# Prediction of the Coding Sequences of Unidentified Human Genes. III. The Coding Sequences of 40 New Genes (KIAA0081-KIAA0120) Deduced by Analysis of cDNA Clones from Human Cell Line KG-1 

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#### Abstract

We isolated full-length cDNA clones from size-fractionated cDNA libraries of human immature myeloid cell line KG-1, and the coding sequences of 40 genes were newly predicted. A computer search of the GenBank/EMBL databases indicated that the sequences of 14 genes were unrelated to any reported genes, while the remaining 26 genes carried some sequences with similarities to known genes. Significant transmembrane domains were identified in 17 genes, and protein motifs that matched those in the PROSITE motif database were identified in 11 genes. Northern hybridization analysis with 18 different cells and tissues demonstrated that 10 genes were apparently expressed in a cell-specific or tissue-specific manner. Among the genes predicted, half were isolated from the medium-sized cDNA library and the other half from the small-sized cDNA library, and their average sizes were 4 kb and 1.4 kb , respectively. As judged by Northern hybridization profiles, small-sized cDNAs appeared to be expressed more ubiquitously and abundantly in various tissues, compared with that of medium-sized cDNAs.


Key words: full-length cDNA sequence; unidentified human gene; protein motif; mRNA expression; chromosomal location; myeloid cell line KG-1

## 1. Introduction

To accumulate information on the structure of unidentified human genes, we have begun a sequencing project of full-length cDNA clones of human cells. ${ }^{1}$ The cell source used was human immature myeloid cell line KG1. As to the sequencing strategy, size-fractionated cDNA libraries were constructed, from which cDNA clones that carry unreported sequences were first isolated. Then, the sizes of the mRNA corresponding to these clones were analyzed by Northern hybridization, and the entire nucleotide sequences of clones that comprised nearly fulllength transcripts were deduced. ${ }^{1}$ By using this strategy, we have already predicted the coding sequences of 80 new genes. ${ }^{1,2}$ In this paper, we report the coding sequences of an additional 40 genes newly determined and their sequence features as well as expression profiles.

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## 2. Materials and Methods

The source of cDNA libraries and the methods used for sequence analysis, chromosomal mapping of cDNA clones and computer analysis of sequences are identical to those used in the previous study. ${ }^{1}$ Expression profiles in human tissues were examined using human multiple tissue Northern blots (Human MTN blots) from Clontech (California, USA).

## 3. Results and Discussion

### 3.1. Sequence features of analyzed $c D N A$ clones

The clones that carried inserts longer than 2 kb and those that retained $1-2 \mathrm{~kb}$ inserts were randomly selected from the libraries constructed from the mediumand small-sized cDNA classes, respectively. After the $5^{\prime}$-terminal sequences of the inserts were determined by single-run sequencing, similarities of the sequences were searched using the FASTA program, and the clones which showed no significant similarities to reported sequences were selected. The sizes of their inserts were then compared with those of corresponding transcripts by Northern hybridization, and the clones that carried inserts


Figure 1. Physical maps of the 40 cDNA clones analyzed. The horizontal scale represents the cDNA length in kb, and gene numbers are given on the left. Open reading frames (ORFs) within coding regions, untranslated regions and repetitive sequences are indicated by solid, open and dotted boxes, respectively. The positions of the first ATG codon in each ORF are represented by triangles. The repetitive sequences observed are shown by hatched boxes. The major restriction sites are indicated above the sequences by the following abbreviations: B, BamHI; E, EcoRI; H, HindIII; S, SalI. The nucleotide sequence data reported in this paper were deposited in the GSDB, DDBJ, EMBL and NCBI nucleotide sequence databases under the accession numbers shown in Table 3.

Table 1. cDNA clones with similarities to the Genbank/EMBL database files.

| Gene no. <br> (KIAA) | Database files | Accession no. ${ }^{\text {a) }}$ | Identity <br> (\%) | Overlap ${ }^{\text {b) }}$ <br> (amino acid <br> residues) |
| :--- | :--- | :--- | :---: | :---: |
| 0082 | ORF69 (ACNPV) | L22858 | 30.5 | 174 |
| 0083 | Chromosome III Cosmid 9986 (Sc) | U00027 | 41.0 | 458 |
| 0085 | KIAA0108 (H) | D14696 | 31.3 | 261 |
| 0086 | DNA repair protein SNM1 gene (Sc) | X64004 | 33.3 | 192 |
| 0088 | Glucan 1,4- $\alpha$-glucosidase (Sc) | Z36098 | 41.5 | 590 |
| 0089 | Glycerol-3-phosphate dehydrogenase (H) | L34041 | 71.8 | 344 |
| 0090 | Hypothetical protein YCL045C (Sc) | S19374 | 29.8 | 289 |
| 0091 | Subtilisin (Ba) | A00970 | 30.4 | 303 |
| 0092 | Smooth muscle myosin (H) | X69292 | 17.3 | 405 |
| 0093 | NEDD4 gene (M) | D10714 | 84.4 | 488 |
| 0094 | Methionine aminopeptidase 1 (Sc) | M77092 | 51.8 | 369 |
| 0095 | Nucleoporin-interacting protein NIC96 gene (Sc) | X72923 | 22.4 | 840 |
| 0096 | Protein kinase (M) | U11494 | 28.9 | 166 |
| 0098 | Chaperonin containing TCP-1 (M) | Z31555 | 95.8 | 546 |
| 0099 | Pumilio protein (D) | L07943 | 42.1 | 875 |
| 0100 | el protein (M) | X81632 | 95.3 | 1182 |
| 0102 | Signal peptidase complex SPC 25 (Do) | U12687 | 95.1 | 225 |
| 0106 | B15C gene (Hv) | X76605 | 48.9 | 231 |
| 0108 | KIAA0085 (H) | D42042 | 31.3 | 261 |
| 0109 | Clathrin-associated protein (Ce) | L26290 | 81.0 | 253 |
| 0111 | Translation initiation factor nuk34 (H) | X79538 | 99.5 | 411 |
| 0115 | Oligosaccharyltransferase 48 kDa subunit (Do) | M98392 | 95.7 | 443 |
| 0116 | 75 kDa autoantigen (H) | M58460 | 26.7 | 225 |
| 0118 | Rab B (Dd) | L21012 | 59.0 | 161 |
| 0119 | Chlordecone reductase (H) | S68288 | 99.4 | 323 |
| 0120 | Neuronal protein NP25 (R) | M84725 | 69.7 | 195 |

ACNPV, Autographa california nuclear polyhedrosis virus; Ba, Bacillus amyloliquifacience; C, chicken; Ce, Caenorhabditis elegance; D, Drosophila melanogaster, Dd, Dictyostelium discoideum; Do, dog; H, human; Hv, Hordeum vulgare; M, mouse; R, rat; Sc, Saccharomyces cerevisiae.
${ }^{\text {a) }}$ Genbank/EMBL database files are shown except KIAA0090. ${ }^{\text {b) }}$ The size of regions which show similarities. ${ }^{\text {c) }}$ PIR database file.
which were more than $90 \%$ of the length of the corresponding transcripts were selected and subjected to sequence analysis. After the sequence determination, the integrity of the clones was analyzed by Northern hybridization, ${ }^{1}$ and the coding sequences of 40 genes were newly predicted. Among the genes predicted, half belonged to the medium-sized cDNA class and the other half to the small-sized cDNA class, and their average sizes were roughly 4 kb and 1.4 kb , respectively.

In Fig. 1, open reading frames are indicated by solid boxes and the first ATG codon by open triangles above the solid boxes. In-frame termination codons upstream of the first ATG codon were identified in 9 clones in the medium-sized cDNA class and 6 clones in the small-sized cDNA class. It is therefore likely that at least $40 \%$ of the clones analyzed harbor the complete coding region. As can be seen in the patterns in Fig. 1, the medium-sized
cDNAs retain relatively long stretches of $3^{\prime}$-untranslated regions (UTRs). The biological significance of these long UTRs remains to be elucidated.

Computer analysis of the sequences was carried out by using the GCG software package. The results are shown in Tables 1 and 2 and also in the figures in the Supplement section. Sequence features noted are summarized as follows:

1. Sequences of 14 genes were unrelated to any reported genes in the GenBank/EMBL database files, while the remaining 26 genes carried some sequences with similarities to known genes (Table 1).
2. Protein motifs that matched those in the PROSITE motif database were found in 11 genes (Table 2).
3. Significant transmembrane domains were identified

Table 2. cDNA clones with regions that matched motifs in the PROSITE database.

| Motifs | Description | Gene number <br> (KIAA) |
| :--- | :--- | :--- |
| ATP GTP A | ATP/GTP-binding site motif A (P-loop) | 0083,0089 |
| NAD G3PDH | NAD-dependent glycerol-3-phosphate dehydrogenase | 0089 |
| SUBTILASE HIS | Serine proteases, subtilase family, active site | 0091 |
| SUBTILASE SER | Serine proteases, subtilase family, active site | 0091 |
| RECEPTOR CYTOKINES 1 | Growth factor and cytokine receptors family | 0091 |
| C2 DOMAIN | C2 domain | 0093 |
| PRENYLATION | Prenyl group binding site | 0096 |
| TCP1-1 | Chaperonins TCP-1 | 009 |
| TCP1-2 | Chaperonins TCP-1 | 009 |
| TCP1-3 | Chaperonins TCP-1 | 009 |
| MITOCH CARRIER | Mitochondrial energy transfer proteins | 0098 |
| CLAT ADAPTOR M 2 | Clathrin adaptor complexes medium chain | 0106 |
| DEAD ATP HELICASE | DEAD and DEAH box families ATP-dependent helicases | 0109 |
| ATP A | ATP/GTP-binding site motif A | $\mathbf{0 1 1}$ |
| ALDOKETO REDUCTASE 1 | Aldo/keto reductase family | 0115 |

in 17 genes, 10 in which harbored multiple hydrophobic regions, as judged by the methods of Engelman et al. ${ }^{3}$ and of Kyte and Doolittle ${ }^{4}$ (see figures in the Supplement section).
4. Repetitive sequences were identified in the $3^{\prime}$-UTR of 4 genes.
5. Three genes (KIAA0096, 0099 and 0118) were related to signal transducing genes on the basis of sequence similarities and characteristic protein motifs. Particularly, it was noted that the KIAA0096 gene carried sequences with similarities to the genes in the protein kinase family and harbored a possible prenylation site, which is often observed in signal transducing membrane proteins.
6. The product of the KIAA0091 gene retained a possible leader peptide, a single transmembrane domain and a motif for a cytokine receptor, suggesting that the gene belongs to the cytokine receptor family.

### 3.2. Expression profiles in tissues

The expression profiles of the sequenced genes were examined in 16 different human tissues and in 2 cell lines including the KG- 1 cell as a positive control. The results are summarized in Table 3. On the basis of the Northern hybridization profiles, the genes can be categorized into several types. Thirty genes in total, 11 genes in the medium-sized and 19 in the small-sized cDNA class, were expressed ubiquitously in all the cells and tissues examined. This indicates that the small cDNA class is more ubiquitously expressed in various tissues. The hybridization profiles also showed that expression of the
small cDNA class is relatively abundant. Among the genes assigned to the ubiquitous class, 10 genes produced a few discrete bands which varied in size depending on the tissues (KIAA0084, HeLa: 0092, testis: 0093, skeletal muscle: 0094, testis: 0095, brain: 0097, KG-1: 0111, skeletal muscle: 0114, placenta: 0116, skeletal muscle: 0118 , testis). This may be due to alternative splicing or alternative initiation of transcription. Typical expression patterns of the genes in this category are shown in Fig. 2E-G. In the remaining genes, the expression of 8 genes was tissue-specific and that of 2 genes (KIAA0083 and 0087) was specific for the KG-1 cell line, although slight expression of KIAA0083 was detected in the thymus. Typical expression profiles of the genes belonging to this category are shown in Fig. 2A-E, in comparison with that of the ubiquitously expressed $\beta$-actin gene (Fig. 2 H ).

Including our previously reported cDNA sequences, ${ }^{1,2}$ we have predicted the coding sequences of 120 genes in total, and found that expression of 31 genes ( $26 \%$ ) was cell- or tissue-specific. By continuation of this type of analysis, therefore, it is possible to find many new genes with tissue-specific expression.

The chromosomal location of these genes were determined using a panel of human-rodent hybrid cell lines (see Table 3).

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Table 3. Summary of the cDNA sequence data and the expression patterns of the cloned genes in human tissues and cell lines.

He, heart; Br , brain; Pl , placenta; Lu, lung; Li, liver; Sk.m., skeletal muscle; Ki , kidney; Pa , pancreas; Sp , spleen; Th , thymus; Pr , prostate; Te , testis; Ov , ovary; $\mathrm{Sm.i}$, small intestine; Co, colon; Pe.b, peripheral blood leukocytes. a) Values excluding poly(A) sequences. b) Expression of mRNA in indicated cells and human tissues (Clontech, USA) was examined by Northern hybridization, and the relative strength of the positive signals are indicated ( $\pm,+,++,+++$ ). ${ }^{\text {c) }}$ Accession number of GSDB, DDBJ, EMBL and NCBI
nucleotide sequence databases. d) The presence of possible transmembrane domains was revealed (see Supplemental pages). e) Similarities to known genes were identified (see Table 1 and Supplemental pages).


Figure 2. The expression patterns of representative clones. cDNA fragments were randomly labeled and hybridization was carried out as described previously. Human MTN blots were purchased from Clontech Laboratories. A, KIAA0101; B, 0084; C, 0089; D, 0090; E, 0093; F, 0095; G, 0114; H, $\beta$-actin gene. Lane 1, heart; 2, brain; 3, placenta; 4, lung; 5, liver; 6, skeletal muscle; 7 , kidney; 8 , pancreas; 9 , spleen; 10, thymus; 11 , prostate; 12, testis; 13, ovary; 14, small intestine; 15, colon; 16, peripheral blood leukocyte.

## References

1. Nomura, N., Miyajima, N., Sazuka, T. et al. 1994, Prediction of the coding sequences of unidentified human genes. I. The coding sequences of 40 new genes (KIAA0001-KIAA0040) deduced by analysis of randomly sampled cDNA clones from human immature myeloid cell
line KG-1, DNA Res., 1, 27-35.
2. Nomura, N., Nagase, T., Miyajima, N. et al. 1994, Prediction of the coding sequences of unidentified human genes. II. The coding sequences of 40 new genes (KIAA0041-KIAA0080) deduced by analysis of cDNA clones from human cell line KG-1, DNA Res., 1, 223229.
3. Engelman, D. M., Steize, T. A., and Goldman, A. 1986, Identifying nonpolar transbilayer helices in amino acid sequences of membrane proteins, Annu. Rev. Biophys. Biophys. Chem., 15, 321-353.
4. Kyte, J. and Doolittle, R. F. 1982, A simple method for displaying the hydropathic character of a protein, J. Mol. Biol., 157, 105-132.
5. Walker, J. E., Saraste, M., Runswick, M. J., and Gay, N. J. 1982, Distantly related sequences in the $\alpha$ - and $\beta$-subunits of ATP synthase, myosin, kinases and other ATP-requiring enzymes and a common nucleotide binding fold, $E M B O$ J., 1, 945-951.
6. Otto, J., Argos, P., and Rossmann, M. G. 1980, Prediction of secondary structural elements in glycerol-3phosphate dehydrogenase by comparison with other dehydrogenases, Eur. J. Biochem., 109, 325-330.
7. Barr, P. J. 1991, Mammalian subtilisins: The long-sought dibasic processing endoproteases, Cell, 66, 1-3.
8. D'Andrea, A. D., Fasman, G. D., and Lodish, H. F. 1989, Erythropoietin receptor and interleukin-2 receptor $\beta$ chain: A new receptor family, Cell, 58, 1023-1024.
9. Perin, M. S., Fried, V. A., Mignery, G. A., Jahn, R., and Sudhol, T. C. 1990, Phospholipid binding by a synaptic
vesicle protein homologous to the regulatory region of protein kinase C, Nature, 345, 260-263.
10. Lowy, D. R. and Willumsen, B. M. 1989, New clue to ras lipid glue, Nature, 341, 384-385.
11. Ellis, J. 1992, Cytosolic chaperonin confirmed, Nature, 358, 191-192.
12. Nelson, D. R., Lawson, J. E., Klingerberg, M., and Douglas, M. G. 1993, Site-directed mutagenesis of the yeast mitochondrial ADP/ATP translocator, J. Mol. Biol., 230, 1159-1170.
13. Lee, J., Jongeward, G. D., and Sternberg, P. W. 1994, unc-101, a gene required for many aspects of Canorhabditis elegans development and behavior, encodes a clathrinassociated protein, Genes Dev., 8, 60-73.
14. Linder, P., Lasko, P., Ashburner, M. et al. 1989, Birth of the DEAD box, Nature, 337, 121-122.
15. Bohren, K. M., Bullock, B., Wermuth, B., and Gabbay, K. H. 1989, The aldo-keto reductase family, J. Biol. Chem., 264, 9547-9551.
16. Nomura, N., Takahashi, M., Matsui, M. et al., 1988, Isolation of human cDNA clones of myb-related genes, A$m y b$ and B-myb, Nucleic Acids Res., 16, 11075-11089.

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