, 3157. [[CrossRef](#)]
113. Chang, L.; Ruiz, P.; Ito, T.; Sellers, W.R. Targeting Pan-Essential Genes in Cancer: Challenges and Opportunities. *Cancer Cell* **2021**, *39*, 466–479. [[CrossRef](#)]
114. Ocaña, A.; García-Alonso, S.; Amir, E.; Pandiella, A. Refining Early Antitumoral Drug Development. *Trends Pharm. Sci.* **2018**, *39*, 922–925. [[CrossRef](#)] [[PubMed](#)]
115. Cimas, F.J.; Niza, E.; Juan, A.; Noblejas-López, M.d.M.; Bravo, I.; Lara-Sanchez, A.; Alonso-Moreno, C.; Ocaña, A. Controlled Delivery of BET-PROTACs: In Vitro Evaluation of MZ1-Loaded Polymeric Antibody Conjugated Nanoparticles in Breast Cancer. *Pharmaceutics* **2020**, *12*, 986. [[CrossRef](#)]
116. Ahmad, A.; Khan, F.; Mishra, R.K.; Khan, R. Precision Cancer Nanotherapy: Evolving Role of Multifunctional Nanoparticles for Cancer Active Targeting. *J. Med. Chem.* **2019**, *62*, 10475–10496. [[CrossRef](#)] [[PubMed](#)]
117. Zou, W.; Sarisozen, C.; Torchilin, V.P. The Reversal of Multidrug Resistance in Ovarian Carcinoma Cells by Co-Application of Tariquidar and Paclitaxel in Transferrin-Targeted Polymeric Micelles. *J. Drug Target.* **2017**, *25*, 225–234. [[CrossRef](#)] [[PubMed](#)]
118. Xu, L.; Xu, S.; Wang, H.; Zhang, J.; Chen, Z.; Pan, L.; Wang, J.; Wei, X.; Xie, H.; Zhou, L.; et al. Enhancing the Efficacy and Safety of Doxorubicin against Hepatocellular Carcinoma through a Modular Assembly Approach: The Combination of Polymeric Prodrug Design, Nanoparticle Encapsulation, and Cancer Cell-Specific Drug Targeting. *ACS Appl. Mater. Interfaces* **2018**, *10*, 3229–3240. [[CrossRef](#)]
119. Voltan, R.; Secchiero, P.; Ruozi, B.; Forni, F.; Agostinis, C.; Caruso, L.; Vandelli, M.A.; Zauli, G. Nanoparticles Engineered with Rituximab and Loaded with Nutlin-3 Show Promising Therapeutic Activity in B-Leukemic Xenografts. *Clin. Cancer Res.* **2013**, *19*, 3871–3880. [[CrossRef](#)] [[PubMed](#)]
120. Sun, B.; Ranganathan, B.; Feng, S.-S. Multifunctional Poly(D,L-Lactide-Co-Glycolide)/Montmorillonite (PLGA/MMT) Nanoparticles Decorated by Trastuzumab for Targeted Chemotherapy of Breast Cancer. *Biomaterials* **2008**, *29*, 475–486. [[CrossRef](#)] [[PubMed](#)]
121. Qiao, Y.; Wan, J.; Zhou, L.; Ma, W.; Yang, Y.; Luo, W.; Yu, Z.; Wang, H. Stimuli-Responsive Nanotherapeutics for Precision Drug Delivery and Cancer Therapy. *Wiley Interdiscip. Rev. Nanomed. Nanobiotechnol.* **2019**, *11*, e1527. [[CrossRef](#)] [[PubMed](#)]
122. Xu, X.; Wu, J.; Liu, Y.; Yu, M.; Zhao, L.; Zhu, X.; Bhasin, S.; Li, Q.; Ha, E.; Shi, J.; et al. Ultra-PH-Responsive and Tumor-Penetrating Nanoplatfor for Targeted siRNA Delivery with Robust Anti-Cancer Efficacy. *Angew. Chem. Int. Ed. Engl.* **2016**, *55*, 7091–7094. [[CrossRef](#)] [[PubMed](#)]
123. Yin, T.; Wang, P.; Li, J.; Wang, Y.; Zheng, B.; Zheng, R.; Cheng, D.; Shuai, X. Tumor-Penetrating Codelivery of siRNA and Paclitaxel with Ultrasound-Responsive Nanobubbles Hetero-Assembled from Polymeric Micelles and Liposomes. *Biomaterials* **2014**, *35*, 5932–5943. [[CrossRef](#)] [[PubMed](#)]
124. Indoria, S.; Singh, V.; Hsieh, M.-F. Recent Advances in Theranostic Polymeric Nanoparticles for Cancer Treatment: A Review. *Int. J. Pharm.* **2020**, *582*, 119314. [[CrossRef](#)] [[PubMed](#)]
125. Zhang, C.; Zeng, Z.; Cui, D.; He, S.; Jiang, Y.; Li, J.; Huang, J.; Pu, K. Semiconducting Polymer Nano-PROTACs for Activatable Photo-Immunometabolic Cancer Therapy. *Nat. Commun.* **2021**, *12*, 2934. [[CrossRef](#)] [[PubMed](#)]