

1 **Predatory zooplankton on the move:**

2 ***Themisto* amphipods in high-latitude marine pelagic food webs**

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16 <https://doi.org/10.1016/bs.amb.2019.02.002>

17 **ABSTRACT**

18 Hyperiid amphipods are predatory pelagic crustaceans that are particularly prevalent in high-latitude
19 oceans. Many species are likely to have co-evolved with soft-bodied zooplankton groups such as salps
20 and medusae, using them as substrate, for food, shelter or reproduction. Compared to other pelagic
21 groups, such as fish, euphausiids and soft-bodied zooplankton, hyperiid amphipods are poorly studied
22 especially in terms of their distribution and ecology. Hyperiids of the genus *Themisto*, comprising seven
23 distinct species, are key players in temperate and cold-water pelagic ecosystems where they reach
24 enormous levels of biomass. In these areas, they are important components of marine food webs, and
25 they are major prey for many commercially important fish and squid stocks. In northern parts of the
26 Southern Ocean, *Themisto* are so prevalent that they are considered to take on the role that Antarctic

27 krill play further south. Nevertheless, although they are around the same size as krill, and may also
28 occur in swarms, their feeding behaviour and mode of reproduction are completely different, hence
29 their respective impacts on ecosystem structure differ. *Themisto* are major predators of meso- and
30 macrozooplankton in several major oceanic regions covering shelves to open ocean from the polar
31 regions to the subtropics. Based on a combination of published and unpublished occurrence data, we
32 plot out the distributions of the seven species of *Themisto*. Further, we consider the different predators
33 that rely on *Themisto* for a large fraction of their diet, demonstrating their major importance for higher
34 trophic levels such as fish, seabirds and mammals. For instance, *T. gaudichaudii* in the Southern Ocean
35 comprises a major part of the diets of around 80 different species of squid, fish, seabirds and marine
36 mammals, while *T. libellula* in the Bering Sea and Greenland waters is a main prey item for
37 commercially exploited fish species. We also consider the ongoing and predicted range expansions of
38 *Themisto* species in light of environmental changes. In northern high latitudes, sub-Arctic *Themisto*
39 species are replacing truly Arctic, ice-bound, species. In the Southern Ocean, a range expansion of *T.*
40 *gaudichaudii* is expected as water masses warm, impacting higher trophic levels and biogeochemical
41 cycles. We identify the many knowledge gaps that must be filled in order to evaluate, monitor and
42 predict the ecological shifts that will result from the changing patterns of distribution and abundance
43 of this important pelagic group.

44 **KEYWORDS**

45 Hyperiidea, biogeography, range shifts, food web, life cycle, Antarctic krill, salps, climate change

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80 1. BACKGROUND

81 Five major groups of zooplankton are characteristic of high-latitude oceans, copepods, soft-bodied
82 zooplankton (e.g. tunicates, cnidarians), pelagic amphipods, euphausiids and chaetognaths (Longhurst,
83 1985). Of these groups, amphipods are amongst the least known (e.g. Murphy et al., 2007). Unlike the
84 chaetognaths and euphausiids that comprise relatively few species with little variation in morphology
85 and feeding behaviour, pelagic amphipods are highly diverse. This is reflected in their wide range of
86 feeding habits, which is as diverse as that of copepods, and comprises carnivory, omnivory and even
87 herbivory in certain developmental stages. There are also parasitic and commensal forms. Such varying
88 lifestyles is manifested in pronounced morphological diversity which is comparable to that of
89 cnidarians. This diversity is far from being fully described and understood and deserves much greater
90 attention.

91 The Hyperiidea represent the most dominant group of pelagic amphipods, comprising exclusively
92 pelagic species. They are believed to be the most ancient amphipod colonizers of the pelagic realm, as
93 opposed to the Gammaridea, of which only about 30% of the species inhabit the pelagial which they
94 colonized much later in evolutionary history (Vinogradov, 1999a). Hyperiids span the size range of
95 around 2 mm adult size to a maximum of 10 cm recorded for the genus *Megalanceola* (Zeidler, 1992).
96 They contribute up to 20% of all zooplankton biomass in some regions, but generally are in about the
97 same range as other so-called raptorial planktonic predators: the chaetognaths, which in total
98 comprise 4% of the global ocean's zooplankton biomass (Longhurst, 1985). So far, 286 hyperiid species
99 belonging to 32 families and 77 genera (De Broyer, 2010) have been described from the open ocean,
100 the majority of which inhabit the epipelagic zone, however several are mesopelagic and deep-water
101 species (Vinogradov, Volkov & Semenova, 1996; Vinogradov, 1999a).

102 Recently their phylogenetic relationships have been investigated with modern molecular tools which
103 confirmed the presence of two monophyletic groups: the Physosomata, mainly confined to
104 bathypelagic depths and the Physocephalata, inhabiting primarily epi- and mesopelagic depths (Hurt,

105 Haddock & Browne, 2013). This independent radiation, segregated on the bathymetric scale, is
106 reflected in the morphological characteristics of both groups. Whilst the Physosomata often show an
107 overall reduction in the size of the head and eyes relative to the body as well as a cryptic coloration
108 typical of deep-sea organisms, most Physocephalata have large heads and eyes relative to their body
109 length and are often transparent (Hurt et al., 2013). Despite these generalizations, the range in
110 variation of hyperiid morphology can reach bizarre proportions in some highly specialised species and
111 contrasts with the relatively similar body shapes across an order of magnitude size scale in other
112 pelagic crustacea: copepods, euphausiids and decapods. Many species may have coevolved alongside
113 large-volume zooplankton, in particular cnidarians that themselves exhibit a broad range of body
114 plans. Indeed, hyperiids are considered as an entirely pelagic group but are described as having a
115 “quasi-benthic lifestyle” where soft-bodied (often lumped under the term gelatinous) zooplankton
116 such as salps and jellyfish function as moving substrate. These are often indispensable to the
117 completion of the hyperiid’s life cycle for shelter, reproduction, food and predator avoidance (Laval,
118 1980). Many reports exist on a commensal or parasitic relationship with ctenophores, cnidarians and
119 salps (e.g. Harbison, Diggs & Madin, 1977; Gasca & Haddock, 2004). The co-evolution with other
120 plankton can also be illustrated by the example of two Antarctic *Hyperiella* species that carry live
121 pteropods (*Clione* and *Spongiobranchaea*) on their backs, holding these between their elongated
122 pereopods as an efficient chemical defence against fish predators (Havermans et al., 2018).

123 Soft-bodied zooplankton are classically regarded as a trophic ‘dead end’ in the pelagic food web: even
124 though the disparate groups, e.g. cnidarians and tunicates, that fall in this category can build up an
125 enormous biomass very rapidly by asexual reproduction, few pelagic predators seem to benefit from
126 their abundances. However, this is contested; besides a relatively small number of specialists on a soft-
127 bodied plankton diet (e.g. Harbison, 1993; Mianzan et al., 1996), a majority of predators use soft-
128 bodied zooplankton as part of their diet (Arai, 2005) as so-called ‘survival-food’ when preferred prey
129 items are limited (e.g. anchovies feeding on salps, Mianzan et al., 2001). Hyperiid amphipods, with
130 their grappling and tearing mouthparts, are particularly well adapted to feeding on soft-bodied

131 zooplankton and parasitizing them for completing (part of) their life cycle. This is confirmed by a high
132 predation pressure on hydromedusae by hyperiids (e.g. Mills, 1993). Regional studies have clearly
133 demonstrated a relationship between the distribution of several species of hyperiids and the presence
134 of salps (e.g. Young, 1989) and other groups (e.g. radiolarians, ctenophores, siphonophores, e.g.
135 Colebrook, 1977). Burrige et al. (2017) linked the distribution and diversity of hyperiids sampled
136 throughout the Atlantic with those of soft-bodied zooplankton. On the other hand, the importance of
137 parasitic hyperiids has recently been emphasized as an important energy transfer pathway, with fish
138 preying on hyperiids within jellyfish and hence, as a hitherto unstudied link between the so-called
139 trophic dead end and fishes in pelagic ecosystems (Riascos et al., 2012). In the context of hypothesized
140 synergistic events of the overfished fish stocks and increasing blooms of soft-bodied zooplankton,
141 these interactions in the shape of parasitism, commensalism and predation urgently need a more
142 concentrated research effort.

143 Hyperiid amphipods of the genus *Themisto* Guérin, 1825 (a senior synonym of *Parathemisto*, Bowman
144 et al., 1982) play an important role in high-latitude and temperate waters where they often represent
145 a major trophic link between zooplankton secondary production and higher trophic levels such as
146 squid, fish, seabirds and marine mammals (see section VI in this review). *Themisto* amphipods are
147 believed to be voracious visual predators using their large compound eyes to detect and feed upon
148 meso- and macrozooplankton in the epipelagic layer. *Themisto* feeds upon the most abundant
149 zooplankton species in the water column and can control the mesozooplankton standing stock.
150 However, a phytoplankton diet has been proposed for the juvenile life stages (see section IV). The
151 genus is currently represented by seven species (Zeidler, 2004): *T. gaudichaudii* Guérin, 1825, the most
152 abundant amphipod in the southern hemisphere, *T. japonica* (Bovallius, 1887) and *T. pacifica*
153 (Stebbing, 1888) from North Pacific waters and *T. australis* (Stebbing, 1888) from the colder waters of
154 the Southwest Pacific and *T. libellula* (Lichtenstein in Mandt, 1822), *T. compressa* Goës, 1865 and *T.*
155 *abyssorum* (Boeck, 1871), which inhabit temperate Atlantic and Arctic waters. *T. gaudichaudii* was
156 previously believed to be an amphitrophic species, occurring in both hemispheres, but has been

157 revised to comprise *T. gaudichaudii* in the southern hemisphere and *T. compressa* in the northern
158 hemisphere (Schneppenheim & Weigmann-Haas, 1986). Synonymized species are *T. bispinosa* Boeck,
159 1871 that is now accepted as *T. compressa* and *T. gracilipes* (Norman, 1869), now *T. gaudichaudii*.
160 However, records of *T. gracilipes* north of the Southern Ocean, such as those in Australian and New
161 Zealand waters, may refer to *T. australis*.

162 Climate change, proceeding at an unprecedented pace, is currently redistributing life on Earth (Pecl et
163 al., 2017). Warming of the upper ocean layer and the atmosphere have altered sea ice extent and
164 seasonal dynamics in the Arctic (Screen & Simmonds, 2010; Stroeve et al., 2014), and similar changes
165 are observed in the Atlantic sector of the Southern Ocean, the western Antarctic Peninsula and
166 Bellingshausen Sea (Meredith & King, 2005; Gille, 2008; Whitehouse et al., 2008; Stammerjohn et al.,
167 2012). This has a strong impact on stocks of key pelagic species such as Antarctic krill (*Euphausia*
168 *superba* Dana, 1850). In light of these environmental changes, range expansions or shifts in the polar
169 pelagic realm are ongoing or predicted for some species whilst others, e.g. ice-dependent species, are
170 undergoing poleward range contractions. Within the SW Atlantic sector of the Southern Ocean, a
171 decline of Antarctic krill densities is hypothesized (although still debated) concomitant with an increase
172 in salps (mainly *Salpa thompsoni* Foxton, 1961), which is often attributed to bottom-up factors such as
173 alterations in summer phytoplankton blooms and winter sea-ice extent (Loeb et al., 1997; Atkinson et
174 al., 2004; Meyer, 2012). In the Arctic Ocean and surrounding seas, changes in the distributional range
175 of *Themisto libellula* have also been reported (Marion et al., 2008; Volkov, 2012), while *T. compressa*
176 has recently invaded the Arctic Ocean in the Fram Strait (Kraft et al., 2013). Hence, in order to make
177 reliable predictions of the consequences of such distributional shifts and the effects of environmental
178 changes, we feel a stock-take of the information available on *Themisto* amphipods is urgently needed,
179 as well as highlighting what needs to be studied to determine the future status and role of this key
180 group in global plankton communities. Therefore, we will discuss the knowns and known unknowns of
181 *Themisto* amphipods regarding distributional patterns, life history traits, feeding habits and their role
182 in regional food webs and biogeochemical cycles and develop hypotheses on their ecology and biology

183 based on literature and observations. In doing so, we provide both the current status of this group and
184 move towards predicting the consequences of range shifts of *Themisto* species for high-latitude
185 ecosystems.

186 **2. DISTRIBUTIONAL PATTERNS AND SPECIES ZONATION OF *THEMISTO***

187 Distributional ranges of macrozooplankton are often linked with oceanographic features and the
188 distribution of their major prey, or both. Some species, such as *Themisto libellula* and *T. abyssorum*,
189 are assumed to be indicators of particular water masses: *T. libellula* is a typical species of cold Arctic
190 waters in different sub-Arctic regions, whilst *T. abyssorum* is more associated with warmer Atlantic
191 waters (e.g. Mumm et al., 1998; Dalpadado, 2002; Volkov, 2012). Nonetheless, *T. libellula* is not only
192 thriving in the Arctic but also in its marginal seas (Fig. 1), where water layers < 3°C are present
193 throughout summer, including the Bering and Okhotsk seas, as well as in southern Alaskan fjords,
194 Prince William Sound and the Gulf of St Lawrence (Marion et al., 2008; Pinchuk et al., 2013). In the
195 southern Alaskan fjords, as well as in Prince William Sound, no extensive cold layers persist, and also
196 in the Bering Sea, the upper layers are 9°C in the coldest years and 14°C in the warmest (Pinchuk et al.,
197 2013). *T. libellula*'s upper lethal temperature (at which 50% of the animals die) has been
198 experimentally determined to be 9.4°C for the Canadian Arctic populations (Baffin Bay), whereas it is
199 between 13 – 15°C for individuals of Alaskan populations (Percy, 1993). This shows that some
200 populations are physiologically adapted to warmer waters by shifting their thermal ranges (Percy,
201 1993), which may be the case for other geographic populations as well.

202 Similarly, *Themisto abyssorum* is also found in the Arctic Barents Sea, although in tenfold lower
203 abundances than in waters of Atlantic origin (e.g. Dalpadado, 2002), indicating a broad temperature
204 tolerance. However, contrary to *T. libellula*, it is absent from the Bering Sea and Pacific (Fig. 1). The
205 species supposedly prefers deeper waters (> 50m), mostly linked to the presence of deep Atlantic
206 water in the Arctic Ocean, possibly explaining its absence in the shallow Bering Sea. However, surface

207 records of this species also exist (Dalpadado, 2002; Havermans C., unpublished data), and it is likely
208 that other bottom-up or top-down factors are having an impact on *T. abyssorum*'s realized distribution.

209 *Themisto australis* is present in the southwestern Pacific, but seemingly absent from the eastern part
210 (Fig. 2), the reasons for which being still unclear. Both *T. compressa* and *T. gaudichaudii* are
211 characterized by a very wide distribution encompassing both polar and temperate regions (Figs. 1 and
212 2). *T. compressa* is distributed in the western Atlantic from 40°N to about 66°N in the Davis Strait whilst,
213 in the eastern Atlantic, it can be found as far north as the northern Barents Sea (79°N), down to about
214 30°N off the Moroccan coast. It is also present in the Mediterranean Sea from Gibraltar to about 24°W.

215 In the southern Atlantic and Southern Ocean, *Themisto gaudichaudii* can be found in waters to the
216 North and South of the Polar Front. The species occurs in waters from subzero temperatures around
217 the Antarctic Peninsula and Weddell Sea (66 – 70°S) to as far north as the Benguela upwelling system
218 (Kane, 1966, Auel & Ekau, 2009) and the Patagonian shelf and coast (Ramírez & Viñas, 1985; Padovani
219 et al., 2012) (Fig. 2). *T. gaudichaudii* is regarded as a species typical of the warmer (surface) waters of
220 the Antarctic (Mackintosh, 1934) and is more common in the northern Scotia Sea to as far south as the
221 Bransfield Strait (Jazdzewski & Presler, 1988). In contrast to the high abundances of *Themisto* species
222 observed throughout Arctic water masses, Southern Ocean distributions seem to be very patchy with
223 only particular areas harbouring high amphipod concentrations. This can be explained by the fact that
224 the Southern Ocean itself is a mosaic of high and low productivity regions, with the coastal and
225 continental shelf zones being amongst the most productive (Constable, Nicol & Strutton, 2003). The
226 Antarctic Polar Frontal (APF) Zone, situated between the Polar and sub-Antarctic fronts is also
227 characterized by an elevated primary production and intense eddy and frontal activities (Constable et
228 al., 2003). In both areas, *T. gaudichaudii* has high abundances, e.g. around South Georgia, the
229 Kerguelen, Heard, Crozet and Prince Edward Islands, and in the APF zone (Ealy, 1954; Kane, 1966;
230 Labat, Mayzaud & Sabini, 2005; Pakhomov & Froneman, 1999; Froneman, Pakhomov & Treasure,

231 2000; Watts & Tarling, 2012). Whether these patchy distributions can be linked with temperature,
232 particular prey abundances or concentrations of predators needs to be further examined.

233 The interesting feature of the distribution of *Themisto* species is that it extends across several
234 latitudinal zones of prey species. In the case of *T. gaudichaudii*, its southern range overlaps with the
235 northern range of *Euphausia superba* and covers the ranges of the euphausiid species *E. frigida*
236 Hansen, 1911, *E. triacantha* Holt & Tattersall, 1906, *Thysanoessa macrura* G.O. Sars, 1883 and *T. vicina*
237 Hansen, 1911 (Brinton, 1985). Furthermore, several *Themisto* species have overlapping geographic
238 distributions. This is the case for example for *T. libellula*, *T. abyssorum* and *T. compressa* in the Arctic
239 Ocean and shelf seas (Fig. 1), for *T. libellula* and *T. pacifica* in the Sea of Okhotsk (Gorbatenko, Grishan
240 & Dudkov, 2017) and for *T. pacifica* and *T. japonica* in the western sub-Arctic Pacific (Bowman, 1960;
241 Yamada, Ikeda & Tsuda, 2004). Where distributions overlap, each species occupies a distinct ecological
242 niche. For instance, both the sub-Arctic boreal *Themisto abyssorum* and the high-Arctic *T. libellula* are
243 present sympatrically in the Arctic Ocean and surrounding seas, but they feed on different prey (Auel
244 et al., 2002; Kohlbach et al., 2016).

245 Nevertheless, the genus *Themisto* is in urgent need of a taxonomic revision and the biogeographic
246 limits of the species must be tested with molecular tools. *T. gaudichaudii* has been shown to consist of
247 at least three distinct genetic lineages throughout the Atlantic sector of the Southern Ocean
248 (Havermans C. et al., in preparation) and in-depth population genetic studies should be carried out to
249 evaluate the extent of gene flow between these populations. Within the Southern Ocean at least two
250 morphospecies have been distinguished (Zeidler & De Broyer, 2014). Populations along the Patagonian
251 shelf consist of *T. gaudichaudii* (Havermans C. et al., in preparation), whilst the populations from the
252 Benguela upwelling system have not yet been revised according to their differing morphology and
253 genetic connectivity. The morphological differences between *T. pacifica* and *T. japonica* are minute
254 (Yamada et al., 2004) and only a century after their description has a study pinpointed characters
255 allowing immature specimens of these two species to be distinguished from each other (Yamada &

256 Ikeda, 2004). Furthermore, several characters used to distinguish mature adults of both species
257 (Yamada et al., 2004) are also prone to vary according to sex and developmental stage (e.g. length of
258 second antennae), which may lead to further identification errors. Within *T. libellula*, several distinct
259 genetic lineages have been revealed, linked to regional variation (Tempestini et al., 2017). Only after
260 the genus *Themisto* has been thoroughly revised with an integrative approach combining morphology
261 and genetics, can further conclusions be made regarding species' zonation and distributional patterns.

262 **3. LIFE-HISTORY TRAITS AND SMALL-SCALE DISTRIBUTIONAL PATTERNS OF *THEMISTO***

263 **3.1. Life cycles of the different *Themisto* species**

264 In the genus *Themisto*, the number of generations per year decreases with increasing latitude: the
265 respective boreal and Arctic species *T. libellula* and *T. abyssorum* have one generation every year or
266 every two years, whilst warmer-water species such as *T. japonica*, *T. pacifica* and *T. compressa* have
267 several generations per year (Ikeda, Hirakawa & Imamura, 1992) (Table 1). This does not hold true for
268 *T. gaudichaudii*, for which the number of generations varies throughout its distributional range.
269 Around South Georgia, it has two recruitment events per year (Watts & Tarling, 2012) but only one
270 around the sub-Antarctic Kerguelen Islands (Labat et al., 2005). However, this statement is subject to
271 the validity of the current species delimitation (see above). *Themisto* populations off South Africa have
272 a life cycle of less than a year and females become mature when reaching 6 mm of length (Siegfried,
273 1965) whilst in Antarctic waters, *T. gaudichaudii* grows to a larger size with a maturity of around 12
274 mm or more, displaying slower growth rates (Barnard, 1932). Hence, it has been argued that growth
275 and maturation rates depend on food availability and temperature (Sheader, 1981; Auel & Ekau, 2009).
276 Breeding periods and number of generations per year also differ amongst sympatric species. *T. libellula*
277 has a prolonged breeding period from January to March, however, breeding females have been
278 recorded as early as July to September in Svalbard fjords (Dale, 2006). *T. libellula* females release
279 juveniles in a time frame (March to May) matching the spring blooms in the Marginal Ice zones of the
280 Arctic Ocean (Dalpadado, 2002). In the Bering Sea, this peak release occurs much later in June (Pinchuk

281 et al., 2013). *T. abyssorum*, strongly associated with the Atlantic inflow in the Arctic, breeds later and
282 over a shorter time period (May and June) (Dalpadado et al., 1994; Dalpadado, 2002). For *T. japonica*,
283 experimentally determined life cycles varied with temperature and almost doubled in duration upon
284 exposure to waters at 1°C compared to those at 5°C (Ikeda, 1990). Individuals of *T. libellula*, as a typical
285 Arctic species, appear to be smaller in Atlantic waters (Dalpadado, 2002). Hence, growth and
286 maturation rates depend on temperature and food availability (Sheader, 1981; Yamada et al., 2004;
287 Auel & Ekau, 2009). In most species, peaks of hatched juveniles seem to be synchronized with the
288 increase of seawater temperatures in spring and its associated phytoplankton blooms followed by
289 increases in zooplankton abundances (e.g. *T. gaudichaudii*, Labat et al., 2005; *T. libellula*, Noyon,
290 Gasparini & Mayzaud, 2009). This timing allows juveniles, reported to feed both herbivorously as well
291 as on mesozooplankton (see below), to take advantage of increased food supply and pass through the
292 more vulnerable life stages quickly.

293 **3.2. Do *Themisto* species swarm?**

294 *Themisto* amphipods are very motile and have been reported to occur in large swarms (e.g. Vinogradov
295 et al., 1982). Net catch data reported hundreds of *Themisto* individuals per square meter (e.g. *T.*
296 *abyssorum*: 269 ind.m⁻², Dalpadado, 2002; *T. japonica*: 622 ind.m⁻², Ikeda et al., 1992). However, more
297 research is needed to find out whether these high densities represent just locally aggregating
298 individuals feeding upon patchily distributed prey or true schooling behaviour (Hamner, 1984). For the
299 hyperiids *Hyperoche* and *Themisto*, Westernhagen & Rosenthal (1976) suggest chemical or visual
300 detection of copepod prey, but they hypothesize that predation depends on random encounters,
301 therefore requiring a minimum density of prey. Hence, active hunting of copepods may be facilitated
302 by the formation of swarms. Swarms have also been suggested to be linked to certain reproductive
303 stages. Anecdotal underwater observations around the sub-Antarctic Snares Islands have reported the
304 occurrence of *T. gaudichaudii* and *T. australis* in loose swarms in the neuston layer (down to 3 m depth)
305 (Fenwick, 1973). Camera images from subsurface layers in the Fram Strait showed many but rather

306 spaced out encounters of *T. libellula* (Havermans C., unpublished data). From submersibles, near-
307 bottom swarms of hundreds of *T. abyssorum* mature females have been observed several times at
308 1700 m depth. Acoustic records have shown diel vertical migrations of swarms of zooplankton,
309 including *T. gaudichaudii*, that forage in near surface waters at night and descend to the seafloor after
310 sunrise (Pakhomov & Froneman, 1999). The swarming or aggregating habit of *Themisto* may explain
311 its dominant role in the macrozooplankton compared with other hyperiids with similar morphological
312 and ecological traits. Two other abundant hyperiid species in the Southern Ocean are *Cylopus lucasii*
313 and *Primno macropa*. In common with *Themisto*, they are good swimmers, and are not commensal or
314 parasitic on soft-bodied zooplankton (Zeidler & De Broyer, 2014). Logically, species adapted to a
315 commensal life style depend on their host and remain solitary rather than form dense aggregations.
316 However, *P. macropa* and *C. lucasii* are not known to swarm (Vinogradov, 1999b) but can still be found
317 in sufficient biomass to represent a major food source for top predators (Zeidler & De Broyer, 2014).
318 Combining optical with advanced acoustic methods may be pivotal for understanding to what extent
319 swarming occurs in pelagic amphipods and its selective advantage over the more solitary lifestyles of
320 other hyperiid species.

321 **3.3. Vertical distributions and diel migrations: do all species exhibit the same patterns?**

322 Diel vertical migrations (DVM), of ascent at night and descent during the day, have been well
323 documented for *Themisto* species (Ikeda et al., 1992). It is yet unknown what triggers the diel vertical
324 migration of *Themisto* species. If these migrations are determined by the vertical distribution of their
325 prey, in this case copepods, would *Themisto* feed on these during diurnal aggregation at depth, by
326 looking upward, or chasing them in the surface layer at night? *Themisto* species are assumed to be
327 visual predators based on the large size of their eyes, but they are still capable of capturing copepods
328 during imposed periods of darkness when kept in aquaria (Pakhomov & Perissinotto, 1996). The optical
329 structure of *Themisto's* eyes, in comparison to other hyperiids, reveals an increased resolution
330 particularly in the forward-pointing part of the lower eye (Land, 1989). Hence, *Themisto* not only uses

331 the dorsal upward looking direction but, in addition, has enhanced visual acuity looking forwards (Land,
332 1989). This, together with a better understanding of its hunting habits, could provide an answer to the
333 questions above.

334 Whether the ascent to surface layers during the night is a consistent pattern for all *Themisto* species
335 across regional populations still needs to be ascertained. For instance, *T. gaudichaudii* has been
336 reported at the surface during day time: e.g. off Terra Nova (Barnard, 1930), as well as along the
337 Patagonian shelf (Havermans C., unpublished data). In many sampling localities in New Zealand and
338 sub-Antarctic waters, *T. australis* was not found in any catch at the surface at night but was there
339 during day time, rising to the surface in the afternoon (Fenwick, 1978). Also a large portion of the *T.*
340 *compressa* population spent more time in surface layers, independent of day/night time (Lampitt et
341 al., 1993). Similarly, in the shelf regions of the Prince Edward Islands, part of the *T. gaudichaudii*
342 population did not display a clear diel vertical pattern and remained in the upper 100 m whereas
343 another fraction of the population descended to depths between 200 – 400 m (Pakhomov &
344 Froneman, 1999). This was also visible in the acoustic record, where small swarms occurring between
345 50 and 100 m tended to descend after sunrise, to greater depths, sometimes to the shelf floor
346 (Pakhomov & Froneman, 1999). A sinking behaviour towards deeper depths straight after feeding may
347 also explain these descents, similar to the satiation sinking behaviour discovered for Antarctic krill
348 (Tarling & Thorpe, 2017). In another study, nighttime abundance of *T. gaudichaudii* was consistently
349 higher than day time levels on the Prince Edward Islands' shelf, and no vertical variation in distribution
350 between size classes was observed (Pakhomov & Froneman, 1999). Juveniles and immatures of *T.*
351 *japonica* migrate to depths of 150 – 200 m at daytime, whereas smaller size classes of the co-occurring
352 *T. pacifica* stay in shallower waters both at night- and daytime. Furthermore, in both species, a
353 segregation exists between mature males and females (Yamada et al., 2004). For *T. japonica*, males
354 were never found in daytime samples, indicating a deeper descent (beyond 500 m) and an ascent
355 during daytime to depths < 100 m (Yamada et al., 2004). Between these two species, the extent of
356 vertical migration, as well as the daytime distribution depth, also differ, as a function of the superior

357 swimming abilities of *T. japonica* and a higher risk of predation associated with its larger body size
358 (Yamada et al., 2004). Hence, there is much variation in DVM behaviour between *Themisto* species and
359 even between regional populations of the same species. Furthermore, we presently have little
360 understanding of what triggers DVM in *Themisto*. A more frequent use of opening/closing nets will
361 decrease the uncertainties about the precise depth distributions of the different species.

362 **3.4. Local and regional segregation of juveniles, males and females**

363 Many hyperiid species are known to form single-sex swarms, particularly during the reproductive
364 period, but *Themisto* species were considered to be one of the exceptions in forming mixed swarms
365 during this time (Laval, 1980 and references therein). However, both for *T. gaudichaudii* and *T. libellula*,
366 several authors have reported males to be absent, or only present in low densities in their samples
367 (Barnard, 1930, 1932; Schneppenheim & Weigmann-Haass, 1986), which may indicate separate
368 swarms outside reproductive periods. For *T. japonica* and *T. pacifica*, males and females show distinct,
369 but overlapping, vertical distributions (Yamada et al., 2004). Active migrations associated with growth
370 stages have also been proposed (e.g. Labat et al., 2005). In Toyama Bay in the Sea of Japan, adult
371 females only appeared in spring (Ikeda et al., 1992). In Arctic Kongsfjorden where a year-round
372 presence of *T. libellula* has been recorded, mature females have never been caught, however, juveniles
373 are found in high abundances (Noyon et al., 2011). Vast numbers of *T. compressa* (then: *Parathemisto*
374 *gaudichaudii*) were found washed upon the shore of Northeast England, turning the beaches white,
375 which consisted of females carrying young, eggs, and many recently hatched juveniles (Gray &
376 McHardy, 1967). These examples may corroborate other previous findings (Labat et al., 2005; Noyon
377 et al., 2011) that females release their brood nearshore, entering bays or fjords and subsequently
378 leaving these “nursery” areas. Around Svalbard, first- and second-year specimens of *T. libellula* have
379 been found in different fjords (Noyon et al., 2011) and, for *T. gaudichaudii* in the Kerguelen
380 archipelago, younger individuals dominate the sheltered sites between the islands and segregate from
381 larger-sized individuals offshore (Labat et al., 2005).

382 *Themisto* juveniles seem to be segregated vertically, being distributed in the top 100 m layer (daytime:
383 0 – 100 m, nighttime: 0 – 50 m, e.g. Yamada et al., 2004) and, in some cases, appear not to perform
384 DVM, possibly because of surface layer temperatures (Ikeda et al., 1992). Size segregation may avoid
385 competition or cannibalism on newly hatched juveniles. A geographic separation of
386 recruitment/nursery areas from the feeding grounds of mature individuals, known to be the case for
387 Antarctic krill (Meyer et al., 2017), may enhance recruitment success and dispersal dynamics of the
388 different populations. Investigations of patterns of gene flows may be one means of determining
389 whether such segregation is also commonly prevalent in *Themisto* species.

390 **3.5. Commensalism or parasitism on soft-bodied zooplankton**

391 In many species, the use of a planktonic host is assumed to ensure food availability when juveniles
392 hatch. In other species, juveniles are capable of catching pelagic prey directly upon release from the
393 brood pouch: *Hyperoche medusarum* (Krøyer, 1838) juveniles immediately prey on herring larvae
394 when leaving the the marsupium, as observed in aquaria (von Westernhagen & Rosenthal, 1976). They
395 have been observed clinging onto herring larvae, after having grasped them by the tail, and then
396 sinking together to the bottom where they continue feeding on them (von Westernhagen & Rosenthal,
397 1976). Juveniles of *Themisto pacifica* have been collected from medusae (*Calyropsis nematomorpha*
398 Bigelow, 1913) in the sub-Arctic Pacific Ocean (Renshaw, 1965). Juveniles of the same species have
399 been reported to infest *Aequorea* medusae, living inside their stomachs where they feed on partially
400 digested prey, whilst larger individuals have been found burrowed in the jelly or grazing on
401 subumbrellar structures (Mills, 1993). Similarly, *T. australis* was associated with the scyphozoan
402 *Cyanea capillata* (Linnaeus, 1758). The amphipods did not seem to feed on the jellies but rather use
403 them as a substrate to attach to (Condon & Norman, 1999), likely facilitating dispersal. Some salps
404 (*Pegaea, lasis*) collected in the Atlantic were covered with recently hatched *Themisto* juveniles, which
405 has been interpreted as a close association between juveniles and salps (Madin & Harbison, 1977).
406 Despite these observations, this relationship is thought to be much more tenuous than most other

407 interactions documented for hyperiids (Zeidler & De Broyer, 2014) and many authors argue that
408 *Themisto* release juveniles into the pelagic environment without the presence of a host (e.g. Dunbar,
409 1957; Siegfried, 1965; Kane, 1963, 1966). After hatching, juveniles likely colonize the salps
410 independently, to which they commonly attach using their pereopods as shown in Fig. 3c. In other
411 hyperiids, the females actively find salps or other gelatinous zooplankton and demarsupiate their
412 brood into their tissues. This does not seem to be the case for most *Themisto* species, with the
413 potential exception of *T. pacifica*, of which specimens were found inside medusae. In the Southern
414 Ocean, at a sampling site where hundreds of *T. gaudichaudii* juveniles were recovered, salps were
415 absent. On the contrary, where many adults were found, salps densities were high (Havermans,
416 Schöbinger & Schröter, 2017). This observation does not support the hypothesis that salps are hosts
417 for juvenile stages but adults likely feed on salps. However, an algal bloom was observed at the site
418 where juveniles were abundant (Havermans et al., 2017), which supports their herbivorous feeding
419 habits and the synchronization of juvenile hatching and spring blooms, observed for *Themisto* species
420 (e.g. Dalpadado, 2002).

421 **4. THEMISTO'S FEEDING ECOLOGY**

422 **4.1. From herbivory to carnivory: which trophic niches do *Themisto* species occupy?**

423 *Themisto* amphipods are believed to be roving predators, feeding on the most abundant taxa in the
424 water column. In the southern hemisphere, gut content analyses of *T. gaudichaudii* have shown that
425 it feeds non-selectively and opportunistically, on copepods, chaetognaths, euphausiids and pteropods,
426 amongst other taxa (Siegfried, 1965; Hopkins, 1985; Gibbons, Stuart & Verheye, 1992; Pakhomov &
427 Perissinotto, 1996). In the Benguela Upwelling system, it was shown to consume the most abundant
428 copepod and chaetognath species (Gibbons et al., 1992). Nonetheless, other studies focusing on the
429 feeding dynamics of *T. gaudichaudii* are surprisingly scarce: two studies have been carried out in
430 nearshore waters of (sub-) Antarctic islands (Pakhomov & Perissinotto, 1996; Froneman et al., 2000),
431 one study was done off the West Coast of South Africa (Siegfried, 1965) and one in the Polar Frontal

432 Zone (Lange, 2006). Virtually nothing is known about *T. gaudichaudii*'s feeding ecology elsewhere, e.g.
433 on the Patagonian shelf.

434 Salps have been reported in gut contents of *Themisto gaudichaudii* collected near the Antarctic
435 Peninsula (Hopkins, 1985), and on the basis of its well-suited grappling appendages it has been
436 hypothesized that the species is a major predator of salps more widely (Smetacek, Assmy & Henjes,
437 2004). Unfortunately, conventional gut content analyses with microscopy often fail to find soft-bodied
438 zooplankton due to their rapid degradation in the stomach and lack of hard features for identification
439 (Arai et al., 2003). Feeding experiments of *T. gaudichaudii* have shown that adults feed on salps,
440 particularly on their stomachs (see Fig. 3d), a habit which may be held responsible for the presence of
441 biomarkers for herbivory in adult *Themisto* (e.g. Stowasser et al., 2012). Based on both morphological
442 stomach analyses and stable isotopes, Kruse et al. (2015) hypothesized an extensive feeding of *T.*
443 *gaudichaudii* on salps in the Polar Frontal zone. *Salpa thompsoni* DNA has also been successfully
444 amplified from stomach contents of *T. gaudichaudii* sampled in the Polar Frontal Zone (Havermans C.,
445 unpublished data). During an *in-situ* iron-fertilization experiment carried out in the same region, *T.*
446 *gaudichaudii* was the dominant macrozooplankton species that colonized the fertilized patch, showing
447 a two-fold higher abundance within the patch (Mazzocchi et al., 2010). *T. gaudichaudii* may have been
448 attracted to the phytoplankton bloom within the patch to prey on salps, which would explain the low
449 numbers of salps observed. In this case, *Themisto* would form an efficient link between the gelatinous
450 and muscular food chains (Verity & Smetacek, 1996).

451 The position of *Themisto* species in Arctic food webs is better understood, particularly in the European
452 Arctic. Trophic studies have been carried out both in open waters (Fram Strait, Auel et al., 2002;
453 Kohlbach et al., 2016) and coastal regions (Svalbard fjords, Noyon et al., 2009, 2011) as well as in
454 temperate ecosystems (Gulf of St. Lawrence, Marion et al., 2008). Both *T. libellula* and *T. abyssorum*
455 are known to feed predominantly on copepods. Only one account of feeding on gelatinous
456 zooplankton has been reported for Arctic species, despite "jellies" being ubiquitous and occurring in

457 high abundances (e.g. Rascoff et al., 2010). Only one specimen of *T. abyssorum* investigated from
458 slurpgun samples taken with submersibles had a jellyfish tentacle in its stomach (Vinogradov, 1999b).
459 Despite their co-occurrence, *T. abyssorum* and *T. libellula* populations occupy distinct ecological
460 niches. *T. libellula* feeds on herbivorous copepods that are dependent on the cryo-pelagic pathway
461 involving ice algae (sympagic diatoms) (Auel et al., 2002; Kohlbach et al., 2016). By contrast, *T.*
462 *abyssorum*'s feeding involves a more variable, less ice-dependent, trophic pathway where a variety of
463 mesozooplankton grazing on flagellates and *Phaeocystis* seems to be the main prey (Auel et al., 2002;
464 Kohlbach et al., 2016). Biomarker analyses indicated a higher trophic level for *T. abyssorum* than *T.*
465 *libellula*, suggesting greater feeding on omnivorous and carnivorous prey (Auel et al., 2002). *T. libellula*
466 seems to prefer copepodite stages CIII of *Calanus* species but can also feed on smaller copepods such
467 as *Oithona* and *Pseudocalanus* species, when abundances reach a certain threshold (Noyon et al.,
468 2009). In the St Lawrence system, stomach content analyses indicate feeding on copepodite stages CIV
469 and CV of *Calanus finmarchicus* (Gunnerus, 1770), complemented by euphausiids, chaetognaths,
470 amphipods and mysids (Marion et al., 2008). In the North Atlantic, *T. abyssorum*'s diet, investigated
471 with molecular methods, consisted mainly of crustaceans but detritus also appeared to be an
472 important food source (Olsen et al., 2013). *T. compressa* and *T. abyssorum* have both been
473 hypothesized to feed on particles in the water column. In the Arctic, *T. libellula* and *T. abyssorum*
474 accumulate high amounts of wax esters (> 40% of total lipids), with their proportion increasing with
475 individual size (Auel et al., 2002). This contrasts with *T. gaudichaudii*, which has virtually no wax esters
476 (0.1%). Despite the comparatively more complete knowledge of *Themisto* feeding ecology on the
477 Northern hemisphere, the scarcity of reports of feeding on gelatinous zooplankton should not be taken
478 as evidence of its absence in *Themisto* diet due to the high probability of false negatives until
479 investigated with methods that are not misled by the absence of persistent hard structures identifiable
480 in amphipod stomachs.

481 **4.2. Do functional morphologies indicate distinct prey preferences in *Themisto*?**

482 In the pelagic realm, there are two kinds of predators: engulfers (e.g. fish) and grapplers (e.g.
483 cephalopods). *Themisto* belongs to the latter type. As well as using its appendages to grapple and
484 manipulate prey items, aquarium observations have shown that *T. gaudichaudii* uses its posterior long
485 legs (pereopods) to manoeuvre: for stopping, turning sharply or making movements towards food
486 items in immediate proximity (Kane, 1963). The grasping and holding on of prey is mainly achieved by
487 the posterior pereopods, in particular the fifth pair, which is longer than the others (Nemato & Yoo,
488 1970). In the case of *T. gaudichaudii*, *T. australis* and *T. libellula*, the fifth pereopods also have well-
489 developed spines and setae along their anterior edges. The third and fourth pereopods are generally
490 characterized by sickle-shaped terminal segments apparently used to hold the prey (as seen in Fig. 3a,
491 b) and to direct food items towards the gnathopods (Nemato & Yoo, 1970) that tear apart pieces and
492 push them towards the mouthparts (Kane, 1963). *T. gaudichaudii* is capable of hooking onto larger
493 prey such as small fish (Kane, 1963) and euphausiids and to start feeding on their stomach content
494 while attached (Havermans C., unpublished data). The long and spiny fifth pereopods of *T.*
495 *gaudichaudii*, used for grasping prey, are supposedly linked to feeding on larger prey items (see below).

496 When comparing the mouthparts of *Themisto pacifica* to those of the gammarid-type pelagic predator
497 *Cyphocaris challengerii* Stebbing, 1888, Haro-Garay (2003) found that the mandibular palps of *T.*
498 *pacifica* appeared weaker and the toothed, more comb-like incisors indicated a less pronounced
499 predatory lifestyle suggesting a diet that combines microphagous and carnivorous feeding.

500 Investigating the functional anatomy of mouthparts as well as the alimentary canals may reveal more
501 about feeding habits than the actual gut content analyses regarding the prevalence of soft-bodied
502 zooplankton in the diet (Coleman, 1994). When comparing internal foreguts of gammarids and
503 hyperiids, Coleman (1994) noted an impressive variation in morphologies as well as several presumed
504 adaptations to handle larger food particles in the latter group, likely for feeding on larger gelatinous
505 zooplankton. Therefore, a comparative analysis of the mouthparts of the different *Themisto* species
506 may give insights into the importance of salps or other gelatinous zooplankton in their respective diets.
507 One caveat here is that it is at present unknown to what extent non-exclusive feeding on gelatinous

508 zooplankton requires adaptations of external and internal functional morphology of *Themisto* and
509 what those may look like.

510 Within several species, e.g. *Themisto gaudichaudii* (Schneppenheim and Weigmann-Haass, 1986) and
511 *T. compressa* (Stephensen, 1924; McHardy, 1970; Sheader, 1975), both “long-legged” and “short-
512 legged” morphs, differing in the length of the fifth pereopod, have been observed to occur in sympatry.
513 Experiments have shown that these different morphs arise depending on temperature and nutrition
514 (Sheader, 1975). Phylogeographic analyses have shown that these morphs are independent of the
515 different mitochondrial DNA lineages observed and that the ratio of the fifth versus the sixth pereopod
516 decreases with increasing latitude (and hence decreasing temperature) (Havermans C., unpublished
517 data). Within populations in the Southern Ocean, these two morphs have been linked to different
518 feeding strategies. Recent findings confirm that long-legged *bispinosa* morphotypes feed on a slightly
519 higher trophic level than short-legged *compressa* morphotypes and it is hypothesized that the length
520 of the pereopod plays a role in the efficiency with which bigger types of prey are caught (Kruse et al.,
521 2015).

522 **4.3. Herbivory? Grazing by juveniles and feeding on prey stomach contents by adults**

523 Some trophic studies of *Themisto gaudichaudii* based on stable isotope analyses of the pelagic food
524 web confirmed a high degree of omnivory (Gurney et al., 2001), whereas others place adults of this
525 species at similar trophic levels to herbivorous zooplankton (Stowasser et al., 2012). Gut content
526 analyses of *T. gaudichaudii* and *T. japonica* juveniles revealed significantly higher pigment
527 concentrations than in adults’ stomachs and hence juveniles are believed to feed substantially on
528 phytoplankton (Siegfried, 1965; Nemoto & Yoo, 1970; Hopkins, 1985; Sugisaki et al., 1991). In the
529 faecal pellets of *T. compressa* from the Northeast Atlantic, a marine snow signature was clearly
530 distinguished by Lampitt et al. (1993). In incubation experiments, *T. compressa* individuals also fed to
531 a great extent on aggregates (Lampitt et al., 1993). For *T. japonica*, results suggest that feeding
532 behaviour switches from herbivory to carnivory as they grow (Sugisaki et al., 1991). The time of

533 hatching of the juveniles also often matches with the onset of the spring bloom, e.g. for *T. libellula* in
534 the Arctic (Dalpadado, 2002). Nelson et al. (2001) revealed a source of phytoplankton present in the
535 fatty acid profiles of both juvenile and adult *T. gaudichaudii*. Large amounts of phytoplankton pigments
536 were also reported in the gut contents of adult *T. gaudichaudii*, however, it remains unclear whether
537 these were ingested by *Themisto* themselves or originated from digested prey (Pakhomov &
538 Perissinotto, 1996). Indeed, adults have been observed to feed preferably on stomach contents of
539 salps, euphausiids and conspecifics (Havermans et al., 2017; Figs. 3d, e, f).

540 **4.4. Explaining *Themisto*'s visits to the seafloor**

541 Various observations on migrations by adults to the deep-sea floor (depths around 1000 – 3000 m)
542 have been reported for the northern *Themisto* species, *T. abyssorum* and *T. libellula*. These were
543 explained by the animals feeding on detritus or phytoplankton (Vinogradov, 1999b and references
544 herein). Furthermore, in Svalbard waters, *T. libellula* seems to be the major food item of the Atlantic
545 spiny lump sucker (*Eumicrotremus spinosus* (Fabricius, 1776)), a slow-moving benthic fish that is
546 unlikely to prey upon fast-swimming migrating amphipods in the water column (Berge & Nahrgang,
547 2013). Apparently, *T. libellula* migrates to the bottom during the day where it aggregates, as was
548 observed by submersible imaging (Vinogradov, 1999b) and temporarily makes up a major component
549 of the hyperbenthos. This may also be the case for *T. gaudichaudii*, since individuals have been
550 collected by epibenthic sled catches at depths of more than 3000 m in the Polar Frontal Zones
551 (Havermans C., unpublished data). On the shelf around the Prince Edward Islands, *T. gaudichaudii* has
552 frequently been sampled with near bottom trawls (Pakhomov & Froneman, 1999). Also *T. japonica*
553 adults have been recorded at depths of 3000 m (Semenova, 1974). Hence, feeding by juveniles and
554 adults on phytoplankton in the water column or on the deep-sea floor should be further explored given
555 that it may have profound implications for pelago-benthic coupling processes and the biological pump.
556 Migrations to the seafloor can also stem from moulting and reproductive behaviour including the
557 release of juveniles by brooding females (see above).

558 **5. THEMISTO, DRESSED FOR SUCCESS?**

559 *Themisto*'s omnivorous and flexible feeding habits alone do not justify its abundance and status as the
560 most abundant of the pelagic amphipods found in temperate or high latitude oceans. Other hyperiids
561 seem to be equally voracious predators, for example, *Hyperoche medusarum* from the Pacific appears
562 to have a similar diet composition and raptorial behaviour as *T. gaudichaudii*, feeding on a variety of
563 mesozooplankton such as copepods, juvenile decapods, euphausiids, medusae and clupeid fish larvae.
564 The latter appears to be its preferred prey and it exerts a high predation pressure on newly hatched
565 herring larvae and hence herring stocks in British Columbia waters (von Westernhagen & Rosenthal,
566 1976). Why *Themisto* alone reaches these high biomass levels needs further consideration:

567 **5.1. A body fit for hunting and escaping?**

568 Studying morphological differences and similarities between zooplankton species, i.e. identifying
569 features retained from ancestors versus unique adaptations newly evolved within *Themisto*, is one way
570 to consider the influence of competition and the ability to colonize new niches, amongst other
571 processes. In the case of *Themisto*, one could argue that its morphology reflects a development
572 towards a shrimp-like morphotype. Within the genus, species bear a well-developed fan-like urosome
573 that reaches its maximum in *T. libellula* and *T. gaudichaudii* that could provide a tail-flip escape
574 response capability as seen in euphausiids and decapods. However, the urosome appears to function
575 more as armour, given that the amount of muscle tissue it contains appears insufficient to provide a
576 strong tail-flip. *Themisto* differs from all other hyperiids in that it bears many spines on the dorsum
577 and urosome (posterior part, in particular on the uropods), which could provide protection from
578 predation. When feeling threatened, *T. gaudichaudii* spreads its spiny uropods upwards, which may
579 indicate they serve as a primary defence apparatus (Fig. 4). Despite their armour, *Themisto* amphipods
580 are fast swimmers: swimming speeds of 30 cm.s⁻¹ have been measured for *T. japonica* (Hiroki, 1988).
581 Nonetheless, *Themisto* seems to escape less from predators compared to euphausiids and
582 chaetognaths (Volkov, 2012). Most other hyperiid amphipods lack conspicuous morphological

583 attributes such as spines which would ward off predators. For the many hyperiid species associated
584 with, and often residing inside, soft-bodied zooplankters, a smoother body surface facilitates the
585 interactions of juveniles or adults with their hosts. The dorsal spines on the back of *T. gaudichaudii* and
586 *T. australis* compared with the absence of spines in *T. abyssorum* and *T. pacifica* may reflect different
587 predator avoidance strategies or interactions with hosts. Variation in diet and predation pressure may
588 account for the intra-specific occurrence of morphotypes with and without dorsal spines (e.g. in *T.*
589 *gaudichaudii*, Havermans C., unpublished data).

590 **5.2. Adaptations to life in the mesopelagial**

591 The highly developed eyes of many hyperiidean species suggest selection for finding their transparent,
592 widely scattered prey. Nevertheless, it is often the case that one cannot see without being seen and
593 the large eyes of *Themisto* must be visible to predators. However common in pelagic animals (Buskey,
594 1992), bioluminescent properties have not yet been reported from *Themisto* amphipods, but its dark
595 coloration and opacity could reduce its detectability by predators in deeper waters, contrary to the
596 epipelagic waters. Pelagic taxa are transparent in shallower waters and become more opaque in the
597 deep, with colorations turning to uniform black (fish) or scarlet red (crustaceans) and with reduced
598 reflectance over the gut to mask their bioluminescent prey. Forms of *T. gaudichaudii* with different
599 pigments (from partly transparent to almost totally brown) have been discovered at sampling sites in
600 close proximity to each other (Havermans C., unpublished data). They may be linked to different stages
601 after moulting, turning darker with time. At hatching, juveniles appear to be almost completely
602 transparent except for pleonites covering the stomach region (which are light brown) and the eyes
603 (Havermans C., unpublished data), which may protect them from predation after release from the
604 brood pouch, after which they seem to remain in the surface layer.

605 **6. UP THE FOOD CHAIN: THE IMPORTANCE OF THEMISTO FOR HIGHER TROPHIC LEVELS**

606 **6.1. *Themisto* sustaining a variety of top consumers in polar and boreal food webs**

607 Environmental change has winners and losers and *Themisto* are considered to be the major
608 replacement of krill both in the Arctic (e.g. Dalpadado et al., 2001) and the Southwest Atlantic Ocean
609 (Padovani et al., 2012). A list of predators of the different *Themisto* species worldwide has been
610 compiled (Supplementary Material, Table S1), demonstrating their major importance as key species
611 for higher trophic levels such as fish, seabirds and marine mammals in boreal and polar food webs. In
612 the adjacent seas and gateways of the Arctic Ocean, *T. abyssorum* and *T. libellula* are the main prey of
613 birds, fish, whales and seals (Supplementary Material, Table S1). In the Bering Sea, *T. libellula* sustains
614 commercially important fish stocks such as the walleye pollock, the Pacific herring and cod, and the
615 most commonly exploited salmonid species (Fig. 5a). The species varies in abundance from year to
616 year due to natural climatic oscillations. In colder waters, it reaches enormous numbers, being a major
617 prey item for several of these fish species and impacting the entire food web structure (Volkov, 2012;
618 Pinchuk et al., 2013). Also on the Greenland shelf and in the Barents Sea, *T. libellula* is preyed upon by
619 fish stocks such as capelin, Atlantic cod and Greenland halibut (Fig. 5b). Similarly, *T. gaudichaudii* along
620 the Patagonian shelf and around the Falkland Islands has also been referred to as the krill of the
621 northern Southern Ocean and supports millions of tons of commercially exploited fish and squid
622 (Arkhipkin et al., 2012, Padovani et al., 2012, Arkhipkin, 2013) (Fig. 6a). Across the whole Southern
623 Ocean, *T. gaudichaudii* comprises a major share of the diet of at least 80 different species of squid,
624 fish, seabirds and marine mammals (Supplementary Material, Table S1, Fig. 6b).

625 **6.2. *Themisto* amphipods are not the preferred food of all predators**

626 Compared to euphausiids, hyperiid amphipods have a tough exoskeleton, which is reflected when
627 comparing the chitin content of *Euphausia superba* and *Themisto gaudichaudii* (Ikeda, 1974). This may
628 serve both for feeding by providing a stiff skeleton to exert strength, with the long pereopods serving
629 as lever arms to rip off pieces of soft-bodied plankters, as well as to deter predators which prefer to
630 feed on the “muscular” food chain and not the “armoured” one. The known predators of *T.*
631 *gaudichaudii* are summarized in Table S1. Certain species appear actively to avoid eating *Themisto*

632 when encountering them. For instance, although humpback (*Megaptera novaeangliae*, Clapham,
633 2002; Findlay et al., 2017) and fin whales (*Balaenoptera physalus*, Aguilar, 2002) do consume *T.*
634 *gaudichaudii*, other baleen whales do not, including blue whales and minke whales, which appear to
635 avoid *Themisto* swarms despite high abundances in their feeding grounds (Kawamura, 1994).
636 Observations from the *Discovery Reports* state that: “the whales caught at South Georgia (excluding
637 the Sperm Whale, *Physeter macrocephalus*) feed exclusively on *Euphausia superba* and have no other
638 food whatever in their stomachs apart from a few specimens of the amphipod *Euthemisto*, which is so
639 abundant in the plankton round South Georgia that the whales can hardly help swallowing a certain
640 quantity” (Mackintosh & Wheeler, 1929).

641 **7. IMPACTS OF ONGOING AND ANTICIPATED RANGE SHIFTS IN BOTH HEMISPHERES**

642 **7.1. Range shifts in the Arctic and adjacent oceans**

643 In the last decades, range shifts have been observed for *Themisto* species in the northern hemisphere
644 as a likely consequence of environmental changes. In the Fram Strait, the temperate species, *T.*
645 *compressa*, started to appear in high abundances in the long-term sediment trap record in 2004 and a
646 reproductive event in the region was first documented in 2011 (Kraft et al., 2012, 2013, Schröter et al.,
647 submitted). Furthermore, *T. abyssorum* has become more abundant whilst *T. libellula* has decreased,
648 both in the Barents Sea and Fram Strait (the Atlantic gateway to the Arctic, CAFF, 2017). Other – less
649 monitored regions – of the Arctic are likely undergoing similar changes. An expansion of the range of
650 *T. abyssorum* and a corresponding range contraction of *T. libellula* is very likely to be a manifestation
651 of the ongoing Atlantification of the Arctic with corresponding reduced levels of sea-ice (Overland &
652 Wang, 2013; Polyakov et al., 2017). *T. libellula* depends on the cryo-pelagic pathway involving ice algae
653 and herbivorous copepods (Auel et al., 2002; Kohlbach et al., 2016) and is likely to suffer from these
654 changes. In contrast, *T. abyssorum* and *T. compressa* may benefit due to their shorter life cycles and a
655 more varied diet (Auel et al. 2002; Kohlbach et al., 2016). These shifts in distributional range and
656 abundance may cause difficulties for higher trophic levels specializing on *T. libellula*, which is larger

657 than *T. abyssorum* and *T. compressa*. Top predators that rely on larger sized prey, such as little auks
658 (*Alle alle*) that specialize on feeding on only the largest *T. libellula* size class (Lønne & Gabrielsen, 1992),
659 face an uncertain future if climate change leads to a shift towards equally nutritive but smaller
660 *Themisto* in the near future.

661 Nevertheless, climatic shifts have also allowed *Themisto libellula* to inhabit new environments. For
662 instance, since the 1990s, *T. libellula* appeared in the Gulf and Estuary of St. Lawrence where it had
663 not been recorded before and it is now an abundant full-time resident of the system (Marion et al.,
664 2008). In the Bering Sea, *T. libellula* has periodically spread further south with the southward inflow of
665 colder northern waters, whilst in years characterized by a higher inflow of Pacific waters, the species
666 disappears again (Volkov, 2012). These changes impact trophic pathways in these waters: when *T.*
667 *libellula* is present, the major fish species (Pacific herring *Clupea pallasii*, Pacific cod *Gadus*
668 *macrocephalus* and juvenile pink, chum and sockeye salmon *Oncorhynchus* spp.) switch from piscivory
669 to planktivory (Pinchuk et al., 2013). In the southeastern Bering Sea shelf ecosystem, it is believed that
670 *T. libellula* was once an ever-present key component of the system but now is only present in cold
671 years (Pinchuk et al., 2013). Combined, these examples highlight the importance of these amphipods
672 to entire Arctic food webs and emphasise the need to predict their response to rapid ongoing changes
673 of the Arctic system.

674 **7.2. *Themisto* in the Southern Ocean**

675 *Themisto* is an opportunistic predator and may impact the abundances and recruitment of both krill
676 and salps through consumption of the smaller larval stages. For example, in waters around South
677 Georgia, *T. gaudichaudii* can consume up to 70% of daily secondary production (Pakhomov &
678 Perissinotto, 1996) – over 200 krill larvae per square metre per day – and by doing so, can significantly
679 influence the local recruitment of Antarctic krill (Tarling et al., 2007). Salps are also part of *Themisto*'s
680 diet, as demonstrated by Kruse et al. (2015) with stomach content analyses and by Havermans et al.

681 (2017) with experimental observations. Hence, changes in the distributional range and abundance of
682 *T. gaudichaudii* may represent a significant top-down control of other biomass dominant species.

683 Mackey et al. (2012) took a climatic envelope approach to considering historical and present-day
684 abundance distributions of a number of macrozooplankton species in the Atlantic sector of the
685 Southern Ocean, including that of *Themisto gaudichaudii*. Historical distributions were determined
686 from *Discovery Investigations* records (1925 – 1951), from which species-specific temperature-
687 envelope models were determined. Projections to the present day were made through assuming a 1°C
688 increase in upper water column temperature over the past 70 years (a conservative estimate based on
689 measured changes over that time by Meredith & King, 2005 and Whitehouse et al., 2008). From being
690 limited to mostly the northern sections of the Atlantic sector in the *Discovery* era, temperature
691 envelope projections predict *T. gaudichaudii* establishing itself even in the vicinity of the Antarctic
692 Peninsula. Indeed, recent macrozooplankton surveys at the Peninsula report *T. gaudichaudii* as a minor
693 component of the total catch (Ross et al., 2008, Loeb et al., 2009, Steinberg et al., 2015). A continued
694 upward trend in abundances of *T. gaudichaudii* in southern parts of the Southern Ocean may have
695 direct predatory impacts to both Antarctic krill and salps to go alongside the environmental pressures
696 on these biomass dominant species. The further impacts to krill-consuming higher predators may also
697 be profound. Even though some krill consumers can efficiently switch from a krill-based to an
698 amphipod-based diet in years of low krill availability (e.g. Macaroni penguins *Eudyptes chrysolophus*),
699 most species seem to be truly krill-dependent for their breeding success and even adult survival, which
700 was shown to be the case for the black-browed and grey-headed albatrosses (*Thalassarche* spp.), the
701 Gentoo penguin (*Pygoscelis papua*) and the Antarctic fur seal (*Arctocephalus gazella*) (e.g. Croxall, Reid
702 & Prince, 1999; Forcada et al., 2005). The limited palatability of *T. gaudichaudii* as a food source for
703 some baleen whale species (Kawamura, 1994) may also limit their population recovery in these
704 regions.

705 **7.3. Implications for the biological carbon pump and biogeochemical cycling**

706 Zooplankton may play an important role in the biological pump by the vertical flux of particulate
707 organic matter (POC) in the form of both faecal pellets, moults and dead bodies that sink to the ocean
708 floor (Turner, 2015). However, the faecal pellet production of pelagic amphipods remains largely
709 unstudied. Lampitt et al. (1993) studied the presence of marine snow derived from zooplankton faecal
710 pellets, focusing on *Themisto compressa* (a key species of the Northeast Atlantic). A large variety of
711 material in the pellets was noted from black, densely packed material to white, apparently empty
712 “ghost” pellets. Sinking rates were in the range of 108 and 215 m day⁻¹, depending on size (Lampitt et
713 al., 1993).

714 In our own observations of faecal pellet production by *Themisto libellula* from the Arctic, faecal pellets
715 of individuals that had been feeding on copepods consisted of rather loose and fluffy orange-coloured
716 material (Fig. 7). They easily fell apart and were prone to degradation by other organisms, in contrast
717 to the compact faecal pellets of copepods and euphausiids and the large fast-sinking pellets of salps.
718 In several instances, we observed *T. libellula* individuals feeding on their own faeces (coprophagy, Fig.
719 8). The faecal pellet produced was transferred from the urosome to the gnathopods after the individual
720 had swirled on its axis several times and then bent its body so that the faecal pellet could be grasped
721 by the feeding appendages directly from the posterior part of the body. This handling and ingestion of
722 own faecal material could explain why the pellets are very loose and occur in aggregates. The partial
723 degradation of pellets into smaller, slow-sinking pellets (called coprorhexy) will affect the vertical flux
724 as they can be more easily degraded by other organisms (Iversen & Poulsen, 2007). Also the fate of
725 limiting micronutrients (e.g. iron in the Southern Ocean) during the passage of food through the gut
726 remains unexplored in amphipods. As shown for Antarctic krill (Schmidt et al., 2016), the breaking
727 down and release of iron by grazers feeding on primary producers could be an important means of
728 sustaining productivity in iron deplete regions such as the Southern Ocean.

729 **8. Conclusions**

730 (1) We compiled existing knowledge on the distributions of the different *Themisto* species which
731 dominate boreal and polar pelagic ecosystems. In the northern hemisphere, boreal, sub-Arctic and
732 Arctic species often show overlapping distributions whereas, in the southern hemisphere, one species
733 i.e. *T. gaudichaudii*, dominates, while the distribution of *T. australis* is more restricted. There is a strong
734 relationship between the geographic limits of a number of *Themisto* species and water mass
735 boundaries such that *Themisto* species can be used as water mass indicators. *Themisto* species also
736 exhibit fluctuations in abundance as a consequences of natural climate oscillations, and are likely to
737 be impacted by ocean warming.

738 (2) At a smaller scale, *Themisto* species exhibit distinct diel vertical migration patterns; patterns vary
739 even within populations, with segregation according to sex and age being evident in both vertical and
740 horizontal scales. Nearshore areas may function as “nursery” areas, where females release their brood
741 from their brood pouch, as is evident in *T. libellula* and *T. gaudichaudii*. In contrast to other hyperiids,
742 *Themisto* species seem to be less dependent on soft-bodied zooplankton as hosts, but rather use them
743 as food, e.g. salps in the Southern Ocean. However, *T. pacifica* and *T. australis* use jellies as a holdfast
744 during several life stages.

745 (3) *Themisto* is not an exclusive carnivore as previously suggested, but feeds on a variety of food types,
746 displaying detritivorous, herbivorous and carnivorous feeding habits. Juveniles benefit from algal
747 blooms after their release, whereas adults also feed on algal fluff and detritus on the seafloor. This
748 previously underestimated flexibility further contributes to the ever-growing importance of *Themisto*
749 in a changing ocean. These hitherto unconsidered trophic links also change our view on the trophic
750 role of *Themisto* species, although further quantification of these feeding habits is now required.

751 (4) Through reviewing the available literature, we demonstrate that *Themisto* is a major trophic link in
752 boreal and polar food webs, sustaining a variety of predators, of which many are commercially
753 exploited. Both in the Arctic and Antarctic, many fish, bird and marine mammal species are dependent

754 on *Themisto* as prey. Nevertheless, it may not always be a preferred prey and its consumption as a
755 secondary prey item can indicate stress within regional food-webs.

756 (5) The distributional ranges of *Themisto* appears to be changing in line with environmental shifts. The
757 consequences of the replacement of the larger, lipid-rich *T. libellula* by the smaller and less nutritious
758 sub-Arctic and temperate species *T. abyssorum* and *T. compressa* in the Arctic will significantly impact
759 fish, whale and seabird populations. In the Southern Ocean, a poleward range expansion of *T.*
760 *gaudichaudii* will generate an overlap of distribution with Antarctic krill and salps which may impact
761 their levels of recruitment. However, the consequences remain hypothetical and will depend on the
762 feeding habits and prey preferences of *Themisto*. Hence, a better understanding of the biology of this
763 key pelagic group is crucial to predicting its future impact on food web structure, energy flow and
764 biogeochemical cycles.

765 **ACKNOWLEDGEMENT**

766 The first author is funded by the Deutsche Forschungsgemeinschaft (DFG, German Science Foundation)
767 with the projects HA 7627/1-1 and HA 7627/1-2 within the Priority Programme 1158 on Antarctic
768 Research with Comparable Investigations in Arctic Sea Ice Areas. GT was supported by the Ecosystems
769 and SCOOBIES programmes at BAS. Thanks to Laura Gerrish (MAGIC group at British Antarctic Survey)
770 for the generation of the distribution maps. We thank Prof. Victor Smetacek (AWI) for extensive
771 discussions on the topic. This publication is a result of the *Polarstern* Grant-Nos: AWI_PS103_03
772 (project “InterPelagic”), AWI-PS100_07 (project “AMICA”) and AWI_PS107_10.

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1104 **FIGURE LEGENDS**

1105 **Figure 1.** Occurrence maps of *Themisto* species from the northern hemisphere: *T. libellula* (Tli), *T.*
1106 *abyssorum* (Tab), *T. compressa* (Tco), *T. japonica* (Tja), *T. pacifica* (Tpa).

1107 **Figure 2.** Occurrence maps of *Themisto* species from the southern hemisphere: *T. gaudichaudii* (Tga)
1108 and *T. australis* (Tau).

1109 **Figure 3.** Documented feeding of a) *T. libellula* on an ostracod; b) on a pteropod (*Limacina*); c) *T.*
1110 *gaudichaudii*'s typical position when holding onto a salp or siphonophore; d) *T. gaudichaudii* specimens
1111 were observed at several instances to feed directly on the salp's stomach content; e) behavioural
1112 observations documented cannibalism of *T. gaudichaudii*: it was observed attacking other individuals
1113 of a similar size class and starting to feed on them and in particular the stomach region; f) When
1114 *Themisto* was placed in an aquarium together with large individuals of Antarctic krill (> 5 cm), it
1115 predominantly fed on the head, in particular the eyes, as well as on the stomach region.

1116 **Figure 4.** a) When feeling threatened, *T. gaudichaudii* spreads its uropods upwards, which is likely
1117 serving as a primary defence apparatus. b) Its spiny urosome, or posterior part offers protection from
1118 predation, and is a distinguishing character from other abundant hyperiid species.

1119 **Figure 5.** *Themisto libellula* as major prey item for a variety of top consumers, many of which are of
1120 commercial importance, in a) the Bering Sea and b) the Greenland and Barents seas.

1121 **Figure 6.** *Themisto gaudichaudii* as major prey item for a variety of top consumers, many of which are
1122 of commercial importance, in a) the Southwest Atlantic Ocean (Patagonian shelf and slope and
1123 Falkland Islands) and b) the Scotia Sea and Antarctic Peninsula region.

1124 **Figure 7.** Faecal pellets produced by an individual of *Themisto libellula*.

1125 **Figure 8.** The phenomenon of autocoprophagy was observed several times when keeping *T. libellula*
1126 in aquaria. A swirling movement was carried out to produce the faecal pellet (indicated with a black

1127 arrow) and subsequently transfer it from its position between the uropods to the mouthparts and
1128 gnathopods.

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1130 **SUPPORTING INFORMATION**

1131 **Table S1:** Importance of *Themisto* species as major prey in the diet of cephalopods, fish, birds and
1132 marine mammals. A non-exhaustive list compiled from literature. Predator species followed by an *
1133 represent a species at the basis of a (major) commercial fishery, raised for aquaculture, or an important
1134 bycatch species. Past fisheries are indicated by their time span between brackets. Abbreviations: NZ –
1135 New Zealand, SA – South Africa.

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1143 **Table 1.** Life-history characteristics of the different *Themisto* species from distinct geographic areas,
 1144 including the life cycle and the number of generations per year.

Species	Geographic distribution	Life cycle	Generations yr ⁻¹	References
<i>T. gaudichaudii</i>	Kerguelen Islands	-	one	Bocher et al., 2001, Labat et al., 2005
	0-20°E sector SO	1 year		Kane, 1966
	South Georgia region	-	two	Watts & Tarling, 2012
<i>T. libellula</i>	Off W coast South Africa	-	several	Siegfried, 1965
	Fram Strait – Greenland Sea	4 years	one	Kraft, 2010
	Northern and central Barents Sea	Up to 3 years	one	Dalpadado, 2002
	Eastern Barents Sea	2 years		Koszteyn et al., 1995
	Hudson Bay, SE Baffin Island	Up to 2 years		Dunbar, 1957
	NW Greenland Sea	2-3 years		Koszteyn et al., 1995
<i>T. abyssorum</i>	Greenland Sea, Fram Strait	At least 3 years		Auel & Werner, 2003
	Baffin Bay	1 year	-	Dunbar, 1946
	Western and southern Barents Sea	Up to 2 years		Bogorov, 1940; Koszteyn et al., 1995
	Northern and central Barents Sea	1 (-2) years	one	Dalpadado, 2002
	NW Greenland Sea	2 years	-	Koszteyn et al., 1995
<i>T. compressa</i>	Greenland, Norwegian, Barents seas	1 year	one-two	Koszteyn et al., 1995
	Gulf of St Lawrence	1 year	one	Hoffer, 1971
	North Sea		several	Sheader, 1981
<i>T. japonica</i>	Fram Strait – Greenland Sea	2 years	two	Kraft et al., 2013
	Japan Sea	195-593 days		Ikeda, 1990
<i>T. pacifica</i>	southern Japan Sea		three	Ikeda et al., 1992
	Western North Pacific	8.5 – 12 months	-	Yamada et al., 2004
	Western North Pacific		four	Yamada et al., 2004
	Southeastern Alaska		four-five	Wing et al., 1976

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Figure 1.

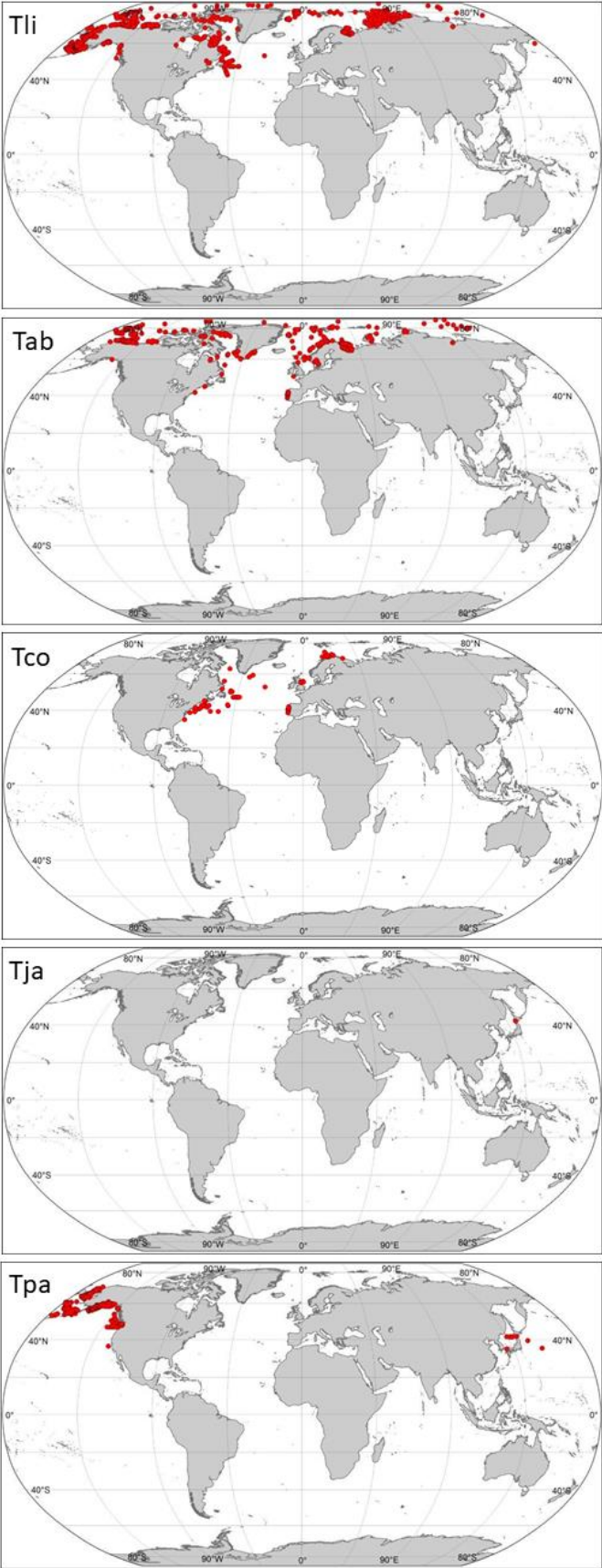


Figure 2.

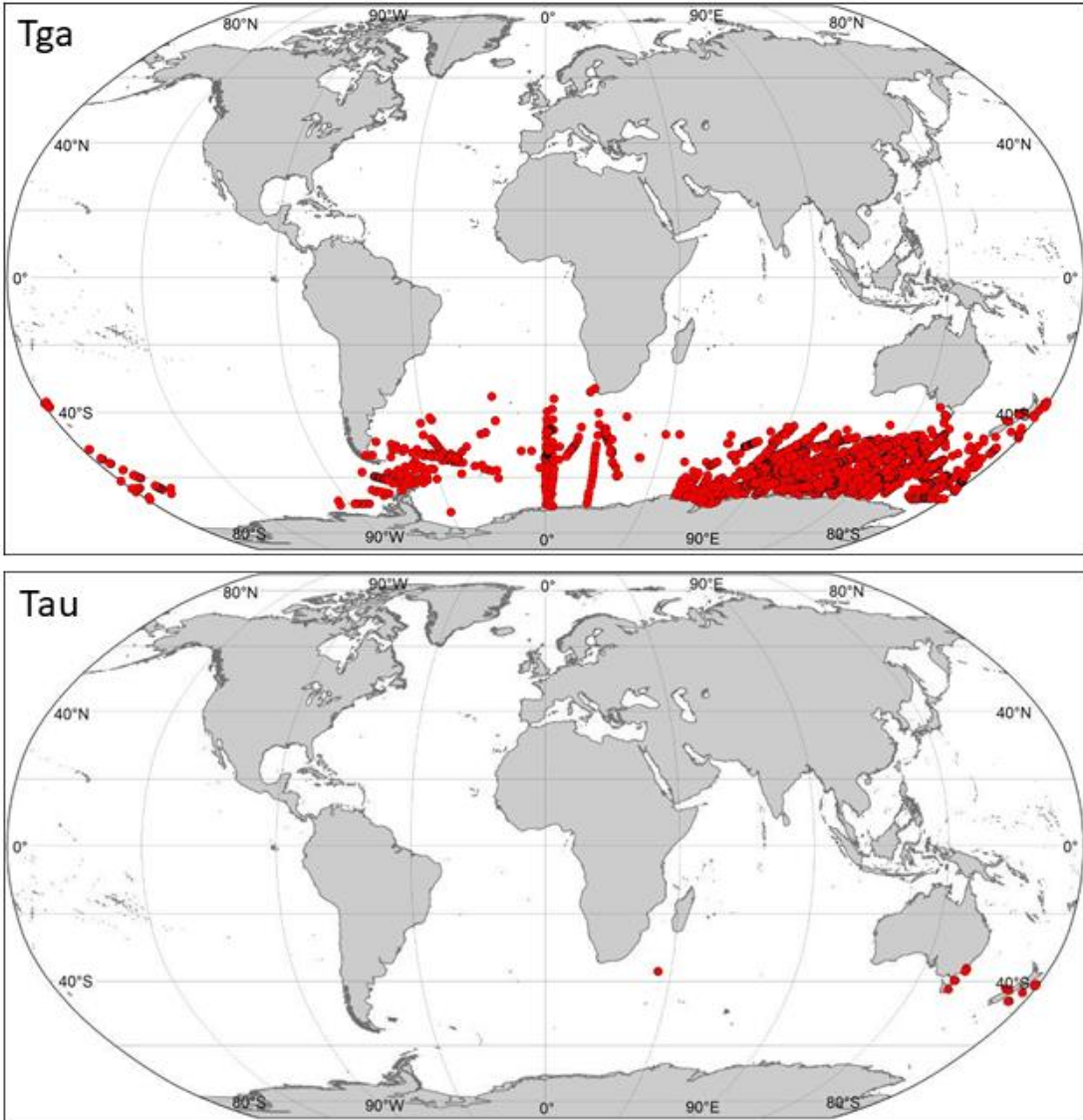


Figure 3.



Figure 4.



Figure 5.

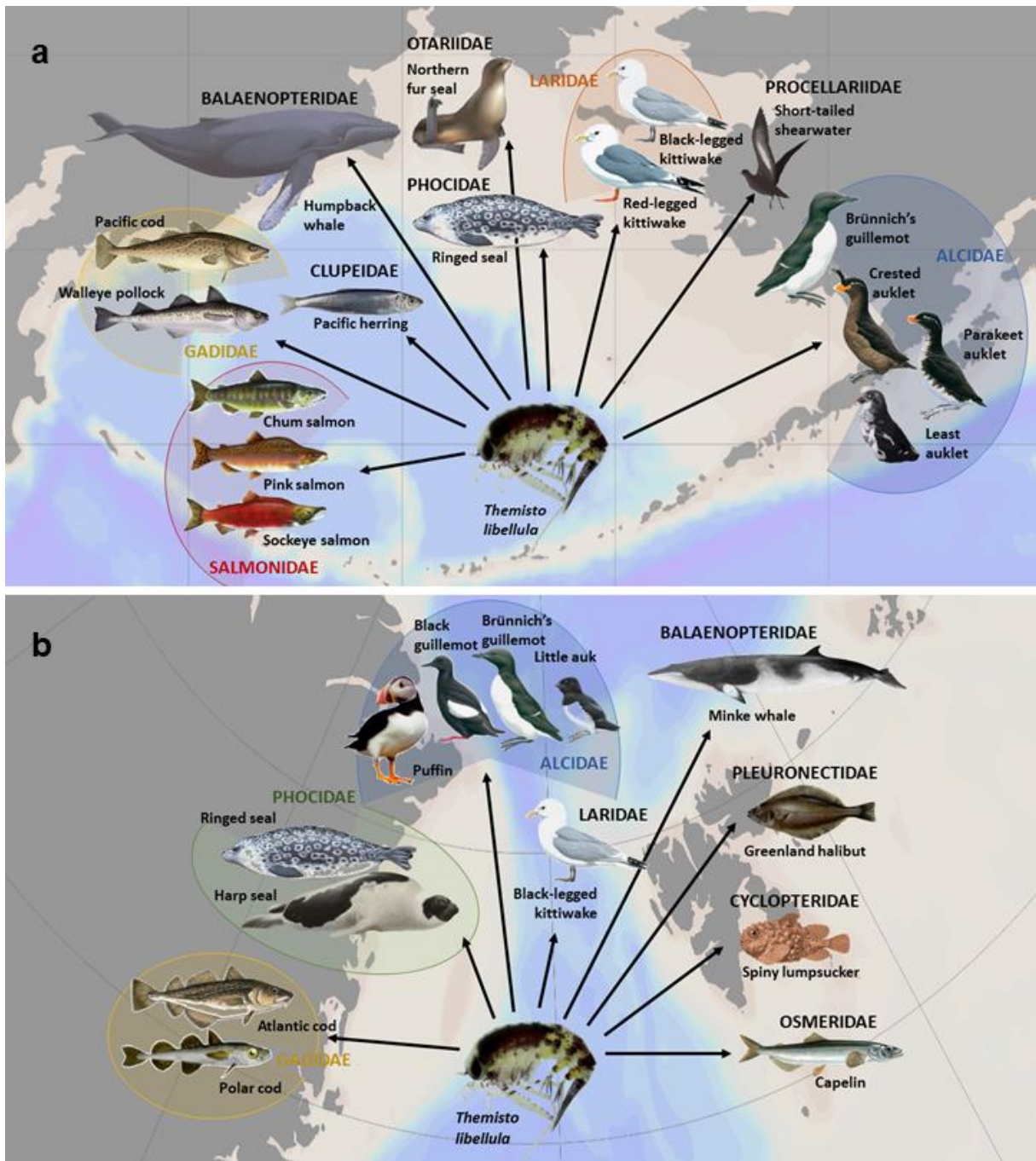


Figure 6.

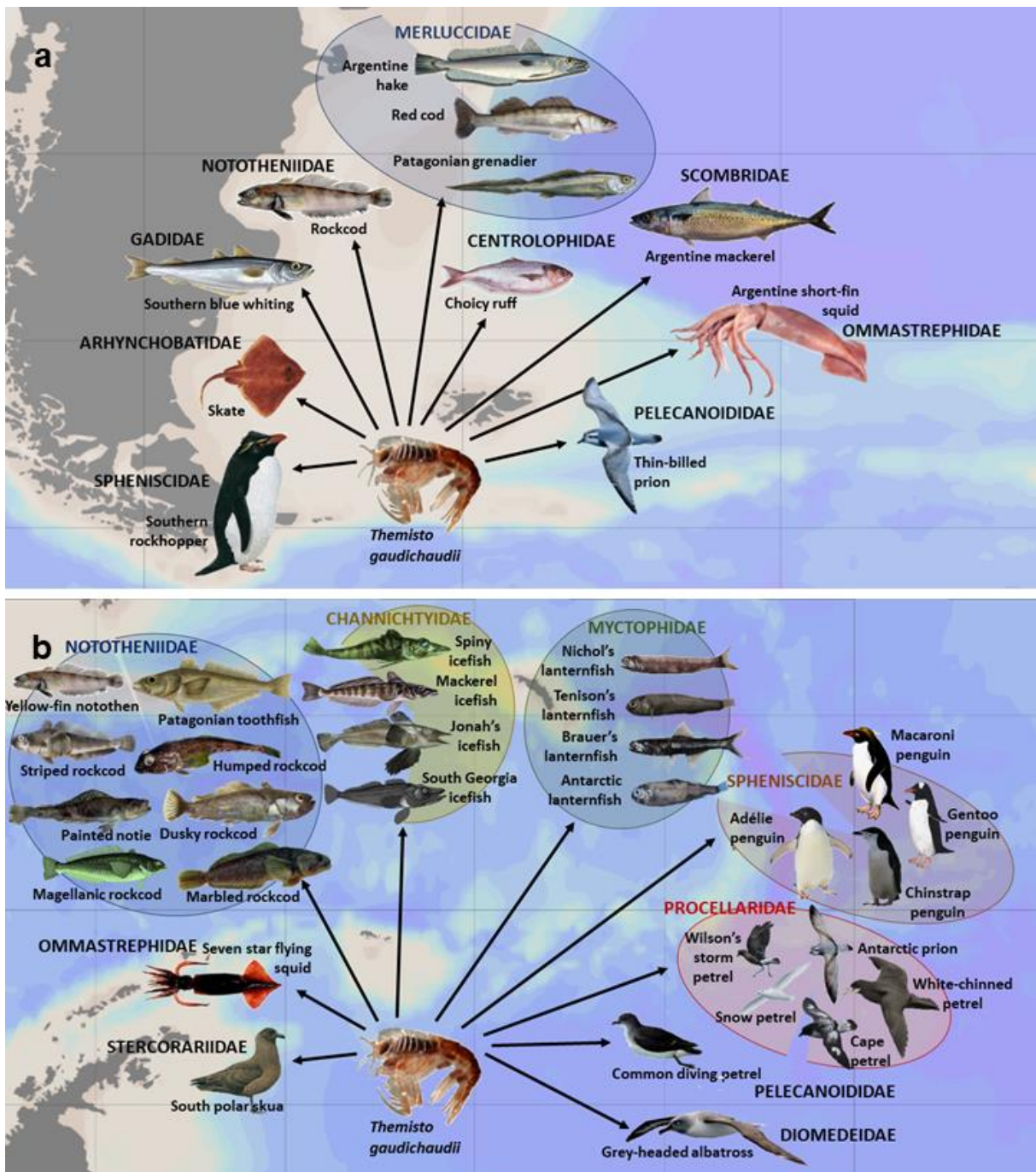
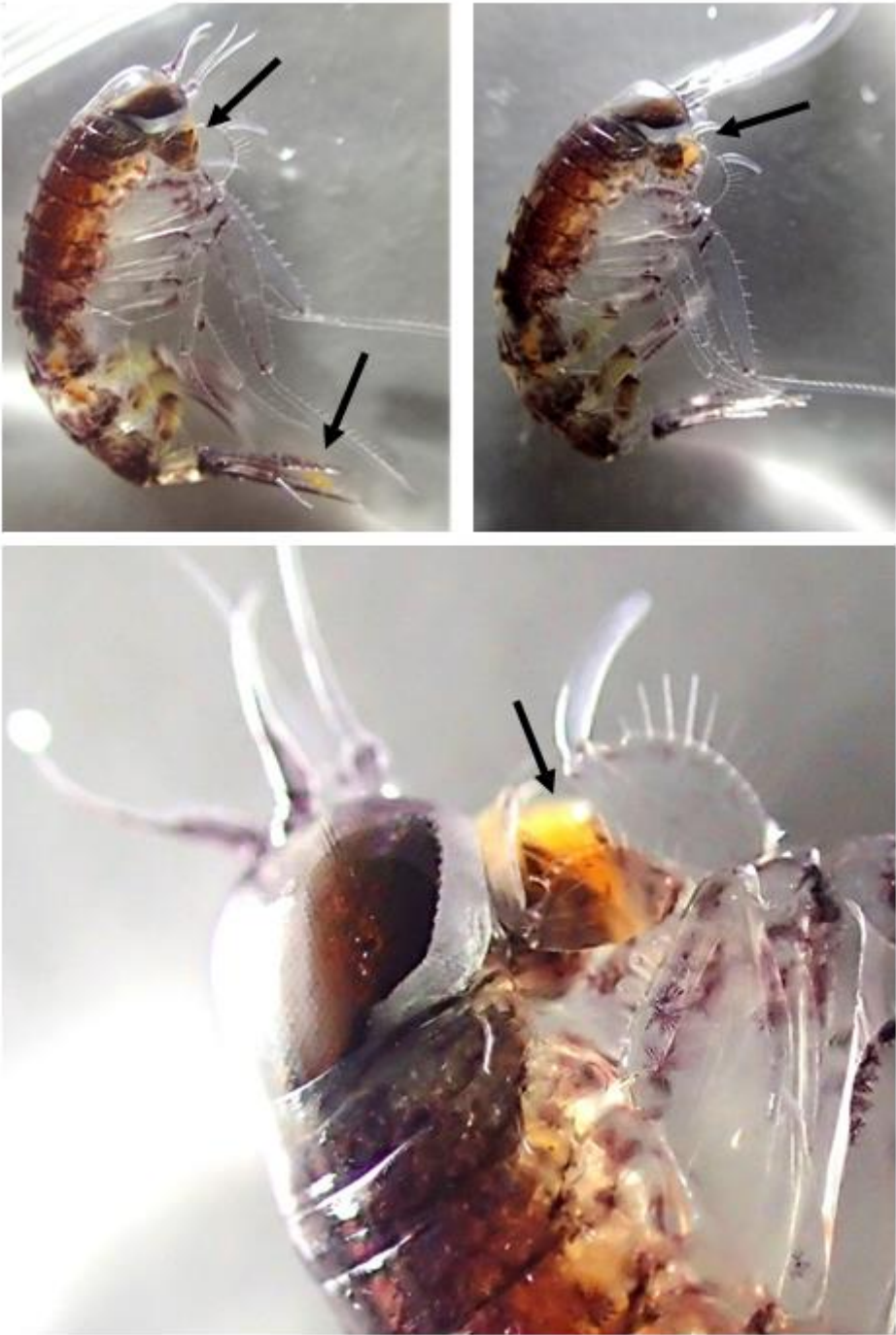


Figure 7.



Figure 8.



Predatory zooplankton on the move: *Themisto* amphipods in high-latitude pelagic food webs

Advances in Marine Biology

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Supporting Information

Table S1: Importance of *Themisto* species as major prey in the diet of cephalopods, fish, birds and marine mammals. A non-exhaustive list compiled from literature. Predator species followed by an * represent a species at the basis of a (major) commercial fishery, raised for aquaculture, or an important bycatch species. Past fisheries are indicated by their time span between brackets. Abbreviations: NZ – New Zealand, SA – South Africa.

Species	Region	References
<i>Themisto gaudichaudii</i>		
Cephalopods		
OMMASTREPHIDAE		
<i>Illex argentinus</i> (Argentine shortfin squid)*	Southern Patagonian Shelf	Ivanovic & Brunetti, 1994
<i>Illex argentinus</i> (Argentine shortfin squid)*	Falkland Islands	Mouat et al., 2001; Laptikovskiy, 2002
<i>Martialia hyadesi</i> (Seven star flying squid)*	South Georgia	Dickson et al., 2004
<i>Martialia hyadesi</i> (Seven star flying squid)*	NE Falkland Islands	González et al., 1997
CRANCHIIDAE		
<i>Galiteuthis glacialis</i>	South Shetlands	Nemoto et al., 1985
ONYCHOTEUTHIDAE		
<i>Onykia ingens</i> (Greater hooked squid)	Falkland Islands	Phillips et al., 2003
<i>Kondakovia longimana</i> (Giant warty squid)	Southern Ocean (90°-50°W)	Nemoto et al., 1985
Fish		
NOTOTHENIIDAE – Cod icefishes		
<i>Gobionotothen gibberifrons</i> (Humped rockcod)	South Georgia	Targett, 1981; Jażdżewski & Presler, 1988
<i>Gobionotothen gibberifrons</i> (Humped rockcod)	southern Scotia Sea	Rembiszewski et al., 1978
<i>Gobionotothen gibberifrons</i> (Humped rockcod)	South Orkneys	Targett, 1981
<i>Gobionotothen gibberifrons</i> (Humped rockcod)	Bransfield Strait	Jażdżewski & Presler, 1988
<i>Notothenia rossii</i> (Marbled rockcod)	South Georgia	Hoshiai, 1979; Jażdżewski & Presler, 1988
<i>Notothenia rossii</i> (Marbled rockcod)	Heard & McDonald Islands	Williams, 1983
<i>Notothenia coriiceps</i> (Black rockcod)	McDonald Island	Williams, 1963
<i>Notothenia microlepidota</i> (Small-scaled cod)	Campbell Plateau (NZ)	Clark, 1985
<i>Paranotothenia magellanica</i> (Magellanic rockcod)	Scotia Sea	Rembiszewski et al., 1978
<i>Lepidonotothen larseni</i> (Painted notie)	South Georgia	Targett, 1981; Jażdżewski & Presler, 1988
<i>Lepidonotothen larseni</i> (Painted notie)	South Orkneys	Targett, 1981
<i>Trematomus hansonii</i> (Striped rockcod)	South Georgia	Jażdżewski & Presler, 1988
<i>Trematomus lepidorhinus</i> (Slender scalyhead)	Western Ross Sea	Takahashi & Nemoto, 1984
<i>Trematomus newnesi</i> (Dusky rockcod)	South Orkneys	Targett, 1981
<i>Dissostichus eleginoides</i> (Patagonian toothfish)*	Shag Rocks & South Georgia	Collins et al., 2007
<i>Patagonotothen ramsayi</i> (Rock cod)*	Southern Patagonian Shelf	Padovani et al., 2012
<i>Patagonotothen ramsayi</i> (Rock cod)*	Falkland Islands	Laptikovskiy & Arkhipkin, 2003
<i>Patagonotothen guntheri</i> (Yellow-fin notothen)*(1978-1990)	Shag Rocks & South Georgia	Collins et al., 2008
<i>Pleuragramma antarctica</i> (Antarctic silverfish)	Ross Sea	Takahashi & Nemoto, 1984; La Mesa et al., 2004
CHANNICHTYIDAE – Icefishes		
<i>Chaenodraco wilsoni</i> (Spiny icefish)	Joinville Island	Kock et al., 2004
<i>Champocephalus gunnari</i> (Mackerel icefish)*(1975-1990)	Shag Rocks, South Georgia	Kock et al., 1994; Jażdżewski & Presler, 1988
<i>Neopagetopsis ionah</i> (Ionah's icefish)	Scotia Sea	Rembiszewski et al., 1978
<i>Pseudochaenichthys georgianus</i> (South Georgia icefish)	South Georgia	Clarke et al., 2008
MERLUCCIIDAE		
<i>Merluccius capensis</i> and <i>M. paradoxus</i> (Cape Hakes)*	Benguela Upwelling System	Pillar & Barange, 1997
<i>Merluccius hubbsi</i> (Argentine hake)*	Southern Patagonian Shelf	Padovani et al., 2012
<i>Merluccius hubbsi</i> (Argentine hake)*	Southern Atlantic Ocean	Angelescu & Cousseau, 1969
<i>Merluccius hubbsi</i> (Argentine hake)*	San Gorge Gulf	Temperoni et al., 2013
<i>Salilota australis</i> (Red Cod)*	Falkland Islands	Arkhipkin et al., 2001
<i>Macruronus magellanicus</i> (hoki, Patagonian grenadier)*	Southern Patagonian Shelf	Padovani et al., 2012
<i>Macruronus magellanicus</i> (hoki, Patagonian grenadier)*	Southwest Atlantic	Giussi et al., 2016
<i>Macruronus magellanicus</i> (hoki, Patagonian grenadier)*	Falklands Isl./Islas Malvinas	Brickle et al., 2009
<i>Macruronus novaezelandicae</i> (hoki)	Campbell Plateau (NZ)	Clark, 1985
MACROURIDAE – grenadiers or rattails		

<i>Lepidorhynchus denticulatus</i> (Javelin fish)	Campbell Plateau (NZ)	Clark, 1985
<i>Malacocephalus laevis</i> (Armed grenadier)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Coelorinchus braueri</i> (Shovelnose grenadier)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Coelorinchus matamua</i> (Mahia whiptail)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Coelorinchus simorhynchus</i>	W coast SA, Agulhas Bank	Anderson, 2005
<i>Coryphaenoides striaturus</i> (Striate whiptail)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Lucigadus ori</i> (Bronze whiptail)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Nezumia micronychodon</i> (Small-tooth grenadier)	W coast SA, Agulhas Bank	Anderson, 2005
<i>Nezumia umbracincta</i>	W coast SA, Agulhas Bank	Anderson, 2005
<i>Macrourus carinatus</i>	Southwest Atlantic	Giussi et al., 2010
ARHYNCHOBATIDAE		
<i>Bathyraja</i> spp. (Skates)*	Falkland Isl./Islas Malvinas	Brickle et al., 2003
ARGENTINIDAE		
<i>Argentina elongata</i> (Silverside)	Campbell Plateau (NZ)	Clark, 1985
CENTROLOPHIDAE - Medusafishes		
<i>Seriolella porosa</i> (Choicy ruff)*	Southern Patagonian Shelf	Padovani et al., 2012
GADIDAE – Codfishes		
<i>Micromesistius australis</i> (Southern blue whiting)*	Southern Patagonian Shelf	Padovani et al., 2012
<i>Micromesistius australis</i> (Southern blue whiting)*	Campbell Plateau (NZ)	Clark, 1985
<i>Micromesistius australis</i> (Southern blue whiting)*	Falkland Islands	Brickle et al., 2009
SCOMBRIDAE		
<i>Scomber japonicas marplatensis</i> (Argentine mackerel)*	Argentine Sea	Angelescu, 1979
<i>Allothunnus fallai</i> (Slender tuna)*	S Pacific, S Peru current	Yatsu, 1995
MYCTOPHIDAE – Lanternfishes		
<i>Electrona antarctica</i> (Antarctic lanternfish)	northern Scotia Sea	Shreeve et al., 2009
<i>Electrona antarctica</i> (Antarctic lanternfish)	South Georgia	Rowedder, 1979
<i>Metelectrona herwigii</i> (Herwig lanternfish)	Sub-Tropical Front	Pakhomov et al., 1996
<i>Protomyctophum choriodon</i> (Gaptooth lanternfish)	36-51°S	Pakhomov et al., 1996
<i>Gymnoscopelus braueri</i> (Brauer's lanternfish)	northern Scotia Sea	Jażdżewski & Presler, 1988; Shreeve et al., 2009
<i>Gymnoscopelus nicholsi</i> (Nichol's lanternfish)	Scotia Sea	Rembiszewski et al., 1978
<i>Gymnoscopelus nicholsi</i> (Nichol's lanternfish)	Sub-Tropical Front to APF	Pakhomov et al., 1996
<i>Protomyctophum tenisoni</i> (Tenison's lanternfish)	Scotia Sea	Rembiszewski et al., 1978
NOTACANTHIDAE		
<i>Notacanthus sexspinis</i> (Spiny-back eel)	W coast SA, Agulhas Bank	Anderson, 2005
OPHIDIIDAE – Cusk eels		
<i>Genypterus blacodes</i> (Pink cusk-eel)	Falkland Islands	Nyegaard et al., 2004
SERRANIDAE		
<i>Lepidoperca aurantia</i> (Orange Perch)	Chatman Rise, NZ	Horn et al., 2013
Birds		
SPHENISCIDAE – Penguins		
<i>Eudyptes chrysolophus</i> (Macaroni penguin)	South Georgia	Croxall et al., 1997, 1999; Waluda et al., 2012
<i>Eudyptes chrysolophus</i> (Macaroni penguin)	Crozet Islands	Ridoux, 1994
<i>Eudyptes chrysolophus</i> (Macaroni penguin)	Marion Island	Brown & Klages, 1987
<i>Pygoscelis adeliae</i> (Adélie penguin)	King George Island	Volkman et al., 1980; Jażdżewski, 1981
<i>Pygoscelis adeliae</i> (Adélie penguin)	Adélie Land	Ridoux & Offredo, 1989
<i>Pygoscelis adeliae</i> (Adélie penguin)	Laurie Island, S Orkneys	Libertelli et al., 2003
<i>Pygoscelis adeliae</i> (Adélie penguin)	Shirley Island, E Antarctica	Kent et al., 1998
<i>Pygoscelis antarcticus</i> (Chinstrap penguin)	King George Island	Volkman et al., 1980; Jażdżewski, 1981
<i>Pygoscelis papua</i> (Gentoo penguin)	Kerguelen Islands	Bost et al., 1994; Lescroëil et al., 2004
<i>Pygoscelis papua</i> (Gentoo penguin)	South Georgia	Williams, 1991; Xavier et al., 2018
<i>Pygoscelis papua</i> (Gentoo penguin)	Heard Island	Ealey, 1954; Green & Wong, 1992
<i>Pygoscelis papua</i> (Gentoo penguin)	Kerguelen Islands	Bost et al., 1994
<i>Eudyptes chrysocome chrysocome</i> (Southern rockhopper)	Tierra del Fuego	Schiavini & Raya Rey, 2004
<i>Eudyptes chrysocome</i> (Rockhopper penguin)	Kerguelen Islands	Bocher et al., 2001; Tremblay & Cherel, 2003
<i>Eudyptes chrysocome</i> (Rockhopper penguin)	Heard Island	Ealey, 1954
<i>Eudyptes chrysocome</i> (Rockhopper penguin)	Macquarie Island	Horne, 1985
<i>Eudyptes schlegelii</i> (Royal penguin)	Macquarie Island	Horne, 1985
STERCORARIIDAE – Skuas		
<i>Stercorarius maccormicki</i> (South polar skua)	Ant. Peninsula, S Scotia Sea	Reinhardt et al., 2000
PELECANOIDIDAE		
<i>Pelecanoides urinatrix</i> (Common diving petrel)	Kerguelen Islands	Bocher et al., 2000, 2001
<i>Pelecanoides urinatrix</i> (Common diving petrel)	Crozet Islands	Ridoux, 1994
<i>Pelecanoides urinatrix</i> (Common diving petrel)	South Georgia	Reid et al., 1997
PROCELLARIDAE		
<i>Pachyptila belcheri</i> (Thin-billed prion)	Falkland Islands	Quillfeldt et al., 2010, 2011
<i>Pachyptila belcheri</i> (Thin-billed prion)	Kerguelen Islands	Bocher et al., 2001; Cherel et al., 2002a
<i>Pachyptila desolata</i> (Antarctic prion)	Kerguelen Islands	Bocher et al., 2001; Cherel et al., 2002a
<i>Pachyptila desolata</i> (Antarctic prion)	South Georgia	Croxall et al., 1997, 1999
<i>Pachyptila salvini</i> (Salvin's prion)	Crozet Islands	Ridoux, 1994
<i>Pachyptila turtur</i> (Fairy prion)	Crozet Islands	Ridoux, 1994
<i>Pachyptila crassiostriis</i> (Fulmar prion)	Heard Island	Ealey, 1954
<i>Procellaria aequinoctialis</i> (White-chinned petrel)	Marion Island	Cooper et al., 1992
<i>Procellaria aequinoctialis</i> (White-chinned petrel)	Crozet Islands	Ridoux, 1994; Catard et al. 2000; Connan et al. 2007
<i>Procellaria aequinoctialis</i> (White-chinned petrel)	South Georgia	Berrow et al., 2000
<i>Pterodroma mollis</i> (Soft-plumaged petrel)	Crozet Islands	Ridoux, 1994
<i>Daption capense</i> (Cape petrel)	Crozet Islands	Ridoux, 1994
<i>Daption capense</i> (Cape petrel)	South Orkneys, Signy Isl.	Fijn et al., 2012

<i>Halobaena caerulea</i> (Blue petrel)	Crozet Islands	Ridoux, 1994
<i>Halobaena caerulea</i> (Blue petrel)	Kerguelen Islands	Bocher et al., 2001; Cherel et al., 2002b
<i>Halobaena caerulea</i> (Blue petrel)	Marion Island	Steele & Klages, 1986
<i>Puffinus tenuirostris</i> (Short-tailed shearwater)	Bruny Island, Tasmania	Weimerskirch & Cherel, 1998; Connan et al., 2010
<i>Fulmares glacialis</i> (Antarctic fulmar)	Adélie Land	Ridoux & Offredo, 1989
<i>Pagodroma nivea</i> (Snow petrel)	Adélie Land	Ridoux & Offredo, 1989
<i>Pagodroma nivea</i> (Snow petrel)	South Orkneys, Signy Isl.	Fijn et al., 2012
HYDROBATIDAE – Storm petrels		
<i>Oceanites oceanicus</i> (Wilson's storm petrel)	Crozet Islands	Ridoux, 1994
<i>Oceanites oceanicus</i> (Wilson's storm petrel)	South Georgia	Croxall et al., 1988
<i>Fregetta tropica</i> (Black-bellied storm petrel)	Crozet Islands	Ridoux, 1994
DIOMEDEIDAE – Albatrosses		
<i>Thalassarche chrysostoma</i> (Grey-headed albatross)	Crozet & Kerguelen Islands	Ridoux, 1994; Cherel et al., 2002c
<i>Thalassarche chrysostoma</i> (Grey-headed albatross)	South Georgia	Xavier et al., 2003
<i>Thalassarche chrysostoma</i> (Grey-headed albatross)	Marion Island	Connan et al., 2014
<i>Thalassarche melanophrys</i> (Black-browed albatross)	Kerguelen Islands	Cherel et al., 2000; 2002c
<i>Thalassarche melanophrys</i> (Black-browed albatross)	Diego Ramirez Isl, Chile	Arata & Xavier, 2003
<i>Phoebastria fusca</i> (Sooty albatross)	Marion Island	Cooper & Klages, 1995
<i>Phoebastria palpebrata</i> (Light-mantled sooty albatross)	Marion Island	Cooper & Klages, 1995
<i>Phoebastria palpebrata</i> (Light-mantled sooty albatross)	Macquarie Island	Green et al., 1998
Marine mammals		
PHOCIDAE – True seals		
<i>Mirounga leonina</i> (southern Elephant Island)	Macquarie Island	Green & Burton, 1993
OTARIIDAE – Eared seals		
<i>Arctocephalus gazella</i> (Antarctic fur seal)	Kerguelen Islands	Lea et al. 2002, 2008
BALAENOPTERIDAE – Rorquals		
<i>Balaenoptera borealis</i> (Sei whale)	Polar Frontal zone	Nemoto, 1970
<i>Balaenoptera borealis</i> (Sei whale)	Sub-Antarctic 170°W-170°E	Nemoto, 1962
<i>Balaenoptera borealis</i> (Sei whale)	Indian sector SO	Bottino, 1978
<i>Themisto libellula</i>		
Fish		
CLUPEIDAE		
<i>Clupea pallasii</i> (Pacific herring)*	SE Bering Sea	Pinchuk et al., 2013
GADIDAE – Codfishes		
<i>Gadus macrocephalus</i> (Pacific cod)*	SE Bering Sea	Pinchuk et al., 2013
<i>Gadus chalcogrammus</i> (Walleye pollock)*	Bering Sea	Yoshida, 1984; Pinchuk et al., 2013
<i>Boreogadus saida</i> (Polar cod)	W Barents Sea, Svalbard	Lønne & Gulliksen, 1989
<i>Boreogadus saida</i> (Polar cod)	Canadian Beaufort Sea	Majewski et al., 2016
<i>Boreogadus saida</i> (Polar cod)	NE Greenland	Christiansen et al., 2012
<i>Gadus morhua</i> (Atlantic cod)*	Barents Sea	Bogstad & Mehl, 1997
<i>Arctogadus glacialis</i> (Arctic cod)	NE Greenland (polynya)	Süfke et al., 1998; Christiansen et al., 2012
SALMONIDAE – Salmon		
<i>Oncorhynchus keta</i> (Chum salmon)*	Bering Sea	Pinchuk et al., 2013
<i>Oncorhynchus keta</i> (Chum salmon)*	Sea of Okhotsk	Karpenko et al., 2007
<i>Oncorhynchus nerka</i> (Sockeye salmon)*	Bering Sea	Pinchuk et al., 2013
<i>Oncorhynchus gorbuscha</i> (Pink salmon)*	Bering Sea	Pinchuk et al., 2013
<i>Oncorhynchus gorbuscha</i> (Pink salmon)*	Sea of Okhotsk	Karpenko et al., 2007
<i>Salvelinus alpinus</i> (Arctic char)*	Baffin Island	Moore & Moore, 1974
<i>Salmo salar</i> (Atlantic salmon)	Canadian high Arctic	Neilson & Gills, 1979
OSMERIDAE		
<i>Mallotus villosus</i> (Capelin)*	Barents Sea	Lund, 1981; Ajiad & Pushaeva, 1991
PLEURONECTIDAE – Righteye flounders		
<i>Reinhardtius hippoglossoides</i> (Greenland halibut)*	Greenland waters	Smidt, 1969; Haug et al. 1989; Michalsen et al., 1998
<i>Reinhardtius hippoglossoides</i> (Greenland halibut)*	Kara Sea	Dolgov & Benzik, 2017
CYCLOPTERIDAE		
<i>Eumicrotremus spinosus</i> (Spiny lump sucker)	Svalbard waters	Berge & Nahrgang, 2013
Birds		
ALCIDAE – Auks		
<i>Uria lomvia</i> (Brünnich's guillemot)	Gulf of Anadyr, N Bering Sea	Ogi & Hamanaka, 1982
<i>Uria lomvia</i> (Brünnich's guillemot)	E Bering Sea	Hunt et al., 1981
<i>Uria lomvia</i> (Brünnich's guillemot)	W Barents Sea (ice-covered)	Lønne & Gabrielsen, 1992
<i>Uria lomvia</i> (Brünnich's guillemot)	Svalbard, Barents Sea	Lydersen et al., 1989
<i>Cepphus grylle</i> (Black guillemot)	W Barents Sea (ice-covered)	Lønne & Gabrielsen, 1992
<i>Cepphus grylle</i> (Black guillemot)	Svalbard, Barents Sea	Lydersen et al., 1989
<i>Aethia pusilla</i> (Least auklet)	St Lawrence Island, Alaska	Bédard, 1969
<i>Aethia cristatella</i> (Crested auklet)	St Lawrence Island, Alaska	Bédard, 1969
<i>Aethia psittacula</i> (Parakeet auklet)	St Lawrence Island, Alaska	Bédard, 1969
<i>Alle alle</i> (Little Auk)	W Barents Sea (ice-covered)	Lønne & Gabrielsen, 1992
<i>Alle alle</i> (Little Auk)	W Barents Sea, Svalbard	Lydersen et al., 1989; Steen et al., 2007
<i>Alle alle</i> (Little Auk)	W Greenland	Pedersen & Falk, 2001
<i>Alle alle</i> (Little auk)	Bear Isl., Barents Sea	Weslawski et al., 1999
<i>Fratercula arctica</i> (Puffin)	W Barents Sea, Svalbard	Lydersen et al., 1989
PROCELLARIIDAE (puffins)		
<i>Puffinus tenuirostris</i> (Short-tailed shearwater)	NE Bering Sea shelf	Ogi et al., 1980
<i>Puffinus tenuirostris</i> (Short-tailed shearwater)	E Bering Sea	Hunt et al., 1981

LARIDAE		
<i>Rissa tridactyla</i> (Black-legged kittiwakes)	E Bering Sea	Sinclair et al., 2008
<i>Rissa tridactyla</i> (Black-legged kittiwakes)	Svalbard	Lydersen et al., 1989
<i>Rissa tridactyla</i> (Black-legged kittiwakes)	W Barents Sea (ice-covered)	Lønne & Gabrielsen, 1992
<i>Rissa brevirostris</i> (Red-legged kittiwakes)	E Bering Sea	Sinclair et al., 2008
Marine mammals		
PHOCIDAE – True seals		
<i>Phoca hispida</i> (Ringed seal)	Canadian High Arctic	Bradstreet & Finley, 1983
<i>Phoca hispida</i> (Ringed seal)	Point Barrow, Alaska	Lowry et al., 1978
<i>Phoca hispida</i> (Ringed seal)	Canadian Western Arctic	Smith, 1987
<i>Phoca hispida</i> (Ringed seal)	Canadian Eastern Arctic	Dunbar, 1941; McLaren, 1958
<i>Phoca hispida</i> (Ringed seal)	Svalbard fjord	Weslawski et al., 1994
<i>Phoca hispida</i> (Ringed seal)	NW Greenland	Vibe, 1950
<i>Phoca hispida</i> (Ringed seal)	Kara Sea	Chapskii, 1940
<i>Phoca hispida</i> (Ringed seal)	Chukchi Peninsula	Fedoseev, 1976
<i>Phoca hispida</i> (Ringed seal)	Alaskan and Canadian Arctic	Dehn et al., 2007
<i>Phoca hispida</i> (Ringed seal)	Bering Sea	Lowry et al., 1982
<i>Phoca hispida</i> (Ringed seal)	Barents Sea	Whatne et al., 2000
<i>Erignathus barbatus</i> (Bearded seal)	Alaskan and Canadian Arctic	Dehn et al., 2007
<i>Phoca largha</i> (Spotted seal)	Alaskan and Canadian Arctic	Dehn et al., 2007
<i>Pagophilus groenlandicus</i> (Harp seal)	Barents Sea	Lydersen et al., 1991, Nilsen et al. 1991,1992, 1995
<i>Pagophilus groenlandicus</i> (Harp seal)	NW, central W Greenland	Finley et al., 1990; Kapel, 2000
<i>Pagophilus groenlandicus</i> (Harp seal)	Greenland Sea, E Greenland	Enoksen, 2014
OTARIIDAE – Eared seals		
<i>Callorhinus ursinus</i> (Northern fur seal)	E Bering Sea Shelf	Harry & Hartley, 1981
BALAENOPTERIDAE – Rorquals		
<i>Megaptera novaeangliae</i> (Humpback whale)	Bering Strait, Chukchi Sea	Tomilin, 1957
<i>Balaenoptera acutorostrata</i> (Minke whale)	Barents Sea	Haug et al., 1993
<i>Balaena mysticetus</i> (Bowhead whale)	Beaufort Sea	Lowry & Frost, 1984; Lowry et al., 2004
<i>Themisto abyssorum</i>		
Squid		
OMMASTREPHIDAE		
<i>Todarodes sagittatus</i> (European flying squid)*	North Norwegian waters	Breiby & Jobling, 1985
Fish		
OSMERIDAE		
<i>Mallotus villosus</i> (Capelin)*	Barents Sea	Lund, 1981; Ajiad & Pushaeva, 1991
GADIDAE – Codfishes		
<i>Gadus morhua</i> (Atlantic cod)*	Barents Sea	Bogstad & Mehl, 1997
<i>Arctogadus glacialis</i> (Arctic cod)	NE Greenland (polynya)	Süfke et al., 1998; Christiansen et al., 2012
<i>Boreogadus saida</i> (Polar cod)	NE Greenland fjords	Christiansen et al., 2012
<i>Boreogadus saida</i> (Polar cod)	Canadian Beaufort Sea	Majewski et al., 2016
MYCTOPHIDAE – Lanternfishes		
<i>Lampanyctus macdonaldi</i> (MacDonald's lanternfish)	Greenland Sea	Klimpel et al., 2006
Birds		
ALCIDAE – Auks		
<i>Alle alle</i> (Little auk)	W Spitsbergen fjord	Steen et al., 2007
<i>Alle alle</i> (Little auk)	Bear Isl., Barents Sea	Weslawski et al., 1999
<i>Themisto australis</i>		
Fish		
MACROURIDAE – grenadiers or rattails		
<i>Lepidorhynchus denticulatus</i> (Javelin fish)	Chatman Rise, NZ	Stevens & Dunn, 2011
<i>Coelorhynchus oliverianus</i> (Hawknose grenadier)	Chatman Rise, NZ	Stevens & Dunn, 2011
Birds		
PELECANOIDIDAE		
<i>Pelecanoides urinatrix urinatrix</i> (Common diving petrel)	SE Australia	Schumann et al., 2008
PROCELLARIIDAE		
<i>Puffinus tenuirostris</i> (Short-tailed shearwater)	Bruny Island, Tasmania	Weimerskirch & Chereil, 1998
<i>Puffinus griseus</i> (Sooty shearwater)	New Zealand	Cruz et al., 2001
<i>Themisto pacifica</i>		
Squid		
OMMASTREPHIDAE		
<i>Todarodes pacificus</i> (Japanese common squid)*	Sea of Japan	Okiyama, 1965; Uchikawa & Kidokoro, 2014
GONATIDAE		
<i>Berryteuthis anonychus</i> (Minimal armhook squid)	NE Pacific	Uchikawa et al., 2004
Fish		
SALMONIDAE – Salmon		
<i>Oncorhynchus kisutch</i> (Coho salmon)*	Northern California current	Brodeur, 1989; Brodeur et al., 2013
<i>Oncorhynchus mykiss</i> (Steelhead salmon)	Northern California current	Brodeur et al., 2013
<i>Oncorhynchus gorbuscha</i> (Pink salmon)*	Offshore Japan Sea	Fukataki, 1967
<i>Oncorhynchus masou</i> (Masu salmon)*	Offshore Japan Sea	Fukataki, 1969
<i>Oncorhynchus</i> spp.	Central Gulf of Alaska	Kaeriyama et al., 2004
<i>Oncorhynchus keta</i> (Chum salmon)*	Aleutian Basin	Kosenok & Naidenko, 2008

<i>Oncorhynchus keta</i> (Chum salmon)*	Central Bering sea	Sakai et al., 2012
GADIDAE – Codfishes		
<i>Gadus macrocephalus</i> (Pacific cod)*	Eastern coast N Japan	Yamamura, 1993
<i>Gadus chalcogrammus</i> (Alaska/Walleye pollock)*	Off Iwate, E Japan	Fujita et al., 1995
<i>Gadus chalcogrammus</i> (Alaska/Walleye pollock)*	Gulf of Alaska	Brodeur, 1998
<i>Gadus chalcogrammus</i> (Alaska/Walleye pollock)*	Northern Japan	Yamamura et al., 2002
MACROURIDAE – Grenadiers		
<i>Albatrossia pectoralis</i> (Giant grenadier)	NW Pacific	Chuchukalo & Napazakov, 2012
MYCTOPHIDAE – Lanternfishes		
<i>Stenobrachius leucopsarus</i> (Northern lanternfish)	Central Bering Sea	Tanimata et al., 2008
<i>Stenobrachius leucopsarus</i> (Northern lanternfish)	Western North Pacific	Moku et al., 2000
<i>Stenobrachius leucopsarus</i> (Northern lanternfish)	Northern California current	Suntsov & Brodeur, 2008
<i>Tarletonbeania crenularis</i>	Northern California current	Suntsov & Brodeur, 2008
<i>Diaphus theta</i> (California headlightfish)	Western North Pacific	Moku et al., 2000
<i>Diaphus theta</i> (California headlightfish)	Northern California current	Suntsov & Brodeur, 2008
SEBASTIDAE – Rockfishes		
<i>Sebastes entomelas</i> (Widow rockfish)	Off Oregon	Bosley et al., 2014
<i>Sebastes flavidus</i> (Yellowtail rockfish)	Off Oregon	Bosley et al., 2014
<i>Sebastes crameri</i> (Darkblotched rockfish)	Off Oregon	Bosley et al., 2014
GONOSTOMATIDAE		
<i>Sigmops gracilipes</i>	NW North Pacific	Uchikawa et al., 2001b
BATHYLAGIDAE		
<i>Leuroglossus schmidti</i> (Northern smoothtongue)	Sea of Okhotsk	Beamish et al., 1999
<i>Themisto japonica</i>		
Fish		
SALMONIDAE – Salmon		
<i>Oncorhynchus</i> spp.	Central Gulf of Alaska	Kaeriyama et al., 2004
<i>Oncorhynchus keta</i> (Chum salmon)*	Sea of Okhotsk	Karpenko et al., 2007
<i>Oncorhynchus keta</i> (Chum salmon)*	Bering Sea	Karpenko & Koval, 2007
<i>Oncorhynchus gorbuscha</i> (Pink salmon)*	Sea of Okhotsk	Karpenko et al., 2007
STERNOPTYCHIDAE		
<i>Maurollicus japonicus</i> (North Pacific lightfish)	Pacific coast N Japan	Uchikawa et al., 2001a
GADIDAE – Codfishes		
<i>Gadus chalcogrammus</i> (Alaska/Walleye pollock)*	Northern Japan	Yamamura et al., 2002
<i>Gadus macrocephalus</i> (Pacific cod)*	Eastern coast N Japan	Yamamura, 1993
GONOSTOMATIDAE		
<i>Sigmops gracilipes</i>	NW North Pacific	Uchikawa et al., 2001b
MYCTOPHIDAE – Lanternfishes		
<i>Stenobrachius leucopsarus</i> (Northern lanternfish)	Western North Pacific	Moku et al., 2000
<i>Diaphus theta</i> (California headlightfish)	Western North Pacific	Moku et al., 2000
CYCLOPTERIDAE		
<i>Eumicrotremus asperrimus</i> (Spiny lumpfish)	NW Sea of Japan	Antonenko et al., 2008
HEMITRIPTERIDAE		
<i>Blepsias cirrhosis</i> (Silverspotted sculpin)	Sea of Okhotsk	Kolpakov & Dolganova, 2006
PLEURONECTIDAE – Righteye flounders		
<i>Hippoglossoides dubius</i> (Flathead flounder)	Japan coast	Kimura et al., 2004
<i>Themisto</i> sp./spp.		
Fish		
PLEURONECTIDAE		
<i>Reinhardtius hippoglossoides</i> (Greenland halibut)*	West Greenland	Pedersen & Riget, 1993
<i>Reinhardtius hippoglossoides</i> (Greenland halibut)*	Barents Sea	Nizovtsev, 1991
SEBASTIDAE – Rockfishes		
<i>Sebastes</i> spp. (Redfish)	West Greenland	Pedersen & Riget, 1993
HEXAGRAMMIDAE		
<i>Pleurogrammus monopterygius</i> (Atka mackerel)	Aleutian Islands	Yang, 1999
SQUALIDAE		
<i>Squalus acanthias</i> (Spiny dogfish)	South Island, NZ	Hanchet, 1991
CLUPEIDAE		
<i>Clupea harengus</i> (Atlantic herring)*	Western Norwegian Sea	Dalpadado et al., 2000
MYCTOPHIDAE – Lanternfishes		
<i>Lampanyctus macdonaldi</i> (MacDonald's lanternfish)	Greenland Sea, Irminger Sea	Kimpel et al., 2006
SCIAENIDAE – Croakers		
<i>Larimichthys polyactis</i> (Small yellow croaker)*	Central Yellow Sea	Xue et al., 2005

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