

 Open access • Journal Article • DOI:10.1097/MOG.000000000000151

Prebiotics, probiotics, synbiotics, and the immune system: experimental data and clinical evidence. — Source link

Remo Frei, Mübeccel Akdis, Liam O'Mahony, Liam O'Mahony

Institutions: University of Zurich, Swiss Institute of Allergy and Asthma Research

Published on: 15 Jan 2015 - Current Opinion in Gastroenterology (Curr Opin Gastroenterol)

Topics: Synbiotics and Immune system

Related papers:

- [Dietary Modulation of the Human Colonic Microbiota: Introducing the Concept of Prebiotics](#)
- [Expert consensus document. The International Scientific Association for Probiotics and Prebiotics consensus statement on the scope and appropriate use of the term probiotic](#)
- [Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis](#)
- [CURRENT OPINION Prebiotics, probiotics, synbiotics, and the immune system: experimental data and clinical evidence](#)
- [Probiotics, prebiotics and synbiotics- a review](#)

Share this paper:    

View more about this paper here: <https://typeset.io/papers/prebiotics-probiotics-synbiotics-and-the-immune-system-1iiqxvo9dd>



**University of
Zurich**^{UZH}

**Zurich Open Repository and
Archive**

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2015

Prebiotics, probiotics, synbiotics, and the immune system: experimental data and clinical evidence

Frei, Remo ; Akdis, Mübeccel ; O'Mahony, Liam

Abstract: **PURPOSE OF REVIEW** The intestinal immune system is constantly exposed to foreign antigens, which for the most part should be tolerated. Certain probiotics, prebiotics, and synbiotics are able to influence immune responses. In this review, we highlight the recent publications (within the last 2 years) that have substantially progressed this field. **RECENT FINDINGS** The immunological mechanisms underpinning probiotics, prebiotics, and synbiotics effects continue to be better defined with novel mechanisms being described for dendritic cells, epithelial cells, T regulatory cells, effector lymphocytes, natural killer T cells, and B cells. Many of the mechanisms being described are bacterial strain or metabolite specific, and should not be extrapolated to other probiotics or prebiotics. In addition, the timing of intervention seems to be important, with potentially the greatest effects being observed early in life. **SUMMARY** In this review, we discuss the recent findings relating to probiotics, prebiotics, and synbiotics, specifically their effects on immunological functions.

DOI: <https://doi.org/10.1097/MOG.000000000000151>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-107856>

Journal Article

Published Version

Originally published at:

Frei, Remo; Akdis, Mübeccel; O'Mahony, Liam (2015). Prebiotics, probiotics, synbiotics, and the immune system: experimental data and clinical evidence. *Current Opinion in Gastroenterology*, 31(2):153-158.

DOI: <https://doi.org/10.1097/MOG.000000000000151>



Prebiotics, probiotics, synbiotics, and the immune system: experimental data and clinical evidence

Remo Frei^{a,b}, Mübeccel Akdis^a, and Liam O'Mahony^a

Purpose of review

The intestinal immune system is constantly exposed to foreign antigens, which for the most part should be tolerated. Certain probiotics, prebiotics, and synbiotics are able to influence immune responses. In this review, we highlight the recent publications (within the last 2 years) that have substantially progressed this field.

Recent findings

The immunological mechanisms underpinning probiotics, prebiotics, and synbiotics effects continue to be better defined with novel mechanisms being described for dendritic cells, epithelial cells, T regulatory cells, effector lymphocytes, natural killer T cells, and B cells. Many of the mechanisms being described are bacterial strain or metabolite specific, and should not be extrapolated to other probiotics or prebiotics. In addition, the timing of intervention seems to be important, with potentially the greatest effects being observed early in life.

Summary

In this review, we discuss the recent findings relating to probiotics, prebiotics, and synbiotics, specifically their effects on immunological functions.

Keywords

adaptive immune system, innate immune system, prebiotics, probiotics, synbiotics

INTRODUCTION

The mammalian gastrointestinal tract is a highly evolved system specialized to perform the essential functions of nutrient digestion, absorption, and waste disposal. The intestinal immune system has the unenviable task of maintaining intestinal integrity in the presence of vast quantities of external or foreign antigens. Sophisticated cellular and molecular networks need to be constantly coordinated in order to tolerate nonpathogenic antigens, while also protective immune responses to potential pathogens must be maintained and can be induced effectively on demand. Inappropriate immune response to bacterial or dietary antigens is a significant component in several intestinal diseases, including inflammatory bowel disease, irritable bowel syndrome, and food allergies [1,2].

The balance between immune tolerance and inflammation is regulated in part by the crosstalk between innate and adaptive immune cells and the intestinal microbiota. Disrupted communication between the microbiome and the host because of altered composition and metabolism is thought to negatively influence the intestinal immune

homeostatic networks. The deliberate modification of microbial species and their metabolism has led to the probiotic and prebiotic concepts [3].

Probiotics can be defined as live microorganisms which, when administered in adequate amounts, confer a health benefit on the host. Notably, the definition of a probiotic does not differentiate between the wide range of potential health benefits and it is clear that not all probiotics will influence the immune system in the same way. In this review, we have focused on the probiotic studies with immunological endpoints, but these findings cannot be extrapolated to other probiotic strains. In addition, we have included microbial species not currently used in the industry as probiotics, even

^aSwiss Institute of Allergy and Asthma Research, University of Zurich and
^bChristine Kühne – Center for Allergy Research and Education (CK-CARE), Davos, Switzerland

Correspondence to Dr Liam O'Mahony, SIAF, Obere Strasse 22, 7270 Davos Platz, Switzerland. Tel: +41 81 4100853; fax: +41 81 4100840; e-mail: liam.omahony@siaf.uzh.ch

Curr Opin Gastroenterol 2015, 31:153–158

DOI:10.1097/MOG.0000000000000151

KEY POINTS

- Specific probiotics, prebiotics, or their combination significantly influence host immunological networks.
- The immune responses to microbial components (e.g. polysaccharides) and their metabolites (e.g. SCFA) are being better characterized at a molecular level.
- The immune-modulatory effects of probiotics, prebiotics, or synbiotics may be most potent when administered early in life.

though these microbes fulfill the definition provided above.

Prebiotics can be defined as selectively fermented ingredients that allow specific changes, both in the composition and in the activity of the gastrointestinal microflora, which confer benefits upon host well being and health. Prebiotics typically are fibers that cannot be digested by the host, but are metabolized by the colonic microbiome resulting in expansion of certain bacterial species and the release of metabolites, such as short-chain fatty acids (SCFAs). As with the probiotic definition, not all prebiotics will have the same effect on immunological functions.

The combination of probiotics and prebiotics is termed 'synbiotics'.

INNATE IMMUNE SYSTEM

The innate immune system is comprised of many different cell types, and these cells are often the first cells to come in contact with intestinal microbes and their metabolic products. The most commonly described cell types in the probiotics and prebiotics literature are dendritic cells and epithelial cells.

Dendritic cells

Intestinal dendritic cells are located within specific intestinal lymphoid tissues, collectively termed gut-associated lymphoid tissues (GALT), or diffusely distributed throughout the intestinal lamina propria [4]. Dendritic cells are the primary cell type involved as 'sensors' of microbial ligands through activation of innate immune receptors (e.g. Toll-like receptors and c-type lectin receptors). The signaling pathways triggered by bacterial-derived molecules allow for changes in dendritic cell phenotypes and cytokine secretion, which underlie the integration of microbial and host metabolism with immune functions. Metabolism of vitamin A to retinoic acid is a key immunomodulatory activity associated with

intestinal dendritic cells [5[■]]. Certain, but not all, probiotic microbes can induce retinoic acid metabolism by human dendritic cells *in vitro* and by murine CD103⁺ dendritic cells within the small intestine lamina propria [6[■],7]. In addition to vitamin A metabolism, induction of another dendritic cell metabolic enzyme, heme oxygenase-1 (HO-1), was shown to be required for the induction of mucosal T regulatory (T_{REG}) cells within the mesenteric lymph nodes by *Lactobacillus rhamnosus* [8].

Bacterial cell wall components and metabolites have been associated with the immunoregulatory effects on dendritic cells. For example, major histocompatibility complex (MHC)-II-dependent presentation of segmented filamentous bacteria antigens by intestinal CD11c⁺ dendritic cells is crucial for the local induction of T_H17 lymphocytes [9[■]]. In addition, capsular polysaccharide A (PSA) from *Bacteroides fragilis* has been shown to interact directly with mouse plasmacytoid dendritic cells via TLR-2. PSA-exposed plasmacytoid dendritic cells express molecules involved in protection against colitis and stimulated CD4⁺ cells to secrete IL-10 [10[■]]. An exopolysaccharide from *Bacillus subtilis* prevents gut inflammation stimulated by *Citrobacter rodentium*, which is dependent on TLR-4 and MyD88 signaling [11[■]].

The production of SCFAs occurs in the colon following fermentation of dietary fibers, such as prebiotics [12]. Abnormalities in the production of these metabolites (because of dietary factors and dysbiosis) might play a role in the pathogenesis of type 2 diabetes, obesity, inflammatory bowel disease, colorectal cancer, and allergies [13]. Among the SCFAs, butyrate seems to be more potent than acetate or propionate in inducing immunomodulatory effects. Butyrate influences the activity of histone deacetylases (HDAC), which is responsible for decreasing dendritic cell IL-12 and IL-6 cytokine secretion and allows dendritic cells to promote T_{REG} cells. Propionate can also contribute to the induction of Foxp3 expression by dendritic cells, whereas acetate does not have this activity possibly because of the lack of HDAC activity [14[■]]. Butyrate also inhibits intestinal macrophage HDAC [15[■]]. Another recent study has confirmed and extended the observation that butyrate promotes dendritic cell regulatory activity, resulting in the induction of T_{REG} cells and IL-10-secreting T cells. These effects were mediated by the G-protein-coupled receptor Gpr109a on colonic dendritic cells and macrophages [16[■]]. In contrast, butyrate has also been shown to promote IL-23 secretion by murine dendritic cells, which may promote T_H17 responses under certain circumstances [17].

Histamine is another important mucosal metabolite secreted by the gut microbes, and mucosal histamine levels are increased in patients with irritable bowel syndrome and inflammatory bowel disease [18[¶]]. Histamine is able to decrease chemokine and proinflammatory cytokine secretion induced by the Toll-like-receptor-stimulated dendritic cells, while increasing IL-10 production [19[¶]]. Histamine exerted this effect by activating the histamine 2 receptor (H₂R) on dendritic cells and the signaling mechanism required cyclic adenosine monophosphate (cAMP) and exchange protein directly activated by cAMP (EPAC). Administration of a histamine-secreting *Lactobacillus* strain to mice resulted in rapid weight loss and enhanced Peyer's patch cytokine secretion, which was exaggerated in H₂R-deficient animals [20^{¶¶}].

Epithelial cells

Epithelial cells play an essential role in nutrient absorption. Pathogen-induced reductions in epithelial cell digestive enzyme activity can be blocked by *Bifidobacterium infantis* 35624, possibly via modulation of mucosal inflammatory responses [21,22[¶]]. In addition to their absorptive function, epithelial cells form a mucosal barrier that protects host tissue from damaging agents such as luminal pathogens and toxic products. One protective barrier mechanism is the production and secretion of antimicrobial peptides, such as defensins and cathelicidins. Probiotic strains have been shown to differentially regulate defensin expression and protein secretion, which is influenced by local inflammatory mediators [23]. Autophagy is an important adaptive response to stress, which promotes cell survival and is required for the maintenance of the epithelial barrier. A number of *Bifidobacteria* have been recently described that promote autophagy in an intestinal cell line [24]. The mucus layer coating the gastrointestinal tract is an important barrier component and probiotics have been shown to promote mucin production by goblet cells in the intestine. Recently, p40 from *Lactobacillus* GG was demonstrated to be sufficient for the stimulation of mucin production through transactivation of the epidermal growth factor receptor [25[¶]]. Excessive epithelial cell responses to microbial ligands result in local inflammatory responses, which disrupt the epithelial barrier. A wide range of probiotic microbes have been demonstrated to suppress epithelial cell proinflammatory chemokine responses [26[¶],27,28]. However, not all chemokine responses are impacted to the same extent by every probiotic strain and a single probiotic strain may reduce the expression of certain chemokines, while increasing the expression

of others. For example, *Bifidobacterium bifidum* PRL2010 suppresses CCL22 expression but enhances CCL19 expression, suggesting that strain-specific and chemokine-specific responses are induced by probiotics [29[¶]]. In addition, prebiotics themselves, or SCFAs, can also modulate epithelial barrier function, production of antimicrobial peptides, and secretion of proinflammatory mediators [30,31].

ADAPTIVE IMMUNE SYSTEM

The adaptive immune system receives polarizing signals from the innate cells to expand an appropriately controlled lymphocyte response to bacterial and metabolic factors. Recent evidence has suggested a role for probiotic and prebiotic effects on T_{REG} cells, effector T cells, natural killer T (NKT) cells, and B cells.

T lymphocytes

The beneficial effect of prebiotics, probiotics, and synbiotics against diseases such as allergy or colitis is often associated with enhancement of T_{REG} cells [32[¶],33]. Specific probiotics, but not all, have been shown to induce an increase in T_{REG} cells. Notably, consumption of *B. infantis* 35624 by healthy human volunteers resulted in an increased proportion of Foxp3⁺ lymphocytes in peripheral blood, whereas administration of this probiotic to psoriasis patients, chronic fatigue syndrome patients, or ulcerative colitis patients consistently resulted in reduced levels of serum proinflammatory biomarkers such as C-reactive protein, possibly mediated by increased numbers of T_{REG} cells [7,34^{¶¶}].

In addition to probiotics, effects on lymphocytes can also be mediated by SCFAs such as acetate, propionate, butyrate, and *n*-butyrate. Oral administration of a mixture of 17 *Clostridia* strains to mice attenuated the severity of colitis and allergic diarrhea in a T_{REG}-TGF- β -dependent mechanism. This process is most likely because of SCFAs produced by the *Clostridia* strains [35,36^{¶¶}]. G-protein-coupled receptor (GPCR) 43 has been described as receptor for SCFAs. GPR43 signaling ameliorates diseases such as colitis, inflammatory arthritis, and allergic airway diseases [35,37[¶]]. In addition to the expression on neutrophils and eosinophils, GPCR43 is expressed on colonic inducible T_{REG} cells and promotes their expansion and IL-10 secretion.

As already described above for dendritic cells, SCFAs inhibit HDAC activity also in lymphocytes enhancing histone H3 acetylation in the promoter and conserved noncoding sequence regions of the *Foxp3* locus [35,38[¶]].

Probiotic and commensal bacteria have been shown to suppress T_H17 responses through both direct and indirect mechanisms. T_H17 cells secrete IL-17 that induces tissue inflammation. Probiotic strains inhibit T_H17 and IL-17 activity by inducing T_{REG} and T_H1 subsets, by induction of IL-27 production, which suppress the generation of IL-17 and induce IL-10 or by stimulation of TLR9 on T_H17 cells [39].

Natural killer T cells

NKT cells are central mediators of intestinal inflammation and pathogenic NKT cell activation is mediated by CD1d⁺ bone-marrow-derived cells, whereas CD1d⁺ epithelial cells protect against intestinal inflammation [40]. It has been shown that NKT cells can be influenced by the gut microflora and certain probiotic lipid antigens may directly activate NKT cells [41,42]. Recently, a healthy human volunteer study showed that the combination of xylo-oligosaccharide with *Bifidobacterium animalis* reduced CD16/CD56 expression on NKT cells and reduced IL-10 secretion from peripheral blood mononuclear cells in response to lipopolysaccharide [43[¶]]. The functional consequences of altered NKT cell activation by the microbiome in humans remain to be determined.

B cells

B lymphocytes have an essential role in humoral immune responses via their secretion of antigen-specific antibodies. In addition, B cells can limit aggressive immune reactivity. B cells regulate immune responses mainly via IL-10, which has been shown in the experimental models of infection, allergic inflammation, and tolerance [44].

B-cell-dependent modulation of the microbiome was shown in IgA-deficient mice. IgA-deficient mice had persistent intestinal colonization with γ -Proteobacteria that cause sustained intestinal inflammation and increased susceptibility to neonatal and adult models of intestinal injury. The group also identified an IgA-dependent mechanism responsible for the maturation of the intestinal microbiota in mice [45]. Recently, another group showed that the number of gut-homing IgG⁺ and IgA⁺ B cells was significantly higher in infants compared with adults. This suggests that activation of naïve B cells in the gut overlaps with the establishment of the gut microbiota in humans [46]. Oral administration of *Lactobacillus gasseri* SBT2055 (LG2055) induced IgA production and increased the number of IgA⁺ cells in Peyer's patches and in the lamina propria. Combined stimulation of B cells

with B-cell activating factor and LG2055 enhanced the induction of IgA production. IgA plays an important role in host defense against mucosally transmitted pathogens, prevents commensal bacteria from binding to epithelial cells, and neutralizes their toxins to maintain homeostasis at the mucosal surfaces [47].

It was recently shown that the gut microbiota induces dendritic cells and macrophages to produce IL-1 β and IL-6, which both drive T_H17 differentiation and arthritis. The same signals also induced the differentiation of IL-10-producing B regulatory cells. These data suggest that the commensal microbiota is important for inducing both proinflammatory and regulatory responses in order to rapidly clear infections and minimize the inflammation-associated tissue damage [48^{¶¶}]. Interestingly, supplementation with xylooligosaccharide and *B. animalis* in a human study led to reduced expression of CD19 on B cells [43[¶]].

AGE-DEPENDENT EFFECTS OF PROBIOTICS, PREBIOTICS, AND SYNBIOTICS

The timing of bacterial colonization early in life is thought to be important for appropriate immune education and the transmission from mother to the fetus during pregnancy and birth is being better described. Cultures of meconium have shown diverse groups of Gram-positive and Gram-negative bacteria, possibly not all derived after delivery. The development of the gut microbiome is a dynamic process and early colonization with *Bacteroides* and *Bifidobacterium* species might play a crucial role in the development of immune regulation [49[¶]]. Factors that can influence early-life colonization include antibiotic treatment, method of delivery, maternal and infant diet, and biodiversity in the home, surrounding environment and in family members.

The immune system at birth is dominated by the T_H2 cells. However, the human fetus has a functional immune system at a relative early status of development comprising not only CD4⁺ and CD8⁺ T cells, but also FOXP3⁺ T_{REG} cells. One concept gaining support is that the developing fetus may become educated by whole bacteria or their genetic material that is provided via maternal serum. DNA from *Bifidobacteria* and *Lactobacilli*, two genera typically used as probiotics, are found in human placenta. In contrast, in-utero exposure to potentially pathogenic bacteria such as *Ureaplasma* species leads to immune dysregulation, commonly ending in fatal complications [50]. Maternal consumption

of probiotic-containing food components may reduce the risk for childhood allergic diseases, and mouse models demonstrate a reduced risk of inflammatory bowel diseases [50]. Epigenetic mechanisms may be critical as application of *Acinetobacter lwoffii* to pregnant mice reduced the airway hypersensitivity response of the offspring. The promoter region of IFN- γ in CD4⁺ T cells of the offspring had high levels of histone-4 acetylation, associated with enhanced transcription, whereas the IL-4 promoter region had lower levels of histone-4 acetylation. Moreover, exposure of pregnant mothers to the farm environment, which has high levels of *A. lwoffii*, was associated with DNA demethylation of the *Foxp3* locus and methylation of the T_H2-associated genes RAD50 and IL-13 [50].

As the gut microbiota composition during the first months of life seems to be important for the development of appropriate immune regulatory networks and thereby influences later life disease risk, intervention with probiotics, prebiotics, or synbiotics might be most effective at this age or even during pregnancy.

CONCLUSION

Appropriately selected probiotics, prebiotics, or their combination exert potent effects on the immune system. Of particular interest are the recent findings on the molecular mechanisms underpinning the effects of SCFAs on immune cells. However, it is highly likely that SCFAs are only one example of the many bacterial metabolites which influence immune cells. A better description of the bacterial strains and metabolites which influence immune function will allow for improved design of future probiotic and prebiotic cocktails for the prevention and treatment of immunological disorders.

Acknowledgements

None.

Financial support and sponsorship

The authors are supported by the Swiss National Foundation grants (project numbers: 310030-127356, 310030-144219, and 320030-140772), Allergiestiftung Ulrich Müller-Gierok, European Union Marie Curie grants, and CK-CARE. M.A. received funding from the European Union research grant numbers 260895 and 261357.

Conflicts of interest

L.O.M. is a consultant to Alimentary Health Ltd. and has received research funding from GSK. R.F. and M.A. have no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Wang ZK, Yang YS, Chen Y, *et al.* Intestinal microbiota pathogenesis and fecal microbiota transplantation for inflammatory bowel disease. *World J Gastroenterol* 2014; 20:14805–14820.
 2. Shanahan F, Quigley EM. Manipulation of the microbiota for treatment of IBS and IBD-challenges and controversies. *Gastroenterology* 2014; 146:1554–1563.
 3. Frei R, Lauener RP, Cramer R, *et al.* Microbiota and dietary interactions: an update to the hygiene hypothesis? *Allergy* 2012; 67:451–461.
 4. Schiavi E, Smolinska S, O'Mahony L. Intestinal dendritic cells. *Curr Opin Gastroenterol* 2015; 31:000–000.
 5. Bakdash G, Vogelpoel LT, van Capel TM, *et al.* Retinoic acid primes human dendritic cells to induce gut-homing, IL-10-producing regulatory T cells. *Mucosal Immunol* 2014. [Epub ahead of print]
- This study shows that retinoic acid induces IL-10-producing T cells and postulates a novel mechanism for IL-10 in maintaining tolerance to the intestinal microbiome.
6. Konieczna P, Ferstl R, Ziegler M, *et al.* Immunomodulation by *Bifidobacterium infantis* 35624 in the murine lamina propria requires retinoic acid-dependent and independent mechanisms. *PLoS One* 2013; 8:e62617.
- This study describes the induction of retinoic acid metabolism in CD103⁺ dendritic cells by a specific probiotic within the gut.
7. Konieczna P, Groeger D, Ziegler M, *et al.* *Bifidobacterium infantis* 35624 administration induces Foxp3 T regulatory cells in human peripheral blood: potential role for myeloid and plasmacytoid dendritic cells. *Gut* 2012; 61:354–366.
 8. Karimi K, Kandiah N, Chau J, *et al.* A *Lactobacillus rhamnosus* strain induces a heme oxygenase dependent increase in Foxp3⁺ regulatory T cells. *PLoS One* 2012; 7:e47556.
 9. Goto Y, Panea C, Nakato G, *et al.* Segmented filamentous bacteria antigens presented by intestinal dendritic cells drive mucosal Th17 cell differentiation. *Immunity* 2014; 40:594–607.
- This report describes the complex role of dendritic cells and innate lymphoid cells in the regulation of intestinal Th17 cell homeostasis in response to a gut microbe.
10. Dasgupta S, Erturk-Hasdemir D, Ochoa-Reparaz J, *et al.* Plasmacytoid dendritic cells mediate anti-inflammatory responses to a gut commensal molecule via both innate and adaptive mechanisms. *Cell Host Microbe* 2014; 15:413–423.
- This study shows that plasmacytoid dendritic cells can orchestrate the beneficial immunoregulatory interaction of commensal microbial molecules via both innate and adaptive immune mechanisms.
11. Jones SE, Paynich ML, Kearns DB, *et al.* Protection from intestinal inflammation by bacterial exopolysaccharides. *J Immunol* 2014; 192:4813–4820.
- This study demonstrates that bacterial exopolysaccharides prevent colitis in a TLR4-dependent manner, which requires myeloid cells.
12. Tan J, McKenzie C, Potamitis M, *et al.* The role of short-chain fatty acids in health and disease. *Adv Immunol* 2014; 121:91–119.
 13. Thorburn AN, Macia L, Mackay CR. Diet, metabolites, and 'western-lifestyle' inflammatory diseases. *Immunity* 2014; 40:833–842.
 14. Arpaia N, Campbell C, Fan X, *et al.* Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 2013; 504:451–455.
- This study shows that butyrate, produced by commensal microorganisms, facilitated extrathymic generation of T regulatory cells.
15. Chang PV, Hao L, Offermanns S, *et al.* The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc Natl Acad Sci USA* 2014; 111:2247–2252.
- These authors elucidate a novel pathway in which the host may maintain tolerance to intestinal microbiota by rendering lamina propria macrophages hyporesponsive.
16. Singh N, Gurav A, Sivaprakasam S, *et al.* Activation of Gpr109a, receptor for niacin and the commensal metabolite butyrate, suppresses colonic inflammation and carcinogenesis. *Immunity* 2014; 40:128–139.
- This study demonstrates that Gpr109a has an essential role in mediating the beneficial effects of gut microbiota and dietary fiber in colon.
17. Berndt BE, Zhang M, Owyang SY, *et al.* Butyrate increases IL-23 production by stimulated dendritic cells. *Am J Physiol Gastrointest Liver Physiol* 2012; 303:1384–1392.
 18. Smolinska S, Jutel M, Cramer R, *et al.* Histamine and gut mucosal immune regulation. *Allergy* 2014; 69:273–281.
- This report comprehensively describes the cellular sources and immunological effects of histamine within the gut.
19. Frei R, Ferstl R, Konieczna P, *et al.* Histamine receptor 2 modifies dendritic cell responses to microbial ligands. *J Allergy Clin Immunol* 2013; 132:194–204.
- This study describes the immunoregulatory molecular mechanisms associated with histamine modulation of dendritic cell activation following TLR stimulation.

20. Ferstl R, Frei R, Schiavi E, *et al.* Histamine receptor 2 is a key influence in immune responses to intestinal histamine-secreting microbes. *J Allergy Clin Immunol* 2014; 134:744–746.
- This report highlights the immunoregulatory role for histamine secreted by gut microbes *in vivo* and the detrimental effects associated with a high histamine-secreting *Lactobacillus* strain.
21. Symonds EL, O'Mahony C, Lapthorne S, *et al.* *Bifidobacterium infantis* 35624 protects against Salmonella-induced reductions in digestive enzyme activity in mice by attenuation of the host inflammatory response. *Clin Transl Gastroenterol* 2012; 3:e15.
22. Scully P, Macsharry J, O'Mahony D, *et al.* *Bifidobacterium infantis* suppression of Peyer's patch MIP-1 α and MIP-1 β secretion during Salmonella infection correlates with increased local CD4⁺CD25⁺ T cell numbers. *Cell Immunol* 2013; 281:134–140.
- This study suggests that multiple mechanisms may underpin the anti-inflammatory effects of this probiotic.
23. Habil N, Abate W, Beal J, *et al.* Heat-killed probiotic bacteria differentially regulate colonic epithelial cell production of human β -defensin-2: dependence on inflammatory cytokines. *Benef Microbes* 2014; 5:483–495.
24. Lin R, Jiang Y, Zhao X, *et al.* Four types of Bifidobacteria trigger autophagy response in intestinal epithelial cells. *J Dig Dis* 2014; 15:597–605.
25. Wang L, Cao H, Liu L, *et al.* Activation of epidermal growth factor receptor mediates mucin production stimulated by p40, a *Lactobacillus rhamnosus* GG-derived protein. *J Biol Chem* 2014; 289:20234–20244.
- This study demonstrates that a single isolated protein from this probiotic strain can enhance mucin production.
26. Boonma P, Spinler JK, Venable SF, *et al.* *Lactobacillus rhamnosus* L34 and *Lactobacillus casei* L39 suppress *Clostridium difficile*-induced IL-8 production by colonic epithelial cells. *BMC Microbiol* 2014; 14:177.
- These results suggest that both probiotic strains can produce factors capable of modulating inflammation stimulated by *C. difficile*.
27. Ren DY, Li C, Qin YQ, *et al.* Lactobacilli reduce chemokine IL-8 production in response to TNF- α and Salmonella challenge of Caco-2 cells. *Biomed Res Int* 2013; 2013:925219.
28. Sibartie S, O'Hara AM, Ryan J, *et al.* Modulation of pathogen-induced CCL20 secretion from HT-29 human intestinal epithelial cells by commensal bacteria. *BMC Immunol* 2009; 10:54.
29. Turroni F, Taverniti V, Ruas-Madiedo P, *et al.* *Bifidobacterium bifidum* PRL2010 modulates the host innate immune response. *Appl Environ Microbiol* 2014; 80:730–740.
- This report presents global transcription profiles of cells exposed to *Bifidobacterium bifidum* PRL2010, demonstrating that this bacterium modulates the innate immune response of the host.
30. Johnson-Henry KC, Pinnell LJ, Waskow AM, *et al.* Short-chain fructooligosaccharide and inulin modulate inflammatory responses and microbial communities in Caco2-bbe cells and in a mouse model of intestinal injury. *J Nutr* 2014; 144:1725–1733.
31. Jiang W, Sunkara LT, Zeng X, *et al.* Differential regulation of human cathelicidin LL-37 by free fatty acids and their analogs. *Peptides* 2013; 50:129–138.
32. Kim HJ, Kim YJ, Lee SH, *et al.* Effects of *Lactobacillus rhamnosus* on allergic march modulated by suppressing Th2, Th17, and TSLP responses via CD4(+)CD25(+)Foxp3(+) Tregs. *Clin Immunol* 2014; 153:178–186.
- Oral application of this probiotic prevented the development of atopic dermatitis and allergic asthma by suppressing Th2, Th17, and TSLP responses via a mechanism that may involve regulatory T cells.
33. Liu Y, Fatheree NY, Mangalat N, *et al.* Human-derived probiotic *Lactobacillus reuteri* strains differentially reduce intestinal inflammation. *Am J Physiol Gastrointest Liver Physiol* 2010; 299:1087–1096.
34. Groeger D, O'Mahony L, Murphy EF, *et al.* *Bifidobacterium infantis* 35624 modulates host inflammatory processes beyond the gut. *Gut Microbes* 2013; 4:325–339.
- These authors demonstrate the ability of a single probiotic to reduce systemic proinflammatory biomarkers in both gastrointestinal and nongastrointestinal conditions in humans.
35. Arpaia N, Rudensky AY. Microbial metabolites control gut inflammatory responses. *Proc Natl Acad Sci USA* 2014; 111:2058–2059.
36. Atarashi K, Tanoue T, Oshima K, *et al.* Treg induction by a rationally selected mixture of Clostridia strains from the human microbiota. *Nature* 2013; 500:232–236.
- Seventeen bacterial strains were shown to act as a community *in vivo* and to provide bacterial antigens, and a TGF- β -rich environment to help expansion and differentiation of regulatory T cells.
37. Trompette A, Gollwitzer ES, Yadava K, *et al.* Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat Med* 2014; 20:159–166.
- This study shows that dietary fermentable fiber and SCFAs can shape the immunological environment in the lung and influence the severity of allergic inflammation.
38. Furusawa Y, Obata Y, Fukuda S, *et al.* Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. *Nature* 2013; 504:446–450.
- These authors showed that butyrate enhanced histone H3 acetylation in the promoter and conserved noncoding sequence regions of the Foxp3 locus, suggesting a novel mechanism for how microbial-derived butyrate regulates the differentiation of regulatory T cells.
39. Tanabe S. The effect of probiotics and gut microbiota on Th17 cells. *Int Rev Immunol* 2013; 32:511–525.
40. Olszak T, Neves JF, Dowds CM, *et al.* Protective mucosal immunity mediated by epithelial CD1d and IL-10. *Nature* 2014; 509:497–502.
41. Olszak T, An D, Zeissig S, *et al.* Microbial exposure during early life has persistent effects on natural killer T cell function. *Science* 2012; 336:489–493.
42. Liang S, Webb T, Li Z. Probiotic antigens stimulate hepatic natural killer T cells. *Immunology* 2014; 141:203–210.
43. Childs CE, R yti  H, Alhoniemi E, *et al.* Xylo-oligosaccharides alone or in synbiotic combination with *Bifidobacterium animalis* subsp. *lactis* induce bifidogenesis and modulate markers of immune function in healthy adults: a double-blind, placebo-controlled, randomised, factorial cross-over study. *Br J Nutr* 2014; 111:1945–1956.
- This study suggests that a synbiotic may confer further health benefits compared with a probiotic or probiotic alone.
44. Stanic B, van de Veen W, Wirz OF, *et al.* IL-10-overexpressing B cells regulate innate and adaptive immune responses. *J Allergy Clin Immunol* 2014. [Epub ahead of print]
45. Mirpuri J, Raetz M, Sturge CR, *et al.* Proteobacteria-specific IgA regulates maturation of the intestinal microbiota. *Gut Microbes* 2014; 5:28–39.
46. Lundell AC, Rabe H, Quiding-J rbrink M, *et al.* Development of gut-homing receptors on circulating B cells during infancy. *Clin Immunol* 2011; 138:97–106.
47. Sakai F, Hosoya T, Ono-Ohmachi A, *et al.* *Lactobacillus gasseri* SBT2055 induces TGF- β expression in dendritic cells and activates TLR2 signal to produce IgA in the small intestine. *PLoS One* 2014; 9:e105370.
48. Rosser EC, Oleinika K, Tonon S, *et al.* Regulatory B cells are induced by gut microbiota-driven interleukin-1 β and interleukin-6 production. *Nat Med* 2014.
- This study shows that inflammatory signals induced by the gut flora result in increased numbers of B regulatory cells, which restrain excessive inflammation.
49. Abrahamsson TR, Wu RY, Jenmalm MC. Gut microbiota and allergy: the importance of the pregnancy period. *Pediatr Res* 2014.
- These authors suggest that a possible reason for the initial exposure of bacterial molecular patterns to the fetus *in utero* is to prime the immune system and the epithelium to respond appropriately to pathogens and commensals after birth.
50. Romano-Keeler J, Weitkamp JH. Maternal influences on fetal microbial colonization and immune development. *Pediatr Res* 2014.