

Preanalytical management: serum vacuum tubes validation for routine clinical chemistry

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Abstract

Introduction: The validation process is essential in accredited clinical laboratories. Aim of this study was to validate five kinds of serum vacuum tubes for routine clinical chemistry laboratory testing.

Materials and methods: Blood specimens from 100 volunteers in five different serum vacuum tubes (Tube I: VACUETTE[®], Tube II: LABOR IMPORT[®], Tube III: S-Monovette[®], Tube IV: SST[®] and Tube V: SST II[®]) were collected by a single, expert phlebotomist. The routine clinical chemistry tests were analyzed on cobas[®] 6000 <501> module. The significance of the differences between samples was assessed by paired Student's t-test after checking for normality. The level of statistical significance was set at $P < 0.005$. Finally, the biases from Tube I, Tube II, Tube III, Tube IV and Tube V were compared with the current desirable quality specifications for bias (B), derived from biological variation.

Results and conclusions: Basically, our validation will permit the laboratory or hospital managers to select the brand's vacuum tubes validated according him/her technical or economical reasons, in order to perform the following laboratory tests: glucose, total cholesterol, high density lipoprotein-cholesterol, triglycerides, total protein, albumin, blood urea nitrogen, uric acid, alkaline phosphatase, aspartate aminotransferase, gamma-glutamyltransferase, lactate dehydrogenase, creatine kinase, total bilirubin, direct bilirubin, calcium, iron, sodium and potassium. On the contrary special attention will be required if the laboratory already performs creatinine, amylase, phosphate and magnesium determinations and the quality laboratory manager intend to change the serum tubes. We suggest that laboratory management should both standardize the procedures and frequently evaluate the quality of in vitro diagnostic devices.

Key words: preanalytical variability; blood collection; serum samples; vacuum tubes; validation process

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Introduction

Laboratory testing is an integral part of the decision-making process, and results of laboratory testing often strongly influence medical diagnoses and therapies. There is a long history of quality requirements in laboratory medicine, which have mainly concerned the analytic phase of this process (1). Owing to the substantial advances in technology, laboratory automation and analytic quality, there is increasing evidence that further quality improvements should be targeted to extra-analyt-

ic phases of laboratory testing (2-6). Clinical laboratories routinely use commercial diagnostic products during the testing process. Diagnostic products can be divided into two major categories: in vitro diagnostic (IVD) devices, such as laboratory instruments, reagents, assays and blood collection tubes, and medical devices, such as specimen collection devices (needles and sets) (7). Necessary improvements and potential sources of nonconformities, either technical or concerning the quali-

ty management system, shall be identified and all laboratory process shall be validated (8). Some IVD devices (e.g. blood collection vacuum tubes) are not validated before the quality laboratory managers decide to start using or to change the brand. The aim of this study was to validate five kinds of serum vacuum tubes for routine clinical chemistry laboratory testing.

Materials and methods

Study design

A group of 100 adult ambulatory patients of both genders, volunteers for this study, from Dante Pazzanese Cardiology Institute, São Paulo city, Brazil, were evaluated between 1st September and 1st October 2011. This study was submitted to the Internal Review Board and approved by the local Human Research Ethics Committee. All volunteers signed an informed consent.

Collection of diagnostic blood specimens

The collection of all diagnostic blood specimens was performed from 8.00 to 9.00 AM during one week by a single, expert phlebotomist, according to the recommendations of the Clinical Laboratory Standard Institute (CLSI) (9). All volunteers, after 12-hours fasting, were maintained seated for 15 minutes prior to phlebotomy in order to eliminate possible interferences of blood distribution due to the posture (10). After this time interval, a vein was located on forearm by a subcutaneous tissue transilluminator device (Venoscópio IV plus, Duan do Brasil, Sao Paulo, Brazil) to prevent interference from venous stasis (11-13), and 23.9 mL of blood was collected by venipuncture with a 20 G straight needle (Terumo Europe NV, Leuven, Belgium) directly into five serum vacuum tubes with clot activator and gel separator of different brands, as follows:

- Tube I: VACUETTE® 4.0 mL (lot C080818, Greiner Bio-one GmbH, Kremsmünster, Austria);
- Tube II: LABOR IMPORT® 6.0 mL (lot C38005-6, Guangzhou Improve Medical Instruments Co. Ltda, Zhejiang, China);
- Tube III: S-Monovette® 4.9 mL (lot 8092506, Sarstedt, Nümbrecht, Germany);

- Tube IV: SST® 4.0 mL (lot 8308434, Becton, Dickinson and Company Franklin Lakes, NJ, USA); and
- Tube V: SST II Advance® 5.0 mL (lot 8225174, Becton, Dickinson and Company Franklin Lakes, NJ, USA).

To eliminate any potential interference due to either the contact phase or the tissue factor, ~2 mL of blood were preliminarily collected in a discard tube without additive (Vacuette® lot A101004D, Greiner Bio-One GmbH, Kremsmünster, Austria). The exact composition of the gel separator and amount of clot activator inside the vacuum tubes was not communicated by manufactures, as patented. Blood collection was accurately standardized, including the use of needles and vacuum tubes of the same lot.

Processing of diagnostic blood specimens

All the sample tubes were left in upright position for 45 min at room temperature (20 °C) to allow complete blood clotting before centrifugation (14). After centrifugation at 1500 x g for 10 min at room temperature (according to the instructions of the manufacturers), serum was separated, stored in aliquots and kept frozen at -70 °C until measurement. All samples did not show any sign of haemolysis by visual inspection. No specimen was discarded due to unsatisfactory attempts, difficulty in locating venous access, missing veins, manifest haemolysis or lipaemia.

Laboratory testing

All serum aliquots were thawed at the same time. The routine clinical biochemistry tests were performed in duplicate immediately after thawing on the same instrument cobas® 6000 <c501> module (Roche Diagnostics GmbH, Penzberg, Germany), according to the manufacturer's specifications and using proprietary reagents. The panel of tests included the following: glucose (GLU), total cholesterol (COL), high density lipoprotein-cholesterol (HDL), triglycerides (TG), total protein (TP), albumin (ALB), blood urea nitrogen (UREA), creatinine (CRE), uric acid (AU), alkaline phosphatase (ALP), amylase (AMYL), aspartate aminotransferase (AST), alanine aminotransferase (ALT), gamma-glutamyltransferase (GGT), lactate dehydrogenase (LDH), creatine kinase (CK), total bilirubin (BT), direct bilirubin (BD),

phosphate (P), calcium (CA), magnesium (MG), iron (FE), sodium (NA) and potassium (K). The instrument was calibrated against appropriate proprietary reference standard material and verified with

the use of proprietary quality controls. Our evaluation of the within-run precision by internal quality control on cobas® 6000 <c501> module showed low coefficients of variation (Table 1).

TABLE 1. Variability in routine clinical biochemistry testing from five different brands of serum vacuum tubes with clot activator and gel separator.

Tests	Desirable Bias (%)	CVa	Mean % difference (P value)									
			Tube I vs.				Tube II vs.			Tube III vs.		Tube IV vs.
			Tube II	Tube III	Tube IV	Tube V	Tube III	Tube IV	Tube V	Tube IV	Tube V	Tube V
CRE*	3.8	2.5	0.0 (0.234)	6.6 (0.038)	-2.7 (0.001)	-1.3 (0.032)	6.6 (0.019)	-2.7 (0.001)	-1.3 (0.022)	-9.9 (0.001)	-8.4 (0.013)	1.3 (0.008)
AMYL**	7.4	0.8	-6.6 (0.001)	7.8 (0.001)	2.1 (0.003)	5.1 (0.001)	13.5 (0.001)	8.2 (0.001)	11.0 (0.001)	-6.2 (0.001)	-2.9 (0.002)	3.1 (0.001)
P**	3.2	3.0	0.8 (0.107)	0.0 (0.104)	-0.8 (0.876)	-0.8 (0.856)	-0.8 (0.002)	-1.6 (0.001)	-1.6 (0.001)	-0.8 (0.429)	-0.8 (0.667)	0.0 (0.331)
MG**	1.8	1.2	4.9 (0.001)	4.9 (0.001)	4.9 (0.001)	4.9 (0.001)	0.0 (0.666)	0.0 (0.666)	0.0 (0.666)	0.0 (1.000)	0.0 (1.000)	0.0 (1.000)
ALT*	12.0	1.3	4.1 (0.887)	10.2 (0.981)	-2.0 (0.841)	4.1 (0.173)	6.4 (0.353)	-6.4 (0.386)	0.0 (0.130)	-13.6 (0.043)	-6.8 (0.182)	6.0 (0.113)

* Non-normal distribution; P value represents the significance by Wilcoxon ranked-pairs test.

**Normal distribution; P value represents the significance by paired Student's t-test.

The bold P values are statistically significant (P < 0.005) and bold mean % differences represent clinically significant variations, when compared with desirable bias (15).

Tube I: VACUETTE® 4.0 mL (lot C080818, Greiner bio-one, Kremstünster, Austria); Tube II: LABOR IMPORT® 6.0 mL (lot C38005-6, Guangzhou Improve Medical Instruments Co. Ltda, Zhejiang, China); Tube III: S-Monovette® 4.9 mL (lot 8092506, Sarstedt, Nümbrecht, Germany); Tube IV: SST® 4.0 mL (lot 8308434, Becton, Dickinson and Company, Franklin Lakes, NJ, USA); Tube V: SST II Advance® 5.0 mL (lot 8225174, Becton, Dickinson and Company, Franklin Lakes, NJ, USA).

CVa (%): Analytical coefficient (within-run precision), by internal quality control on cobas® 6000 <c501> module.

CRE - creatinine; AMYL - amylase; P - phosphate; MG - magnesium; ALT - alanine aminotransferase.

Statistical analysis

The significance of the differences between samples was assessed by repeated measures using RM ANOVA and paired Student's t-test after checking for normality (with D'Agostino-Pearson's omnibus test). As non-normal distribution was found for GLU, CRE, AU, ALT, GGT, CK, BT, FE and NA results were assessed by Friedman test and Wilcoxon ranked-pairs test using licensed statistical software (GraphPad Prism® version 5.01, La Jolla, CA, USA). Based on screening five different vacuum tubes in parallel, a P value < 0.005 was considered statistically significant according to Bonferroni correction for multiple comparisons. Finally, the biases from Tube I, Tube II, Tube III, Tube IV and Tube V were compared with the current desirable quality specifications for bias (B), derived from biological varia-

tion according to the formula $B < 0.25 (CV_w^2 + CV_g^2)^{1/2}$ where CV_w and CV_g are within- and between-subject CVs (15).

Results

The main results of this study are shown in tables 1 and 2. Basically, significantly differences could be recorded for the following: CRE when comparing Tube I vs. Tube IV, Tube II vs. Tube IV and Tube III vs. Tube IV; AMYL when comparing Tube I vs. Tube II, Tube I vs. Tube III, Tube I vs. Tube IV, Tube I vs. Tube V, Tube II vs. Tube III, Tube II vs. Tube IV, Tube II vs. Tube V, Tube III vs. Tube IV, Tube III vs. Tube V and Tube IV vs. Tube V; P when comparing Tube II vs. Tube III, Tube II vs. Tube IV and Tube II vs. Tube V;

MG when comparing Tube I vs. Tube II, Tube I vs. Tube III, Tube I vs. Tube IV and Tube I vs. Tube V. No significant differences ($P > 0.005$) were observed for: GLU, COL, HDL, TG, TP, ALB, UREA, AU, ALP, AST, ALT, GGT, LDH, CK, BT, BD, CA, FE, NA and K. Clinically significant variations as compared with the current desirable quality specifications (15) were found for: CRE when comparing Tube I vs. Tube III (without statistical significance, $P = 0.038$), Tube II vs. Tube III (without statistical significance, $P = 0.019$), and Tube III vs. Tube IV and Tube III vs. Tube V (without statistical significance, $P = 0.013$); AMYL when comparing Tube I vs. Tube III, Tube II vs. Tube III, Tube II vs. Tube IV, and Tube II vs. Tube V; ALT only when comparing Tube III vs. Tube IV (without statistical significance, $P = 0.043$); MG when comparing Tube I vs. Tube II, Tube I vs. Tube III, Tube I vs. Tube IV and Tube I vs. Tube V.

Discussion

The validation process is essential in accredited clinical laboratories (8,16). Recently article published by Stankovic *et al.* (7) showed that IVD companies that are concerned about total quality in laboratory diagnostics have dedicated resources to assure total system performance. These individuals collaborate with colleagues from instrument/assay/tube companies to address particular customer complaints and find root cause and ways to mitigate it (7). In opposite way Bowen *et al.* showed that components from blood collection tube e.g. surfactants, stopper, stopper lubricant, separator gel and clot activator interact with blood affecting laboratory tests (17). Our results showed that both Stankovic *et al.* and Bowen *et al.* are partially correct. Basically, our validation will permit the laboratory or hospital managers to select the brand's vacuum tubes validated according him/her technical or economical reasons, in order to perform the following laboratory tests: GLU, COL, TG, TP, ALB, UREA, AU, ALP, AST, CK, BT, BD, CA, FE, NA and K. On the contrary special attention will be required in the following situations: a) if the laboratory performs CRE determination and the quality laboratory manager intends to change the serum Tube IV to Tube- I, II or III; b) if the laboratory performs AMYL determination and the quality laboratory

manager intends to change any of the tested brands of serum vacuum tubes; c) if the laboratory performs P determination and the quality laboratory manager intends to change the serum Tube II to Tube- III, IV or V; and d) if the laboratory already performs MG determination and the quality laboratory manager intends to change the serum Tube I to Tube- II, III, IV or V (Table 1). When looking at the above CRE, AMYL, P and MG results (Table 2) one might regard them as clinically irrelevant, though such a conclusion is far from correct in respect of the current quality specifications for bias, derived from biological variation (15). Obviously the quality specifications derived from biological variation (15) are considered both very important and useful in the daily practice by the quality managers of the medical laboratories (18-21). Even in this case caring physicians unaware of the real patient situation might abstain from appropriate treatments as a consequence of change in serum vacuum tubes brands. The National Kidney Foundation published the Kidney Disease Outcomes Quality Initiative (K/DOQI) Clinical Practice Guidelines for chronic kidney disease years ago (22). These guidelines recommend the use of estimated glomerular filtration rate (eGFR) equations to estimate a patient's renal function. At the present time several articles have been published about, evidencing the impact of these guidelines in clinical practices daily (23-25). When considering that accurate serum creatinine determination is necessary for deriving the above eGRF equations, changes of serum vacuum tubes principally from Tube III to Tube IV (with clinically significant difference) can induce diagnostic errors regarding patient real conditions. Acute pancreatitis is an acute inflammatory condition of the pancreas, which might extend to local and distant extra pancreatic tissues. Diagnosis of acute pancreatitis is substantially based on a combination of clinical signs and symptoms, imaging techniques and laboratory investigations (26). A host of serum enzymes such as amylase, lipase, trypsinogen, elastase, phospholipase A₂, ribonuclease, etc are available to diagnose acute pancreatitis and/or to assess the severity, but elevated amylase levels continue to be the "gold standard" among the serum markers (27). Our results show that the changes in serum vacuum

TABLE 2. Comprehensive results of routine clinical biochemistry testing from five different brands of serum vacuum tubes with clot activator and gel separator.

Test	Tube I	Tube II	Tube III	Tube IV	Tube V	P value
GLU (mmol/L)*	4.63 (4.45 - 4.76)	4.61 (4.41 - 4.76)	4.61 (4.43 - 4.72)	4.61 (4.40 - 4.76)	4.58 (4.44 - 4.74)	0.178
COL (mmol/L)**	4.64 ± 0.20	4.66 ± 0.20	4.61 ± 0.20	4.64 ± 0.20	4.66 ± 0.20	0.078
HDL (mmol/L)**	1.37 ± 0.08	1.37 ± 0.09	1.35 ± 0.08	1.35 ± 0.09	1.37 ± 0.09	0.041
TG (mmol/L)**	1.39 ± 0.16	1.38 ± 0.16	1.39 ± 0.16	1.39 ± 0.15	1.39 ± 0.15	0.286
TP (g/L)**	73.0 ± 1.0	73.0 ± 1.0	73.0 ± 1.0	73.0 ± 1.0	73.0 ± 1.0	0.432
ALB (g/L)**	42.0 ± 1.0	42.0 ± 1.0	42.0 ± 1.0	42.0 ± 1.0	42.0 ± 1.0	0.155
UREA (mmol/L)**	11.0 ± 0.8	10.9 ± 0.7	10.9 ± 0.7	10.9 ± 0.7	10.9 ± 0.7	0.450
CRE (μmol/L)*	67.2 (57.5 - 76.0)	67.2 (56.6 - 76.0)	62.8 (53.9 - 76.0)	69.0 (60.1 - 78.7)	68.1 (60.1 - 76.9)	< 0.001
AU (μmol/L)*	273.6 (220.1 - 315.2)	273.6 (220.1 - 315.2)	4.5 (220.1 - 315.2)	273.6 (220.1 - 315.2)	273.6 (220.1 - 321.2)	0.736
ALP (U/L)**	70.0 ± 3.8	69.8 ± 3.6	70.4 ± 3.8	70.6 ± 3.8	70.1 ± 3.8	0.322
AMYL (U/L)**	66.6 ± 5.6	71.0 ± 5.9	61.4 ± 5.2	65.2 ± 5.4	63.2 ± 5.4	< 0.001
AST (U/L)**	23.2 ± 1.2	23.3 ± 1.3	23.6 ± 1.3	23.7 ± 1.3	23.6 ± 1.3	0.260
ALT (U/L)*	24.5 (17.2 - 35.0)	23.5 (18.5 - 34.0)	22.0 (17.5 - 36.5)	25.0 (16.8 - 32.8)	23.5 (16.8 - 35.5)	0.026
GGT (U/L)*	23.0 (17.0 - 35.0)	22.5 (16.2 - 34.0)	23.0 (17.0 - 34.2)	24.0 (16.2 - 34.0)	23.0 (17.0 - 34.2)	0.003
LDH (U/L)**	407 ± 12.4	404 ± 13.8	416 ± 12.2	412 ± 11.4	414 ± 11.4	0.039
CK (U/L)*	71 (50 - 142)	71 (50 - 140)	70 (53 - 142)	70 (53 - 144)	72 (54 - 148)	0.051
BT (μmol/L)*	8.55 (6.33 - 10.09)	8.21 (6.33 - 9.92)	8.55 (5.98 - 9.40)	8.38 (6.50 - 9.58)	8.21 (5.64 - 9.58)	0.216
BD (μmol/L)**	2.22 ± 0.34	2.39 ± 0.17	2.05 ± 0.34	2.22 ± 0.34	2.05 ± 0.34	0.417
P (mmol/L)**	1.24 ± 0.04	1.23 ± 0.04	1.24 ± 0.04	1.25 ± 0.03	1.25 ± 0.04	0.044
CA (mmol/L)**	2.36 ± 0.02	2.34 ± 0.02	2.34 ± 0.02	2.34 ± 0.02	2.34 ± 0.02	0.119
MG (mmol/L)**	0.82 ± 0.02	0.78 ± 0.02	0.78 ± 0.02	0.78 ± 0.02	0.78 ± 0.02	< 0.001
FE (μmol/L)*	18.0 (15.5 - 19.9)	18.0 (15.2 - 19.3)	18.0 (15.5 - 20.0)	18.3 (15.4 - 19.8)	18.0 (15.3 - 19.6)	0.293
NA (mmol/L)*	138.0 (138.0 - 139.0)	139.0 (138.0 - 139.0)	139.0 (138.0 - 139.8)	139.0 (138.0 - 139.0)	139.0 (138.0 - 139.8)	0.265
K (mmol/L)**	4.30 ± 0.06	4.24 ± 0.06	4.24 ± 0.05	4.27 ± 0.06	4.27 ± 0.05	0.095

*Non-normal distribution; the values are presented as median (interquartile range); P value represents the significance by Friedman test

**Normal distribution; the values are presented as mean ± standard deviation; P value represents the significance by RM ANOVA.

The bold P values are statistically significant (P < 0.005). Tube I: VACUETTE® 4.0 mL (lot C080818, Greiner bio-one, Kremsmünster, Austria); Tube II: LABOR IMPORT® 6.0 mL (lot C38005-6, Guangzhou Improve Medical Instruments Co. Ltda, Zhejiang, China); Tube III: S-Monovette® 4.9 mL (lot 8092506, Sarstedt, Nümbrecht, Germany); Tube IV: SST® 4.0 mL (lot 8308434, Becton, Dickinson and Company, Franklin Lakes, NJ, USA); Tube V: SST II Advance® 5.0 mL (lot 8225174, Becton, Dickinson and Company, Franklin Lakes, NJ, USA).

GLU - glucose; COL - total cholesterol; HDL - high density lipoprotein-cholesterol; TG - triglycerides; TP - total protein; ALB - albumin; UREA - blood urea nitrogen; CRE - creatinine; AU - uric acid; ALP - alkaline phosphatase; AMYL - amylase; AST - aspartate aminotransferase; ALT - alanine aminotransferase; GGT - gamma-glutamyltransferase; LDH - lactate dehydrogenase; CK - creatine kinase; BT - total bilirubin; BD - direct bilirubin; P - phosphate; CA - calcium; MG - magnesium; FE - iron; NA - sodium; K - potassium.

tubes brands principally from Tube II to Tube- III or V can manifest clinically significant impact in medical decisions based on laboratory diagnosis. As for phosphate, the vacuum tubes change from Tube II to Tube- III, IV or V can significantly influence phosphates levels; in this respect we must consider that: a) Ferrari *et al.* (28) showed that serum phosphate is an important determinant for correcting

serum calcium in end-stage kidney disease, b) this retains even if the target concentrations for phosphorus and calcium × phosphorus product are sometimes close to the normal range even in patients with end-stage kidney disease (28), c) a relationship between serum phosphate and cardiovascular risk factors was demonstrated by Lippi *et al.* (29). The fourth most abundant cation in the

human body, magnesium is, like potassium, predominantly intracellular. It is critically involved in energy metabolism, enzyme functions and participates in the regulation of PTH synthesis, release, and action (30). Low magnesium levels have been associated with impairment of myocardial contractility, intradialytic hemodynamic instability, and hypotension. In addition, low MG has been also linked to carotid intima-media thickness, a marker of atherosclerotic vascular disease and a predictor of vascular events (31). Magnesium sulphate is also acknowledged as the preferred anticonvulsant for eclamptic women, since it reduces the risk ratio of recurrence of seizures, probably reduces the risk of maternal death, and improves outcome for the children (32). Even for MG, inappropriately high values due to changes in vacuum tubes brands (from Tube I to Tube- II, III, IV or V) might induce diagnostic errors regarding patient real conditions. The 5 kinds of serum vacuum tubes evaluated were validated for 83.3% of the tests but obviously

16.7% of the routine clinical chemistry testing needs more attention when the lab intends to change the serum vacuum tubes brands. In the same way Lima-Oliveira *et al.* have shown recently that different manufacturing source of syringes is a new source of extra analytical variability in blood gas analyses (33). Similarly, as the concentration of clot activator additives as well as gel composition in the serum tubes are patented, the implicit "industrial secret" does not aid to clarify the causes of the bias observed in our study though it is strongly hypothesized, basing on Bowen *et al.* (17) arguments. Future investigations should be planned to better understand the nature of this intriguing observations; in the meantime we suggest that every laboratory management should both standardize the procedures and frequently evaluate the quality of IVD devices.

Potential conflict of interest

None declared.

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Upravljanje kvalitetom prijeanalitičke faze: validacija vakumskih serumskih epruveta kod rutinskih biokemijskih pretraga

Sažetak

Uvod: Proces validacije ključan je u akreditiranim kliničkim laboratorijima. Cilj ovog istraživanja bio je validirati pet vrsta vakumskih serumskih epruveta koje se koriste pri rutinskim biokemijskim pretragama u laboratoriju.

Materijali i metode: Iskusni laboratorijski tehničar sakupio je uzorke krvi od 100 dobrovoljaca u pet različitim vakumskih serumskih epruveta (epruveta I: VACUETTE®, epruveta II: LABOR IMPORT®, epruveta III: S-Monovette®, epruveta IV: SST® i epruveta V: SST II®). U uzorcima su provedene rutinske biokemijske pretrage na analizatoru cobas® 6000 modul c501. Statistička značajnost razlika između rezultata svakog uzorka procijenjena je Studentovim t-testom nakon provjere normalnosti raspodjele. Razina statističke značajnosti postavljena je na $P < 0,005$. Naposljetku su sustavne pogreške dobivene rezultatima iz epruvete I, epruvete II, epruvete III, epruvete IV i epruvete V uspoređene s preporučenim vrijednostima odstupanja (engl. *desirable quality specifications for bias*, B) koja su izvedena iz biološke varijacije.

Rezultati i zaključak: Naša će validacija omogućiti voditeljima laboratorija ili bolnice odabrati vakumske epruvete određenog proizvođača validirane prema vlastitim tehničkim ili ekonomskim zahtjevima, za sljedeće pretrage: glukozu, ukupni kolesterol, HDL-kolesterol, trigliceride, ukupne proteine, albumin, ureju, mokraćnu kiselinu, alkalnu fosfatazu, aspartat-aminotransferazu, gama-glutamilttransferazu, laktat-dehidrogenazu, kreatinin-kinazu, ukupni bilirubin, direktni bilirubin, kalcij, željezo, natrij i kalij. Posebna se pažnja treba obratiti ukoliko laboratorij već vrši pretrage za određivanje koncentracije kreatinina, fosfora i magnezija te aktivnosti amilaze, ukoliko voditelj upravljanja kvalitetom laboratorija želi promijeniti serumske epruvete. Predlažemo da voditelj laboratorija standardizira postupke i redovito procjenjuje kvalitetu *in vitro* dijagnostičkih uređaja.

Ključne riječi: prijeanalitička varijabilnost; uzorkovanje; uzorci seruma; vakumske epruvete; proces validacije