

Poor sleep is associated with CSF biomarkers of amyloid pathology in cognitively normal adults

Kate E. Sprecher, MS
Rebecca L. Kosciak, PhD
Cynthia M. Carlsson,
MD
Henrik Zetterberg, PhD,
MD
Kaj Blennow, PhD, MD
Ozioma C. Okonkwo,
PhD
Mark A. Sager, MD
Sanjay Asthana, MD
Sterling C. Johnson, PhD
Ruth M. Benca, MD,
PhD
Barbara B. Bendlin, PhD

Correspondence to
K.E. Sprecher:
ksprecher@wisc.edu

ABSTRACT

Objective: To determine the relationship between sleep quality and CSF markers of Alzheimer disease (AD) pathology in late midlife.

Methods: We investigated the relationship between sleep quality and CSF AD biomarkers in a cohort enriched for parental history of sporadic AD, the Wisconsin Registry for Alzheimer's Prevention. A total of 101 participants (mean age 62.9 ± 6.2 years, 65.3% female) completed sleep assessments and CSF collection and were cognitively normal. Sleep quality was measured with the Medical Outcomes Study Sleep Scale. CSF was assayed for biomarkers of amyloid metabolism and plaques (β -amyloid 42 [A β 42]), tau pathology (phosphorylated tau [p-tau] 181), neuronal/axonal degeneration (total tau [t-tau], neurofilament light [NFL]), neuroinflammation/astroglial activation (monocyte chemoattractant protein-1 [MCP-1], chitinase-3-like protein 1 [YKL-40]), and synaptic dysfunction/degeneration (neurogranin). To adjust for individual differences in total amyloid production, A β 42 was expressed relative to A β 40. To assess cumulative pathology, CSF biomarkers were expressed in ratio to A β 42. Relationships among sleep scores and CSF biomarkers were assessed with multiple regression, controlling for age, sex, time between sleep and CSF measurements, and CSF assay batch.

Results: Worse subjective sleep quality, more sleep problems, and daytime somnolence were associated with greater AD pathology, indicated by lower CSF A β 42/A β 40 and higher t-tau/A β 42, p-tau/A β 42, MCP-1/A β 42, and YKL-40/A β 42. There were no significant associations between sleep and NFL or neurogranin.

Conclusions: Self-report of poor sleep was associated with greater AD-related pathology in cognitively healthy adults at risk for AD. Effective strategies exist for improving sleep; therefore sleep health may be a tractable target for early intervention to attenuate AD pathogenesis.

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GLOSSARY

A β 42 = β -amyloid 42; **AD** = Alzheimer disease; **BMI** = body mass index; **ESS** = Epworth Sleepiness Scale; **MCP-1** = monocyte chemoattractant protein-1; **MOS** = Medical Outcomes Study; **NFL** = neurofilament light; **OSA** = obstructive sleep apnea; **p-tau** = phosphorylated tau; **PiB** = Pittsburgh compound B; **t-tau** = total tau; **WRAP** = Wisconsin Registry for Alzheimer's Prevention; **YKL-40** = chitinase-3-like protein 1.

To delay or prevent dementia due to Alzheimer disease (AD), it is critical to identify modifiable risk factors. Sleep quality is a promising target for intervention during the preclinical phase of AD, when pathogenesis has begun but cognition is still intact.¹ Sleep is associated with AD brain pathophysiology, including amyloid mis-metabolism and deposition, tau hyperphosphorylation and aggregation, and neuronal and synaptic dysfunction and degeneration. We previously found that self-report of poor sleep was associated with greater brain amyloid burden, as measured by PET with Pittsburgh compound B (PiB).² In young men, sleep deprivation diminished the diurnal fluctuation of CSF amyloid levels.³ In rodent models of AD, sleep restriction alters amyloid metabolism and tau phosphorylation,⁴⁻⁶ and tau-deficient mice show

From the Department of Medicine and Neuroscience Training Program (K.E.S.) and Wisconsin Alzheimer's Disease Research Center (C.M.C., O.C.O., M.A.S., S.A., S.C.J., B.B.B.), University of Wisconsin-Madison; Wisconsin Alzheimer's Institute (R.L.K., C.M.C., O.C.O., M.A.S., S.A., S.C.J., B.B.B.); Geriatric Research Education and Clinical Center (C.M.C., O.C.O., S.A., S.C.J., B.B.B.), Wm. S. Middleton Veterans Hospital, Madison, WI; Institute of Neuroscience and Physiology (H.Z., K.B.), University of Gothenburg; Clinical Neurochemistry Laboratory (H.Z., K.B.), Sahlgrenska University Hospital, Mölndal, Sweden; Department of Molecular Neuroscience (H.Z.), UCL Institute of Neurology, Queen Square, London, UK; and Department of Psychiatry and Human Behavior (R.M.B.), University of California, Irvine.

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disturbed sleep architecture.⁷ Maintenance of axons⁸ and synapses occurs during sleep,⁹ and sleep loss elevates inflammation and microglial activation.¹⁰

Preclinical AD brain pathology can be detected via CSF biomarkers; however, little is known of their relationship with sleep. CSF β -amyloid 42 (A β 42, a marker of amyloid deposition) may be associated with sleep-disordered breathing¹¹ and sleep fragmentation.¹² However, less is known about other markers of AD pathophysiology. Furthermore, CSF biomarkers combined in ratios have superior diagnostic and prognostic power compared to single biomarkers.^{13,14} Therefore we examined the relationship between self-reported sleep and CSF biomarkers of amyloid deposition and plaque formation (A β 42), tau phosphorylation state and tau pathology (phosphorylated tau [p-tau]), axonal degeneration (total tau [t-tau], neurofilament light [NFL]), neuroinflammation (monocyte chemoattractant protein-1 [MCP-1], chitinase-3-like protein 1 [YKL-40]), and synaptic dysfunction/degeneration (neurogranin) in cognitively healthy adults in late middle age.

METHODS **Participants and study design.** Participants were drawn from a longitudinal cohort enriched with parental

history of AD, the Wisconsin Registry for Alzheimer's Prevention (WRAP).¹⁵ Beginning in 2001, 1,500+ WRAP participants were recruited from the community via advertisements and word of mouth and were aged 40–65 years at study entry. Participants underwent extensive cognitive testing and medical history assessment at baseline, 4 years later, and every 2 years subsequently at the University of Wisconsin–Madison. Beginning in 2010, participants were sequentially recruited by letter and telephone into substudies that included CSF collection. Recruitment into CSF substudies was based on temporal proximity to recent or upcoming evaluations for the parent study (average interval was 0.84 years), as well as self-reported interest in substudies that were advertised at WRAP participant events. All WRAP participants were included in the present analysis who had assayed CSF samples, completed sleep questionnaires, and were cognitively normal. When CSF was available from multiple time points, the data from the CSF sample collected closest to the sleep questionnaire were used.

Standard protocol approvals, registrations, and patient consents. The study was approved by the University of Wisconsin–Madison Health Sciences Institutional Review Board and participants provided informed written consent.

CSF collection and quantification. CSF was collected in the morning (mean 10:22 AM \pm 1 hour 12 minutes SD) following a 12-hour fast. Lumbar puncture was performed with a Sprotte 25- or 24-gauge spinal needle at L3/4 or L4/5, using gentle extraction into polypropylene syringes. Approximately 22 mL of CSF was gently mixed to remove gradient effects and centrifuged at 2,000 *g* for 10 minutes. A total of 0.5 mL aliquots of supernatants was frozen in polypropylene tubes and stored at -80°C . The samples were sent in 2 batches to the University of Gothenburg in Sweden to be assayed. For the A β 42/A β 40 ratio, CSF A β 42 and A β 40 were measured by electrochemiluminescence using an A β triplex assay (MSD Human A β peptide Ultra-Sensitive Kit, Meso Scale Discovery, Gaithersburg, MD). For

Table 1 Medical Outcomes Study Sleep Scale

	Sleep adequacy	Somnolence	Sleep problems index	Snoring
During the last 4 weeks...				
1. How long did it usually take for you to fall asleep? ^a			○	
2. On average, how many hours did you sleep each night? ^b				
How often did you... ^c				
3. Feel that your sleep was not quiet (moving restlessly, feeling tense, speaking, et cetera, while sleeping)?			●	
4. Get enough sleep to feel rested upon waking in the morning?	●		○	
5. Awaken short of breath or with a headache?			●	
6. Feel drowsy or sleepy during the day?		●	●	
7. Have trouble falling asleep?			●	
8. Awaken during your sleep time and have trouble falling asleep again?			●	
9. Have trouble staying awake during the day?		●	●	
10. Snore during your sleep?				●
11. Take naps (5 minutes or longer) during the day?		●		
12. Get the amount of sleep you needed?	●		○	

Circles indicate items included in each score. Filled circles indicate item was reversed before computing score.

^aPossible responses to question 1 were 15-minute increments from 1 = 0–15 minutes to 5 = more than 60 minutes.

^bResponses to question 2 were free entry.

^cResponses to questions 3–12 were on a 6-point scale ranging from 1 = all of the time to 6 = none of the time.

all other ratios, CSF A β 42 was quantified with a sandwich ELISA, as were t-tau and p-tau181 (INNOTEST β -amyloid1-42, hTAU-Ag, and phospho-tau[181P], respectively; Fujirebio Europe, Ghent, Belgium). Sandwich ELISAs were also used to

assay CSF YKL-40 (R&D Systems, Minneapolis, MN) and NFL (NF-light ELISA kit, UmanDiagnostics AB, Umeå, Sweden). MCP-1 was measured using the Meso Scale Discovery technique (MSD Human MCP-1; Meso Scale Discovery). All assays were conducted by board-certified laboratory technicians blinded to participant clinical characteristics. Assays were completed in a single round of analyses using one batch of reagents, yielding intra-assay coefficients of variation below 10%.

CSF measures were selected for this analysis based on their ability to predict subsequent amyloid accumulation, as we have reported previously in an overlapping sample.¹³ For statistical analyses, A β 42 was expressed in ratio to A β 40 (the more abundant, nontoxic fragment), to assess the pathologic species (A β 42) while accounting for individual differences in amyloid production. T-tau, p-tau181, NFL, MCP-1, YKL-40, and neurogranin were expressed in ratios to A β 42 to reflect coincident pathologies and because these ratios have better diagnostic and prognostic power than each species expressed alone.^{13,14} CSF A β 42 decreases as plaque burden increases, whereas the other CSF markers analyzed are elevated when pathology is greater. Therefore lower A β 42 and A β 42/A β 40 indicate greater pathology, while ratios of other CSF markers to A β 42 indicate greater pathology when they are elevated.¹³

Sleep assessment. Sleep was assessed with the Medical Outcomes Study (MOS) Sleep Scale and the Epworth Sleepiness Scale (ESS). The MOS Sleep Scale was selected because it assesses multiple sleep domains with low participant burden and good internal consistency.¹⁶ It gives scores in 6 sleep domains, derived from 12 questions (table 1). Responses were elicited for the last 4 weeks using a 6-point scale, then converted to a 0–100 scale before being summed to give sleep scores. Primary analyses focused on scores we previously found to be associated with brain amyloid burden measured by PiB-PET: sleep adequacy, somnolence, and sleep problems index.² Secondary analyses of symptoms of sleep-disordered breathing (snoring and waking short of breath) were conducted to better understand the relationship between sleep and CSF biomarkers. Higher scores indicate more of the construct being measured, such that worse sleep is reflected by lower sleep adequacy and higher somnolence and sleep problems index. The ESS was selected because it is a widely used tool in clinical sleep research for assessing daytime sleepiness and sleep propensity, with good internal validity and test–retest reliability.¹⁷ Participants use a 4-point scale to rate their likelihood of dozing off in 8 common situations, with higher scores indicating greater sleepiness (range 0–24).

APOE, family history, and cognitive assessment. Participants were classified as carriers or noncarriers of one or more APOE ϵ 4 alleles, determined by standard PCR techniques. Positive parental history of AD was defined as having one or both parents with AD as determined by a validated interview¹⁸ or autopsy-confirmed or probable AD as outlined by research criteria.¹⁹ Detailed medical history and phone interviews were conducted to confirm family history negative participants. Global cognitive function was assessed with the Mini-Mental State Examination,²⁰ and learning and memory with the Rey Auditory Verbal Learning Test.²¹ Clinical diagnosis of cognitively normal was determined by consensus of a multidisciplinary panel, based on physical examination, medical and social history, neuropsychological testing, and self-reports and informant reports of cognitive and functional status.

Statistical analysis. Separate multiple regression models were run for each combination of CSF and sleep measures, with CSF as the dependent variable and sleep as the independent variable,

Table 2 Participant characteristics

	No.	Mean (\pm SD)
Demographics		
Age at sleep Q, y	101	63.76 (\pm 6.18)
Age at CSF sample, y	101	62.95 (\pm 6.21)
Interval from CSF sample to sleep Q, y	101	0.82 (\pm 1.23)
Female, %	101	65.30
APOE ϵ 4 positive, %	99	29.70
Family history positive, %	101	73.30
CES-D	101	6.45 (\pm 6.61)
BMI, kg/m ²	101	28.92 (\pm 6.20)
Cognition		
MMSE	101	29.29 (\pm 0.97)
Education, y	100	16.60 (\pm 2.8)
RAVLT total	101	51.71 (\pm 7.73)
RAVLT delayed	101	10.64 (\pm 2.66)
Sleep		
Sleep adequacy	101	67.33 (\pm 20.78)
Somnolence	100	22.40 (\pm 16.51)
Sleep problems index	100	22.01 (\pm 12.75)
Snoring	101	31.09 (\pm 27.64)
ESS	99	6.43 (\pm 3.48)
CSF amyloid		
A β 42 (TRIPLEX), ng/L	101	686.36 (\pm 405.26)
A β 42 (INNOTEST), ng/L	101	733.32 (\pm 222.20)
A β 42/A β 40	101	0.09 (\pm 0.02)
CSF neural injury and tangles		
T-tau, ng/L	101	325.95 (\pm 115.63)
P-tau, ng/L	100	48.44 (\pm 15.20)
NFL, ng/L	96	701.92 (\pm 278.40)
Neurogranin, ng/L	91	376.17 (\pm 169.91)
T-tau/A β 42, ng/L	101	0.50 (\pm 0.32)
P-tau/A β 42, ng/L	100	0.07 (\pm 0.05)
NFL/A β 42, ng/L	96	1.05 (\pm 0.58)
Neurogranin/A β 42, ng/L	91	0.58 (\pm 0.36)
CSF inflammation		
MCP-1, ng/L	100	447.50 (\pm 143.68)
YKL-40, ng/L	99	159,786.56 (\pm 54,914.49)
MCP-1/A β 42, ng/L	98	0.67 (\pm 0.36)
YKL-40/A β 42, ng/L	99	239.83 (\pm 126.15)

Abbreviations: A β = β -amyloid; BMI = body mass index; CES-D = Center for Epidemiologic Studies Depression Scale; ESS = Epworth Sleepiness Scale; MCP-1 = monocyte chemoattractant protein-1; MMSE = Mini-Mental State Examination; NFL = neurofilament light; p-tau = phosphorylated tau; Q = questionnaire; RAVLT = Rey Auditory Verbal Learning Test; t-tau = total tau; YKL-40 = chitinase-3-like protein 1.

Table 3 Results of multiple regression of CSF biomarkers on sleep measures, controlling for sex, age at CSF sample, CSF assay batch, and time between CSF sample and sleep assessment (all mean centered)

	No.	β	R ²	R ² change	p Value
Aβ42/Aβ40					
Sleep adequacy	101	0.247	0.269	0.052	0.010 ^a
ESS	99	0.166	0.241	0.024	0.087
Somnolence	100	-0.070	0.221	0.004	0.463
Sleep problems	100	-0.148	0.238	0.021	0.114
Log(t-tau/Aβ42)					
Sleep adequacy	101	-0.310	0.167	0.083	0.003 ^a
ESS	99	0.016	0.085	0.000	0.880
Somnolence	100	0.242	0.139	0.054	0.017 ^a
Sleep problems	100	0.230	0.135	0.050	0.021 ^a
Log(p-tau181/Aβ42)					
Sleep adequacy	100	-0.281	0.112	0.068	0.007 ^a
ESS	98	-0.037	0.090	0.001	0.726
Somnolence	99	0.211	0.130	0.041	0.038 ^a
Sleep problems	99	0.214	0.132	0.044	0.033 ^a
Log(NFL/Aβ42)					
Sleep adequacy	96	-0.136	0.272	0.016	0.164
ESS	94	-0.068	0.260	0.004	0.483
Somnolence	95	0.228	0.304	0.048	0.015 ^a
Sleep problems	95	0.135	0.273	0.017	0.148
Log(MCP-1/Aβ42)					
Sleep adequacy	98	-0.158	0.257	0.022	0.105
ESS	96	-0.104	0.245	0.010	0.287
Somnolence	97	0.207	0.275	0.040	0.028 ^b
Sleep problems	97	0.104	0.246	0.010	0.266
Log(YKL-40/Aβ42)					
Sleep adequacy	99	-0.242	0.222	0.050	0.016 ^a
ESS	97	-0.064	0.175	0.004	0.529
Somnolence	98	0.184	0.203	0.031	0.060
Sleep problems	98	0.208	0.213	0.041	0.031 ^b
Log(neurogranin/Aβ42)					
Sleep adequacy	91	-0.174	0.070	0.026	0.126
ESS	89	-0.086	0.050	0.007	0.451
Somnolence	90	0.166	0.069	0.026	0.133
Sleep problems	90	0.097	0.052	0.009	0.376
Log(1/Aβ42)					
Sleep adequacy	101	-0.219	0.096	0.041	0.040 ^b
ESS	99	-0.115	0.067	0.012	0.281
Somnolence	100	0.117	0.068	0.013	0.262
Sleep problems	100	0.124	0.070	0.015	0.226
T-tau					
Sleep adequacy	101	-0.215	0.089	0.040	0.045 ^b
ESS	99	0.068	0.054	0.004	0.523

Continued

with covariates sex, age at CSF sample, CSF assay batch, and time between CSF sample and sleep assessment (all mean centered). Due to missing data, the number of participants included in each analysis ranged from 94 to 101. Ratios of CSF values to A β 42 were log-transformed to achieve normal distributions. Casewise diagnostics did not reveal any outliers (± 3 SD from the mean) that warranted removal, based on influence (Cook distance < 0.039) or leverage (DFBETA < 0.197). To account for multiple comparisons, a false discovery rate approach was used, which controls error while preserving power by adjusting the p value criterion for significance based on the number of tests performed.²² All analyses were conducted with IBM SPSS 23 (SPSS Inc., Chicago, IL).

RESULTS A total of 101 members of the WRAP cohort met criteria for inclusion (completed CSF assays, MOS Sleep Scale, and ESS). Participant characteristics are summarized in table 2. The sample was in late midlife (age 63.76 ± 6.18 years at time of sleep questionnaires), cognitively normal, and enriched with AD risk (73.3% with family history of AD, 29.7% *APOE* $\epsilon 4+$).

Regression results are summarized in table 3 and significant relationships between CSF biomarkers and sleep scores are plotted in the figure. Lower A β 42/A β 40 was associated with lower sleep adequacy (figure, A), indicating that less adequate sleep was associated with greater amyloid pathology. A β 42/A β 40 was not significantly associated with somnolence, sleep problems index, or ESS. Elevated NFL/A β 42 was associated with greater somnolence (figure, B). There was no significant relationship between NFL/A β 42 and the other sleep measures examined here. T-tau/A β 42 and p-tau/A β 42 were significantly higher with lower sleep adequacy and greater somnolence and sleep problems index (figure, C–H). Neither t-tau/A β 42 nor p-tau/A β 42 was related to ESS. Higher YKL-40/A β 42 was associated with lower sleep adequacy and greater sleep problems index (figure, I and J). There was no significant relationship between YKL-40/A β 42 and daytime sleepiness (somnolence or ESS). Higher MCP-1/A β 42 was associated with greater somnolence (figure, K) but no other sleep measures. Neurogranin/A β 42 was not associated with any of the sleep measures. Greater tau was associated with lower sleep adequacy. There were no other significant relationships between sleep and CSF biomarkers expressed alone (table e-1 at Neurology.org). There were no significant interactions with *APOE* $\epsilon 4$ status or age. There was no significant association between CSF biomarkers and self-report of sleep-disordered breathing symptoms or sleep duration. Depression, body mass index (BMI), education, cardiovascular disease, and sleep-affecting medications (table e-2) did not explain any additional variance in CSF biomarkers (table e-3), except for BMI with log (NFL/A β 42). The relationship between somnolence and log(NFL/A β 42) remained statistically significant

Table 3 Continued

	No.	β	R^2	R^2 change	p Value
Somnolence	100	0.183	0.081	0.031	0.078
Sleep problems	100	0.174	0.079	0.029	0.090

Abbreviations: A β = β -amyloid; β = standardized regression coefficient; ESS = Epworth Sleepiness Scale; MCP-1 = monocyte chemoattractant protein-1; NFL = neurofilament light; p-tau = phosphorylated tau; R^2 = overall model fit; R^2 change = contribution of sleep term to model fit; t-tau = total tau; YKL-40 = chitinase-3-like protein 1.

^ap (false discovery rate) < 0.05.

^bp < 0.05.

and effect size (model R^2) did not change substantially after controlling for BMI (supplemental data).

DISCUSSION Self-report of less adequate sleep, greater daytime sleepiness, and more sleep problems were associated with CSF biomarkers of amyloid deposition in combination with tau pathology, axonal degeneration, and neuroinflammation.

We have previously shown that self-report of poor sleep was associated with greater brain amyloid burden, measured with PiB-PET.² The CSF results reported here mirror the PiB-PET findings, and reveal further relationships between sleep and CSF biomarkers of cumulative AD pathology. CSF biomarkers give complimentary information to amyloid PET scans. First, abnormalities may become apparent in CSF before PET.²³ Second, CSF can be collected and assayed using widely available methods, whereas PET remains more expensive and requires substantial infrastructure. Finally, whereas a PET scan assesses only a single form of pathology, CSF can be assayed for multiple pathologic species simultaneously.

CSF biomarker ratios indexing multiple pathologies capture the temporal relationship between pathologies, which is a better indicator of disease stage than absolute levels of individual pathologies because AD is a progressive and multifactorial neurodegenerative disease.¹ Multipathology ratios have superior predictive power for development of neuropathologic features of AD¹³ and appear to provide superior diagnostic markers of AD.¹⁴ Our findings confirm their utility in the context of sleep, given that multipathology biomarkers were associated with more sleep domains than single-pathology markers. In particular, the only single-pathology markers associated with sleep were amyloid and tau, which were associated with lower sleep adequacy. Yet when expressed in ratio to each other, they were also associated with daytime somnolence and sleep problems. The value 1/A β 42 was associated with sleep scores in a similar but not identical pattern to the ratios, confirming that the ratios capture information that is different from single markers and that our results were not driven by A β 42 in the denominator. This is further supported

by effect sizes, which were greater for CSF ratios than 1/A β 42.

There is evidence that sleep affects AD pathology through multiple pathways. Sleep disturbance may promote amyloid plaque formation through increased amyloid production or reduced amyloid clearance. Orexin, an important regulator of the sleep-wake cycle, has been shown to drive production and deposition of A β in transgenic mice, possibly by promoting wakefulness.³ It has also been shown in mice that clearance of exogenous amyloid is greater during sleep, as a result of increased glymphatic flow.⁴ Thus sleep disruption could promote a buildup of soluble amyloid, leading to aggregation into plaques. These hypotheses remain to be tested in humans.

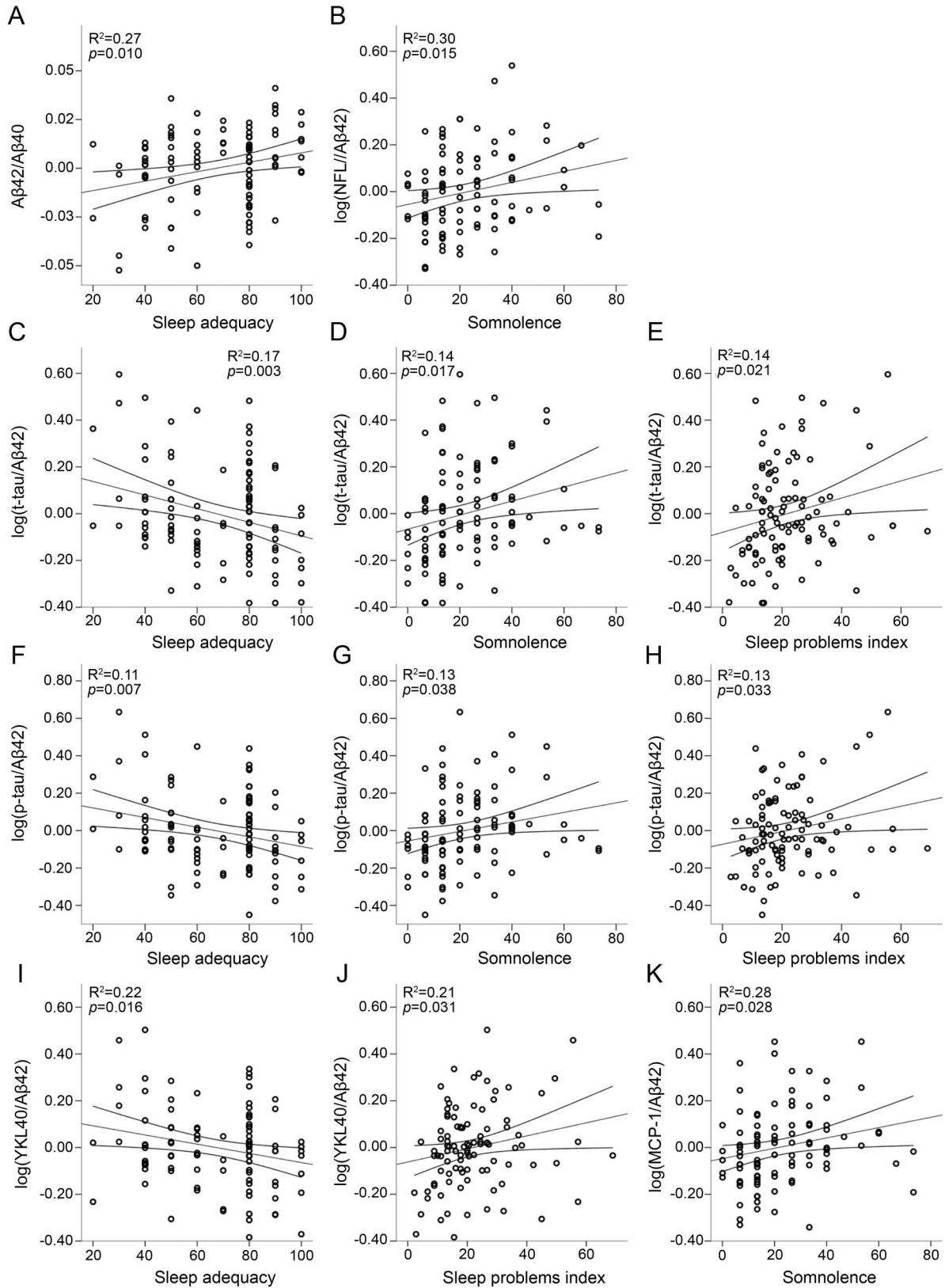
Interestingly, CSF biomarkers of AD pathology were associated with daytime sleepiness indexed by MOS somnolence but not ESS. We previously observed the same distinction with PET-measured amyloid burden.² The difference could be due to the nature of the daytime sleepiness: ESS indexes irresistible sleepiness (falling asleep in inappropriate situations such as driving) whereas somnolence may index resistible sleepiness (feeling drowsy and deliberate napping). Orexin, a wake promoter, is elevated in patients with mild cognitive impairment and associated with tau pathology.²⁴ It is possible that the sleep disturbance reported here in conjunction with resistible, but not irresistible, sleepiness reflects an abnormal upregulation of the orexin system.

YKL-40 and MCP-1 in CSF are indicative of neuroinflammation and astroglial activation associated with amyloid plaques, with diagnostic and prognostic utility for AD.²⁵⁻²⁷ Sleep restriction broadly promotes proinflammatory cytokines, possibly by disrupting diurnal fluctuations of growth and stress hormone release, and their modulation by slow-wave sleep.¹⁰ Both neurotoxic and neuroprotective roles have been proposed for YKL-40 and MCP-1 during early stages of amyloid plaque formation²⁷ and further study is required to interpret their relationship with poor sleep.

Degeneration of synapses is an early feature of AD indexed by neurogranin, a postsynaptic protein with diagnostic utility for AD.²⁸ Maintenance of membranes and synapses is upregulated during sleep^{8,9}; therefore we hypothesized that sleep disturbance would increase synaptic injury. However, we found no relationship between neurogranin and sleep. Synaptic degeneration in AD may be secondary to amyloid and tau pathology²⁹; therefore it may be that a relationship between synaptic injury and sleep is not yet detectable in this relatively young healthy cohort.

We did not find a relationship between CSF biomarkers and symptoms of obstructive sleep apnea

Figure Association of CSF biomarkers with sleep scores



(A-K) Regression results and 95% confidence intervals are plotted for the significant relationships between CSF biomarkers and sleep scores. CSF measures are adjusted for age, sex, CSF batch, and time between CSF sample and sleep questionnaire. Higher sleep scores indicate more of the construct being measured. $A\beta$ = β -amyloid; MCP-1 = monocyte chemoattractant protein-1; NFL = neurofilament light; p-tau = phosphorylated tau; t-tau = total tau; YKL-40 = chitinase-3-like protein 1.

(OSA). This lack of relationship is surprising, given consistent findings that OSA is a risk factor for dementia, possibly promoting AD pathogenesis via sleep fragmentation³⁰ or hypoxia.³¹ It is possible, however, that OSA impedes the transfer of proteins into the CSF, altering the relationship between CSF concentrations and CNS pathology in patients with OSA.¹¹ Alternatively, OSA severity in this sample may have been too low to detect a relationship with CSF, as on average participants rated their snoring frequency as low. Sleep-disordered breathing often goes undetected by patients, and subjective reports are not reliable in detecting sleep apnea. Neither snoring nor waking short of breath was significantly correlated with sleep adequacy, somnolence, ESS, or each other. This fits with literature showing that snoring and daytime sleepiness add distinct information to predict SDB.³² Given that we did not directly test for OSA, further studies measuring sleep and breathing are needed to understand the contribution of sleep-disordered breathing to the relationship between poor sleep and CSF biomarkers of AD.

Sleep disturbances may have an effect on AD pathology, but on the other hand, AD pathology may affect sleep quality. Animal models show that increasing amyloid plaque burden is accompanied by increasingly fragmented sleep, which is rescued by elimination of A β deposits in the mouse brain by active immunization.³³ In humans, there is evidence that amyloid plaques disrupt slow-wave sleep, impairing memory consolidation.³⁴ Slow-wave sleep is important for feeling refreshed³⁵; therefore our finding that perceptions of inadequate sleep were related to more AD pathology could reflect an inability to obtain sufficient slow-wave sleep as a result of amyloid plaques or possibly elevated presence of oligomeric forms of A β .

Limitations of this study include the cross-sectional design and use of subjective sleep measures. There is evidence of bidirectional relationships between sleep and amyloid^{3,33} that cannot be disentangled by this cross-sectional study design. Longitudinal follow-up within individuals will be important to determine the course of sleep and brain pathology in preclinical AD. Variability in CSF collection time may have contributed some unaccounted for variance in amyloid levels, and mean collection time was 10:22 AM, close to a 10 AM nadir in CSF amyloid reported by one study.³⁶ However, the diurnal fluctuation was smaller in older adults, and we found no correlation between CSF collection time and amyloid levels.

This study measured sleep through self-report. Objective sleep measures such as actigraphy and polysomnography would clarify the contribution of sleeping brain activity, breathing, and sleep-wake rhythmicity to

AD pathogenesis. However, the investigation of self-reported sleep quality is itself an important clinical question. There is substantial interindividual variability in the effect of sleep disturbance on cognitive and physiologic functioning.³⁷ Furthermore, sleep health is multidimensional,³⁸ and current objective techniques (e.g., polysomnography and actigraphy) do not fully capture “adequate” sleep.³⁹ Thus self-report provides important complementary information on functional outcomes and perceptions.

These results add to a growing body of evidence linking sleep quality with AD neuropathology. Our findings demonstrate that the relationship between sleep and AD pathology is present in late midlife in the absence of cognitive impairment. Furthermore, we show that the relationship is detectable using self-report and CSF biomarkers, tools that are relatively inexpensive and accessible, making them appealing for further research in large cohorts and clinical trials. ESS is a simple and widely used measure of sleep dysfunction in clinical sleep research. Given the lack of associations shown here with CSF, and previously with PET biomarkers of AD,² we recommend that other sleep measures be included in studies of the relationship between sleep and AD. Findings were not altered by race, education, depressive symptoms, BMI, or use of sleep medications.

Sleep may be a modifiable risk factor for AD during the earliest stages of the disease, before dementia symptoms appear. Although the effect sizes reported here are modest, it is estimated that delaying AD onset by a mere 5 years would reduce AD cases by 5.7 million and save \$367 billion in health care spending in the United States.⁴⁰ Many effective pharmaceuticals, devices, and behavioral interventions are already available in the clinic for improving sleep quality. Follow-up studies are needed to identify the aspects of sleep that are most amenable to modification and most effective in affecting AD pathology, to ultimately delay AD or diminish AD symptoms.

AUTHOR CONTRIBUTIONS

Kate Sprecher: design of the study, analysis and interpretation of the data, drafting and revising the manuscript. Rebecca L. Kosciak: design of the study, interpretation of the data, revising the manuscript. Cynthia M. Carlsson: interpretation of the data, revising the manuscript. Henrik Zetterberg: analysis and interpretation of the data, revising the manuscript. Kaj Blennow: analysis and interpretation of the data, revising the manuscript. Ozioma C. Okonkwo: interpretation of the data, revising the manuscript. Mark A. Sager: design of the study, revising the manuscript. Sanjay Asthana: revising the manuscript. Sterling C. Johnson: interpretation of the data, revising the manuscript. Ruth M. Benca: design of the study, interpretation of the data, revising the manuscript. Barbara B. Bendlin: design of the study, interpretation of the data, revising the manuscript.

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DISCLOSURE

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On being sick: Musings about kindness, side effects, and slowing down (see p. 414)

This podcast begins and closes with Dr. Robert Gross, Editor-in-Chief, briefly discussing highlighted articles from the August 1, 2017, issue of *Neurology*. In the first segment, Dr. Alberto Espay talks with Dr. Ted Burns about his Special Editorial, “On being sick: Musings about kindness, side effects, and slowing down.” In the second part of the podcast, Dr. Andy Southerland focuses his interview with Dr. John Millichap on the annual Resident & Fellow writing award.

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