Open access • Posted Content • DOI:10.1101/449363

Polygenic scores for major depressive disorder and depressive symptoms predict response to lithium in patients with bipolar disorder — Source link

Azmeraw T. Amare, Klaus Oliver Schubert, Klaus Oliver Schubert, Liping Hou ...+122 more authors

Institutions: University of Adelaide, Mental Health Services, United States Department of Health and Human Services, Ludwig Maximilian University of Munich ...+44 more institutions

Published on: 22 Oct 2018 - bioRxiv (Cold Spring Harbor Laboratory)

Topics: Major depressive disorder, Bipolar disorder and Lithium (medication)

Polygenic scores for major depressive disorder and depressive symptoms predict response to lithium in patients with bipolar disorder

Azmeraw T. Amare, MPH, MSc, PhD[1,2*]; Klaus Oliver Schubert, MD, PhD[1,3*]; Liping Hou, PhD[4]; Scott R. Clark, MBBS, PhD [1]; Sergi Papiol, PhD[5,6]; Micah Cearns, BSc (Hons)[1]; Urs Heilbronner, PhD[5,7]; Franziska Degenhardt, MD[8]; Fasil Tekola-Ayele, PhD[9]; Yi-Hsiang Hsu, PhD[10,11]; Tatyana Shekhtman, MSc[12]; Mazda Adli, MD[13]; Nirmala Akula, PhD[4]; Kazufumi Akiyama, MD[14]; Raffaella Ardau, MD[15]; Bárbara Arias, PhD[16]; Jean-Michel Aubry, MD[17]; Lena Backlund, MD, PhD[18]; Abesh Kumar Bhattacharjee, MD[12]; Frank Bellivier, MD, PhD[19]; Antonio Benabarre, MD, PhD[20]; Susanne Bengesser, MD[21]; Joanna M. Biernacka, PhD[22,23]; Armin Birner, MD[21]; Clara Brichant-Petitjean, MD[19]; Pablo Cervantes, MD[24]; Hsi-Chung Chen, MD, PhD[25]; Caterina Chillotti, MD[15]; Sven Cichon, PhD[8,26]; Cristiana Cruceanu, PhD[27]; Piotr M. Czerski, PhD[28]; Nina Dalkner, MSc[21]; Alexandre Dayer, MD[17]; Maria Del Zompo, MD[29]; J. Raymond DePaulo, MD[30]; Bruno Étain, MD, PhD[19]; Peter Falkai, MD[32]; Andreas J. Forstner, MD[8,26,33]; Louise Frisen, MD[18]; Mark A. Frye, MD[23]; Janice M. Fullerton, PhD[34, 35]; Sébastien Gard, MD[36]; Julie S. Garnham, BN[37]; Fernando S. Goes, MD[30]; Maria Grigoroiu-Serbanescu, PhD[38]; Paul Grof, MD, PhD[39]; Ryota Hashimoto, MD, PhD[40,41]; Joanna Hauser, MD[28]; Stefan Herms, Dipl.Biol.[8,26]; Per Hoffmann, PhD[8,26]; Andrea Hofmann, PhD[8]; Stephane Jamain, PhD[31]; Esther Jiménez, PhD[20]; Jean-Pierre Kahn, MD, PhD[42]; Layla Kassem, PhD[4]; Po-Hsiu Kuo, PhD[43]; Tadafumi Kato, MD, PhD[44]; John Kelsoe, MD[12]; Sarah Kittel-Schneider, MD[45]; Sebastian Kliwicki, MD[46]; Barbara König, MSc[47]; Ichiro Kusumi, MD[48]; Gonzalo Laje, MD[4]; Mikael Landén, MD[49,50]; Catharina Lavebratt, PhD[18]; Marion Leboyer, MD, PhD[51]; Susan G. Leckband, BSc[52]; Alfonso Tortorella, MD[53]; Mirko Manchia, MD, PhD[54,55]; Lina Martinsson, MD[56]; Michael J. McCarthy, MD, PhD[12,57]; Susan McElroy, MD[58]; Francesc Colom, PhD[59,60,]; Marina Mitjans, PhD[60,61]; Francis M. Mondimore, MD[30]; Palmiero Monteleone, MD[62,63]; Caroline M. Nievergelt, PhD[12]; Markus M. Nöthen, MD[8]; Tomas Novák, MD[64]; Claire O'Donovan, MB[37]; Norio Ozaki, MD[65]; Urban Ösby, MD, PhD[66]; Andrea Pfennig, MD[67]; James B. Potash, MD, MPH[30]; Andreas Reif, MD[45]; Major Depressive Disorder Working Group of the Psychiatric Genomics Consortium[74], Eva Reininghaus, MD[21]; Guy A. Rouleau, MD[68]; Janusz K. Rybakowski, MD[46]; Martin

Schalling, MD[18]; Peter R. Schofield, PhD, DSc[34,35]; Barbara W. Schweizer, RN[30]; Giovanni Severino, MD[29]; Paul D. Shilling, PhD[12]; Katzutaka Shimoda, MD[69]; Christian Simhandl, MD[70]; Claire M. Slaney, RN[37]; Alessio Squassina, PhD[29]; Thomas Stamm, MD[13]; Pavla Stopkova, MD[64]; Mario Maj, MD[63]; Gustavo Turecki, MD[27]; Eduard Vieta, MD, PhD[20]; Julia Veeh, PhD[45]; Stephanie H. Witt, PhD[71]; Adam Wright, MCP[72]; Peter P. Zandi, PhD[73]; Philip B. Mitchell, MD[72]; Michael Bauer, MD,PhD[67]; Martin Alda, MD[37]; Marcella Rietschel, MD[71]; Francis J. McMahon, MD[4]; Thomas G. Schulze, MD[4,5,7,30,71]; Bernhard T. Baune, MD, PhD[75]

* The authors Azmeraw T Amare and K. Oliver Schubert contributed equally to this study and are regarded as shared first authors.

AFFILIATIONS

[1]Discipline of Psychiatry, School of Medicine, University of Adelaide, Adelaide, SA, Australia

[2]South Australian Academic Health Science and Translation Centre, South Australian Health and Medical Research Institute (SAHMRI), Adelaide, South Australia, Australia

[3]Northern Adelaide Local Health Network, Mental Health Services, Adelaide, SA, Australia

[4]Intramural Research Program, National Institute of Mental Health, National Institutes of Health, US Department of Health & Human Services, Bethesda, MD, USA

[5]Institute of Psychiatric Phenomics and Genomics (IPPG), University Hospital, LMU Munich, Munich, Germany

[6]Department of Psychiatry and Psychotherapy, Ludwig-Maximilian-University Munich, Munich, Germany

[7]Department of Psychiatry and Psychotherapy, University Medical Center (UMG), Georg-August University Göttingen, Göttingen, Germany

[8]Institute of Human Genetics, University of Bonn and Department of Genomics, Life & Brain Center, Bonn, Germany

[9]Epidemiology Branch, Division of Intramural Population Health Research, Eunice Kennedy Shriver National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, MD, USA

[10]HSL Institute for Aging Research, Harvard Medical School, Boston, MA, United States

[11]Program for Quantitative Genomics, Harvard School of Public Health, Boston, MA, United States

[12]Department of Psychiatry, University of California San Diego, San Diego, CA, United States

[13]Department of Psychiatry and Psychotherapy, Charité - Universitätsmedizin Berlin, Campus Charité Mitte, Berlin, Germany

[14]Department of Biological Psychiatry and Neuroscience, Dokkyo Medical University School of Medicine, Mibu, Tochigi, Japan

[15]Unit of Clinical Pharmacology, Hospital University Agency of Cagliari, Cagliari, Italy

[16]Unitat de Zoologia i Antropologia Biològica (Dpt. Biologia Evolutiva, Ecologia i Ciències Ambientals), Facultat de Biologia and Institut de Biomedicina (IBUB), University of Barcelona, CIBERSAM, Barcelona, Spain

[17]Department of Psychiatry, Mood Disorders Unit, HUG - Geneva University Hospitals, Geneva, Switzerland

[18]Department of Molecular Medicine and Surgery, Karolinska Institutet, Stockholm, Sweden, and Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden

[19]INSERM UMR-S 1144, Université Paris Diderot, Département de Psychiatrie et de Médecine Addictologique, AP-HP, Groupe Hospitalier Saint-Louis-Lariboisière-F.Widal, Paris, France

[20]Bipolar Disorder Program, Institute of Neuroscience, Hospital Clinic, University of Barcelona, IDIBAPS, CIBERSAM, Barcelona, Catalonia, Spain

[21]Department of Psychiatry and Psychotherapeutic Medicine, Research Unit for bipolar affective disorder, Medical University of Graz, Graz, Austria

[22]Department of Health Sciences Research, Mayo Clinic, Rochester, MN, United States

[23]Department of Psychiatry and Psychology, Mayo Clinic, Rochester, MN, United States

[24]The Neuromodulation Unit, McGill University Health Centre, Montreal, Canada

[25]Department of Psychiatry & Center of Sleep Disorders, National Taiwan University Hospital, Taipei, Taiwan

[26]Human Genomics Research Group, Department of Biomedicine, University Hospital Basel, Basel, Switzerland

[27]Douglas Mental Health University Institute, McGill University, Montreal, Canada

[28]Psychiatric Genetic Unit, Poznan University of Medical Sciences, Poznan, Poland

[29]Department of Biomedical Sciences, University of Cagliari, Cagliari, Italy

[30]Department of Psychiatry and Behavioral Sciences, Johns Hopkins University, Baltimore, MD, United States

[31]Inserm U955, Translational Psychiatry laboratory, Fondation FondaMental, Créteil, France

[32]Department of Psychiatry and Psychotherapy, Ludwig-Maximilian-University Munich, Munich, Germany

[33]Department of Psychiatry (UPK), University of Basel, Basel, Switzerland

[34]Neuroscience Research Australia, Sydney, NSW, Australia

[35]School of Medical Sciences, University of New South Wales, Sydney, NSW, Australia

[36]Service de psychiatrie, Hôpital Charles Perrens, Bordeaux, France

[37]Department of Psychiatry, Dalhousie University, Halifax, Nova Scotia, Canada

[38]Biometric Psychiatric Genetics Research Unit, Alexandru Obregia Clinical Psychiatric Hospital, Bucharest, Romania

[39]Mood Disorders Center of Ottawa, Ontario, Canada

[40]Molecular Research Center for Children's Mental Development, United Graduate School of Child Development, Osaka University, Osaka, Japan

[41]Department of Psychiatry, Osaka University Graduate School of Medicine, Osaka, Japan

[42]Service de Psychiatrie et Psychologie Clinique, Centre Psychothérapique de Nancy - Université de Lorraine, Nancy, France

[43]Department of Public Health & Institute of Epidemiology and Preventive Medicine, College of Public Health, National Taiwan University, Taipei, Taiwan

[44]Laboratory for Molecular Dynamics of Mental Disorders, RIKEN Brain Science Institute, Saitama, Japan

[45]Department of Psychiatry, Psychosomatic Medicine and Psychotherapy, University Hospital Frankfurt, Frankfurt, Germany

[46]Department of Adult Psychiatry, Poznan University of Medical Sciences, Poznan, Poland

[47]Department of Psychiatry and Psychotherapeutic Medicine, Landesklinikum Neunkirchen, Neunkirchen, Austria

[48]Department of Psychiatry, Hokkaido University Graduate School of Medicine, Sapporo, Japan

[49]Institute of Neuroscience and Physiology, the Sahlgrenska Academy at the Gothenburg University, Gothenburg, Sweden

[50]Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden

[51]Inserm U955, Translational Psychiatry laboratory, Université Paris-Est-Créteil, Department of Psychiatry and Addictology of Mondor University Hospital, AP-HP, Fondation FondaMental, Créteil, France

[52]Office of Mental Health, VA San Diego Healthcare System, San Diego, CA, United States

[53]Department of Psychiatry, University of Perugia, Italy

[54]Section of Psychiatry, Department of Medical Sciences and Public Health, University of Cagliari, Cagliari, Italy

[55]Department of Pharmacology, Dalhousie University, Halifax, NS, Canada

[56]Department of Clinical Neurosciences, Karolinska Institutet, Stockholm, Sweden

[57]Department of Psychiatry, VA San Diego Healthcare System, San Diego, CA, United States

[58]Department of Psychiatry, Lindner Center of Hope / University of Cincinnati, Mason, OH, United States

[59]Mental Health Research Group, IMIM-Hospital del Mar, Barcelona, Catalonia, Spain

[60]Centro de Investigación Biomédica en Red de Salud Mental (CIBERSAM), Instituto de Salud Carlos III, Madrid, Spain

[61]Clinical Neuroscience, Max Planck Institute of Experimental Medicine, Göttingen, Germany

[62]Neurosciences Section, Department of Medicine, Surgery and Dentistry “Scuola Medica Salernitana”, University of Salerno, Salerno, Italy

[63]Department of Psychiatry, University of Campania “Luigi Vanvitelli”, Naples, Italy

[64]National Institute of Mental Health, Klecany, Czech Republic

[65]Department of Psychiatry & Department of Child and Adolescent Psychiatry, Nagoya University Graduate School of Medicine, Nagoya, Japan

[66]Department of Neurobiology, Care Sciences, and Society, Karolinska Institutet and Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden

[67]Department of Psychiatry and Psychotherapy, University Hospital Carl Gustav Carus, Medical Faculty, Technische Universität Dresden, Germany

[68]Montreal Neurological Institute and Hospital, McGill University, Montreal, Canada

[69]Department of Psychiatry, Dokkyo Medical University School of Medicine, Mibu, Tochigi, Japan

[70]Bipolar Center Wiener Neustadt, Sigmund Freud University, Medical Faculty, Vienna, Austria

[71]Department of Genetic Epidemiology in Psychiatry, Central Institute of Mental Health, Medical Faculty Mannheim, University of Heidelberg, Mannheim, Germany

[72]School of Psychiatry, University of New South Wales, and Black Dog Institute, Sydney, Australia

[73]Department of Mental Health, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, United States

[74] For a full list of MDD working group of the PGC investigators, see the Supplementary Material

[75] Department of Psychiatry, Melbourne Medical School, University of Melbourne, Parkville, Victoria, Australia

Correspondence to:

Bernhard T. Baune,

Department of Psychiatry

Melbourne Medical School

University of Melbourne

Parkville VIC 3010

E-Mail: bbaune@unimelb.edu.au

ABSTRACT

Background: Lithium is a first-line medication for bipolar disorder (BD), but only ~30% of patients respond optimally to the drug. Since genetic factors are known to mediate lithium treatment response, we hypothesized whether polygenic susceptibility to the spectrum of depression traits is associated with treatment outcomes in patients with BD. In addition, we explored the potential molecular underpinnings of this relationship.

Methods: Weighted polygenic scores (PGSs) were computed for major depressive disorder (MDD) and depressive symptoms (DS) in BD patients from the Consortium on Lithium Genetics (ConLi^{+}Gen; n=2,586) who received lithium treatment. Lithium treatment outcome was assessed using the ALDA scale. Summary statistics from genome-wide association studies (GWAS) in MDD (130,664 cases and 330,470 controls) and DS (n=161,460) were used for PGS weighting. Associations between PGSs of depression traits and lithium treatment response were assessed by binary logistic regression. We also performed a cross-trait meta-GWAS, followed by Ingenuity® Pathway Analysis.

Outcomes: BD patients with a low polygenic load for depressive traits were more likely to respond well to lithium, compared to patients with high polygenic load (MDD: OR =1.64 [95%CI: 1.26-2.15], lowest vs highest PGS quartiles; DS: OR=1.53 [95%CI: 1.18-2.00]). Associations were significant for type 1, but not type 2 BD. Cross-trait GWAS and functional characterization implicated voltage-gated potassium channels, insulin-related pathways, mitogen-activated protein-kinase (MAPK) signaling, and miRNA expression.

Interpretation: Genetic loading to depression traits in BD patients lower their odds of responding optimally to lithium. Our findings support the emerging concept of a lithium-responsive biotype in BD.

Funding: see attached details

Keywords: lithium treatment, Major depressive disorder, depressive symptoms, depressive traits, bipolar disorder, polygenic score, pharmacogenomics, Voltage-gated potassium channel, insulin, MAPK.

Word count
Abstract: 243
main text: 3199

INTRODUCTION

Bipolar disorder (BD) is a chronic and severe psychiatric illness characterized by episodic, abnormal manic and depressive mood states. An estimated 48.8 million people are affected by BD globally[1]. The disorder accounts for 9.9 million years of life lived with disability worldwide[1], and substantially increases all-cause mortality and risk of suicide[2].

Amongst available treatments, lithium is regarded as a gold standard by several clinical guidelines[3,4]. Lithium uniquely protects against both manic and depressive illness phases, has demonstrated protective effects against suicide[5-7], and is particularly effective in preventing rehospitalisation[8]. However, not all patients with BD fully benefit from lithium, and only about 30% show full response to the drug[5-7]. In current psychiatric practice, no biological or clinical markers exist that could reliably predict responsiveness to lithium[9], and prescribing cannot be targeted to patients who benefit most while avoiding side effects and sub-optimal treatment for poor responders[10 11 12,13].

In order to develop objective response markers and to move towards personalized prescribing of lithium for BD patients, a better understanding of the biological mechanisms underlying lithium response is urgently required. Recent genome-wide association studies (GWAS) carried out by our International Consortium on Lithium Genetics (ConLi^{+}Gen)[5] and others[14,15] have indicated that genetic variation could be an important mediator of response to long-term lithium treatment response in BD patients. Additionally, we have recently demonstrated that high genetic loading for schizophrenia (SCZ) risk variants in people with BD decreases the likelihood of favorable response to lithium[16], suggesting that polygenic score (PGS) analysis of mental and physical traits could yield important information on the genetic architecture of BD phenotypes[17 18 19]. In the current study, we

address the question of whether genetic loading for major depressive disorder (MDD) and depressive symptoms (DS) contribute to treatment outcomes in BD.

BD and MDD show 47% genetic overlap[20-22], and shared risk genes and biological pathways have been described[22,23]. Lithium can be effective as an augmentation strategy in MDD patients who have experienced an insufficient response to first-line antidepressants[24,25] and is protective against further MDD episodes after symptom remission has been achieved[20,26]. Moreover, a large observational study based on the Finnish registry showed that lithium is the most effective agent preventing rehospitalization in MDD[26].

On the other hand, in BD, lithium is more effective in preventing manic than depressive episodes[27,28], leading to the notion that better lithium responders might be more likely to experience manic predominant polarity, as opposed to depressive predominant polarity[29]. In support of this view, one study found that excellent lithium responders were characterized by a manic but not depressive polarity of the index episode [30]. Another study described an episodic illness pattern of 'mania-depression-interval' as a predictor for good response, whereas a 'depression-mania-interval' predicted poorer outcomes[31]. Inter-episode residual mood symptoms, as opposed to full remission[6,7,32], a rapid cycling pattern[31,32], and a history of mixed episodes[33,34] have also been described as predictors of poor response.

On the background of these complex interactions between BD, MDD, and lithium treatment, we asked whether BD patients with a high genetic susceptibility for depression (MDD and DS), expressed by their PGS for these traits, would respond better or worse to lithium than BD patients with a low genetic loading[35 36]. To explore potential genetic and molecular drivers of any detected polygenic association, we carried out a cross-trait GWAS meta-analysis, combining summary statistics from the largest available GWASs for MDD[35] and

DS[36] with GWASs for response to lithium treatment in patients with BD[5]. Overlapping SNPs that met genome-wide significance in the meta-GWAS were subsequently analyzed for biological context using the Ingenuity® Pathway Analysis platform (IPA®).

METHODS AND MATERIALS

Discovery GWAS summary data sets

The polygenic score and cross-trait meta-analysis for this study were based on genetic data from the International Consortium on Lithium Genetics (ConLi^{+}Gen)[5], and the summary statistics of three largest GWASs available for MDD[35], DS[36] and treatment response to lithium in patients with BD[5].

Major depressive disorder

The most recent GWAS meta-analysis of 9.6 million SNPs (Psychiatric Genomics Consortium-PGC; http://www.med.unc.edu/pgc/), obtained from 7 cohorts (deCODE, Generation Scotland, GERA, iPSYCH, UK Biobank, CONVERGE and 23andMe) containing 130,664 MDD cases and 330,470 healthy controls, identified 44 independent loci that reached the criteria for statistical significance. Details on this study are available elsewhere[35].

Depressive symptoms

The GWAS on DS (N = 161,460) used data from the PGC, the UK Biobank (UKB), the Resource for Genetic Epidemiology Research on Aging (GERA) Cohort and the Social Science Genetic Association Consortium (SSGAC) https://www.thessgac.org/. The summary statistics were made publically available for scientific usage[36].

Lithium treatment response in BD

The summary GWAS on lithium treatment response was produced through a combined analysis of 2,563 patients collected by 22 participating sites from the International Consortium on Lithium Genetics (ConLi^{+}Gen) http://www.conligen.org/. In our analysis, we used the data analyzed on the categorical scale for lithium response[5].

Target Study Sample

For the PGS analysis, clinical data on lithium treatment response and genetic information were obtained from the International Consortium on Lithium Genetics (ConLi^{+}Gen; www.ConLiGen.org) for n=2,586 patients (including 23 patients in the replication sample)[3,5,16]. A series of quality control procedures were implemented on the genotype data before and after imputation as described below.

Target outcome

Lithium treatment response was assessed using the validated "Retrospective Criteria of Long-Term Treatment Response in Research Subjects with Bipolar Disorder" scale, also known as the ALDA scale [7,37,38]. This scale quantifies symptom improvement over the course of treatment (A score, range 0–10), which is then weighted against five criteria (B score) that assess confounding factors[5]. Patients with a total score of 7 or higher were categorized as "good responders", and the remainder were categorized as poor responders[5,38]. In addition to the ALDA scale scores, information on covariates such as age and gender was collected, as described in detail elsewhere[5].

Genotyping and quality control

The genome-wide genotypes, as well as clinical and demographic data, were collected by 22 participating sites. Quality control (QC) procedures were implemented on the genotype data using PLINK, version 1.09 prior to imputation[39]. Samples with low genotype rates <95%,

sex inconsistencies (based on X-chromosome heterozygosity), and one of a pair of genetically related individuals were excluded. SNPs were excluded based on the following criteria: a poor genotyping rate (<95%), strand ambiguity (A/T and C/G SNPs), a low minor allele frequency (MAF<1%), or those deviated from genotype frequency expectations under the Hardy-Weinberg Equilibrium ($p<10^{-6}$).

Imputation

The genotype data passing QC were imputed on the Michigan server[40] (https://imputationserver.sph.umich.edu) separately for each genotype platform using reference data from the 1000 Genomes Project Phase 3 (Version 5). The European reference panel was used for all the samples except for those from Japan and Taiwan, for which an East Asian reference population data was used. After excluding low-frequency SNPs (MAF<10%); low-quality variants (imputation INFO < 0.9); and indels, the imputed dosages were converted to best guess genotypes. The subsequent polygenic analyses were performed using these best guess genotypes.

STATISTICAL ANALYSES

Polygenic score (PGS) association analysis

PGSs were calculated using the approach previously described by the International Schizophrenia Consortium[41]. Prior to PGS computation, independent SNPs were identified through a clumping procedure implemented in PLINK software, version 1.09 run on Linux[39]. Quality-controlled SNPs were clumped for linkage disequilibrium based on GWAS association p-value informed clumping at $r^2 = 0.1$ within a 250- kilobase window to create a SNP-set in linkage equilibrium (*plink --clump-p1 1 --clump-p2 1 --clump-r2 0.1 --clump-kb 250*). Polygenic risk scores were calculated for MDD and DS in the ConLi$^+$Gen sample at

ten GWAS association p-value thresholds ($<1x10^{-4}$, $<1x10^{-3}$, <0.01, <0.05, <0.1, <0.2, <0.3, <0.4, <0.5, <1).

Cross-trait meta-analysis of genome-wide association studies

Having identified a significant polygenic association that indicated the presence of genetic overlap, we conducted cross-trait meta-analyses of GWASs to identify genetic polymorphisms that were likely to increase the susceptibility to both MDD and DS as well as influence lithium treatment response in patients with BD. The cross-trait meta-analyses were performed by combining the summary statistics for GWAS on lithium response [5] and GWAS on MDD[35] and DS[36]. We applied the O'Brien's (OB) method and the direct Linear Combination of dependent test statistics (dLC) approach[42,43], which are implemented in the C^{++} eLX package (further details in supplementary methods).

Ingenuity® Pathway Analysis (IPA®)

To characterize the biological context of the discovered SNPs from the cross-trait meta-analyses, we implemented a functional analysis using QIAGEN's Ingenuity® Pathway Analysis (IPA®, QIAGEN Redwood City, CA, USA, www.qiagen.com/ingenuity). For details see supplementary methods.

RESULTS

Sample characteristics and lithium treatment response rate

After QC, 2,586 patients (3,193 before QC) remained for analysis. While 2,366 were of European ancestry, the remaining were of Asian ancestry. In all, 704 (27.2%) responded to lithium treatment (ALDA score ≥7). Detailed sample and demographics details have been described previously[16].

MDD and DS PGS are associated with lithium treatment response in BD

Associations between the PGSs for MDD and DS with lithium treatment response were found at various p-value thresholds. The strongest association were found for MDD (p= 0.0003) at $P_T < 5x10^{-2}$, $R^2 = 0.7\%$ and for DS (p= 0.0003) at $P_T < 1x10^{-2}$, $R^2 = 0.7\%$) (Figure 1).

[insert figure 1 about here]

High genetic loadings for MDD and DS are associated with poorer response to lithium in BD

We divided the study population into quartiles, according to their polygenic loading for MDD and DS, respectively. As shown in Figure 2 and Table 1, BD patients who carry a lower polygenic load (1st quartile) for MDD or DS have higher odds of favorable lithium treatment response, compared to patients carrying a high polygenic load (4th quartile). The odds ratio (OR) of favorable response for patients in the 1st quartile compared with those in

the 4th quartile was 1.64 [95%CI: 1.26-2.15] for MDD PGS, and 1.53 [95%CI: 1.18-2.00]) for DS PGS (Table 1 & Figure 2).

[insert table 1 and figure 2 about here]

Exploratory analyses were conducted to investigate whether the associations between MDD/DS PGSs and lithium response are driven by BD type 1 or type 2 (type 1 n=2,044; type 2 n=506). The observed inverse effects for MDD and DS PGS in the entire sample remained statistically significant for BD type 1 patients (eTable 3) only. For the BD type 2 group, an opposite non-significant trend for MDD PGSs, i.e. higher MDD loading was associated with *better* response to lithium, was observed (eTable 3).

Cross-trait meta-analysis of GWAS on lithium treatment response and GWAS on MDD and depressive symptoms yields 7 significant loci

Subsequent to the PGS analysis, we performed a SNP-based cross-trait meta-analysis by combining the summary statistics for the GWASs on: a) MDD and lithium treatment response; and b) DS and lithium treatment response — with the aim of identifying individual genetic variants implicated in the genetic susceptibility to both depression traits and lithium treatment response. These analyses yielded 7 loci with p-values below the genome-wide significance level ($p<5x10^{-8}$). These loci, and their nearby genes, were rs2327713: *PUM3* [OMIM: 609960], rs7134419: *KSR2* [OMIM: 610737], rs59659806: *RASGRP1* [OMIM: 603962], rs7405404: *ERCC4* [OMIM: 133520], rs11657502: *MYO18A* [OMIM: 610067], rs8099160: *DCC* [OMIM: 120470], rs6066909: *ARFGEF2* [OMIM: 605371] (Figure 3, Table 2)

[insert table 2 and figure 3 about here]

Functional and biological characterization of genetic loci associated with lithium response and MDD/depressive symptoms

To characterize the functional implications of the SNPs identified by cross-trait meta-analysis, we first explored the functional genetic context of these variations by examination of SNPs with high linkage disequilibrium, characterization of their nearby hosting genes, and eQTL lookup from published databases. This approach yielded a list of 39 genes with potential functional significance [eTable 1]. Second, we investigated the biological roles of these 39 genes using IPA® analysis.

IPA® identified cellular development and cellular growth and proliferation as the top cellular functions associated with the 39 genes. These associations were driven by only a handful of genes, including micro RNA (miR) -144, miR-451, regulatory factor X3 (RFX), phosphatidylinsositol-3,4,5-trisphosphate dependent Rac exchange factor 1 (PREX1), and RAS guanyl releasing protein 1 (RASGRP1).

The top IPA®-identified functional networks containing dataset genes pointed to 'hub' functions for insulin, ERK, JNK (network 1), ELAV like RNA binding protein 1 (ELAV1)(network 2), and nuclear RNA export factor 1 (NXF1) (network 3), [eFigures 1 A-C, eTable 2]. The IPA® top hits for upstream regulators were potassium voltage-gated channel subfamily A member 1 (KCNA1) and leucine-2-alanine encephalin, both of which impact on the dataset gene potassium voltage-gated channel subfamily B member 1 (KCNB1) (Table 3).

[Insert Table 3 about here]

DISCUSSION

Our study represents the first direct evidence of an association between a genetic predisposition for depression and poorer response to lithium treatment in patients with BD. Using PGS analyses of genetic variants related to MDD and DS, we found that BD patients with low genetic loading for these variants were about 1.6 times *more* likely to have favorable long-term outcomes following lithium treatment compared to BD patients with high MDD/DS genetic loading. To explore which genes might functionally drive these effects, we carried out a cross-trait meta-analysis of lithium response and MDD/DS. Pathway analyses of variants associated with both traits implicated roles for voltage-gated potassium channels, for insulin-related mechanisms, for the ERK and JNK signaling pathways, and for the micro RNAs miR-144 and miR-451.

Our findings could form part of a genetic explanation for the previously described clinical observations in relation to mania, depression and lithium response in BD[6,7,27-34] and supports the notion that lithium responsiveness could be associated with a 'core' bipolar phenotype in the *Kraepelinian* form of manic depression[34,44]. Such a concept is further supported by our previous finding of an inverse association of lithium response and schizophrenia PGS in BD[16].

Although these results have to be interpreted with caution due to smaller subgroup sample sizes, the exploratory analyses indicate that BD type 2 patients may differ from type 1 patients with regards to the depression PGS on lithium response association. A non-significant trend for *improved* lithium treatment response was found in type 2 patients with high MDD PGS. Genetically, differences between type 1 and type 2 BD cohorts have been suggested [45 46], and type 2 patients show substantially higher genetic overlap with MDD[47]. Lithium's effectiveness as an adjunct antidepressant treatment for people with treatment-resistant MDD is well

established[48-54]; therefore, our finding raises the intriguing possibility of MDD-specific mechanisms of action, which might be different from the mechanisms underlying the more 'anti-manic' response in BD type 1.

Our cross-trait GWAS meta-analysis yielded 7 loci that exceeded a genome-wide significance level of 5×10^{-8}. Amongst the nearby genes of these loci, the *DCC* gene and its encoded netrin 1 receptor has previously been shown to play an important role in mediating axonal growth in developing human brain[55,56]. Additionally, genetic variation within the *DCC* gene has previously been shown to be associated with depressive symptoms[36].

Based on the 7 loci identified by cross-trait GWAS meta-analysis, we generated a list of 39 functionally related genes by examination of SNPs with high linkage disequilibrium, characterization of their nearby hosting genes, and eQTL lookup from published databases (eTable 1). Functional exploration of these genes by IPA® implicated the voltage-gated potassium channel (K^+v), KCNA1 as a top upstream regulator (Table 3). The family of K^+v proteins plays a role in the regulation of the excitability of neurons, and genetic variations in these channels have been linked to epilepsy[57,58]. Recent experiments with inducible pluripotent stem cells from patients with BD suggest that neural hyperexcitability could be a core pathophysiological trait in BD, which is reversible by lithium in a subset of patients[59,60]. Remarkably, regulation of KCNA1 gene expression by lithium was shown to be involved in these 'therapeutic' lithium effects[59]. Therefore, a role for K^+v's in the genetic architecture of lithium response in BD appears plausible.

Further functional characterization by IPA® suggested that genes regulating insulin homeostasis could be important mediators of the MDD-lithium relationship (eFigure 1). These genes included

regulating factor X3 (RFX3), Phosphatidylinositol 3,4,5-trisphosphate-dependent Rac exchanger 1 (PREX 1), and K^+v subfamily B member 1(KCNB1). Interestingly, previous clinical studies have shown that BD patients with impaired glucose tolerance or diabetes mellitus type 2 are over 8-times less likely to benefit from lithium and have an overall less favorable illness course [19].

Functionally, the genes *RFX3*, *PREX1*, and *KCNB1* are involved in insulin regulation in various ways. The transcription factor RFX3 is required for the differentiation and function of insulin-producing, mature pancreatic beta cells, and regulates the beta-cell promotor of the glucokinase gene[61]. Interestingly, *RFX3* variants were also implicated in a recent GWAS examining sleeplessness/insomnia[62], a condition with aetiological relationships to BD[63], and depression[64]. Variants of the PREX1 gene on chromosome 20q12-13.1were associated with increased risk of diabetes mellitus type 2 and increased BMI in a cohort of European Americans[65], through mechanisms are yet insufficiently understood[66]. Variants of the KCNB1 gene in humans are associated with increases of waist to hip ratio, fasting insulin, and triglycerides, as well as decreased insulin sensitivity[67,68]. Mechanistically, the KCNB1-encoded Kv2.1 and other K^+v are important for the fine-tuning of the release of cellular insulin and other hormones or neurotransmitters, and have both inhibitory (through re-polarization of the membrane potential[69]) and stimulating (through interaction with the soluble N-ethylmaleimide-sensitive factor attachment protein receptor (SNARE) complex)[70-73] effects on exocytotic mechanisms. A previous study in rats suggested that treatment with lithium directly stimulates the expression of SNARE protein in brain tissue[74].

Genetic variations in genes regulating ERK and JNK expression were identified as additional contributors to the effects of MDD PGS on lithium response. Belonging to a family of protein kinases in the mitogen-activated protein kinases (MAPK) pathway, these molecules are highly

interactive with insulin-signaling mechanisms. Previous evidence has indicated that MAPK and insulin signaling could be activated by lithium, to enhance insulin-stimulated glucose transport and glycogen synthesis[75]. Additionally, lithium is known to stimulate MAPK-mediated neurite growth, neuronal survival, and neurogenesis[76], and regulate circadian rhythms[77]. Therefore, it is possible that variation in MAPK-associated genes interferes with these potentially therapeutic effects of lithium.

MicroRNAs (miRNAs) regulate messenger RNA (mRNA) translation in a sequence-specific manner and are emerging as critical regulators of central nervous system plasticity. We found that genetic effects on miR-144 and miR-451 expression could play a role in mediating lithium response in BD. Previous animal studies have shown that lithium treatment in vivo induces changes in miRNA expression, specifically miR-144[78]. It is possible that variations of miRNA genes influence their contribution to lithium's therapeutic mechanisms.

The main limitation of our study is that PGSs for MDD and DS explain only a small proportion of the variance in lithium treatment response (~1%), and have on their own no utility as clinical tests. Our cross-trait analysis provides a clue for a potential genetic overlap; however, no formal pleiotropy analyses were employed to confidently conclude about the effect of each genetic variant on the phenotypes tested. In addition, our pathway analysis findings are of an explorative nature and have not been validated on the transcript- or protein level or with experimental procedures in cellular models. Further, the current version of the Alda scale assesses only overall lithium efficacy but not effects specific to predominant illness polarity. Availability and incorporation of such information would have refined our results. The centrality of insulin-associated pathways in our findings could be a result of high representation of these genes within curated tools such as IPA®. However, these tools are powerful for hypothesis generation and

indicate plausible molecular targets to be tested. Since our sample size already detected significant effects, it is likely that in the future, an increased sample size will further improve the predictive power of PGSs[79].

In conclusion, we demonstrated that high genetic loadings for MDD and DS are predictive of unfavorable long-term response to lithium in patients with BD. Our study underscores the potential of PGS analysis to contribute to predictive models for medication response in psychiatry, and to uncover novel molecular pathways that drive these effects. While our findings, in isolation, are not yet ripe for clinical applications, they could serve as a component of multimodal predication models incorporating clinical and other biological data. The results of our study support clinical observations that have pointed to better lithium responsiveness in a BD subtype characterized by predominantly manic features. The study raises the possibility that mood-stabilizing- and anti-depressant properties of lithium are mediated through separate biological mechanisms.

Conflict of interest

All authors declare that they have no competing interests.

Acknowledgment

The authors are grateful to all patients who participated in the study and we appreciate the contributions of clinicians, scientists, research assistants and study staffs who have helped with patient recruitment, data collection and sample preparation for the studies. We are also indebted to the members of the ConLi^{+}Gen Scientific Advisory Board (http://www.conligen.org/) for critical input over the course of the project.

The analysis of this study was carried out using the high-performance computational capabilities of the University of Adelaide, Phoenix supercomputer https://www.adelaide.edu.au/phoenix/.

Funding

ATA received a Postgraduate Research Scholarship support from the University of Adelaide through the Adelaide Scholarship International (ASI) program. The primary sources of funding were the Deutsche Forschungsgemeinschaft (DFG; grant no.RI 908/7-1; grant FOR2107, RI 908/11-1 to Marcella Rietschel, NO 246/10-1 to Markus M. Nöthen, WI3429/3-1 to Stephanie H. Witt) and the Intramural Research Program of the National Institute of Mental Health (ZIA-MH00284311; ClinicalTrials.gov identifier: NCT00001174). The genotyping was in part funded by the German Federal Ministry of Education and Research (BMBF) through the Integrated Network IntegraMent (Integrated Understanding of Causes and Mechanisms in Mental Disorders), under the auspices of the e:Med Programme (grants awarded to Thomas G. Schulze, Marcella Rietschel, and Markus M. Nöthen).

Some data and biomaterials were collected as part of eleven projects (Study 40) that participated in the National Institute of Mental Health (NIMH) Bipolar Disorder Genetics Initiative. From 2003–2007, the Principal Investigators and Co-Investigators were: Indiana University, Indianapolis, IN, R01 MH59545, John Nurnberger, M.D., Ph.D., Marvin J. Miller, M.D., Elizabeth S. Bowman, M.D., N. Leela Rau, M.D., P.Ryan Moe, M.D., Nalini Samavedy, M.D., Rif El-Mallakh, M.D. (at University of Louisville), Husseini Manji, M.D.(at Johnson and Johnson), Debra A.Glitz, M.D.(at Wayne State University), Eric T. Meyer, Ph.D., M.S.(at Oxford University, UK), Carrie Smiley, R.N., Tatiana Foroud, Ph.D., Leah Flury, M.S., Danielle M.Dick, Ph.D (at Virginia Commonwealth University), Howard Edenberg, Ph.D.; Washington University, St. Louis, MO, R01 MH059534, John Rice, Ph.D, Theodore Reich, M.D., Allison Goate, Ph.D., Laura Bierut, M.D.K02 DA21237; Johns Hopkins University, Baltimore, M.D., R01 MH59533, Melvin McInnis, M.D., J.Raymond DePaulo, Jr., M.D., Dean F. MacKinnon, M.D., Francis M. Mondimore, M.D., James B. Potash, M.D., Peter P. Zandi, Ph.D, Dimitrios Avramopoulos, and Jennifer Payne; University of Pennsylvania, PA, R01 MH59553, Wade Berrettini, M.D., Ph.D.; University of California at San Francisco, CA, R01 MH60068, William Byerley, M.D., and Sophia Vinogradov, M.D.; University of Iowa, IA, R01 MH059548, William Coryell, M.D., and Raymond Crowe, M.D.; University of Chicago, IL, R01 MH59535, Elliot Gershon, M.D., Judith Badner, Ph.D., Francis McMahon, M.D., Chunyu Liu, Ph.D., Alan Sanders, M.D., Maria Caserta, Steven Dinwiddie, M.D., Tu Nguyen, Donna Harakal; University of California at San Diego, CA, R01 MH59567, John Kelsoe, M.D., Rebecca McKinney, B.A.; Rush University, IL, R01 MH059556, William Scheftner, M.D., Howard M. Kravitz, D.O., M.P.H., Diana Marta, B.S., Annette Vaughn-Brown, M.S.N., R.N., and Laurie Bederow, M.A.; NIMH Intramural Research Program, Bethesda, MD, 1Z01MH002810-01, Francis J. McMahon,

M.D., Layla Kassem, Psy.D., Sevilla Detera-Wadleigh, Ph.D, Lisa Austin, Ph.D, Dennis L. Murphy, M.D.; Howard University, William B. Lawson, M.D., Ph.D., Evarista Nwulia, M.D., and Maria Hipolito, M.D. This work was supported by the NIH grants P50CA89392 from the National Cancer Institute and 5K02DA021237 from the National Institute of Drug Abuse. The Canadian part of the study was supported by the Canadian Institutes of Health Research to MA grant #64410 to MA. Collection and phenotyping of the Australian UNSW sample, by Philip B. Mitchell, Peter R. Schofield, Janice M. Fullerton and Adam Wright, was funded by an Australian NHMRC Program Grant (No.1037196)), with personnel supported by NHMRC project grants (No. 1063960, 1066177) and the Janette Mary O'Neil Research Fellowship to JMF. The collection of the Barcelona sample was supported by the Centro de Investigación en Red de Salud Mental (CIBERSAM), IDIBAPS, the CERCA Programme / Generalitat de Catalunya, Miguel Servet II and Instituto de Salud Carlos III (grant numbers PI080247, PI1200906, PI12/00018, 2017 SGR 1577 and 2017 SGR 1365). Dr. Colom was funded by an unrestricted grant from CIBERSAM and a tenure-track grant Miguel Servet II, Instituto de Salud Carlos III (MSII14/00030). The Swedish Research Council, the Stockholm County Council, Karolinska Institutet and the Söderström-Königska Foundation supported this research through grants awarded to Lena Backlund, Louise Frisén, Catharina Lavebratt and Martin Schalling. The collection of the Geneva sample was supported by the Swiss National Foundation (grants Synapsy 51NF40-158776 and 32003B-125469). The work by the French group was supported by INSERM (Institut National de la Santé et de la Recherche Médicale); AP-HP (Assistance Publique des Hôpitaux de Paris); the Fondation FondaMental (RTRS Santé Mentale) and the labex Bio-PSY (Investissements d'Avenir program managed by the ANR under reference ANR-11-IDEX-0004-02). The German Research Foundation (DFG, grant FOR2107 DA1151/5-1 and

DA1151/5-2 to UD; SFB-TRR58, Project C09 and Z02 to UD), and the Interdisciplinary Center for Clinical Research (IZKF) of the Medical Faculty of the University of Münster (grant Dan3/012/17 to UD) supported this work. The collection of the Romanian sample was supported by U.E.F.I.S.C.D.I., Romania, grant awarded to Maria Grigoroiu-Serbanescu. The collection of the Czech sample was supported by the project Nr. LO1611 with a financial support from the MEYS under the NPU I program and by the Czech Science Foundation, grant Nr. 17-07070S.

Figures and Tables

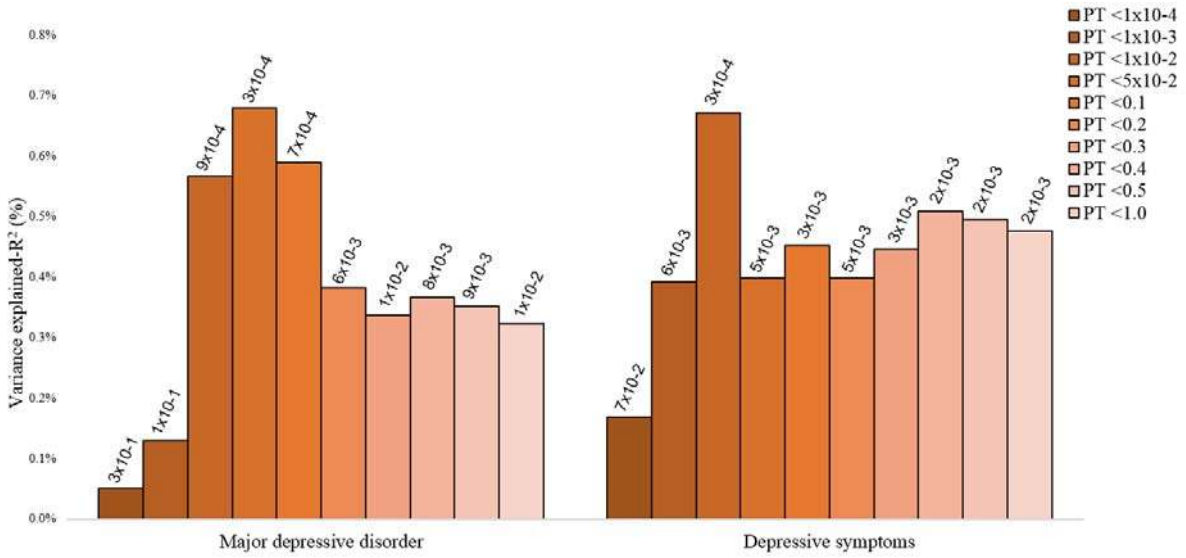


Figure 1: The association of PGS for depression traits (MDD and DS) and lithium treatment response at different GWAS p-value thresholds. The y-axis refers to the percentage of variance in treatment response to lithium accounted for by the PGSs for depressive traits at particular P-value thresholds. On the x-axis, are the GWAS P-value thresholds used to select single-nucleotide polymorphisms for the PGSs. On the top of each bar are the p-values for the association between the PGSs for depressive traits and lithium treatment response.

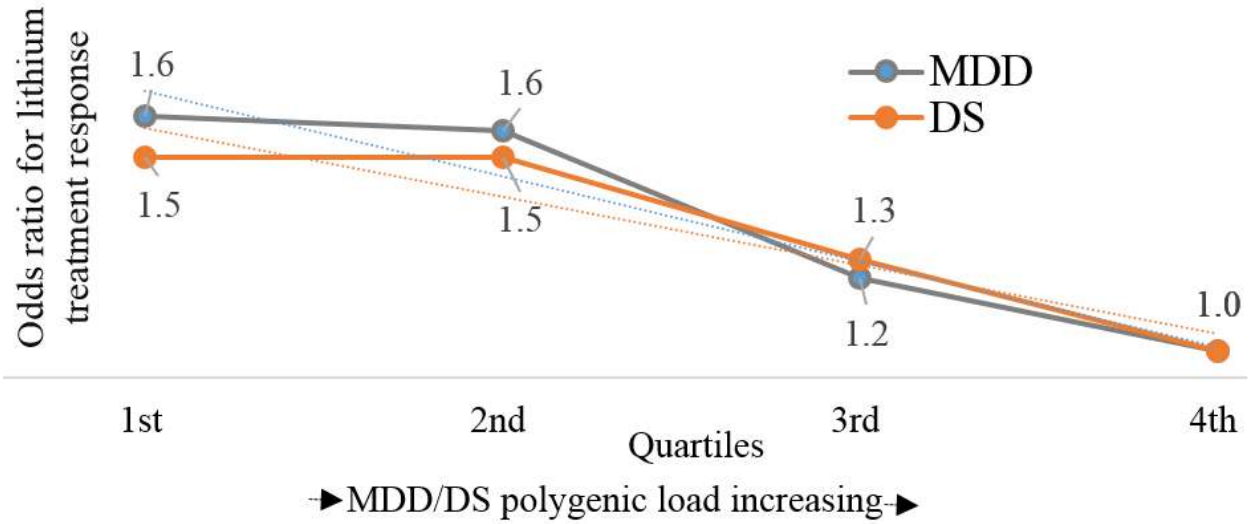


Figure 2: Odds ratios (ORs) for favorable treatment response to lithium for patients with BD in the low depression polygenic load quartiles (1st to 3rd), compared with patients in the highest depression polygenic load quartile (4th), estimated at the most significant p-value thresholds (n = 2586).

Table 1: Odds ratios of favorable lithium treatment response in patients with BD, comparing the response status of patients in the low PGS quartile for MDD and DS with patients with the highest polygenic load for MDD/DS (4th quartile).

Categories	Patients with BD (n=2,586)		
PGS quartiles	Responders/ Non-responders	unadjusted OR (95% CI)	¥Adjusted OR (95% CI)
MDD			
1st lowest score	197/449	1.56 (1.21-2.00)	1.64 (1.26-2.15)
2nd	202/444	1.62 (1.26-2.07)	1.60 (1.22-2.08)
3rd	163/485	1.19 (0.92-1.54)	1.20 (0.91-1.57)
4th highest score	142/504	1 (reference)	1 (reference)
DS			
1st lowest score	195/452	1.50 (1.17-1.93)	1.53 (1.18-2.00)
2nd	199/447	1.55 (1.21-1.99)	1.53 (1.18-1.99)
3rd	166/481	1.20 (0.93-1.55)	1.25 (0.96-1.64)
4th highest score	144/502	1 (reference)	1 (reference)

Legend: reference quartile (4th quartile) is the PGS category with the highest polygenic load for MDD/DS at the most significant threshold.

¥adjusted for age, sex, genotyping platform and 7-principal components. MDD: Major depressive disorder, DS: Depressive symptoms, PGS: polygenic score, OR: odds ratio

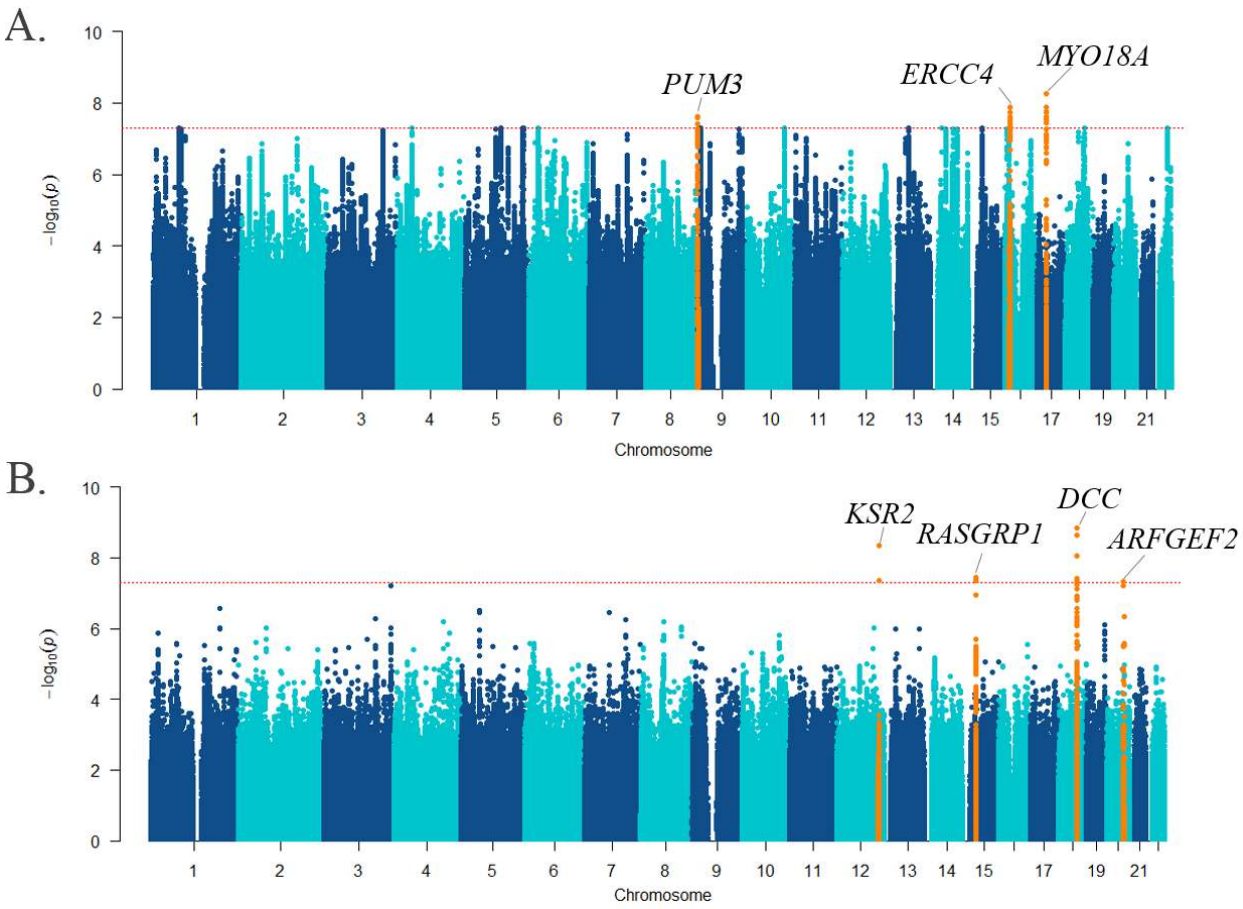


Figure 3: Manhattan plot of the cross-trait meta-analyses of GWASs on lithium treatment response and A) MDD; and B) DS. The loci that showed genome-wide significance are highlighted (orange), and their nearest genes indicated (top). The −log10 (cross-trait p-value) is plotted against the physical position of each SNP on each chromosome. The threshold for genome-wide significance (cross-trait p-value<$5x10^{-8}$) is indicated by the red dotted horizontal line.

Table 2: Loci resulting from a cross-trait meta-analysis of GWASs for lithium treatment response in patients with BD patients and GWAS for MDD and DS (cross-trait $p<5x10^{-8}$).

SNP	CHR	BP	A1	A2	GWAS P-value			Nearby gene
					MDD	**Lithium**	**Cross-trait**	
rs2327713	9	2974953	T	C	$3.19 x10^{-8}$	$3.81 x10^{-2}$	$2.35 x10^{-8}$	*PUM3*
rs7405404	16	13749859	T	C	$1.91 x10^{-7}$	$5.27 x10^{-3}$	$1.33 x10^{-8}$	*ERCC4*
rs11657502	17	27490977	A	G	$9.87 x10^{-7}$	$4.53 x10^{-4}$	$1.35 x10^{-8}$	*MYO18A*
					DS			
rs7134419	12	118383133	A	C	$9.39 x10^{-8}$	$5.64 x10^{-1}$	$4.34 x10^{-8}$	*KSR2*
rs59659806	15	38919964	T	C	$6.01 x10^{-7}$	$5.12 x10^{-2}$	$3.58 x10^{-8}$	*RASGRP1*
rs8099160	18	50752610	A	G	$2.68 x10^{-8}$	$4.67 x10^{-1}$	$1.42 x10^{-9}$	*DCC*
rs6066909	20	47529913	T	C	$9.23 x10^{-7}$	$2.21 x10^{-1}$	$4.66 x10^{-8}$	*ARFGEF2*

Legend: A1, effect allele; A2, alternative allele; CHR: chromosome; BP: position in base-pairs based on build 37; ED: Effect direction, +, increased susceptibility to MDD/DS or positive effect on lithium treatment response oriented to the effect allele (A1); -, decreased susceptibility to MDD/DS or negative effect on lithium treatment response oriented to the effect allele (A1); Nearest genes were based on RefSeq genes (build 37).

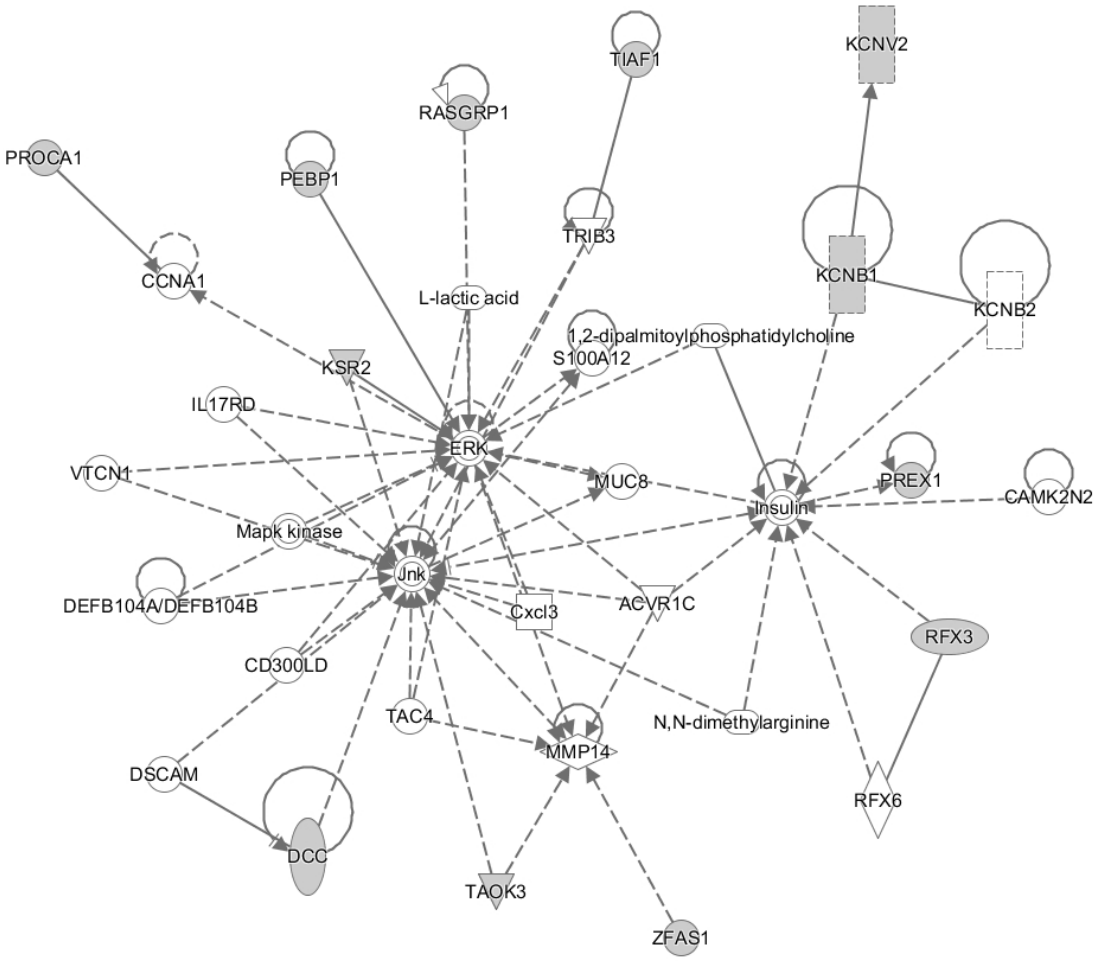
Table 3: Top upstream regulators for genes identified in the cross-trait meta-analyses

Upstream Regulator	Molecule type	P-value of overlap	Target molecules in dataset	Pathway for the regulator
KCNA1	ion channel	9.31X10-4	KCNB1	Voltage-gated potassium channel (function)
leucine-2-alanine enkephalin	chemical drug	2.79X10-3	KCNB1	opioid peptide (selective delta opioid receptor agonist) with analgesic properties
rubitecan	chemical drug	3.72X10-3	PEBP1	A semisynthetic agent with antitumor and antiviral properties
CAMKK2	kinase	7.43X10-3	KCNB1	AMPK Signaling; Calcium Signaling; Dopamine-DARPP32 Feedback in cAMP Signaling
EGR2	transcription regulator	8.69X10-3	FLOT2, TAOK1	Adipogenesis pathway
DAB1	other	8.35X10-3	DCC	Reelin Signaling in Neurons

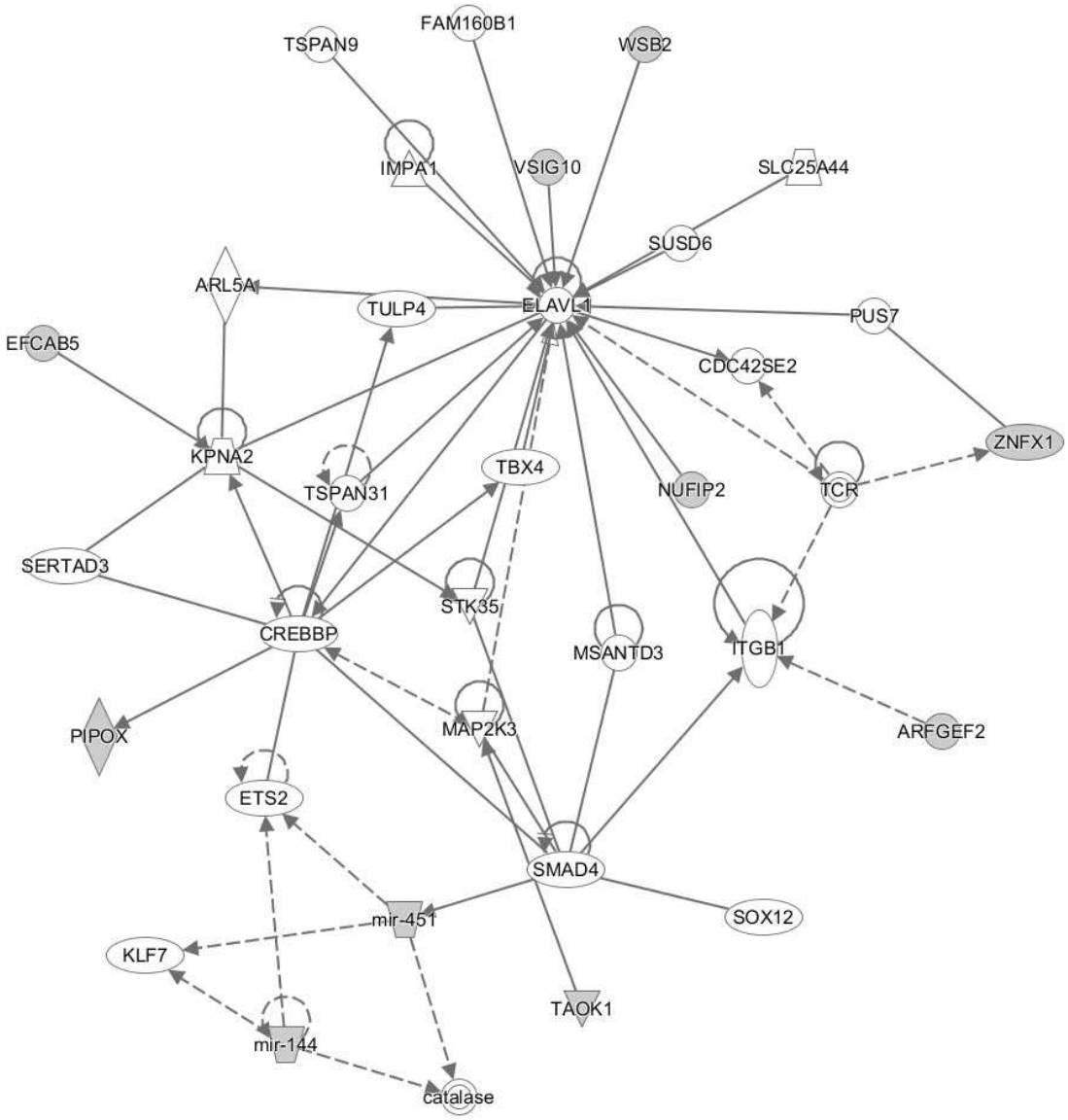
Legend: [a]P-values were adjusted by Benjamini & Hochberg (BH) method[80].The p-value of overlap was determined at BH adjusted p-value ≤0.01.

eFigure 1: Top IPA® functional network of genes associated with lithium response in BD and MDD/depressive symptoms.

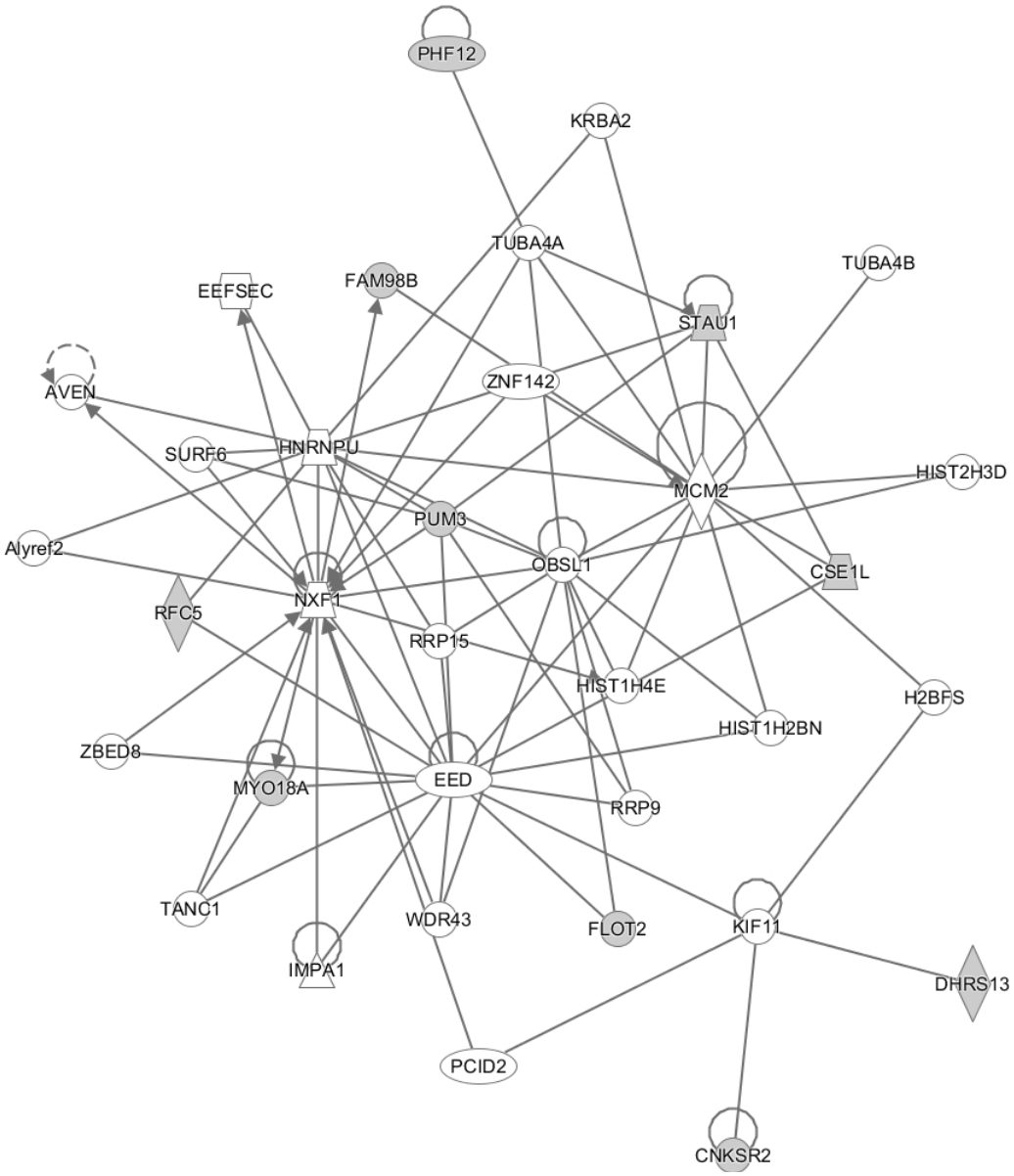
A) Network-1



B) Network-2



C) Network-3



eFigure 1 legend: IPA generates the network using a proprietary algorithm, and included genes that could contribute to the network, even if they were not contained in the original dataset.

eTable 1: Combined list of genes used for an input in the Ingenuity Pathway Analysis (IPA).

Hosting gene hGenes	eQTL genes reported in			All combined
	Westra	BRAINEAC	GTEx	
ARFGEF2	*CSE1L*	*STAU1*	*KCNB1*	*ARFGEF2*
C15orf53	*ZNFX1*	*ZNFX1*	*KCNB1*	*C15orf53*
CRYBA1	*NUFIP2*	*PYY2*	*KCNB1*	*CRYBA1*
CSE1L	*C15orf53*	*MYO18A*	*PROCA1*	*CSE1L*
DCC	*STAU1*	*EFCAB5*	*ZFAS1*	*DCC*
DHRS13	*ARFGEF2*	*TIAF1*	*SNORD12B*	*DHRS13*
ERAL1			*STAU1*	*EFCAB5*
FAM222B			*TIAF1*	*ERAL1*
FAM98B			*TIAF1*	*FAM222B*
FLOT2			*TIAF1*	*FAM98B*
KCNV2			*SNORD12B*	*FLOT2*
KSR2				*KCNB1*
MIR144				*KCNV2*
MIR451A				*KSR2*
MYO18A				*CNKSR2*
NUFIP2				*MIR144*
PEBP1				*MIR451A*
PHF12				*MYO18A*
PIPOX				*NUFIP2*
PREX1				*PEBP1*
PUM3				*PHF12*
RASGRP1				*PIPOX*
RFC5				*PREX1*
RFX3				*PROCA1*
SEZ6				*PUM3*
STAU1				*PYY2*
TAOK1				*RASGRP1*
TAOK3				*RFC5*
TIAF1				*RFX3*
VSIG10				*SEZ6*
WSB2				*SNORD12B*
				STAU1
				TAOK1
				TAOK3
				TIAF1
				VSIG10

	WSB2
	ZFAS1
	ZNFX1

eTable 2: Molecules within the three-top significant functional networks

Molecules in Network	Score	Focus Molecules
1, 2-dipalmitoylphosphatidylcholine, ACVR1C, CAMK2N2, CCNA1, CD300LD, Cxcl3, DCC, DEFB104A/DEFB104B, DSCAM, ERK, IL17RD, Insulin, Jnk, KCNB1**, KCNB2,** KCNV2**,** KSR2**, L-lactic acid, Mapk kinase, MMP14, MUC8, N, N-dimethylarginine,** PEBP1, PREX1, PROCA1, RASGRP1, RFX3, **RFX6, S100A12, TAC4,** TAOK3, TIAF1**, TRIB3, VTCN1,** ZFAS1	27	12
ARFGEF2**, ARL5A, catalase, CDC42SE2, CREBBP,** EFCAB5**, ELAVL1, ETS2, FAM160B1, IMPA1, ITGB1, KLF7, KPNA2, MAP2K3,** mir-144, mir-451**, MSANTD3,** NUFIP2, PIPOX**, PUS7, SERTAD3, SLC25A44, SMAD4, SOX12, STK35, SUSD6,** TAOK1**, TBX4, TCR, TSPAN9, TSPAN31, TULP4,** VSIG10**,** WSB2, ZNFX1	21	10
Alyref2, AVEN, CNKSR2**,** CSE1L, DHRS13**, EED, EEFSEC,** FAM98B, FLOT2**, H2BFS, HIST1H2BN, HIST1H4E, HIST2H3D, HNRNPU, IMPA1, KIF11, KRBA2, MCM2,** MYO18A**, NXF1, OBSL1, PCID2,** PHF12, PUM3, RFC5**, RRP9, RRP15,** STAU1, **SURF6, TANC1, TUBA4A, TUBA4B, WDR43, ZBED8, ZNF142**	21	10

Legend: The molecules represented in **bold** are derived from the cross-trait meta-GWAS (Table 1) and post-GWAS analysis (eTables). The p-score is calculated by IPA, and estimates, the probability of finding eleven (group 1) or ten (group 2) or more focus molecules in a network of 35 molecules randomly selected from IPA's Global Molecular Network. The p-score = –log10 (p-value); the p-value is calculated by Fisher's exact test.

eTable 3: The association of PGS for depression traits (MDD and DS) and lithium treatment response in patients with BD type 1 versus BD type 2 at different GWAS p-value thresholds (P_T).

MDD PGS		**BD type 1**		**BD type 2**	
P_T	No of SNPs	P-value	R^2 (%)	P-value	R^2 (%)
P_T <1x10^{-4}	852	1.83x10^{-1}	0.1%	0.43	0.2%
P_T <1x10^{-3}	2987	1.09x10^{-1}	0.2%	0.98	0.0%
P_T <1x10^{-2}	11892	4.34x10^{-4}	0.8%	0.85	0.0%
P_T <5x10^{-2}	31712	2.94x10^{-5}	1.2%	0.50	0.1%
P_T <0.1	47716	8.86x10^{-5}	1.0%	0.50	0.1%
P_T <0.2	70784	1.08x10^{-3}	0.7%	0.39	0.2%
P_T <0.3	88552	2.61x10^{-3}	0.6%	0.40	0.2%
P_T <0.4	102503	1.33x10^{-3}	0.7%	0.33	0.2%
P_T <0.5	114071	1.72x10^{-3}	0.6%	0.35	0.2%
P_T <1.0	147596	2.97x10^{-3}	0.6%	0.37	0.2%
DS PGS		**BD type 1**		**BD type 2**	
P_T	No of SNPs	P-value	R^2 (%)	P-value	R^2 (%)
P_T <1x10^{-4}	229	4.08x10^{-2}	0.3%	2.67x10^{-1}	0.3%
P_T <1x10^{-3}	1352	1.18x10^{-2}	0.4%	1.08x10^{-1}	0.7%
P_T <1x10^{-2}	8057	7.26x10^{-3}	0.5%	6.26x10^{-2}	0.9%
P_T <5x10^{-2}	25918	5.51x10^{-2}	0.2%	4.68x10^{-2}	1.0%
P_T <0.1	41865	1.46x10^{-1}	0.1%	4.86x10^{-2}	1.0%
P_T <0.2	65634	1.54x10^{-1}	0.1%	5.51x10^{-2}	0.9%
P_T <0.3	83845	1.99x10^{-1}	0.1%	5.48x10^{-2}	0.9%
P_T <0.4	98217	1.61x10^{-1}	0.1%	5.16x10^{-2}	1.0%
P_T <0.5	109888	1.95x10^{-1}	0.1%	4.96x10^{-2}	1.0%
P_T <1.0	143345	2.16x10^{-1}	0.1%	4.42x10^{-2}	1.0%

References

1. Ferrari AJ, Stockings E, Khoo JP, et al. The prevalence and burden of bipolar disorder: findings from the Global Burden of Disease Study 2013. *Bipolar disorders* 2016; **18**(5): 440-50.
2. Chesney E, Goodwin GM, Fazel S. Risks of all-cause and suicide mortality in mental disorders: a meta-review. *World Psychiatry* 2014; **13**(2): 153-60.
3. Schulze TG, Alda M, Adli M, et al. The International Consortium on Lithium Genetics (ConLiGen): An Initiative by the NIMH and IGSLI to Study the Genetic Basis of Response to Lithium Treatment. *Neuropsychobiology* 2010; **62**(1): 72-8.
4. Malhi GS, Bassett D, Boyce P, et al. Royal Australian and New Zealand College of Psychiatrists clinical practice guidelines for mood disorders. *Aust N Z J Psychiatry* 2015; **49**(12): 1087-206.
5. Hou L, Heilbronner U, Degenhardt F, et al. Genetic variants associated with response to lithium treatment in bipolar disorder: a genome-wide association study. *The Lancet* 2016; **387**(10023): 1085-93.
6. Grof P, Duffy A, Cavazzoni P, et al. Is response to prophylactic lithium a familial trait? *The Journal of clinical psychiatry* 2002; **63**(10): 942-7.
7. Garnham J, Munro A, Slaney C, et al. Prophylactic treatment response in bipolar disorder: results of a naturalistic observation study. *J Affect Disord* 2007; **104**(1-3): 185-90.
8. Lahteenvuo M, Tanskanen A, Taipale H, et al. Real-World Effectiveness of Pharmacologic Treatments for the Prevention of Rehospitalization in a Finnish Nationwide Cohort of Patients With Bipolar Disorder. *JAMA Psychiatry* 2018.
9. Grande I, Berk M, Birmaher B, Vieta E. Bipolar disorder. *Lancet* 2016; **387**(10027): 1561-72.
10. Bauer M, Gitlin M. Practical Management of Lithium. The Essential Guide to Lithium Treatment. Cham: Springer International Publishing; 2016: 113-28.
11. Oedegaard KJ, Alda M, Anand A, et al. The Pharmacogenomics of Bipolar Disorder study (PGBD): identification of genes for lithium response in a prospective sample. *BMC Psychiatry* 2016; **16**: 129.
12. Severus E, Taylor MJ, Sauer C, et al. Lithium for prevention of mood episodes in bipolar disorders: systematic review and meta-analysis. *Int J Bipolar Disord* 2014; **2**: 15.
13. Joas E, Karanti A, Song J, Goodwin GM, Lichtenstein P, Landen M. Pharmacological treatment and risk of psychiatric hospital admission in bipolar disorder. *Br J Psychiatry* 2017; **210**(3): 197-202.
14. Chen CH, Lee CS, Lee MT, et al. Variant GADL1 and response to lithium therapy in bipolar I disorder. *The New England journal of medicine* 2014; **370**(2): 119-28.
15. Song J, Bergen SE, Di Florio A, et al. Genome-wide association study identifies SESTD1 as a novel risk gene for lithium-responsive bipolar disorder. *Mol Psychiatry* 2016; **21**(9): 1290-7.
16. Amare AT, Schubert KO, Hou L, et al. Association of Polygenic Score for Schizophrenia and HLA Antigen and Inflammation Genes With Response to Lithium in Bipolar Affective Disorder: A Genome-Wide Association Study. *JAMA psychiatry* 2017.
17. Goldberg JF, Garno JL, Leon AC, Kocsis JH, Portera L. A history of substance abuse complicates remission from acute mania in bipolar disorder. *The Journal of clinical psychiatry* 1999; **60**(11): 733-40.
18. Calkin C, van de Velde C, Ruzickova M, et al. Can body mass index help predict outcome in patients with bipolar disorder? *Bipolar disorders* 2009; **11**(6): 650-6.

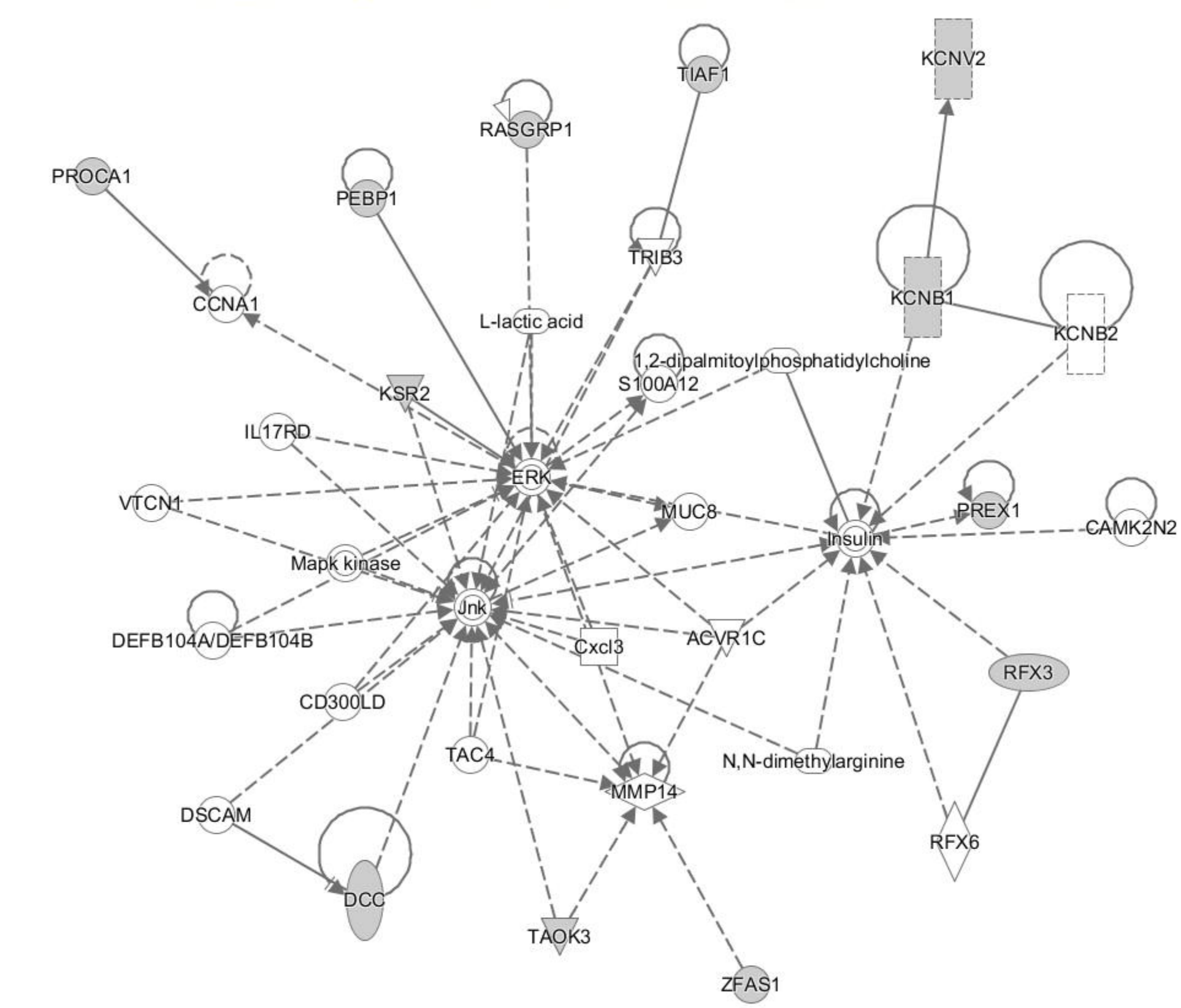
19. Calkin CV, Ruzickova M, Uher R, et al. Insulin resistance and outcome in bipolar disorder. *The British Journal of Psychiatry* 2015; **206**(1): 52-7.
20. Mitchell PB, Frankland A, Hadzi-Pavlovic D, et al. Comparison of depressive episodes in bipolar disorder and in major depressive disorder within bipolar disorder pedigrees. *The British Journal of Psychiatry* 2011; **199**(4): 303-9.
21. Cross-Disorder Group of the Psychiatric Genomics C, Lee SH, Ripke S, et al. Genetic relationship between five psychiatric disorders estimated from genome-wide SNPs. *Nat Genet* 2013; **45**(9): 984-94.
22. Cross-Disorder Group of the Psychiatric Genomics C. Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *The Lancet* 2013; **381**(9875): 1371-9.
23. Amare AT, Schubert KO, Klingler-Hoffmann M, Cohen-Woods S, Baune BT. The genetic overlap between mood disorders and cardiometabolic diseases: a systematic review of genome wide and candidate gene studies. *Transl Psychiatry* 2017; **7**(1): e1007.
24. Abou-Saleh MT, Muller-Oerlinghausen B, Coppen AJ. Lithium in the episode and suicide prophylaxis and in augmenting strategies in patients with unipolar depression. *Int J Bipolar Disord* 2017; **5**(1): 11.
25. Zhou X, Ravindran AV, Qin B, et al. Comparative efficacy, acceptability, and tolerability of augmentation agents in treatment-resistant depression: systematic review and network meta-analysis. *The Journal of clinical psychiatry* 2015; **76**(4): e487-98.
26. Tiihonen J, Tanskanen A, Hoti F, et al. Pharmacological treatments and risk of readmission to hospital for unipolar depression in Finland: a nationwide cohort study. *Lancet Psychiatry* 2017; **4**(7): 547-53.
27. Popovic D, Reinares M, Goikolea JM, Bonnin CM, Gonzalez-Pinto A, Vieta E. Polarity index of pharmacological agents used for maintenance treatment of bipolar disorder. *Eur Neuropsychopharmacol* 2012; **22**(5): 339-46.
28. Vieta E, Berk M, Schulze TG, et al. Bipolar disorders. *Nat Rev Dis Primers* 2018; **4**: 18008.
29. Colom F, Vieta E, Daban C, Pacchiarotti I, Sanchez-Moreno J. Clinical and therapeutic implications of predominant polarity in bipolar disorder. *J Affect Disord* 2006; **93**(1-3): 13-7.
30. Kessing LV, Hellmund G, Andersen PK. Predictors of excellent response to lithium: results from a nationwide register-based study. *Int Clin Psychopharmacol* 2011; **26**(6): 323-8.
31. Kleindienst N, Engel R, Greil W. Which clinical factors predict response to prophylactic lithium? A systematic review for bipolar disorders. *Bipolar disorders* 2005; **7**(5): 404-17.
32. Pfennig A, Schlattmann P, Alda M, et al. Influence of atypical features on the quality of prophylactic effectiveness of long-term lithium treatment in bipolar disorders. *Bipolar disorders* 2010; **12**(4): 390-6.
33. Fountoulakis KN, Kontis D, Gonda X, Siamouli M, Yatham LN. Treatment of mixed bipolar states. *Int J Neuropsychopharmacol* 2012; **15**(7): 1015-26.
34. Sportiche S, Geoffroy PA, Brichant-Petitjean C, et al. Clinical factors associated with lithium response in bipolar disorders. *Aust N Z J Psychiatry* 2017; **51**(5): 524-30.
35. Wray NR, Sullivan PF. Genome-wide association analyses identify 44 risk variants and refine the genetic architecture of major depression. *bioRxiv* 2017.
36. Okbay A, Baselmans BML, De Neve J-E, et al. Genetic variants associated with subjective well-being, depressive symptoms and neuroticism identified through genome-wide analyses. *Nature genetics* 2016; **48**(6): 624-33.

37. Duffy A, Alda M, Milin R, Grof P. A consecutive series of treated affected offspring of parents with bipolar disorder: is response associated with the clinical profile? *Can J Psychiatry* 2007; **52**(6): 369-76.
38. Manchia M, Adli M, Akula N, et al. Assessment of Response to Lithium Maintenance Treatment in Bipolar Disorder: A Consortium on Lithium Genetics (ConLiGen) Report. *PloS one* 2013; **8**(6): e65636.
39. Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007; **81**(3): 559-75.
40. Das S, Forer L, Schonherr S, et al. Next-generation genotype imputation service and methods. *Nat Genet* 2016; **48**(10): 1284-7.
41. Purcell SM, Wray NR, Stone JL, et al. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature* 2009; **460**(7256): 748-52.
42. Yang Q, Wu H, Guo CY, Fox CS. Analyze multivariate phenotypes in genetic association studies by combining univariate association tests. *Genet Epidemiol* 2010; **34**(5): 444-54.
43. Yang Q, Wang Y. Methods for Analyzing Multivariate Phenotypes in Genetic Association Studies. *J Probab Stat* 2012; **2012**: 652569.
44. Malhi GS, Tanious M, Das P, Berk M. The science and practice of lithium therapy. *Aust N Z J Psychiatry* 2012; **46**(3): 192-211.
45. Charney AW, Ruderfer DM, Stahl EA, et al. Evidence for genetic heterogeneity between clinical subtypes of bipolar disorder. *Transl Psychiatry* 2017; **7**(1): e993.
46. Song J, Kuja-Halkola R, Sjolander A, et al. Specificity in Etiology of Subtypes of Bipolar Disorder: Evidence From a Swedish Population-Based Family Study. *Biological psychiatry* 2017.
47. Kan C, Pedersen NL, Christensen K, et al. Genetic overlap between type 2 diabetes and depression in Swedish and Danish twin registries. *Molecular Psychiatry* 2016; **21**: 903.
48. Bschor T. Lithium in the Treatment of Major Depressive Disorder. *Drugs* 2014; **74**(8): 855-62.
49. Alevizos B, Alevizos E, Leonardou A, Zervas I. Low dosage lithium augmentation in venlafaxine resistant depression: an open-label study. *Psychiatrike = Psychiatriki* 2012; **23**(2): 143-8.
50. Bauer M, Döpfmer S. Lithium augmentation in treatment-resistant depression: Meta-analysis of placebo-controlled studies. *Prim Care Companion J Clin Psych* 2000; **2**(1): 31.
51. Bauer M, Adli M, Ricken R, Severus E, Pilhatsch M. Role of lithium augmentation in the management of major depressive disorder. *CNS drugs* 2014; **28**(4): 331-42.
52. Bauer M, Bschor T, Kunz D, Berghofer A, Strohle A, Muller-Oerlinghausen B. Double-blind, placebo-controlled trial of the use of lithium to augment antidepressant medication in continuation treatment of unipolar major depression. *Am J Psychiatry* 2000; **157**(9): 1429-35.
53. Bauer M, Adli M, Baethge C, et al. Lithium Augmentation Therapy in Refractory Depression: Clinical Evidence and Neurobiological Mechanisms. *Can J Psychiatry* 2003; **48**(7): 440-8.
54. Bschor T, Bauer M. Efficacy and mechanisms of action of Lithium augmentation in refractory major depression. *Curr Pharm Des* 2006; **12**(23): 2985-92.
55. O'Leary NA, Wright MW, Brister JR, et al. Reference sequence (RefSeq) database at NCBI: current status, taxonomic expansion, and functional annotation. *Nucleic acids research* 2016; **44**(D1): D733-45.

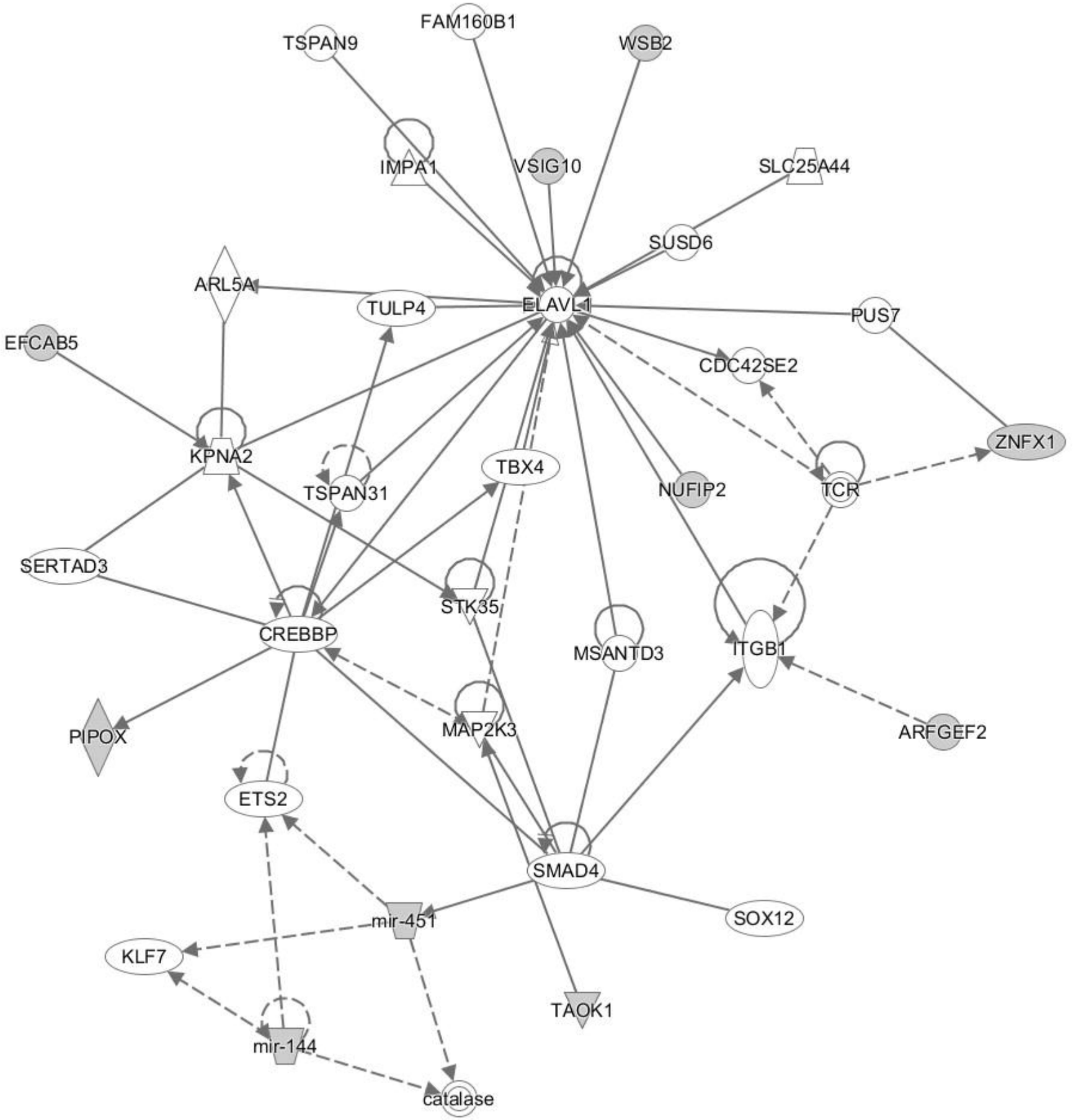
56. Harter PN, Bunz B, Dietz K, Hoffmann K, Meyermann R, Mittelbronn M. Spatio-temporal deleted in colorectal cancer (DCC) and netrin-1 expression in human foetal brain development. *Neuropathology and applied neurobiology* 2010; **36**(7): 623-35.
57. Watanabe H, Nagata E, Kosakai A, et al. Disruption of the epilepsy KCNQ2 gene results in neural hyperexcitability. *J Neurochem* 2000; **75**(1): 28-33.
58. Zhang X, Bertaso F, Yoo JW, et al. Deletion of the potassium channel Kv12.2 causes hippocampal hyperexcitability and epilepsy. *Nature neuroscience* 2010; **13**(9): 1056-8.
59. Mertens J, Wang QW, Kim Y, et al. Differential responses to lithium in hyperexcitable neurons from patients with bipolar disorder. *Nature* 2015; **527**(7576): 95-9.
60. Stern S, Santos R, Marchetto MC, et al. Neurons derived from patients with bipolar disorder divide into intrinsically different sub-populations of neurons, predicting the patients' responsiveness to lithium. *Mol Psychiatry* 2017.
61. Ait-Lounis A, Bonal C, Seguin-Estevez Q, et al. The transcription factor Rfx3 regulates beta-cell differentiation, function, and glucokinase expression. *Diabetes* 2010; **59**(7): 1674-85.
62. Stein MB, McCarthy MJ, Chen CY, et al. Genome-wide analysis of insomnia disorder. *Mol Psychiatry* 2018.
63. Gold AK, Sylvia LG. The role of sleep in bipolar disorder. *Nature and Science of Sleep* 2016; **8**: 207-14.
64. Szklo-Coxe M, Young T, Peppard PE, Finn LA, Benca RM. Prospective associations of insomnia markers and symptoms with depression. *Am J Epidemiol* 2010; **171**(6): 709-20.
65. Lewis JP, Palmer ND, Ellington JB, et al. Analysis of candidate genes on chromosome 20q12-13.1 reveals evidence for BMI mediated association of PREX1 with type 2 diabetes in European Americans. *Genomics* 2010; **96**(4): 211-9.
66. Barrows D, He JZ, Parsons R. PREX1 Protein Function Is Negatively Regulated Downstream of Receptor Tyrosine Kinase Activation by p21-activated Kinases (PAKs). *J Biol Chem* 2016; **291**(38): 20042-54.
67. Yu Y, Wang J, Kang R, et al. Association of KCNB1 polymorphisms with lipid metabolisms and insulin resistance: a case-control design of population-based cross-sectional study in Chinese Han population. *Lipids Health Dis* 2015; **14**: 112.
68. Zhang YX, Liu Y, Dong J, et al. An exploratory study of the association between KCNB1 rs1051295 and type 2 diabetes and its related traits in Chinese Han population. *PloS one* 2013; **8**(2): e56365.
69. Dodson PD, Forsythe ID. Presynaptic K+ channels: electrifying regulators of synaptic terminal excitability. *Trends Neurosci* 2004; **27**(4): 210-7.
70. Singer-Lahat D, Sheinin A, Chikvashvili D, et al. K+ channel facilitation of exocytosis by dynamic interaction with syntaxin. *J Neurosci* 2007; **27**(7): 1651-8.
71. Feinshreiber L, Singer-Lahat D, Ashery U, Lotan I. Voltage-gated potassium channel as a facilitator of exocytosis. *Ann N Y Acad Sci* 2009; **1152**: 87-92.
72. Feinshreiber L, Singer-Lahat D, Friedrich R, et al. Non-conducting function of the Kv2.1 channel enables it to recruit vesicles for release in neuroendocrine and nerve cells. *J Cell Sci* 2010; **123**(Pt 11): 1940-7.
73. Dai XQ, Manning Fox JE, Chikvashvili D, et al. The voltage-dependent potassium channel subunit Kv2.1 regulates insulin secretion from rodent and human islets independently of its electrical function. *Diabetologia* 2012; **55**(6): 1709-20.
74. Carlson SW, Yan H, Dixon CE. Lithium increases hippocampal SNARE protein abundance after traumatic brain injury. *Exp Neurol* 2017; **289**: 55-63.

75. Macko AR, Beneze AN, Teachey MK, Henriksen EJ. Roles of insulin signalling and p38 MAPK in the activation by lithium of glucose transport in insulin-resistant rat skeletal muscle. *Archives of physiology and biochemistry* 2008; **114**(5): 331-9.
76. Engel SR, Creson TK, Hao Y, et al. The extracellular signal-regulated kinase pathway contributes to the control of behavioral excitement. *Mol Psychiatry* 2009; **14**(4): 448-61.
77. McCarthy MJ, Le Roux MJ, Wei H, Beesley S, Kelsoe JR, Welsh DK. Calcium channel genes associated with bipolar disorder modulate lithium's amplification of circadian rhythms. *Neuropharmacology* 2016; **101**: 439-48.
78. Zhou R, Yuan P, Wang Y, et al. Evidence for selective microRNAs and their effectors as common long-term targets for the actions of mood stabilizers. *Neuropsychopharmacology* 2009; **34**(6): 1395-405.
79. Wray NR, Lee SH, Mehta D, Vinkhuyzen AA, Dudbridge F, Middeldorp CM. Research review: Polygenic methods and their application to psychiatric traits. *Journal of child psychology and psychiatry, and allied disciplines* 2014; **55**(10): 1068-87.
80. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B (Methodological)* 1995; **57**(1): 289-300.

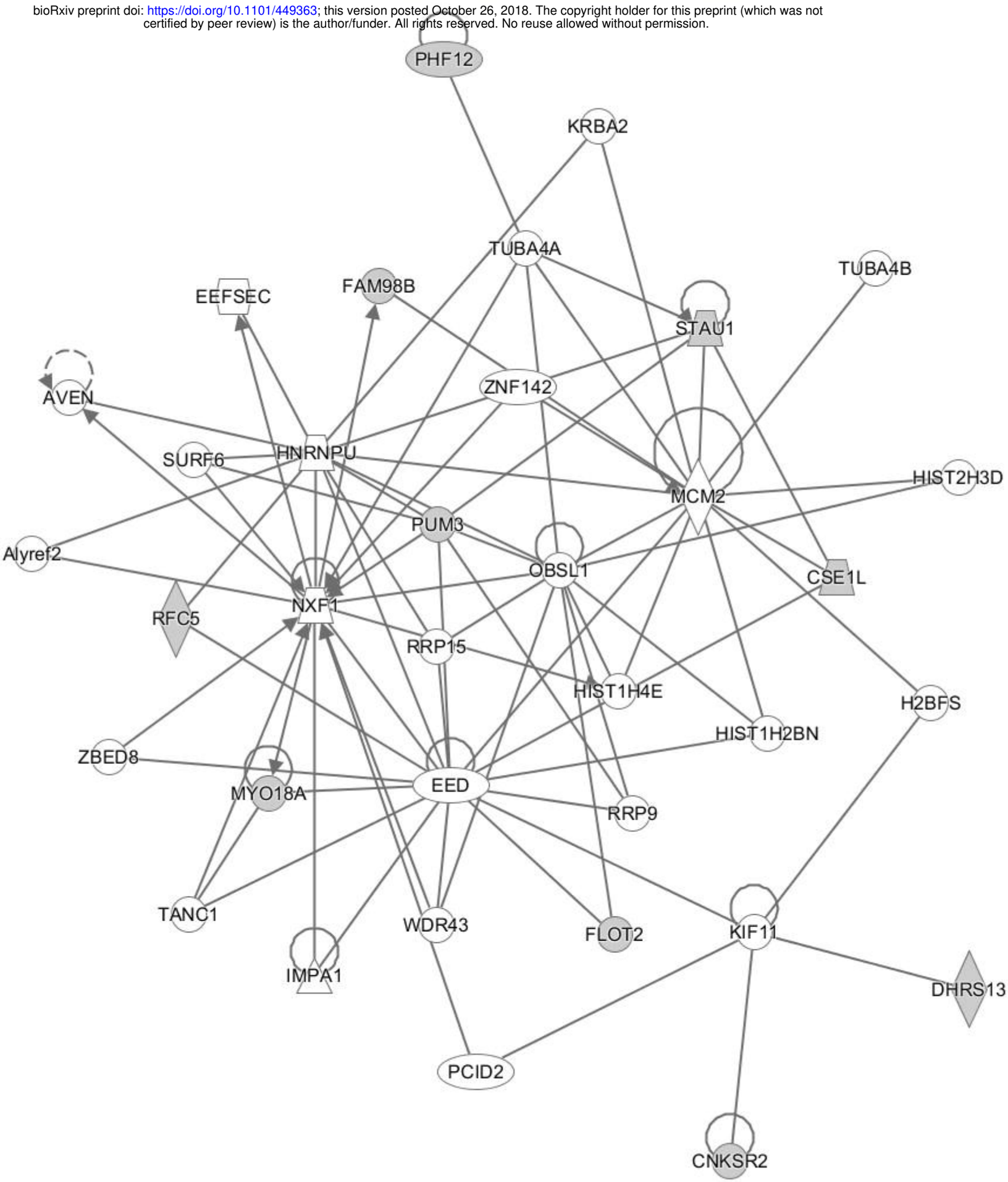
Network 1 : IPA_genes_ALL_Depression_Lithium : Depression_Lithium : IPA_genes_ALL_Depression_Lithium

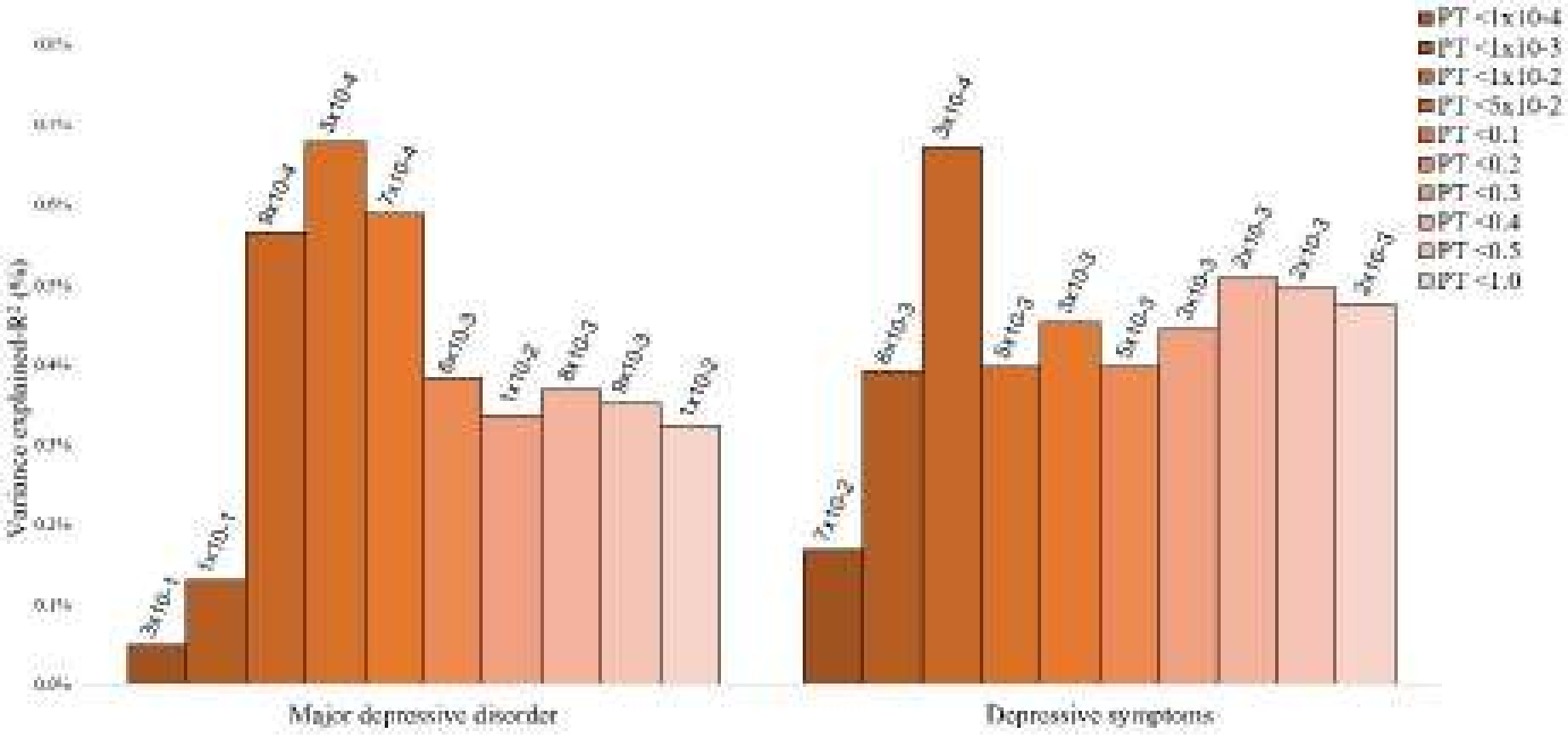


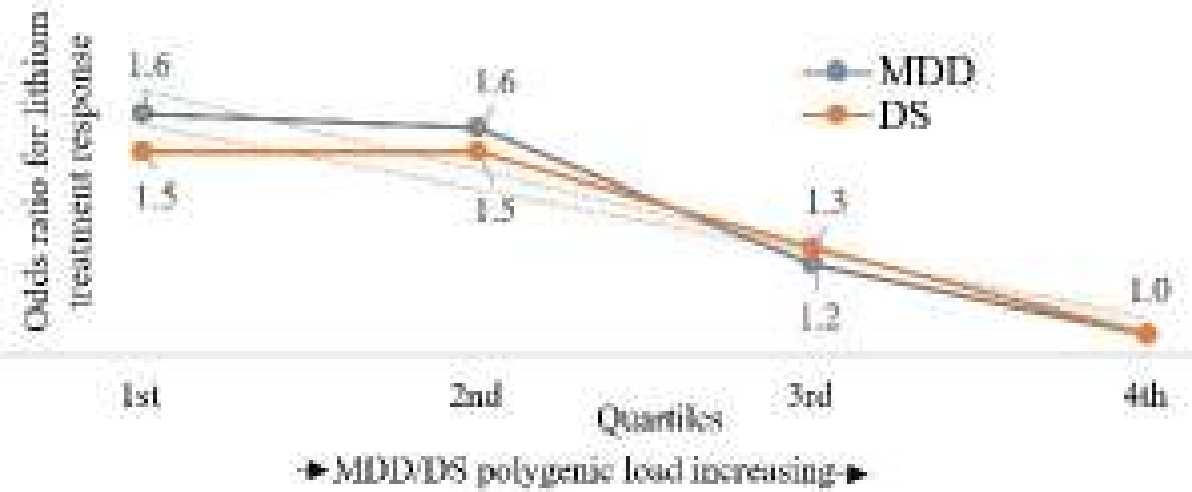
Network 2 : IPA_genes_ALL_Depression_Lithium : Depression_Lithium : IPA_genes_ALL_Depression_Lithium

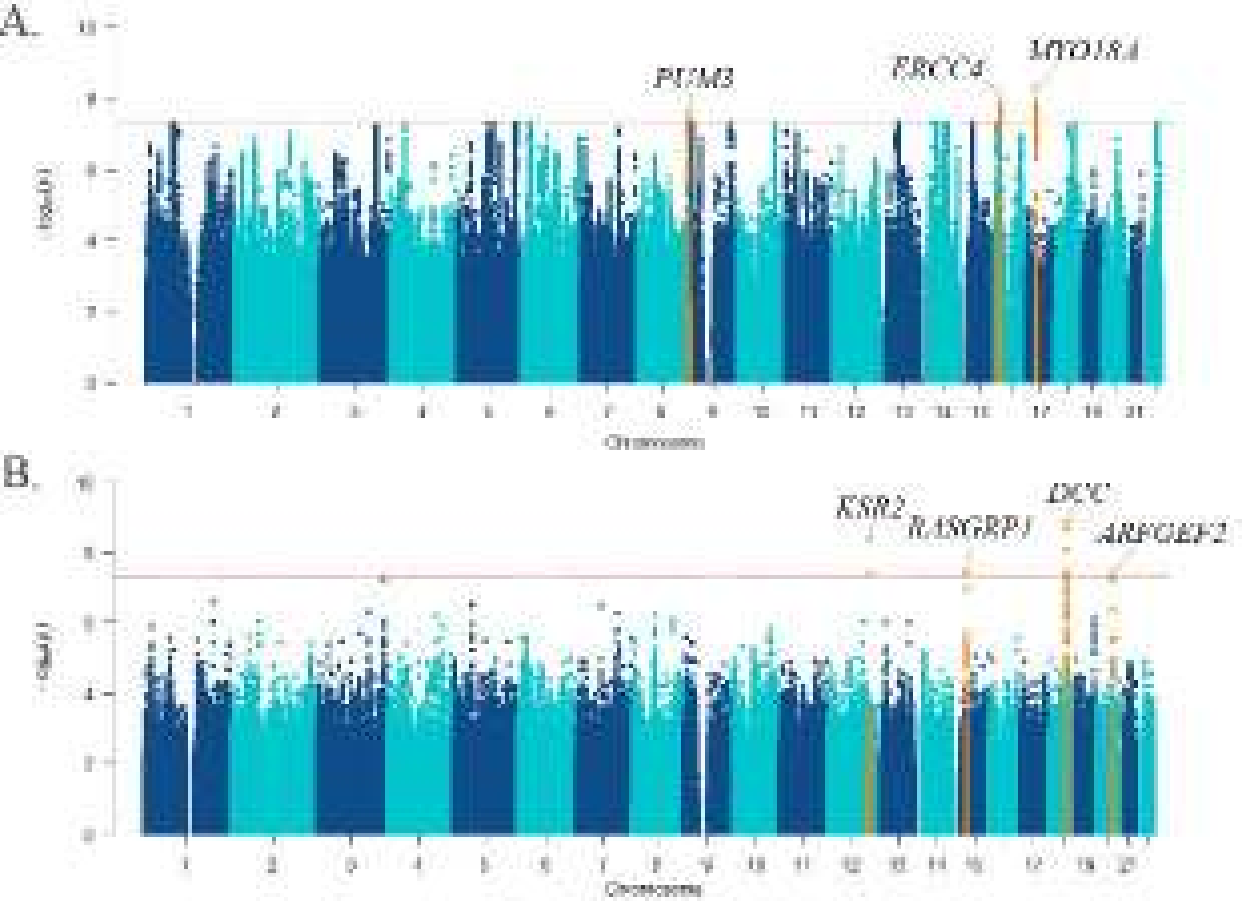


Network 3 : IPA_genes_ALL_Depression_Lithium : Depression_Lithium : IPA_genes_ALL_Depression_Lithium




Variance explained R^2 (%)
0.0%
0.1%
0.2%
0.3%
0.4%
0.5%
0.6%
0.7%
0.8%
PT <1x10-4
PT <1x10-3
PT <1x10-2
PT <5x10-2
PT <0.1
PT <0.2
PT <0.3
PT <0.4
PT <0.5
PT <1.0
3x10-4
1x10-3
8x10-3
1x10-2
8x10-3
7x10-3
1x10-2
8x10-3
8x10-3
1x10-2
Major depressive disorder
7x10-3
3x10-2
8x10-3
1x10-2
8x10-3
1x10-2
8x10-3
1x10-2
2x10-2
2x10-2
2x10-2
Depressive symptoms


Odds ratio for lithium treatment response
1.6
1.6
1.5
1.5
1.3
1.2
1.0
MDD
DS
1st
2nd
3rd
4th
Quantiles
MDD/DS polygenic load increasing


A.
PIM3
ERCC4
MYO18A
B.
KSR2
RANGRP1
DCC
ARFGEF2