

## Preparation and Applications of Chitosan Nanoparticles: A Brief Review

Divya Lanka<sup>1\*</sup>, Vijay Kumar Mittapally<sup>2</sup>

<sup>1</sup>Department of Pharmaceutics, Gland Institute of Pharmaceutical Sciences, Medak, Telangana

<sup>2</sup>Department of Biotechnology, Periyar University, Salem, Tamilnadu

### Review Article

Received: 08/07/2016

Accepted: 01/08/2016

Published: 10/08/2016

#### \*For Correspondence

Divya Lanka, Department of Pharmaceutics, Gland Institute of Pharmaceutical Sciences, Medak, Telangana.

#### E-Mail:

ramanujandivya@gmail.com

**Keywords:** Nanomedicine, Crustaceans, Chitosan, Chitin, Anti-thrombogenic

#### ABSTRACT

Chitosan is a natural polysaccharide well-known for its significant biological and chemical properties such as biodegradability, biocompatibility, bioactivity, and polycationicity. It is widely used in the nanomedicine, biomedical engineering and development of new therapeutic drug delivery systems with enhanced bioavailability, specificity and reduced pharmacological toxicity. Chitosan is a natural polymer obtained by deacetylation of chitin. After cellulose chitin is the second most abundant polysaccharide in nature. Different methods of synthesis of chitosan nanoparticles and their applications in nanomedicine, biomedical engineering, industrial and pharmaceutical fields were discussed in this review.

#### INTRODUCTION

Chitosan nanoparticles are biocompatible, biodegradable, cationic and relatively non-toxic in nature. Hence, they are widely used in drug delivery systems. Chitosan is a natural polysaccharide prepared by the N-deacetylation of chitin, which is a natural carbohydrate polymer found in the skeleton of crustaceans, such as crab, shrimp and lobsters (**Figure 1**) [1-5].



**Figure 1:** Different types of Crustaceans

#### METHODS

Chitin is prepared from crustacean shells through various chemical processes involving demineralization and deacetylation [6-8].

The crustacean shells are rinsed several times with water. Followed by drying in a hot air oven for about 24 hrs at 55 °C, the sample obtained was soaked in boiling 4% sodium hydroxide using 1000 ml beaker for 1 hr. The sample is then allowed to cool at room temperature for half an hour [9-11].

### ***Demineralization***

The sample obtained in the first step is demineralized using 1% hydrogen chloride in a ratio of 1:4 and soaked for 24 hrs. Then the samples are treated with 50 ml of 2% sodium hydroxide for 1 hr [12,13]. The sample remains are washed with deionized water after 24 hrs and then drained off [14-16].

### ***Deacetylation***

50% sodium hydroxide is added to the demineralized sample and boiled for 2 hours at 100 °C. The sample is allowed to cool and washed continuously with 50% sodium hydroxide. The sample obtained is filtered and dried in hot air oven for 6 hrs at 110 °C [17-20].

### ***Purification***

The purification process was performed by removal of insoluble material by filtration. The sample is further re-precipitated with 1 N sodium hydroxide and demetallized [21-25].

## **PREPARATION OF CHITOSAN NANOPARTICLES**

Nanoparticles are defined as colloidal structures with a varying size range of 1–1000 nm. Chitosan nanoparticles can be prepared by various methods including:

1. Ionotropic gelation method
2. Microemulsion method
3. Emulsification solvent diffusion method
4. Emulsion droplet coalescence method
5. Reverse micellisation
6. Desolvation

### ***Ionotropic gelation method***

Ionotropic gelation method involves the formation of chitosan nanoparticles based on the electrostatic interaction between the amine group of chitosan and negatively charged group of polyanions [26,27]. Chitosan is dissolved in acetic acid solution utilizing a stabilizing agent, such as poloxamer. Polyanion polymers are then added at room temperature under mechanical stirring which results in the formation of nanoparticles [28-30].

### ***Microemulsion method***

In this method, N-hexane is dissolved with a surfactant. Then, chitosan dissolved in acetic solution and glutaraldehyde is added dropwise to surfactant/hexane mixture under continuous stirring at room temperature. Nanoparticles are formed in the presence of surfactant [31-34]. The mixture is allowed to stir overnight to complete the crosslinking process. This enhances conjugation of the free amine group of chitosan with glutaraldehyde. The acetic acid solution is removed by evaporation [35-38].

## **APPLICATIONS OF CHITOSAN NANOPARTICLES**

1. Chitosan nanoparticles are used as anti-thrombogenic materials for: controlled release and drugs encapsulation [39,40].
2. Chitosan nanoparticles-based films are used in the food industry for the control of microorganisms in food thereby enhancing the shelf life of the food products.
3. Also used for the delivery of anti-infectives such as antibacterial, antiviral, antifungal drugs [41,42].
4. Chitosan exhibits anti-fungal property against *Aspergillus niger*, *Rhizopus stolonifer*, *Rhizopus oryzae* etc., in its free polymer form.
5. Chitosan nanoparticles can be a major source for the delivery of various drugs like heparin, chondroitin sulphate through oral route [43-45].
6. Chitosan nanoparticles, being extremely low toxic are used as efficient vectors for gene delivery.

7. Used as a potential adjuvant for vaccines against influenza, hepatitis [46,47].
8. Chitosan NP is also used in the dentistry to eliminate caries [48].
9. Used as an additive in antimicrobial textile industry for producing clothes for health care professionals.
10. The nanoparticles have also been proven to show skin regenerative properties, hence they can be used in anti-aging skin care products [49,50].

## REFERENCES

1. Ahamed MEH, Marjanovic L, Mbianda XY (2016) Statistical Optimization, Kinetic and Isotherm Studies on Selective Adsorption of Silver and Gold Cyanocomplexes Using Aminoguanidyl-Chitosan Imprinted Polymers.
2. Jayasinghe PS, Pahalawattaarachchi V, Ranaweera KKDS (2016) Effect of Extraction Methods on the Yield and Physicochemical Properties of Polysaccharides Extracted from Seaweed Available in Sri Lanka. *Poult Fish Wildl Sci* 4:150.
3. Keleher J, Thomas C, Kevin J, Bianca G, Mlynarski A, Brain S, et al. (2016) Synthesis and Characterization of a Chitosan / PVA Antimicrobial Hydrogel Nanocomposite for Responsive Wound Management Materials. *J Microb Biochem Technol* 8:065-070.
4. Bhavani K, Roshan Ara Begum E, Selvakumar S, Shenbagarathai R (2016) Chitosan– A Low Cost Adsorbent for Electroplating Waste Water Treatment. *J Bioremed Biodeg* 7:346.
5. Muniyappan Gandhi R (2016) Recent Advances in Chitosan Based Biosorbent for Environmental Clean-Up. *J Bioremed Biodeg* 7:e173.
6. Feketeföldi B, Cermenek B, Spirk C, Schenk A, Grimmer C, et al. (2016) Chitosan-Based Anion Exchange Membranes for Direct Ethanol Fuel Cells. *J Membra Sci Technol* 6:145.
7. García MA, Casariego A, Arnet Y, de la Paz N, Fernández M, et al. (2016) Evaluation of Chitosan Acid Salts as Clarifying Agents of Orange Nectar. *J Exp Food Chem* 2:106.
8. Jabnoun-Khiareddine H, El-Mohamedy RSR, Abdel-Kareem F, Abdallah RAB, Gueddes-Chahed M, et al. (2015) Variation in Chitosan and Salicylic Acid Efficacy Towards Soil-borne and Air-borne Fungi and their Suppressive Effect of Tomato Wilt Severity. *J Plant Pathol Microbiol* 6:325.
9. Puga CH, Orgaz B, Muñoz S, SanJose C (2015) Cold Stress and Presence of *Pseudomonas fluorescens* Affect *Listeria monocytogenes* Biofilm Structure and Response to Chitosan. *J Mol Genet Med* 9: 180.
10. Saikia C, Gogoi P, Maji TK (2015) Chitosan: A Promising Biopolymer in Drug Delivery Applications. *J Mol Genet Med* S4:006.
11. Dai Y, Zhong Z (2015) The Antioxidant Activities of C3, 6-Dibenzoylated Phenyl-Thiosemicarbazone-Chitosans. *J Develop Drugs* 4:140.
12. Ahmad M, Jayachandran M, Qureshi MA, Ikram S et al. (2015) Chitosan Based Dressings for Wound Care. *Immunochem Immunopathol: Open Access* 1:106.
13. Jana S, Trivedi MK, Tallapragada RM, Branton A, Trivedi D, Nayak G, et al. (2015) Characterization of Physicochemical and Thermal Properties of Chitosan and Sodium Alginate after Biofield Treatment. *Pharm Anal Acta* 6:430.
14. Chiu HT, Hsu XY, Yang HM, Ciou YS (2015) Synthesis and Characteristics of m-TMXDI-based Waterborne Polyurethane Modified by Aqueous Chitosan. *J Textile Sci Eng* 5:218.
15. Zeraatian S, Salehiomran A, Aghdam RM, Tafti SHA, Ghiasi SR (2015) Cardiac Effects of Glucagon-Like Peptide 1 with Chitosan-Based Scaffold after Inducing Myocardial Infarction in Dogs. *Cardiol Pharmacol* 4:159.
16. Pighinelli L, Guimaraes MF, Paz RL (2015) Properties of Hydrochloric Chitosan Solutions Modified With Nano-Calcium Phosphate Complex. *J Tissue Sci Eng* 6:155.
17. Radhakrishnan Y, Gopal G, Lakshmanan CC, Nandakumar KS (2015) Chitosan Nanoparticles for Generating Novel Systems for Better Applications: A Review. *J Mol Genet Med* S4:005.
18. Elango J, Robinson JS, Arumugam VK, Geevaretnama J, Durairaj S (2015) Mechanical and Barrier Properties of Multi-Composite Shark Catfish (*Pangasius fongaseous*) Skin Gelatin Films with the Addition of Sorbitol, Clay and Chitosan Using Response Surface Methodology. *J Mol Genet Med* 9:179.

19. Freitas JHES, Mahnke LC, Estevam-Alves MHM, Santana KV, Campos-Takaki GM, et al. (2015) Evaluation of the Potential of Cadmium and Dyes Removal by Chitosan Obtained from Zygomycetes. *J Mol Genet Med* S4:003.
20. Martins EAN, Baccarin RYA, Moraes APL, Mantovani CF, Machado TSL, et al. (2015) Evaluation of Chitosan-Glycerol Phosphate in Experimental Osteochondral Joint Defects in Horses. *J Mol Genet Med* S4:002.
21. Amouzgar P, Salamatinia B (2015) A Short Review on Presence of Pharmaceuticals in Water Bodies and the Potential of Chitosan and Chitosan Derivatives for Elimination of Pharmaceuticals. *J Mol Genet Med* S4:001.
22. Thirumavalavan M, Lee JF (2015) A Short Review on Chitosan Membrane for Biomolecules Immobilization. *J Mol Genet Med* 9:178.
23. Duan J, Liua Y, Liua L, Jianga J, Lia J (2015) Double-Network Carboxymethyl Chitosan Grafting Polyacrylamide/Alginate Hydrogel Compositions Adapted to Achieve High Stretchable Properties. *J Mol Genet Med* 9:177.
24. Silva DJB, Zuluaga F, Valencia CH (2015) Evaluation of Biocompatibility of Chitosan Films from the Mycelium of *Aspergillus niger* in Connective Tissue of *Rattus norvegicus*. *J Mol Genet Med* 9:174.
25. Kusuma HP, Agasi H, Darmokoeseoemo H (2015) Effectiveness Inhibition of Fermentation Legion using Chitosan Nanoparticles. *J Mol Genet Med* 9:173.
26. Aminabhavi TM (2015) Chitosan-Based Hydrogels in Biomedicine. *J Pharma Care Health Sys* 2:e133.
27. Abul A, Samad SA, Huq D, Moniruzzaman M, Masum M (2015) Textile Dye Removal from Wastewater Effluents Using Chitosan-Zno Nanocomposite. *J Textile Sci Eng* 5:200.
28. Ajalloueiian F, Fransson M, Tavanai H, Massumi M, Hilborn J, et al. (2015) Investigation of Human Mesenchymal Stromal Cells Cultured on PLGA or PLGA/Chitosan Electrospun Nanofibers. *J Bioprocess Biotech* 5:230.
29. Thien DT, An NT, Hoa NT (2015) Preparation of Fully Deacetylated Chitosan for Adsorption of Hg(II) Ion from Aqueous Solution. *Chem Sci J* 6:095.
30. Alsudir S, Lai EPC (2015) Hydroxypropyl Methacrylate Interaction and Chitosan Coating for Enhanced UV Detection Sensitivity of Colloidal Nanoparticles in Capillary Electrophoresis Analysis. *J Anal Bioanal Tech* 6: 242.
31. Hadwiger LA (2015) Chitosan: The Preliminary Research and the Host/Parasite System that Led to the Discovery of its Antifungal and Gene Inducing Properties. *J Mol Genet Med* 9:158.
32. Perchyonok VT, Zhang S, Basson N, Grobler S, Oberholzer T, et al. (2014) Insights into Functional Erythromycin/Antioxidant Containing Chitosan Hydrogels as Potential Bio-active Restorative Materials: Structure, Function and Antimicrobial Activity. *Adv Tech Biol Med* 2:116.
33. Manikandan A, Sathiyabama M (2015) Green Synthesis of Copper-Chitosan Nanoparticles and Study of its Antibacterial Activity. *J Nanomed Nanotechnol* 5:251.
34. Jebahi S, Oudadesse H, Faouzi FZ, Elleuch J, Rebai T, et al. (2013) Osteoinduction and Antiosteoporotic Performance of Hybrid Biomaterial Chitosan-Bioactive Glass Graft: Effects on Bone Remodeling. *J Material Sci Eng* 2:128.
35. Wang C, Ni K, Zhou X, Wei D, Ren Y (2013) Immobilization of *Gluconobacter oxydans* by Entrapment in Porous Chitosan Sponge. *J Bioprocess Biotech* 3:132.
36. Youwei Y, Yinzhe R (2013) Effect of Chitosan Coating on Preserving Character of Post-Harvest Fruit and Vegetable: A Review. *J Food Process Technol* 4:254.
37. Jesus S, Borchard G, Borges O (2013) Freeze Dried Chitosan/ Poly- e-Caprolactone and Poly-e-Caprolactone Nanoparticles: Evaluation of their Potential as DNA and Antigen Delivery Systems. *J Genet Syndr Gene Ther* 4:164.
38. Ganguly S (2013) Chitosan from Shellfishes having Promising Biomedical Importance: An Editorial. *Adv Pharmacoepidemiol Drug Saf* 2:e122.
39. Nadapdap TP, Lutan D, Arsyad KHM, Ilyas S (2014) Influence of Chitosan from Shrimp Skin to Quality and Quantity of Sperm of Albino Rats after Administration of Lead. *Andrology* 3:114.
40. Oliveira SM, Turner G, Rodrigues SP, Barbosa MA, Alikhani M, et al. (2013) Spontaneous Chondrocyte Maturation on 3D-Chitosan Scaffolds. *J Tissue Sci Eng* 4:124.

41. Jianglian D, Shaoying Z (2013) Application of Chitosan Based Coating in Fruit and Vegetable Preservation: A Review. *J Food Process Technol* 4:227.
42. Hajidoun HA, Jafarpour A (2013) The Influence of Chitosan on Textural Properties of Common Carp (*Cyprinus Carpio*) Surimi. *J Food Process Technol* 4:226.
43. Sarkar K, Debnath M, Kundu PP (2012) Recyclable Crosslinked O-Carboxymethyl Chitosan for Removal of Cationic Dye from Aqueous Solutions. *Hydrol Current Res* 3:138.
44. Mahmoud HA, Melake NA, El-Semary MT (2012) Bactericidal Activity of Various Antibiotics versus Tetracycline-loaded Chitosan Microspheres against *Pseudomonas aeruginosa* Biofilms. *Pharmaceut Anal Acta* S15.
45. Thi Yeu Ly NVSH (2012) Adsorption of U(VI) from Aqueous Solution by Chitosan Grafted with Citric Acid via Crosslinking with Glutraldehyde. *J Chem Eng Process Technol* 3:128.
46. de Assis CF, Melo-Silveira RF, de Oliveira RM, Costa LS, de Oliveira Rocha HA, et al. (2012) Cytotoxicity of Chitosan Oligomers Produced by Crude Enzyme Extract from the Fungus *Metarhizium Anisopliae* in Hepg2 and Hela Cells. *Bioprocess Biotechniq* 2:114.
47. Yu DG, Williams GR, Yang JH, Wang X, Qian W, et al. (2012) Chitosan Nanoparticles Self-Assembled from Electrospun Composite Nanofibers. *J Textile Sci Engg* 2:107.
48. Mahoney C, McCullough MB, Sankar J, Bhattarai N (2012) Nanofibrous Structure of Chitosan for Biomedical Applications. *J Nanomedic Biotherapeu Discover* 2:102.
49. Nassif L, Jurjus A, Nassar J, Ghafari J, Sabban MEI (2012) Enhanced in-Vivo Bone Formation by Bone Marrow Differentiated Mesenchymal Stem Cells Grown in Chitosan Scaffold. *J Bioeng Biomed Sci* 2:106.
50. Moussa SA, Farouk AF, Opwis K, Chollmeyer E (2011) Production, Characterization and Antibacterial Activity of *Mucor rouxii* DSM-119 Chitosan. *J Textile Sci Engg* 1:105.