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Posterior Capsular Opacification

A Problem Reduced but Not Yet Eradicated

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osterior capsular opacification (PCO) is the most frequent complication of cataract surgery. Advances in surgical techniques, intraocular lens materials, and designs have reduced the PCO rate, but it is still a significant problem. The only effective treatment for PCO, Nd:YAG laser capsulotomy carries vision-related complications and risks and puts a significant financial burden on the health care system. This review contains current knowledge about the mechanisms of PCO development. Posterior capsular opacification is caused mainly by remnant lens epithelial cell proliferation and migration, epithelial-mesenchymal transition, collagen deposition, and lens fiber generation. All of these processes are influenced by cytokines, growth factors, and extracellular matrix proteins. We also describe advances and improvements in surgical techniques, intraocular lens materials, and the designs and use of therapeutic agents leading to safe, effective, and less expensive strategies to eradicate PCO.

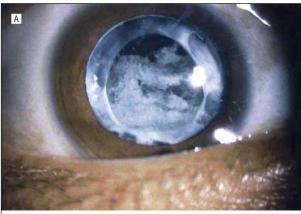
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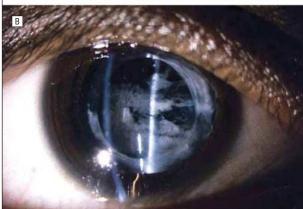
Cataract surgery is currently the most common and well-established ophthalmic surgical procedure in the world. This procedure involves the extracapsular extraction of the natural opaque lens fibers and implantation of an intraocular lens (IOL), which restores good vision. However, posterior capsular opacification (PCO), which is also termed secondary cataract, is a common long-term complication of modern cataract surgery (Figure 1).2-6 The first IOL implantation was performed by Sir Harold Ridley in 1950. Since that time, the technology has undergone a wide variety of improvements that reduced the incidence of PCO but did not eliminate it as a significant clinical problem. Decreased visual acuity induced by PCO is reported to occur in 20% to 40% of patients 2 to 5 years after surgery (Figure 1). Posterior capsular opacification development is age dependent, with a low incidence in older

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patients but high rates in young patients, especially children and infants.^{2,7-9}

At present, the only effective treatment of PCO is Nd:YAG laser capsulotomy, which involves clearing the visual axis by creating a central opening in the opacified posterior capsule. 2,10 Although this procedure is easy and quick, there are complications, including retinal detachment, damage to the IOL, cystoid macular edema, an increase in intraocular pressure, iris hemorrhage, corneal edema, IOL subluxation, and exacerbation of localized endophthalmitis.^{2,10,11} Changes induced by Nd:YAG capsulotomy have been shown to be affected by IOL material and design. Trinavarat et al¹² observed that silicone lenses were more easily damaged by laser capsulotomy than were acrylic or polymethyl methacrylate (PMMA) lenses. In addition, this treatment represents a considerable cost burden to national health care systems, and such laser treatment is not readily available in developing countries. Therefore, a better understanding of the pathogenic mechanism of PCO is highly desirable as a basis for improving the outcome of cataract surgery and eradicating PCO.





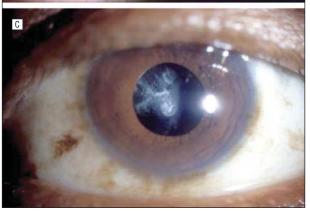


Figure 1. Representative slitlamp photographs of human eyes with posterior capsular opacification (PCO). A, Diffuse overall illumination and high magnification showing a large PCO blocking the visual axis. B, Optical sectioning and background illumination showing that the PCO is located on the central posterior lens capsule. C, Diffuse overall illumination and low magnification showing that the central visual axis is obscured.

Research laboratories worldwide attempting to eliminate the problem of PCO development are focusing on several strategies, including improving surgical techniques, IOL materials, IOL designs, use of therapeutic agents, and combination therapy (**Figure 2**). Recent improvements in surgical techniques and IOL materials and designs have served mainly to delay the onset of PCO rather than eliminate the problem. ^{13,14} The use of cytotoxic agents carries the risk of toxic effects on surrounding ocular tissues.

The aim of this review is to describe current knowledge about the mechanisms of PCO development and strategies to eliminate this sight-threatening problem.

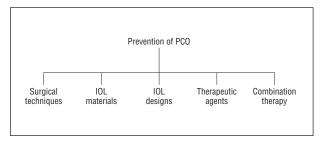


Figure 2. Schematic representation of current strategies toward finding a safe, effective, and less expensive way to eradicate posterior capsular opacification (PCO). IOL indicates intraocular lens.

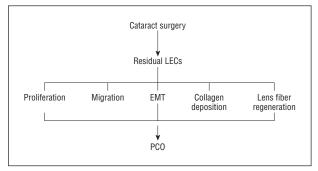


Figure 3. Schematic representation of the mechanism of posterior capsular opacification (PCO) development. Lens epithelial cells (LECs) left behind in the capsular bag after cataract surgery proliferate, migrate, convert from epithelial to mesenchymal cells (EMT), deposit collagen, and generate lens fibers, leading to PCO development.

MECHANISMS OF PCO DEVELOPMENT

Lens epithelial cells (LECs) left behind in the capsular bag after any type of extracapsular cataract surgery are mainly responsible for PCO development.² Proliferation, migration, epithelial-to-mesenchymal transition (EMT), collagen deposition, and lens fiber regeneration of LECs are the main causes of opacification (**Figure 3**). It appears that cataract surgery induces a woundhealing response in the lens, and leftover LECs proliferate and migrate across the posterior capsule and undergo lens fiber regeneration and EMT.^{7,15,16} Clinically, there are 2 morphological types of PCO, the fibrosis type and the pearl type. Fibrosis-type PCO is caused by the proliferation and migration of LECs, which undergo EMT, resulting in fibrous metaplasia and leading to significant visual loss by producing folds and wrinkles in the posterior capsule.⁷ Pearl-type PCO is caused by the LECs located at the equatorial lens region (lens bow) causing regeneration of crystallin-expressing lenticular fibers and forming Elschnig pearls and Soemmering ring, responsible for most cases of PCO-related visual loss. 2,17 The histological features of PCO are now well established, but to date the molecular mechanisms influencing leftover LEC behavior after cataract surgery are not completely clear. In vitro studies and animal models of PCO suggest that several cytokines and growth factors play a major role in the pathogenesis of PCO.^{18,19}

Studies show that levels of several cytokines and growth factors increase in aqueous humor and influence the behavior of the remaining LECs after cataract surgery. These factors include transforming growth factor β (TGF- β), fibroblast growth factor 2 (FGF-2), hepatocyte growth fac-

tor, interleukins 1 and 6 (IL-1 and IL-6), and epithelial growth factor. ^{18,20} Wormstone et al²¹ and Duncan et al²² have studied LEC growth on human capsular bags in a proteinfree medium, which has allowed the autocrine control by individual growth factors to be analyzed.

Transforming growth factor β plays a central role in the cell biology of PCO. It is a multifunctional growth factor with a wide range of opposing effects on cellular processes. ²³ Transforming growth factor β induces EMT²⁴⁻²⁷ and has been shown to suppress LEC proliferation. 26,28 A recent study showed that during PCO formation the TGFβ-activated pathway is influenced by SPARC (secreted protein, acidic, and rich in cysteine), which regulates matrix-cell interactions.²⁹ Members of the FGF family are known to play a critical role in establishing and maintaining normal lens structure and function. Proliferation, migration, and fiber differentiation of normal LECs have been shown to be affected by FGF-2 in vitro. Previous studies have shown that FGF-2 induces proliferation, 30,31 which may contribute to the development of PCO. Also, FGF-2 has been shown to counteract the TGF-β effect.³² Hepatocyte growth factor is highly expressed in human capsular bag cultures in protein-free medium. Studies have shown that hepatocyte growth factor plays an important role in PCO development by inducing proliferation of LECs.33,34 Interleukin 1 is synthesized by LECs in vitro35 and stimulates LEC mitosis and collagen synthesis.36 Interleukin 1 stimulates human LECs to produce prostaglandin-E2, which contributes to increased inflammation after cataract surgery. In human vein endothelial cells, IL-1 induced IL-6 secretion,³⁷ suggesting that the action of IL-1 on LECs may be mediated by IL-6. Jiang et al³⁸ have shown that LEC migration is induced by epithelial growth factor, suggesting its role in PCO development.

Migration of human LECs plays an important role in the remodeling of the lens capsule^{39,40} and is associated with matrix metalloproteinase activity in the lens.⁴¹ Matrix metalloproteinases are a group of proteolytic enzymes, essential for cell migration and cell-mediated contraction after wound healing.⁴² Changes in lens capsule structure during PCO development may include remodeling of the extracellular matrix by matrix metalloproteinases.

PREVENTION OF PCO

There are several urgent reasons to eradicate PCO. First, PCO remains the most common complication of cataract surgery. 43,44 Posterior capsular opacification is even more threatening in young adults and children, with a higher incidence, quicker onset, and greater amblyogenic effect. The rates of PCO in children ranged from 43.7% to 100%, probably because of the high proliferation of LECs. Recent advances in pediatric cataract surgery, such as posterior continuous curvilinear capsulorrhexis and anterior vitrectomy, have decreased PCO incidence.^{2,44-52} Posterior capsular opacification in young children is always dense and may need to be removed with additional surgery, which carries more risks of potential complications. 45,46 In addition, Nd:YAG laser capsulotomy, the only currently available treatment of PCO, has several significant complications, and this procedure imposes a severe financial burden on the health care system. Therefore, investigators are constantly working to advance and improve the following areas to find a safe and effective way to eradicate PCO.

Surgical Techniques

Because PCO is predominantly caused by residual LECs in the capsular bag after cataract surgery, 2,19,53 several surgical techniques have been attempted for the removal of these LECs at the time of lens extraction. These techniques include aspiration of the anterior capsule using an extensive irrigation/aspiration system during cataract surgery, 54-61 pharmacological dispersion and aspiration of the anterior capsule, 62-65 and manual polishing of the anterior and/or posterior capsule. 58,66-68 These studies provided limited and conflicting information about the effect of residual LECs on long-term PCO development. One study showed that ultrasonic vacuuming during cataract surgery reduced the number of patients requiring laser capsulotomy.54 Another study showed that capsule vacuuming reduced but did not eliminate PCO.⁶⁹ Khalifa⁶⁶ determined that vacuuming the posterior capsule had no effect on the long-term development of PCO. Therefore, vacuuming or polishing the capsule may delay the onset of PCO, but the long-term benefit is limited because PCO is mainly caused by germinative LECs in the equatorial region rather than the displaced metaplastic LECs already on the posterior capsule. 70 Also, equatorial capsule vacuuming has been found to be associated with additional surgery time and trauma and risk of capsule tears, damaging the capsular support of the IOL implant. 71,72 Davidson et al 73 suggested that near 100% removal of residual LECs at the time of cataract surgery may be necessary to prevent LEC proliferation on the posterior capsule and development of PCO. Hydrodissectionenhanced cortical cleanup after cataract surgery to remove retained/regenerative cortical material and cells has been shown to be important for PCO prevention. 14 Fine⁷⁴ modified this technique by cleaning the residual cortex in the presence of a posterior chamber IOL, which protected the posterior capsule from disruption. These studies demonstrated that hydrodissection is an effective, practical, and inexpensive method for cortex removal, but alone it does not completely eliminate LECs. The rare complications of this technique are posterior capsule rupture ⁷⁵ and nuclear dislocation into the vitreous. ⁷⁶ Peng et al¹⁴ demonstrated the importance of the barrier effect of the IOL optic in preventing LEC growth and suggested that it can be a second line of defense when cortical cleanup is incomplete.

A sealed capsule irrigation device was introduced by Maloof et al⁷⁷ and other investigators. ⁷⁸⁻⁸¹ This device consists of a foldable suction ring with 2 separate lines, one for vacuum application and the other for irrigation. The device allows the temporary seal of the capsulorrhexis after cataract removal and selective irrigation of the capsular bag with a pharmacological agent without damaging surrounding tissues. This device is minimally invasive and easy to use, fits through a small incision, is relatively inexpensive, and does not add significant time to cataract surgery and therefore shows great promise in

combating PCO. Hara et al⁸² reported the advantages of a closed endocapsular ring to prevent PCO. This approach was shown to be promising in PCO prevention in the eyes of adults and children.^{83,84} Recently, primary capsulorrhexis with anterior vitrectomy has been suggested to be a necessary and effective procedure to lower PCO rates in pediatric cataract surgery.⁴⁹

IOL Materials and Designs

Since IOLs were first implanted, a wide range of improvements has been introduced specifically to prevent PCO. Among these, several advancements have been made in IOL materials and designs to improve biocompatibility assessed in terms of the eye's foreign body reaction against the IOL (uveal biocompatibility) and interaction of the IOL with residual LECs within the capsular bag, which influences LEC proliferation, migration, and EMT, resulting in anterior capsular opacification and PCO (capsular biocompatibility). 85

There are 2 main types of materials used for IOL manufacturing: acrylic and silicone. Acrylic lenses are rigid (made from PMMA) or foldable (made from hydrophobic material such as AcrySof [Alcon Inc, Fort Worth, Texas] and Sensar [Staar Surgical Company, Monrovia, California] or hydrophilic material [also known as hydrogels] such as Hydroview [Bausch and Lomb Surgical, Rochester, New York] and Centriflex [Edge Biosystems, Gaithersburg, Maryland]). Silicone lenses were initially made from polydimethylsiloxane. ^{86,87}

Several clinical and experimental studies have been performed to demonstrate the role of the IOL materials and designs to reduce the incidence of PCO. Comparison of hydrophobic and hydrophilic materials showed that the type of material might influence PCO development.88-90 Although it is well recognized that a hydrophilic acrylic material is more biocompatible, 91 IOLs made of this material have been shown to support LEC adhesion, migration, and proliferation and thus PCO development⁹²⁻⁹⁴ compared with an IOL made of PMMA or hydrophobic acrylic materials. 95,96 Modification of the IOL surface, which can inhibit cell and protein adhesion, has been suggested as one of the most tolerable methods for preventing PCO because it does not require any manipulation within the eye or the use of any active or harmful agents during IOL implantation. Surface modifications of PMMA IOLs by carbon and titanium, 97 heparin, 98 and polytetrafluoroethylene (Teflon; Dupont de Nemours, Wilmington, Delaware)99 and of silicon IOLs by oxygen and carbon dioxide plasma100 or a sulfonate and carboxylate group containing polymer¹⁰¹ have been reported to have higher biocompatibility and effectiveness in prevention of PCO. Recently, IOL surface modification by gas plasma¹⁰² and polyethylene glycol¹⁰³ has been shown to influence LEC behavior and to prevent PCO.

Many advances have been made in IOL designs that have reduced PCO incidence. A higher PCO inhibitory effect has been observed with IOLs that provide a mechanical barrier effect on the posterior lens capsule. ^{2,4,19,104-106} Nishi et al ¹⁰⁶⁻¹¹⁰ demonstrated that the sharpedge optic IOL and the formation of a capsular bend are highly effective in reducing PCO. Adhesion of the IOL

material with the lens capsule also plays a role in PCO prevention by creating a sharp capsular bend, ^{89,109-111} which inhibits LEC migration onto the posterior capsule. Nishi and other investigators^{106,107,112,113} also demonstrated that contact inhibition of migrating LECs is induced at the capsular bend, which leads to PCO prevention. Recently, Zemaitiene et al¹¹⁴ showed that there is no difference in PCO development between 3-piece and 1-piece acrylic hydrophobic IOLs.

Posterior capsular opacification is a multifactorial process. The 3 main factors for PCO development are patient related (eg, age and ocular disease), surgery related (eg, irrigation/aspiration of the capsule, hydrodissectionenhanced cortical cleanup, sealed capsule irrigation, capsulorrhexis size, and in-the-bag IOL fixation), and IOL related. It is well accepted that PCO incidence is greatly influenced by IOL material and design. A recent metanalysis of 23 randomized controlled trials concluded that the sharp-edge optic IOLs made of acrylic and silicone are superior in lowering the rates of PCO and laser capsulotomy. In the sharp-edge optic IOLs made of acrylic and silicone are superior in lowering the rates of PCO and laser capsulotomy.

Therapeutic Agents

Improvements in surgical techniques and IOL materials and designs have lowered PCO rates but have not eradicated the problem. Therefore, despite several complications and the cost burden, Nd:YAG laser capsulotomy is still the most frequently used treatment of PCO. Consequently, the development of an alternative medical treatment of PCO is of critical importance. Alternatives include selectively destroying residual LECs while avoiding toxic effects on other intraocular tissues. Intraocular application of pharmacological agents to prevent PCO has been investigated by several laboratories, and the commonly used methods for this application are direct injection into the anterior chamber, addition to the irrigating solution, or impregnation of the IOL. Pharmacological agents such as thermosetting plastic prepared with phenol formaldehyde resins (Catalin), methotrexate, mitomycin, daunomycin, and fluorouracil have been shown to be effective in preventing PCO in vitro, 19,117,118 but in vivo studies have shown their toxicity to corneal endothelial cells, iris, ciliary body epithelial cells, and retina.¹⁹ Studies tested cytotoxic and therapeutic agents, including diclofenac sodium, ¹¹⁹ saporin, ¹²⁰ thapsigargin, ²² salmosin, ¹²¹ minoxidil, ¹²² a matrix metalloproteinase inhibitor (Ilomostat), 41 and cyclo-oxygenase 2 inhibitors. 123 These studies showed promise for finding medical treatment of PCO by targeting the survival, adhesion, proliferation, migration, and transdifferentiation of residual LECs, but the risk of their toxic effects on surrounding intraocular tissues has restricted their clinical use. Malecaze et al^{124,125} provided a gene therapy approach to target LECs in the capsular bag by inducing therapeutic apoptosis by overexpression of proapoptotic genes. Walker et al¹²⁶ showed the blocking effect of an Src family kinase inhibitor on PCO development in a chick model of the lens capsular bag.

Posterior capsular opacification is a multifactorial disease and is influenced by the increased levels of several cytokines and growth factors, including TGF-β, FGF-2,

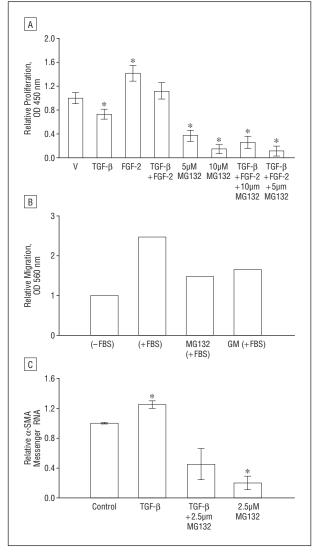


Figure 4. The proteasome inhibitor MG132 inhibits lens epithelial cell (LEC) proliferation, migration, and epithelial-to-mesenchymal transition (EMT) marker production in primary human LECs. A, Cell proliferation assay was performed by treating cells with transforming growth factor β (TGF- β) (1 ng/mL), fibroblast growth factor 2 (FGF-2) (20 ng/mL), or MG132 (5µM or 10μM), alone or in combination. After 12 hours of incubation, proliferation was evaluated with a colorimetric WST-1 assay. B, Cell migration assay was performed using a transwell chamber assay. Serum-starved cells were treated with MG132 (10µM) or matrix metalloproteinase inhibitor GM600 (GM, 10µM) and added in upper polycarbonate membrane inserts in 24-well plates. Lower wells contained Dulbecco Modified Eagle Medium with or without 20% fetal bovine serum (FBS), indicated in parentheses, as chemoattractant. After 6 hours of incubation, migratory cells were stained and quantitated by absorption at 560 nm. C, Expression of the EMT marker α -smooth muscle actin (α -SMA) was observed by means of reverse-transcriptase polymerase chain reaction analysis after 16 hours of treatment with TGF-β (1 ng/mL) and MG132 (2.5μM), alone or in combination. OD indicates optical density; V, vehicle control. Reprinted from Hosler et al¹²⁷ and Awasthi et al^{128,129} with permission from the Association for Research in Vision and Ophthalmology.

hepatocyte growth factor, IL-6, and epithelial growth factor. These cytokines and growth factors differentially affect residual LEC behavior that leads to PCO development. We recently demonstrated that a reversible peptide aldehyde inhibitor of the proteasome MG132 can simultaneously block EMT markers¹²⁷ and proliferation¹²⁸ and migration¹²⁹ of LECs, suggesting that proteasome inhibition might be a good therapeutic strategy for PCO pre-

vention (**Figure 4**¹²⁷⁻¹²⁹). Maloof et al⁷⁷ developed the PerfectCapsule device (Milvella Ltd, Sydney, Australia), which permits cytotoxic agents to be delivered selectively to the capsular bag, thus selectively targeting residual LECs. This device also allows cytotoxic agents to be removed after treatment during the cataract surgery procedure. Duncan et al⁵ demonstrated that the applications of the PerfectCapsule device in conjunction with the cytotoxic drug thapsigargin is effective in the prevention of PCO. A recent study by Kim et al¹³⁰ showed that in rabbit eyes mitomycin is more effective than distilled water for reducing PCO and that the sealed capsule irrigation device protected the surrounding tissue from mitomycin toxicity.

CONCLUSIONS

In recent years, our understanding of mechanisms of PCO development has increased significantly; therefore, several advances have been made to improve cataract surgery techniques, IOL materials and designs, and the use of therapeutic agents. Because of these improvements, PCO occurrence has decreased, or at least PCO onset has been delayed. Nevertheless, PCO remains the most common complication of cataract surgery, especially in young adults and children. Therefore, research aimed at improving surgical techniques to eliminate almost all LECs from the capsular bag at the time of surgery, optimizing IOL biocompatibility, minimizing postoperative inflammation reaction, and targeting residual LECs by therapeutic agents that have minimal or no effect on other ocular tissues is highly desirable.

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REFERENCES

- Beebe DC. The lens. In: Kaufman PL, Alm A, eds. Adler's Physiology of the Eye: Clinical Application. 10th ed. St Louis, MO: Mosby–Year Book; 2003:117-158.
- Apple DJ, Solomon KD, Tetz MR, et al. Posterior capsule opacification. Surv Ophthalmol. 1992;37(2):73-116.
- Pandey SK, Apple DJ, Werner L, Maloof AJ, Milverton EJ. Posterior capsule opacification: a review of the aetiopathogenesis, experimental and clinical studies and factors for prevention. *Indian J Ophthalmol*. 2004;52(2):99-112.
- Spalton DJ. Posterior capsular opacification after cataract surgery. Eye. 1999; 13(pt 3b):489-492.
- Duncan G, Wang L, Neilson GJ, Wormstone IM. Lens cell survival after exposure to stress in the closed capsular bag. *Invest Ophthalmol Vis Sci.* 2007; 48(6):2701-2707.
- 6. Clark DS. Posterior capsule opacification. Curr Opin Ophthalmol. 2000;11(1):56-64.
- McDonnell PJ, Zarbin MA, Green WR. Posterior capsule opacification in pseudophakic eyes. Ophthalmology. 1983;90(12):1548-1553.
- Binkhorst CD, Gobin MH. Injuries to the eye with lens opacity in young children. Ophthalmologica. 1964;148:169-183.

- Hiles DA, Wallar PH. Phacoemulsification versus aspiration in infantile cataract surgery. Ophthalmic Surg. 1974;5(2):13-16.
- Apple DJ, Peng Q, Visessook N, et al. Eradication of posterior capsule opacification: documentation of a marked decrease in Nd:YAG laser posterior capsulotomy rates noted in an analysis of 5416 pseudophakic human eyes obtained postmortem. Ophthalmology. 2001;108(3):505-518.
- Aslam TM, Devlin H, Dhillon B. Use of Nd:YAG laser capsulotomy. Surv Ophthalmol. 2003;48(6):594-612.
- Trinavarat A, Atchaneeyasakul L, Udompunturak S. Neodymium: YAG laser damage threshold of foldable intraocular lenses. J Cataract Refract Surg. 2001; 27(5):775-780.
- Nagamoto T, Eguchi G. Effect of intraocular lens design on migration of lens epithelial cells onto the posterior capsule. J Cataract Refract Surg. 1997; 23(6):866-872.
- Peng Q, Visessook N, Apple DJ, et al. Surgical prevention of posterior capsule opacification, part 3: intraocular lens optic barrier effect as a second line of defense. J Cataract Refract Surg. 2000;26(2):198-213.
- Cobo LM, Ohsawa E, Chandler D, Arguello R, George G. Pathogenesis of capsular opacification after extracapsular cataract extraction: an animal model. Ophthalmology. 1984;91(7):857-863.
- Wormstone IM. Posterior capsule opacification: a cell biological perspective. Exp Eye Res. 2002;74(3):337-347.
- Saika S. Relationship between posterior capsule opacification and intraocular lens biocompatibility. *Prog Retin Eye Res.* 2004;23(3):283-305.
- Meacock WR, Spalton DJ, Stanford MR. Role of cytokines in the pathogenesis of posterior capsule opacification. Br J Ophthalmol. 2000;84(3):332-336.
- Nishi O. Posterior capsule opacification, part 1: experimental investigations. J Cataract Refract Surg. 1999;25(1):106-117.
- Wallentin N, Wickström K, Lundberg C. Effect of cataract surgery on aqueous TGF-β and lens epithelial cell proliferation. *Invest Ophthalmol Vis Sci.* 1998; 39(8):1410-1418.
- Wormstone IM, Liu CS, Rakic JM, Marcantonio JM, Vrensen GF, Duncan G. Human lens epithelial cell proliferation in a protein-free medium. *Invest Oph-thalmol Vis Sci.* 1997:38(2):396-404.
- Duncan G, Wormstone IM, Liu CS, Marcantonio JM, Davies PD. Thapsigargincoated intraocular lenses inhibit human lens cell growth. *Nat Med.* 1997; 3(9):1026-1028.
- Massagué J. The transforming growth factor-beta family. Annu Rev Cell Biol. 1990;6:597-641.
- Liu J, Hales AM, Chamberlain CG, McAvoy JW. Induction of cataract-like changes in rat lens epithelial explants by transforming growth factor β. Invest Ophthalmol Vis Sci. 1994;35(2):388-401.
- Hales AM, Chamberlain CG, McAvoy JW. Cataract induction in lenses cultured with transforming growth factor-β. *Invest Ophthalmol Vis Sci.* 1995;36(8): 1709-1713
- Saika S, Okada Y, Miyamoto T, Ohnishi Y, Ooshima A, McAvoy JW. Smad translocation and growth suppression in lens epithelial cells by endogenous TGFβ2 during wound repair. Exp Eye Res. 2001;72(6):679-686.
- Saika S, Miyamoto T, Ishida I, et al. TGFβ-Smad signalling in postoperative human lens epithelial cells. Br J Ophthalmol. 2002;86(12):1428-1433.
- Kurosaka D, Nagamoto T. Inhibitory effect of TGF-β2 in human aqueous humor on bovine lens epithelial cell proliferation. *Invest Ophthalmol Vis Sci.* 1994; 35(9):3408-3412.
- Gotoh N, Perdue NR, Matsushima H, Sage EH, Yan Q, Clark JI. An in vitro model
 of posterior capsular opacity: SPARC and TGF-β2 minimize epithelial-tomesenchymal transition in lens epithelium. *Invest Ophthalmol Vis Sci.* 2007;
 48(10):4679-4686.
- McAvoy JW, Chamberlain CG. Fibroblast growth factor (FGF) induces different responses in lens epithelial cells depending on its concentration. *Development*. 1989;107(2):221-228.
- Tanaka T, Saika S, Ohnishi Y, et al. Fibroblast growth factor 2: roles of regulation of lens cell proliferation and epithelial-mesenchymal transition in response to injury. Mol Vis. 2004;10:462-467.
- Mansfield KJ, Cerra A, Chamberlain CG. FGF-2 counteracts loss of TGFβ affected cells from rat lens explants: implications for PCO (after cataract). Mol Vis. 2004;10:521-532.
- Wormstone IM, Tamiya S, Marcantonio JM, Reddan JR. Hepatocyte growth factor function and c-Met expression in human lens epithelial cells. *Invest Ophthalmol Vis Sci.* 2000;41(13):4216-4222.
- Choi J, Park SY, Joo CK. Hepatocyte growth factor induces proliferation of lens epithelial cells through activation of ERK1/2 and JNK/SAPK. *Invest Ophthalmol Vis Sci.* 2004;45(8):2696-2704.
- Nishi O, Nishi K, Imanishi M. Synthesis of interleukin-1 and prostaglandin E₂ by lens epithelial cells of human cataracts. Br J Ophthalmol. 1992;76(6): 338-341

- Nishi O, Nishi K, Fujiwara T, Shirasawa E, Ohmoto Y. Effects of the cytokines on the proliferation of and collagen synthesis by human cataract lens epithelial cells. Br J Ophthalmol. 1996;80(1):63-68.
- May LT, Torcia G, Cozzolino F, et al. Interleukin-6 gene expression in human endothelial cells: RNA start sites, multiple IL-6 proteins and inhibition of proliferation. *Biochem Biophys Res Commun.* 1989;159(3):991-998.
- Jiang Q, Zhou C, Bi Z, Wan Y. EGF-induced cell migration is mediated by ERK and PI3K/AKT pathways in cultured human lens epithelial cells. *J Ocul Phar-macol Ther*. 2006;22(2):93-102.
- Beck R, Nebe B, Guthoff R, Rychly J. Inhibition of lens epithelial cell adhesion by the calcium antagonist Mibefradil correlates with impaired integrin distribution and organization of the cytoskeleton. *Graefes Arch Clin Exp Ophthalmol*. 2001;239(6):452-458.
- Sugita M, Kato S, Sugita G, Oshika T. Migration of lens epithelial cells through haptic root of single-piece acrylic-foldable intraocular lens. Am J Ophthalmol. 2004;137(2):377-379.
- Wong TT, Daniels JT, Crowston JG, Khaw PT. MMP inhibition prevents human lens epithelial cell migration and contraction of the lens capsule. *Br J Ophthalmol*. 2004;88(7):868-872.
- 42. Lemaître V, D'Armiento J. Matrix metalloproteinases in development and disease. Birth Defects Res C Embryo Today. 2006;78(1):1-10.
- Dewey S. Posterior capsule opacification. Curr Opin Ophthalmol. 2006;17(1):45-53
- Wilson ME Jr, Trivedi RH. The ongoing battle against posterior capsular opacification. Arch Ophthalmol. 2007;125(4):555-556.
- BenEzra D, Cohen E, Rosa L. Traumatic cataract in children: correction of aphakia by contact lens or intraocular lens. Am J Ophthalmol. 1997;123(6):773-782.
- Atkinson CS, Hiles DA. Treatment of secondary posterior capsular membranes with the Nd:YAG laser in a pediatric population. Am J Ophthalmol. 1994; 118(4):496-501.
- Gimbel HV, Neuhann T. Development, advantages, and methods of the continuous circular capsulorhexis technique. J Cataract Refract Surg. 1990; 16(1):31-37.
- Gimbel HV. Posterior continuous curvilinear capsulorhexis and optic capture of the intraocular lens to prevent secondary opacification in pediatric cataract surgery. J Cataract Refract Surg. 1997;23(suppl 1):652-656.
- Luo Y, Lu Y, Lu G, Wang M. Primary posterior capsulorhexis with anterior vitrectomy in preventing posterior capsule opacification in pediatric cataract microsurgery. *Microsurgery*. 2008;28(2):113-116.
- Ram J, Brar GS, Kaushik S, Gupta A, Gupta A. Role of posterior capsulotomy with vitrectomy and intraocular lens design and material in reducing posterior capsule opacification after pediatric cataract surgery. *J Cataract Refract Surg.* 2003;29(8):1579-1584.
- Mackool RJ, Chhatiawala H. Pediatric cataract surgery and intraocular lens implantation: a new technique for preventing or excising postoperative secondary membranes. J Cataract Refract Sura. 1991;17(1):62-66.
- Guo S, Wagner RS, Caputo A. Management of the anterior and posterior lens capsules and vitreous in pediatric cataract surgery. *J Pediatr Ophthalmol Strabismus*. 2004;41(6):330-337, 356-357.
- Frezzotti R, Caporossi A, Mastrangelo D, et al. Pathogenesis of posterior capsular opacification, II: histopathological and in vitro culture findings. *J Cata*ract Refract Surg. 1990;16(3):353-360.
- Nishi O, Nishi K. Intercapsular cataract surgery with lens epithelial cell removal, III: long-term follow-up of posterior capsular opacification. *J Cataract Refract Surg.* 1991;17(2):218-220.
- Hara T, Hara T. Observations on lens epithelial cells and their removal in anterior capsule specimens. Arch Ophthalmol. 1988;106(12):1683-1687.
- Nishi O. Intercapsular cataract surgery with lens epithelial cell removal, II: effect on prevention of fibrinous reaction. J Cataract Refract Surg. 1989;15(3): 301-303
- Nishi O. Intercapsular cataract surgery with lens epithelial cell removal, I: without capsulorhexis. J Cataract Refract Surg. 1989;15(3):297-300.
- Mathey CF, Kohnen TB, Ensikat HJ, Koch HR. Polishing methods for the lens capsule: histology and scanning electron microscopy. *J Cataract Refract Surg*. 1994:20(1):64-69.
- Nishi O. Lens epithelial cell removal by ultrasound: access to 12 o'clock. J Cataract Refract Surg. 1989;15(6):704-706.
- Nishi O. Removal of lens epithelial cells by ultrasound in endocapsular cataract surgery. Ophthalmic Surg. 1987;18(8):577-580.
- 61. Green WT, Boase DL. How clean is your capsule? Eye. 1989;3(pt 6):678-684.
- Humphry RC, Davies EG, Jacob TJ, Thompson GM. The human anterior lens capsule: an attempted chemical debridement of epithelial cells by ethylenediaminetetracetic acid (EDTA) and trypsin. Br J Ophthalmol. 1988;72(6):406-408.

- Nishi O, Nishi K, Hikida M. Removal of lens epithelial cells by dispersion with enzymatic treatment followed by aspiration. *Ophthalmic Surg.* 1991;22(8): 444-450
- Nishi O, Nishi K, Hikita M. A new approach to lens epithelial cell removal: dispersion aspiration. Dev Ophthalmol. 1991;22:101-105.
- Nishi O, Nishi K, Hikida M. Removal of lens epithelial cells following loosening of the junctional complex. J Cataract Refract Surg. 1993;19(1):56-61.
- Khalifa MA. Polishing the posterior capsule after extracapsular extraction of senile cataract. J Cataract Refract Surg. 1992;18(2):170-173.
- Dahan E, Allarakhia L. Irrigation, aspiration, and polishing cannula. J Cataract Refract Surg. 1991;17(1):97-98.
- Turtz AI. Sandblasted end-cutting tip for posterior capsular vacuuming and polishing. J Am Intraocul Implant Soc. 1985;11(1):73.
- Nishi O, Nakai Y, Mizumoto Y, Yamada Y. Capsule opacification after refilling the capsule with an inflatable endocapsular balloon. *J Cataract Refract Surg.* 1997:23(10):1548-1555.
- Liu CS, Wormstone IM, Duncan G, Marcantonio JM, Webb SF, Davies PD.
 A study of human lens cell growth in vitro: a model for posterior capsule opacification. *Invest Ophthalmol Vis Sci.* 1996;37(5):906-914.
- O'Donnell FE Jr, Santos B. Posterior capsular-zonular disruption in planned extracapsular surgery. Arch Ophthalmol. 1985;103(5):652-653.
- Osher RH, Cionni RJ. The torn posterior capsule: its intraoperative behavior, surgical management, and long-term consequences. *J Cataract Refract Surg*. 1990;16(4):490-494.
- Davidson MG, Morgan DK, McGahan MC. Effect of surgical technique on in vitro posterior capsule opacification. J Cataract Refract Surg. 2000;26(10):1550-1554
- Fine IH. Cortical cleaving hydrodissection. J Cataract Refract Surg. 1992;18(5): 508-512.
- Yeoh R. The "pupil snap" sign of posterior capsule rupture with hydrodissection in phacoemulsification. Br J Ophthalmol. 1996;80(5):486.
- Ota I, Miyake S, Miyake K. Dislocation of the lens nucleus into the vitreous cavity after standard hydrodissection. Am J Ophthalmol. 1996;121(6):706-708
- Maloof A, Neilson G, Milverton EJ, Pandey SK. Selective and specific targeting
 of lens epithelial cells during cataract surgery using sealed-capsule irrigation.

 J Cataract Refract Surg. 2003;29(8):1566-1568.
- Agarwal A, Agarwal S, Agarwal A, Maloof A. Sealed-capsule irrigation device. J Cataract Refract Surg. 2003;29(12):2274-2276.
- Maloof AJ, Pandey SK, Neilson G, Milverton EJ. Selective death of lens epithelial cells using demineralized water and Triton X-100 with PerfectCapsule sealed capsule irrigation: a histological study in rabbit eyes. *Arch Ophthalmol*. 2005; 123(10):1378-1384.
- Rabsilber TM, Limberger IJ, Reuland AJ, Holzer MP, Auffarth GU. Secondary cataract prevention. *Ophthalmology*. 2007;114(2):397-398.
- Rabsilber TM, Limberger IJ, Reuland AJ, Holzer MP, Auffarth GU. Long-term results of sealed capsule irrigation using distilled water to prevent posterior capsule opacification: a prospective clinical randomised trial. *Br J Ophthalmol*. 2007; 01(7):012-015.
- Hara T, Hara T, Hara T. Preventing posterior capsular opacification with an endocapsular equator ring in a young human eye: 2-year follow-up. Arch Ophthalmol. 2007;125(4):483-486.
- Dick HB. Closed foldable capsular rings. J Cataract Refract Surg. 2005;31 (3):467-471
- Dick HB, Schwenn O, Pfeiffer N. Implantation of the modified endocapsular bending ring in pediatric cataract surgery using a viscoadaptive viscoelastic agent. J Cataract Refract Surg. 1999;25(11):1432-1436.
- Werner L. Biocompatibility of intraocular lens materials. Curr Opin Ophthalmol. 2008;19(1):41-49.
- Christ FR, Fencil DA, Van Gent S, Knight PM. Evaluation of the chemical, optical, and mechanical properties of elastomeric intraocular lens materials and their clinical significance. *J Cataract Refract Surg.* 1989;15(2):176-184.
- Christ FR, Buchen SY, Deacon J, et al. Biomaterials used for intraocular lenses.
 In: Wise DL, Trantolo DJ, Altobelli DE, eds, et al. Encyclopedic Handbook of Biomaterials and Bioengineering, B: Applications. Vol 2. New York, NY: Marcel Dekker; 1995:1261-1313.
- Schmidbauer JM, Vargas LG, Apple DJ, et al. Evaluation of neodymium: yttriumaluminum-garnet capsulotomies in eyes implanted with AcrySof intraocular lenses. *Ophthalmology*. 2002;109(8):1421-1426.
- Linnola RJ, Werner L, Pandey SK, Escobar-Gomez M, Znoiko SL, Apple DJ. Adhesion of fibronectin, vitronectin, laminin, and collagen type IV to intraocular lens materials in pseudophakic human autopsy eyes, part 1: histological sections. *J Cataract Refract Surg.* 2000;26(12):1792-1806.
- Linnola RJ, Werner L, Pandey SK, Escobar-Gomez M, Znoiko SL, Apple DJ. Adhesion of fibronectin, vitronectin, laminin, and collagen type IV to intraocu-

- lar lens materials in pseudophakic human autopsy eyes, part 2: explanted intraocular lenses. *J Cataract Refract Surg.* 2000;26(12):1807-1818.
- Hollick EJ, Spalton DJ, Ursell PG. Surface cytologic features on intraocular lenses: can increased biocompatibility have disadvantages? *Arch Ophthalmol.* 1999; 117(7):872-878.
- Müllner-Eidenböck A, Amon M, Schauersberger J, Abela C, Petternel V, Zidek T. Cellular reaction on the anterior surface of 4 types of intraocular lenses. J Cataract Refract Surg. 2001;27(5):734-740.
- Schauersberger J, Amon M, Kruger A, Abela C, Schild G, Kolodjaschna J. Lens epithelial cell outgrowth on 3 types of intraocular lenses. *J Cataract Refract Surg.* 2001:27(6):850-854.
- Tognetto D, Toto L, Ballone E, Ravalico G. Biocompatibility of hydrophilic intraocular lenses. J Cataract Refract Surg. 2002;28(4):644-651.
- Hollick EJ, Spalton DJ, Ursell PG, Meacock WR, Barman SA, Boyce JF. Posterior capsular opacification with hydrogel, polymethylmethacrylate, and silicone intraocular lenses: two-year results of a randomized prospective trial. Am J Ophthalmol. 2000;129(5):577-584.
- Hayashi K, Hayashi H, Nakao F, Hayashi F. Anterior capsule contraction and intraocular lens decentration and tilt after hydrogel lens implantation. Br J Ophthalmol. 2001;85(11):1294-1297.
- Yuan Z, Sun H, Yuan J. A 1-year study on carbon, titanium surface-modified intraocular lens in rabbit eyes. *Graefes Arch Clin Exp Ophthalmol*. 2004; 242(12):1008-1013.
- Larsson R, Selén G, Björdklund H, Fagerholm P. Intraocular PMMA lenses modified with surface-immobilized heparin: evaluation of biocompatibility in vitro and in vivo. *Biomaterials*. 1989;10(8):511-516.
- Werner L, Legeais JM, Nagel MD, Renard G. Evaluation of teflon-coated intraocular lenses in an organ culture method. *J Biomed Mater Res.* 1999;46 (3):347-354.
- Hettlich HJ, Otterbach F, Mittermayer C, Kaufmann R, Klee D. Plasma-induced surface modifications on silicone intraocular lenses: chemical analysis and in vitro characterization. *Biomaterials*. 1991;12(5):521-524.
- Yammine P, Pavon-Djavid G, Helary G, Migonney V. Surface modification of silicone intraocular implants to inhibit cell proliferation. *Biomacromolecules*. 2005:6(5):2630-2637.
- Yuen C, Williams R, Batterbury M, Grierson I. Modification of the surface properties of a lens material to influence posterior capsular opacification. *Clin Experiment Ophthalmol.* 2006;34(6):568-574.
- Bozukova D, Pagnoulle C, De Pauw-Gillet MC, et al. Improved performances of intraocular lenses by poly(ethylene glycol) chemical coatings. *Biomacromolecules*. 2007;8(8):2379-2387.
- Hollick EJ, Spalton DJ, Meacock WR. The effect of capsulorhexis size on posterior capsular opacification: one-year results of a randomized prospective trial. *Am J Ophthalmol.* 1999;128(3):271-279.
- Hollick EJ, Spalton DJ, Ursell PG, et al. The effect of polymethylmethacrylate, silicone, and polyacrylic intraocular lenses on posterior capsular opacification 3 years after cataract surgery. *Ophthalmology*. 1999;106(1):49-54.
- 106. Nishi O, Nishi K, Sakanishi K. Inhibition of migrating lens epithelial cells at the capsular bend created by the rectangular optic edge of a posterior chamber intraocular lens. *Ophthalmic Surg Lasers*. 1998;29(7):587-594.
- Nishi O, Nishi K. Preventing posterior capsule opacification by creating a discontinuous sharp bend in the capsule. J Cataract Refract Surg. 1999;25 (4):521-526.
- Nishi O, Nishi K, Wickström K. Preventing lens epithelial cell migration using intraocular lenses with sharp rectangular edges. *J Cataract Refract Surg.* 2000; 26(10):1543-1549.
- Nishi O, Nishi K, Akura J, Nagata T. Effect of round-edged acrylic intraocular lenses on preventing posterior capsule opacification. J Cataract Refract Surg. 2001;27(4):608-613.
- Nishi O, Nishi K, Akura J. Speed of capsular bend formation at the optic edge of acrylic, silicone, and poly(methyl methacrylate) lenses. *J Cataract Refract* Surg. 2002;28(3):431-437.
- Oshika T, Nagata T, Ishii Y. Adhesion of lens capsule to intraocular lenses of polymethylmethacrylate, silicone, and acrylic foldable materials: an experimental study. Br J Ophthalmol. 1998;82(5):549-553.
- 112. Nishi O, Nishi K, Mano C, Ichihara M, Honda T. The inhibition of lens epithelial cell migration by a discontinuous capsular bend created by a band-shaped circular loop or a capsule-bending ring. *Ophthalmic Surg Lasers*. 1998;29(2):119-125.
- 113. Nishi O, Yamamoto N, Nishi K, Nishi Y. Contact inhibition of migrating lens epithelial cells at the capsular bend created by a sharp-edged intraocular lens after cataract surgery. *J Cataract Refract Surg.* 2007;33(6):1065-1070.
- 114. Zemaitiene R, Jasinskas V, Auffarth GU. Influence of three-piece and single-piece designs of two sharp-edge optic hydrophobic acrylic intraocular lenses on the prevention of posterior capsule opacification: a prospective, randomised, long-term clinical trial. Br J Ophthalmol. 2007;91(5):644-648.

- Ursell PG, Spalton DJ, Pande MV, et al. Relationship between intraocular lens biomaterials and posterior capsule opacification. J Cataract Refract Surg. 1998; 24(3):352-360.
- Cheng JW, Wei RL, Cai JP, et al. Efficacy of different intraocular lens materials and optic edge designs in preventing posterior capsular opacification: a meta-analysis. Am J Ophthalmol. 2007;143(3):428-436.
- 117. Biswas NR, Mongre PK, Das GK, Sen S, Angra SK, Vajpayee RB. Animal study on the effects of catalin on aftercataract and posterior capsule opacification. *Ophthalmic Res.* 1999;31(2):140-142.
- Power WJ, Neylan D, Collum LM. Daunomycin as an inhibitor of human lens epithelial cell proliferation in culture. *J Cataract Refract Surg.* 1994;20(3): 287-290
- Cortina P, Gómez-Lechón MJ, Navea A, Menezo JL, Terencio MC, Diaz-Llopis M.
 Diclofenac sodium and cyclosporin A inhibit human lens epithelial cell proliferation in culture. Graefes Arch Clin Exp Ophthalmol. 1997;235(3):180-185.
- Behar-Cohen FF, David T, D'Hermies F, et al. In vivo inhibition of lens regrowth by fibroblast growth factor 2-saporin. *Invest Ophthalmol Vis Sci.* 1995;36 (12):2434-2448.
- Kim JT, Lee DH, Chung KH, Kang IC, Kim DS, Joo CK. Inhibitory effects of salmosin, a disintegrin, on posterior capsular opacification in vitro and in vivo. *Exp Eye Res.* 2002;74(5):585-594.
- Ishida I, Saika S, Ohnishi Y. Effect of minoxidil on rabbit lens epithelial cell behavior in vitro and in situ. Graefes Arch Clin Exp Ophthalmol. 2001;239(10): 770-777.

- Chandler HL, Barden CA, Lu P, Kusewitt DF, Colitz CM. Prevention of posterior capsular opacification through cyclooxygenase-2 inhibition. *Mol Vis.* 2007; 13:677-691
- 124. Malecaze F, Lubsen NH, Serre B, et al. Lens cell targeting for gene therapy of prevention of posterior capsule opacification. *Gene Ther.* 2006;13(19):1422-1429.
- Malecaze F, Decha A, Serre B, et al. Prevention of posterior capsule opacification by the induction of therapeutic apoptosis of residual lens cells. *Gene Ther*. 2006;13(5):440-448.
- Walker JL, Wolff IM, Zhang L, Menko AS. Activation of SRC kinases signals induction of posterior capsule opacification. *Invest Ophthalmol Vis Sci.* 2007; 48(5):2214-2223.
- 127. Hosler MR, Wang-Su ST, Wagner BJ. Role of the proteasome in TGF-β signaling in lens epithelial cells. *Invest Ophthalmol Vis Sci.* 2006;47(5):2045-2052.
- Awasthi N, Wagner BJ. Suppression of human lens epithelial cell proliferation by proteasome inhibition, a potential defense against posterior capsular opacification. *Invest Ophthalmol Vis Sci.* 2006;47(10):4482-4489.
- 129. Awasthi N, Wang-Su ST, Wagner BJ. Downregulation of MMP-2 and -9 by proteasome inhibition: a possible mechanism to decrease LEC migration and prevent posterior capsular opacification. *Invest Ophthalmol Vis Sci.* 2008;49 (5):1998-2003.
- Kim SY, Kim JH, Choi JS, Joo CK. Comparison of posterior capsular opacification in rabbits receiving either mitomycin-C or distilled water for sealedcapsule irrigation during cataract surgery. *Clin Experiment Ophthalmol*. 2007; 35(8):755-758.

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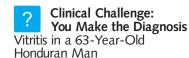
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