

Profiling of Flavonol Derivatives for the Development of Antitrypanosomatidic Drugs

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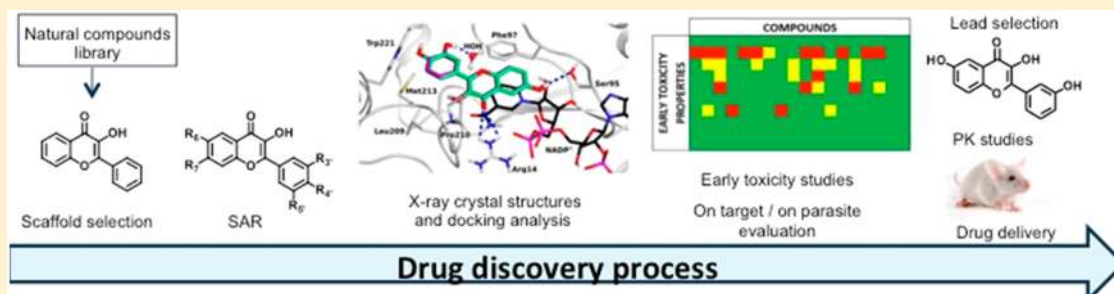
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Supporting Information



ABSTRACT: Flavonoids represent a potential source of new antitrypanosomatidic leads. Starting from a library of natural products, we combined target-based screening on pteridine reductase 1 with phenotypic screening on *Trypanosoma brucei* for hit identification. Flavonols were identified as hits, and a library of 16 derivatives was synthesized. Twelve compounds showed EC₅₀ values against *T. brucei* below 10 μM. Four X-ray crystal structures and docking studies explained the observed structure–activity relationships. Compound 2 (3,6-dihydroxy-2-(3-hydroxyphenyl)-4H-chromen-4-one) was selected for pharmacokinetic studies. Encapsulation of compound 2 in PLGA nanoparticles or cyclodextrins resulted in lower in vitro toxicity when compared to the free compound. Combination studies with methotrexate revealed that compound 13 (3-hydroxy-6-methoxy-2-(4-methoxyphenyl)-4H-chromen-4-one) has the highest synergistic effect at concentration of 1.3 μM, 11.7-fold dose reduction index and no toxicity toward host cells. Our results provide the basis for further chemical modifications aimed at identifying novel antitrypanosomatidic agents showing higher potency toward PTR1 and increased metabolic stability.

INTRODUCTION

Protozoan parasites of the Trypanosomatidae family are the etiological agents of several significant neglected tropical diseases including human African trypanosomiasis (HAT),

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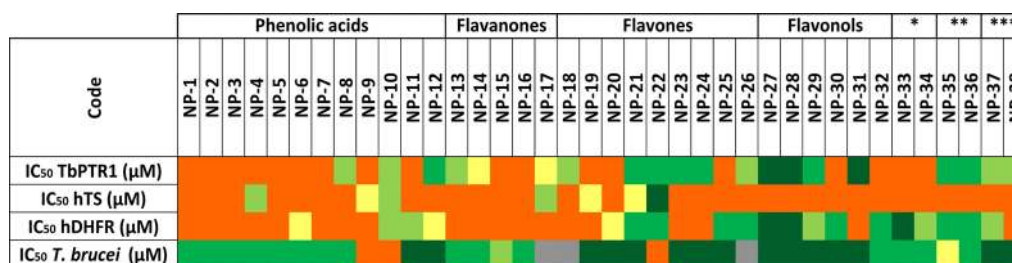


Figure 1. Activity profile of the 38 phytochemicals screened against *TbPTR1*, hTS, and hDHFR and against the *T. brucei* parasite. The IC₅₀ is indicated by the color: dark-green, 0–30 µM; green, 31–90 µM; light-green, 90–150 µM; yellow, 151–250 µM; red, >250 µM; gray, not tested. *, Catechins; **, triterpenes; ***, anthraquinones.

Chagas' disease, and leishmaniasis, which collectively affect nearly 10 million people worldwide. HAT is caused by the bloodstream form of *Trypanosoma brucei*.¹ *Leishmania* spp. infect macrophages and cause a wide spectrum of symptoms ranging from cutaneous lesions to potentially fatal visceral infections.² *Trypanosoma cruzi*, the etiological agent of Chagas' disease, is an obligate intracellular parasite that invades different internal organs. Since effective vaccines are lacking, the control of these diseases is based on vector control and chemotherapy. However, the drugs currently available are characterized by high toxicity, limited efficacy, and require a long period of treatment. The first line drugs have been used for over half a century, and their efficacy is compromised by widespread resistance.^{3–5} Therefore, there is an urgent requirement for new, safe, and effective drugs. The drug discovery process against these parasites is hampered by the intrinsic biology of these organisms that can include intracellular stages or central nervous system involvement.⁶

Although there are very few validated drug targets for leishmaniasis and HAT, a target-based approach is extensively used in the drug discovery process for neglected tropical diseases.⁷ The folate pathway is an established target for the treatment of bacterial infections and some parasitic diseases such as malaria.⁸ Drugs targeting folate-dependent enzymes represent useful candidates against trypanosomatid infections and key enzymes of the folate metabolism are thymidylate synthase (TS) and dihydrofolate reductase (DHFR), which is the target of the antifolate drugs methotrexate (MTX), pyrimethamine (PYR), and trimethoprim (TMP). However, the activity of classical inhibitors of DHFR against *Leishmania* and *Trypanosoma* is reduced because of pteridine reductase 1 (PTR1).⁹ PTR1 is responsible for the salvage of pterins in parasitic trypanosomatids and has overlapping activity with DHFR, providing a metabolic bypass to alleviate DHFR inhibition.^{10–12} Under physiological conditions, PTR1 is responsible for only 10% of the reduction of folic acid required by the cell, but when classic antifolate drugs inhibit DHFR, the gene encoding the NADPH-dependent PTR1 is upregulated and PTR1 provides the reduced folates necessary for parasite survival.¹³ Therefore, PTR1 is considered a promising target for the development of improved therapies. This protein is considered a validated target in *T. brucei*, even though a correlation between inhibition of *T. brucei* PTR1 (*TbPTR1*) and inhibition of parasite growth has not been demonstrated yet. The combination with MTX, a DHFR inhibitor, has been shown to improve the efficacy and the potency of PTR1 inhibitors.^{14,15}

In the drug discovery process, natural compounds represent a potential source of new leads.^{16–19} Different natural com-

pounds from plants including flavonoids have shown antitrypanosomatid activity.^{20,21} Although flavonoids have been found to inhibit polyamine biosynthesis, the mechanism of action has not been fully elucidated.^{21–24} Flavonoids exert pleiotropic effects, and different targets have been suggested to explain at least part of their antiparasitic activity.^{25–27} Flavonoids have been proposed as PTR1 inhibitors only in silico.²⁸ Although some flavonoids are very effective in vitro, they lack notable in vivo activity due to their low bioavailability.²² However, an efficient drug delivery strategy together with a clear definition of their molecular targets could lead to the exploitation of the antiparasitic potential of these natural products.²²

The purpose of the present study was to identify new PTR1 inhibitors showing antitrypanosomal (*T. brucei*, *T. cruzi*) and antileishmanial (*Leishmania infantum*) activity by screening a library of natural products and then by using a medicinal chemistry approach to design, synthesize, and biologically evaluate a new library. Assessment of early ADME and toxicity properties provided selection criteria for further studies. MTX has previously been combined with PTR1 inhibitors to achieve synergistic/additive effects on *T. brucei*.¹⁴ In our study, we evaluated the antiparasitic activity of the compounds both alone and in combination with MTX. Pharmacokinetic studies using BALB/c mice showed that one of our compounds (compound 2) was characterized by a short half-life, and therefore it was loaded in poly(lactic-co-glycolic acid) (PLGA) nanoparticles and solubilized with cyclodextrins, aiming to increase its stability and bioavailability.

RESULTS AND DISCUSSION

On-Target Screening of a Natural Product Library and Hit Identification. To identify potential new drugs for treating parasitic diseases, we investigated a library of natural products. Starting from a library of 98 compounds, a computational docking analysis against *TbPTR1* together with consideration of structural diversity allowed the restriction of the library to 38 compounds that were tested for PTR1 inhibition. These 38 phytochemicals (NP-1–NP-38), consisting of twelve phenolic acids, five flavanones, nine flavones, six flavonols, two catechins, two triterpenes, and two anthraquinones, were investigated for in vitro inhibition of *TbPTR1* and for growth inhibition of *T. brucei*. Due to their ability to bind similar folate-related metabolites, PTR1 inhibitors can also interact with other folate-dependent enzymes such as TS and DHFR both from parasite and human cells. Both human enzymes can be considered potential off-targets for PTR1 inhibitors, therefore all the compounds were assessed for their selectivity toward the human TS and human DHFR. The data obtained are depicted

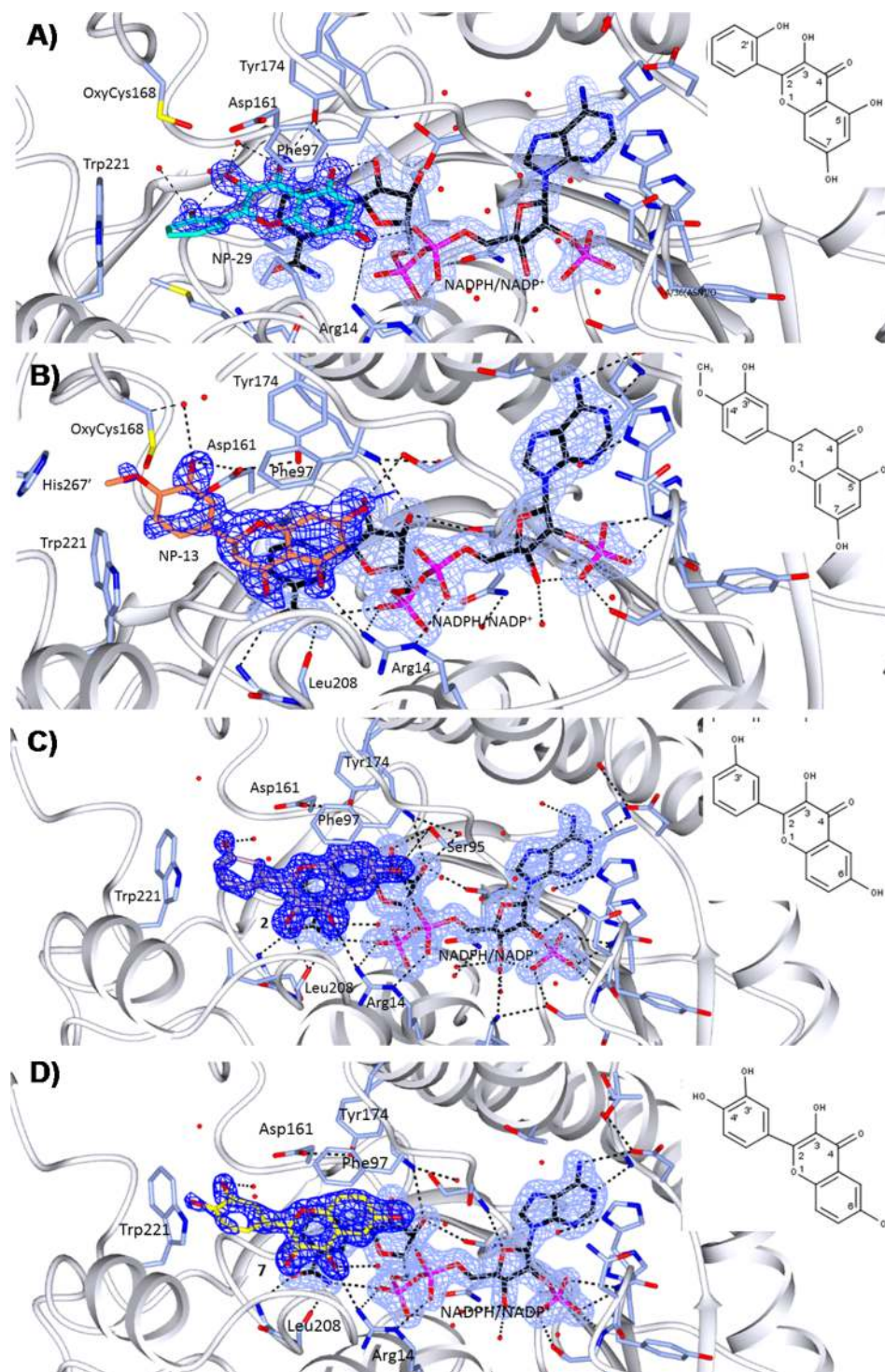


Figure 2. Crystal structures of *TbPTR1* (gray cartoon, interacting residues in sticks) in complex with NADPH/NADP⁺ (in sticks, black carbon atoms) and four inhibitors (in sticks) (A) NP-29 (cyan), (B) NP-13 (orange), (C) compound 2 (lilac), and (D) compound 7 (yellow). Hydrogen bond interactions (dashed lines) in the active site are shown. The $2F_o - F_c$ electron density maps corresponding to the inhibitors (dark-blue wire) and NADPH/NADP⁺ (light-blue wire), contoured at the 1σ level are shown. The chemical structures and atom names of the ligands are specified in the insets.

in Figure 1. All chemical structures and IC₅₀ values are reported in Supporting Information, Table S1.

With the exception of quercetin dehydrate (NP-30), all flavon-3-ol type aglycones showed a significant *TbPTR1* inhibition, with 3-hydroxyflavone (NP-27, IC₅₀ = 12.8 μM) and isorhamnetin (NP-31, IC₅₀ = 13.9 μM) being the most potent, whereas the glycosylated flavonoid (NP-32) did not

inhibit *TbPTR1*. Most phenolic acids showed no inhibitory activity toward *TbPTR1*, highlighting the importance of a condensed ring (chromen-4-one) for protein inhibition. The presence of two rings, one aromatic and the other one aliphatic (NP-8) or both aromatic (NP-10), slightly increased the inhibitory activity with respect to the other phenolic acid derivatives.

The simplest flavonol (NP-27, $IC_{50} = 12.8 \mu M$) was 10 times more potent than the corresponding flavone (NP-18, $IC_{50} = 120.8 \mu M$). This implies the importance of OH at position R3. Moreover, isorhamnetin (NP-31, $IC_{50} = 13.9 \mu M$) was over 35 times more active against *TbPTR1* than the corresponding flavone (chrysoeriol, NP-25, $IC_{50} > 490 \mu M$). The insertion of a hydroxyl group at position R5 led to a decrease in the inhibitory activity. Indeed, the simplest flavone (NP-18, $IC_{50} = 120.8 \mu M$) is more than 20-fold more active than primuletin (NP-19, $IC_{50} > 2450 \mu M$), which has a hydroxyl group at position 5. On target screening of the NP series indicates that flavonols are the most promising compounds, with NP-31 being the most active and selective hit (selectivity index: SI $hTS/TbPTR1 > 35$; SI $hDHR/TbPTR1 = 36$). Nevertheless, a clear structure–activity relationship (SAR) could not be established from this data set, and we could not reliably determine how the number and the pattern of hydroxyl/methoxy substituents influenced the activity.

The 38 compounds were also assessed for their inhibitory activity against *T. brucei* parasite (Figure 1, Supporting Information, Table S1). With the exception of NP-9, NP-10, NP-22, and NP-35, all the natural flavonoids screened showed antitrypanosomal activity with IC_{50} lower than $30 \mu M$. Figure 1 shows that there is no direct correlation between the antitrypanosomal activity of the NP compounds and their *PTR1* inhibition effect.

X-ray Crystallographic Studies of the Natural Products. To understand the flavonoid interactions into the protein binding site, the 18 compounds showing IC_{50} against *TbPTR1* lower than $150 \mu M$ (compounds NP-8, NP-10, NP-12, NP-13, NP-18, NP-21, NP-22, NP-23, NP-24, NP-26, NP-27, NP-28, NP-29, NP-31, NP-35, NP-36, NP-37, and NP-38; Figure 1) were selected for structural analysis. We obtained crystal structures of the *TbPTR1*–NADPH/NADP⁺ complexes for two moderately active natural products (NP-29 (datisctin) $IC_{50} TbPTR1 = 76.9 \mu M$ and NP-13 (hesperetin) $IC_{50} = 104.2 \mu M$) only. These compounds show inhibitory activity against *TbPTR1* and selectivity against at least one of the human proteins (TS and DHFR; Figure 1). Statistics for data collection and refinement are reported in the Supporting Information, Tables S2 and S3. In both cases, the crystal asymmetric unit contains the functional unit of the enzyme, the *TbPTR1* tetramer. The tertiary structure of the *TbPTR1* subunits is typical of the short-chain dehydrogenases/reductases (SDR) superfamily and shows a single α/β domain consisting of a seven-stranded parallel β -sheet sandwiched between two sets of α -helices.²⁹ The *TbPTR1* active site is mainly formed by a single chain, with one end blocked by the C-terminus of the partner subunit. In this L-shaped depression, the cofactor binds in an extended conformation entrapped by a network of highly conserved hydrogen bonds.²⁹ The substrate-binding loop (residues 207–215) interacts with both the cofactor and the substrate. Two surface-exposed parts of the sequence, residues 104–112 and 143–151, are usually poorly visible in *TbPTR1* crystal structures and they were not included in our models. The overall structure of each subunit of the *TbPTR1* complexes is the same as indicated by the small root-mean-square deviations (RMSDs) after $C\alpha$ superimposition of the four subunits in each tetramer (0.16–0.26 Å for NP-13 and 0.11–0.41 Å for NP-29). The binding of the cofactor is essential to create both the catalytic site and the substrate-binding pocket, indicating an ordered sequential reaction mechanism with NADPH binding before the substrate.³⁰ The

pterin moiety of substrates and of pterin-like inhibitors binds in a π -sandwich between the nicotinamide ring of NADPH/NADP⁺ and the aromatic side chain of Phe97.³¹ Pterin also interacts with the NADPH/NADP⁺ β -phosphate, and this interaction is strongly conserved in pterin-like inhibitors.³¹

***TbPTR1*–NADPH/NADP⁺–NP-29.** The chromen-4-one core of NP-29 binds in the biopterin binding pocket as described above. The ligand is supported by a network of H-bonds involving the cofactor and the surrounding residues in the active site cavity (see Figure 2A). The hydroxyl group at position 7 on the chromen-4-one of NP-29 (the NP-29 atom numbering is given in the inset in Figure 2A) donates a H-bond to the NADPH/NADP⁺ β -phosphate and accepts a H-bond from an amino group on Arg14. The hydroxyl group at position 5 accepts a H-bond from NADPH/NADP⁺ ribose OH₂, and donates a H-bond to the hydroxyl group of Tyr174, which donates a H-bond to the inhibitor carbonyl oxygen at position 4. Furthermore, the carbonyl oxygen and the hydroxyl group at position 3 of NP-29 are both H-bonded to a water molecule linked by a further H-bond to the side chain of Asp161. An additional water molecule, which is highly conserved throughout all *TbPTR1* structures, mediates the interaction between the position 3 hydroxyl group of NP-29 and the backbone carbonyl oxygen of Gly205 (not shown). The *o*-phenol moiety of NP-29 is located in a hydrophobic pocket of the *TbPTR1* active site cavity, surrounded by the side chains of Val206, Leu209, Pro210, Met213, and Trp221. The 2' hydroxyl group on this ring is close (~ 4 Å) to the carbonyl oxygen of Gly205 and forms an intramolecular H-bond (2.4 Å) with the hydroxyl group at position 3 of the chromen-4-one moiety.

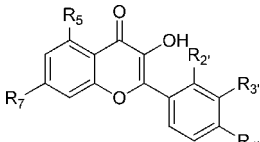
***TbPTR1*–NADPH/NADP⁺–NP-13.** The chromen-4-one core of NP-13 also binds in the biopterin binding pocket, but it is rotated by about 180° with respect to its orientation in the *TbPTR1*–NP-29 complex (Figure 2B). The ether oxygen at position 1 on the chromen-4-one moiety of NP-13 points toward the side chains of Asp161 and Tyr174. The hydroxyl group at position 7 of the NP-13 chromen-4-one is within H-bonding distance from the NADPH/NADP⁺ ribose OH₂, and the side chain of Ser95. The NP-13 hydroxyl group at position 5 accepts a H-bond from an amino group of Arg14 and donates a H-bond to the backbone carbonyl oxygen of Leu208. The *o*-methoxy-phenol (*o*-guaiacol) ring in position 2 on the chromen-4-one makes a T-shaped stacking interaction with the aromatic side chain of Trp221. Furthermore, the 3' phenolic moiety is within H-bonding distance from the Asp161 side chain and forms two water-mediated interactions with the backbone nitrogen of Cys168 and the amide nitrogen of Asn175 (not shown). The methoxy oxygen at position 4' interacts with the sulfur of the modified Cys168 (S-oxy-cysteine), whereas the terminal methyl makes van der Waals contacts with the side chains of Trp221 and His267 of the partner subunit.

In summary, structural comparison of the two *TbPTR1* complexes reveals two opposite orientations of the natural flavonoid inhibitors NP-13 and NP-29 in the biopterin binding pocket in which the chromen-4-one substituents interact with different residues surrounding the cavity. A double binding mode was previously observed for pteridine-like inhibitors and MTX.¹⁵

Flavonoid Library Design and Synthesis. The crystal structures showed that flavonoids occupy the biopterin binding site of *TbPTR1*, suggesting that the flavonol moiety could provide a new scaffold for the development of *PTR1* inhibitors.

The flavonoids selected as starting points for the development of compounds to treat trypanosomatidic diseases are depicted in Table 1. Even though the two crystal structures showed that

Table 1. Chemical Structures and IC₅₀ Values for Selected Compounds from the Screened Natural Products Library



compd	R ₅	R ₇	R _{2'}	R _{3'}	R _{4'}	IC ₅₀ <i>Tb</i> PTR1 (μM)	IC ₅₀ <i>T. brucei</i> (μM)
NP-27	H	H	H	H	H	12.8	9.1
NP-28	OH	OH	H	H	OH	28.0	9.0
NP-29	OH	OH	OH	H	H	76.9	27.0
NP-31	OH	OH	H	OCH ₃	OH	13.9	11.2

the hydroxyl group at position 5 could establish H-bonds with the protein and the cofactor, the inhibitory data on natural flavonoids (Figure 1) suggested that a hydroxyl group in this position leads to a decrease in the inhibitory activity against *Tb*PTR1. Therefore, we decided to synthesize compounds with hydroxyl substituents at positions 6 or 7 of the chromenone A-ring because, according to the crystal structures, these should be able to establish H-bonds with the NADPH/NADP⁺ ribose and the side chain of Ser95 or with the NADPH/NADP⁺β-phosphate and Arg14 (Figure 3). Moreover, we performed modifications at positions 3', 4', and 5' on ring B to explore the SAR. As shown in Figure 3, hydroxyl groups in positions 3', 4', and 5' could form a H-bonding interaction with the Asp161 side chain and two water-mediated interactions with the backbone nitrogen of Cys168 and the amide nitrogen of Asn175. Many natural products have both hydroxyl and methoxy groups, thus we synthesized and biologically evaluated eight hydroxylated (1–8) and eight methoxylated (9–16) compounds aiming to investigate how the number and the

pattern of hydroxyl/methoxy substituents influenced the activity.

The synthesis of compounds 1–16 is depicted in Scheme 1. All the compounds have been already reported in literature, but as far as we know, they have never been proposed as PTR1 inhibitors. The intermediate chalcones (17–24) were synthesized by Claisen–Schmidt condensation using substituted acetophenones and benzaldehydes in the presence of NaOH as the base in ethanol. The reaction was carried out as previously reported in literature for similar compounds.²⁶ Afterward, the chalcones (17–24) were converted into the corresponding methoxylated flavonols (9–16), using the Flynn–Algar–Oyamada method for epoxidation and subsequent intramolecular cyclization of the open-chain structure.²⁵ The reaction was performed with hydrogen peroxide in aqueous base (1 M NaOH). Cleavage of methoxy protecting groups with boron tribromide gave the hydroxylated flavonols 1–8 in high yield. All synthesized compounds were characterized by ¹H NMR, ¹³C NMR, and mass analysis. The obtained NMR and mass data are reported in the Supporting Information (pp 33–37) and were compared with those available in literature.

Target Compound Profile. In our study, a panel of assays was performed and activity/toxicity data were considered during the hit selection/prioritization.³² Together with the potency data (measured as IC₅₀, i.e., the compound concentration producing 50% reduction of targeted enzyme activity and parasite cell growth), we considered the selectivity index (IC₅₀ toward the parasite compared with compound cytotoxicity IC₅₀ on host cells) and early toxicity properties. We have assessed the entire profile of the 16 synthesized compounds, and the whole data panel has been evaluated before selecting compounds for pharmacokinetic studies.

Testing of Synthetic Flavonols against PTR1 Enzymes. All flavonols were investigated for their activity toward *T. brucei* (*Tb*PTR1) and *L. major* PTR1 (*Lm*PTR1). The results are shown in Figure 4 and Supporting Information, Table S4. All hydroxylated compounds except compound 5 showed significant inhibitory activity against *Lm*PTR1 and *Tb*PTR1 at 50 μM concentration, with compound 2 being the most potent.

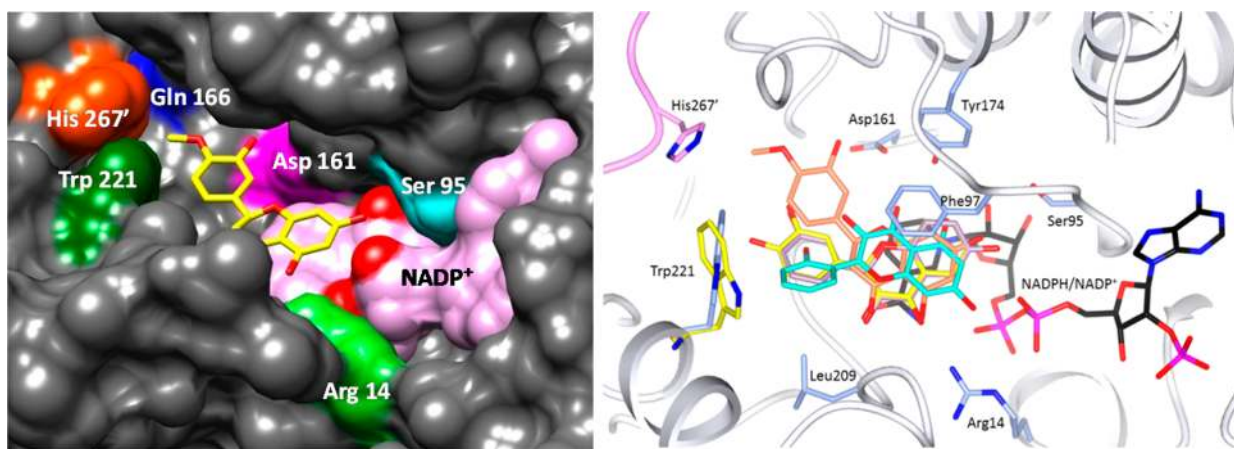
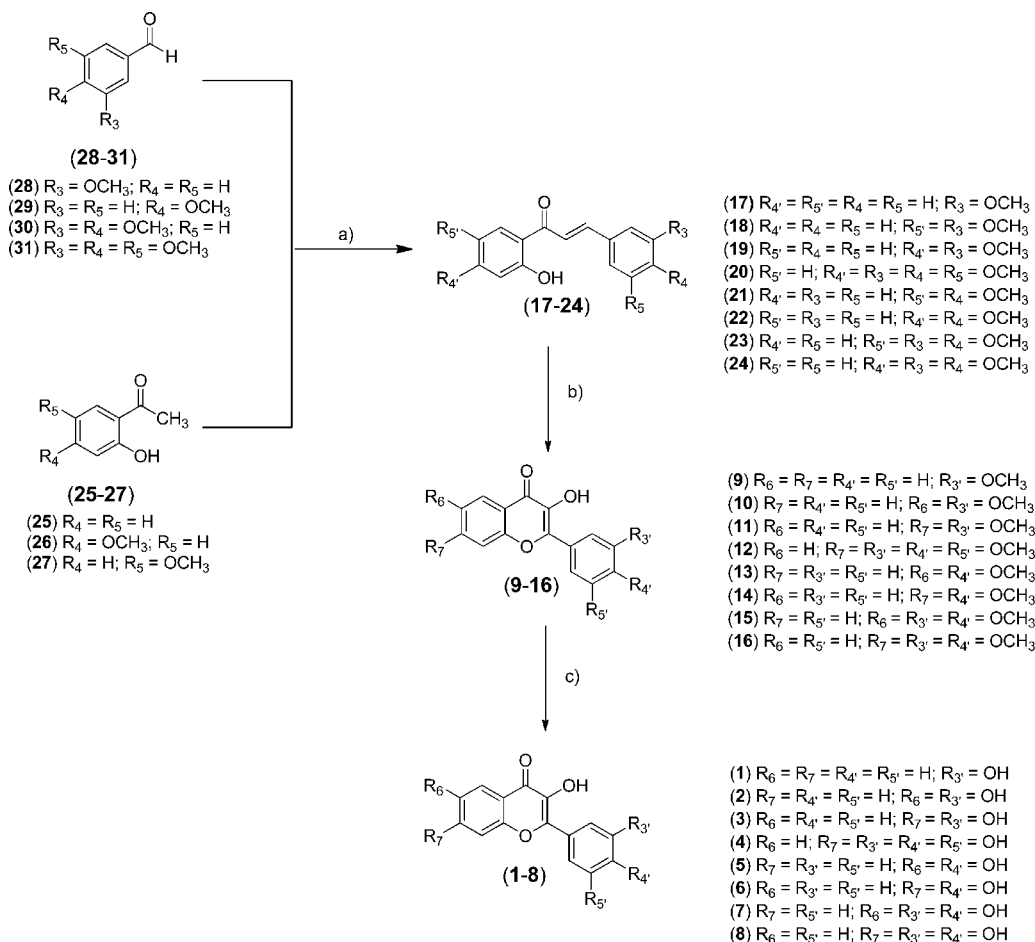


Figure 3. Design of the synthetic library (left). NP-13 (in yellow) into *Tb*PTR1 (in gray). The amino acids involved in the interactions of the designed compounds are shown in different colors. For clarity reasons, two residues involved in the interactions are not shown (Cys168 and Asn175). Superimposition of the four crystal structures of *Tb*PTR1 (right) (chain A gray ribbon; chain D pink ribbon; relevant active site residues as light-blue sticks) in complex with NADPH/NADP⁺ (sticks, black carbon atoms) and the inhibitors (stick) NP-13 (orange), NP-29 (cyan), compound 2 (lilac), and compound 7 (yellow). The three different binding modes adopted by the ligands can be appreciated as well as the movement of Trp221 (yellow sticks) upon binding of compound 7.

Scheme 1. Synthesis of the Flavonols 1–16^a

^aReaction conditions: (a) NaOH (3 M), EtOH, rt; (b) H₂O₂, NaOH (1 M), EtOH, rt; (c) BBr₃ (1 M in dry DMC), dry DMC, 0 °C → rt.

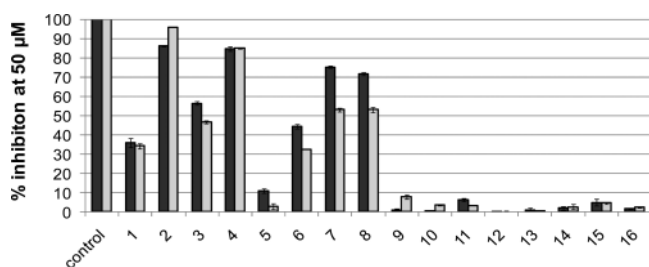


Figure 4. Inhibitory activity against *TbPTR1* (in gray) and *LmPTR1* (in black). The control compound was pyrimethamine, a PTR1 inhibitor (100% inhibition at 50 μM against both PTR1 enzymes).

Methylation resulted in almost complete loss of inhibitory activity. The most noteworthy examples are represented by two pairs: compounds **10** and **2** (4% and 96% inhibition of *TbPTR1* at 50 μM, respectively) and compounds **12** and **4** (no inhibition of *TbPTR1* at 50 μM and 85% inhibition at 50 μM, respectively). The introduction of a hydroxyl group on ring A in compound **2** led to an increase of the inhibitory potency against both enzymes of almost 3-fold with respect to compound **1**, which bears an unsubstituted ring (34% inhibition of *TbPTR1* at 50 μM). To explain a complete SAR, X-ray crystallographic studies and docking analyses were carried out. The compounds were evaluated also toward *TcPTR2*, and none of the compounds significantly inhibited *TcPTR2* activity (data not shown).

Crystal Structures of Synthetic Flavonoid–*TbPTR1* Complexes. Compounds **2** and **7**, which show inhibitory activity against *TbPTR1*, were submitted to X-ray crystallographic characterization aimed at comparing their binding poses in the PTR1 active site with those of the natural flavonoids and thus gaining further information for compound optimization.

***TbPTR1*-NADPH/NADP⁺-2.** The structure of *TbPTR1* in complex with NADPH/NADP⁺ and compound **2** was determined at 1.38 Å resolution. Ligand placement and key interactions within the active site cavity are reported in Figure 2C. Compound **2**, like NP-13 and NP-29, binds in the substrate binding pocket, with the chromen-4-one in a π -sandwich interaction between the nicotinamide ring of NADPH/NADP⁺ and the aromatic side chain of Phe97. The chromen-4-one core of compound **2** adopts the binding mode observed for NP-13, with the oxygen atom at position 1 directed toward the side chain of Asp161. The R6 hydroxyl group on the chromen-4-one is within H-bonding distance from the hydroxyl group of Ser95 and the NADPH/NADP⁺-phosphate oxygen. The carbonyl oxygen at position 4 completes the interactions made by the chromen-4-one by receiving two H-bonds from an amino group of Arg14 and a water molecule that is H-bonded to the NADPH/NADP⁺-phosphate O2. These interactions, together with the stacking of the chromen-4-one described above, bring its 3 position hydroxyl group within H-bonding distance from the backbone

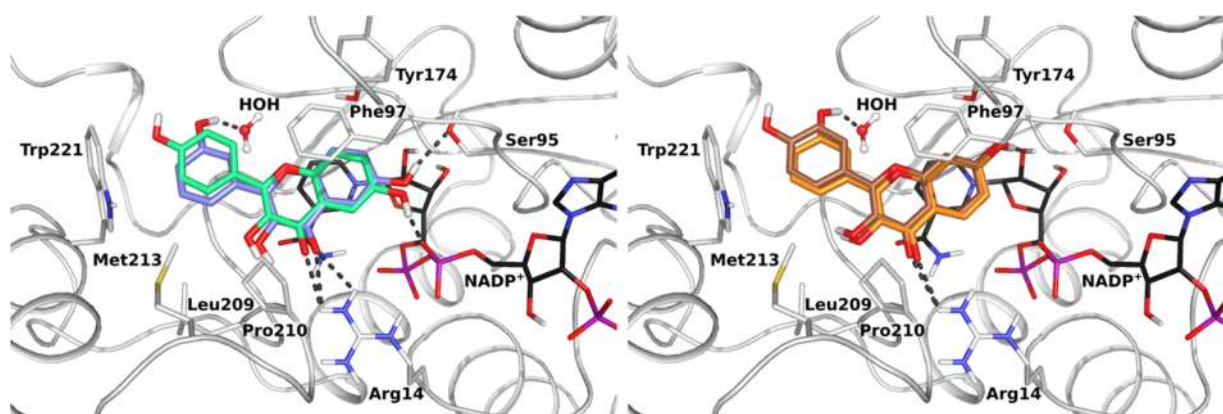


Figure 5. Comparison of the effects of hydroxyl substituents at the R6 (left) and the R7 (right) positions of ring A on the binding of the synthetic flavonoids to *TbPTR1*. Superimposition of constraint docking poses for compound 2 (in sticks, lilac carbons) and compound 5 (in sticks, pale-green carbons) (left) and compound 3 (in sticks, orange carbons) and compound 6 (in sticks, brown carbons) (right) in *TbPTR1* (in cartoon with interacting residues in sticks with gray carbons) in complex with NADPH/NADP⁺ (in sticks, black carbon atoms). A conserved water molecule is shown in ball-and-stick representation. Hydrogen bonds are indicated by dark-gray dotted lines.

carbonyl oxygen of Leu208. The phenyl ring in position 2 on the chromen-4-one establishes a T-shaped stacking interaction with the aromatic side chain of Trp221 and forms van der Waals interactions with the side chains of Val206 and Leu209 (not shown). The *meta*-hydroxyl group at the 3' position on the phenyl moiety of 2 is only involved in a water mediated H-bond network linking it to the backbone carbonyl of Gly205 and the side chain of Asp161 (not shown).

***TbPTR1*-NADPH/NADP⁺-7.** The structure of *TbPTR1* in complex with NADPH/NADP⁺ and compound 7 has been determined at 1.76 Å resolution. Ligand placement and key interactions within the active site cavity are shown in Figure 2D, where it can be observed that compound 7 adopts exactly the same pose as compound 2, making the same interactions with the enzyme and cofactor. The only difference consists in the steric repulsion of the additional hydroxyl group at position 4' that forces the side chain of Trp221 to move away by about 1.4 Å. The 4' hydroxyl group is not involved in H-bonds but is solvent exposed and presumably can form a H-bond with water.

The superposition of the four crystal structures is shown in Figure 3. The orientation of compounds 2 and 7 in the *TbPTR1* cavity is the same as for NP-13, with the inhibitor bicyclic core directed toward the opposite side of the cavity with respect to NP-29. Due to the larger steric hindrance of the substituents on the 2-*o*-methoxy-phenolring of NP-13, a little rearrangement of the whole molecule occurs with a rotation of about 35° of the bicyclic core with respect to 2 and 7 that allows the side chain of Trp221 to remain in the position observed in the *TbPTR1*-2 complex (Figure 2C). The rotation of NP-13 allows direct H-bonding between the 3' hydroxyl group of NP-13 and Asp161, while the interaction of the corresponding group of 2 and 7 is mediated by a water molecule. The displacement of the Trp221 side chain occurring upon binding of compound 7 can be appreciated (Trp221 as yellow sticks in Figure 3, right).

A puzzling aspect of all four crystal structures is that the different inhibitors have been systematically observed bound to three of the four *TbPTR1* subunits, whereas the NADPH/NADP⁺ cofactor is found bound to all subunits in three instances out of four, although with lower occupancy in those subunits where the inhibitor is absent. The absence of the inhibitor is correlated with larger disorder in the substrate binding loop that brings Trp221 into the active site cavity.

However, this loop always shows temperature factors higher than average, even in the subunits where both cofactor and inhibitor are bound. The disorder of this region is independent of the crystal packing because, when the loop is disordered (e.g., in subunit C in *TbPTR1*-NP-29), it approaches the symmetry-related subunits in the same way as the ordered loop (e.g., in subunit A in *TbPTR1*-NP-29) located on the opposite face of the *TbPTR1* tetramer. Confirming this observation, the other two ordered loops (e.g., in subunits B and D in *TbPTR1*-NP-29) do not make close intermolecular contacts. We can conclude that stabilization due to the π -stacking between the R2-aromatic ring of the inhibitor and the side chain of Trp221 is critical for the ordering of the substrate binding loop.

Computational Docking of Synthetic Compounds to *TbPTR1* and *LmPTR1* and SAR Analysis. Docking studies were conducted to further expand the SAR to all the hydroxylation/methoxylation patterns in the synthesized compounds. In addition, computational docking allowed the evaluation of possible binding modes and a corresponding SAR for *LmPTR1*, for which no crystal structure with these compounds could be solved.

Like the crystallographic studies, computational docking showed multiple possible binding modes for the chromen-4-one system, which roughly group into three classes: (1) a typical substrate-like binding mode involving H-bonding with the NADPH/NADP⁺ cofactor and Ser95 of *TbPTR1* (or Ser111 of *LmPTR1*), as observed in the complexes of compounds 2, 7, and NP-13; (2) an alternative binding mode with chromen-4-one flipped by roughly 180° as observed in the complex with NP-29; and (3) an inverse binding mode solely observed in docking studies, with ring B rather than the chromen-4-one in a stacking orientation between the NADPH/NADP⁺ nicotinamide and Phe97 of *TbPTR1* (or Phe113 of *LmPTR1*). While three of the four crystal structures feature binding mode (1), a general dependence of the binding mode on the substituent pattern is notable both from crystallographic and docking studies. In docking, many of the observed binding modes were found to have highly similar contacts and H-bonding partners, thus creating a favorable entropic contribution to the binding of this compound class (see also Supporting Information, Tables S5 and S6). However, due to their large number of H-bond donors/acceptors, flavonoids can adopt

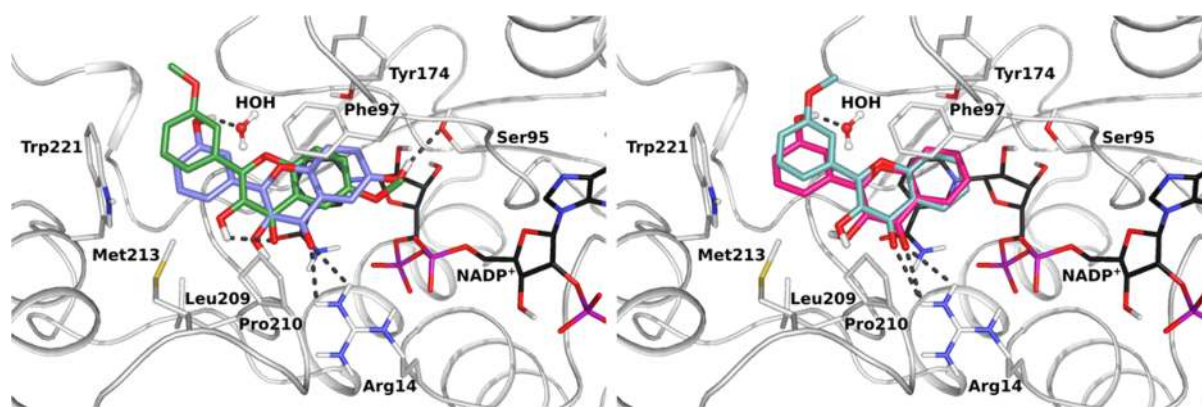


Figure 6. Comparison of the effects of hydroxyl and methoxy substituents at the R6 (left) and the R3' (left and right) positions on the binding of the synthetic flavonoids to *TbPTR1*. Constraint docking poses are shown for compound 2 (in sticks, lilac carbons), compound 10 (in sticks, dark-green carbons) (left), compound 9 (in sticks, pale-cyan carbons), and compound 1 (in sticks, magenta carbons) (right) in *TbPTR1* (in cartoon with interacting residues in sticks with gray carbons) in complex with NADPH/NADP⁺ (in sticks, black carbon atoms). A conserved water molecule is shown in ball-and-stick representation. Hydrogen bonds are indicated by dark-gray dotted lines.

additional binding modes with different, often water mediated, interactions. We therefore used a ligand constraint docking approach to favor arrangements close to the crystal complexes to assess the potential of other synthetic derivatives to bind in similar orientations.

The crystal structures and docking indicate that hydroxylation patterns on ring A influence the orientation of chromen-4-one in the pocket. Precisely, compounds hydroxylated at position R6 show different H-bonding partners (mostly NADPH/NADP⁺ phosphate and Ser95/Ser111 of *TbPTR1*/*LmPTR1*, respectively) compared with those hydroxylated at R7 (mostly NADPH/NADP⁺ ribose and Tyr174/Tyr194 or *TbPTR1*/*LmPTR1*, respectively), see Figure 5. This difference also affected the compound activity (compare R6-hydroxylated compound 2 (96% *TbPTR1* inhibition at 50 μ M) and the corresponding R7-hydroxylated compound 3 (47% *TbPTR1* inhibition at 50 μ M)). While the geometry of compound 2 fits ideally in the *TbPTR1* binding pocket, the slight reorientation observed for compound 3 leads to a loss of H-bonds and stabilizing hydrophobic contacts, halving the compound activity. Thus, hydroxylation of ring A can fine-tune the arrangement of the chromen-4-one system within the pocket and critically influence the interaction pattern of rings B and C.

Methoxylations, in contrast to hydroxylations, were found to lead to activity loss. For example, the inhibitory activity against *TbPTR1* and *LmPTR1* drops substantially when comparing compound 2 with the corresponding methoxylated compound 10 (96% and 4% *TbPTR1* inhibition at 50 μ M; 86% and no *LmPTR1* inhibition, respectively). Methoxylations on the A-ring cause a displacement of the chromen-4-one from the primary stacking geometry to accommodate the bulkier methoxy group in close proximity to the NADPH/NADP⁺ cofactor and Ser95 of *TbPTR1* (Ser111 of *LmPTR1*), see Figure 6, left. Notably, this adverse effect was also apparent in the comparison of compounds 1 and 9, bearing no substitution on ring A but a hydroxylation or methoxylation on R3' of the B-ring. Whereas the activity of compound 9 is negligible, compound 1 shows about 35% inhibition against both *TbPTR1* and *LmPTR1* (at 50 μ M). A H-bond donor functionality on R3' has an important stabilizing effect, as it can interact with a water molecule bridging Asp161 and Gly205 of *TbPTR1* (see Figure 6, right) or Asp181 and Gly225 of *LmPTR1*. As this water, according to our WatCH analysis, is almost 100%

conserved in both *TbPTR1* and *LmPTR1*, it provides a clear advantage for R3' hydroxylated compounds. This is further supported by the crystal structures of *TbPTR1* with compounds 2 and 7, where the R3' hydroxylation is in H-bonding distance to this water site. In agreement with this observation, compound 2 with the *m*-hydroxyphenyl was over 30 times more active than compound 5 with a *p*-hydroxyphenyl ring (96% and 3% *TbPTR1* inhibition at 50 μ M, respectively). In contrast to a *meta*-hydroxylation, a *para*-hydroxylation cannot interact with the structural water, see Figure 5. Again, this effect was sensitive to the hydroxylation pattern of ring A, as can be observed by comparing compounds 3 (R3' and R7 hydroxylation, 47% *TbPTR1* inhibition at 50 μ M) and compound 6 (R4' and R7 hydroxylation, 33% *TbPTR1* inhibition at 50 μ M), see Figure 5. While *p*-substituted compounds generally show a drop in inhibitory activity compared to their respective *m*-substituted counterparts, the effect is weaker for the second pair with the R7 hydroxylation on ring A, possibly because R7 substitutions lead to a slight rotation of the stacking chromen-4-one, directing the *p*-hydroxyl toward a more solvent-exposed region of the pocket. In conclusion, the effects of individual hydroxyl substituents are not purely local and do not show a completely additive effect; rather, the combinations of ring A and B hydroxylations will determine the final binding mode.

On the basis of the binding modes observed for *TbPTR1*, we evaluated possible binding orientations to *LmPTR1* by docking. Notably, all compounds with R4' hydroxylation (compounds 5 and 6), compounds with R3' and R4' hydroxylations (compounds 7 and 8) and one compound with R3' hydroxylation combined with an R7 substitution on ring A (compound 3) were found to be slightly more active toward *LmPTR1* than *TbPTR1*. While the depth of the biopterin binding pocket is well conserved and the interactions for chromen-4-one are practically identical in both proteins, there are notable differences in the opening of the pocket that can explain the observed activity differences (see sequence alignment of interaction partners in Supporting Information, Figure S2). While in *TbPTR1*, R3' hydroxylations on the B-ring mainly come into contact with the conserved water molecule and Cys168 (see Figure 5) and R4' hydroxylated compounds can hardly form direct H-bonds with the receptor, *LmPTR1* generally has a more polar opening part of the pocket. Three

major differences are crucially important: His241, Tyr283, and Arg287 from the neighboring subunit in *Lm*PTR1, correspond to Trp221, Leu263, and His267 from the neighboring subunit in *Tb*PTR1, thereby providing several additional polar contact points. As the side chain of Arg287 in *Lm*PTR1 is much bigger than that of His267 in *Tb*PTR1, arginine is much more accessible to the ligands than histidine. Furthermore, as observed in crystal structures of *Tb*PTR1, Trp221 is rather motile and different rotamers lead to an open or a rather closed form of the pocket, thereby potentially blocking ligand access to His267. While Trp221 of *Tb*PTR1 did not provide a H-bonding contact to any of the studied compounds, induced fit docking studies of the compounds to *Lm*PTR1 showed movement of His241 to allow additional H-bonding interactions in the most stable structure, as also shown in Figure 7 (see also Supporting Information, Table S5). Overall,

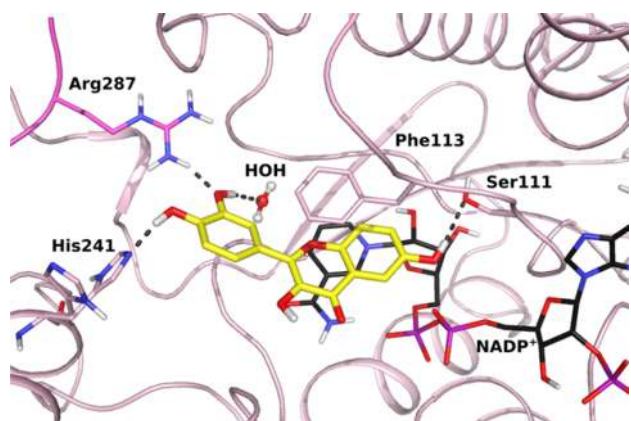


Figure 7. Superimposition of the crystal structure of *Lm*PTR1 (PDB ID 1E92) in cartoon representation and interacting residues in sticks representation. Chains A and D (containing Arg287) are colored in pale-pink and magenta, respectively) and the best predicted receptor conformation obtained in the induced-fit docking study starting from this crystal structure (His241 in H-bonding contact to compound 7) in complex with NADPH/NADP⁺ (in sticks, black carbons) and compound 7 (in sticks, yellow carbons). A conserved water molecule is shown in ball-and-stick representation. Hydrogen bonds are indicated by dark-gray dotted lines.

π - π stacking and hydrophobic contacts are clearly the main stabilizing factors for flavonoids in *Tb*PTR1, whereas a tendency toward a higher number of H-bond donor/acceptor contacts may stabilize hydroxylations in both the *meta*- and *para*-positions of ring B slightly better in *Lm*PTR1.

While addition of a *para*-hydroxylation for the R7 substituted ring A did not lead to a significant change in activity against *Tb*PTR1 (compound 3, 47%; compound 8, 53% *Tb*PTR1 inhibition at 50 μ M), compound 4 with hydroxylations at R3', R4', and R5' yielded a clear activity increase (85% *Tb*PTR1 inhibition at 50 μ M). Considering the lack of direct polar contact points in *Tb*PTR1, this is surprising and there may be two possible explanations: (1) bridging water molecules may enable additional contacts for the second *meta*-hydroxylation, e.g., with Trp221, or (2) the compound may adopt an inverted binding mode, placing the pyrogallol moiety in a stacking orientation with Phe97 and the cofactor, where it could make strong H-bonds and thereby keep the more hydrophobic chromen-4-one in the hydrophobic *Tb*PTR1 subpocket, where it is able to form additional hydrophobic contacts.

Testing of Synthetic Flavonols against *Tb*DHFR, *Lm*DHFR, hDHFR, and hTS. The compounds showing inhibitory activity against PTR1 (1–8) were evaluated for their activity against *Tb*DHFR, *Lm*DHFR, and hDHFR. The results are reported in the Supporting Information, Table S7. With the exception of compounds 1 and 2, all the tested compounds had inhibitory activities against *Tb*DHFR and *Lm*DHFR below 25% at 50 μ M. Compounds 1 and 2 slightly inhibit, respectively, *Tb*DHFR and *Lm*DHFR (42–43% at 50 μ M). Therefore, we can conclude that compounds 1–8 have low inhibitory activity against the parasitic DHFRs. With the exception of compound 2, all the compounds had inhibitory activities against hDHFR below 30% at 50 μ M. The IC₅₀ of compound 2 against hDHFR is 50 μ M and against *Tb*PTR1 is 4.3 μ M, thus compound 2 has a selectivity index higher than 10. None of the tested compounds inhibited hTS at 50 μ M (data not shown).

Early Toxicity Studies. To characterize the compounds properties and to define their toxicological profile, we used an assay panel typical of those employed in drug discovery projects for hit/lead selection.³² Our panel includes hERG, five cytochrome P450s, (CYP1A2, CYP2C9, CYP2C19, CYP2D6, and CYP3A4), Aurora B kinase, A549 (human lung adenocarcinoma epithelial) and WI-38 (fetal lung fibroblasts) cell lines and mitochondrial toxicity. The results of the screening are reported in Figure 8 as a traffic light code

compound	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	i. comp.
% Inhibition hERG	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition CYP1A2	Red	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition CYP2C9	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition CYP2C19	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition CYP2D6	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition CYP3A4	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition Aurora B	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Cell Growth A549	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Cell Growth WI-38	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Tox Mitochondria	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green
% Inhibition <i>T. brucei</i>	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green	Green

Figure 8. Early toxicity properties combined with inhibitory activity against *T. brucei*. The data are reported as a traffic light system. An ideal compound (I. comp.) should have all the parameters green. The cells are colored in green when the percentage of inhibition of *T. brucei* and the percentage of A549 and WI-38 cell growth is between 60 and 100, while the percentage of inhibition of CYP isoforms, hERG, Aurora B kinase and mitochondrial toxicity is between 0 and 30. Cells are colored in red when data indicates toxicity or inactivity. Yellow stands for a borderline value (30–60%): moderately active or slightly toxic compound. Gray: not tested.

(from red through yellow to green for increasing biological effect and decreasing toxicity of the compounds) and in Supporting Information, Table S8. An ideal lead compound should have all parameters colored in green. In detail, the percentage of mitochondrial toxicity and of inhibition toward hERG, CYP isoforms, and Aurora B kinase should be below 30%, while the percentage of A549/WI-38 cells growth, measuring cytotoxicity, should be above 70%. A few compounds showed a percentage of inhibition toward hERG > 10% at 10 μ M. Apart from compounds 4 and 8, all the compounds inhibited at least one isoform of cytochrome P450 (mostly CYP1A2), while only two compounds (4 and 8) exhibited a significant inhibitory activity against Aurora B kinase

at 10 μM . None of the compounds was either cytotoxic or showed mitochondrial toxicity.

Testing of Synthetic Compounds on Cultured Parasites. With the exception of compound 8, all the molecules were tested at 10 μM against *T. brucei* bloodstream form, intracellular *T. cruzi*, and *L. infantum* intracellular amastigotes (Figure 9). Almost all tested compounds were active against *T.*

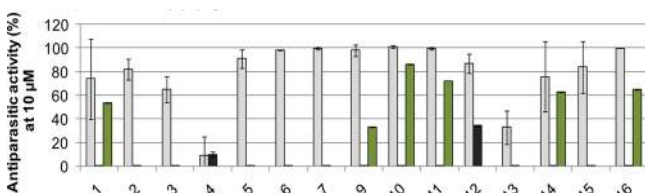


Figure 9. Antiparasitic activity of the synthesized compounds against *Trypanosoma brucei* (in gray), *Trypanosoma cruzi* (in green), and *Leishmania infantum* (in black) at 10 μM . The reference compounds were pentamidine ($\text{IC}_{50} = 1.55 \pm 0.24 \text{ nM}$) for *T. brucei*, miltefosine ($\text{IC}_{50} = 2.65 \pm 0.4 \mu\text{M}$) for *L. infantum*, and nifurtimox ($\text{IC}_{50} = 2.2 \pm 0.4 \mu\text{M}$) for *T. cruzi*.

brucei, while only two compounds, 4 and 12, were slightly active against *L. infantum*, showing inhibitory activities of 10 and 35%, respectively. Twelve compounds (1, 2, 5–12, 14–16) presented EC_{50} values against *T. brucei* between 1 and 8 μM (Table 2), while compounds 3, 4, and 13 showed EC_{50}

Table 2. EC_{50} against *T. brucei*, NOAEL, and Selectivity Index of the Synthesized Compounds^a

compd	EC_{50} <i>T. brucei</i> (μM) \pm SD	$\text{CC}_{50} \pm$ SD or NOAEL	selectivity index ($\text{CC}_{50}/\text{EC}_{50}$)
1	5.18 \pm 1.10	20	4
2	7.56 \pm 0.51	53 \pm 2	7
3	12.29 \pm 2.82	80 \pm 2	6
4	18.04 \pm 0.50	100	5
5	2.32 \pm 0.42	20	7
6	4.29 \pm 0.71	20	4
7	1.36 \pm 0.57	10	6
9	2.32 \pm 1.01	10	4
10	1.43 \pm 0.42	10	7
11	1.14 \pm 0.24	10	8
12	1.17 \pm 1.07	20	17
13	20.12 \pm 2.27	20	1
14	1.10 \pm 0.47	10	9
15	2.02 \pm 0.51	10	5
16	2.99 \pm 1.86	25	8
pentamidine	0.00155 \pm 0.00024	10	6440

^a EC_{50} and NOAEL represent the arithmetic average of at least two independent determinations done in triplicate.

values of 12.29, 18.04, and 20.12 μM , respectively. Only the methoxylated compounds 10, 11, 14, and 16 showed antiparasitic activity toward *T. cruzi* trypomastigotes similar to that of nifurtimox (NFX), the reference drug for the treatment of Chagas' disease. Compound 10 is the most active ($\text{EC}_{50} = 4.5 \mu\text{M}$, NFX $\text{EC}_{50} = 1.6 \mu\text{M}$).

The series was assessed for cytotoxicity on THP1 macrophage-like cells to evaluate the NOAEL (no observed adverse effect level). The reference compound was pentamidine with a NOAEL of 10 μM and a selectivity index of 6440. The selectivity indexes of our compounds, given by the ratio

between the EC_{50} toward *T. brucei* and CC_{50} toward THP1, were lower than 10 with the exception of compound 12 (SI = 17) (Table 2). The selectivity index acceptable for a compound to be considered non toxic³³ is at least 10, thus our compounds showed slight toxicity.

Combination Studies and Evaluation of Synergy. All compounds were evaluated in combination with MTX independently of their PTR1 inhibitory activity against the recombinant protein. We aimed to assess the potential gain in potency and the reduction in toxicity through the combination of our compounds with MTX, a well-known DHFR inhibitor. In trypanosomatids, MTX is expected to exert a synergistic behavior in combination with PTR1 inhibitors. Moreover, MTX can show a synergistic effect in combination with non-PTR1 inhibitors through a multitarget inhibition. Preliminary combination experiments in *T. brucei* were conducted by fixing MTX at 4 μM (MTX average EC_{50} against *T. brucei*) and varying the concentration of the selected compounds between 1.25 and 20 μM (Supporting Information, Table S10). The synergy coefficients of all the compounds tested at the different concentrations (1.25, 2.5, 5, 10, 20 μM) are reported in Supporting Information, Table S11. Eight compounds presented a synergistic effect (synergy coefficient >1) when combined with MTX. The antiparasitic activity against *T. brucei* of the eight compounds both alone and in combination with MTX is shown in Figure 10. Compounds 5, 6, and 13 consistently presented the highest synergy coefficients (maximum synergy coefficient >1.5). Compound 13 was the most synergic with a calculated maximum synergy coefficient of 3.1 at 1.25 μM (Supporting Information, Table S11). Since it showed the highest synergy coefficient together with a safe toxicological profile (Figure 8, the only poor (red) parameter is for CYP1A2 (72% inhibition at 10 μM)), it was selected for further synergistic combination studies.

To better evaluate the gain in potency of the combination MTX–13, we employed the software Compusyn and a constant ratio between the concentrations of the two compounds chosen from their EC_{50} values (EC_{50} 13 = 20 μM ; EC_{50} MTX = 15; constant ratio = 1.3).³⁴ This enabled us to quantify synergism between the two compounds (combination index, CI) and to determine the dose reduction (dose reduction index, DRI) needed to observe the studied effect in the combination compared with the dose of each drug alone. The combination of MTX and compound 13 at the EC_{50} concentration had a CI of 0.174 ± 0.047 . When CI is lower than 1, the combination can be considered synergistic, therefore the observed CI shows strong synergism and confirms the initial data (Supporting Information, Table S10). At higher EC values, CI becomes lower, indicating very strong synergism (Table 3). The EC_{50} determined for the combination showing CI of 0.147 was $3.15 \pm 0.33 \mu\text{M}$. This enabled an average DRI at the EC_{50} of 14.6 ± 3.7 for MTX and of 11.7 ± 2.7 for compound 13 (IC_{50} MTX = 1.3 μM and IC_{50} 13 = 1.7 μM) (Supporting Information, Figure S3C). At the EC_{90} of the mixture, the predicted DRI is over 100-fold for both compounds when compared with the EC_{50} of the compounds alone. The isobologram is reported in Supporting Information, Figure S3. The toxicity of the MTX–13 mixture on THP1 was determined at three different concentrations (12.5, 25, 50 μM), and no synergistic toxicity on human cells was observed (Supporting Information, Figure S4 and Table S12). Therefore, the identified combination MTX–13 is selective for the inhibition of the parasite growth, while there is no synergy in

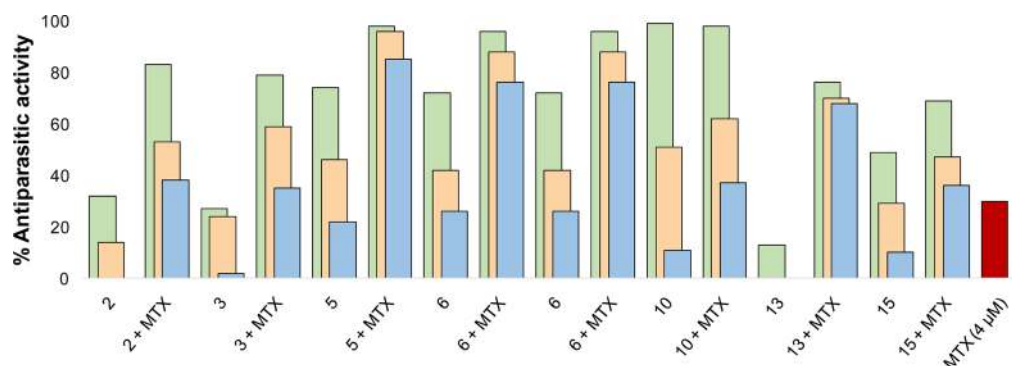


Figure 10. Trypanocidal activity of eight synthesized compounds alone and in combination with MTX. The compounds were tested at three different concentrations: 5 μM (in green), 2.5 μM (in light-pink), 1.25 μM (in light-blue). Antiparasitic activity of MTX is shown in dark-red.

the toxicity toward the host cells. These data together with the predicted DRI indicate that the combination improves the selectivity indexes for both MTX and compound 13.

Pharmacokinetic Studies, in Vivo Assays, and Compound Delivery. Compound 2 was selected for pharmacokinetic studies because it was the most active compound against *TbPTR1* ($\text{IC}_{50} = 4.3 \mu\text{M}$) that showed activity against the parasite *T. brucei* ($\text{EC}_{50} = 7.6 \mu\text{M}$) and it presented a safe profile, see Figure 8. The hERG IC_{50} (99.3 μM) was more than 20-fold higher than the target IC_{50} . With the exception of the CYP1A2 IC_{50} (6.1 μM), the IC_{50} against the cytochrome isoforms was higher than 10 μM . The solubility of compound 2 was evaluated using UV–Vis spectroscopy. In vivo bioavailability and half-life of compound 2 were evaluated in BALB/c mice treated IV with 1 mg/kg based on its low solubility. The molecule showed a short half-life ($t_{1/2} = 7.6 \text{ min}$), and it was not detected in blood after 30 min (the plasma levels are shown in Supporting Information, Figure S4). With the aim of improving the plasma levels of compound 2, we used two different drug delivery systems: the molecule was encapsulated in PLGA nanoparticles and solubilized with hydroxypropyl- β -cyclodextrins. The encapsulation did not change the physical properties of the PLGA nanoparticles as reported in Supporting Information, Table S13. The unloaded and loaded NPs were characterized by dynamic light scattering (DLS) in terms of size, polydispersity index, and ζ potential. We evaluated the in vivo bioavailability of compound 2 encapsulated in nanoparticles in BALB/c mice (IV and per os administration) and formulated with cyclodextrins after per os administration to NMRI mice. From preliminary data, both formulations did not significantly increase the levels of compound 2 in plasma samples from mice (data not shown). However, the in vitro activity of compound 2 both in PLGA and in cyclodextrins solution was preserved when compared to the free compound while the toxicity on THP1 was diminished (Table 4).

CONCLUSION

This is the first study in which we have confirmed the potential of flavonols as PTR1 inhibitors. Apart from compounds 4 and 8, none of the 16 synthesized flavonols have previously been reported in literature for their antiparasitic activity.²² Twelve of these compounds show EC_{50} values against *T. brucei* below 10 μM . We performed combination studies with MTX and compound 13, a non-PTR1 inhibitor, which was the best-performing compound. Although the reason for the synergy is unknown, compound 13 is an interesting compound for drug development due to the synergistic behavior with MTX and the

low toxicity on host cells. Target identification studies could be carried out aiming to explain the mechanism of action. The synthesized compounds were also evaluated for the activity against *T. cruzi* trypomastigotes, and compound 10 turned out to have a potency comparable to that of nifurtimox, the drug currently used to treat Chagas' disease (EC_{50} compound 10 = 4.5 μM , NFX $\text{EC}_{50} = 1.6 \mu\text{M}$). We carried out early in vitro toxicity assays, and overall the library showed a satisfactory profile. Combining the activity/toxicity data, we selected compound 2 for pharmacokinetic studies and we observed a very quick turnover of the molecule in BALB/c mice. Neither encapsulation in PLGA nanoparticles nor solubilization with cyclodextrins increased the plasma level of compound 2; however, both formulations allowed to maintain the in vitro activity of the compound and to reduce the toxicity. Our crystal structures and SAR analysis provide a basis for structure-based drug design aimed at identifying novel flavonol-like compounds with increased potency and selectivity toward PTR1 and able to overcome the problems of classical flavonols.

EXPERIMENTAL SECTION

Preparation of the Natural Products Library. A library of 98 natural compounds, consisting of 8 flavanones, 19 flavones, 20 flavonols, 2 dihydroflavonols, 13 anthocyanins, 2 catechins, 2 isoflavones, 4 chalcones, 18 phenolic acids and derivatives, 1 aurone, 5 anthraquinones, 3 triterpenes, and 1 phloroglucinol was purchased from Extrasynthese (Genay, France) and Sigma-Aldrich-Fluka (Milan, Italy) for drug screening. The degree of purity of all these molecules was checked by HPLC. The HPLC analyses were performed on an Agilent Technologies (Waldbronn, Germany) modular model 1100 system, consisting of a vacuum degasser, a quaternary pump, an autosampler, a thermostated column compartment, and a diode array detector (UV/DAD). The chromatograms were recorded using an Agilent Chemstation for LC and LC-MS systems (Rev. B.01.03). An Ascentis C_{18} column (250 mm \times 4.6 mm I.D., 5 μm , Supelco, Bellefonte, PA, USA) was used, with a mobile phase composed of (A) water (H_2O) and (B) acetonitrile (ACN). The gradient elution was modified as follows: 0–3 min 25% B, 3–10 min from 25 to 30% B, from 10–40 min from 30 to 40% B, which was held for 5 min. The postrunning time was 5 min. The flow rate was 1.0 mL/min. The column temperature was set at 30 $^\circ\text{C}$. The sample injection volume was 5 μL . The UV/DAD acquisitions were carried out in the range 190–600 nm, and chromatograms were acquired at 310, 330, 370, and 520 nm. All the compounds tested were found to be stable, and degradation products were not detected in the HPLC chromatograms. Supporting Information, Table S1 shows the purity (>95%) of the 38 natural compounds screened. The natural compounds were stored at low temperature ($-80 \text{ }^\circ\text{C}$), protected from light and humidity.

Computational Studies. *Virtual Screening of the Natural Product Library.* The library of 98 natural products was subjected to

Table 3. Summary Report of Computer-Simulated CI and DRI Values for MTX and Compound 13 Combinations at 50%, 75%, 90%, and 95% Inhibition of *T. brucei* Growth^a

drug combination	combination ratio	CI values at the reported EC				DRI values at the reported EC			
		EC ₅₀	EC ₇₅	EC ₉₀	EC ₉₅	EC ₅₀	EC ₇₅	EC ₉₀	EC ₉₅
MTX + 13	(2:1.5)	0.174 ± 0.047	0.075 ± 0.036	0.034 ± 0.023	0.020 ± 0.016	14.58 ± 3.71 ^b 11.72 ± 2.79 ^b	41.95 ± 10.52 ^b 27.45 ± 6.90 ^b	127.36 ± 60.20 ^b 65.56 ± 16.42 ^b	278.17 ± 172.13 ^b 119.72 ± 29.15 ^b

^aData reported are the average of three independent experiments. Data analysis was carried out using the CompuSyn software.³⁴ Values on top are for the MTX and on the bottom are for compound 13

Table 4. *In Vitro* Activity against *T. brucei* of Compound 2 in Three Different Formulations^a

preparation	<i>T. brucei</i> EC ₅₀ (μM)	THP1 CC ₅₀ ± SD or NOAEL
2	7.56 ± 0.51	53 ± 2
PLGA-2	5.70 ± 0.23	>100
cyclodextrin-2	3.27 ± 0.16	>100

^aEC₅₀ and NOAEL are the arithmetic mean of at least two independent determinations done in triplicate.

virtual screening using the GOLD software (version 5.1)^{35,36} following the protocol validated in previous studies on PTR1.¹³ The compounds were docked into the binding sites of the targets, *Lm*PTR1 (PDB ID 1E92) and *Tb*PTR1 (PDB ID 3JQ9), and the off-targets, hDHFR (PDB ID 1U72) and hTS (PDB ID 1HVY). Ten docking poses were generated for each ligand in each target, and these were visually inspected and manually ranked, considering the Gold fitness score, hydrogen bonding, and π - π aromatic interactions. A search of BioAssay data in PubChem was done to filter out promiscuous compounds. The set of 38 compounds given in Supporting Information, Table S1 was selected for inhibition assays considering the above properties and structural diversity.

Docking of Synthetic Flavonoid Derivatives. The 3D structures of the compounds were created from SMILES strings and optimized with the OPLS_2005 force field using Maestro.³⁷ Ionization states and tautomers were generated at pH 7.0 ± 0.5 using Epik.³⁷ Up to eight stereoisomers and one low-energy ring conformation were generated per compound. Important structural water sites were identified using the WatCH clustering approach.³⁸ All available chains of the *Lm*PTR1 crystal structures and 99 chains from *Tb*PTR1 crystal structures (see Supporting Information, Tables S14 and S15 for a list of structures) were superimposed and their water sites were clustered with a distance criterion of 2.4 Å. Water sites with at least 50% conservation were considered conserved.

The PTR1 structures for docking consisted of chain A of the relevant crystal structure and a C-terminal tripeptide from the neighboring subunit pointing into the chain A active site (Val266 to Ala268 in *Tb*PTR1 and Thr286 to Ala288 in *Lm*PTR1). The side-chain oxygen atom on the modified Cys 168 in the crystal structure of *Tb*PTR1 was removed. Crystallographic solvent molecules were removed and replaced by the conserved water molecules identified by WatCH clustering. The *Lm*PTR1 structure (PDB ID 1E92) was aligned to the *Tb*PTR1 template (PDB ID 3JCJ). PrepWizard was used to assign bond orders and add hydrogen atoms.³⁷ The N- and C-termini of chain A were capped with *N*-acetyl and *N*-methyl amide groups, respectively. The protonation state of the NADPH/NADP⁺ cofactor was computed at pH 7.0 ± 0.5. Protein protonation states were assigned by PROPKA⁴³ at pH 7.0. The H-bond network was optimized and all hydrogens were subjected to a restrained minimization. A 20 Å × 20 Å × 20 Å docking grid was centered on Phe97 of *Tb*PTR1 or Phe113 of *Lm*PTR1. The following hydroxyl groups were set rotatable: Ser95 and Tyr174 in *Tb*PTR1 and Ser111, Thr184, Tyr191, Tyr194, Thr195, and Tyr283 in *Lm*PTR1, as well as those of the NADPH/NADP⁺ ribose.

Docking was performed with and without ligand-based constraints using the Glide software.^{39–42} For the ligand constraints, protein preparation was performed including compound 7 (substrate-like binding mode, PDB ID 3JCJ) or compound NP-29 (inhibitor-like binding mode, PDB ID 3JCX) or the highest scoring docking solution from unconstrained docking of compound 4 with ring B in a stacking orientation (alternate binding mode). The ligands were then separated from the structure to provide reference geometries. The van der Waals radii of the ligand atoms were scaled by 0.80, and a partial charge cutoff of 0.15 was used. XP (extra precision) docking and flexible ligand sampling were set. Nitrogen inversions and ring conformations were sampled, and biased sampling of torsions was performed for amides only and set to penalize a nonplanar conformation. Epik state penalties were added to the docking score, intramolecular H-bonds were rewarded, and the planarity of conjugated π groups was

enhanced. A core pattern comparison with a tolerance of 3 Å was used for the ligand-based constraint. The core was defined as all non-hydrogen atoms of the chromen-4-one core ring system except the R3 hydroxyl group. Twenty poses per ligand were subjected to postdocking minimization with a threshold of 0.5 kcal/mol for rejecting a minimized pose. Up to 10 final docking solutions were reported. Unconstrained docking was performed analogously, omitting only the core pattern comparison. For *LmPTR1*, the core constraint docking was based on the binding poses in *TbPTR1*. In addition, induced-fit docking was carried out using the same settings except that the size of the docking grid was chosen automatically and a ligand and receptor van der Waals scaling of 0.5 was applied. Refinement was carried out with Prime³⁷ for residues within 5 Å of the ligand, followed by XP redocking into structures within 30 kcal/mol of the best structure and within the top 20 structures overall. Up to 20 final docking solutions were reported.

Synthesis. General. All commercial chemicals and solvents were reagent grade and were used without further purification. Reaction progress was monitored by TLC on precoated silica gel 60 F254 plates (Merck), and visualization was accomplished with UV light (254 nm). ¹H and ¹³C NMR spectra were recorded on a Bruker FT-NMR AVANCE 400. Chemical shifts are reported as δ values (ppm) referenced to residual solvent (CHCl₃ at δ 7.26 ppm, DMSO at δ 2.50 ppm, MeOD at δ 3.31 ppm); *J* values are given in Hz. When peak multiplicities are given, the following abbreviations are used: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broadened signal. Silica gel Merck (60–230 mesh) was used for column chromatography. All the reactions were monitored using TLC (Merck 890 F-254 silica gel). Mass spectra were obtained on a 6520 Accurate-Mass Q-TOF LC/MS and 6310A Ion Trap LC-MS(n). The detailed NMR and mass data of the synthesized compounds are reported in the Supporting Information (pp 33–37).

General Procedure for the Synthesis of (2E)-1-(2-Hydroxyphenyl)-3-phenylprop-2-en-1-ones. An aqueous solution of NaOH (3 M, 1.6 mL) was added to a solution of aromatic ketone (1 mmol) and substituted benzaldehyde (1.2 equiv) in EtOH (1–2 mL). The reaction was stirred at room temperature overnight. The reaction mixture was cooled in an ice–water bath and acidified to pH 2 with concentrated HCl (37%). The solid formed was filtered, washed with ethanol, and then further purified by recrystallization from EtOH.

General Procedure for the Synthesis of Methylated Flavonols (Compounds 9–16). An aqueous solution of H₂O₂ (30%, 250 μ L) was added to an ice-cold suspension of the chalcone (1 mmol) in ethanol (5 mL) and 1 M NaOH (2 mL). The mixture was allowed to warm to room temperature and was stirred for 5–18 h. Then the reaction mixture was cooled in an ice bath, and distilled water (2–4 mL) was added. Concentrated HCl (37%) was added until pH 2, and the precipitate was filtered, washed with EtOH, and further purified by recrystallization from EtOH. When no precipitate occurred, the reaction mixture was extracted with dichloromethane and washed with water and brine. The organic layer was dried over Na₂SO₄ and concentrated under reduced pressure. Column chromatography was utilized to purify the desired product.

General Procedure for the Synthesis of Demethylated Flavonols (Compounds 1–8). To a stirring solution of **15** (0.500 g, 1.52 mmol) in anhydrous dichloromethane (30 mL) under nitrogen at 0 °C, boron tribromide in dichloromethane (1.0 M, 13.7 mL, 13.7 mmol, 9 equiv) was added. The mixture was allowed to warm at room temperature and was stirred for 2 days. The reaction mixture was then cooled to 0 °C, and methanol (10 mL) was added. The reaction mixture was concentrated in vacuo. Water (10 mL) was added, and the reaction sonicated and then left to stand. Compound **7** (0.440 g, quantitative yield) was collected as a red solid.

The procedure followed for the synthesis of all demethylated flavonols is analogue to that of compound **7** by considering 3 equiv of BBr₃ for each C–O bond cleavage.

Protein Expression and Purification. Recombinant *TbPTR1* was expressed and purified by established methods¹⁰ with minor revisions. The plasmid was supplied by Prof. William N. Hunter, University of Dundee. Briefly, protein expression was carried out in

Escherichia coli BL21(DE3) cultured at 37 °C in SuperBroth (SB) medium to mid log phase and induced with 1 mM isopropyl- β -D-thiogalactopyranoside (IPTG) overnight at 30 °C.²⁷ Cells were resuspended in 50 mM Tris-HCl, pH 7.5, 250 mM NaCl and disrupted by sonication. The target protein purification was achieved in a single step using a HisTrap FF 5 mL column (GE-Healthcare) and an imidazole concentration of 250 mM in the same buffer. The resulting protein sample was dialyzed overnight in 20 mM Tris-HCl pH 7.5 at 8 °C (membrane cutoff 10 kDa). The high purity of the sample was confirmed by SDS-PAGE analysis and MALDI-TOF mass spectrometry. The final protein yield of enzyme was established as approximately 50 mg/L bacterial culture.

The plasmid pET15b-*LmPTR1* was supplied by Prof. William N. Hunter, University of Dundee. Protein expression was performed in the *E. coli* strain BL21(DE3) cultured at 37 °C in SB medium (supplemented with 100 mg/L ampicillin) to mid log phase and induced with 0.4 mM IPTG for 16 h at 28 °C. Cells, harvested by centrifugation, were resuspended in buffer A (50 mM Tris-HCl, pH 7.6, 20 mM imidazole, and 250 mM NaCl) and disrupted by sonication. The supernatant, collected by centrifugation, was further purified by nickel-affinity chromatography (HisTrap FF 5 mL column, GE-Healthcare), and the target protein was eluted using a 250–500 mM imidazole concentration in the same buffer. Fractions containing the protein (identified by SDS-PAGE) were pooled and combined with thrombin protease and then dialyzed overnight in 50 mM Tris-HCl, pH 7.6, at 25 °C. The mature protein was purified by a second nickel-affinity chromatography (HisTrap FF 5 mL column, GE-Healthcare) and eluted as weakly bound protein in 50–100 mM imidazole and 50 mM Tris-HCl, pH 7.6. The resulting protein sample was dialyzed in 20 mM sodium acetate pH 5.3 and 10 mM DTT. The high purity of the sample was confirmed by SDS-PAGE analysis and MALDI-TOF mass spectrometry, and the final protein yield established as approximately 30 mg/L bacterial culture.

Recombinant *LmDHFR-TS* was cloned in our lab. The gene coding sequence for *LmDHFR-TS*, cloned in pET-15b expression vector, was introduced by thermal shock into the *E. coli* strain ArcticExpress-(DE3). Bacterial culture was grown at 30 °C in ZYP-5052 autoinduction medium (supplemented with ampicillin 100 mg/L) to OD_{600 nm} value of about 1, and then cell growth was continued for 60 h at 12 °C under vigorous aeration. Cells, harvested by centrifuge, were resuspended in buffer A (50 mM sodium citrate pH 5.5 and 0.25 M NaCl) and disrupted by sonication. The supernatant of the resulting crude extract was collected by centrifuge and purified by nickel-affinity chromatography. The target protein was eluted using a linear gradient 20–250 mM imidazole in the same buffer. Fractions containing the target protein (identified by SDS-PAGE) were pooled and dialyzed overnight in buffer A at 8 °C. The resulting protein sample was further purified on a gel filtration column (HiLoad 16/600 Superdex 200pg, GE Healthcare) equilibrated with 50 mM sodium citrate, pH 5.5, and 0.25 M NaCl. The high purity of the target protein was confirmed by SDS-PAGE analysis and the final yield established as approximately 10 mg/L bacterial culture.

The recombinant *TbDHFR-TS* was cloned in our lab. The synthetic gene encoding for *TbDHFR-TS*, optimized for *E. coli* expression and cloned in pET-15b expression vector, was purchased from GenScript USA Inc. The target protein was expressed and purified using the same protocol described for *LmDHFR-TS*, with minor modification. Briefly, cells were resuspended in buffer A (50 mM phosphate buffer, pH 7.5, 0.25 M NaCl, 10% glycerol, and 20 mM imidazole) and disrupted by sonication. The supernatant of the resulting crude extract was collected by centrifuge and purified by nickel-affinity chromatography (HisTrap FF 5 mL column, GE-Healthcare). The target protein was eluted using a 400 mM imidazole concentration in the same buffer. Eluted fractions were pooled and dialyzed overnight in buffer A at 8 °C. The resulting protein sample was concentrated and applied to a gel filtration column (HiLoad 16/600 Superdex 200pg, GE Healthcare) equilibrated in buffer A. The high purity of the sample was confirmed by SDS-PAGE and the final protein yield established as approximately 8 mg/L bacterial culture. hTS and hDHFR were purified as reported in literature.^{15,44}

The kinetic characterization (K_m and K_{cat}) of all the proteins purified and employed for the target/off target screening is reported in Supporting Information, Table S16.

X-ray Crystallography. Crystals of histidine-tagged *TbPTR1* were obtained by the vapor diffusion hanging drop method technique at room temperature.⁴⁵ Drops were prepared by mixing equal volumes of protein (6–10 mg/mL in 20 mM Tris-HCl, pH 7.5, and 5–10 mM DTT) and precipitant (1.5–2.5 M sodium acetate and 0.1 M sodium citrate pH 5) solutions and equilibrated over a 600 μ L reservoir. Well-ordered monoclinic crystals grew in 3–6 days. The *TbPTR1*–cofactor–inhibitor complexes were obtained by soaking crystals with each compound solubilized either in 1,4-dioxane or DMSO (without exceeding a dioxane-DMSO/crystal solution ratio of 1:9). After 4–21 h, crystals were transferred to a cryoprotectant, prepared by adding 30% v/v glycerol to the precipitant solution, and flash frozen in liquid nitrogen. Diffraction data for the complexes with compound NP-29 and NP-13 were measured using synchrotron radiation at the Diamond Light Source (DLS, Didcot, United Kingdom) beamline I04 equipped with a Pilatus 6M-F detector, using a wavelength of 0.9795 Å, a 0.25° rotation, and a 0.4 s exposure time/image. Data for the complex *TbPTR1*-NADPH/NADP⁺-2 were collected at the DLS beamline I03 equipped with a Pilatus3 6 M detector and using a wavelength of 0.9173 Å, a 0.25° rotation, and a 0.15 s exposure time/image. Data relative to the complex with compound 7 were collected at the European Synchrotron Radiation Facility (ESRF, Grenoble, France) beamline ID30A-1, equipped with a Dectris Pilatus3 2 M detector and by using a wavelength of 0.9660 Å, a 0.1° rotation, and a 0.08 s exposure time/image. X-ray images were integrated using Mosflm and scaled with SCALA within the CCP4 software suite.^{46–48} *TbPTR1* crystals belong to the monoclinic space group $P2_1$, showing only slight variations in cell parameters among the different crystals. Data collection and processing statistics are reported in Supporting Information, Table S2. The structures were solved by molecular replacement using the software Molrep⁴⁹ and a whole tetramer of *TbPTR1* (PDB code 2X9G) as searching model.⁵⁰ Structural models were refined using REFMAC5,⁵¹ whereas the program Coot⁵² was used for electron density inspection, model manipulation, and placement of the solvent molecules. The final models were inspected manually and checked with the programs Coot and Procheck.⁵³ Data refinement statistics are reported in Supporting Information, Table S3. Figures were generated with CCP4 mg.⁵⁴

***TbPTR1*, *LmPTR1*, *TbDHFR*, *LmDHFR*, *hTS*, and *hDHFR* Target Enzyme Assays.** The *in vitro* assay used for *PTR1* enzymes is based on the coupled assay reported by Shanks et al.⁵⁵ *PTR1* uses H2B as a substrate and requires also NADPH for the reaction, and the reduction of H2B to H4B by *PTR1* is nonenzymatically linked with the reduction of cytochrome *c* in this assay, which is detected at 550 nm. The formation of cyt *c* Fe²⁺ results in a signal increase in the photometric readout.

TbPTR1 and *LmPTR1* activity was assayed in a buffer containing 20 mM sodium citrate (pH 6.0). The final reaction mixture contained the test compound at a range of concentrations and *TbPTR1*/*LmPTR1* (6.0 nM/12 nM), H2B (0.3 μ M/3 μ M), cytochrome *c* (100 μ M/100 μ M), and NADPH (500 μ M/500 μ M). The final assay volume was 50 μ L in 384-well clear plates (Greiner Bio-One, 781101). Compound screening was performed by addition of compound to assay plates (in 100% DMSO) followed by addition of 45 μ L of Reaction Mix (enzyme, H2B cytochrome *c* in 20 mM sodium citrate buffer). A pre-read was made at 550 nm using an EnVision Multilabel Reader 2103 (PerkinElmer Inc., US), followed by incubation of the assay plates at 30 °C for 10 min. The reaction was initiated by the addition of 5 μ L of NADPH (5 mM in ultrapure water), followed by the kinetically reading of the assay plates at 550 nm using the EnVision MultilabelReader at 10, 20, 30, 40, and 50 min, and the slope of each assay well was calculated. The screening data was analyzed using ActivityBase (IDBS) and for outlier elimination in the control wells the 3- σ method was applied. On the basis of the slope, data was normalized to the positive control methotrexate for *TbPTR1*/*LmPTR1* (1 μ M/50 μ M, yielding 100% inhibition) and negative controls (1% DMSO, yielding 0% inhibition) and % inhibition

calculated for all samples. The measurement at time 0 min was used to flag optically interfering samples. Each compound was tested in triplicate and the pIC₅₀ value, standard deviation, Hill slope, minimum signal, and maximum signal for each dose–response curve were obtained using a four-parameter logistic fit in the XE module of ActivityBase (IDBS).

LmDHFR, *hDHFR*, and *hTS* activities were assessed spectrophotometrically.¹³ The K_m for the substrates and k_{cat} for all enzymes purified and used for the inhibition studies are reported in Supporting Information, Table S16.

In Vitro Biological Assays. A cloned line of *L. infantum* (MOM/MA671TMAP263) promastigotes were maintained in RPMI 1640 medium supplemented with 10% heat-inactivated fetal bovine serum (FBS), 2 mM L-glutamine, 20 mM HEPES, and 1% Penn/Strep. Maintenance of promastigotes was done in T-25 flasks at 26 °C by subpassage at 10⁶ parasites/mL every 5–6 days. *L. infantum* axenic amastigotes expressing episomal luciferase were maintained in MAA/20 (axenic amastigote medium) at 37 °C under a 5% CO₂ environment with subpassages every 5 days. *LUC*-positive parasites were selected by addition of Geneticin sulfate (G418) to the culture at 60 μ g/mL. *Trypanosoma brucei brucei* Lister 427 bloodstream forms were grown at 37 °C, 5% CO₂ in complete HMI-9 medium supplemented with 10% fetal calf serum (FCS) and 100 UI/mL of penicillin/streptomycin. Cultures were diluted before a cell density of 2 \times 10⁶/mL was reached.

In Vitro Evaluation of Activity against *L. infantum* Intra-macrophage Amastigotes. The efficacy of compounds against *L. infantum* intracellular amastigotes was determined according to literature⁵⁶ but with slight modifications. Briefly, 1 \times 10⁶ THP-1-derived macrophages were infected with luciferase-expressing *L. infantum* axenic amastigotes in a macrophage:amastigotes ratio of 1:10 for 4 h at 37 °C, 5% CO₂. Noninternalized parasites were washed, and compounds were added at different concentrations (20–0.6 μ M). After 72 h of incubation at 37 °C and 5% CO₂, media was substituted by PBS. Macrophages were lysed by addition of 25 μ L of Glo-lysis buffer (Promega). Steady-Glo reagent (Promega) was then added, and the content of each well was transferred to white-bottom 96-well plates. Luminescence intensity was read using a Synergy 2 multi-mode reader (Biotek). The antileishmanial effect was evaluated by the determination of IC₅₀ value (concentration required to inhibit growth in 50%) and calculated by nonlinear regression analysis using GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego, California, www.graphpad.com.

In Vitro Evaluation of Activity against *T. brucei*. The efficacy of compounds against *T. brucei* bloodstream forms was evaluated using a modified resazurin-based assay previously described.⁵⁷ Mid log bloodstream forms were added to an equal volume of serial dilutions of compounds in supplemented complete HMI-9 medium at a final cell density of 5 \times 10³/mL. Following incubation for 72 h at 37 °C 5% CO₂, 20 μ L of a 0.5 mM resazurin solution was added and plates were incubated for further 4 h under the same conditions. Fluorescence was measured at 540 and 620 nm excitation and emission wavelength, respectively, using a Synergy 2 multi-mode reader (Biotek). The antitrypanosomatid effect was evaluated by the determination of IC₅₀ value (concentration required to inhibit growth in 50%) and calculated by nonlinear regression analysis using GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego California, www.graphpad.com. Half-maximal inhibitory concentrations (IC₅₀) reported correspond to the averages of the results obtained in at least two independent experiments. Compusyn software 1.0 was used to evaluate synergy.³⁴

In Vitro Evaluation of Activity against *T. cruzi*. Infections were performed in 6-well plates (3 \times 10⁶ HG39 cells/well). Confluent HG39 cells were infected with sanguineous trypomastigotes of *T. cruzi* Y strain at a 1:1 ratio. After 4 h, the cells were washed three times with PBS and fresh medium was added (RPMI supplemented with Glut/Pen/Strep and 20% heat-inactivated FCS, Sigma-Aldrich). The infected cells were incubated at 37 °C and 5% CO₂. Then 24 h post infection, the cells were treated with compounds and incubation was continued for 48 h. The gDNAs of infected cells were isolated

using the LGC Genomics Mag Maxi kit following the manufacturer's (LGC Genomics, Berlin) protocol and used as a template for a TaqMan probe-based quantitative real-time PCR.⁵⁸ The reference gene and the gene of interest were amplified using the KAPA Probe Fast qPCR Master Mix Kit (VWR) following the manufacturer protocol. Template DNA was used as 10% of the final reaction volume. Primers specific for human β -actin (reference gene) were human- β -AcF2 (CCCCTCTACGAGGGGTATG) and human- β -AcR3 (TCGGTGAGGATCTTCATG). The parasite actin-specific primers were *T. cruzi*-AcF (CGTGAGAAGATGACACAG) and *T. cruzi*-AcR (GGGAGAGAGTATCCCTCG). All primers were used at a final concentration of 300 nM. The probe-specific for human β -actin was human- β -Ac probe (Cy5-CCTGGCTGGCCGGGACCTGAC-BHQ-3) labeled with the dye Cy5 at the 5' end and with the quencher BHQ-3 at the 3' end. The probe specific for parasite actin was *T. cruzi* Ac probe (FAM-CACGCCATCACCAGCATCAAG-BHQ-1) labeled with the dye FAM at the 5' end and with the quencher BHQ-1 at the 3' end. Probes were used in a final concentration of 200 nM. The program was adapted to the use with the KAPA Probe Fast qPCR Master Mix Kit.

Cytotoxicity Assessment against THP-1 Macrophages. The effect of compounds 1–16 on THP-1-derived macrophages was assessed by the colorimetric MTT assay (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide). Briefly, 1×10^6 THP-1 cells were differentiated into macrophages by addition of 20 ng/mL of phorbol-myristate 13-acetate (PMA, Sigma-Aldrich) for 18 h, followed by replacement with fresh medium for 24 h. Cells were incubated with compounds ranging from 100 to 3 μ M after dilution in RPMI complete medium containing a maximum amount of 1% DMSO. After incubation for 72 h at 37 °C, 5% CO₂, medium was removed and a 0.5 mg/mL MTT solution was added. Plates were incubated for additional 4 h to allow viable cells to convert MTT into a purple formazan product. Solubilization of formazan crystals was achieved by addition of 2-propanol, and absorbance was read at 570 nm using a Synergy 2 multi-mode reader (Biotek). Cytotoxicity was evaluated by the determination of CC₅₀ value (drug concentration that reduced the percentage of viable cells in 50%) and calculated by nonlinear regression analysis using GraphPad Prism version 5.00 for Windows, GraphPad Software, San Diego California, www.graphpad.com.

hERG Cardiotoxicity Assay. The Invitrogen Predictor hERG fluorescence polarization assay was used in 384-well assay format (Greiner Bio-One, 784076) to test compounds. To each well of the assay plate, 100 nL of the test/control compound was added followed by addition of 5 μ L of homogenized membrane solution (undiluted), followed by a further addition of 5 μ L of tracer (1 nM final concentration in assay). The plates were incubated for 2 h at 25 °C in a humidity controlled incubator and the fluorescence polarization measured using an EnVision multilabel reader 2103 (Perkin Elmer Inc., US). The screening data was analyzed using ActivityBase (IDBS), and for outlier elimination in the control wells the 3- σ method was applied. The negative controls (0% inhibition) and positive controls E-4031, a blocker of hERG-type potassium channels (yielding 100% inhibition), were used to normalize the raw data. Each compound was tested in triplicate, and the pIC₅₀ value, standard deviation, Hill slope, minimum signal, and maximum signal for each dose–response curve were obtained using a four-parameter logistic fit in the XE module of ActivityBase (IDBS). All IC₅₀ data were associated with <10% SD.

Cytochrome P450 1A2, 2C9, 2C19, 2D6, and 3A4 Assay. The luminescence based P450-Glo (Promega) was used in 384-well assay format (Greiner Bio-One, 784076) to test compounds. The selected cytochrome P450 panel included microsomal preparations of cytochromes P450 1A2, 2C9, 2C19, 2D6, and 3A4 (Corning) from baculovirus infected insect cells (BTL-TN-5B1–4) which express cytochromes P450 (CYP) and cytochrome c reductase (and cytochrome b5 for 3A4). Compounds/controls were added into the empty 384-well plate (100 nL/well v/v 100% DMSO) using the Echo 550 Liquid Handler (Labcyte Inc., US), followed by addition of 5 μ L/well of CYP/Luciferin substrate, and incubated for 30 min at 37 °C. The reaction was initiated by addition of 5 μ L/well NADPH regeneration mixture. After 30 min at 37 °C, the CYP reaction was

stopped and the luciferase reaction was initiated by addition of 10 μ L/well of luciferin reagent. After additional 30 min at 37 °C, the luminescence readout was performed using the Infinite M1000 PRO plate reader (Tecan). Outliers in the control wells were eliminated according to the 3- σ method. Negative controls (0% inhibition) included only vehicle (v/v 1% DMSO), standard specific CYP inhibitors [CYP 1A2 inhibitor α -naphtho-flavone (Sigma-Aldrich, 15 nM), CYP 2C9 inhibitor sulfaphenazole (Sigma-Aldrich, 67 nM), CYP 2C19 inhibitor troglitazone (Sigma-Aldrich, 3.2 μ M), CYP 2D6 inhibitor quinidine (Sigma-Aldrich, 2 nM), and CYP 3A4 inhibitor ketoconazole (Sigma-Aldrich, 54 nM)] were used as positive controls (100% inhibition). Each compound was tested in triplicate, and the pIC₅₀ value, standard deviation, Hill slope, minimum signal, and maximum signal for each dose–response curve were obtained using a four-parameter logistic fit in the XE module of ActivityBase (IDBS). All IC₅₀ data were associated with <10% SD.

Cytotoxicity Assay against A549 and WI38 Cells. These assays were performed using the CellTiter-Glo assay from Promega Corp. A549 cells obtained from DSMZ (German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany) and WI-38 cells were obtained from ATCC (ATCC CCL-75) and were grown on surface modified T175 cell culture flasks (cat. no. 660175, Greiner Bio-One GmbH, AT) in DMEM (cat. no. 41966–052, Life Technologies, US) with 10% FCS, streptomycin (100 μ g/mL) and 100 U/mL penicillin G (cat. no. P-11-010, PAA Laboratories GmbH, AT). Cells were incubated at 37 °C in the presence of 5% CO₂ and were harvested at 80–90% confluency. Each test compound (200 nL of 10 mM in 100% v/v DMSO) was added to polystyrene 384-well cell culture microtiter plates (cat. no. 781073, 384 CellStar, Greiner Bio One, AT) using the Echo 550 Liquid Handler (Labcyte Inc., US). To harvest the cells, 1.5 mL of trypsin/EDTA (cat. no. L11-004, 0.5 and 0.22 mg/mL, respectively, PAA Laboratories GmbH, AT) was added per T175 flask and incubated at 37 °C in the presence of 5% CO₂ for 2 min. Detached cells were then resuspended in prewarmed media to a density of 0.2×10^6 cells/mL. From this cell suspension, 20 μ L was added per well of a 384-well microtiter plate, thereby giving a final test compound concentration of 100 μ M and 0.1% v/v DMSO. After 48 h of incubation at 37 °C in the presence of 5% CO₂, 20 μ L of CellTiter-Glo reagent (cat. no. G7571, Promega Inc., US) was added per well and the plate placed upon a linear shaker for 1 min at room temperature and further incubated at room temperature without shaking for 10 min. The luminescence readout was performed using the EnVision multilabel reader 2103 (PerkinElmer Inc., US) with a 0.5 s read time per well. Each assay plate also contained 16 wells for the positive control (cells prepared by treatment with cisplatin: 200 nL of 300 mM stock solution in 100% v/v DMSO, giving a final concentration of 3 mM cisplatin). The 16 wells for the negative controls per assay plate were prepared by treatment of cells with DMSO only (200 nL). Each compound was tested in triplicate, and the pIC₅₀ value, standard deviation, Hill slope, minimum signal, and maximum signal for each dose–response curve were obtained using a 4-parameter logistic fit in the XE module of ActivityBase (IDBS).

Assessment of Mitochondrial Toxicity. The 786-O (renal carcinoma) cell line was used for mitochondrial toxicity screening. Cells were maintained using RPMI-1640 medium containing 2 mM glutamine, 100 U/mL penicillin G, 100 mg/mL streptomycin, and 10% FCS. Cells were counted and diluted to a concentration of 75000 cells/mL, and 20 μ L of this suspension (1500 cells/well) were added to Cell Carrier 384-well TC plates (PerkinElmer Inc.) and incubated for 36 h at 37 °C and 5% CO₂. Compound dilutions were prepared using a 384-well PP predilution plate (Greiner Bio-One), and 20 μ L of the dilution were transferred to the 786-O cells. The low controls were valinomycin (final concentration of 1 μ M and 1% DMSO v/v in column 23) and the high controls were 1% DMSO v/v in column 24. After 6 h incubation at 37 °C and 5% CO₂, 10 μ L of a 200 nM solution of MitoTracker Red CMXRos (final concentration 50 nM) in prewarmed cell culture media (RPMI-1640 supplemented with 10% FCS, 100 U/mL penicillin, and 100 μ g/mL streptomycin) was added to each well and the 786-O cells were incubated for an additional 45 min at 37 °C and 5% CO₂. MitoTracker Red CMXRos uptake was

measured using an Opera HCS system (PerkinElmer), and image analysis was performed using Columbus 2.4.0 (Perkin Elmer). All screening experiments were performed for the compounds as 11-point dose–responses in triplicates. The raw data were processed using GraphPad Prism (GraphPad Software Inc.), with outliers in control wells eliminated according to the 3- σ method. The dose–response curves were analyzed using four-parameter sigmoidal fit to obtain plausible results in the case of nonsigmoidal dose–response curves.

Aurora B Kinase Assay. The assay used was the ADP-Glo Kinase Enzyme System from PromegaCorp. The assay was performed in 384-well white low volume plate (Corning Inc., NY, 3824). The control compound used was SU6656 (Calbiochem, Germany, 572635) at a final concentration of 1 μ M in column 23 and DMSO in column 24 at the same concentration (v/v) as the SU6656 control and the compound area. DMSO concentration is tolerated up to 2% final (v/v). An enzyme Mastermix containing 1 \times buffer, 50 μ M DTT, and 17.5 ng/ μ L (35 μ L/well) Aurora B [all reagents provided in the kit] was prepared. A substrate Mastermix containing 1 \times buffer, 36 μ M ATP, and 7.5 ng/ μ L (15 ng/well) MBP protein as substrate [buffer, MBP provided in the Aurora B Kinase Enzyme System kit; ultrapure ATP provided in the ADP-Glo Kinase Assay kit] was prepared. Then 2 μ L of the enzyme Mastermix and 2 μ L of the substrate Mastermix were added to each well of a 384 low volume plate, and the plate was sealed using Thermowell sealing tape (Corning Inc. NY, USA) and incubated for 45 min at RT. The enzymatic reaction was stopped by adding 4 μ L of ADP-Glo reagent [provided in the ADP-Glo Kinase Assay kit], and the plate is sealed again using Thermowell sealing tape (Corning Inc. NY) and incubated for 40 min at RT. Afterward, 8 μ L of detection reagent were added to each well and the plate was sealed again. The plate was incubated for 45 min at RT, and afterward luminescence was measured using the EnVision multilabel reader 2103 (Perkin Elmer Inc., US). Each compound was tested in triplicate, and the pIC₅₀ value, standard deviation, Hill slope, minimum signal, and maximum signal for each dose–response curve were obtained using a four-parameter logistic fit in the XE module of ActivityBase (IDBS). All IC₅₀ data were associated with <10% SD.

Solubility Assays. UV–visible spectroscopy was used to measure the solubility of compound 2. The experiments were carried out with the instrument CARY 50. Six different solutions (3.125, 6.25, 12.5, 25, 50, and 100 μ M) were prepared. The maximum of absorbance was found at 334 nm, and epsilon was calculated. Four saturated solutions were prepared: 100 μ M in water, 100 μ M in PBS, 200 μ M in PBS + 10% DMSO, and 10 mM in PBS + 50% DMSO. The suspensions were left in incubation at 25 $^{\circ}$ C for 18 h, then filtered (0.2 μ m). The absorbance at 334 nm was measured. One blank and three replicates of the same sample were performed for each solution. Using the calibration curve ($Y = 0.0143x + 0.0033$), we calculated the maximum solubility. Maximum solubility in water: 2.6 μ M. Max solubility in PBS: 0.5 μ M. Max solubility in PBS + 10% DMSO = 85.5 μ M. Max solubility in PBS + 50% DMSO = 7.0 mM.

Compound 2 Encapsulation in PLGA and Solubilization with Cyclodextrins. The encapsulation of compound 2 on PLGA nanoparticles was obtained by a nanoprecipitation methodology. The polymer was dissolved in acetone at 20 mg/kg for the diffusing phase. Then 0.5 mg of free compound were added for drug loading. This phase was then added to 10 mL of Pluronic F127 (0.1% in water) under moderate magnetic stirring at room temperature. For the evaporation of the organic solvent, the emulsion was stirred for 2 h. The formed NPs were recovered and washed by centrifugation and stored in PBS at 4 $^{\circ}$ C until use. Quantification of incorporated compound was done by the indirect method using absorption spectrophotometry. Standard curves for each compound were performed using 334 nm. The encapsulation efficiency was calculated as the ratio between the initial quantity of compound added to produce the NPs and the quantity of nonencapsulated compound present in the recovery and washing supernatants over the initial quantity of compound added to produce the NPs. Particles were characterized by dynamic light scattering (DLS); the size, polydispersity index, and ζ potential were determined using a Zetasizer Nano ZS (Malvern Instruments, Malvern, UK) with a detection angle of 173 $^{\circ}$.

Measurements were made in triplicate at 25 $^{\circ}$ C. Compound 2 was solubilized with hydroxypropyl- β -cyclodextrin (50% w/vol)/(Cavaso W7 HP Pharma, Ashland).

Pharmacokinetics of Compound 2. Plasma samples from 6 BALB/c mice treated with compound 2 administered alone (1 mg/kg IV) (5, 15, and 30 min post administration, pa), in PLGA nanoparticles (1 mg/kg/IV and oral) (0, 5, 15, 30, 45, 60 min and then 1 min, 3 min, 24 h, 48 h, and 72 h pa) and solubilized with cyclodextrins (NMRI mice, 4 mg/kg/per os) (1, 6, and 24 h pa) were analyzed by HPLC. Analysis was carried out with a modular Jasco HPLC fitted with a Hypersil BDS C18 column (250 mm \times 4.6 mm, 5 μ m) (Thermo Scientific) as stationary phase. Samples were treated with acetonitrile (ACN), vortexed, centrifuged (15 min, 6000 rpm), filtered through a 0.45 μ m filter (Pall GHP Acrodisc GF), and assayed. Mobile phase was ACN: pure water (35:65), flow rate was 1.5 mL/min, injection loop was 100 μ L, and detection at a wavelength of 335 nm. Sulfate and glucuronide conjugation of compound 2 was tested.⁵⁹

Ethics Statement. All experiments performed at IBMC were carried out in accordance with the IBMC and INEB Animal Ethics Committees and the Portuguese National Authorities for Animal Health guidelines, according to the statements on the directive 2010/63/EU of the European Parliament and of the Council.

The experimental design and housing conditions at UCM were approved by the Committee of Animal Experimentation (Universidad Complutense de Madrid) and regional authorities (Community of Madrid) (ref. PROEX 169/15). Experiments were carried out at the animal house with official identification code ES280790001164 following the 3Rs principles. Animal handling and sampling were performed by trained and officially qualified personnel.

■ ASSOCIATED CONTENT

📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jmedchem.6b00698.

Compound characterization checklist (XLS)

Molecular formula strings (XLSX)

Compound 1–TbPTR1 (PDB)

Compound 2–TbPTR1 (PDB)

Compound 3–TbPTR1 (PDB)

Compound 4–TbPTR1 (PDB)

Compound 5–TbPTR1 (PDB)

Compound 6–TbPTR1 (PDB)

Compound 7–TbPTR1 (PDB)

Compound 8–TbPTR1 (PDB)

Compound 9–TbPTR1 (PDB)

Compound 10–TbPTR1 (PDB)

Compound 11–TbPTR1 (PDB)

Compound 12–TbPTR1 (PDB)

Compound 13–TbPTR1 (PDB)

Compound 14–TbPTR1 (PDB)

Compound 15–TbPTR1 (PDB)

Compound 16–TbPTR1 (PDB)

Compound 1–LmPTR1 (PDB)

Compound 2–LmPTR1 (PDB)

Compound 3–LmPTR1 (PDB)

Compound 4–LmPTR1 (PDB)

Compound 5–LmPTR1 (PDB)

Compound 6–LmPTR1 (PDB)

Compound 7–LmPTR1 (PDB)

Compound 8–LmPTR1 (PDB)

Compound 9–LmPTR1 (PDB)

Compound 10–LmPTR1 (PDB)

Compound 11–LmPTR1 (PDB)

Compound 12–LmPTR1 (PDB)

Compound 13–*LmPTR1* (PDB)

Compound 14–*LmPTR1* (PDB)

Compound 15–*LmPTR1* (PDB)

Compound 16–*LmPTR1* (PDB)

General structures of the flavonoids; code, chemical structure, purity, and IC₅₀ values of the 38 natural compounds screened; X-ray crystallographic data; inhibitory activity of the synthetic flavonols against *TbPTR1* and *LmPTR1*; docking analysis; sequence alignment of *TbPTR1* and *LmPTR1*; inhibitory activity of the synthetic flavonols against hDHFR, *TbDHFR* and *LmDHFR*; ADME-Tox data; antiparasitic activity of the synthesized compounds alone and in combination with MTX and synergy coefficients; isobologram and dose–effect curves for the combination of MTX and compound 13; toxicity of the combination on THP-1 cells; plasma concentration of compound 2 in BALB/c mice; nanoparticles characterization by dynamic light scattering; crystal structures used for conserved water analysis in *TbPTR1* and *LmPTR1*; kinetic characterization of *PTR1* and *DHFR*; characterization of the compounds 1–16 and of the intermediates 17–24 (PDF)

Accession Codes

PDB code 5JCJ was used for docking of compounds 1–16 (*TbPTR1*). PDB code 1E92 was used for docking of compounds 1–16 (*LmPTR1*). The Protein Data Bank accession codes of the X-ray crystallographic structures of *TbPTR1* in complex with NP-13, NP-29, 2, and 7 are 5JDC, 5JCX, 5JDI, and 5JCJ, respectively. Authors will release the atomic coordinates and experimental data upon article publication.

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Notes

The authors declare no competing financial interest.

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ABBREVIATIONS USED

WHO, World Health Organization; NTDs, neglected tropical diseases; HAT, human African trypanosomiasis; DNDi, Drugs for Neglected Diseases Initiative; DHFR, dihydrofolate reductase; *TbPTR1*, *Trypanosoma brucei* pteridine reductase 1; *LmPTR1*, *Leishmania major* pteridine reductase 1; MTX, methotrexate; *L. infantum*, *Leishmania infantum*; *L. donovani*, *Leishmania donovani*; *T. brucei*, *Trypanosoma brucei*; *T. cruzi*, *Trypanosoma cruzi*; CDCl₃, deuterated trichloromethane; EtOH, ethanol; MeOH, methanol; NaOH, sodium hydroxide; TMS, trimethylsilane; DLS, dynamic light scattering; NPs, nanoparticles; PLGA, poly(lactic-co-glycolic acid); IV, intravenous; PBS, phosphate-buffered saline; ACN, acetonitrile; FBS, fetal bovine serum; EC₅₀, half-maximal effective concentration; CC₅₀, half-maximal cytotoxicity concentration; THP1, human monocytic cell line; A549, human lung adenocarcinoma epithelial cell line; WI-38, fetal lung fibroblasts cell lines; DRC, dose–response curve

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