

POTASSIUM AND SODIUM OF RED BLOOD CELLS IN SICKLE CELL ANEMIA¹

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Normal human red cells contain approximately 100 mEq./l. of potassium (K) and 10 mEq./l. of sodium (Na). The K composition of sickle cell anemia (hereinafter designated SCA) red cells was first investigated by Erickson and her co-workers (1) who found that red cell [K] in SCA was approximately 10% less than normal red cell [K].

Our interest in the distribution of cations in SCA blood was stimulated by the observation of elevated venous serum [K] in three patients with SCA. Erickson and her co-workers did not report what effect varying blood oxygen content had on [K] and [Na] in SCA red cells. Since oxygen is known to affect other properties of SCA red cells (2, 3), it was decided to investigate the effect of controlling this variable. Therefore measurements of SCA red cell [Na] and [K] were made in both oxygenated and de-oxygenated states, *in vivo* and *in vitro*.

METHODS

1. Analytical techniques

K and Na were measured with a barrier layer cell flame photometer operated with a lithium internal standard (4). It is estimated that these measurements are accurate to $\pm 2\%$. When measured, water content of the red cells was determined by weighing a sample before and after drying in an oven at 110° C. for 24 or more hours. In some instances, pO_2 and pCO_2 in the gas phase of the experiments were measured by the method of Scholander (5), and the O_2 and CO_2 content of the blood phase by the technique of Van Slyke and Neill (6). The red cells for K and Na analyses were pipetted by syringe suction with an Ostwald pipette calibrated to contain the desired volume.

2. Experimental procedures

(a) *Short term experiments in vitro.* Forty to fifty ml. of blood were drawn from the antecubital vein of nor-

mal subjects and patients with SCA. The blood was immediately heparinized with approximately 50 cat units of crystalline heparin per 10 ml. blood. Within one-half to one hour, 15-20 ml. of this heparinized venous blood were placed in each of two Barcroft tonometers (300 ml. capacity), one containing 30% O_2 -5% CO_2 -65% N_2 , and the other containing 0% O_2 -5% CO_2 -95% N_2 . The tonometers were sealed with Hg and rotated at 20 rpm. in a water bath at 37° C. for 20-30 minutes. The blood was then removed under oil and, in some instances, whole blood samples taken for gas analysis. The remainder of the blood was centrifuged under oil at 14×10^3 g. for one hour, after which samples of plasma and red cells were taken for K, Na, and water analysis as indicated.

In order to test the reversibility of the cation changes observed in the experiments described above, a 60 ml. sample of fresh heparinized venous SCA blood was first de-oxygenated by equilibration for one-half hour in a Barcroft tonometer at 37° C. with 95% N_2 -5% CO_2 , as before. Then a 20 ml. sample was removed for analysis, the gas phase was flushed out with 95% O_2 -5% CO_2 , and the remaining 40 ml. of blood re-equilibrated for one-half hour with this gas. Next, a second sample was taken out, the final 20 ml. of blood were again deoxygenated by equilibrium in 95% N_2 -5% CO_2 . This blood was then the third sample. All blood samples were collected and centrifuged under oil at 800 g. for five minutes and plasma samples taken. The remaining loosely packed red cells were then centrifuged at 14×10^3 g. as before, the remaining plasma removed, and red cell samples taken for measurement of Na, K, and water content.

Since carbon monoxide prevents the occurrence of sickling of SCA red cells in a hypoxic medium (2), it was decided to utilize this fact to study the effect of hypoxia on cation changes in the absence of sickling. Forty ml. of fresh heparinized venous SCA or normal blood were gently shaken in a flask containing approximately 25% CO -72% N_2 -3% CO_2 , until cherry red in color. Twenty ml. samples of this blood were then placed in each of two Barcroft tonometers, one containing 95% O_2 -5% CO_2 , the other containing 95% N_2 -5% CO_2 . The tonometers were then sealed and rotated at 37° C. for one-half hour, after which samples were taken under oil and prepared for analysis as before.

(b) *Long term in vitro experiments.* In order to investigate changes in SCA red cell [K] over a longer period of time, a different technique was employed for several reasons: 1) SCA red cells kept in the de-oxygenated state for several hours become extremely viscous and

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TABLE I

Results of typical experiments on one normal subject and one patient with SCA
Each blood sample was incubated for one-half hour in the oxygenated and de-oxygenated state.

Subject	Gas		Blood gas		Red blood cell					Plasma			
	pO ₂	pCO ₂	O ₂	CO ₂	Na	K	Na+K	H ₂ O	pH	Na	K	Na+K	pH
	mm.Hg	mm.Hg	vol. %	vol. %	mEq./l.	mEq./l.	mEq./l.	%	calc.	mEq./l.	mEq./l.	mEq./l.	calc.
Normal	215	45	22.8	46	11	98	109	66.0	7.19	136	3.7	140	7.37
	12	42	3.4	50	11	96	107	66.0	7.25	136	3.7	140	7.44
Sickle cell anemia	237	42	8.5	50	27	83	110	64.1	7.19	134	4.5	139	7.37
	8	41	0.4	51	40	60	100	63.4	7.19	133	7.0	140	7.38

difficult to pipette, 2) heparinized SCA blood in a Barcroft tonometer rotated at 37° C. for 24 hours shows considerable hemolysis, 3) the vigorous centrifugation used to separate red cells and plasma might conceivably affect the results.

Forty ml. of fresh heparinized venous SCA blood were placed in a flask containing 60 ml. of a bicarbonate-phosphate buffered electrolyte solution, (pH 7.4; [K] = 4.4 mEq./l.; [Na] = 135 mEq./l.) containing 4.0 gm./l. of glucose. This solution is known to maintain normal hu-

man red cells in a relatively stable state for 48 hours (7). In our experiments visible hemolysis was present in only one sample and then was barely detectable by the naked eye. Twenty-five ml. of the resultant diluted blood were placed in each of four Parafilm sealed Erlenmeyer flasks with open side-arms to allow gas flow. Ninety-five% O₂-5% CO₂ was flushed through the flasks which were then rocked 12 × /min. through an arc of 30-40° in a water bath at 37° C. for one-half hour. One flask was then taken from the bath and the diluted blood removed under oil. Either 95% O₂-5% CO₂ or 95% N₂-5% CO₂ was flushed through the remaining three flasks which were then rocked in the bath as before. Samples of diluted blood were taken under oil at four, 12, and 24 hours. Hematocrit was determined on all samples by centrifugation at 800 g. for one hour. K in diluted blood and diluted plasma was measured by flame photometer, as before. Red cell [K] was calculated from these data by the following formula: $K_e = 1/\gamma K_b - K_p (1-\gamma)$ when K_e = red cell [K], K_b = diluted blood [K], K_p = diluted plasma [K], and γ = hematocrit corrected for incomplete packing (8).

(c) *In vivo* experiments. In order to investigate the initial phase of the cation changes *in vivo*, 20 ml. samples of blood were drawn simultaneously, under oil, from the antecubital vein and opposite brachial or femoral artery. Drawing of the venous sample was preceded by five to 30 minutes of venous congestion produced by a blood pressure cuff at 90 mm. Hg. The samples were immediately heparinized and centrifuged under oil at 14×10^3 g. for one hour. Samples of red cells and plasma were analyzed for K, Na, and water as before.

RESULTS

(a) Short-term experiments *in vitro*

In one experiment (Table I) typical of four with SCA red cells, [K] was about 23 mEq./l. RBC less and [Na] about 13 mEq./l. RBC more, in the de-oxygenated than in the oxygenated state. This loss of K and gain of Na was not evident upon de-oxygenation of normal cells. Even in the oxygenated state, SCA red cell [K] was somewhat less and [Na] somewhat more than the values in nor-

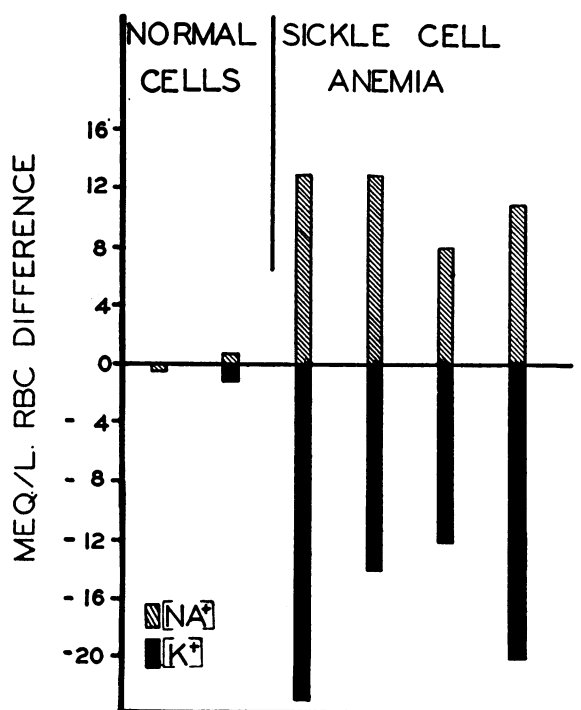


FIG. 1. EXPERIMENTS ON TWO NORMAL SUBJECTS AND FOUR PATIENTS WITH SCA—EACH BLOOD SAMPLE INCUBATED FOR ONE-HALF HOUR IN THE OXYGENATED AND DE-OXYGENATED STATE

Each bar represents the difference between oxygenated and de-oxygenated red cell sodium or potassium concentration.

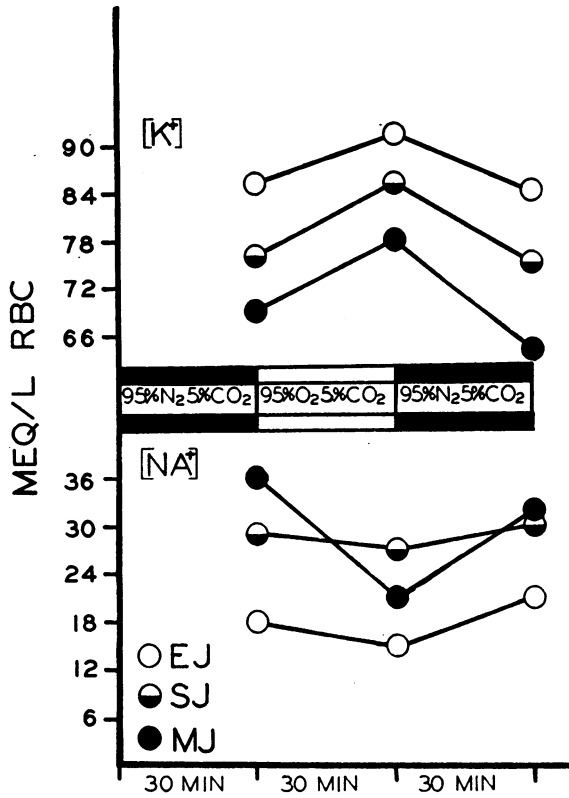


FIG. 2. EXPERIMENTS ON THREE PATIENTS WITH SCA—EACH BLOOD SAMPLE INCUBATED CONSECUTIVELY FOR ONE-HALF HOUR IN THE DE-OXYGENATED, OXYGENATED, AND DE-OXYGENATED STATE

mal red cells. Since the loss of K exceeded the gain of Na by de-oxygenated SCA red cells, the sum of [K⁺] + [Na⁺] was slightly less in the de-oxygenated than in the oxygenated state.

The water content of SCA red cells was slightly less than the value for normal red cells in the oxygenated state. De-oxygenation resulted in a slight fall in SCA red cell water content but did not affect normal red cell water content. Since the water content of de-oxygenated SCA red cells was, if anything, slightly less than the oxygenated value, it is unlikely that increased plasma contamination due to incomplete packing of the sickled de-oxygenated cells caused the observed reduced [K⁺] and increased [Na⁺] in these cells. pH as calculated by the method of Henderson (9) was essentially the same in normal and SCA red cells in oxygenated and de-oxygenated states. The Donnan r value for H⁺ was assumed to be the same for normal and SCA blood (10).

Figure 1 presents a graphic summary of data obtained from four SCA patients and two normal subjects. The results qualitatively confirmed all the findings described above for a single experiment. The blood of every SCA patient studied showed significantly less red cell K and more Na in the de-oxygenated than in the oxygenated state. These differences were not present in eight experiments on two normal subjects.

Figure 2 presents data which demonstrate the reversibility of the K loss and Na gain of previously de-oxygenated SCA red cells upon re-oxygenation. In experiments on three SCA patients, the mean rates of K gain by re-oxygenated and K loss by de-oxygenated red cells were both about 20 mEq./l. RBC/hr. The mean rates of Na loss by re-oxygenated and gain by de-oxygenated red cells were both about 14 mEq./l. RBC/hr.

Figure 3 shows that CO blocked the K loss and Na gain by de-oxygenated SCA red cells. CO did not affect the cation concentrations of normal red

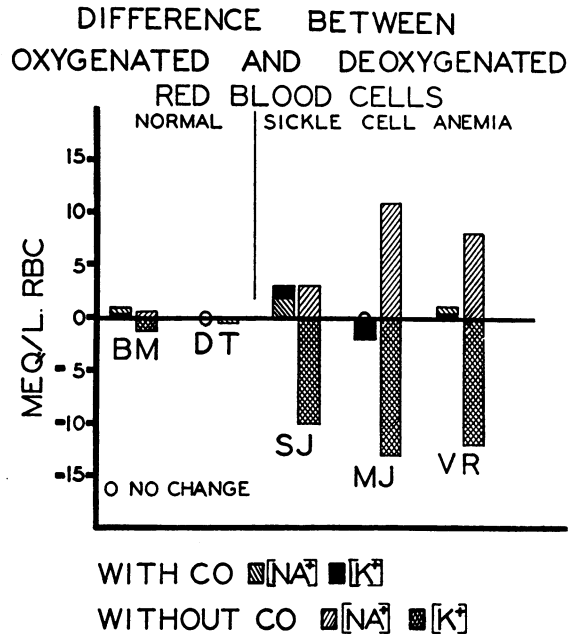


FIG. 3. EXPERIMENTS ON TWO NORMAL SUBJECTS AND THREE PATIENTS WITH SCA—EACH BLOOD SAMPLE INCUBATED FOR ONE-HALF HOUR IN 95% O₂ AND 95% N₂, WITH AND WITHOUT PRE-EXPOSURE TO 25% CO

The bar on the left of each pair represents the difference between oxygenated and de-oxygenated red cell sodium or potassium concentration with pre-exposure to CO. The bar on the right of each pair represents the same quantity without pre-exposure to CO.

cells. Thus, hypoxia *per se* does not cause the K-Na exchange observed in de-oxygenated SCA red cells.

(b) Long-term experiments *in vitro*

In the experiments with slow gentle shaking (see Methods) de-oxygenated SCA red cells continued to lose K for at least 24 hours (Figure 4). K loss from oxygenated SCA red cells was small and irregular in the two cases studied. The rate of K loss during the first four hours from de-oxygenated SCA red cells in this system was about 4 mEq./l. RBC/hr. The rate of loss decreased with time but was still appreciable between 12 and 24 hours after the start of the experiment. In a system identical with that employed here Raker and his associates (7) working with normal cells did not observe accelerated K loss upon de-oxygenation.

(c) Short-term experiments *in vivo*

Figure 5 shows the cation difference in four SCA patients between red cells from congested

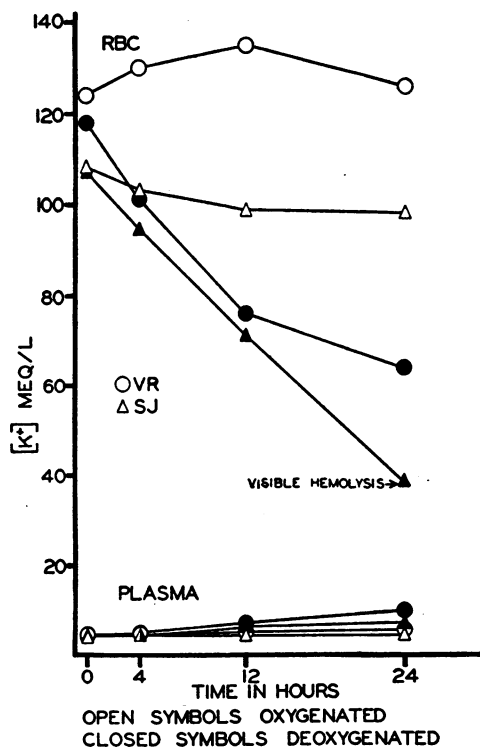


FIG. 4. EXPERIMENTS ON TWO PATIENTS WITH SCA—EACH SAMPLE OF DILUTED BLOOD INCUBATED FOR 24 HOURS IN THE OXYGENATED AND DE-OXYGENATED STATE

The de-oxygenated 24-hour sample of SJ showed hemolysis just detectable by the naked eye.

A-V DIFFERENCE IN RBC

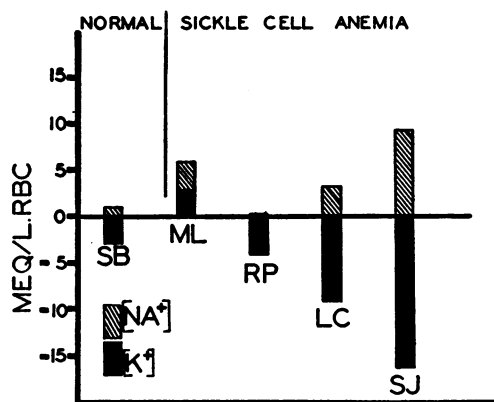


FIG. 5. EXPERIMENTS ON ONE NORMAL SUBJECT AND FOUR PATIENTS WITH SCA—EACH BAR REPRESENTING THE ARTERIOVENOUS DIFFERENCE IN RED CELL NA OR K CONCENTRATION

veins and arterial red cells. In general, venous red cell [K] was less and the [Na] more than the arterial values. The A-V differences varied from a maximum of -17 mEq. K/l. RBC and +9 mEq. Na/l. RBC in one patient to a minimum of +3 mEq. K/l. RBC and 0 mEq. Na/l. RBC in another. There was no A-V difference in plasma [K] so that we were unable to produce experimentally the clinically observed elevation of venous plasma [K]. In the normal subject, A-V differences in red cell and plasma [K] and [Na] were small.

DISCUSSION

Human red cells are known to lose K and gain Na on exposure to cold (11), X-rays (12), iodoacetate (13) and fluoride (13-15), in the absence of glucose (14, 15), and possibly upon infestation with malarial parasites (16). Ponder describes red cell K loss and Na uptake prior to hemolysis in a variety of *in vitro* hemolytic systems (17). The mechanism of such "prolytic" K-Na exchange is obscure. The electrolyte shift which we found during hypoxia is, in our experience, unique to SCA and possibly sickle cell trait red cells. Whether this process is also "prolytic" is beyond the scope of this paper.

The initial rate of K loss from SCA red cells de-oxygenated in a Barcroft tonometer (about 20 mEq./l. RBC/hr.) was considerably faster than the value for SCA red cells de-oxygenated in an

Erlenmeyer flask (about 4 mEq./l. RBC/hr.). This difference was perhaps due to the slower rate of agitation or of equilibration of blood with gas phase in the latter system. During at least the first one-half hour of de-oxygenation the K loss could be reversed by re-oxygenation. In this circumstance, K uptake also occurred at an initial rate of about 20 mEq./l. RBC/hr. Since these rates of net K uptake and loss were considerably faster than the rate of turnover of K in normal red cells in the steady state (2 mEq./l. RBC/hr.) (7, 18), it is probable that under certain conditions K turnover in SCA red cells is considerably faster than in normal red cells. Experiments are now in progress to determine the effect of variations in temperature, concentration gradient, etc. on the rates of net K and Na transfer and to measure with appropriate isotopes the rate of turnover of these ions in SCA red cells.

Why do these electrolyte changes occur in de-oxygenated SCA red cells but not in normal cells? Two general hypotheses are plausible:

(1) K-Na exchange is due to an abnormality in SCA plasma. For example, "prolytic" ion exchange as described by Ponder is particularly large in hemolytic systems containing bile salts. Most SCA patients were slightly jaundiced but accurate determinations were not made of the concentration of bile salts in their plasma. It does not seem possible that plasma bile salts *per se* cause the cation exchange observed in SCA red cells because (a) it is unlikely that reduction in pO_2 would accelerate such a process, and (b) red cells taken from a patient with severe obstructive jaundice due to carcinoma of the head of the pancreas did not lose K or gain Na during 24 hours of incubation at 37° C. in the oxygenated or de-oxygenated state. Since the shift occurred in de-oxygenated SCA red cells suspended in diluted plasma and (in one experiment) Ringer's solution, evidence for a plasma abnormality responsible for the process is at present lacking.

(2) K-Na exchange is due to an abnormality in SCA red cells. The most obvious difference between the responses of normal and SCA red cells to hypoxia is the occurrence of sickling in the latter. Several facts suggest that K loss and Na uptake by de-oxygenated SCA red cells is intimately associated with the process of sickling: (a) CO prevents both sickling and K-Na exchange in de-oxy-

genated SCA red cells, (b) both processes begin almost immediately after de-oxygenation of SCA blood.

What is the mechanism relating the processes of sickling and K-Na exchange in SCA red cells? The change of shape and probable mechanical distortion of the red cell membrane in the sickled state are obvious. These changes could accelerate the diffusion of K and Na in the direction of their concentration gradients. In addition, recently demonstrated differences between ionic (19), and physical (3, 12, 20) properties of SCA and normal hemoglobin may be related to cation exchange in sickled SCA red cells. Specifically, the work of Harris (3) and of Perutz and Mitchison (20) suggests that unsaturated SCA hemoglobin is in a crystalline or para-crystalline state. It is not yet clear whether the observed changes in cation composition of *sickled* SCA red cells are primarily related to the physical properties of desaturated SCA hemoglobin or to alteration of the SCA red cell membrane incident to the sickle shape.

SUMMARY AND CONCLUSIONS

1. In the de-oxygenated but not in the oxygenated state, red cells from patients with sickle cell anemia (SCA) begin to lose K and gain Na almost immediately and continue to do so for at least 24 hours. This process does not occur in de-oxygenated normal red cells.
2. During the first half hour at least, K-Na exchange in de-oxygenated SCA red cells can be reversed by re-oxygenation.
3. K loss and Na gain by red cells does not occur in the absence of O_2 if sickling is prevented by CO.

REFERENCES

1. Erickson, B. N., Williams, H. H., Hummel, F. C., Lee, P., and Macy, I. G., The lipid and mineral distribution of the serum and erythrocytes in the hemolytic and hypochromic anemias of childhood. *J. Biol. Chem.*, 1937, **118**, 569.
2. Hahn, E. V., and Gillespie, E. B., Sickle cell anemia: report of a case greatly improved by splenectomy. Experimental study of sickle cell formation. *Arch. Int. Med.*, 1927, **39**, 233.
3. Harris, J. W., Studies on the destruction of red blood cells. VIII. Molecular orientation in sickle cell hemoglobin solutions. *Proc. Soc. Exper. Biol. & Med.*, 1950, **75**, 197.

4. Berry, J. W., Chapell, D. G., and Barnes, R. B., Improved method of flame photometry. *Indust. & Engin. Chem. (Analyt Ed.)*, 1946, 18, 19.
5. Scholander, P. F., Analyzer for accurate estimation of respiratory gases in one-half cubic centimeter samples. *J. Biol. Chem.*, 1947, 167, 235.
6. Van Slyke, D. D., and Neill, J. M., The determination of gases in blood and other solutions by vacuum extraction and manometric measurement. *J. Biol. Chem.*, 1924, 61, 523.
7. Raker, J. W., Taylor, I. M., Weller, J. M., and Hastings, A. B., Rate of potassium exchange of the human erythrocyte. *J. Gen. Physiol.*, 1950, 33, 691.
8. MILLAR, W. G., Observations on the haematocrite method of measuring the volume of erythrocytes. *Quart. J. Exper. Physiol.*, 1925, 15, 187.
9. Henderson, L. J., *Blood, a Study in General Physiology*. Yale Univ. Press, New Haven, 1928.
10. Dill, D. B., Edwards, H. T., and Consolazio, W. V., Blood as a physico-chemical system. XI. Man at rest. *J. Biol. Chem.*, 1937, 118, 635.
11. Smith, M. E., Tuthill, E., Drew, C. R., and Scudder, J., Studies in blood preservation. Some effects of carbon dioxide. *J. Biol. Chem.*, 1940, 133, 499.
12. Sheppard, C. W., and Beyl, G. E., Cation exchange in mammalian erythrocytes. III. The prolytic effect of x-rays on human cells. *J. Gen. Physiol.*, 1951, 34, 691.
13. Wilbrandt, W., A relation between the permeability of the red cell and its metabolism. *Tr. Faraday Soc.*, 1937, 33, 956.
14. Harris, J. E., The influence of the metabolism of human erythrocytes on their potassium content. *J. Biol. Chem.*, 1941, 141, 579.
15. Danowski, T. S., The transfer of potassium across the human blood cell membrane. *J. Biol. Chem.*, 1941, 139, 693.
16. Overman, R. R., Hill, T. S., and Wong, Y. T., Physiological studies in the human malarial host. I. Blood, plasma, extracellular fluid volumes and ionic balance in therapeutic *P. vivax* and *P. falciparum* infections. *J. Nat. Malaria Soc.*, 1949, 8, 14.
17. Ponder, E., *Hemolysis and Related Phenomena*. Grune & Stratton, New York, 1948.
18. Sheppard, C. W., and Martin, W. R., Cation exchange between cells and plasma of mammalian blood. I. Methods and application to potassium exchange in human blood. *J. Gen. Physiol.*, 1950, 33, 703.
19. Pauling, L., Itano, H. A., Singer, S. J., and Wells, I. C., Sickle cell anemia, a molecular disease. *Science*, 1949, 110, 543.
20. Perutz, M. F., and Mitchison, J. M., State of haemoglobin in sickle-cell anaemia. *Nature*, 1950, 166, 677.