

## Preparation of Mosquito Salivary Gland Extract and Intradermal Inoculation of Mice

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**[Abstract]** Mosquito-transmitted pathogens are among the leading causes of severe disease and death in humans. Components within the saliva of mosquito vectors facilitate blood feeding, modulate host responses, and allow efficient transmission of pathogens, such as Dengue, Zika, yellow fever, West Nile, Japanese encephalitis, and chikungunya viruses, as well as *Plasmodium* parasites, among others. Here, we describe standardized methods to assess the impact of mosquito-derived factors on immune responses and pathogenesis in mouse models of infection. This protocol includes the generation of mosquito salivary gland extracts and intradermal inoculation of mouse ears. Ultimately, the information obtained from using these techniques can help reveal fundamental mechanisms of interaction between pathogens, mosquito vectors, and the mammalian host. In addition, this protocol can help establish improved infection challenge models for pre-clinical testing of vaccines or therapeutics that take into account the natural route of transmission via mosquitoes.

**Keywords:** Mosquito, Saliva, Salivary gland, Extract, Virus transmission, Intradermal, Needle, Mouse

**[Background]** While probing for blood, the mosquito inoculates saliva that facilitates feeding but can also contain pathogens, if the mosquito has previously fed on an infected individual. Mosquito saliva plays an important role in establishing infection, facilitating dissemination, modulating immune responses, and exacerbating pathogenesis during West Nile virus (Schneider *et al.*, 2006; Styer *et al.*, 2011), Dengue virus (Cox *et al.*, 2012; Conway *et al.*, 2014; McCracken *et al.*, 2014; Schmid *et al.*, 2016), chikungunya virus (Agarwal *et al.*, 2016), Semliki Forest virus (Pingen *et al.*, 2016), Rift Valley Fever virus (Le Coupanec *et al.*, 2013) and *Plasmodium* parasite (Schneider *et al.*, 2011) infection. Many important questions yet remain and call for improved animal models.

Whereas inoculation via infected mosquitoes best mimics natural transmission, high variability in the inoculated dose and limited availability of insectary facilities result in restricted use of such procedures. In addition, the amount of saliva and the presence or absence of mosquito-derived components cannot be controlled when using infected mosquitoes. As an alternative, ‘spot feeding’ of uninfected female mosquitoes followed by intradermal inoculation of the pathogen via a needle mimics the natural deposition of saliva into mouse skin and delivers a defined dose of pathogen. The ‘spot feeding’ model has successfully been used to study Dengue virus (Cox *et al.*, 2012; McCracken *et al.*, 2014) and West

Nile virus (Moser *et al.*, 2015) infection but still requires the concomitant use of live mosquitoes and mice, and cannot control for the amount of saliva delivered. To separately control for mosquito and mouse experiments, inserting the mosquito proboscis into a sucrose solution in a capillary tube can serve to collect mosquito saliva artificially. The saliva collected during sugar feeding, however, differs qualitatively from mosquito saliva that is inoculated into the host skin during natural blood feeding (Marinotti *et al.*, 1990; Moser *et al.*, 2015).

Here, we describe the use of a simplified model of needle-inoculating mosquito salivary gland extract (SGE) from non-infected mosquitoes that can be delivered with a pathogen in a controlled manner at a defined dose. This method allows for independent handling of live mosquitoes and mice between collaborators and can more easily be standardized between assays and research groups. Use of SGE has proven useful to study infection with West Nile virus (Schneider *et al.*, 2006; Moser *et al.*, 2015), Dengue virus (Conway *et al.*, 2014; Schmid *et al.*, 2016), Rift Valley fever virus (Le Coupanec *et al.*, 2013), and Sindbis virus (Schneider *et al.*, 2004). Injection of SGE does not precisely mimic inoculation via the mosquito proboscis and likely contains non-secreted components of salivary glands. Nevertheless, this method allows collection of higher quantities of mosquito-derived factors, contains all secreted proteins, and can also be used in *in vitro* assays. Overall, the procedures described here should facilitate collaboration between entomologists, immunologists, and researchers studying pathogens of interest.

## **Materials and Reagents**

*Note: For the rearing of mosquitoes and general maintenance of a mosquito colony, see Bio-protocol Kauffman et al. (2017).*

1. Cotton-tipped swab, 15 cm handle (Puritan, catalog number: 25-826 5WC)
2. Petri dish, sterile, 60 mm (*e.g.*, 60 mm TC-Treated Cell Culture Dish, Corning, Falcon®, catalog number: 353002)
3. Plastic wrap
4. 50-ml conical centrifuge tubes (CELLTREAT Scientific Products, catalog number: 229422)
5. 15-ml conical centrifuge tubes (CELLTREAT Scientific Products, catalog number: 229412)
6. 5-ml tubes, 12 x 75 mm (Corning, Falcon®, catalog number: 352054)
7. 96-well plates (Thermo Fisher Scientific, Thermo Scientific™, catalog number: 15041)
8. 5-ml serological pipets (CELLTREAT Scientific Products, catalog number: 229206B)
9. 10-ml serological pipets (CELLTREAT Scientific Products, catalog number: 229211B)
10. Microcentrifuge tubes, 1.5 ml (Corning, Axygen®, catalog number: MCT-150-C)
11. Needle: 30-gauge, small RN hub, custom-made, point style 4 (10°-12° bevel), 25-mm length (Hamilton, catalog number: 7803-07)

*Note: Alternative needles: use with disposable hypodermic needle, e.g., 30 G x 1 inch (BD, catalog number: 305128).*

12. Frosted microscope slides, 25 x 75 x 1.0 mm (*e.g.*, Fisher Scientific, catalog number: 12-550-343)
13. Wooden applicator sticks, 15 cm (Puritan, catalog number: 807)
14. Insect Pins Morpho Black enameled No.000 (BioQuip, catalog number: 1208B000)
15. Dissecting probes fabricated from wooden applicator sticks and insect pins  
*Note: Dissecting probes are fabricated from 15-cm wooden applicators and insect pins. Soak the applicator sticks in hot water for at least 30 min, and cut the heads off the pins and discard. Using pliers, hold the pin, avoiding damage to the pointy end, and push the blunt (cut) end into the stick. Let the probe dry.*
16. Transfer pipets, 1 ml, bulk, nonsterile (BioLogix, catalog number: 30-0135)
17. pH test strips (*e.g.*, Sigma-Aldrich, catalog number: P4536-100EA)
18. Female mosquitoes
19. Mosquito holding carton (see Bio-protocol Kauffman *et al.*, 2017 for description and construction)
20. Mice
21. Triethylamine (Sigma-Aldrich, catalog number: T0866)
22. Ethyl alcohol
23. Phosphate-buffered saline (PBS), low endotoxin  $\leq 0.25$  EU/ml (*i.e.*, Mediatech, catalog number: 21-030-CV)
24. Micro BCA Protein Assay Kit (Thermo Fisher Scientific, Thermo Scientific™, catalog number: 23235)
25. Endpoint Chromogenic Limulus Amebocyte Lysate QCL-1000 Assay (Lonza, catalog number: 50-647U)
26. Disinfectant, such as Bleach or Umonium<sup>38</sup> (Huckert's Laboratoire International, catalog number: PF 12209)
27. Isoflurane, Iso-Vet (Chanelle, catalog number: CDS019936) for anesthesia of mice
28. 70% ethanol: Combine 74 ml of ethyl alcohol with 26 ml of water. Label as flammable and store at room temperature for up to 3 months
29. Alexa Fluor 680
30. Monoclonal antibody 4G2

## **Equipment**

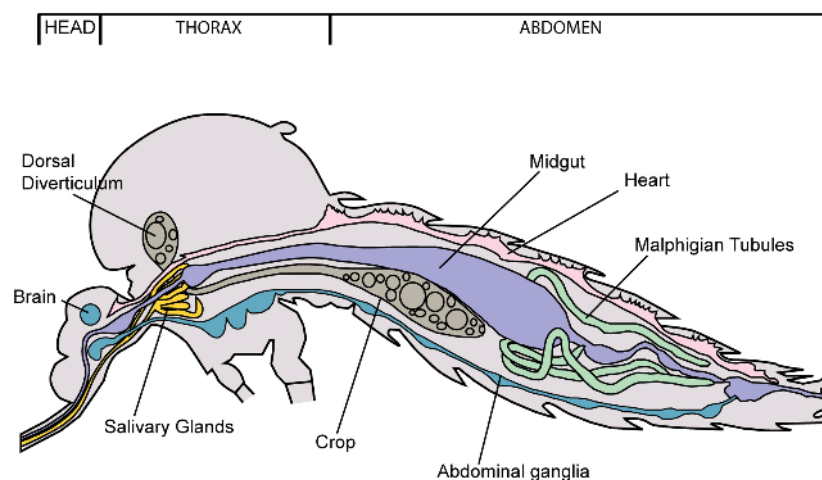
1. Stir plate and stir bar
2. Floating microtube rack (*i.e.*, Thermo Fisher Scientific, Thermo Scientific™, catalog number: 5974-4015)
3. Forceps, Superfine Tips, Swiss Style #5 (BioQuip, catalog number: 4535)
4. Cover slip forceps (Fisher Scientific, catalog number: S17328C)  
*Manufacturer: Medco Instruments, catalog number: S17328C.*

*Note: It is important that the flat parts of the forceps touch each other over the entire surface. If the opposing surfaces touch at only one point, the ear skin is more likely to generate folds when inserting the needle.*

5. Stereoscopic Zoom Microscope (e.g., Nikon Instruments, model: SMZ1500)
6. Branson Sonifier Model 450 (Branson 450 Analog Sonifier with 1/2" Horn, 400 W, 120 VAC) (Cole-Parmer, catalog number: EW-04715-03)  
*Manufacturer: Emerson Electric, BRANSON, model: Model 450.*
7. Cup Horn for Ultrasonic Processors, 3" dia (Cole-Parmer, catalog number: EW-04715-39)  
*Manufacturer: Emerson Electric, BRANSON, catalog number: 109-116-1760.*
8. Branson Sound-Proof Enclosure for Ultrasonic Processors (Cole-Parmer, catalog number: EW-04715-44)  
*Manufacturer: Emerson Electric, BRANSON, catalog number: 101-063-275.*
9. Refrigerated centrifuge (e.g., Eppendorf, model: 5417 R)
10. Reusable glass microinjection syringe type 702RN, no needle, volume 25  $\mu$ l (Hamilton, catalog number: 7636-01)  
*Note: Alternative syringe: Reusable glass microinjection syringe model 702 LT (Luer tip), SYR, NDL sold separately, no needle, volume 25  $\mu$ l (Hamilton, catalog number: 80401).*
11. Heat block capable of heating to 37 °C and 60 °C (e.g., Digital Dry Block Heater, Single Block, 240 V, Thermo Fisher Scientific, Thermo Scientific™, catalog number: 88870004)
12. Plate reader capable of reading absorption at 562 nm and 405-410 nm, and temperature control capable of heating to 37 °C (e.g., BioTek Instruments, model: ELx808)

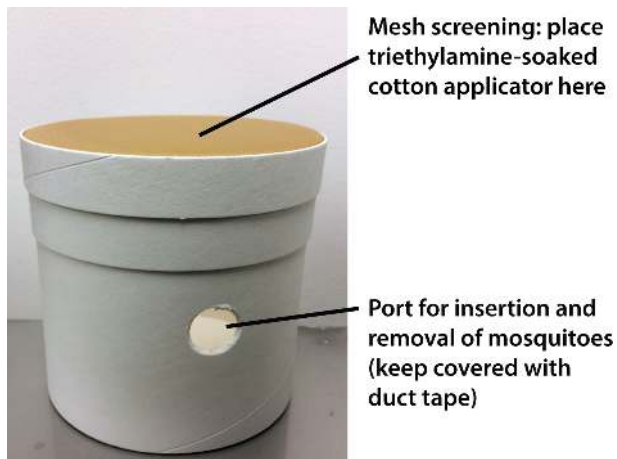
## **Procedure**

### **A. Dissection of salivary glands of female mosquitoes (Figure 1)**



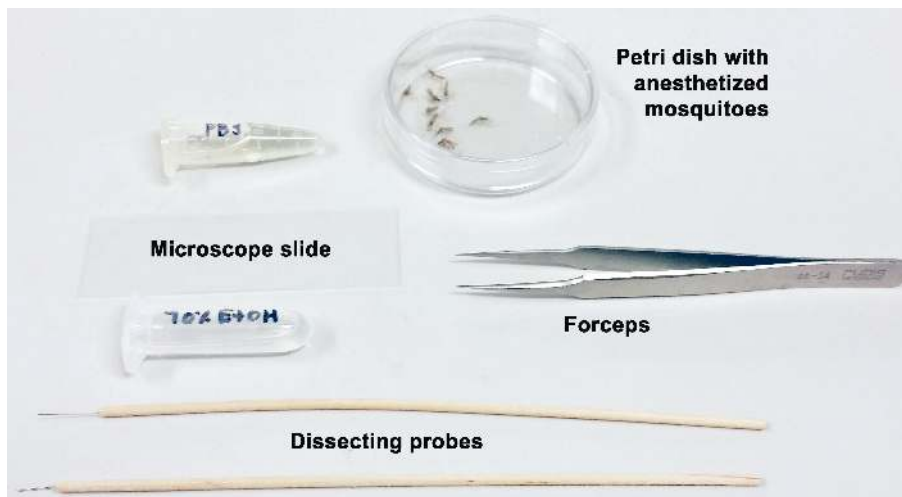
**Figure 1. Diagram of *Aedes* mosquito showing location of salivary glands**

1. Anesthetize mosquitoes by resting a cotton swab applicator moistened with triethylamine on the mesh lid of the mosquito holding carton; cover with plastic wrap or Petri dish to contain the triethylamine (Figure 2).



**Figure 2. Mosquito holding carton used for anesthetizing mosquitoes**

2. Once the mosquitoes are anesthetized, transfer them to a Petri dish and cover. Remove a female from the dish using Superfine tip forceps and dip into a microcentrifuge tube containing 70% ethanol to sterilize the body surface (Figure 3).



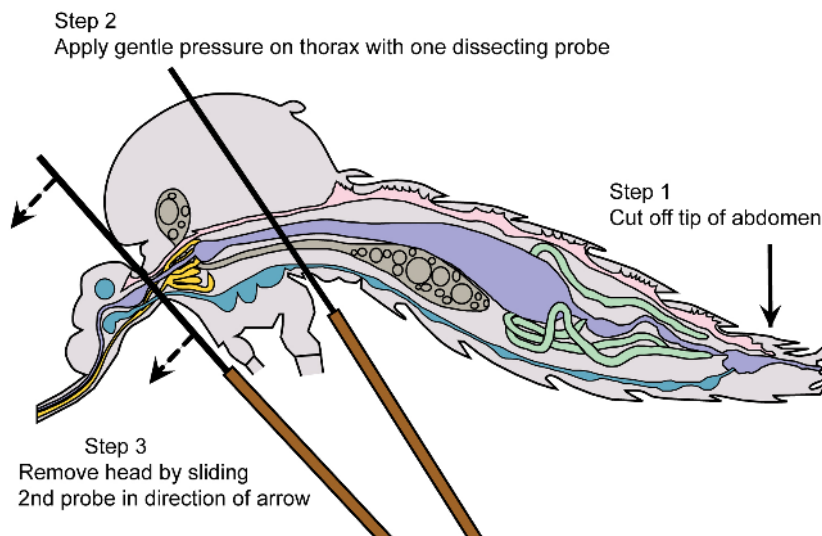
**Figure 3. Tools for salivary gland removal**

3. Place a drop (approximately 50  $\mu$ l) of PBS on a microscope slide using a transfer pipet and spread the drop around.
4. Place the slide on the stage of the dissecting microscope.
5. Remove the legs of the mosquito, then place the body on its side in the PBS (see Video 1).

### Video 1. Dissection of mosquito salivary glands

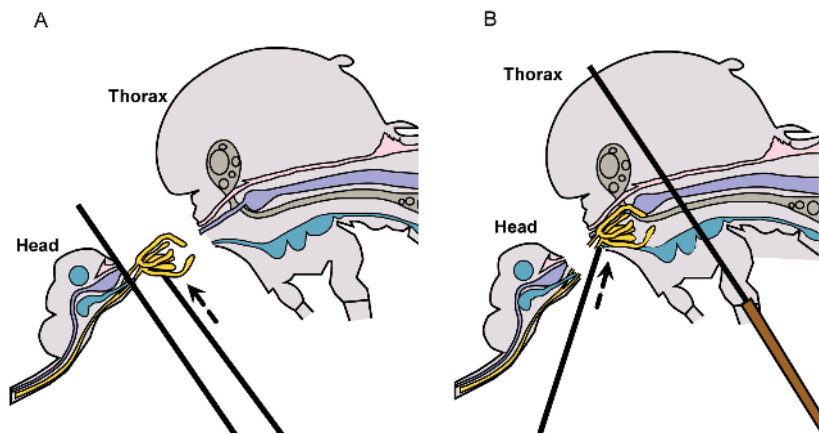


6. Remove the tip of the abdomen with a sharp cut from the dissecting probe; do not pull.
7. Place one dissecting probe on the thorax of the mosquito and the other between the head and the thorax. Apply gentle pressure to the thorax, then slowly remove the head by gently pulling away from the thorax (Figure 4 and Video 1).



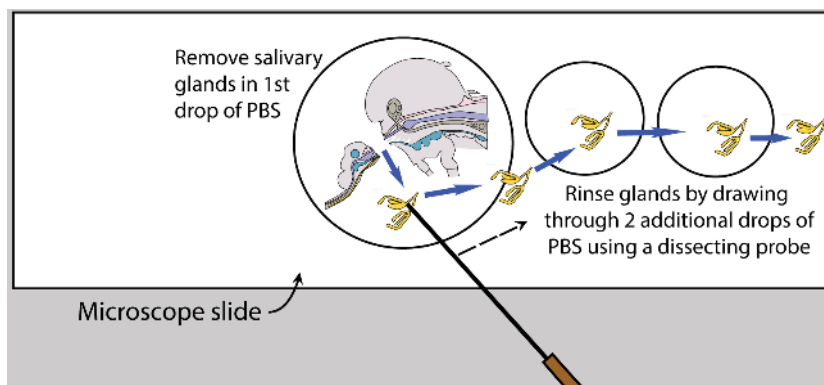
**Figure 4. Mosquito diagram: positioning for salivary gland removal**

8. The salivary glands will be apparent, if they remain with the head and may be removed by lifting them up and away from the head with the probe, while holding one probe between the glands and the head (Figure 5A and Video 1). If the glands remain within the thorax, apply gentle pressure with one probe held flat against the thorax and use the other probe to locate and gently tease out the glands (Figure 5B and Video 1).



**Figure 5. Mosquito diagram: removal of salivary glands.** A. Lift salivary glands up & away from head; B. Tease salivary glands from thorax.

9. Once the glands are removed, place an additional two drops of fresh PBS separately on the slide, near but not touching the initial drop, and carefully rinse the glands by drawing them sequentially through the PBS drops (Figure 6).



**Figure 6. Rinse salivary glands after removal**

10. Place one pair of salivary glands in 10  $\mu$ l of PBS, pooling up to 50 pairs in a volume of 500  $\mu$ l. Because mosquitoes inoculate volumes < 5  $\mu$ l while probing for blood vessels, one may choose to prepare SGE at a higher concentration (using lower volumes of PBS) for later being able to reduce the volume of intradermal inoculation.
11. Store at -20  $^{\circ}$ C until ready to prepare the SGE.

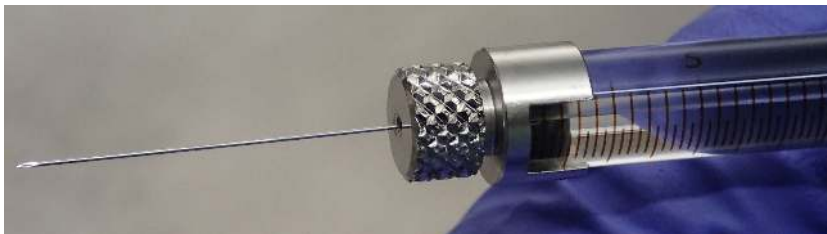
## B. Preparation of SGE

1. Remove frozen glands and thaw quickly at 37  $^{\circ}$ C.
2. Add water and ice chips to the horn of the sonicator.
3. Place a microfuge tube containing salivary glands into the floating microtube rack.
4. Program the sonicator with the following parameters: 100 mV, 3 bursts of 20 sec, 1 min cooling between bursts.

5. Centrifuge at 5,000  $\times g$  at 4 °C for 10 min.
6. Remove the supernatant to a fresh microcentrifuge tube.
7. Determine the protein concentration of the SGE using the Micro BCA Protein Assay Kit, following the manufacturer's microplate procedure. The typical amount of protein obtained per one pair of salivary gland is 0.8-1.2  $\mu\text{g}$ , which we obtained in a volume of 10  $\mu\text{l}$  PBS.
8. Use pH strips to test pH of SGE stocks, which should be neutral pH of 7.0 due to the buffering capacity of PBS. In the case that the pH is not neutral, interpretation of the data should take this into account. On the one hand, pH may be an intrinsic property of saliva from the used vector species. On the other hand, differences in pH may change infectivity and transmission of a virus or another pathogen used in the experiments.
9. Confirm that SGE stock is free of endotoxin or contains very low levels that are well below the FDA-approved limit for injection solutions (< 5 endotoxin units per kg per hour), as tested using the endpoint chromogenic Limulus Amebocyte Lysate kit (Lonza) and 10  $\mu\text{l}$  SGE stock, which corresponds to approximately 1  $\mu\text{g}$  of protein. If endotoxin levels are high, the SGE preparation should be discarded because observed effects cannot clearly be attributed to components of the SGE or the presence of endotoxin.
10. Store at -80 °C.

#### C. Intradermal inoculation

1. **Dilute** the SGE in sterile PBS in a 1.5-ml tube. Adjust concentration to contain the extract of 0.2 to 1 salivary gland (each mosquito has one pair of salivary glands) per 20  $\mu\text{l}$  PBS. If testing the effect of SGE on arbovirus infection, mix SGE with a concentrated virus stock of *e.g.*, DENV ( $10^4$ - $10^6$  plaque forming units, see [Schmid and Harris, 2014; Schmid *et al.*, 2016]). Use virus diluted in PBS as control for the absence of mosquito saliva. As further controls, use non-injected (untouched) ears or intradermal inoculation of PBS alone.  
If titers of the virus and SGE stock solutions are high enough, it may be beneficial to inoculate a total volume of less than 20  $\mu\text{l}$  because mosquitoes typically inoculate < 5  $\mu\text{l}$ .
2. **Assemble and sterilize** the reusable glass syringe and needle (Figure 7) by flushing at least three times with 70% ethanol. Rinse three times with sterile PBS.



**Figure 7. Hamilton glass syringe and needle**



3. For a 20  $\mu$ l injection, **take up** a total of 25  $\mu$ l of the salivary gland extract into the glass syringe, while excluding air bubbles. Avoid touching the surface of the tube to avoid blunting of the tip of the reusable needle.
4. In the meantime, **anesthetize** mice via inhalation of 2-4% isoflurane in a stream of 2 L/min oxygen. Adjust inhalation time and percent isoflurane until the mouse breathes approximately once per second. After removing from the anesthesia, the mouse will still be anesthetized for approximately one minute, to allow the intradermal inoculation.

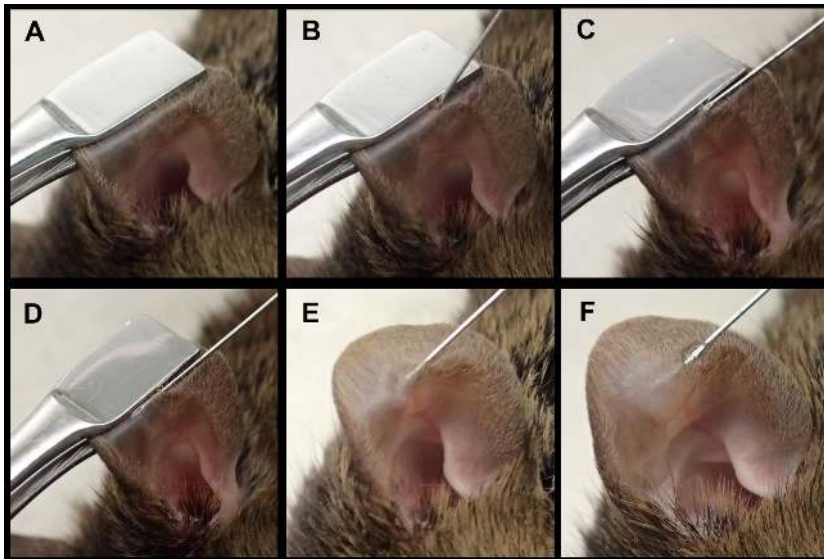
*Note: Anesthesia of mice via isoflurane inhalation must be performed by trained laboratory personnel.*

5. **Immobilize** one ear of an anesthetized mouse using cover slip forceps so that the ventral part of the ear faces up. The bottom of the forceps should end approximately in the center of the ear at the site of injection. Tilt the ear backwards and remove folds within the ear skin (Figure 8A).

*Note: Make sure that the ear skin is dry prior to injection in order to avoid sticking of the ear to the forceps. If necessary, dry the skin with tissue or gauze. See also Video 2, Intradermal Inoculation.*

#### Video 2. Intradermal inoculation of the mouse ear





**Figure 8. Intradermal inoculation of the mouse ear**

6. **Force** the part of the forceps that faces up towards the top and the part at the dorsal side of the ear (facing away) towards the bottom. Thus forming a 'bench' or 'ledge' will provide a solid support for the injection site (Figure 8A).
7. Hold the needle with the bevel pointing up. Approach the 'ledge' with the needle held at a ~45° angle and gently **penetrate** the surface of the skin (Figure 8B).
8. **Drop** the 45° angle of the needle until it is parallel to the forceps and skin, while precisely maintaining the position of the tip of the needle. Being then parallel to the skin, one should feel the support of the forceps along the length of the needle (Figure 8C).
9. **Insert** the needle ~3 mm while moving parallel to the surface within the skin tissue. One should feel substantial resistance and continuously see the bevel of the needle through the surface of the skin (Figure 8D). If the resistance suddenly decreases or the bevel can no longer be seen, the needle may have poked through the back of the skin (if this occurs, start a second attempt after removing the forceps and needle).
10. Carefully **let go** of the forceps without moving the ear skin relative to the needle (Figure 8E).
11. Move the hand that held the forceps over the other hand to **reach the plunger** of the syringe.
12. Slowly **inoculate** 20 µl of solution and monitor the appearance of a small blister underneath the skin (Figure 8F). Successful injection leads to a transient stiffening of the ear (induced by the small blister), and some resistance should be felt while injecting. If no **blister** develops or no stiffening occurs, the inoculum may have leaked through the back of the ear. If this occurs, remove the needle and reinsert at a slightly different site of the ear.
13. If a small amount of inoculum **leaks** through the front or back of the skin, compensate this volume with the extra 5 µl that have been taken up into the syringe to ensure injection of exactly 20 µl. If the inoculated volume is still < 20 µl, inoculate more at a different site of the same ear skin.

14. After inoculation, hold the side of the ear (not where the blister has formed) with the forceps and slowly **remove** the needle.

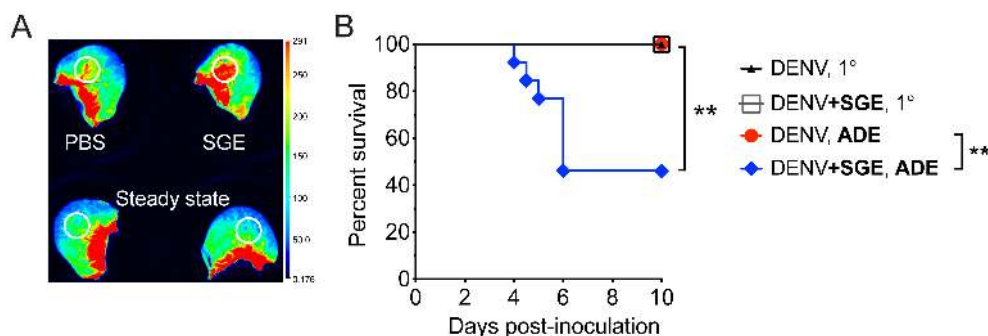
*Note: Start using a new needle once initial puncture of the skin becomes more difficult (every 6-10 experiments).*

15. Throughout the procedure, closely **monitor** the breathing of the anesthetized mouse. Also, monitor the mice regularly during and after waking up from anesthesia.

16. After use, **clean** the glass syringe by flushing several times with Bleach or other disinfectant, followed by PBS and then 70% ethanol. For reusing the glass syringes, designate one syringe per reagent (*e.g.*, SGE alone, virus alone, SGE + virus, PBS alone) to avoid cross-contamination. The glass syringes and needles can also be autoclaved.

## Data analysis

For details on animal experiments and ethical approvals, methods used, data processing and analysis, and statistical tests that were used to generate the example data in Figure 9, see our original publication (under the open access Creative Commons Attribution license [Schmid *et al.*, 2016]), from which the representative example of data (below) was extracted:



**Figure 9. Intradermal inoculation of SGE induces vascular leak into the skin and exacerbates Dengue disease in a mouse model of infection.** A. Wild-type mice were inoculated intravenously with dextran that was labeled with Alexa Fluor 680. Immediately thereafter, 15  $\mu$ l SGE or PBS was inoculated intradermally. After 30 min, mice were euthanized, and ears were scanned using the Odyssey CLx Infrared Imaging System (Licor). Representative scan showing fluorescence that leaked with plasma into the skin at sites of intradermal inoculation or corresponding areas in steady-state ears (white circles). B. Mice deficient in the interferon- $\alpha/\beta$  receptor (*Ifnar*<sup>-/-</sup>) were inoculated intradermally with 10<sup>5</sup> plaque forming units of Dengue virus-2 strain D220 in the absence or presence of enhancing antibodies (5  $\mu$ g of monoclonal antibody 4G2, targeting Dengue virus E protein) to model antibody-dependent enhancement (ADE). Dengue virus was inoculated alone or after mixing with *Ae. aegypti* SGE (+SGE). Kaplan-Meier curves showing survival of mice. Data were pooled from three experiments. Statistically significant differences were tested between the presence and absence of SGE using the Log-rank (Mantel-Cox) test and are marked as \*\* for  $P < 0.01$ .

## **Acknowledgments**

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## **References**

1. Agarwal, A., Joshi, G., Nagar, D. P., Sharma, A. K., Sukumaran, D., Pant, S. C., Parida, M. M. and Dash, P. K. (2016). [Mosquito saliva induced cutaneous events augment Chikungunya virus replication and disease progression](#). *Infect Genet Evol* 40: 126-135.
2. Conway, M. J., Watson, A. M., Colpitts, T. M., Dragovic, S. M., Li, Z., Wang, P., Feitosa, F., Shepherd, D. T., Ryman, K. D., Klimstra, W. B., Anderson, J. F. and Fikrig, E. (2014). [Mosquito saliva serine protease enhances dissemination of Dengue virus into the mammalian host](#). *J Virol* 88(1): 164-175.
3. Cox, J., Mota, J., Sukupolvi-Petty, S., Diamond, M. S. and Rico-Hesse, R. (2012). [Mosquito bite delivery of Dengue virus enhances immunogenicity and pathogenesis in humanized mice](#). *J Virol* 86(14): 7637-7649.
4. Kauffman, E., Payne, A., Franke, M. A., Schmid, M. A., Schmid, E. and Kramer, L. D. (2017). [Rearing of \*Culex\* spp. and \*Aedes\* spp. mosquitoes](#). *Bio Protoc* 7(17): e2542.
5. Le Coupanec, A., Babin, D., Fiette, L., Jouvion, G., Ave, P., Misse, D., Bouloy, M. and Choumet, V. (2013). [Aedes mosquito saliva modulates Rift Valley fever virus pathogenicity](#). *PLoS Negl Trop Dis* 7(6): e2237.
6. Marinotti, O., James, A. A. and Ribeiro, J. (1990). [Diet and salivation in female \*Aedes aegypti\* mosquitoes](#). *J Insect Physiol* 36(8): 545-548.
7. McCracken, M. K., Christofferson, R. C., Chisenhall, D. M. and Mores, C. N. (2014). [Analysis of early dengue virus infection in mice as modulated by \*Aedes aegypti\* probing](#). *J Virol* 88(4): 1881-1889.
8. Moser, L. A., Lim, P. Y., Styer, L. M., Kramer, L. D. and Bernard, K. A. (2015). [Parameters of Mosquito-enhanced West Nile virus infection](#). *J Virol* 90(1): 292-299.

9. Pingen, M., Bryden, S. R., Pondeville, E., Schnettler, E., Kohl, A., Merits, A., Fazakerley, J. K., Graham, G. J. and McKimmie, C. S. (2016). [Host inflammatory response to mosquito bites enhances the severity of arbovirus infection.](#) *Immunity* 44(6): 1455-1469.
10. Schmid, M. A., Glasner, D. R., Shah, S., Michlmayr, D., Kramer, L. D. and Harris, E. (2016). [Mosquito saliva increases endothelial permeability in the skin, immune cell migration, and dengue pathogenesis during antibody-dependent enhancement.](#) *PLoS Pathog* 12(6): e1005676.
11. Schmid, M. A. and Harris, E. (2014). [Monocyte recruitment to the dermis and differentiation to dendritic cells increases the targets for dengue virus replication.](#) *PLoS Pathog* 10(12): e1004541.
12. Schneider, B. S., Mathieu, C., Peronet, R. and Mecheri, S. (2011). [Anopheles stephensi saliva enhances progression of cerebral malaria in a murine model.](#) *Vector Borne Zoonotic Dis* 11(4): 423-432.
13. Schneider, B. S., Soong, L., Girard, Y. A., Campbell, G., Mason, P. and Higgs, S. (2006). [Potentiation of West Nile encephalitis by mosquito feeding.](#) *Viral Immunol* 19(1): 74-82.
14. Schneider, B. S., Soong, L., Zeidner, N. S. and Higgs, S. (2004). [Aedes aegypti salivary gland extracts modulate anti-viral and TH1/TH2 cytokine responses to sindbis virus infection.](#) *Viral Immunol* 17(4): 565-73.
15. Styer, L. M., Lim, P. Y., Louie, K. L., Albright, R. G., Kramer, L. D. and Bernard, K. A. (2011). [Mosquito saliva causes enhancement of West Nile virus infection in mice.](#) *J Virol* 85(4): 1517-1527.