

Postconditioning and protection from reperfusion injury: where do we stand?

Position Paper from the Working Group of Cellular Biology of the Heart of the European Society of Cardiology

Michel Ovize¹, Gary F. Baxter², Fabio Di Lisa³, Péter Ferdinandy^{4,5},
David Garcia-Dorado⁶, Derek J. Hausenloy⁷, Gerd Heusch⁸, Jakob Vinten-Johansen⁹,
Derek M. Yellon⁷, and Rainer Schulz^{8*}

¹Service d'Explorations Fonctionnelles Cardiovasculaires and Inserm U886, Hospices Civils de Lyon, University of Lyon, France; ²Division of Pharmacology, The Welsh School of Pharmacy, Cardiff University, King Edward VII Avenue, Cardiff CF10 3NB, UK; ³Dipartimento di Chimica Scienze Biomediche Sperimentali, Università di Padova, Viale G. Colombo 3, Padova 35131, Italy; ⁴Cardiovascular Research Group, Department of Biochemistry, University of Szeged, Dóm tér 9, H-6720 Szeged, Hungary; ⁵Pharmahungary Group, Department of Biochemistry, University of Szeged, Hajnóczy utca 6, H-6722 Szeged, Hungary; ⁶Hospital Universitario Vall d'Hebron, P masculina Vall d'Hebron 119, Barcelona 08035, Spain; ⁷Hatter Cardiovascular Institute, Centre for Cardiology & Research Department of Cardiovascular Medicine, University College London Hospital & Medical School, 67 Chenies Mews, London WC1E 6HX, UK; ⁸Institute for Pathophysiology, University of Essen, Medical School, Hufelandstrasse 55, 45122 Essen, Germany; and ⁹Carlyle Fraser Heart Center, Emory University Hospital Midtown and Emory University, 550 Peachtree Street NE, Atlanta, GA 30308-2225, USA

Received 25 February 2010; revised 19 April 2010; accepted 3 May 2010; online publish-ahead-of-print 6 May 2010

Ischaemic postconditioning (brief periods of ischaemia alternating with brief periods of reflow applied at the onset of reperfusion following sustained ischaemia) effectively reduces myocardial infarct size in all species tested so far, including humans. Ischaemic postconditioning is a simple and safe manoeuvre, but because reperfusion injury is initiated within minutes of reflow, postconditioning must be applied at the onset of reperfusion. The mechanisms of protection by postconditioning include: formation and release of several autacoids and cytokines; maintained acidosis during early reperfusion; activation of protein kinases; preservation of mitochondrial function, most strikingly the attenuation of opening of the mitochondrial permeability transition pore (MPTP). Exogenous recruitment of some of the identified signalling steps can induce cardioprotection when applied at the time of reperfusion in animal experiments, but more recently cardioprotection was also observed in a proof-of-concept clinical trial. Indeed, studies in patients with an acute myocardial infarction showed a reduction of infarct size and improved left ventricular function when they underwent ischaemic postconditioning or pharmacological inhibition of MPTP opening during interventional reperfusion. Further animal studies and large-scale human studies are needed to determine whether patients with different co-morbidities and co-medications respond equally to protection by postconditioning. Also, our understanding of the underlying mechanisms must be improved to develop new therapeutic strategies to be applied at reperfusion with the ultimate aim of limiting the burden of ischaemic heart disease and potentially providing protection for other organs at risk of reperfusion injury, such as brain and kidney.

Keywords Postconditioning • Ischaemia • Reperfusion

1. Historical development of the concept of infarct size limitation

Since the early 1970s,^{1,2} the prognostic impact of the size of an acute myocardial infarction (AMI) has been recognized; infarct size (IS) correlates with arrhythmia severity, the development of heart failure and mortality.^{3–5} Hence, the reduction of IS is a worthy goal in the treatment of AMI. Notably, the time-dependent development of infarction and the potential for limitation of experimental IS by various interventions were recognized.

A systematic study by Reimer *et al.*⁶ confirmed that necrosis progressed in a 'wavefront' pattern related to the duration of ischaemia, which firmly established the concept that myocardium could be salvaged by initiating reperfusion as early as possible. A microvascular no-reflow phenomenon is associated with, and possibly contributes causally to, the development of AMI and its final extent.⁷ Pioneering experimental studies from John Ross's laboratory first demonstrated that early reperfusion could salvage myocardium from infarction and reduce IS.^{8,9} This experimental concept of early reperfusion was quickly translated to clinical use, first in Europe^{10,11} and then also in

* Corresponding author. Tel: +49 2017234521, Fax: +49 2017234481, Email: rainer.schulz@uk-essen.de

the USA.¹² Subsequently, early reperfusion therapy was further promoted by the introduction of primary percutaneous coronary intervention (PCI) in myocardial infarction.¹³

Today, reperfusion strategies remain the keystone therapies to reduce IS. Improvements in outcomes have been made through facilitated reperfusion in conjunction with thrombolysis and PCI;¹⁴ no pharmacological agents have been proven to reduce IS clinically. However, they may reduce major adverse coronary events or mortality by other means. Despite better symptom recognition and transport to hospital in order to initiate reperfusion in a timely manner, ‘... it is unlikely that additional substantive improvements in morbidity and mortality can be achieved by reperfusion therapy without ... new adjunctive therapies’.¹⁵

1.1 Contribution of reperfusion injury to infarct size

Reperfusion contributes to lethal injury following prolonged periods of ischaemia (Figure 1A). The idea of reperfusion injury was first introduced by Jennings *et al.*¹⁶ as significant morphological alterations appearing after the onset of reperfusion, including cardiomyocyte swelling, mitochondrial clarification, amorphous/flocculent densities representing calcium phosphate deposits, hypercontracture, and loss of sarcomere organization. They proposed that reperfusion could hasten the progression of necrosis, but did not suggest that reperfusion caused *de novo* injury. The very existence of lethal reperfusion injury was vigorously debated¹⁷ by those who maintained that lethal injury was expressed or hastened by reperfusion on the one hand,¹⁸ and those who advocated reperfusion as a contributor to *de novo* injury.¹⁹

The existence of lethal reperfusion injury is strongly supported by evidence of reduced IS achieved by interventions when applied at the onset of reperfusion. Lethal reperfusion injury follows only after severe ischaemic injury and does not occur following fully reversible ischaemic episodes. Reperfusion initiates a cascade of events within the first minutes after restoration of flow, and causes injury in a relatively short time (Figure 1A and B), leading Piper *et al.*²⁰ to conclude that ‘what happens first must be treated first’. The obvious implication of this concept of reperfusion injury is that more myocardium can be salvaged by adding adjunct reperfusion therapies to early reperfusion.

The mechanisms of reperfusion injury have been under intense investigation for several decades, and have been reviewed elsewhere.²¹ Numerous agents and mechanical manoeuvres have demonstrated robust IS reduction when given at reperfusion in experimental studies. However, clinical trials on cardioprotective strategies and drugs were largely negative²² except for those which will be mentioned below.¹⁵ These negative results may be related, in part, to not applying the intervention at the start of reperfusion, delaying its application beyond the brief window within which salvage is achievable, or ignoring confounders and co-morbidities in animal studies when compared with the clinical reality.

Although the term ‘postconditioning’ was first introduced by Na *et al.*²³ with respect to prevention of arrhythmias, Zhao *et al.*²⁴ were the first to report in 2003 the application of postconditioning to limit lethal reperfusion injury in experimental AMI. Postconditioning differed from its preconditioning counterpart in that the mechanical intervention focused exclusively on events occurring during

reperfusion (Figure 2) and identified the early moments of reperfusion as a key therapeutic window. Later studies by Zhao *et al.* and others^{24,25} revealed that postconditioning also reduced cardiomyocyte apoptosis and contracture, coronary endothelial dysfunction, microvascular injury, tissue oedema, and organelle dysfunction. We now know that postconditioning reduces lethal reperfusion injury in multiple species including humans, and also in organs other than the heart, such as brain and kidney.

2. Models and algorithms of ischaemic postconditioning

Two recent analyses have systematically addressed models and algorithms of ischaemic postconditioning.^{26,27} For translation to humans, the temporal and spatial evolution of myocardial infarction is of critical importance. Myocardial infarction develops over time, with close dependence on the residual flow through the occluded artery and collateral blood flow. In rodents, infarction develops more quickly than in larger mammals such as dogs and pigs.²⁸ Primates exhibit a surprising resistance against infarct development.^{29,30} In humans, infarct development—assessed by enzyme release—falls in between the time course observed in dogs and pigs.³¹ As in humans, preferentially subendocardial infarction is seen in dogs, pigs, and primates, while in mice, infarction occurs preferentially in the outer layers. It is obvious that more reductionist experimental approaches, e.g. isolated, buffer perfused hearts, isolated pieces of tissue, isolated cells, or even isolated mitochondria are even further away from the true translational endpoint, i.e. IS in humans.

2.1 Duration of index ischaemia and protection

The duration of ischaemia—in addition to risk zone size and the extent of residual blood flow—is a critical determinant of IS, and also determines the reduction of IS by postconditioning. In rats after an index ischaemia of 60 min or longer, IS reduction by ischaemic postconditioning was largely diminished. In contrast, with the same postconditioning algorithm applied after an index ischaemia of 30 or 45 min, there was still significant IS reduction.²⁶ Also, with too brief index ischaemia, postconditioning failed to reduce IS in rats³² and pigs.³³ In one study in rats with very short duration of index ischaemia, postconditioning even slightly increased IS³⁴ (Figure 1B).

2.2 Postconditioning manoeuvre and protection

The postconditioning algorithm consists of at least three factors: the delay after which the first re-occlusion is established; the duration and number of re-occlusions; and the duration of the interspersed reperfusion. There is consensus that the delay in applying the first re-occlusion can only be short, but the available data are surprisingly sparse. In rats *in vivo*, IS reduction by postconditioning was lost when the first re-occlusion was shifted from 10 to 60 s reperfusion.³⁵ In rabbits *in vivo*, protection was achieved when the postconditioning manoeuvre was initiated at 30 s reperfusion but was lost at 60 s³⁶ or 10 min³⁷ reperfusion. However, in several studies in mice, rabbits, dogs, and also in studies in patients, ischaemic postconditioning still reduced IS when the delay to the first

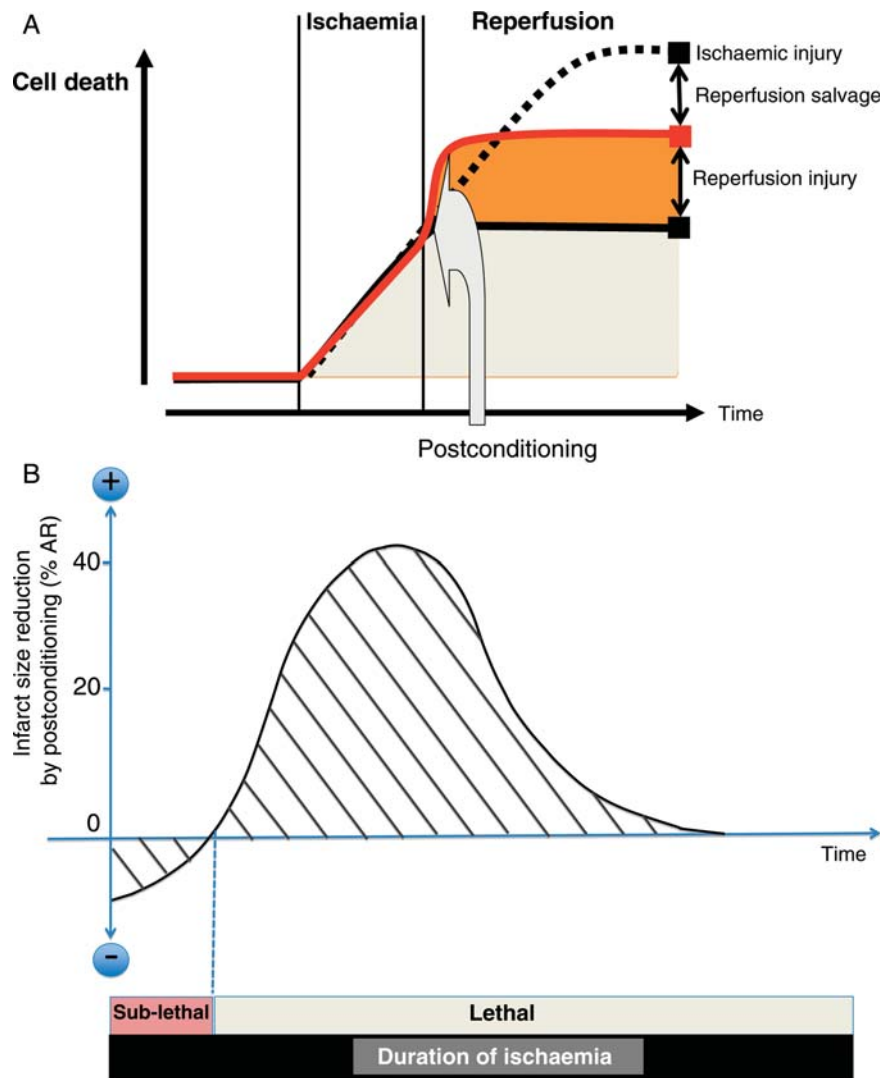


Figure 1 (A) The concept of lethal reperfusion injury. During ischaemia, irreversible cell injury leading to cell death occurs within the ischaemic risk zone in a time-dependent manner. In the absence of reperfusion, ischaemic injury would progressively kill more and more cells, eventually accounting for near total cell death (broken line). Reperfusion halts the process of ischaemic cell death but in its early stages imposes injury that results in further cell death, beyond that due to the ischaemic period: this is lethal reperfusion injury. The net result, however, is that the reperfused tissue sustains less cell death than would occur in ischaemic tissue without reperfusion. Hence, targeting cell death due to reperfusion has the potential to maximize cell salvage. Postconditioning applied at the onset of reperfusion limits the extent of reperfusion injury and maximizes reperfusion salvage. Adapted from Garcia-Dorado and Piper.²⁰⁹ (B) Attenuation of lethal reperfusion injury as a function of the duration of the preceding index ischaemia. Infarct size reduction by protective interventions performed after reperfusion (e.g. ischaemic postconditioning) may vary according to the duration of the preceding ischaemia. Experimental studies show that with prolonged periods of index ischaemia, the potential to minimize tissue salvage (infarct size reduction) may be limited probably because the extent of lethal ischaemic injury is so severe. With briefer lethal periods of ischaemia, interventions at reperfusion are able to confer marked reduction in infarct size. When the index ischaemic event is so brief that no infarction occurs (sub-lethal ischaemia), the application of ischaemic postconditioning at the onset of reperfusion may prove injurious and cause a small degree of infarction.

re-occlusion ranged from 60 to 180 s.²⁶ A delay of the postconditioning manoeuvre until 5 min of reperfusion was probably the major reason for the failure of the first postconditioning experiments in 1992.³⁸

The IS reduction by ischaemic postconditioning clearly depends on the 'stimulus' strength. Too few cycles and/or too brief ischaemia/reperfusion within each cycle fail to reduce IS. Increasing the number of postconditioning cycles in rats *in vivo* from three to six³⁴ and in pigs *in vivo* from four to eight³⁹ was needed in some studies to establish protection. In rabbits *in vivo*, the prolongation of the

ischaemia/reperfusion cycle duration from 10 to 30 s also established protection.⁴⁰

Whether or not postconditioning provides as powerful protection as preconditioning is unclear at present,²⁶ and efficacy might be dependent on the postconditioning protocol and the duration of the preceding ischaemia. A technical problem which may result from repeated occlusion of the target vessel, specifically in small animal models, is the failure to achieve complete reperfusion, and such failure of reperfusion may confound any potential protection.⁴¹

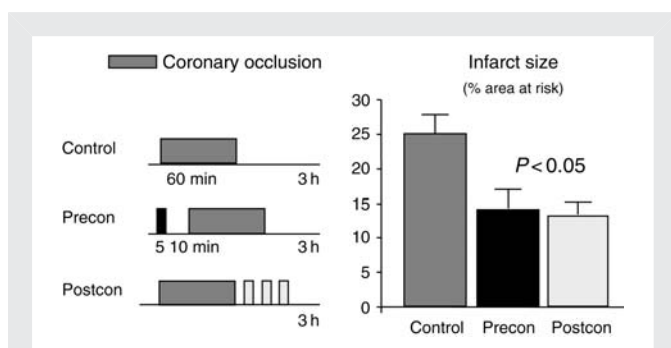


Figure 2 Infarct size reduction by pre- and postconditioning. Coronary occlusion, followed by reperfusion is associated with infarct development in the ischaemic risk zone (Control). Brief periods of ischaemia and reperfusion performed either before index coronary occlusion (Precon: preconditioning) or immediately after index coronary occlusion (Postcon: postconditioning) can significantly reduce infarct size. In animal models such as the dog subjected to coronary artery occlusion and reperfusion, either preconditioning or postconditioning can confer equivalent protection. Data from Zhao *et al.*²⁴

2.3 Remote ischaemic postconditioning

The clinical utility of ischaemic postconditioning may be limited by the need to apply an invasive treatment algorithm to the heart itself, and by the requirement to intervene at the onset of myocardial reperfusion. In this regard, remote ischaemic conditioning, in which the heart is protected by applying a conditioning stimulus of one or more cycles of brief ischaemia and reperfusion to a remote organ or tissue, may be preferable.

Kerendi *et al.*⁴² first reported that one 5 min cycle of renal artery occlusion/reperfusion applied immediately before myocardial reperfusion could reduce myocardial IS by 50% in an *in vivo* rat model. Myocardial IS limitation, attenuated apoptotic cell death, and less oxidative stress have been observed with remote ischaemic postconditioning using *in vivo* rabbit models in which the protective stimulus has been applied to either the renal artery (three cycles of 30 s reperfusion/occlusions at the end of the index ischaemia),⁴³ the femoral artery,⁴⁴ or the carotid artery (four cycles of 60 s reperfusion/occlusions just prior to myocardial reperfusion).⁴⁵ Andreka *et al.*⁴⁶ applied the remote ischaemic postconditioning stimulus (four 5 min cycles of hind-limb ischaemia/reperfusion) at the immediate onset of myocardial reperfusion and observed myocardial IS reduction at 72 h using an *in vivo* pig model.

2.4 Postconditioning effects on stunning and reperfusion arrhythmias

Although IS limitation is the primary focus of this review, several studies have examined postconditioning effects on other reperfusion pathologies, namely myocardial stunning and reperfusion arrhythmias. A beneficial effect on stunning is not clearly identified. Different postconditioning protocols failed to improve contractile recovery after brief coronary occlusion not resulting in detectable cell death in dogs.⁴⁷ Postconditioning also did not improve contractile recovery in rabbits submitted to more prolonged ischaemic episodes, although it limited IS, suggesting dissociation between the anti-infarct and anti-stunning effects. Even if there is no specific effect of postconditioning on stunning,^{48,49} some indirect beneficial effect on ventricular function could be expected from IS limitation. Studies in isolated rat hearts

have shown that the benefit of postconditioning on functional recovery was paralleled by its ability to limit necrosis.^{50,51} Importantly, the short-term and more long-term beneficial effect of postconditioning protocols which limit IS on left ventricular function has been also observed in humans.^{52,53} In an early report, an intermittent form of reperfusion termed 'postconditioning' markedly reduced the incidence of reperfusion-induced ventricular fibrillation (VF) following a 20 min episode of coronary artery occlusion in anaesthetized cats.²³ Several further studies in rats support an anti-arrhythmic action of postconditioning in this species. Galagudza *et al.*⁵⁴ reported that in isolated rat hearts displaying persistent VF 15 min after reperfusion, a single 2 min global ischaemic episode resulted in defibrillation. A similar report of the ability of 5 min global ischaemia (instituted 1 min after reperfusion) to terminate reperfusion arrhythmias in the isolated rat heart was reported by Sasaki *et al.*⁵⁵ Kloner *et al.*⁵⁶ reported that a more classical ischaemic postconditioning protocol (four 20 s episodes) following a 5 min index episode of coronary artery occlusion (insufficient to cause necrosis) in the rat *in vivo* reduced the incidence of ventricular tachycardia during reperfusion. This protective effect of postconditioning was also evident in the senescent (24 months old) rat heart.⁵⁷ The mechanism of this anti-arrhythmic effect is unknown: pharmacological inhibition studies suggest that the effect in rat is not mediated by adenosine, PI3-kinase, K_{ATP} channels, or mitochondrial permeability transition pore (MPTP) opening.⁵⁸ In pigs subjected to 48 min of coronary occlusion, postconditioning induced a clear reduction in IS, but had no significant effect on reperfusion VF (32 vs. 22%).³³

2.5 Non-cardiac ischaemic postconditioning

Ischaemic postconditioning has also been shown to confer protection in a variety of non-cardiac tissues and organs including the brain and kidney.

Ischaemic postconditioning was first demonstrated in rat brain by Zhao *et al.*⁵⁹ who reported that applying three cycles of 30 s/10 s reperfusion/occlusion to the common carotid artery reduced cerebral IS following an episode of focal cerebral ischaemia. There was attenuated apoptotic cell death and reduced oxidative stress production but the protective effect diminished with increasing ischaemic time. By applying a conventional ischaemic postconditioning protocol of 6 × 10 s reperfusion/occlusion in a rat *in vivo* model of renal ischaemia/reperfusion injury, Liu *et al.*⁶⁰ demonstrated reduced renal injury following 45 min ischaemia and 24 h reperfusion and less apoptotic cell death. Renal protection was confirmed in a subsequent study using a more staggered ischaemic postconditioning protocol (comprising 3, 6, and 12 min reperfusion interspersed with 5 min occlusion), findings which were associated with less mitochondrial oxidative stress and improved mitochondrial respiration.⁶¹

Many of the mechanisms underlying ischaemic postconditioning in brain and kidney are similar to those discussed for the heart below.^{62–65}

3. Mechanisms involved in protection by postconditioning

The current mechanistic paradigm for ischaemic postconditioning invokes the activation of signal transduction cascades by autacoid triggers; these accumulate extracellularly in response to the

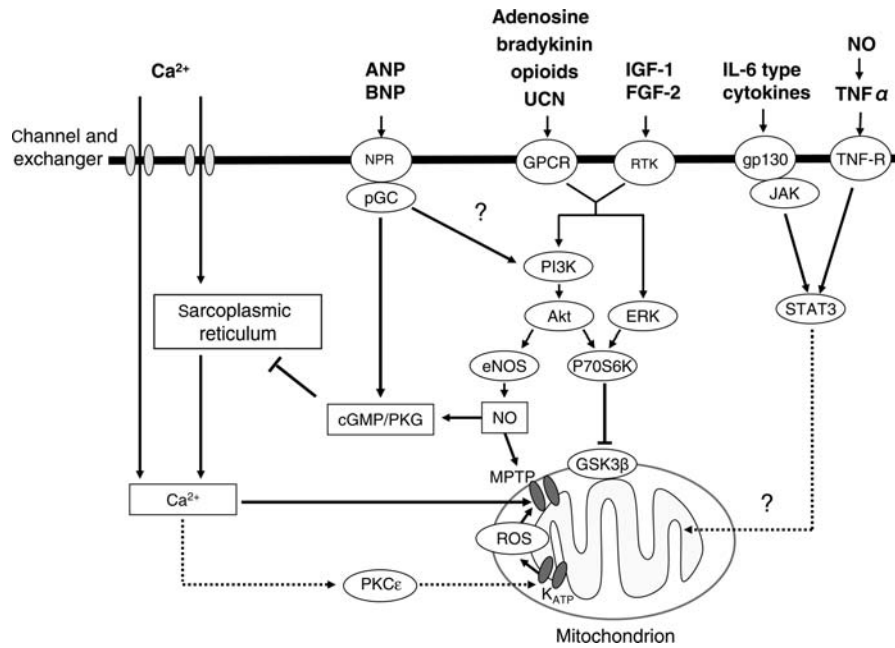


Figure 3 Signalling mechanisms in postconditioning. Several extracellular factors produced endogenously are known to play an essential role in ischaemic postconditioning (adenosine, bradykinin, and opioid peptides). However, other additional autacoids could play a role, since their exogenous administration at reperfusion mimics the effect of ischaemic postconditioning. These include natriuretic peptides (ANP and BNP), peptide growth factors (IGF-1 and FGF-2), and TNF- α . After binding to cell surface receptors, these autacoids promote the activation of kinase signalling pathways. The precise sequence of elements in these pathways and the extent of interaction between different pathways are unclear. However, evidence from some models implicates the activation of PI3K/Akt and p42/p44 ERKs. This pathway, known as the RISK pathway, is proposed to result in inhibition of mPTP opening at reperfusion, via distal components of the cascade which include NO and inhibition of GSK3 β . The extent to which cGMP accumulation and PKG activation contribute to ischaemic postconditioning is not clearly defined at present, but several pieces of evidence support the hypothesis that the activation of cGMP/PKG, either by NO or by other factors such as natriuretic peptides, is protective during reperfusion by attenuating Ca^{2+} cycling which may be a stimulus for mPTP opening. Furthermore, it has been proposed that the activation of an intramitochondrial pool of PKC ϵ might cause opening of the mitochondrial K_{ATP} channel (mito K_{ATP}), resulting in a slight increase in reactive oxygen species (ROS) formation which eventually causes MPTP inhibition. An alternative pathway, the so-called SAFE pathway, has been proposed to play a role in ischaemic postconditioning. The major components of the SAFE pathway are TNF- α , the kinase JAK which phosphorylates the transcription factor STAT3. It is proposed that after translocation to the nucleus, STAT3 controls the transcription of factors that confer cardioprotection. Also a mitochondrial localization of STAT3 has been suggested; however, both actions of STAT3 need to be finally proven. Key: eNOS, endothelial nitric oxide synthase; GPCR, G-protein coupled receptor; GSK3 β , glycogen synthase kinase-3 β ; MPTP, mitochondrial permeability transition pore; ERK, p42/p44 extracellular regulated kinase; NPR, natriuretic peptide receptor; pGC, particulate guanylyl cyclase; PKG, cGMP-dependent protein kinase; RTK, receptor tyrosine kinase; SR, sarcoplasmic reticulum; TNF-R, TNF receptor; ?, unclear at present.

postconditioning stimulus and act on cell surface receptors or other molecular targets (Figure 3).

The identification of these mediators owes much to the extensive investigations of the endogenous triggers of ischaemic preconditioning, and it seems possible that, as in ischaemic preconditioning, multiple simultaneous triggers evoke the postconditioning response. The general approaches to investigation have included: assessment of the effectiveness of ischaemic postconditioning under specific pharmacological antagonism or inhibition; modification or loss of postconditioning in receptor- or autacoid-deficient mice; the induction of protection by exogenous autacoids or by selective synthetic agonists when given immediately prior to, or at the onset of, reperfusion.

Definition of the role of an endogenous mediator in ischaemic postconditioning ideally requires satisfaction of all of the following criteria: abolition of the postconditioning effect by specific receptor blockade or by inhibition of the mediator's production; absence of postconditioning in animals, tissues, or cells with genetic disruption of the mediator's production or its receptor(s); induction of a

pharmacological postconditioning effect by exogenous administration of the mediator at the time of reperfusion. Increased production or maintenance of extracellular concentrations of the mediator(s) as a direct effect of ischaemic postconditioning might be added to this list although in practice this is the most difficult criterion to satisfy experimentally.

Although isolated cardiomyocytes can be postconditioned, the cellular and/or subcellular origin of most cardioprotective signalling molecules is not firmly known, and it is probably too simplistic to attribute the entire cardioprotective program to cardiomyocytes. In fact, the vascular wall with its endothelium, smooth muscle cells, nerve endings, and (notably in the case of atherosclerosis) inflammatory cells may be a significant source of pertinent signalling molecules: e.g. adenosine,^{66,67} bradykinin, and nitric oxide (NO) originate from endothelium and cardiomyocytes, TNF- α originates not only from cardiomyocytes,⁶⁸ but also from mast cells and macrophages,⁶⁹ and norepinephrine, opioids, and other peptides may originate from perivascular nerves.⁷⁰ The formation and release of all these signalling

molecules is probably altered with coronary artery disease and its treatment, including postconditioning. Indeed, postconditioning reduces the activation of the coronary vascular endothelium, the production of pro-inflammatory cytokines, the production of reactive oxygen species (ROS), and adherence of neutrophils to the ischaemic–reperfused coronary artery.^{24,25} Preliminary data in abstract form⁷¹ also suggest that postconditioning attenuates the superoxide anion generation by neutrophils in coronary venous blood from the ischaemic–reperfused area at risk. The above discussion supports some role for an anti-neutrophil effect in postconditioning's cardioprotection. The overall picture of reduced neutrophil adherence to the post-ischaemic coronary vascular endothelium, and the reduced accumulation of neutrophils in the infarcted myocardium lend support for an anti-neutrophil effect, although the precise mechanisms are not known at this point. However, postconditioning also reduces IS or cell necrosis in isolated perfused (Langendorff) hearts and in *in vitro* cell culture models, which may argue against a significant anti-neutrophil component in postconditioning's cardioprotection, and raises the question of 'what are mechanisms of injury that are independent of neutrophils that postconditioning addresses in these blood-free models?' In these neutrophil- and red blood cell-free systems, post-ischaemic injury can be attributed, in part, to ROS^{72,73} produced by vascular endothelium (NADPH oxidase), cardiomyocytes (xanthine oxidase), and mitochondria (electron transport chain, monoaminooxidases). Since the net effect of oxidants is related to the balance of oxidants and endogenous antioxidants, oxidant-mediated damage may be exacerbated in *in vitro* models by lack of anti-oxidants in plasma (glutathione peroxidase, ascorbate, uric acid, and some amino acids) and red blood cells (glutathione, glutathione peroxidase, catalase, and superoxide dismutase). An IS-sparing effect of postconditioning may be derived from decreased oxidant generation^{74–76} and/or increased endogenous antioxidants.⁷⁶

3.1 Autacoid factors and receptor-mediated mechanisms

A number of locally acting mediators of postconditioning, acting in an autocrine–paracrine fashion, have been identified. The best-characterized are classical autacoids that trigger the cardioprotective effects of postconditioning by receptor-mediated mechanisms, namely adenosine, opioids, and bradykinin. There is also less comprehensive evidence in *in vivo* models for the participation of other locally produced endogenous triggers that contribute to protection by postconditioning. These include ROS, NO, reactive nitrogen species,⁷⁷ calcitonin gene-related peptide, hydrogen sulfide (H₂S), and epoxyeicosatrienoic acids. Finally, there is a diverse group of naturally occurring mediators that can produce a pharmacological postconditioning effect when given exogenously but with little evidence of participation of the native ligands in ischaemic postconditioning. These include natriuretic peptides, adrenomedullin, erythropoietin, adipocytokines (e.g. apelin, visfatin, and leptin), insulin, peptide growth factors (e.g. transforming growth factor- β , and insulin-like growth factor), and urocortins.

The present discussion will be limited to evidence implicating adenosine, opioids, and bradykinin in the injury-limiting effects of postconditioning.

3.1.1 Adenosine

Interstitial adenosine is increased during ischaemia and is washed out during reperfusion.⁷⁸ Multiple adenosine receptor subtypes (A₁, A_{2A},

A_{2B}, and A₃) are expressed in the myocardium and the coronary vasculature. For many years, the ability of adenosine and selective receptor agonists to limit lethal reperfusion injury has been controversial.^{79,80} Recent research has identified a role of endogenous adenosine in ischaemic postconditioning and has renewed the interest in the potential benefits of adenosine receptor activation at reperfusion.

3.1.1.1 Antagonist and receptor deletion approaches

In mouse hearts, postconditioning delayed the adenosine washout during early reperfusion and enhanced myocardial interstitial accumulation of the nucleoside.⁸¹ In rat hearts, involvement of A₂ and A₃, but not A₁, receptor activation was demonstrated by the use of receptor-selective pharmacological agents.⁸¹ In rabbit hearts, postconditioning was abolished with the A₁ receptor antagonist DPCPX,⁸² consistent with a report that postconditioning was abolished in mice with targeted deletion of A₁ receptors.⁸³ However, other studies in rabbits demonstrated that although non-selective adenosine receptor antagonism abolished protection by postconditioning,⁸⁴ selective A_{2B} receptor activation, but not A₁ or A_{2A} activation was involved.⁸⁵ In mouse hearts, the selective A_{2A} receptor antagonist ZM-241385 abrogated postconditioning-induced improvement in functional recovery, and postconditioning failed to improve functional recovery in mouse hearts with targeted deletion of A_{2A} receptors.⁸⁶ In isolated rat hearts, the protection achieved by the adenosine A₁/A₂ agonist NECA at reperfusion was mediated through both A_{2A} and A_{2B} receptor activation, as evidenced by abrogation of protection with selective A_{2A} and A_{2B} antagonists.⁸⁷

3.1.1.2 Pharmacological postconditioning with adenosine and selective agonists

There is no consistency of an IS-limiting effect in studies using adenosine or A₁ receptor agonists.^{79,80} There is, however, a more concordant experimental literature on the ability of A₂ receptor agonists at reperfusion to limit IS among several species. In a recent study in isolated rat hearts, both a selective A_{2A} and a selective A_{2B} agonist reduced IS, and their effect was additive.⁸⁷ Some uncertainty is related to the mixed pharmacological profiles of many adenosine receptor agonists. Additionally, several studies have confirmed the protective effect at reperfusion of selective A₃ receptor agonists such as IB-MECA and chloro-IB-MECA in various models.

Thus, a general role of adenosine receptor activation by endogenous and exogenous adenosine has been demonstrated using a combination of pharmacological antagonism and gene deletion, but the relative contribution of different adenosine receptor subtypes remains unresolved and might be species- and model-dependent.⁸⁰

3.1.2 Opioids

Opioid peptides, including met-enkephalins, leu-enkephalins, and dynorphins, and three major opioid receptors (μ -, δ -, and κ -subtypes) are present in myocardium. Ischaemic postconditioning maintained myocardial opioid peptide concentrations during early reperfusion.⁸⁸ In rats, the non-selective opioid receptor antagonist naloxone or selective antagonists of δ -, κ -, or μ -opioid receptors abrogated protection by postconditioning.^{88,89} Conversely, pharmacological postconditioning was achieved with morphine (non-selective agonist),^{88,90,91} selective δ -opioid,^{90,92,93} or κ -opioid receptor agonists.⁹³ Chen *et al.*⁹¹ implicated κ -opioid receptor activation, but not δ -opioid receptor activation, in morphine-induced postconditioning.

Taken together, opioid receptor activation contributes to the effect of ischaemic postconditioning in rodents. Exogenous administration of opioid receptor agonists induces a pharmacological postconditioning effect, independent of the opioid receptor subtypes stimulated.

3.1.3 Bradykinin

Bradykinin is a peptide mediator of the kinin family produced from precursor kininogens by vascular and cardiac endothelium. Two bradykinin receptor subtypes, B₁ and B₂, are recognized of which B₂ is constitutively expressed in cardiovascular tissues, while B₁ is up-regulated under hypoxic and inflammatory conditions.⁹⁴ The B₂ receptor antagonists icatibant or WIN64338 abrogated protection by postconditioning,⁹⁵ and this finding was confirmed by ablation of B₂ receptors in knockout mice; interestingly, there was partial attenuation in B₁ receptor knockout hearts as well.⁸³ Conversely, administration of bradykinin at reperfusion limits IS in mouse, rat, and rabbit hearts.^{95–98} The mechanisms underlying the protective effect of bradykinin-induced postconditioning are unclear, but participation of both NO and prostaglandin (PGI₂) synthesis has been described in rodent myocardium.^{97,98} Thus, endogenous and exogenous bradykinin can mediate protection by postconditioning through the activation of both bradykinin receptors.

Taken together, ischaemic postconditioning enhances the concentration of endogenous autacoids during early reperfusion. The enhanced autacid concentrations, through activation of their specific receptors, contribute to the reduction in lethal reperfusion injury. Blocking the effect of any of the endogenous autacoids abrogates protection by ischaemic postconditioning while exogenously administering each autacid is capable of inducing postconditioning-like protection.

3.2 Ionic homeostasis in protection by postconditioning

As is the case with autacid accumulation, myocardial ischaemia causes accumulation (increased production/reduced washout) of protons and subsequently a progressive fall in intra- and extra-cellular pH, usually within a few minutes. Reperfusion removes extracellular protons and corrects intracellular acidosis, mainly through the activity of the sarcolemmal Na⁺/H⁺ exchanger (NHE1) and the Na⁺-bicarbonate co-transporter. The activity of both transporters contributes to intracellular Na⁺ accumulation—and subsequently Ca²⁺ overload—as the increased cytosolic Na⁺ concentration stimulates the reverse-mode activity of the sarcolemmal Na⁺/Ca²⁺ exchanger (NCE).^{99,100} Studies in isolated cardiomyocytes identified the rapid correction of intracellular acidosis as an important determinant of reperfusion injury, which favours Na⁺ and Ca²⁺ overload,¹⁰¹ but also allows the activation of systems that remain otherwise inhibited during low pH (see below).¹⁰² The time course of correction of the intracellular Ca²⁺ homeostasis and tissue pH during reperfusion decides between cell death (recovery of pH occurs first) and survival (recovery of Ca²⁺ occurs first).¹⁰⁰ The rapid normalization of the intracellular Ca²⁺ concentration is critically dependent on the rapid recovery of the activity of the sarcolemmal Na⁺/K⁺ ATPase upon re-energization.¹⁰³

The protective effect of acidosis during initial reperfusion is not surprising in view of its effects on many molecular mechanisms of reperfusion injury. During the initial minutes of reperfusion, when Ca²⁺ oscillations between the cytosol and the sarcoplasmic reticulum

occur, low pH inhibits contractile activity and hypercontracture, and also reduces gap-junction communication and thereby the spreading of cell death.²⁰ Moreover, low pH during reperfusion prevents opening of the MPTP (see below),¹⁰² calpain activation, and calpain-mediated proteolysis. NHE inhibition at the time of reperfusion does not delay the recovery of intracellular pH *in vivo* because of the compensating activities of other ionic systems (mainly, the Na⁺-bicarbonate co-transporter), which might explain the negative results of NHE inhibition given at reperfusion in AMI patients.¹⁰⁴

3.2.1 Intracellular pH in postconditioning

Heusch¹⁰⁵ was the first to propose that the effect of postconditioning is secondary to the maintenance of acidosis. Cohen et al.¹⁰⁶ provided the first experimental evidence for this hypothesis; acidic reperfusion caused equivalent protection as postconditioning in isolated rabbit hearts, and the protection was related to inhibition of MPTP opening.^{106,107}

NMR spectroscopy studies in rat hearts showed that postconditioning delays pH recovery for up to 3 min depending on the postconditioning protocol used.⁵¹ In fact, these studies demonstrated that only postconditioning protocols able to induce a significant delay in pH recovery were able to afford cardioprotection. Furthermore, a close correlation between the delay in pH recovery and the magnitude of myocardial salvage was documented.⁵¹ The pH value and duration of infusion that afforded optimal protection was found to be 6.4 and 3 min, respectively, in perfused rat hearts.¹⁰⁸ Interestingly, acidic infusion for less than 2 min was ineffective,¹⁰⁷ and prolongation of infusion beyond 3 min resulted in progressive loss of benefit. The protocol of acidic intracoronary reperfusion in pigs submitted to transient coronary occlusion was as effective as postconditioning in limiting IS, but was accompanied by an increased risk of ventricular arrhythmias.³³

3.2.2 Calcium homeostasis and calpains in postconditioning

Despite the recognized importance of free ionized Ca²⁺ in lethal reperfusion injury, there is little information on calcium homeostasis in postconditioning. The cytosolic and mitochondrial Ca²⁺ accumulation following hypoxia and re-oxygenation was reduced by hypoxic postconditioning in isolated cardiomyocytes.⁷⁴ On the contrary, mitochondria isolated from postconditioned rabbit hearts following ischaemia/reperfusion contained more total (free plus bound) calcium when compared with mitochondria from non-postconditioned hearts.¹⁰⁹ The controversy might be explained in part by ionic⁷⁴ vs. total¹⁰⁹ mitochondrial calcium measurements.

Although the issue of calcium handling in postconditioning is unresolved at present, postconditioning might interfere with Ca²⁺-dependent proteolysis by calpains. Calpains are ubiquitous cytosolic Ca²⁺-dependent proteases which act on a large number of substrates including structural myofibrillar and sarcolemmal proteins, and calpain activation contributes to sarcolemmal fragility and cell rupture during reperfusion,¹¹⁰ to detachment of the Na⁺ pump,¹¹⁰ to cleavage of the anti-apoptotic protein Bid, and activation of the pro-apoptotic Bad/Bax pathway.¹¹¹ More recently, calpain has been shown to be a target of postconditioning's protection.⁵¹

Calpain activation depends on the intracellular pH; calpain activity is maximal at pH close to 7.2 and absent at pH of 6.4.¹¹² Therefore, it can be assumed that pH normalization during reperfusion contributes to calpain-mediated proteolysis. In fact, a close correlation between the myocardial calpain activity during initial reperfusion and IS in

postconditioned rat hearts exists.⁵¹ The observation that transient acidosis at the time of reperfusion effectively reduces calpain-mediated proteolysis and reperfusion injury suggests that calpain is not activated during ischaemia despite the increased cytosolic Ca^{2+} concentration, and that low pH during ischaemia is responsible for this effect. This opens the possibility of IS limitation by administering calpain inhibitors immediately before reperfusion, as recently shown in rats submitted to transient coronary occlusion.¹¹³

3.3 Signal transduction by protein kinases

Certain protein kinase cascades are implicated in the transduction of signals from the receptor to the potential end-effector; the major kinase pathways identified so far are discussed below.

3.3.1 Reperfusion Injury Salvage Kinase (RISK) pathway

In 2002, Yellon and co-authors¹¹⁴ introduced the concept of a pro-survival reperfusion signalling pathway. In a study examining the cardioprotective effect of urocortin they demonstrated that the p42/p44 (ERK 1/2) mitogen activated protein kinase (MAPK)-dependent signalling pathway represented an important survival mechanism against reperfusion injury, and suggested that the heart possessed pro-survival 'Reperfusion Injury Salvage Kinase' (RISK) pathways. Yellon and co-author¹¹⁵ went on to further expand the concept of the RISK pathway, and subsequently they and others demonstrated that the pharmacological activation of pro-survival kinases, such as PI3 kinase-Akt and ERK1/2, at the immediate onset of myocardial reperfusion reduced IS by 40–50%.¹¹⁵ This involvement of the kinases was demonstrated using a diverse variety of agents, including G-protein coupled receptor agonists and natriuretic peptides, but also pharmacological agents such as statins. Successive studies have also confirmed the role for Akt and ERK1/2 in the setting of postconditioning in both non-diseased animal hearts, as well as in post-infarct remodelling.^{37,116,117} Importantly, *ex vivo* studies using human atrial muscle have now confirmed the cardioprotective role of the RISK pathway in the setting of simulated ischaemia/reperfusion injury.¹¹⁸

Although the concept of the RISK pathway has gained widespread attention, it is important to appreciate that it is a broad ranging concept which is inclusive of many prosurvival signalling kinase pathways that are upregulated in response to a stress, such as ischaemia/reperfusion, and it is not limited to ERK1/2 or PI3 kinase-Akt. Glycogen synthase kinase-3 β (GSK-3 β), a protein kinase linked to the regulation of a variety of cellular functions including glycogen metabolism, gene expression, and cellular survival, could either be considered as a specific downstream target of the RISK pathway or indeed as a component of the RISK pathway. Phosphorylation and thereby inhibition of GSK-3 β confers cardioprotection through its potential mitochondrial effects which include the inhibition of MPTP opening¹¹⁹ (see below) and the control of mitochondrial adenine nucleotide transport through the outer mitochondrial membrane.¹²⁰ However, there is conflicting information regarding the role of GSK-3 β as an important mediator of postconditioning.^{121,122} Mice containing a mutant form of GSK-3 β , which cannot be phosphorylated and thereby inhibited, were resistant to the myocardial infarct-limiting effects of postconditioning, suggesting that GSK-3 β inactivation is required for postconditioning.¹²¹ In contrast, mice with a mutant form of both GSK-3 β and GSK-3 α , in which the Akt phosphorylation sites were changed rendering them resistant to inactivation, were still amenable to the myocardial infarct-limiting effects of both pre- and postconditioning,

suggesting that GSK-3 β and GSK-3 α inactivation is not necessary for cardioprotection.¹²²

Although most interest has focused on the ERK member of the MAPK—this being the first kinase to be associated with the RISK pathway¹¹⁴—two other members of the MAPK family¹²³ have been examined and debated in the context of cardioprotection, namely JNK and p38MAPK.^{124–126} With regard to ischaemic postconditioning, however, the only study to investigate the role of these kinases, in which adult rat cardiomyocytes subjected to simulated ischaemia were used, suggests that these MAPKs are inhibited and that their activation at the onset of myocardial reperfusion is detrimental.¹²⁷

From the evidence presented above, it appears that the RISK pathway is fundamental to myocardial protection. However, recent evidence suggests that there are additional and independent pathways which can also be solicited and available to protect the myocardium from ischaemia/reperfusion injury.¹²⁸ In pigs, postconditioning activated Akt and ERK 1/2 but did not reduce IS.¹²⁹ In mice and rabbits, protection by postconditioning was associated only with ERK but not with Akt activation.^{130,131} Recently, Heusch's group¹³² demonstrated that in anaesthetized pigs postconditioning reduced IS. However, this protection was not associated with greater phosphorylation of Akt, ERK, p70S kinase, or GSK-3 β than reperfusion *per se*, and the protection was not abrogated by combined pharmacological inhibition of the PI3 kinase or ERK1/2 pathways; also protection by gentle reperfusion was not mediated by RISK activation in pigs.¹³³

3.3.2 Survivor Activating Factor Enhancement (SAFE) pathway

The activation of the JAK-STAT pathway has been proposed as an alternative cardioprotective pathway, apart from and in addition to the RISK pathway.¹³⁴ Lecour and co-authors has termed this alternative pathway 'Survivor Activating Factor Enhancement pathway (SAFE)'.^{135,136} The JAK-STAT pathway conveys extracellular stress signals from cytokine receptors on the plasma membrane to the nucleus. Here an assortment of proteins is transcribed relating to a variety of cellular processes, including those involved in cardioprotection. Pharmacological inhibition of the JAK-STAT pathway at the onset of myocardial reperfusion, or its genetic ablation, can abrogate the infarct-limiting effects of postconditioning.^{137,138} However, mice with a cardiac-specific STAT3 deletion were still amenable to the infarct-limiting effects of postconditioning, providing a suitable protocol was used. For example, postconditioning using 5 \times 5 s cycles of ischaemia/reperfusion reduced myocardial IS but 3 \times 10 s cycles did not.¹³⁷ In mice with the same cardiac-restricted STAT3 deletion, a postconditioning protocol consisting of 3 \times 10 s cycles of ischaemia/reperfusion improved LV function.¹³⁸ Lecour and co-authors demonstrated in isolated mouse hearts that the protective effects of TNF- α depended on both STAT3 and specifically the TNF receptor 2, but not on the TNF receptor 1¹³⁹ or on the activation of PI3 kinase-Akt or ERK.^{135,136,140} Both the RISK and SAFE pathways seem to converge on the mitochondria which may be the target or end effector for the protection.⁷⁰

3.3.3 Sphingosine kinase

Sphingosine kinase (SPhK), which generates sphingosine-1-phosphate (S1P), regulates cell mitosis, apoptosis, cytoskeletal rearrangement, and survival.¹⁴¹ Hearts isolated from mice lacking SPhK1 sustained larger myocardial infarcts were resistant to protection by postconditioning, and were unable to demonstrate the activation of Akt and ERK 1/2.¹⁴² Potentially, the S1P formed by SPhK in early reperfusion

moves into the extracellular space and activates the S1P-receptor, which then recruits other components of the RISK pathway. On the other hand, sphingosine is a downstream mediator of TNF- α ;¹⁴³ therefore, the relation of sphingosine kinase to the other cardioprotective kinase pathways is not really clear at present.

3.3.4 Protein kinases C and G

The information on protein kinases C and G in protection by ischaemic postconditioning is limited at present. Penna et al.¹⁴⁴ first demonstrated that a non-specific PKC inhibitor abolished the IS-limiting effects of postconditioning in perfused rat hearts, suggesting that postconditioning required the activation of PKC to confer cardioprotection. A subsequent study by Zatta et al.¹⁴⁵ found that protection by postconditioning was abolished by pharmacological inhibition of the PKC- ϵ isoform in early reperfusion. The mechanism through which postconditioning activates PKC is unclear.

Although protein kinase G (PKG) has emerged as a mediator of cardioprotection,¹⁴⁶ its role in IS limiting in ischaemic postconditioning is unclear. Protection by postconditioning has been described to be sensitive to pharmacological inhibition of the NO-sGC-cGMP-PKG pathway,^{84,147} although there might be PKG-independent effects of NO including a wide variety of direct actions on mitochondria.

Thus, in summary, ischaemic postconditioning may alter activation of multiple protein kinase and/or phosphatase signalling pathways during reperfusion. The relative importance of each pathway for protection is still controversial and might depend on the species, model, and protocol used.

3.4 Mitochondria in postconditioning

Mitochondria are both a central target of processes triggered by ischaemia, such as elevation in intracellular calcium and ROS, and a pivotal site for determining the preservation or loss of cell viability. Strategies aimed at protecting against ischaemia/reperfusion damage have focused on mitochondria, especially on the MPTP.^{148–150}

The MPTP can be described as a voltage- and Ca²⁺-dependent, high-conductance channel located in the inner mitochondrial membrane, and sensitive to inhibition by cyclosporine A (CsA). Its opening causes a sudden increase in inner mitochondrial membrane permeability to solutes with molecular masses up to 1500 Da.¹⁵¹ Oddly, the molecular identity of MPTP has not been elucidated yet. Adenine nucleotide translocase and the voltage-dependent anion channel (VDAC) have been proposed to represent MPTP components. However, genetic studies have demonstrated that MPTP opening can still be observed in mitochondria devoid of these proteins that might modulate MPTP function.^{152,153} On the other hand, MPTP opening is facilitated by binding of the matrix protein cyclophilin D (Cyp-D) to the inner mitochondrial membrane in a process modulated by both Ca²⁺ and inorganic phosphate.¹⁵¹ Notably, Cyp-D binding to the inner mitochondrial membrane is prevented by CsA and other molecules interacting with Cyp-D that are usually described as MPTP inhibitors. However, since increasing Ca²⁺ concentration still results in MPTP opening even in the presence of these molecules, MPTP desensitization describes better the effect of this family of compounds. The same applies to mitochondria devoid of Cyp-D.¹⁵⁴

MPTP opening causes an immediate collapse of the mitochondrial membrane potential ($\Delta\psi_m$) that is followed by ATP depletion. When opening is prolonged, the initial uncoupling-like effect is rapidly followed by respiratory inhibition caused by the loss of pyridine nucleotides and of cytochrome c; the latter is potentially

dependent on the rupture of the outer mitochondrial membrane caused by MPTP-dependent matrix swelling. The resulting inhibition of electron flow might explain the increased ROS formation induced by MPTP opening. Since the latter event is favoured by ROS,^{155,156} a vicious cycle of injury amplification is likely to be established, especially at the onset of reperfusion. MPTP opening is favoured by mitochondrial Ca²⁺ elevation, depolarization, and increases in ROS and inorganic phosphate. These factors are counteracted by physiological MPTP antagonists, such as elevated values of $\Delta\psi_m$ and high concentrations of protons (H⁺), magnesium (Mg²⁺), and adenine nucleotides, especially ADP.¹⁵¹ During ischaemia, intracellular acidosis, along with high levels of Mg²⁺ and ADP, overrides the MPTP-promoting conditions established by $\Delta\psi_m$ decrease and increases in Ca²⁺ and inorganic phosphate levels. Conversely, upon reperfusion, the recovery of pH together with a burst in ROS formation in the presence of high matrix concentrations of Ca²⁺ and inorganic phosphate creates optimal conditions for MPTP opening despite the antagonizing effect of $\Delta\psi_m$ recovery.

MPTP opening is prevented by endogenous self-defence mechanisms aimed at maintaining cell viability. This is the case with both ischaemic preconditioning and postconditioning.^{157–161} This notion is supported by the following major lines of evidence: (i) ischaemic postconditioning reduces MPTP susceptibility in mitochondria isolated during reperfusion¹⁵⁷ and (ii) treatments that antagonize MPTP opening elicit cardioprotective effects similar to those induced by ischaemic postconditioning.^{158,162} The inhibition of MPTP opening by CsA was shown to afford significant cardioprotection against ischaemia/reperfusion injury,^{153,155,163–165} and this concept has been successfully translated to the clinical setting.¹⁶⁶

Signalling pathways activated by postconditioning, in particular the RISK pathway, have been assumed to modulate the open probability of MPTP. However, a relevant piece of information that is still missing is how processes occurring in the cytosol modulate MPTP opening in the inner mitochondrial membrane. This might be through (de)phosphorylation of critical proteins in the outer mitochondrial membrane and/or by translocation of cytosolic proteins into mitochondria. For instance, the phosphorylation status of the VDAC appears to affect adenine nucleotide traffic between mitochondria and cytosol and/or the binding of the anti-apoptotic protein Bcl-2.¹⁶⁷

The relationship between MPTP and oxidative stress might be modulated by additional processes. In this respect, an interesting observation is that postconditioning protection is abolished by treatment with the antioxidant N-acetyl-cysteine during the initial 3 min of reperfusion, but not during the subsequent phase.¹⁴⁴ This early ROS formation would then trigger protective mechanisms that appear to include the activation of mitochondrial K_{ATP} channel (mitoK_{ATP}) and PKC, since their inhibition by 5-hydroxydecanoate and chelerythrine, respectively, abrogates protection. Although the source and the moieties of ROS, and PKC isoform(s) were not elucidated, mitoK_{ATP} activation might underlie postconditioning-induced cardioprotection^{37,95,144} by reducing the susceptibility to MPTP opening. In fact, it has been proposed that the activation of an intra-mitochondrial pool of PKC ϵ might cause opening of the mitoK_{ATP}, resulting in a slight increase in H₂O₂ formation which eventually causes MPTP inhibition.¹⁶⁸

Although mitochondria are considered the end effectors of protective pathways, it is likely that differences exist between the modalities through which the various conditioning stimuli affect mitochondrial function and structure. So far the only characterized example of

these possible differences is connexin 43. Mitochondrial connexin 43, which is causally involved in ischaemic preconditioning,^{169,170} is not a prerequisite for ischaemic postconditioning.¹⁷¹

4. Co-morbidities and co-treatments and postconditioning

Many of the above signalling elements might be affected by confounders, co-morbidities, and co-treatments. For example, the bradykinin concentration is increased by ACE inhibition, the NO/ROS balance is affected by atherosclerosis, and age reduces the expression of protein kinases and STAT3. Therefore, it is of utmost importance to assess the effects of these entities on protection by postconditioning.

4.1 Gender and age as confounders of protection by postconditioning

Whether cardioprotection by postconditioning is gender-dependent is not really clear. After 30 min ischaemia, IS was smaller in female than male hearts. However, postconditioning reduced IS in both genders, though the effect was smaller in females³² and dependent on the duration of the sustained ischaemia.¹⁷² In a single study in female rats,¹⁷³ IS *per se* was smaller than expected from male rats, but ischaemic postconditioning failed to protect and even increased IS.

Age has a major impact on IS reduction by ischaemic postconditioning in mice, but not in rats.^{174,175} In isolated hearts from mice older than 80 weeks, postconditioning failed to reduce IS.¹³¹ Postconditioning by three cycles of 10 s/10 s ischaemia/reperfusion reduced IS in young (<12 weeks) mice *in vivo*, but failed to do so in mice older than 52 weeks. However, changing the postconditioning algorithm to five cycles of 5 s/5 s ischaemia/reperfusion in these mice fully re-established the protection.¹³⁷ Interestingly, the biological age of these mice was similar to the biological age of patients in which postconditioning still reduced IS considerably. The induction of postconditioning protection may be a question of optimization of the protocol.

4.2 Underlying conditions and protection by postconditioning

Most experimental studies on cardioprotection have been undertaken in young and healthy animals. However, ischaemic heart disease in humans is a complex disorder caused by or associated with known cardiovascular risk factors including smoking, obesity, hyperlipidaemia, diabetes and hypertension, or pre-existing diseases (e.g. heart failure). In addition, patients with CAD vulnerable to myocardial infarction may be on various pharmacological treatments. These conditions are associated with fundamental molecular alterations that can potentially affect the development of ischaemia/reperfusion injury *per se* and responses to cardioprotective interventions such as postconditioning.

Hyperlipidaemia, especially hypercholesterolaemia, is regarded as an independent risk factor for the development of CAD, increases the severity of myocardial ischaemia/reperfusion injury, and interferes with cellular mechanisms of cardioprotection.^{176,177} However, little is known about the effect of postconditioning in hyperlipidaemia. Iliodromitis *et al.*¹⁷⁸ showed the loss of the IS limiting effect of postconditioning in rabbits fed a cholesterol-enriched diet for 6 weeks. The loss of the IS limiting effect of postconditioning was confirmed in hearts isolated from chronically cholesterol-fed rats.⁷⁷ However, Donato

*et al.*⁸² found no alteration in the IS limiting effect of postconditioning in rabbits fed 1% cholesterol for 4 weeks. The reasons for the conflicting results are unknown, but might be related to the severity and duration of the disease state and extracardiac pathologies in the different experimental hyperlipidaemia models. Apart from the accumulation of tissue/membrane cholesterol and alterations in NO-cGMP and peroxynitrite signalling,^{176,177,179} the exact cellular mechanism by which hyperlipidaemia may influence postconditioning is not known.

Very little is known on the interaction of diabetes with postconditioning. Both diabetic as well as obese mice have been reported to be resistant to protection by postconditioning, a finding that has been associated with insufficient activation of the RISK pathway.^{180,181} Similarly, in pre-diabetic rats with metabolic syndrome, sevoflurane-induced postconditioning no longer reduced IS and even CsA given at reperfusion failed to protect the pre-diabetic heart.¹⁸²

In spontaneously hypertensive rats, postconditioning failed to reduce IS.¹⁸³ In rats with pressure overload-induced LV hypertrophy, postconditioning was still capable of reducing IS. Also, ischaemic postconditioning, as well as pharmacological postconditioning with isoflurane, was effective in reducing IS and activating the RISK pathway in rat hearts 6 weeks after MI.^{117,184}

Although protection by postconditioning was reduced in animals with a single risk factor, whether or not and how much any of these risk factors blunts IS reduction by postconditioning in CAD patients needs to be determined (see below). Acute MI patients treated by postconditioning will have received several pharmacological treatments because of (i) pre-existing risk factors and/or (ii) treatment of AMI; e.g. aspirin, heparin, clopidogrel, β -blockers, morphine, nitrates, etc. Whether these treatments affect IS reduction by postconditioning in patients is currently unknown.

Some pharmacological treatments have been shown to impact on postconditioning's IS reduction in animal models. For example, β -adrenergic signalling is involved in protection by desflurane-postconditioning,¹⁸⁵ and accordingly β -blockade can abolish such protection.¹⁸⁵ Whether or not this holds true for ischaemic postconditioning is currently unknown. On the other hand, ischaemic postconditioning is lost in rat hearts with chronic coronary stenoses, and protection is restored by carvedilol.¹⁸⁶ The protection by the selective bradycardic agent ivabradine is still achieved when given in a postconditioning modality just before reperfusion, and is not dependent on heart rate reduction.¹⁸⁷ Statins may also interfere with IS-limiting effect of postconditioning in rats.¹⁸⁸ Ischaemic (see above) and sevoflurane-induced postconditioning has been shown to be blocked by anti-diabetic K_{ATP} -channel blockers in rats.¹⁸⁹

5. Clinical application of ischaemic or pharmacological postconditioning

In 2005, the experimentally proven concept of ischaemic postconditioning was translated into the clinical setting (*Figure 4* and *Table 1*). In a proof-of-concept trial, patients with a first-time ST elevation myocardial infarction (STEMI) having chest pain for less than 6 h and a need for angioplasty revascularization underwent direct stenting. Control patients underwent no additional intervention after reperfusion, while postconditioned patients underwent four cycles of 1 min inflation and 1 min deflation of the angioplasty balloon at low pressure within the first minute after stent implantation, with placement of the

balloon just upstream of the stent. Area under the curve of creatine kinase release over the first 3 days of reperfusion (as a surrogate for IS) was significantly reduced by 36% in the postconditioned vs. control group.¹⁹⁰ This study confirmed that lethal reperfusion injury represents a significant amount of the overall myocardial damage in AMI patients, and that this tissue destruction can be prevented by a timely intervention. IS reduction by PCI postconditioning persisted 6 months after AMI (IS assessment by ¹⁰¹Thallium SPECT imaging), and resulted in a significant improvement of contractile function at 1 year after infarction.¹⁹¹ These findings are in agreement with a retrospective study by Darling *et al.*¹⁹² who reported that AMI patients who underwent four or more balloon inflations–deflations developed smaller infarcts than patients who received less than four of these brief cycles of ischaemia/reperfusion. Similarly, Wang *et al.*⁵² reported in a retrospective analysis of a cohort of 433 STEMI patients that those with more than three balloon inflations at the time of reperfusion displayed a lower peak of creatine kinase release, a lower LV end-systolic volume, and a higher ejection fraction, than those with one or two inflations.

Previous experimental studies have indeed clearly demonstrated that staged reperfusion provides some protection,^{193,194} although different from that observed with postconditioning (i.e. staged reperfusion protects from stunning, whereas postconditioning does not). To our knowledge, there has been no report directly comparing staged reperfusion vs. ischaemic postconditioning. This issue is of importance in clinical practice, mainly because reperfusion may be obtained using two different techniques, i.e. angioplasty and fibrinolysis. Whether ischaemic postconditioning is efficient when reperfusion is obtained by fibrinolysis is still unknown. This latter mode of

restoration of coronary flow may possibly modify the efficiency of postconditioning. Fibrinolysis may cause a more gentle reflow than angioplasty due to both a slower disaggregation of the thrombus, and mainly, the persistence of a coronary artery stenosis which attenuates hyperaemia.⁴¹ Also, while the timing of reflow by angioplasty is sharp, the exact time of reopening of the coronary artery by thrombolysis may be difficult to determine accurately, and it may only be suggested by the reduction of chest pain and some signs on the ECG such as ST-segment shift reduction or the occurrence of some types of arrhythmias. Timing, however, is of major importance since experimental evidence indicates that any protective intervention can be effective only if performed within the first minutes of reflow.

5.1 Remote ischaemic conditioning

Loukogeorgakis *et al.*¹⁹⁵ demonstrated that remote ischaemic postconditioning could be produced in both human volunteers and patients with CAD in a manner which was sensitive to inhibition by glibenclamide (a K_{ATP} channel blocker) using endothelial function, but not IS, to assess protection. In a recent proof-of-concept clinical study, Botker *et al.*¹⁹⁶ have demonstrated that four 5 min cycles of cuff inflation/deflation to the arm during the ongoing ischaemia administered in the ambulance, improved the myocardial salvage index in STEMI patients receiving primary PCI.

5.2 Potential for pharmacological postconditioning

Although coronary angioplasty is largely used for the treatment of AMI, a large number of patients are reperfused by thrombolysis or

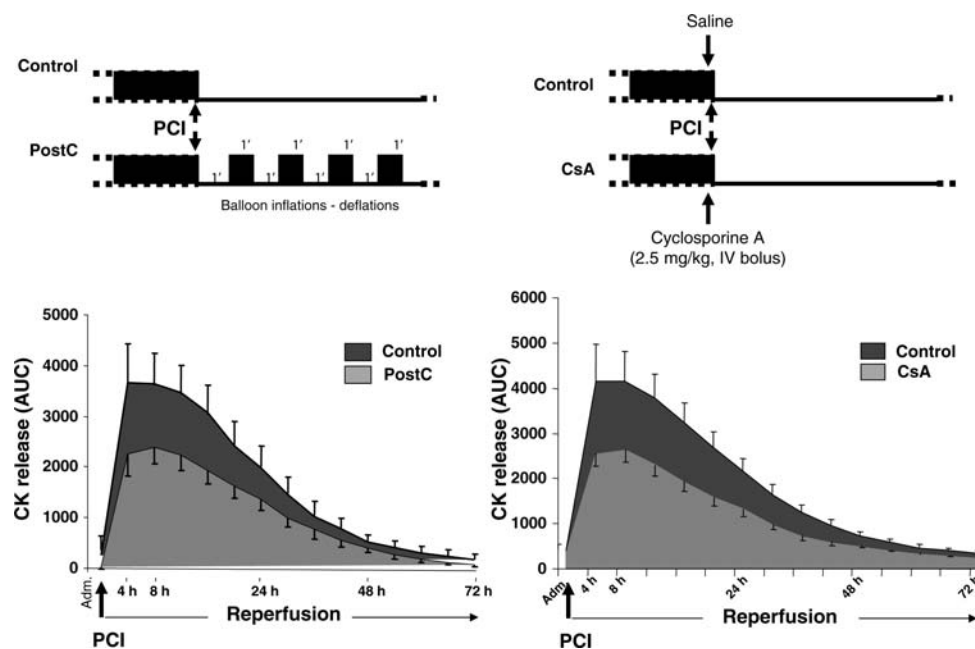


Figure 4 Ischaemic or pharmacological (cyclosporine A) postconditioning in STEMI patients. Left panel (top): STEMI patients treated by PCI (primary coronary intervention) were randomly assigned to receive either no further intervention (control) or four cycles of 1 min angioplasty balloon inflation followed by 1 min balloon deflation, starting less than 1 min after direct stenting. Right panel (top): STEMI patients treated by PCI (primary coronary intervention) were randomly assigned to receive either an IV injection of saline (control) or an IV bolus administration of 2.5 mg/kg of cyclosporine A (CsA). Left and right panels (bottom): area under the curve of creatine kinase (CK) release during the first 3 days of reperfusion in control vs. postconditioned (PostC: left) or cyclosporine A (CsA: right) treated patients.

Table 1 Clinical studies of ischaemic and pharmacological postconditioning in ST-elevation MI patients

Reference	Patient selection	Time (min)	n	Protocol	Effect
210	<12 h LAD, RCA, LCx	341	17	2 cycles of 90 s R/I	Improved ST-segment resolution. Improved coronary flow velocity. No difference in peak CK
190	<6 h LAD or RCA	318	30	4 cycles of 60 s R/I	Improved ST-segment resolution. Improved MBG, 36% reduction in 72 h CK
211	<12 h LAD, RCA, LCx	396	94	3 cycles of 30 s R/I	Improved WMSI, endothelial function. Less CK (P = NS)
212	<12 h LAD, RCA, LCx	312	41	3 cycles of 30 s R/I	27% reduction in 72 h CK. 27% reduction in MI SPECT 1 week. LVEF from 44 to 54% at 1 week (P = NS)
191	<6 h LAD or RCA	283	38	4 cycles of 60 s R/I	40% reduction in CK. 47% reduction in troponin I. 39% reduction in MI SPECT 6 months. 7% increase in LVEF at 12 months
213	<6 h LAD	228	24	2 cycles of 90 s R/I	Improved ST-segment resolution. Improved coronary flow velocity
214	<12 h LAD, RCA, LCx	264	75	3 cycles of 30 or 60 s R/I	Reduced apoptosis at 7 days (serum fas ligand)
215	<12 h LAD, RCA, LCx	255	118	4 cycles of 30 s R/I	19% relative reduction of MI size 3 at months. 31% increase in myocardial salvage ratio. 41% reduction in patients developing heart failure
216	<12 h LAD, RCA, LCx	128	21	GLP-1 infusion for 72 h post-PCI	Improved LVEF
217	<12 h LAD, RCA, LCx	240	20 201	Glucose-insulin-potassium pre-PCI	No beneficial effect
200	<3.17 h LAD	128	1066	50 or 70 mg/kg/min adenosine IV <15 min of PCI	Reduction in 1-month mortality (9.4 vs. 4.2%). Reduction in 6-month mortality (11.2 vs. 7.3%). Reduction in 6-month composite clinical endpoint of death, in-hospital CHF, or rehospitalization for CHF (17.2 vs. 12.0%)
202	<12 h LAD, RCA, LCx	240	569	72 h carperitide (ANP) IV infusion post-PCI	15% reduced total CK. No difference in troponin T. Improved reperfusion
202	<12 h LAD, RCA, LCx	210	545	Nicorandil IV bolus, then 24 h infusion	No difference in total CK or LVEF
218	<6 h LAD	127	94	0.05–5.0 mg IV KAI-9803 pre-PCI	Trend for reduced CK, troponin I, SPECT MI
166	<12 h LAD, RCA	292	58	2.5 mg/kg CsA IV bolus pre-PCI	40% reduction in CK. 13% reduction in troponin I (P = NS). 20% reduction in MI by CMR (n = 27)
53	<12 h LAD, RCA	292	28	2.5 mg/kg CsA IV bolus pre-PCI	24% reduction in MI size on CMR at 6 months. Smaller increase in LV end-systolic volume
219	<12 h LAD, RCA, LCx	341	30	33 000 IU bolus prior to PCI then 24, 48 h later	30% reduction in CK-MB
220	<12 h LAD	341	57	50 IU/kg EPO bolus prior to PCI	No difference in infarct size by CK, CK-MB, or CMR at 4 days
221	<12 h LAD	241	171	80 vs. 10 mg oral atorva-statin prior to PCI	No difference in infarct size by CK-MB or troponin

Patient selection-time from onset of chest pain. ST-elevation myocardial infarction in LAD (left anterior descending) artery, RCA (right coronary artery), and Cx (circumflex) artery. Time denotes the average time from onset of chest pain to intervention. R/I, cycle of reperfusion/ischaemia; CK, creatine kinase; MBG, myocardial blush grade; WMSI, wall motion score index; MI, myocardial infarction; SPECT, single-photon emission computed tomography; LVEF, left ventricular ejection fraction; PCI, percutaneous coronary intervention; CMR, cardiovascular magnetic resonance; CHF, chronic heart failure; ANP, atrial natriuretic peptide; CsA, Cyclosporine-A; EPO, erythropoietin.

cannot benefit from PCI postconditioning for technical reasons. There is, therefore, an obvious need for a pharmacological mimetic to ischaemic postconditioning (Table 1). According to the narrow time window as defined in experimental studies, one has to consider that such a postconditioning drug should be administered before reperfusion, in order to be circulating at the time of onset of coronary reflow. In theory, it could be administered to all patients with ongoing AMI, either as an adjunct to thrombolysis or even just before coronary angioplasty (as an ischaemic postconditioning mimetic).

Clinical trial data from the AMISTAD-I, AMISTAD-II, and ADMIRE trials provide an uncertain picture with regard to the clinical use of adenosine and synthetic adenosine agonist AMP579.^{197–199} The largest of these trials, AMISTAD-II, has been comprehensively

re-analysed.²⁰⁰ The administration of adenosine as an adjunct to reperfusion was of greatest benefit in patients receiving early reperfusion [<3.17 h (median) from onset of chest pain]. Benefit was seen in terms of greater early and late survival, and a reduction in the composite endpoint of death or CHF at 6 months. However, it is not clear that this benefit of adenosine treatment was entirely attributable to IS limitation.

A number of other naturally occurring mediators display postconditioning properties when administered exogenously in experimental AMI, although their roles in ischaemic postconditioning are not established. One example which has been translated to clinical trial is the use of natriuretic peptides. Prevention of reperfusion injury by stimulation of the cGMP synthesis was proposed by García-Dorado.²⁰¹ In the J-WIND trial, recombinant ANP administration as adjunct to

reperfusion therapy was shown to produce a small but significant reduction in IS, and improved 6-month ejection fraction in a non-selected population of patients with AMI.²⁰² Although aspects of the trial design were unsatisfactory, the result nevertheless is in general agreement with experimental studies showing the postconditioning potential of natriuretic peptides.²⁰³ Of interest, nicorandil, which activates K_{ATP} channels and releases NO, was without any beneficial effect in the second arm of the J-WIND trial.

Pharmacological inhibitors of MPTP (including CsA) reduce IS in experimental *in vivo* preparations to a similar extent as ischaemic postconditioning,^{162,204} making MPTP inhibition an interesting therapeutic target. Recently, Piot et al.¹⁶⁶ addressed whether inhibition of MPTP may attenuate IS in AMI patients. They reported that CsA, administered as an intravenous bolus immediately before coronary artery reperfusion by PCI, could significantly reduce IS measured by cardiac enzyme release during the first 3 days of reperfusion and by MRI at day 5 after reflow. This suggests that pharmacological postconditioning can be as efficient as PCI postconditioning, and opens a new therapeutic area for all AMI patients.

The equivalence between ischaemic and pharmacological postconditioning is based on: (i) comparable experimental protocols with the protective intervention being made either immediately before or within the first minute after reperfusion, and (ii) a comparable benefit in terms of IS reduction. In clinical practice, the only available data in the postconditioning era come from the cyclosporine study by Piot et al.,¹⁶⁶ which can be compared with angioplasty postconditioning data. The IS reduction was quite similar in these trials. Whether a postconditioning drug may be more efficient than angioplasty postconditioning could be worth looking at in future trials; angioplasty postconditioning could then be considered a reference protective intervention for the potential of IS reduction in AMI patients.

6. Conclusions/positions

- (1) There is accumulating evidence that lethal reperfusion injury is reduced by ischaemic or pharmacological postconditioning in animals and in small-size clinical trials.
- (2) The evidence remains, however, insufficient to recommend a widespread clinical use of ischaemic or pharmacological postconditioning to reduce IS in clinical practice, and the enthusiasm arising from these encouraging proof-of-concept studies must be tempered by the need to demonstrate an actual clinical benefit in larger scale clinical studies.
- (3) The extent of protection by ischaemic postconditioning is dependent on the duration of index ischaemia and the postconditioning protocol used. Species- and inter-individual variations are obvious. Whether the same is true for pharmacological postconditioning is unknown.
- (4) The signalling pathway(s) involve triggers, protein kinases, and end-effector(s): the contribution of each of these players may vary depending on the experimental model (cell type, *in-vitro*, *in-vivo*) and, again, the postconditioning stimulus used. More importantly, there appear to be redundant pathways which are activated and interact.
- (5) At present, there is insufficient information to judge how and to what extent confounding factors, including the classical CAD risk factors, drug treatments, gender, and age influence postconditioning efficacy.

- (6) There are various approaches to conditioning the heart and other organs against reperfusion injury.

7. Perspectives

- (1) There is a need to define the optimal protective protocol for a given duration of the index ischaemia.
- (2) There is a need to define the interdependence of the signalling pathways (all-or-nothing phenomenon) and the optimal molecular signalling target for protection.
- (3) There is a critical need for pre-clinical and clinical studies that examine cardioprotection specifically in relation to complicating disease states and their drug treatments.
- (4) There is a critical need for clinical research, including large-multicentre studies, to determine the impact of postconditioning on patient outcomes.

8. Designing future infarct size reduction trials

Before designing clinical studies, it is necessary to perform a thorough analysis of the conditions of success of these positive small-size trials, and put these into perspective with the numerous negative IS reduction studies performed during the past two decades (for review see^{22,205}). It would be insufficient to simply transfer a postconditioning regimen to the AMI clinical setting without taking into consideration the major determinants of IS, namely: the timing of application of the protective intervention with respect to the onset of reflow; the duration of ischaemia; the size of the area at risk; and the extent of the collateral circulation.⁶

As mentioned earlier, the time window for ischaemic postconditioning is a critical factor.³⁵ During PCI postconditioning, the first angioplasty balloon re-inflation was always performed within the first minute after direct stenting of the culprit coronary artery; in the case of CsA-induced protection, the drug was administered before re-opening of the coronary artery. Also, only patients with a fully occluded culprit coronary artery (TIMI flow grade at admission coronary angiography of 0 or 1) were included, thereby eliminating all patients who had undergone spontaneous reperfusion before PCI, i.e. those who had already been exposed to reperfusion injury before the protective intervention.

Taking into account the size of the area at risk is a key issue for all IS reduction studies. The studies by Staat et al.,¹⁹⁰ Thibault et al.,¹⁹¹ and Piot et al.,¹⁶⁶ who used area at risk as a covariate for analysing IS reduction, demonstrated that the larger the area at risk, the greater the myocardial salvage in patients with AMI. One may even question whether IS reduction by postconditioning is a real issue in patients with a small area at risk, who will anyway develop small infarcts and have a very good cardiovascular prognosis. In contrast, when the area at risk is larger, any additional myocardial salvage becomes clinically relevant. One can expect that postconditioned patients will less often develop adverse LV remodelling and possibly have a lower risk to develop heart failure; this latter issue needs to be addressed in future large-scale trials.⁵³ Obviously, the technique used to measure area at risk in patients with AMI is critical and remains an unresolved issue in the clinical settings of STEMI.

Although SPECT imaging with ⁹⁹Tc-sestamibi may be of interest here, its application remains difficult on a daily basis for the care of

AMI patients.¹⁹⁶ Recent pre-clinical reports suggest that T2-weighted MRI is able to measure area at risk, based on the detection of oedema in the early hours after reperfusion.²⁰⁶ Although very encouraging, this approach might not be appropriate for IS reduction studies since therapeutic interventions that reduce IS, including ischaemic postconditioning, also reduce reperfusion oedema.^{207,208} Any strategy based on the assessment of oedema may then underestimate the size of area at risk in the treated, but not in the control group, and thus bias the evaluation of myocardial salvage. Clearly, further work is needed to find the appropriate technique to assess this major determinant of IS in AMI patients.

Collateral flow is the third major determinant of IS. Patients with visible collaterals at admission coronary angiography are protected and develop smaller infarcts even in the absence of any protective interventions. Since it remains almost impossible to assess myocardial blood flow in acute clinical situations, these patients should be excluded from trials exploring the efficiency of IS-reduction interventions. Interestingly, when considering these confounding factors, the patients who benefitted most from the therapeutic intervention (postconditioning or CsA) were those who displayed larger areas at risk, lower collateral flow, and a fully occluded coronary artery at admission.^{166,196} Taking into account these major determinants of IS into the design of future trials will help us improve their accuracy and power to explore future treatments aimed at preventing lethal reperfusion injury.

Conflict of interest: none declared.

Funding

The work was supported by a travel grant of the European Society of Cardiology.

References

- Page DL, Caulfield JB, Kastor JA, Desanctis RW, Sanders CA. Myocardial changes associated with cardiogenic shock. *N Engl J Med* 1971;**285**:133–137.
- Sobel BE, Bresnahan GF, Shell WE, Yoder RD. Estimation of infarct size in man and its relation to prognosis. *Circulation* 1972;**46**:640–648.
- Burns RJ, Gibbons RJ, Yi Q, Roberts RS, Miller TD, Schaer GL et al. The relationships of left ventricular ejection fraction, end-systolic volume index and infarct size to six-month mortality after hospital discharge following myocardial infarction treated by thrombolysis. *J Am Coll Cardiol* 2002;**39**:30–36.
- Bellandi F, Maioli M, Gallopin M, Toso A, Dabizzi RP. Increase of myocardial salvage and left ventricular function recovery with intracoronary abximizab downstream of the coronary occlusion in patients with acute myocardial infarction treated with primary coronary intervention. *Catheter Cardiovasc Interv* 2004;**62**:186–192.
- Sarpe N, Doughty R. Epidemiology of heart failure and ventricular dysfunction. *Lancet* 1998;**352**(Suppl. 1):S13–S17.
- Reimer KA, Lowe JE, Rasmussen MM, Jennings RB. The wavefront phenomenon of ischemic cell death. 1. Myocardial infarct size vs duration of coronary occlusion in dogs. *Circulation* 1977;**56**:786–794.
- Kloner RA, Ganote CE, Jennings RB. The 'no-reflow' phenomenon after temporary coronary occlusion in the dog. *J Clin Invest* 1974;**54**:1496–1508.
- Maroko PR, Libby P, Ginks WR, Bloor CM, Shell WE, Sobel BE et al. Coronary artery reperfusion. I. Early effects on local myocardial function and the extent of myocardial necrosis. *J Clin Invest* 1972;**51**:2710–2716.
- Ginks WR, Sybers HD, Maroko PR, Covell JW, Sobel BE, Ross J Jr. Coronary artery reperfusion. II. Reduction of myocardial infarct size at 1 week after the coronary occlusion. *J Clin Invest* 1972;**51**:2717–2723.
- Chazov EI, Matveeva LS, Mazzaev AV, Sargin KE, Sadovskaia GV, Ruda MI. Intracoronary administration of fibrinolysin in acute myocardial infarct. *Ter Arkh* 1976;**48**:8–19.
- Rentrop KP, Blanke H, Karsch KR, Wiegand V, Kosterling H, Oster H et al. Acute myocardial infarction: intracoronary application of nitroglycerin and streptokinase. *Clin Cardiol* 1979;**2**:354–363.
- DeWood MA, Spores J, Notske R, Mouser LT, Burroughs R, Golden MS et al. Prevalence of total coronary occlusion during the early hours of the transmural myocardial infarction. *N Engl J Med* 1980;**303**:897–902.
- Rentrop P, Blanke H. Percutaneous transluminal coronary artery recanalization in evolving myocardial infarction. *Cardiovasc Intervent Radiol* 1982;**5**:194–196.
- Gersh BJ, Stone GW, White HD, Holmes DR Jr. Pharmacological facilitation of primary percutaneous coronary intervention for acute myocardial infarction: is the slope of the curve the shape of the future? *JAMA* 2005;**293**:979–986.
- Bolli R, Becker L, Gross G, Mentzer R Jr, Balshaw D, Lathrop DA. Myocardial protection at a crossroads. The need for translation into clinical therapy. *Circ Res* 2004;**95**:125–134.
- Jennings RB, Sommers HM, Smyth GA, Flack HA, Linn H. Myocardial necrosis induced by temporary occlusion of a coronary artery in the dog. *Arch Pathol* 1960;**70**:68–78.
- Przyklenk K. Lethal myocardial 'reperfusion injury': the opinions of good men. *J Thromb Thrombolysis* 1997;**4**:5–6.
- Braunwald E, Kloner RA. Myocardial reperfusion: a double-edged sword? *J Clin Invest* 1985;**76**:1713–1719.
- Vinten-Johansen J, Johnston WE, Mills SA, Faust KB, Geisinger KR, DeMasi RJ et al. Reperfusion injury after temporary coronary occlusion. *J Thorac Cardiovasc Surg* 1988;**95**:960–968.
- Piper HM, Garcia-Dorado D, Ovize M. A fresh look at reperfusion injury. *Cardiovasc Res* 1998;**38**:291–300.
- Yellon DM, Hausenloy DJ. Myocardial reperfusion injury. *N Engl J Med* 2007;**357**:1121–1135.
- Miura T, Miki T. Limitation of myocardial infarct size in the clinical setting: current status and challenges in translating animal experiments into clinical therapy. *Basic Res Cardiol* 2008;**103**:501–513.
- Na HS, Kim YI, Yoon YW, Han HC, Nahm SH, Hong SK. Ventricular premature beat-driven intermittent restoration of coronary blood flow reduces the incidence of reperfusion-induced ventricular fibrillation in a cat model of regional ischemia. *Am Heart J* 1996;**132**:78–83.
- Zhao Z-Q, Corvera JS, Halkos ME, Kerendi F, Wang N-P, Guyton RA et al. Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 2003;**285**:H579–H588.
- Halkos ME, Kerendi F, Corvera JS, Wang NP, Kin H, Payne CS et al. Myocardial protection with postconditioning is not enhanced by ischemic preconditioning. *Ann Thorac Surg* 2004;**78**:961–969.
- Skyschally A, van Caster P, Iliodromitis EK, Schulz R, Kremastinos DT, Heusch G. Ischemic postconditioning—experimental models and protocol algorithms. *Basic Res Cardiol* 2009;**104**:469–483.
- Iliodromitis EK, Downey JM, Heusch G, Kremastinos DT. What is the optimal post-conditioning algorithm? *J Cardiovasc Pharmacol Ther* 2009;**14**:269–273.
- Schaper W, Görgö G, Winkler B, Schaper J. The collateral circulation of the heart. *Prog Cardiovasc Dis* 1988;**31**:57–77.
- Shen Y-T, Fallon JT, Iwase M, Vatner SF. Innate protection of baboon myocardium: effects of coronary artery occlusion and reperfusion. *Am J Physiol* 1996;**270**:H1812–H1818.
- Yang XM, Liu Y, Liu Y, Tandon N, Kambayashi J, Downey JM et al. Attenuation of infarction in cynomolgus monkeys: preconditioning and postconditioning. *Basic Res Cardiol* 2010;**105**:119–128.
- Tahk SJ, Choi BJ, Choi SY, Yoon MH, Gwon HC, Hong GR et al. Distal protection device protects microvascular integrity during primary percutaneous intervention in acute myocardial infarction: a prospective, randomized, multicenter trial. *Int J Cardiol* 2008;**123**:162–168.
- Penna C, Tullio F, Merlino A, Moro F, Raimondo S, Rastaldo R et al. Postconditioning cardioprotection against infarct size and post-ischemic systolic dysfunction is influenced by gender. *Basic Res Cardiol* 2009;**104**:390–402.
- Rodriguez-Sinovas A, Cabestrero A, Garcia Del BB, Inserte J, Garcia A, Garcia-Dorado D. Intracoronary acid infusion as an alternative to ischemic postconditioning in pigs. *Basic Res Cardiol* 2009;**104**:761–771.
- Manintveld OC, Te Lintel HM, van den Bos EJ, Suurenbroek GM, Dekkers DH, Verdouw PD et al. Cardiac effects of postconditioning depend critically on the duration of index ischemia. *Am J Physiol Heart Circ Physiol* 2007;**292**:H1551–H1560.
- Kin H, Zhao Z-Q, Sun H-Y, Wang N-P, Corvera JS, Halkos ME et al. Postconditioning attenuates myocardial ischemia/reperfusion injury by inhibiting events in the early minutes of reperfusion. *Cardiovasc Res* 2004;**62**:74–85.
- Philipp S, Downey JM, Cohen MV. Postconditioning must be initiated in less than 1 minute following reperfusion and is dependent on adenosine receptors and PI3-kinase. *Circulation* 2004;**110**:III-168.
- Yang X-M, Proctor JB, Cui L, Krieg T, Downey JM, Cohen MV. Multiple, brief coronary occlusions during early reperfusion protect hearts by targeting cell signaling pathways. *J Am Coll Cardiol* 2004;**44**:1103–1110.
- Vinten-Johansen J, Yellon DM, Opie LH. Postconditioning. A simple, clinically applicable procedure to improve revascularization in acute myocardial infarction. *Circulation* 2005;**112**:2085–2088.
- Iliodromitis EK, Georgiadis M, Cohen MV, Downey JM, Bofilis E, Kremastinos DT. Protection from postconditioning depends on the number of short ischemic insults in anesthetized pigs. *Basic Res Cardiol* 2006;**101**:502–507.
- Chiari PC, Bienengraeber MW, Pagel PS, Krolkowski JG, Kersten JR, Wartler DC. Isoflurane protects against myocardial infarction during early reperfusion by activation of phosphatidylinositol-3-kinase signal transduction: evidence for anesthetic-induced postconditioning in rabbits. *Anesthesiology* 2005;**102**:102–109.

41. Heusch G, Schulz R. Neglect of the coronary circulation: some critical remarks on problems in the translation of cardioprotection. *Cardiovasc Res* 2009;**84**:11–14.
42. Kerendi F, Kin H, Halkos ME, Jiang R, Zhao Z-Q, Guyton RA et al. Remote postconditioning. Brief renal ischemia and reperfusion applied before coronary artery reperfusion reduces myocardial infarct size via endogenous activation of adenosine receptors. *Basic Res Cardiol* 2005;**100**:404–412.
43. Liu S, Wu XF, Zhang WZ, Sun YX, Cai SL. Remote postconditioning by brief renal ischemia and reperfusion reduces acute myocardial ischemia and reperfusion induced myocardial apoptosis in rabbits. *Zhonghua Xin Xue Guan Bing Za Zhi* 2007;**35**:757–760.
44. Li CM, Zhang XH, Ma XJ, Luo M. Limb ischemic postconditioning protects myocardium from ischemia/reperfusion injury. *Scand Cardiovasc J* 2006;**40**:312–317.
45. Gritsopoulos G, Iliodromitis EK, Zoga A, Farmakis D, Demerouti E, Papalois A et al. Remote postconditioning is more potent than classic postconditioning in reducing the infarct size in anesthetized rabbits. *Cardiovasc Drugs Ther* 2009;**23**:193–198.
46. Andreka G, Vertesaljai M, Szantho G, Font G, Piroth Z, Fontos G et al. Remote ischaemic postconditioning protects the heart during acute myocardial infarction in pigs. *Heart* 2007;**93**:749–752.
47. Dixon IM. Much ado about bone marrow stem cells: role in post-myocardial infarct repair. *Cardiovasc Res* 2006;**71**:609–611.
48. Bolli R. Mechanism of myocardial 'stunning'. *Circulation* 1990;**82**:723–738.
49. Guth BD, Schulz R, Heusch G. Time course and mechanisms of contractile dysfunction during acute myocardial ischemia. *Circulation* 1993;**87**(Suppl. 4):IV-35–IV-42.
50. Tawa M, Fukumoto T, Yamashita N, Ohkita M, Ayajiki K, Okamura T et al. Postconditioning improves postischemic cardiac dysfunction independently of norepinephrine overflow after reperfusion in rat hearts: comparison with preconditioning. *J Cardiovasc Pharmacol* 2010;**55**:6–13.
51. Insete J, Barba I, Hernando V, Garcia-Dorado D. Delayed recovery of intracellular acidosis during reperfusion prevents calpain activation and determines protection in postconditioned myocardium. *Cardiovasc Res* 2009;**81**:116–122.
52. Wang G, Zhang S, Joggerst SJ, McPherson J, Zhao DX. Effects of the number and interval of balloon inflations during primary PCI on the extent of myocardial injury in patients with STEMI: does postconditioning exist in real-world practice? *J Invasive Cardiol* 2009;**21**:451–455.
53. Mewton N, Croisille P, Gahide G, Rioufol G, Bonnefoy E, Sanchez I et al. Effect of cyclosporine on left ventricular remodeling after reperfused myocardial infarction. *J Am Coll Cardiol* 2010;**55**:1200–1205.
54. Galagudza M, Kurapeev D, Minasian S, Valen G, Vaage J. Ischemic postconditioning: brief ischemia during reperfusion converts persistent ventricular fibrillation into regular rhythm. *Eur J Cardiothorac Surg* 2004;**25**:1006–1010.
55. Sasaki H, Shimizu M, Ogawa K, Okazaki F, Taniguchi M, Taniguchi I et al. Brief ischemia/reperfusion performed after prolonged ischemia (ischemic postconditioning) can terminate reperfusion arrhythmias with no reduction of cardiac function in rats. *Int J Cardiol* 2007;**48**:205–213.
56. Kloner RA, Dow J, Bhandari A. Postconditioning markedly attenuates ventricular arrhythmias after ischemia/reperfusion. *J Cardiovasc Pharmacol Ther* 2006;**11**:55–63.
57. Dow J, Bhandari A, Kloner RA. Ischemic postconditioning's benefit on reperfusion ventricular arrhythmias is maintained in the senescent heart. *J Cardiovasc Pharmacol Ther* 2008;**13**:141–148.
58. Dow J, Bhandari A, Kloner RA. The mechanism by which ischemic postconditioning reduces reperfusion arrhythmias in rats remains elusive. *J Cardiovasc Pharmacol Ther* 2009;**14**:99–103.
59. Zhao H, Sapolsky RM, Steinberg GK. Interrupting reperfusion as a stroke therapy: ischemic postconditioning reduces infarct size after focal ischemia in rats. *J Cereb Blood Flow Metab* 2006;**26**:1114–1121.
60. Liu X, Chen H, Zhan B, Xing B, Zhou J, Zhu H et al. Attenuation of reperfusion injury by renal ischemic postconditioning: the role of NO. *Biochem Biophys Res Commun* 2007;**359**:628–634.
61. Serviddio G, Romano AD, Gesualdo L, Tamborra R, Di Palma AM, Rollo T et al. Postconditioning is an effective strategy to reduce renal ischaemia/reperfusion injury. *Nephrol Dial Transplant* 2008;**23**:1504–1512.
62. Rehni AK, Singh N. Role of phosphoinositide 3-kinase in ischemic postconditioning-induced attenuation of cerebral ischemia-evoked behavioral deficits in mice. *Pharmacol Rep* 2007;**59**:192–198.
63. Gao X, Zhang H, Takahashi T, Hsieh J, Liao J, Steinberg GK et al. The Akt signaling pathway contributes to postconditioning's protection against stroke; the protection is associated with the MAPK and PKC pathways. *J Neurochem* 2008;**105**:943–955.
64. Pignataro G, Meller R, Inoue K, Ordonez AN, Ashley MD, Xiong Z et al. In vivo and in vitro characterization of a novel neuroprotective strategy for stroke: ischemic postconditioning. *J Cereb Blood Flow Metab* 2008;**28**:232–241.
65. Eldaif SM, Deneve JA, Wang NP, Jiang R, Mosunjac M, Mutrie CJ et al. Attenuation of renal ischemia/reperfusion injury by postconditioning involves adenosine receptor and protein kinase C activation. *Transpl Int* 2010;**23**:217–226.
66. Deussen A, Möser G, Schrader J. Contribution of coronary endothelial cells to cardiac adenosine production. *Pflügers Arch* 1986;**406**:608–614.
67. Faigle M, Seessle J, Zug S, El Kasmi KC, Eltzschig HK. ATP release from vascular endothelia occurs across Cx43 hemichannels and is attenuated during hypoxia. *PLoS ONE* 2008;**3**:e2801.
68. Dörge H, Schulz R, Belosjorow S, Post H, van de Sand A, Konietzka I et al. Coronary microembolization: the role of TNF α in contractile dysfunction. *J Mol Cell Cardiol* 2002;**34**:51–62.
69. Schulz R, Aker S, Belosjorow S, Heusch G. TNF α in ischemia/reperfusion injury and heart failure. *Basic Res Cardiol* 2004;**99**:8–11.
70. Heusch G, Boengler K, Schulz R. Cardioprotection: nitric oxide, protein kinases, and mitochondria. *Circulation* 2008;**118**:1915–1919.
71. Granfeldt A, Jiang R, Wang N-P, Mykytenko J, Eldaif S, Deneve J et al. Inhibition of neutrophils is critical to the in vivo cardioprotection of postconditioning. *Circulation* 2008;**118**:S403.
72. Shandelya S, Kuppusamy P, Weisfeldt ML, Zweier JL. Evaluation of the role of polymorphonuclear leukocytes on contractile function in myocardial reperfusion injury. *Circulation* 1993;**87**:536–546.
73. Kevin LG, Camara AKS, Riess ML, Novalija E, Stowe DF. Ischemic preconditioning alters real-time measure of O $_2$ radicals in intact hearts with ischemia and reperfusion. *Am J Physiol Heart Circ Physiol* 2003;**284**:H566–H574.
74. Sun HY, Wang NP, Kerendi F, Halkos M, Kin H, Guyton RA et al. Hypoxic postconditioning reduces cardiomyocyte loss by inhibiting ROS generation and intracellular Ca $^{2+}$ overload. *Am J Physiol Heart Circ Physiol* 2005;**288**:H1900–H1908.
75. Kin H, Wang NP, Mykytenko J, Reeves J, Deneve J, Jiang R et al. Inhibition of myocardial apoptosis by postconditioning is associated with attenuation of oxidative stress-mediated nuclear factor- κ B translocation and TNF α release. *Shock* 2008;**29**:761–768.
76. Serviddio G, Di Venosa N, Federici A, D'Agostino D, Rollo T, Prigalio F et al. Brief hypoxia before normoxic reperfusion (postconditioning) protects the heart against ischemia/reperfusion injury by preventing mitochondria peroxide production and glutathione depletion. *FASEB J* 2005;**19**:354–361.
77. Kupai K, Csonka C, Fekete V, Odendaal L, van Rooyen RJ, Marais DW et al. Cholesterol diet-induced hyperlipidemia impairs the cardioprotective effect of postconditioning: role of peroxynitrite. *Am J Physiol Heart Circ Physiol* 2009;**297**:H1729–H1735.
78. Schulz R, Rose J, Post H, Heusch G. Involvement of endogenous adenosine in ischaemic preconditioning in swine. *Pflügers Arch* 1995;**430**:273–282.
79. Baxter GF, Hale SL, Miki T, Kloner RA, Cohen MV, Downey JM et al. Adenosine A $_1$ agonist at reperfusion trial (AART): results of a three-center, blinded, randomized, controlled experimental infarct study. *Cardiovasc Drugs Ther* 2000;**14**:607–614.
80. Burley DS, Baxter GF. Pharmacological targets revealed by myocardial postconditioning. *Curr Opin Pharmacol* 2009;**9**:177–188.
81. Kin H, Zatta AJ, Lofye MT, Amerson BS, Halkos ME, Kerendi F et al. Postconditioning reduces infarct size via adenosine receptor activation by endogenous adenosine. *Cardiovasc Res* 2005;**67**:124–133.
82. Donato M, D'Annunzio V, Berg G, Gonzalez G, Schreier L, Morales C et al. Ischemic postconditioning reduces infarct size by activation of A1 receptors and K $^{+}$ (ATP) channels in both normal and hypercholesterolemic rabbits. *J Cardiovasc Pharmacol* 2007;**49**:287–292.
83. Xi L, Das A, Zhao ZQ, Merino VF, Bader M, Kukreja RC. Loss of myocardial ischemic postconditioning in adenosine A1 and bradykinin B2 receptors gene knockout mice. *Circulation* 2008;**118**:S32–S37.
84. Yang X-M, Philipp S, Downey JM, Cohen MV. Postconditioning's protection is not dependent on circulating blood factors or cells but involves adenosine receptors and requires PI3-kinase and guanylyl cyclase activation. *Basic Res Cardiol* 2005;**100**:57–63.
85. Philipp S, Yang X-M, Cui L, Davis AM, Downey JM, Cohen MV. Postconditioning protects rabbit hearts through a protein kinase C-adenosine A $_{2b}$ receptor cascade. *Cardiovasc Res* 2006;**70**:308–314.
86. Morrison RR, Tan XL, Ledent C, Mustafa SJ, Hofmann PA. Targeted deletion of A $_{2A}$ adenosine receptors attenuates the protective effects of myocardial postconditioning. *Am J Physiol Heart Circ Physiol* 2007;**293**:H2523–H2529.
87. Xi J, McIntosh R, Shen X, Lee S, Chanoit G, Criswell H et al. Adenosine A $_{2A}$ and A $_{2B}$ receptors work in concert to induce a strong protection against reperfusion injury in rat hearts. *J Mol Cell Cardiol* 2009;**47**:684–690.
88. Zatta AJ, Kin H, Yoshishige D, Jiang R, Wang N, Reeves JG et al. Evidence that cardioprotection by postconditioning involves preservation of myocardial opioid content and selective opioid receptor activation. *Am J Physiol Heart Circ Physiol* 2008;**294**:H1444–H1451.
89. Jang Y, Xi J, Wang H, Mueller RA, Norfleet EA, Xu Z. Postconditioning prevents reperfusion injury by activating delta-opioid receptors. *Anesthesiology* 2008;**108**:243–250.
90. Gross ER, Hsu AK, Gross GJ. Opioid-induced cardioprotection occurs via glycogen synthase kinase β inhibition during reperfusion in intact rat hearts. *Circ Res* 2004;**94**:960–966.
91. Chen Z, Li T, Zhang B. Morphine postconditioning protects against reperfusion injury in the isolated rat hearts. *J Surg Res* 2008;**145**:287–294.
92. Gross ER, Peart JN, Hsu AK, Auchampach JA, Gross GJ. Extending the cardioprotective window using a novel delta-opioid agonist fentanyl isothiocyanate via the PI3-kinase pathway. *Am J Physiol Heart Circ Physiol* 2005;**288**:H2744–H2749.
93. Peart JN, Gross ER, Reichelt ME, Hsu A, Headrick JP, Gross GJ. Activation of kappa-opioid receptors at reperfusion affords cardioprotection in both rat and mouse hearts. *Basic Res Cardiol* 2008;**103**:454–463.

94. Baxter GF, Ebrahim Z. Role of bradykinin in preconditioning and protection of the ischaemic myocardium. *Brit J Pharmacol* 2002;**135**:843–854.
95. Penna C, Mancardi D, Rastaldo R, Losano G, Pagliaro P. Intermittent activation of bradykinin B(2) receptors and mitochondrial K(ATP) channels trigger cardiac postconditioning through redox signaling. *Cardiovasc Res* 2007;**75**:168–177.
96. Lim SY, Davidson SM, Hausenloy DJ, Yellon DM. Preconditioning and postconditioning: the essential role of the mitochondrial permeability transition pore. *Cardiovasc Res* 2007;**75**:530–535.
97. Penna C, Mancardi D, Tullio F, Pagliaro P. Postconditioning and intermittent bradykinin induced cardioprotection require cyclooxygenase activation and prostacyclin release during reperfusion. *Basic Res Cardiol* 2008;**103**:368–377.
98. Bell RM, Yellon DM. Bradykinin limits infarction when administered as an adjunct to reperfusion in mouse heart: the role of PI3K, Akt and eNOS. *J Mol Cell Cardiol* 2003;**35**:185–193.
99. Vandenberg JJ, Metcalfe JC, Grace AA. Mechanisms of pH recovery after global ischemia in the perfused heart. *Circ Res* 1993;**72**:993–1003.
100. Garcia-Dorado D, Ruiz-Meana M, Piper HM. Lethal reperfusion injury in acute myocardial infarction: facts and unresolved issues. *Cardiovasc Res* 2009;**83**:165–168.
101. Piper HM, Balsler C, Ladilov YV, Schäfer M, Siegmund B, Ruiz-Meana M et al. The role of Na⁺/H⁺ exchange in ischemia/reperfusion. *Basic Res Cardiol* 1996;**91**:191–202.
102. Ruiz-Maena M, Pina P, Garcia-Dorado D, Rodriguez-Sinovas A, Barba I, Miro-Casa E et al. Glycine protects cardiomyocytes against lethal reoxygenation injury by inhibiting mitochondrial permeability transition. *J Physiol* 2004;**558**:873–882.
103. Imahashi K, Kusuoka H, Hashimoto K, Yoshioka J, Yamaguchi H, Nishimura T. Intracellular sodium accumulation during ischemia as the substrate for reperfusion injury. *Circ Res* 1999;**84**:1401–1406.
104. Theroux P, Chaitman BR, Danchin N, Erhardt L, Meinertz T, Schroeder JS et al. Inhibition of the sodium-hydrogen exchanger with cariporide to prevent myocardial infarction in high-risk ischemic situations. Main results of the GUARDIAN trial. *Circulation* 2000;**102**:3032–3038.
105. Heusch G. Postconditioning. Old wine in a new bottle? *J Am Coll Cardiol* 2004;**44**:1111–1112.
106. Cohen MV, Yang XM, Downey JM. The pH hypothesis of postconditioning: staccato reperfusion reintroduces oxygen and perpetuates myocardial acidosis. *Circulation* 2007;**115**:1895–1903.
107. Cohen MV, Yang XM, Downey JM. Acidosis, oxygen, and interference with mitochondrial permeability transition pore formation in the early minutes of reperfusion are critical to postconditioning's success. *Basic Res Cardiol* 2008;**103**:464–471.
108. Inserte J, Barba I, Hernandez V, Abellán A, Ruiz-Meana M, Rodriguez-Sinovas A et al. Effect of acidic reperfusion on prolongation of intracellular acidosis and myocardial salvage. *Cardiovasc Res* 2008;**77**:782–790.
109. Argaud L, Gateau-Roesch O, Augeul L, Couture-Lepetit E, Loufouat J, Gomez L et al. Increased mitochondrial calcium coexists with decreased reperfusion injury in postconditioned (but not preconditioned) hearts. *Am J Physiol Heart Circ Physiol* 2008;**294**:H386–H391.
110. Inserte J, Garcia-Dorado D, Hernandez V, Soler-Soler J. Calpain-mediated impairment of Na⁺/K⁺ -ATPase activity during early reperfusion contributes to cell death after myocardial ischemia. *Circ Res* 2005;**97**:465–473.
111. Chen M, Won DJ, Krajewski S, Gottlieb RA. Calpain and mitochondria in ischemia/reperfusion injury. *J Biol Chem* 2002;**277**:29181–29186.
112. Zhao X, Newcomb JK, Posmantur RM, Wang KK, Pike BR, Hayes RL. pH dependency of mu-calpain and m-calpain activity assayed by casein zymography following traumatic brain injury in the rat. *Neurosci Lett* 1998;**247**:53–57.
113. Hernandez V, Inserte J, Satorio CL, Parra V, Poncelas-Nozal M, Garcia-Dorado D. Calpain translocation and activation as pharmacological targets during ischemia/reperfusion. *J Mol Cell Cardiol* 2010; doi:10.1016/j.yjmcc.2010.02.024. Published online ahead of print 6 March 2010.
114. Schulman D, Latchman DS, Yellon DM. Urocortin protects the heart from reperfusion injury via upregulation of p42/p44 MAPK signaling pathway. *Am J Physiol Heart Circ Physiol* 2002;**283**:H1481–H1488.
115. Hausenloy DJ, Yellon DM. New directions for protecting the heart against ischaemia/reperfusion injury: targeting the reperfusion injury salvage kinase (RISK)-pathway. *Cardiovasc Res* 2004;**61**:448–460.
116. Tsang A, Hausenloy DJ, Mocanu MM, Yellon DM. Postconditioning: a form of 'Modified Reperfusion' protects the myocardium by activating the phosphatidylinositol 3-kinase-Akt pathway. *Circ Res* 2004;**95**:230–232.
117. Zhu M, Feng J, Lucchinetti E, Fischer G, Xu L, Pedrazzini T et al. Ischemic postconditioning protects remodeled myocardium via the PI3K-PKB/Akt reperfusion injury salvage kinase pathway. *Cardiovasc Res* 2006;**72**:152–162.
118. Sivaraman V, Mudaligiri NR, Di SC, Kolvekar S, Hayward M, Yap J et al. Postconditioning protects human atrial muscle through the activation of the RISK pathway. *Basic Res Cardiol* 2007;**102**:453–459.
119. Juhaszova M, Zorov DB, Kim S-H, Pepe S, Fu Q, Fishbein KW et al. Glycogen synthase kinase-3 β mediates convergence of protection signaling to inhibit the mitochondrial permeability transition pore. *J Clin Invest* 2004;**113**:1535–1549.
120. Das S, Wong R, Rajapakse N, Murphy E, Steenbergen C. Glycogen synthase kinase 3 inhibition slows mitochondrial adenine nucleotide transport and regulates voltage-dependent anion channel phosphorylation. *Circ Res* 2008;**103**:983–991.
121. Gomez L, Paillard M, Thibault H, Derumeaux G, Ovize M. Inhibition of GSK3 β by postconditioning is required to prevent opening of the mitochondrial permeability transition pore during reperfusion. *Circulation* 2008;**117**:2761–2768.
122. Nishino Y, Webb IG, Davidson SM, Ahmed AI, Clark JE, Jacquet S et al. Glycogen synthase kinase-3 inactivation is not required for ischemic preconditioning or postconditioning in the mouse. *Circ Res* 2008;**103**:307–314.
123. Michel MC, Li Y, Heusch G. Mitogen-activated protein kinases in the heart. *Naunyn-Schmiedeberg's Arch Pharmacol* 2001;**363**:245–266.
124. Behrends M, Schulz R, Post H, Alexandrov A, Belosjorow S, Michel MC et al. Inconsistent relation of MAPK activation to infarct size reduction by ischemic preconditioning in pigs. *Am J Physiol Heart Circ Physiol* 2000;**279**:H1111–H1119.
125. Schulz R, Belosjorow S, Gres P, Jansen J, Michel MC, Heusch G. p38 MAP kinase is a mediator of ischemic preconditioning in pigs. *Cardiovasc Res* 2002;**55**:690–700.
126. Bassi R, Heads R, Marber MS, Clark JE. Targeting p38-MAPK in the ischaemic heart: kill or cure? *Curr Opin Pharmacol* 2008;**8**:141–146.
127. Sun H-Y, Wang N-P, Halkos M, Kerendi F, Kin H, Guyton RA et al. Postconditioning attenuates cardiomyocyte apoptosis via inhibition of JNK and p38 mitogen-activated protein kinase signaling pathways. *Apoptosis* 2006;**11**:1583–1593.
128. Heusch G. No RISK, no cardioprotection? A critical perspective. *Cardiovasc Res* 2009;**84**:173–174.
129. Schwartz LM, Lagranha CJ. Ischemic postconditioning during reperfusion activates AKT and ERK without protecting against lethal myocardial ischemia/reperfusion injury in pigs. *Am J Physiol Heart Circ Physiol* 2006;**290**:H1011–H1018.
130. Darling CE, Jiang R, Maynard M, Whittaker P, Vinten-Johansen J, Przyklenk K. Postconditioning via stuttering reperfusion limits myocardial infarct size in rabbit hearts: role of ERK1/2. *Am J Physiol Heart Circ Physiol* 2005;**289**:H1618–H1626.
131. Przyklenk K, Maynard M, Darling CE, Whittaker P. Aging mouse hearts are refractory to infarct size reduction with post-conditioning. *J Am Coll Cardiol* 2008;**51**:1393–1398.
132. Skyschally A, van Caster P, Boengler K, Gres P, Musiolik J, Schilawa D et al. Ischemic postconditioning in pigs: no causal role for RISK activation. *Circ Res* 2009;**104**:15–18.
133. Musiolik J, van Caster P, Skyschally A, Boengler K, Gres P, Schulz R et al. Reduction of infarct size by gentle reperfusion without activation of reperfusion injury salvage kinases in pigs. *Cardiovasc Res* 2010;**85**:110–117.
134. Boengler K, Hilfiker-Kleiner D, Drexler H, Heusch G, Schulz R. The myocardial JAK/STAT pathway: from protection to failure. *Pharmacol Therap* 2008;**120**:172–185.
135. Lecour S. Activation of the protective survivor activating factor enhancement (SAFE) pathway against reperfusion injury: does it go beyond the RISK path? *J Mol Cell Cardiol* 2009;**47**:32–40.
136. Lacerda L, Somers S, Opie HL, Lecour S. Ischemic postconditioning protects against reperfusion injury via the SAFE pathway. *Cardiovasc Res* 2009;**84**:201–208.
137. Boengler K, Buechert A, Heinen Y, Roeskes C, Hilfiker-Kleiner D, Heusch G et al. Cardioprotection by ischemic postconditioning is lost in aged and STAT3-deficient mice. *Circ Res* 2008;**102**:131–135.
138. Goodman MD, Koch SE, Fuller-Bicer GA, Butler KL. Regulating RISK: a role for JAK-STAT signaling in postconditioning? *Am J Physiol Heart Circ Physiol* 2008;**295**:H1649–H1656.
139. Schulz R, Heusch G. Tumor necrosis factor- α and its receptors 1 and 2: Yin and Yang in myocardial infarction? *Circulation* 2009;**119**:1355–1357.
140. Lecour S, Suleman N, Deuchar GA, Somers S, Lacerda L, Huisamen B et al. Pharmacological preconditioning with tumor necrosis factor- α activates signal transducer and activator of transcription-3 at reperfusion without involving classic prosurvival kinases (Akt and extracellular signal-regulated kinase). *Circulation* 2005;**112**:3911–3918.
141. Vessey DA, Kelley M, Li L, Huang Y, Zhou HZ, Zhu BQ et al. Role of sphingosine kinase activity in protection of heart against ischemia reperfusion injury. *Med Sci Monit* 2006;**12**:BR318–BR324.
142. Jin ZQ, Karliner JS, Vessey DA. Ischaemic postconditioning protects isolated mouse hearts against ischaemia/reperfusion injury via sphingosine kinase isoform-1 activation. *Cardiovasc Res* 2008;**79**:134–140.
143. Thielmann M, Dörge H, Martin C, Belosjorow S, Schwanke U, van de Sand A et al. Myocardial dysfunction with coronary microembolization: signal transduction through a sequence of nitric oxide, tumor necrosis factor- α and sphingosine. *Circ Res* 2002;**90**:807–813.
144. Penna C, Rastaldo R, Mancardi D, Raimondo S, Cappello S, Gattullo D et al. Postconditioning induced cardioprotection requires signaling through a redox-sensitive mechanism, mitochondrial ATP-sensitive K⁺ channel and protein kinase C activation. *Basic Res Cardiol* 2006;**101**:180–189.
145. Zatta AJ, Kin H, Lee G, Wang N, Jiang R, Lust R et al. Infarct-sparing effect of myocardial postconditioning is dependent on protein kinase C signalling. *Cardiovasc Res* 2006;**70**:315–324.
146. Burley DS, Ferdinandy P, Baxter GF. Cyclic GMP and protein kinase-G in myocardial ischaemia/reperfusion: opportunities and obstacles for survival signaling. *Br J Pharmacol* 2007;**152**:855–869.
147. Penna C, Cappello S, Mancardi D, Raimondo S, Rastaldo R, Gattullo D et al. Postconditioning reduces infarct size in the isolated rat heart: role of coronary flow and pressure and the nitric oxide/cGMP pathway. *Basic Res Cardiol* 2006;**101**:168–179.

148. Zamzami N, Marchetti P, Castedo M, Decaudin D, Macho A, Hirsch T et al. Sequential reduction of mitochondrial transmembrane potential and generation of reactive oxygen species in early programmed cell death. *J Exp Med* 1995;**182**:367–377.
149. Bernardi P, Petronilli V, Di LF, Forte M. A mitochondrial perspective on cell death. *Trends Biochem Sci* 2001;**26**:112–117.
150. Heusch G, Boengler K, Schulz R. Inhibition of mitochondrial permeability transition pore opening: the holy grail of cardioprotection. *Basic Res Cardiol* 2010;**105**:151–154.
151. Di Lisa F, Bernardi P. A CaPful of mechanisms regulating the mitochondrial permeability transition. *J Mol Cell Cardiol* 2009;**46**:775–780.
152. Rasola A, Bernardi P. The mitochondrial permeability transition pore and its involvement in cell death and in disease pathogenesis. *Apoptosis* 2007;**12**:815–833.
153. Baines CP. The mitochondrial permeability transition pore and ischemia/reperfusion injury. *Basic Res Cardiol* 2009;**104**:181–188.
154. Basso E, Fante L, Fowlkes J, Petronilli V, Forte MA, Bernardi P. Properties of the permeability transition pore in mitochondria devoid of Cyclophilin D. *J Biol Chem* 2005;**280**:18558–18561.
155. Halestrap AP, Pasdois P. The role of the mitochondrial permeability transition pore in heart disease. *Biochim Biophys Acta* 2009;**1787**:1402–1415.
156. Di LF, Kaludercic N, Carpi A, Menabo R, Giorgio M. Mitochondrial pathways for ROS formation and myocardial injury: the relevance of p66(Shc) and monoamine oxidase. *Basic Res Cardiol* 2009;**104**:131–139.
157. Argaud L, Gateau-Roesch O, Raisky O, Loufouat J, Robert D, Ovize M. Postconditioning inhibits mitochondrial permeability transition. *Circulation* 2005;**111**:194–197.
158. Hausenloy DJ, Yellon DM, Mani-Babu S, Duchon MR. Preconditioning protects by inhibiting the mitochondrial permeability transition. *Am J Physiol Heart Circ Physiol* 2004;**287**:H841–H849.
159. Hausenloy DJ, Yellon DM. Preconditioning and postconditioning: united at reperfusion. *Pharmacol Ther* 2007;**116**:173–191.
160. Javadov SA, Clarke S, Das M, Griffiths EJ, Lim KH, Halestrap AP. Ischaemic preconditioning inhibits opening of mitochondrial permeability transition pores in the reperfused rat heart. *J Physiol* 2003;**549**:513–524.
161. Halestrap AP, Clarke SJ, Khaliulin I. The role of mitochondria in protection of the heart by preconditioning. *Biochim Biophys Acta* 2007;**1767**:1007–1031.
162. Argaud L, Gateau-Roesch O, Muntean D, Chabalbreysse L, Loufouat J, Robert D et al. Specific inhibition of the mitochondrial permeability transition prevents lethal reperfusion injury. *J Mol Cell Cardiol* 2005;**38**:367–374.
163. Griffiths EJ, Halestrap AP. Protection by Cyclosporin A of ischemia/reperfusion-induced damage in isolated rat hearts. *J Mol Cell Cardiol* 1993;**25**:1461–1469.
164. Griffiths EJ, Halestrap AP. Mitochondrial non-specific pores remain closed during cardiac ischaemia, but open upon reperfusion. *Biochem J* 1995;**307**:93–98.
165. Di Lisa F, Bernardi P. Mitochondrial and ischemia/reperfusion injury of the heart: Fixing a hole. *Cardiovasc Res* 2006;**70**:191–199.
166. Piot C, Croisille P, Staat P, Thibault H, Rioufol G, Mewton N et al. Effect of cyclosporine on reperfusion injury in acute myocardial infarction. *N Engl J Med* 2008;**359**:473–481.
167. Steenbergen C, Das S, Su J, Wong R, Murphy E. Cardioprotection and altered mitochondrial adenine nucleotide transport. *Basic Res Cardiol* 2009;**104**:149–156.
168. Costa AD, Jakob R, Costa CL, Andrukiv K, West IC, Garlid KD. The mechanism by which the mitochondrial ATP-sensitive K⁺ channel opening and H₂O₂ inhibit the mitochondrial permeability transition. *J Biol Chem* 2006;**281**:20801–20808.
169. Boengler K, Dodoni G, Ruiz-Meana M, Cabestrero A, Rodriguez-Sinovas A, Garcia-Dorado D et al. Connexin 43 in cardiomyocyte mitochondria and its increase by ischemic preconditioning. *Cardiovasc Res* 2005;**67**:234–244.
170. Heinzl FR, Luo Y, Li X, Boengler K, Buechert A, Garcia-Dorado D et al. Impairment of diazoxide-induced formation of reactive oxygen species and loss of cardioprotection in connexin 43 deficient mice. *Circ Res* 2005;**97**:583–586.
171. Heusch G, Büchert A, Feldhaus S, Schulz R. No loss of cardioprotection by postconditioning in connexin 43-deficient mice. *Basic Res Cardiol* 2006;**101**:354–356.
172. Crisostomo PR, Wang M, Wairiuko GM, Terrell AM, Meldrum DR. Postconditioning in females depends on injury severity. *J Surg Res* 2006;**134**:342–347.
173. Dow J, Kloner RA. Postconditioning does not reduce myocardial infarct size in an in vivo regional ischemia rodent model. *J Cardiovasc Pharmacol Ther* 2007;**12**:153–163.
174. Boengler K, Schulz R, Heusch G. Loss of cardioprotection with ageing. *Cardiovasc Res* 2009;**83**:247–261.
175. Yin Z, Gao H, Wang H, Li L, Di C, Luan R et al. Ischemic postconditioning protects both adult and aged Sprague Dawley rat hearts from ischemia/reperfusion injury through the PI3-K/Akt and GSK-3beta pathway. *Clin Exp Pharmacol Physiol* 2009;**36**:756–763.
176. Ferdinandy P, Schulz R, Baxter GF. Interaction of cardiovascular risk factors with myocardial ischemia/reperfusion injury, preconditioning, and postconditioning. *Pharmacol Rev* 2007;**59**:418–458.
177. Ferdinandy P. Myocardial ischaemia/reperfusion injury and preconditioning: effects of hypercholesterolaemia/hyperlipidaemia. *Br J Pharmacol* 2003;**138**:283–285.
178. Iliodromitis EK, Zoga A, Vrettou A, Andreadou I, Paraskevaidis IA, Kakkamanis L et al. The effectiveness of postconditioning and preconditioning on infarct size in hypercholesterolemic and normal anesthetized rabbits. *Atherosclerosis* 2006;**188**:356–362.
179. Giricz Z, Gorbe A, Pipis J, Burley DS, Ferdinandy P, Baxter GF. Hyperlipidaemia induced by a high-cholesterol diet leads to the deterioration of guanosine-3',5'-cyclic monophosphate/protein kinase G-dependent cardioprotection in rats. *Br J Pharmacol* 2009;**158**:1495–1502.
180. Wagner C, Kloeting I, Strasser RH, Weinbrenner C. Cardioprotection by postconditioning is lost in WOKW rats with metabolic syndrome: role of glycogen synthase kinase 3beta. *J Cardiovasc Pharmacol* 2008;**52**:430–437.
181. Bouhidel O, Pons S, Souktani R, Zini R, Berdeaux A, Ghaleb B. Myocardial ischemic postconditioning against ischemia/reperfusion is impaired in ob/ob mice. *Am J Physiol Heart Circ Physiol* 2008;**295**:H1580–H1586.
182. Huhn R, Heinen A, Hollmann MW, Schlack W, Preckel B, Weber NC. Cyclosporine A administered during reperfusion fails to restore cardioprotection in prediabetic Zucker obese rats in vivo. *Nutr Metab Cardiovasc Dis* 2009; doi:10.1016/j.numecd.2009.06.010. Published online ahead of print 9 October 2009.
183. Penna C, Tullio F, Moro F, Folino A, Merlino A, Pagliaro P. Effects of a protocol of ischemic postconditioning and/or captopril in hearts of normotensive and hypertensive rats. *Basic Res Cardiol* 2010;**105**:181–192.
184. Feng J, Fischer G, Lucchinetti E, Zhu M, Bestmann L, Jegger D et al. Infarct-remodeled myocardium is receptive to protection by isoflurane postconditioning: role of protein kinase B/Akt signaling. *Anesthesiology* 2006;**104**:1004–1014.
185. Lange M, Redel A, Lotz C, Smul TM, Blomeyer C, Frank A et al. Desflurane-induced postconditioning is mediated by beta-adrenergic signaling: role of beta 1- and beta 2-adrenergic receptors, protein kinase A, and calcium/calmodulin-dependent protein kinase II. *Anesthesiology* 2009;**110**:516–528.
186. Oikawa M, Yaoita H, Watanabe K, Maruyama Y. Attenuation of cardioprotective effect by postconditioning in coronary stenosed rat heart and its restoration by carvedilol. *Circ J* 2008;**72**:2081–2086.
187. Heusch G, Skyschally A, Gres P, van Caster P, Schilawa D, Schulz R. Improvement of regional myocardial blood flow and function and reduction of infarct size with ivabradine—protection beyond heart rate reduction. *Eur Heart J* 2008;**29**:2265–2275.
188. Kocsis GF, Pipis J, Fekete V, Kovacs-Simon A, Odendaal L, Molnar E et al. Lovastatin interferes with the infarct size-limiting effect of ischemic preconditioning and postconditioning in rat hearts. *Am J Physiol Heart Circ Physiol* 2008;**294**:H2406–H2409.
189. Obal D, Dettwiler S, Favocchia C, Scharbatke H, Preckel B, Schlack W. The influence of mitochondrial KATP-channels in the cardioprotection of preconditioning and postconditioning by sevoflurane in the rat in vivo. *Anesth Analg* 2005;**101**:1252–1260.
190. Staat P, Rioufol G, Piot C, Cottin Y, Cung TT, L'Huillier I et al. Postconditioning the human heart. *Circulation* 2005;**112**:2143–2148.
191. Thibault H, Piot C, Staat P, Bontemps L, Sportouch C, Rioufol G et al. Long-term benefit of postconditioning. *Circulation* 2008;**117**:1037–1044.
192. Darling CE, Solari PB, Smith CS, Furman MI, Przyklen K. 'Postconditioning' the human heart: multiple balloon inflations during primary angioplasty may confer cardioprotection. *Basic Res Cardiol* 2007;**102**:274–278.
193. Hori M, Kitakaze M, Sato S, Takashima S, Iwakura K, Inoue A et al. Staged reperfusion attenuates myocardial stunning in dogs. Role of transient acidosis during early reperfusion. *Circulation* 1991;**84**:2135–2145.
194. Takeo S, Liu J-X, Tanonaka K, Nasa Y, Yabe K, Tanahashi H et al. Reperfusion at reduced flow rates enhances postischemic contractile recovery of perfused heart. *Am J Physiol* 1995;**268**:H2384–H2395.
195. Loukogeorgakis SP, Williams R, Panagiotidou AT, Kolvkar SK, Donald A, Cole TJ et al. Transient limb ischemia induces remote preconditioning and remote postconditioning in humans by a KATP channel dependent mechanism. *Circulation* 2007;**116**:1386–1395.
196. Botker HE, Kharbanda RK, Schmidt MR, Bottcher M, Kaltoft AK, Terkelsen CJ et al. Prehospital remote ischaemic conditioning increases myocardial salvage in acute myocardial infarction. *Lancet* 2010;**375**:1090–1099.
197. Mahaffey KW, Puma JA, Barbagelata A, DiCarli MF, Leeser MA, Browne KF et al. Adenosine as an adjunct thrombolytic therapy for acute myocardial infarction. *J Am Coll Cardiol* 1999;**34**:1711–1720.
198. Kopecky SL, Aviles RJ, Bell MR, Lobl JK, Tipping D, Frommell G et al. A randomized, double-blinded, placebo-controlled, dose-ranging study measuring the effect of an adenosine agonist on infarct size reduction in patients undergoing primary percutaneous transluminal coronary angioplasty: the ADMIRE (AmpP579 Delivery for Myocardial Infarction Reduction) study. *Am Heart J* 2003;**146**:146–152.
199. Ross AM, Gibbons R-J, Stone GW, Kloner RA, Alexander RW. A randomized, double-blinded placebo-controlled multicenter trial of adenosine as an adjunct to reperfusion in the treatment of acute myocardial infarction (AMISTAD-II). *J Am Coll Cardiol* 2005;**45**:1775–1780.
200. Kloner RA, Forman MB, Gibbons RJ, Ross AM, Alexander RW, Stone GW. Impact of time to therapy and reperfusion modality on the efficacy of adenosine in acute myocardial infarction: the AMISTAD-2 trial. *Eur Heart J* 2006;**27**:2400–2405.
201. Garcia-Dorado D, Agullo L, Sartorio CL, Ruiz-Meana M. Myocardial protection against reperfusion injury: the cGMP pathway. *Thromb Haemostasis* 2009;**101**:635–642.
202. Kitakaze M, Asakura M, Kim J, Shintani Y, Asanuma H, Hamasaki T et al. Human atrial natriuretic peptide and nicorandil as adjuncts to reperfusion treatment for acute myocardial infarction (J-WIND): two randomised trials. *Lancet* 2007;**370**:1483–1493.

203. Burley DS, Hamid SA, Baxter GF. Cardioprotective actions of peptide hormones in myocardial ischemia. *Heart Fail Rev* 2007;**12**:279–291.
204. Hausenloy DJ, Maddock HL, Baxter GF, Yellon DM. Inhibiting mitochondrial permeability transition pore opening: a new paradigm for myocardial preconditioning? *Cardiovasc Res* 2002;**55**:534–543.
205. Downey JM, Cohen MV. Why do we still not have cardioprotective drugs? *Circ J* 2009;**73**:1171–1177.
206. Aletras AH, Tilak GS, Natanzon A, Hsu LY, Gonzalez FM, Hoyt RF Jr et al. Retrospective determination of the area at risk for reperfused acute myocardial infarction with T2-weighted cardiac magnetic resonance imaging: histopathological and displacement encoding with stimulated echoes (DENSE) functional validations. *Circulation* 2006;**113**:1865–1870.
207. Sanz E, Dorado DG, Oliveras J, Barrabés JA, Gonzalez MA, Ruiz-Meana M et al. Dissociation between anti-infarct effect and anti-edema effect of ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 1995;**37**:H233–H241.
208. Garcia-Dorado D, Oliveras J. Myocardial edema: a preventable cause of reperfusion injury. *Cardiovasc Res* 1993;**27**:1555–1563.
209. Garcia-Dorado D, Piper HM. Postconditioning: Reperfusion of 'reperfusion injury' after hibernation. *Cardiovasc Res* 2006;**69**:1–3.
210. Laskey WK. Brief repetitive balloon occlusions enhance reperfusion during percutaneous coronary intervention for acute myocardial infarction: A Pilot Study. *Cathet Cardiovasc Intervent* 2005;**65**:361–367.
211. Ma X, Zhang X, Li C, Luo M. Effect of postconditioning on coronary blood flow velocity and endothelial function and LV recovery after myocardial infarction. *J Interv Cardiol* 2006;**19**:367–375.
212. Yang XC, Liu Y, Wang LF, Cui L, Wang T, Ge YG et al. Reduction in myocardial infarct size by postconditioning in patients after percutaneous coronary intervention. *J Invasive Cardiol* 2007;**19**:424–430.
213. Laskey WK, Yoon S, Calzada N, Ricciardi MJ. Concordant improvements in coronary flow reserve and ST-segment resolution during percutaneous coronary intervention for acute myocardial infarction: a benefit of postconditioning. *Catheter Cardiovasc Interv* 2008;**72**:212–220.
214. Zhao WS, Xu L, Wang LF, Zhang L, Zhang ZY, Liu Y et al. A 60-s postconditioning protocol by percutaneous coronary intervention inhibits myocardial apoptosis in patients with acute myocardial infarction. *Apoptosis* 2009;**14**:1204–1211.
215. Lonborg J, Kelbaek H, Vejstrup N, Jorgensen E, Helqvist S, Saunamaki K et al. Cardioprotective effects of ischemic postconditioning in patients treated with primary percutaneous coronary intervention, evaluated by magnetic resonance. *Circ Cardiovasc Interv* 2010;**3**:34–41.
216. Nikolaidis LA, Mankad S, Sokos GG, Miske G, Shah A, Elahi D et al. Effects of glucagon-like peptide-1 in patients with acute myocardial infarction and left ventricular dysfunction after successful reperfusion. *Circulation* 2004;**109**:962–965.
217. Mehta SR, Yusuf S, Diaz R, Zhu J, Pais P, Xavier D et al. Effect of glucose-insulin-potassium infusion on mortality in patients with acute ST-segment elevation myocardial infarction: the CREATE-ECLA randomized controlled trial. *JAMA* 2005;**293**:437–446.
218. Bates E, Bode C, Costa M, Gibson CM, Granger C, Green C et al. Intracoronary KAI-9803 as an adjunct to primary percutaneous coronary intervention for acute ST-segment elevation myocardial infarction. *Circulation* 2008;**117**:886–896.
219. Ferrario M, Arbustini E, Massa M, Rosti V, Marziliano N, Raineri C et al. High-dose erythropoietin in patients with acute myocardial infarction: A pilot, randomised, placebo-controlled study. *Int J Cardiol* 2009; doi:10.1016/j.ijcard.2009.10.028. Published online ahead of print 10 November 2009.
220. Suh JW, Chung WY, Kim YS, Kim KI, Jeon EJ, Cho YS et al. The effect of intravenous administration of erythropoietin on the infarct size in primary percutaneous coronary intervention. *Int J Cardiol* 2010; doi:10.1016/j.ijcard.2010.02.002. Published online ahead of print 3 March 2010.
221. Kim J-S, Kim J, Choi D, Lee CJ, Lee SH, Ko Y-G et al. Efficacy of high-dose atorvastatin loading before primary percutaneous coronary intervention in ST-segment elevation myocardial infarction. *JACC Cardiovasc Interv* 2010;**3**:332–339.