

Supplementary Material

Potential of Bacteriophage Proteins as Recognition Molecules for Pathogen Detection

Susana P. Costa^{1,2,3}, Catarina L. Nogueira^{2,3}, Alexandra M. Cunha^{1,2}, Ana Lisac⁴, Carla M. Carvalho^{2*}

1. Centre of Biological Engineering, University of Minho, Campus de Gualtar, 4710-057, Braga, Portugal
2. International Iberian Nanotechnology Laboratory, Av. Mestre José Veiga s/n, 4715-330, Braga, Portugal
3. Instituto de Engenharia de Sistemas e Computadores – Microsistemas e Nanotecnologias (INESC MN) and IN – Institute of Nanoscience and Nanotechnology, Rua Alves Redol, 9 1000-029 Lisbon, Portugal
4. Faculty of Chemistry and Chemical Technology, University of Ljubljana, Kongresni trg 12 1000 Ljubljana, Slovenia

*Corresponding author: carla.carvalho@inl.int

Table S1 - Summary of several RBP-based systems available for the detection of bacterial pathogens.

Targeted pathogen	Phage RBP	Sample	Detection system	Detection limit	Response time	Ref.
<i>Campylobacter</i>	gp47*	Bacterial suspension	RBP-based agglutination assay, immunostaining, and fluorescence microscopy	Not reported	Not reported	(1)
	gp48*	Milk and chicken broth	RBP-based MS with TaqMan real-time PCR assay	10 ² CFU/mL	3 h	(2)
	gp48	Bacterial suspension	RBP-based MS, SPR and fluorescence microscope	10 ² CFU/mL	Not reported	(3)
	gp48	Bacterial suspension	RBP-based microresonator array platform	Single-cell limit	Not reported	(4)
<i>Salmonella</i>	gp37-gp38 complex	Bacterial suspension and spiked chicken, milk, chocolate milk and infant formula	RBP-based MS and ELLTA	1-10 CFU/mL	45 min	(5)
	RBP from phage P22	Bacterial suspension	RBP-NAEBs detection probes with SERS biosensor	Single-cell limit	Not reported	(6)
	RBPs from phage P22	Bacterial suspension	RBP-based SPR biosensor, fluorescence microscopy and scanning electron microscopy (SEM)	10 ³ CFU/mL	Not reported	(7)
	RBPs from phages 9NA and P22	Bacterial suspension	RBP-based ELITA assay, MALDI-MS and flow cytometry	Not reported	Not reported	(8)
	Det7T	Bacterial suspension and spiked apple juice	RBP-based SPR biosensor	5 × 10 ⁴ – 5 × 10 ⁷ CFU/mL	20 min	(9)
<i>Escherichia coli</i>	gp37	Bacterial suspension	RBP-based LPG device	Not reported	Not reported	(10)
	gp37	Bacterial suspension	RBP-based LPG device	Not reported	1.5 h	(11)
	gp37	Bacterial suspension	Label-free RBP-based microwave sensors	Not reported	2 h	(12)
	gp37	Bacterial suspension	LPG-based device nano-coated with Tantalum oxide (TaO _x) thin layer	Not reported	30 min	(13)
	gp37	Bacterial suspension	Low-voltage field-emission scanning electron microscopy (LV-FESEM), energy-dispersive spectroscopy (EDS) and ELISA	Not reported	Not reported	(14)
	J	Bacterial suspension	RBP-based SPR	2 × 10 ⁴ CFU/mL	20 min	(15)

<i>Acinetobacter baumannii</i>	TF2 and TF6	Bacterial suspension	RBP-based MS, MALDI-MS	10 ⁴ -10 ⁵ cells/mL	10 min	(16)
<i>Shigella flexneri</i>	RBP from phage SF6	Bacterial suspension	RBP-based ELITA and fluorescence spectroscopy	10 ³ CFU/mL	Not reported	(17)
<i>Pseudomonas aeruginosa</i>	P069	Bacterial suspension and spiked human urine, glucose injection and rat serum	RBP-based MS with bioluminescence and fluorescence spectroscopy	10 ² CFU mL	Not reported	(18)
	P069	Bacterial suspension and spiked milk, human urine, and physiological saline	Fluorescence microscopy and spectrofluorometry	2 × 10 ³ CFU/mL	100 min	(19)
<i>Yersinia pestis</i>	gpH and gp17	Bacterial suspension	Fluorescence microscopy	Not reported	Not reported	(20)
<i>Bacillus anthracis</i>	RBPs from phages γ , Wip1, AP50c and λ Ba03	Bacterial suspension	Fluorescence microscopy	Not reported	10 min	(21)
<i>Enterococcus</i> and <i>Staphylococcus</i>	gp109 and gp18	Bacterial suspension and spiked blood	Epifluorescence microscopy and spectrofluorometry	10 ⁸ CFU/mL	1.5 h	(22)
	gp109 and gp18	Bacterial suspension	RBP-based MS with MR Platform	10 CFU/mL	2 h	(23)
<i>Klebsiella pneumoniae</i>	gp86	Bacterial suspension	Epifluorescence microscopy and spectrofluorometry	10 ⁸ CFU/mL	1.5 h	(24)

*gp47 and gp48 correspond to the same protein (1,25) but for information purposes are described herein as in the original papers. PCR - Polymerase chain reaction; SPR - Surface plasmon resonance; SERS - Surface-enhanced Raman scattering; NAEBs – Nanoaggregate embedded beads; LPG - Long period grating; MS -Magnetic separation; ELISA -Enzyme-linked immunosorbent assay; ELITA- ELISA-like tail spike absorption assay; ELLTA – Enzyme-linked long tail fibre assay; MR - magnetoresistive.

Table S2 - Summary of several phage endolysin CBD-based systems available for the detection of bacterial pathogens.

Targeted pathogen	Phage CBD	Sample	Detection system	Detection limit	Response time	Ref.
<i>Listeria monocytogenes</i>	CBD118 (Ply118) and CBD500 (Ply500)	Infected PtK2 epithelial cells	Fluorescence microscopy and CBD-based SPR	Not reported	Not reported	(26)
	CBD118 (Ply118) and CBD500 (Ply500)	Meat, poultry, fish, dairy products	CBD-MS	0.1 CFU/g	48 h	(27)
	CBD118 (Ply118) and CBD500 (Ply500)	Spiked raw milk	CBD-based MS, Culture-Based methods, and Real-Time PCR	10 ² - 10 ³ CFU/mL	Not reported	(28)
	CBDP40 (PlyP40), CBDP35 (PlyP35), CBD500 (Ply500), CBDP35 (PlyP35), CBD025 (PlyP025)	Spiked milk and cheese	CBD-based MS, fluorescence microscopy	Not reported	At least 24 h	(29)
	CBD500 (Ply500)	Cell suspension and spiked milk	CBD-based electrochemical impedance biosensor	1.1 x 10 ⁴ CFU/mL (suspension) 1.1 x 10 ⁵ CFU/mL (milk)	Not reported	(30)
<i>Listeria</i> spp.	CBD118 (Ply118) and CBD500 (Ply500)	Cell suspension and 9 different food items	CBD-based MS and bioluminescent reporter phage (spectrophotometry)	10 ² CFU/mL (suspension) 0.1-1 CFU/g (food)	22 h	(31)
<i>Bacillus anthracis</i>	CBD of PlyG endolysin	Encapsulated cells in suspension and spores	CBD-based Membrane direct blot assay	10 ³ CFU/mL	3 h	(32)
	Putative binding motifs of PlyG endolysin (PlyP1-P6)	Spiked plasma	QD based-Fluorometric analysis and Fluorescence microscopy	Not reported (single bacterium for PlyP3 peptide)	Not reported	(33)
<i>Bacillus cereus</i>	CBD of PBC1 endolysin	Spiked cooked rice	SPR-based detection method	10 ² CFU/mL (suspension) 10 ³ CFU/mL (food)	Not reported	(34)
	CBD of endolysin LysB4	Cell suspension	Nitrocellulose-based lateral flow assay	10 ⁴ CFU/mL	20 min	(35)
	CBD of PBC1 endolysin	Cell suspension and blood	Bioluminescent assay (spectrophotometry) with CBD-based MS	10 CFU/mL (suspension) 10 ³ CFU/mL (blood)	At least 30 min	(36)

	CBD and SBD of LysPBC2	Cell suspension and spores	Fluorescence microscopy and spectrometry	Not reported	Not reported	(37)
<i>Clostridium</i> spp.	CBD of endolysin CTP1L	Cheese	GFP based protein - Fluorescence microscopy	3 spores/g	Not reported	(38)
<i>Mycobacterium avium</i> subsp. <i>paratuberculosis</i>	CBD of gp10 endolysin	Spiked milk	Pre-concentration by CBD-IMS and PCR	2.4 x 10 ¹ CFU/mL	24 h	(39)
<i>Paenibacillus larvae</i>	CBD of PlyPI23 endolysin	Homogenized honeybee larvae	Fluorescence microscopy	Not reported	Not reported	(40)
<i>S. aureus</i> , <i>Bacillus anthracis</i> -Sterne and <i>Listeria innocua</i>	lysostaphin CBD, AmiBA2446 CBD and Ply500 CBD	Cell suspension	CBD-based ELISA sandwich coupled with spectrophotometric assay and qPCR	10 ³ CFU/mL (<i>S. aureus</i> and <i>B. anthracis</i>), 10 ⁴ CFU/mL (<i>L. innocua</i>); qPCR: 2 CFU/mL (<i>S. aureus</i>), 3 CFU/mL (<i>B. anthracis</i>), 6 CFU/mL <i>L. innocua</i>)	Not reported	(41)
<i>Staphylococcus aureus</i>	CBD of endolysin PlyV12	Spiked milk	IMS and EL-CBD	4 x 10 ³ CFU/mL	1.5 h	(42)
		Buffer and spiked serum and milk	CBD-based MS and spectrophotometry with fluorescent Amplex red/hydrogen peroxide-based assay	78 CFU/mL (buffer) 525 CFU/mL (milk) 348 CFU/mL (serum)	50 min	(43)
	Full-length LysK and truncated LysK	Cell suspension	Immobilization on silicon wafers, XPS and fluorescence microscopy	5 x 10 ³ CFU/mL	Not reported	(44)
<i>Staphylococcus</i> spp.	AMICBD of E-LM12 endolysin	Cell suspension and spiked blood	AMICBD -based flow cytometer assay	1-5 CFU/mL	18 h	(45)
Methicillin-resistant <i>Staphylococcus aureus</i>	CBD from the P108 phage endolysin	Cell suspension	CBD-based lateral flow sandwich assay	6.6 x 10 ² - 6.6 x 10 ⁷ CFU/mL	10 min	(46)
		physiological saline injection, lake water and human urine	CBD-MS and CBD-based flow cytometry assay	1.5 x 10 ² - 1.5 x 10 ⁶ CFU/mL	Not reported	(47)

PCR - Polymerase chain reaction; IMS - Immunomagnetic separation; MS -Magnetic separation; EL-Enzyme linked; ELISA - Enzyme-linked immunosorbent assay; XPS - X-ray photoelectron spectroscopy; QD - Quantum-dots

References

1. Javed MA, Poshtiban S, Arutyunov D, Evoy S, Szymanski CM. Bacteriophage Receptor Binding Protein Based Assays for the Simultaneous Detection of *Campylobacter jejuni* and *Campylobacter coli*. *PLoS One*. 2013;8(7).
2. Poshtiban S, Javed MA, Arutyunov D, Singh A, Banting G, Szymanski CM, et al. Phage receptor binding protein-based magnetic enrichment method as an aid for real time PCR detection of foodborne bacteria. *Analyst*. 2013;138(19):5619–26.
3. Singh A, Arutyunov D, McDermott MT, Szymanski CM, Evoy S. Specific detection of *Campylobacter jejuni* using the bacteriophage NCTC 12673 receptor binding protein as a probe. *Analyst*. 2011;136(22):4780–6.
4. Poshtiban S, Singh A, Fitzpatrick G, Evoy S. Bacteriophage tail-spike protein derivitized microresonator arrays for specific detection of pathogenic bacteria. *Sensors Actuators, B Chem*. 2013;181:410–6.
5. Denyes JM, Dunne M, Steiner S, Mittelviefhaus M, Weiss A, Schmidt H, et al. Modified bacteriophage S16 long tail fiber proteins for rapid and specific immobilization and detection of *Salmonella* cells. *Appl Environ Microbiol*. 2017;83(12):1–15.
6. Tay LL, Huang PJ, Tanha J, Ryan S, Wu X, Hulse J, et al. Silica encapsulated SERS nanoprobe conjugated to the bacteriophage tailspike protein for targeted detection of *Salmonella*. *Chem Commun*. 2012 Jan 25;48(7):1024–6.
7. Singh A, Arya SK, Glass N, Hanifi-Moghaddam P, Naidoo R, Szymanski CM, et al. Bacteriophage tailspike proteins as molecular probes for sensitive and selective bacterial detection. *Biosens Bioelectron*. 2010 Sep 15;26(1):131–8.
8. Schmidt A, Rabsch W, Broeker NK, Barbirz S. Bacteriophage tailspike protein based assay to monitor phase variable glucosylations in *Salmonella* O-antigens. *BMC Microbiol*. 2016;16(1):1–11.
9. Hyeon SH, Lim WK, Shin HJ. Novel surface plasmon resonance biosensor that uses full-length Det7 phage tail protein for rapid and selective detection of *Salmonella enterica* serovar Typhimurium. *Biotechnol Appl Biochem*. 2020;5:bab.1883.
10. Brzozowska E, Śmietana M, Koba M, Górska S, Pawlik K, Gamian A, et al. Recognition of bacterial lipopolysaccharide using bacteriophage-adhesin-coated long-period gratings. *Biosens Bioelectron*. 2015;67:93–9.
11. Brzozowska E, Koba M, Śmietana M, Górska S, Janik M, Gamian A, et al. Label-free Gram-negative bacteria detection using bacteriophage-adhesin-coated long-period gratings. *Biomed Opt Express*. 2016;7(3):829.
12. Rydosz A, Brzozowska E, Górska S, Wincza K, Gamian A, Gruszczynski S. A broadband capacitive sensing method for label-free bacterial LPS detection. *Biosens Bioelectron*. 2016 Jan 15;75:328–36.
13. Piestrzyńska M, Dominik M, Kosiel K, Janczuk-Richter M, Szot-Karpińska K, Brzozowska E, et al. Ultrasensitive tantalum oxide nano-coated long-period gratings for detection of various biological targets. *Biosens Bioelectron*. 2019;133:8–15.
14. Górska S, Rydosz A, Brzozowska E, Drab M, Wincza K, Gamian A, et al. Effectiveness of

- sensors contact metallization (Ti, Au, and Ru) and biofunctionalization for *Escherichia coli* detection. *Sensors (Switzerland)*. 2018;18(9).
15. Shin HJ, Lim WK. Rapid label-free detection of *E. coli* using a novel SPR biosensor containing a fragment of tail protein from phage lambda. *Prep Biochem Biotechnol*. 2018;48(6):498–505.
 16. Bai YL, Shahed-Al-Mahmud M, Selvaprakash K, Lin NT, Chen YC. Tail Fiber Protein-Immobilized Magnetic Nanoparticle-Based Affinity Approaches for Detection of *Acinetobacter baumannii*. *Anal Chem*. 2019;91(15):10335–42.
 17. Kunstmann S, Scheidt T, Buchwald S, Helm A, Mulard LA, Fruth A, et al. Bacteriophage Sf6 tailspike protein for detection of shigella flexneri pathogens. *Viruses*. 2018;10(8).
 18. He Y, Shi Y, Liu M, Wang Y, Wang L, Lu S, et al. Nonlytic Recombinant Phage Tail Fiber Protein for Specific Recognition of *Pseudomonas aeruginosa*. *Anal Chem*. 2018;90(24):14462–8.
 19. Shi Y, He Y, Zhang L, Wang L, Fu Z. Dual-site recognition of *Pseudomonas aeruginosa* using polymyxin B and bacteriophage tail fiber protein. *Anal Chim Acta*. 2021;1180:338855.
 20. Born F, Braun P, Scholz HC, Grass G. Specific detection of *Yersinia pestis* based on receptor binding proteins of phages. *Pathogens*. 2020;9:1–19.
 21. Braun P, Wolfschläger I, Reetz L, Bachstein L, Jacinto AC, Tocantins C, et al. Rapid microscopic detection of *Bacillus anthracis* by fluorescent receptor binding proteins of bacteriophages. *Microorganisms*. 2020;8(6):1–21.
 22. Santos SB, Cunha AP, Macedo M, Nogueira CL, Brandão A, Costa SP, et al. Bacteriophage-receptor binding proteins for multiplex detection of *Staphylococcus* and *Enterococcus* in blood. *Biotechnol Bioeng*. 2020;117(11):3286–98.
 23. Cunha AP, Henriques R, Cardoso S, Freitas PP, Carvalho CM. Rapid and multiplex detection of nosocomial pathogens on a phage-based magnetoresistive lab-on-chip platform. *Biotechnol Bioeng*. 2021;118(8):3164–3174.
 24. Nogueira CL, Pires DP, Monteiro R, Santos SB, Carvalho CM. Exploitation of a *Klebsiella* Bacteriophage Receptor-Binding Protein as a Superior Biorecognition Molecule. *ACS Infect Dis*. 2021;7:3077–3087.
 25. Kropinski AM, Arutyunov D, Foss M, Cunningham A, Ding W, Singh A, et al. Genome and Proteome of *Campylobacter jejuni* Bacteriophage NCTC 12673. *Appl Environ Microbiol*. 2011;77(23):8265–71.
 26. Loessner MJ, Kramer K, Ebel F, Scherer S. C-terminal domains of *Listeria monocytogenes* bacteriophage murein hydrolases determine specific recognition and high-affinity binding to bacterial cell wall carbohydrates. *Mol Microbiol*. 2002;44(2):335–49.
 27. Kretzer JW, Lehmann R, Schmelcher M, Banz M, Kim K-PP, Korn C, et al. Use of high-affinity cell wall-binding domains of bacteriophage endolysins for immobilization and separation of bacterial cells. *Appl Environ Microbiol*. 2007;73(6):1992–2000.
 28. Walcher G, Stessl B, Wagner M, Eichenseher F, Loessner MJ, Hein I. Evaluation of Paramagnetic Beads Coated with Recombinant *Listeria* Phage Endolysin – Derived Cell-Wall-Binding Domain. *Foodborne Pathog Dis*. 2010;7(9):1019–24.
 29. Schmelcher M, Shabarova T, Eugster MR, Eichenseher F, Tchang VS, Banz M, et al. Rapid multiplex detection and differentiation of *Listeria* cells by use of fluorescent phage endolysin cell wall binding domains. *Appl Environ Microbiol*. 2010;76(17):5745–56.

30. Tolba M, Ahmed MU, Tlili C, Eichenseher F, Loessner MJ, Zourob M. A bacteriophage endolysin-based electrochemical impedance biosensor for the rapid detection of *Listeria* cells. *Analyst*. 2012;137(24):5749–56.
31. Kretzer JW, Schmelcher M, Loessner MJ. Ultrasensitive and Fast Diagnostics of Viable *Listeria* Cells by CBD Magnetic Separation Combined with A511::luxAB Detection. *Viruses*. 2018;10(11).
32. Fujinami Y, Hirai Y, Sakai I, Yoshino M, Yasuda J. Sensitive detection of *Bacillus anthracis* using a binding protein originating from γ -phage. *Microbiol Immunol*. 2007;51(2):163–9.
33. Sainathrao S, Mohan K, Atreya C. Gamma-phage lysin PlyG sequence-based synthetic peptides coupled with Qdot-nanocrystals are useful for developing detection methods for *Bacillus anthracis* by using its surrogates, *B. anthracis*-Sterne and *B. cereus*-4342. *BMC Biotechnol*. 2009;9:67.
34. Kong M, Sim J, Kang T, Nguyen HH, Park HK, Chung BH, et al. A novel and highly specific phage endolysin cell wall binding domain for detection of *Bacillus cereus*. *Eur Biophys J*. 2015;44(6):437–46.
35. Kong M, Shin JH, Heu S, Park JK, Ryu S. Lateral flow assay-based bacterial detection using engineered cell wall binding domains of a phage endolysin. *Biosens Bioelectron*. 2017;96:173–7.
36. Park C, Kong M, Lee JH, Ryu S, Park S. Detection of *Bacillus Cereus* Using Bioluminescence Assay with Cell Wall-binding Domain Conjugated Magnetic Nanoparticles. *Biochip J*. 2018;12(4):287–93.
37. Kong M, Na H, Ha N-C, Ryu S. LysPBC2, a Novel Endolysin Harboring a *Bacillus cereus* Spore Binding Domain. *Appl Envir Microbiol*. 2019;85(5):e02462-18.
38. Gómez-Torres N, Dunne M, Garde S, Meijers R, Narbad A, Ávila M, et al. Development of a specific fluorescent phage endolysin for in situ detection of *Clostridium* species associated with cheese spoilage. *Microb Biotechnol*. 2018;11(2):332–45.
39. Singh U, Arutyunov D, Basu U, Santos Seckler H Dos, Szymanski CM, Evoy S. Mycobacteriophage lysin-mediated capture of cells for the PCR detection of *Mycobacterium avium* subspecies paratuberculosis. *Anal Methods*. 2014;6(15):5682–9.
40. Santos SB, Oliveira A, Melo LDR, Azeredo J. Identification of the first endolysin Cell Binding Domain (CBD) targeting *Paenibacillus larvae*. *Sci Rep*. 2019;9(1):1–9.
41. Kwon SJ, Kim D, Lee I, Nam J, Kim J, Dordick JS. Sensitive multiplex detection of whole bacteria using self-assembled cell binding domain complexes. *Anal Chim Acta*. 2018;1030:156–65.
42. Yu J, Zhang Y, Zhang Y, Li H, Yang H, Wei H. Sensitive and rapid detection of *Staphylococcus aureus* in milk via cell binding domain of lysin. *Biosens Bioelectron*. 2016;77:366–71.
43. Yi Z, Wang S, Meng X, Wu A, Li Q, Song Y, et al. Lysin cell-binding domain-functionalized magnetic beads for detection of *Staphylococcus aureus* via inhibition of fluorescence of Amplex Red/hydrogen peroxide assay by intracellular catalase. *Anal Bioanal Chem*. 2019;411(27):7177–85.
44. Chibli H, Ghali H, Park S, Peter YA, Nadeau JL. Immobilized phage proteins for specific detection of *Staphylococci*. *Analyst*. 2014;139(1):179–86.
45. Costa SP, Dias NM, Melo LDR, Azeredo J, Santos SB, Carvalho CM. A novel flow cytometry

- assay based on bacteriophage-derived proteins for Staphylococcus detection in blood. *Sci Rep.* 2020;10:6260.
46. Yang H, Wang Y, Liu S, Ouyang H, Lu S, Li H, et al. Lateral flow assay of methicillin-resistant Staphylococcus aureus using bacteriophage cellular wall-binding domain as recognition agent. *Biosens Bioelectron.* 2021;182:113189.
 47. Wang Y, He Y, Bhattacharyya S, Lu S, Fu Z. Recombinant Bacteriophage Cell-Binding Domain Proteins for Broad-Spectrum Recognition of Methicillin-Resistant Staphylococcus aureus Strains. *Anal Chem.* 2020;92(4):3340–5.