

# Protective Effects of a Glucocorticoid on Downregulation of Pulmonary $\beta_2$ -Adrenergic Receptors In Vivo

Judith C. W. Mak, Masanori Nishikawa, Hideaki Shirasaki, Kikuo Miyayasu, and Peter J. Barnes

Department of Thoracic Medicine, Royal Brompton National Heart and Lung Institute, London SW3 6LY, United Kingdom

## Abstract

We investigated the in vivo effects of a glucocorticoid on  $\beta$ -agonist-induced downregulation of  $\beta_1$ - and  $\beta_2$ -adrenergic receptors (determined by [ $^{125}$ I]iodocyanopindolol binding), mRNA expression (assessed by Northern blotting), and gene transcription (using nuclear run-on assays) in rat lung. Dexamethasone (Dex) (0.2 mg/kg/d, days 1–8) increased  $\beta_1$ - and  $\beta_2$ -receptor numbers by 70 and 69% above control, respectively, but did not change their mRNA expression. Isoproterenol (Iso) (0.96 mg/kg/d, days 2–8) decreased  $\beta_1$ - and  $\beta_2$ -receptor numbers by 48 and 51%, respectively, and also reduced mRNA expression by 69 and 57%, respectively. The combination of Dex and Iso resulted in no net change in  $\beta_2$ -receptor number and its mRNA expression, although there was a significant reduction in  $\beta_1$ -receptor number and mRNA expression. The mapping of  $\beta_1$ - and  $\beta_2$ -receptors by receptor autoradiography confirmed these findings over alveoli, epithelium, endothelium, and airway and vascular smooth muscle. We also measured the activation of the transcription factor, cyclic AMP response element binding protein (CREB) using an electrophoretic mobility shift assay. CREB-like DNA-binding activity was decreased after Iso treatment but this decrease was prevented after treatment with Dex. Nuclear run-on assays revealed that the transcription rate of the  $\beta_1$ -receptor gene did not alter after Dex treatment, but was reduced after Iso treatment. The transcription rate of the  $\beta_2$ -receptor gene was increased after Dex treatment by approximately twofold, but there was no change after Iso treatment. We conclude that glucocorticoids can prevent homologous downregulation of  $\beta_2$ -receptor number and mRNA expression at the transcriptional level without affecting  $\beta_1$ -receptors and that the transcription factor CREB may be involved in this phenomenon. Such an effect may have clinical implications for preventing the development of tolerance to  $\beta_2$ -agonists in asthmatic patients treated with  $\beta$ -agonist bronchodilators. (*J. Clin. Invest.* 1995; 96:99–106.) Key words: glucocorticosteroid • upregulation • downregulation • mRNA expression •  $\beta$ -adrenergic receptors • CREB

Address correspondence to Prof. Peter J. Barnes, Department of Thoracic Medicine, National Heart and Lung Institute, Dovehouse Street, London SW3 6LY, United Kingdom. Phone: 71-352-8121; FAX: 71-376-3442.

Received for publication 6 June 1994 and accepted in revised form 5 April 1995.

*J. Clin. Invest.*

© The American Society for Clinical Investigation, Inc.

0021-9738/95/07/0099/08 \$2.00

Volume 96, July 1995, 99–106

## Introduction

Glucocorticosteroids and  $\beta_2$ -adrenergic agonists are the mainstay of asthma treatment and are usually given together (1).  $\beta$ -Adrenergic agonists cause bronchodilation predominantly in the activation of  $\beta_2$ -adrenergic receptors ( $\beta_2$ -receptors) on airway smooth muscle cells (2). There has been concern that regular use of inhaled  $\beta_2$ -agonists may result in tolerance to their beneficial effects in asthma. Although there is no loss of bronchodilator response to  $\beta_2$ -agonists, several studies have demonstrated loss of protection against various bronchoconstrictor challenges (3, 4) and this may be relevant to the reduced asthma control seen with the regular use of inhaled  $\beta_2$ -agonists (5, 6). There is a downregulation of  $\beta$ -adrenergic receptors ( $\beta$ -receptors) in lung after chronic administration of  $\beta$ -agonists in animals in vivo, although this is less marked in airway smooth muscle than in lung parenchyma (7, 8). Agonist-promoted downregulation of  $\beta_2$ -receptors may be reversed by treatment with glucocorticoids in vitro (9). Glucocorticoids induce an increase in the synthesis of  $\beta$ -receptors in human and rat lung (10) and restoration of desensitization of  $\beta$ -receptors in human neutrophils and lymphocytes (11, 12). The reversal of agonist-induced downregulation of the  $\beta_2$ -receptor by glucocorticoids has been reported in cultured vas deferens smooth muscle cells (DDT<sub>1</sub>-MF2) at the levels of radioligand binding and of mRNA (13).

Cyclic AMP and glucocorticoid response elements (CREs and GREs)<sup>1</sup> have been identified in the promoter region of the  $\beta_2$ -receptor gene (14, 15), suggesting the involvement of these transcription factors in the regulation of  $\beta_2$ -receptors. CRE binding protein (CREB) appears to maintain the basal transcription of the  $\beta_2$ -receptor gene (16). On the other hand, glucocorticoids appear to increase  $\beta_2$ -receptor mRNA expression, exerted at GREs in the 5'-noncoding region (15, 17, 18).

Chronic  $\beta$ -agonist therapy in asthmatic subjects results in reduction in  $\beta$ -receptor density in circulating polymorphonuclear leukocytes and lymphocytes (19) and the downregulated receptor number is restored with oral prednisone. However, a difference in susceptibility to downregulation between lung and lymphoid tissue has been reported (20). To investigate whether the in vivo treatment of glucocorticoids prevent the  $\beta$ -agonist-promoted downregulation of pulmonary  $\beta_2$ -receptors, we studied the effects of dexamethasone (Dex) and isoproterenol (Iso) on  $\beta_2$ -receptor number, and mRNA expression in rat lung in vivo. In addition, we examined effects on the transcription factor CREB. We also used autoradiographic mapping of  $\beta_2$ -receptors to study the effects of Iso and Dex on different cell types in lung. Direct receptor binding techniques using selective  $\beta$ -antagonists have shown the coexistence of  $\beta_1$ - and  $\beta_2$ -receptor

1. Abbreviations used in this paper: CRE, cAMP response element; CREB, CRE binding protein; Dex, dexamethasone; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; GRE, glucocorticoid response element; ICYP, [ $^{125}$ I]iodocyanopindolol; Iso, isoproterenol.

subtypes in rat and human lung (7, 21). Both Dex and Iso may also affect  $\beta_1$ -receptors as well as  $\beta_2$ -receptors, so we also examined the expression of  $\beta_1$ -receptors under the same conditions.

## Methods

**Experimental procedures.** Male Wistar rats weighing 300–320 g were used. One group ( $n = 7$ ) was injected with Dex (0.2 mg/kg/d) subcutaneously for 1 d, with another group ( $n = 7$ ) once a day for 8 d (days 1–8). A third group ( $n = 7$ ) was treated with Iso (0.96 mg/kg/d) for 7 d (days 2–8) delivered by an osmotic minipump (Alzet model 2001; Alza Corp., Palo Alto, CA), which was implanted subcutaneously under anesthesia and sterile conditions. A fourth group ( $n = 7$ ) was treated with both Dex and Iso simultaneously in the same manner described above. A control group ( $n = 7$ ) was treated with vehicle both by subcutaneous injection and by a minipump simultaneously. Dex was dissolved in sterilized isotonic saline only; Iso was dissolved in sterilized isotonic saline containing 1.1 mM ascorbic acid to prevent its oxidation. On the ninth day the animals were killed by 100% CO<sub>2</sub> exposure and the lungs were quickly removed.

**Radioligand receptor binding assay.** Lung was minced coarsely with scissors and suspended in 10 vol of 25 mM Tris-HCl buffer (pH 7.4) containing 0.32 M sucrose at 4°C, and was then homogenized with a Polytron homogenizer (Kinematica, Basel, Switzerland) at setting 6 in 30-s bursts. The homogenate was centrifuged at 1,000 g for 10 min at 4°C to remove unhomogenized debris, the supernatant was then centrifuged at 40,000 g for 20 min at 4°C, and the resulting pellet was washed and recentrifuged at the same speed. The final homogenate was frozen in liquid nitrogen and stored at –80°C without loss of binding characteristics. Portions of lung membrane at a protein concentration of 10  $\mu$ g/tube were incubated with [<sup>125</sup>I]iodocyanopindolol (ICYP) (sp act: 2,000 Ci/mmol; 3–100 pM) in the presence or absence of excess Iso (200  $\mu$ M) in 25 mM Tris-HCl buffer (pH 7.4) containing 154 mM NaCl and 1.1 mM ascorbic acid (to prevent oxidation of Iso) in a final volume of 250  $\mu$ l. The density of  $\beta_1$ -receptors was analyzed by ICYP saturation binding in the presence of 0.1  $\mu$ M ICI 118551, a  $\beta_2$ -selective antagonist, a concentration at which practically all  $\beta_2$ -receptors are occupied. The density of  $\beta_2$ -receptors by ICYP saturation binding in the presence of 0.1  $\mu$ M CGP 20712 A, a  $\beta_1$ -selective antagonist, a concentration at which practically all  $\beta_1$ -receptors are occupied. Incubation was carried out at 37°C for 120 min, which was found to be optimal for specific binding. Incubations were performed in triplicate. The incubation was terminated by rapid filtration through GF/C glass-fiber filters (Whatman Inc., Clifton, NJ). Each filter was rapidly washed with 3  $\times$  5 ml ice-cold 25 mM Tris-HCl buffer (pH 7.4). The filters were counted in the Auto Gamma Counting System (model 5550; Packard Instruments, Downers Grove, IL) at an efficiency of 80%. Specific binding was calculated by subtracting nonspecific binding from total binding. Protein concentration was determined by the method of Lowry et al. (22), with bovine serum albumin as a standard.

**Receptor autoradiography.** Parenchymal tissue was inflated by bronchial instillation of OCT embedding medium diluted 1:4 with PBS. All tissue samples were snap-frozen in isopentane cooled in liquid nitrogen and stored at –80°C until required. Serial frozen sections (10  $\mu$ m) of parenchymal tissue were cut at –30°C, mounted, and thawed onto gelatinized glass slides. Sections were stored at –80°C as long as 2 wk before use without loss of binding capacity.

Receptor mapping was performed using the method as described previously (8). The slides were warmed to room temperature, washed in incubation buffer (25 mM Tris-HCl, 154 mM NaCl, 0.25% polypeptide, and 1.1 mM ascorbic acid; pH 7.4), and incubated with 25 pM ICYP at 37°C for 120 min. Nonspecific binding was determined by incubating adjacent sections with the same concentration of ICYP and 200  $\mu$ M Iso. For mapping of the  $\beta_1$ -receptors, serial sections were incubated with 25 pM ICYP with and without 0.1  $\mu$ M ICI 118551, and for  $\beta_2$ -receptors with and without 0.1  $\mu$ M CGP 20712 A. After incuba-

tion, slides were washed twice for 15 min in ice-cold buffer (25 mM Tris-HCl, pH 7.4), rinsed in cold distilled water, and then rapidly dried in a stream of cold air. Glass coverslips previously coated with Ilford K-5 emulsion were fixed to one end of the slide with cyanoacrylate adhesive and held in contact with the sections with butterfly clips. Slides were exposed to the emulsion for 4 d. The glass coverslip was developed in Kodak D-19 developer and fixed. Sections were stained with cresyl fast violet and examined under a Zeiss microscope equipped with dark and bright-field illumination. Grain density was measured as optical density with a microscope connected to Image Analysis (Seescan, Cambridge, United Kingdom), using a constant magnification. Values of optical density were corrected for background and nonspecific binding. No correction was applied for a possible nonlinearity of emulsion response, as the range of the measurements was small.

**Northern blot analysis.** Random primer labeling was carried out with the 1.8-kb full-length fragment from rat  $\beta_2$ -receptor cDNA (23), the 851-bp SmaI/PvuII fragment from human  $\beta_1$ -receptor cDNA, and the 1.3-kb PstI fragment from rat glyceraldehyde-3-phosphate dehydrogenase (GAPDH) cDNA using [ $\alpha$ -<sup>32</sup>P]dCTP (3,000 Ci/mmol). GAPDH is a constitutive (housekeeping) gene that is expressed at constant levels in all cells (24) and used as an internal control for the quantity of the RNA loaded into each lane.

Total RNA from rat lung was isolated according to the method of Chromczynski and Sacchi (25). Total cellular RNA (20  $\mu$ g/lane) was subjected to electrophoresis on a 1% wt/vol agarose, 6.6% formaldehyde gel and blotted onto Hybond-N membranes (Amersham International, Amersham, United Kingdom). After prehybridization for 5 h at 42°C in buffer containing 5  $\times$  Denhardt's solution, 5  $\times$  standard saline citrate (SSC), 50 mM Na<sub>2</sub>HPO<sub>4</sub>, 0.1% sodium dodecyl sulfate (SDS), 250  $\mu$ g/ml sonicated denatured salmon sperm DNA, and 50% formamide, the blot was incubated for 20 h at 42°C with either first to a <sup>32</sup>P-labeled rat  $\beta_2$ - or a human  $\beta_1$ -receptor cDNA probe (1  $\times$  10<sup>6</sup> cpm/ml) in prehybridization buffer. Each blot was washed twice with 2  $\times$  SSC/0.1% SDS at room temperature and twice with 2  $\times$  SSC/0.1% SDS for 30 min, once with 1  $\times$  SSC/0.1% SDS for 30 min at 42°C, once with 0.5  $\times$  SSC/0.1% SDS for 30 min at 50°C, and finally with 0.1  $\times$  SSC/0.1% SDS for 30 min at 55°C, and exposed at –80°C for 10–14 d to Kodak OMAT XS film with an intensifying screen. After autoradiography, blots were stripped for reprobing with a <sup>32</sup>P-labeled rat GAPDH cDNA probe.

**Electrophoretic mobility shift assay.** The CREB consensus oligonucleotide was labeled at the 5'-ends using T4 polynucleotide kinase and [ $\gamma$ -<sup>32</sup>P]ATP (> 5,000 Ci/mmol). Nuclear protein from rat lung was isolated according to a method described previously (7). Binding reactions between <sup>32</sup>P-labeled CREB consensus oligonucleotide and nuclear protein were performed in a final volume of 10  $\mu$ l in 4% glycerol, 1 mM EDTA, 1 mM DTT, 100 mM NaCl, 10 mM Tris (pH 7.5), and 0.08 mg/ml sonicated salmon sperm DNA. Binding was allowed to proceed for 20 min at room temperature. To resolve the complexes, the reactions were applied to 6% nondenaturing polyacrylamide gels in 0.25  $\times$  TBE buffer (10  $\times$  TBE is 0.89 M Tris base [pH 8.0], 0.89 M boric acid, and 20 mM EDTA) containing 0.1% ammonium persulfate. The gels were run in 0.25  $\times$  TBE buffer at 100 V/cm for 1 h at room temperature with buffer recirculation, then dried and autoradiographed. The specificity of binding was studied by incubation of <sup>32</sup>P-labeled CREB consensus oligonucleotide with nuclear protein in the presence of excess amount of unlabeled oligonucleotide. We have demonstrated previously that CREB-like DNA-binding activity is increased in a concentration-related manner by  $\beta$ -agonists in rat lung (26).

**Nuclear run-on transcription assay.** To determine whether Dex or Iso changed the transcription rate of  $\beta_1$ - and  $\beta_2$ -receptor gene, nuclear run-on transcription assays were performed as described previously (27). Nuclei from frozen rat lung tissues treated with vehicle, Dex, Iso, or both were isolated and stored at –80°C in Keller storage buffer at 25  $\times$  10<sup>6</sup> nuclei/100  $\mu$ l. Each reaction (final volume, 400  $\mu$ l) was carried out in the presence of 5  $\times$  10<sup>7</sup> isolated nuclei, 40 mM Tris-HCl (pH 8.3), 150 mM NH<sub>4</sub>Cl, 7.5 mM MgCl<sub>2</sub>, 0.625 mM ATP, 0.313 mM GTP, 0.313 mM CTP, 0.5 mCi [ $\alpha$ -<sup>32</sup>P]UTP (800 Ci/mmol), and 120

U/ml recombinant RNasin. Transcription reactions were allowed to proceed for 30 min at 27°C before termination by the addition of 40 U of recombinant RNasin and 75 U of RQ-1 DNase. After DNase and proteinase K treatments, the radiolabeled RNA formed was purified by phenol/chloroform extraction and precipitated with ethanol three times in the presence of 1.33 M ammonium acetate. An equal number of counts from each sample ( $2 \times 10^6$  cpm) was added to slot blots, 4 slots on the same blot of which 10  $\mu$ g of either pGEM-3Z plasmid (as control), plasmid containing inserts of full-length human  $\beta_1$ -receptor cDNA, rat  $\beta_2$ -receptor cDNA, or rat GAPDH cDNA has been immobilized to a nylon filter (Hybond-N). After hybridization for 72 h at 42°C, the filters were washed at a final stringency of  $0.1 \times$  SSC and 0.1% SDS at 55°C, including a 30-min digestion with 1  $\mu$ g/ml RNase A and 10 U/ml RNase T<sub>1</sub> at 37°C to digest any single-stranded RNA not hybridized to DNA. The filters were exposed to Kodak OMAT XS film with an intensifying screen at -80°C for 1-3 d.

**Analysis of results.** The experimental data are given as means  $\pm$  SEM. The significance of difference was tested by an ANOVA;  $P < 0.05$  was considered to be statistically significant. Parameters (dissociation constant,  $K_d$ ; maximal binding capacity,  $B_{max}$ ) of ICYP binding were obtained from individual experiments using the program GraphPAD (ISI Software, San Diego, CA). Data from the autoradiograms of Northern blot analysis, gel shift assays, and nuclear run-on assays were assessed using laser densitometry (Howtek, Hudson, NH) linked to a computer analysis system (PDI, Huntington Station, NY).

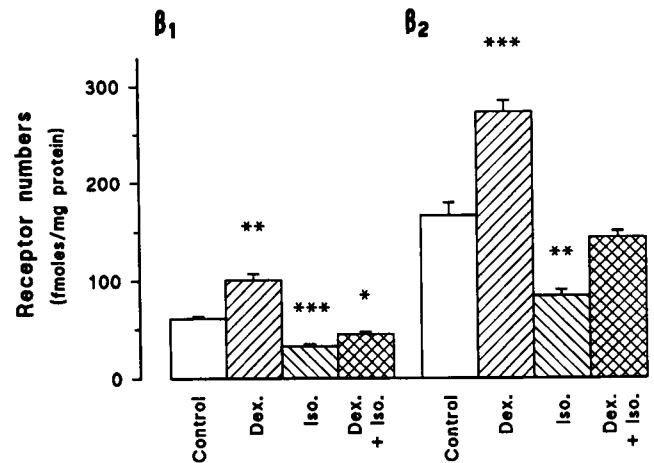
**Materials.** [<sup>125</sup>I]iodocyanopindolol, [ $\alpha$ -<sup>32</sup>P]dCTP, [ $\gamma$ -<sup>32</sup>P]ATP, Hybond-N membranes, and random primer labeling kit were purchased from Amersham International. [ $\alpha$ -<sup>32</sup>P]UTP was obtained from DuPont/New England Nuclear (Stevenage, United Kingdom). Rat  $\beta_2$ -receptor cDNA was obtained from American Type Culture Collection (Rockville, MD). CREB consensus oligonucleotide, recombinant RNasin, RQ-1 DNase, and restriction endonucleases, such as SmaI and PvuII, were purchased from Promega (Southampton, United Kingdom). Dex, Iso, and other substances were obtained from Sigma Chemical Co. (Poole, United Kingdom). CGP-20712A was a gift from Ciba-Geigy (Basel, Switzerland) and ICI-118,551 was from Zeneca Pharmaceuticals (Macclesfield, United Kingdom).

## Results

**Radioligand receptor binding assay.** Chronic injection of Dex (8 d) significantly increased the number of  $\beta_1$ - and  $\beta_2$ -receptor density by  $70 \pm 15\%$  ( $P = 0.001$ ) and  $69 \pm 10\%$  ( $P < 0.001$ ), respectively (Fig. 1), but there was no increase after treatment for 1 d. Continuous infusion of Iso significantly reduced  $\beta_1$ - and  $\beta_2$ -receptor density by  $48 \pm 5\%$  ( $P < 0.001$ ) and  $51 \pm 6\%$  ( $P = 0.001$ ), respectively. Combined treatment of Dex and Iso did not significantly change  $\beta_2$ -receptor density (decrease by  $15 \pm 6\%$ , not significant), but significantly reduced  $\beta_1$ -receptor density by  $23 \pm 5\%$  ( $P = 0.01$ ). These treatments did not change the affinity of binding, nor the ratio of the  $\beta_2$ - and  $\beta_1$ -receptors ( $\sim 3:1$ ).

**Receptor autoradiography.** The distribution of  $\beta_1$ - and  $\beta_2$ -receptors in rat lung (Fig. 2) is in good agreement with previous findings in human and guinea pig lung (8, 28) and confirms and extends the results of the receptor binding study. The increase in  $\beta_1$ - and  $\beta_2$ -receptors after Dex, the decrease in both receptor subtypes after Iso, the decrease in  $\beta_1$ - but no change in  $\beta_2$ -receptor after combined treatment of Dex and Iso were found over alveoli, airway epithelium, vascular endothelium, and airway and vascular smooth muscle (Table I).

**Northern blot analysis.** A single band of 3.2 and 2.2 kb for  $\beta_1$ - and  $\beta_2$ -receptor mRNA was observed in rat lung, respectively, in agreement with previous reports (29). Chronic injection of Dex (8 d) did not change  $\beta_1$ - or  $\beta_2$ -receptor mRNA



**Figure 1.** Effects of treatment with Dex and/or Iso on  $\beta$ -adrenergic receptor subtypes in rat lung. Control, Dex, Iso, and combination of Dex and Iso groups are shown. The density of  $\beta_1$ -receptor subtype was analyzed by ICYP saturation binding in the presence of 0.1  $\mu$ M ICI 118,551 and the density of  $\beta_2$ -receptor subtype by ICYP saturation binding in the presence of 0.1  $\mu$ M CGP 20712 A. Significance of difference from the control value, \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$  ( $n = 7$ ).

expression (Figs. 3 and 4, respectively). Similarly, 1-d treatment with Dex did not change  $\beta_1$ - or  $\beta_2$ -receptor mRNA expression. Continuous infusion of Iso significantly decreased the ratio of  $\beta_1$ - and  $\beta_2$ -receptor mRNA to GAPDH mRNA by  $69 \pm 7$  ( $P < 0.01$ ) and  $57 \pm 8\%$  ( $P = 0.01$ ), respectively. Combined treatment of Dex and Iso did not affect  $\beta_2$ -receptor mRNA expression, but significantly decreased  $\beta_1$ -receptor mRNA expression by  $66 \pm 8\%$  ( $P < 0.01$ ).

**Electrophoretic mobility shift assay.** The specificity of CREB-like DNA-binding activity was demonstrated by the abolition of the single complex formed by excess amount of unlabeled CREB consensus oligonucleotide and the absence of the complex in the control incubation lacking nuclear protein (Fig. 5 A). Chronic injection of Dex did not significantly affect the CREB-like DNA-binding activity, whereas continuous infusion of Iso significantly reduced the CREB-like DNA-binding activity ( $P < 0.05$ ). Combined Dex and Iso treatment showed no change in CREB-like DNA-binding activity (Fig. 5, B and C).

**Nuclear run-on transcription assay.** The transcription rates of  $\beta_1$ - and  $\beta_2$ -receptor genes were measured by nuclear run-on assays in lung tissues from control, Dex-, Iso-, or both treated groups. Chronic injection of Dex showed an increase in transcription rate of  $\beta_2$ -receptor gene without affecting  $\beta_1$ -receptor gene. The transcription rate of  $\beta_2$ -receptor gene, calculated from the ratio of transcription rate of  $\beta_2$ -receptor gene to that of the GAPDH gene, was increased by approximately twofold of control in nuclei from Dex group, whereas continuous infusion of Iso showed a decrease in transcription rate of  $\beta_1$ -receptor gene without affecting the transcription rate of  $\beta_2$ -receptor gene. Combined Dex and Iso treatment did not affect the transcription rate of either  $\beta_1$ - or  $\beta_2$ -receptor genes (Fig. 6, A and B).

## Discussion

In this study we investigated the effect of treatment of a glucocorticoid and a  $\beta$ -agonist, alone and in combination, on the

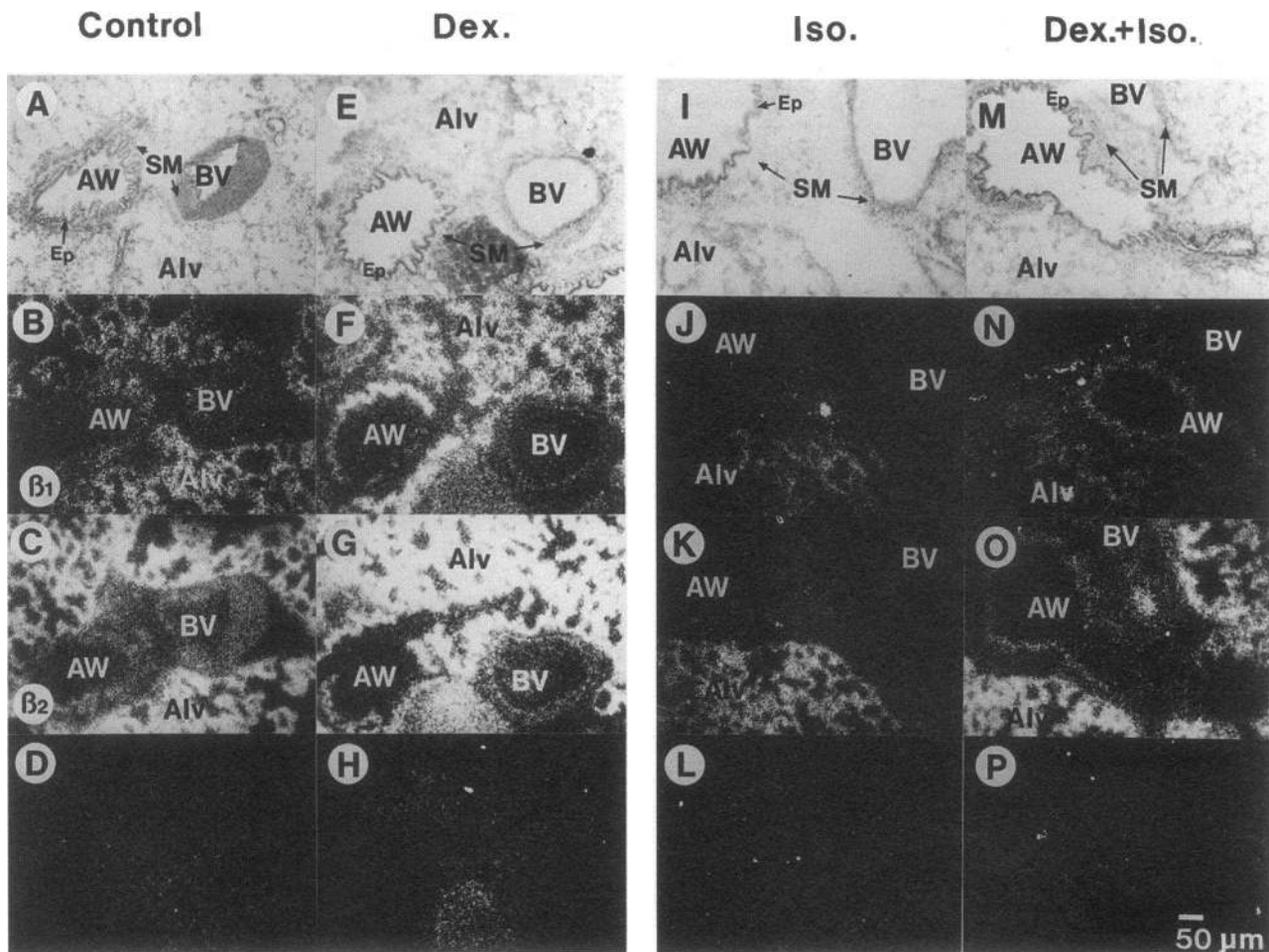


Figure 2. Distribution of  $\beta$ -adrenergic receptor subtypes in rat lung using in vitro receptor autoradiography in control (A–D), Dex-treated (E–H), Iso-treated (I–L), and Dex and Iso-treated (M–P) animals. A, E, I, and M show sections stained with 1% cresyl fast violet. B, F, J, and N show the distribution of  $\beta_1$ -receptor subtype in the presence of 0.1  $\mu$ M ICI 118,551. C, G, K, and O show the distribution of  $\beta_2$ -receptor subtype in the presence of 0.1  $\mu$ M CGP 20712. D, H, L, and P show the nonspecific binding of ICYP to lung sections. Alv, alveoli; AW, airway; BV, blood vessel; SM, airway smooth muscle; and Ep, airway epithelium. Bar, 50  $\mu$ m.

density and distribution of  $\beta_1$ - and  $\beta_2$ -receptors, their mRNA expression, and CREB-like DNA-binding activity in rat lung in vivo. Chronic treatment with Dex resulted in an increased density of both  $\beta_1$ - and  $\beta_2$ -receptors and an increase in the transcription rate of  $\beta_2$ -receptor gene, but no change in their mRNA expression, in the transcription rate of  $\beta_1$ -receptor gene, and in CREB-like DNA-binding activity. Chronic treatment with Iso resulted in a reduced density of both  $\beta_1$ - and  $\beta_2$ -receptors, decreases in their mRNA expression, in the transcription rate of  $\beta_1$ -receptor gene, and in CREB-like DNA-binding activity, but no change in the transcription rate of  $\beta_2$ -receptor gene. Combined treatment of Dex and Iso resulted in no change in  $\beta_2$ -receptor number nor its mRNA expression, while there was a reduction in  $\beta_1$ -receptor number and mRNA expression (Table II). Autoradiographic mapping showed that the changes in  $\beta$ -receptor expression after Dex, Iso, and combined treatment were present on all cell types identified, including airway smooth muscle, epithelium, and alveolar walls.

As observed previously (10), the present study also provides evidence that an increase in pulmonary  $\beta_1$ - and  $\beta_2$ -receptor levels after chronic in vivo dexamethasone treatment can be detected. In vitro, glucocorticoids have been shown to induce

the upregulation of  $\beta$ -receptors in DDT<sub>1</sub>-MF2 (17, 18) and 3T3-F442A (30) cell lines in culture. The downregulation of  $\beta_1$ - and  $\beta_2$ -receptors in rat lung after chronic stimulation by  $\beta$ -agonist is also in good agreement with previous in vivo and in vitro studies (7, 8, 19, 31). When rats were exposed to both Dex and Iso simultaneously,  $\beta_1$ -receptors were reduced, but  $\beta_2$ -receptors were unchanged. This protection of dexamethasone on  $\beta$ -agonist-promoted downregulation of  $\beta_2$ -receptors is consistent with the in vitro study using DDT<sub>1</sub>-MF2 cells (13). However, Dex had no protective effect on  $\beta$ -agonist-promoted downregulation of  $\beta_1$ -receptors.

In agreement with previous reports (7, 8, 31), the present study also provides evidence for a reduction in  $\beta_1$ - and  $\beta_2$ -receptor mRNA after chronic in vivo  $\beta$ -agonist treatment. The fall in  $\beta_1$ -receptor mRNA started at 2 h, maximum at 1 d, and persisted for 7 d while  $\beta_2$ -receptor mRNA did not change at 2 h but was significantly reduced at 1 and 7 d after continuous infusion with Iso (7). Furthermore, our data showed an apparent association between these reductions in mRNA and the reduction in CREB-like DNA-binding activity, suggesting the possible involvement of this transcription factor(s). CREB, a transcription factor, recognizes CRE and stimulates target gene tran-

Table 1. Influence of Treatment on  $\beta$ -Adrenergic Receptors by Receptor Autoradiography on Different Structures in Rat Peripheral Lung, as Determined by Optical Density

Structure	Treatment	$\beta_2$ -Subtype	$\beta_1$ -Subtype
Alveoli	Control	102.4 $\pm$ 3.2	38.0 $\pm$ 1.3
	Dex	184.0 $\pm$ 3.7*	60.0 $\pm$ 0.9*
	Iso	47.3 $\pm$ 1.7*	15.8 $\pm$ 0.4*
	Dex + Iso	103.1 $\pm$ 2.3	26.6 $\pm$ 0.9*
Vascular smooth muscle	Control	26.4 $\pm$ 1.8	14.8 $\pm$ 0.7
	Dex	62.9 $\pm$ 2.1*	26.7 $\pm$ 1.4*
	Iso	7.0 $\pm$ 0.5*	1.4 $\pm$ 0.2*
	Dex + Iso	24.5 $\pm$ 1.1	6.4 $\pm$ 0.6*
Endothelium	Control	18.4 $\pm$ 1.2	9.6 $\pm$ 0.5
	Dex	45.8 $\pm$ 1.3*	22.8 $\pm$ 1.0*
	Iso	4.2 $\pm$ 0.4*	0.6 $\pm$ 0.1*
	Dex + Iso	15.0 $\pm$ 0.7 <sup>‡</sup>	2.0 $\pm$ 0.4*
Airway smooth muscle	Control	18.3 $\pm$ 1.3	8.6 $\pm$ 0.7
	Dex	67.9 $\pm$ 3.1*	19.6 $\pm$ 1.4 <sup>§</sup>
	Iso	5.4 $\pm$ 0.6*	2.4 $\pm$ 0.4 <sup>§</sup>
	Dex + Iso	20.8 $\pm$ 1.3	7.0 $\pm$ 0.5 <sup>‡</sup>
Epithelium	Control	31.2 $\pm$ 1.8	19.4 $\pm$ 1.7
	Dex	92.9 $\pm$ 4.4*	37.6 $\pm$ 2.1*
	Iso	8.0 $\pm$ 0.9*	2.7 $\pm$ 0.4*
	Dex + Iso	27.1 $\pm$ 1.2	9.4 $\pm$ 0.7*

Values are the means $\pm$ SEM from 40 separate optical density measurements ( $\times 10^{-3}$ ) of multiple sections from three control and three treated animals.  $\beta_2$ -Receptor subtype was determined in the presence of 0.1  $\mu$ M CGP 20712 A and  $\beta_1$ -receptor subtype in the presence of 0.1  $\mu$ M ICI 118,551. <sup>‡</sup>  $P < 0.05$ , <sup>§</sup>  $P < 0.01$ , \*  $P < 0.001$ , as compared with the control values.

scription (14). The presence of CRE in the  $\beta_2$ -receptor gene and the ability of  $\beta$ -agonists to increase intracellular cAMP prompted the suggestion that these elements may be involved in the negative control of transcription (30). However, this is unlikely since activation of CRE enhances rather than suppresses transcription rates of target genes (16). Thus, it is possible that the reduced CREB-like DNA-binding activity contributes, in part, to the downregulation of  $\beta_2$ -receptor mRNA after prolonged exposure to a  $\beta$ -agonist, in good agreement with our previous findings in rat and guinea pig lung (7, 8). Recently, the presence of a CRE has also been identified in the 5'-flanking promoter region of the  $\beta_1$ -receptor gene (32) and may be involved in the negative control of transcription of this gene.

Although both  $\beta_1$ - and  $\beta_2$ -receptor densities were increased, neither  $\beta_1$ - nor  $\beta_2$ -receptor mRNA was increased after chronic in vivo glucocorticoid treatment, in agreement with an in vivo study using Sprague-Dawley rats (33). In contrast, induction of  $\beta_2$ -receptor mRNA by glucocorticoids has been demonstrated in human lung and several cell lines in vitro (17, 18, 34). The lack of change in the levels of  $\beta_1$ - and  $\beta_2$ -receptor mRNAs after dexamethasone in this study may be due to the fact that the accumulation of mRNA is transient. Indeed in our previous study in human lung in vitro (34), we observed that the increase in  $\beta_2$ -receptor mRNA was maximal 2 h after exposure to glucocorticoid and returned to baseline by 24 h. Glucocorticoids have the ability to downregulate their own receptors, which act as transcription factors (35). Several putative GREs are identified in the promoter region of the  $\beta_2$ -receptor gene and are obligatory for glucocorticoid regulation of receptor mRNA levels (15).

After combined treatment with both glucocorticoid and  $\beta$ -agonist,  $\beta_1$ -receptor mRNA and the density of  $\beta_1$ -adrenoceptors were reduced, but  $\beta_2$ -receptor mRNA and  $\beta_2$ -receptors remained unchanged, compared with control animals. The differential regulation of  $\beta_1$ - and  $\beta_2$ -receptor gene expression after

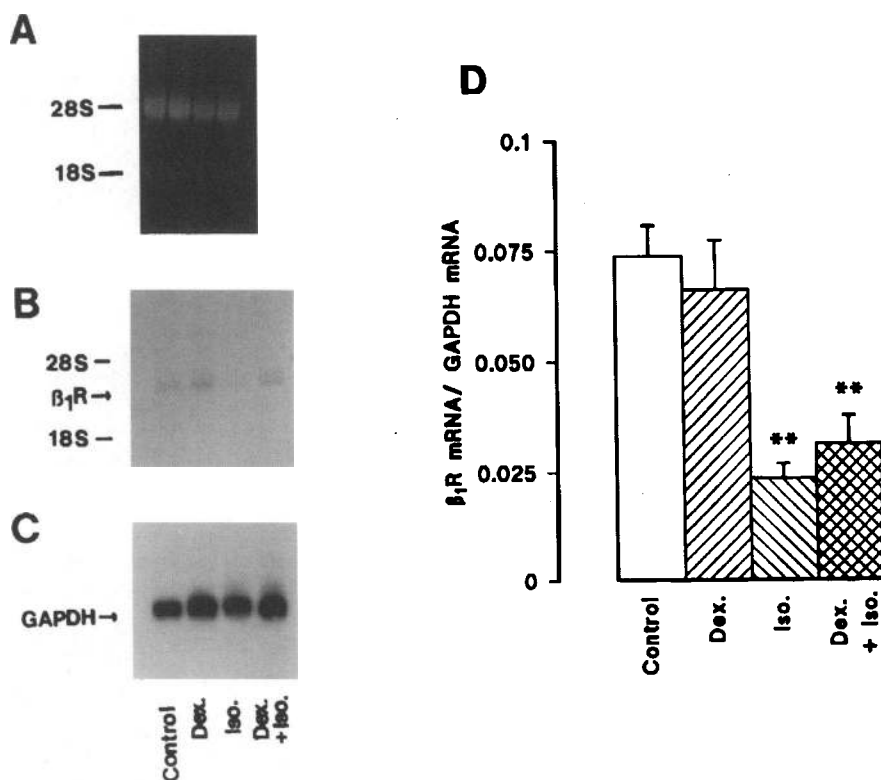


Figure 3. Effects of treatment with Dex and/or Iso on  $\beta_1$ -adrenergic receptor mRNA in rat lung. (A) A photograph of the ultraviolet light-illuminated, ethidium bromide staining of the RNA on the membrane used for Northern blot in B and C. The signal intensity of the 28S and 18S rRNA bands demonstrated the quantity of the RNA loaded into each lane. (B) Representative autoradiogram from Northern blot of rat  $\beta_1$ -receptor mRNA. Total RNA from one control, one Dex, one Iso, and one Dex and Iso-treated rat lung was hybridized with a  $^{32}$ P-labeled SmaI/PvuII fragment human  $\beta_1$ -receptor cDNA probe. The size of the mRNA (in kilobases) was estimated from rRNA markers as 3.2 kb. (C) Representative autoradiogram from Northern blot of rat GAPDH mRNA. The same membrane as in A was later probed with a  $^{32}$ P-labeled rat GAPDH cDNA probe to control for the quantity of the RNA loaded into each lane. (D) Densitometric measurement of  $\beta_1$ -receptor mRNA from control, Dex-treated, Iso-treated, and Dex and Iso-treated rat lungs ( $n = 7$  in each group).  $\beta_1$ -receptor mRNA was normalized to that for GAPDH mRNA. Significance of difference from the control value, \*\*  $P < 0.01$ .

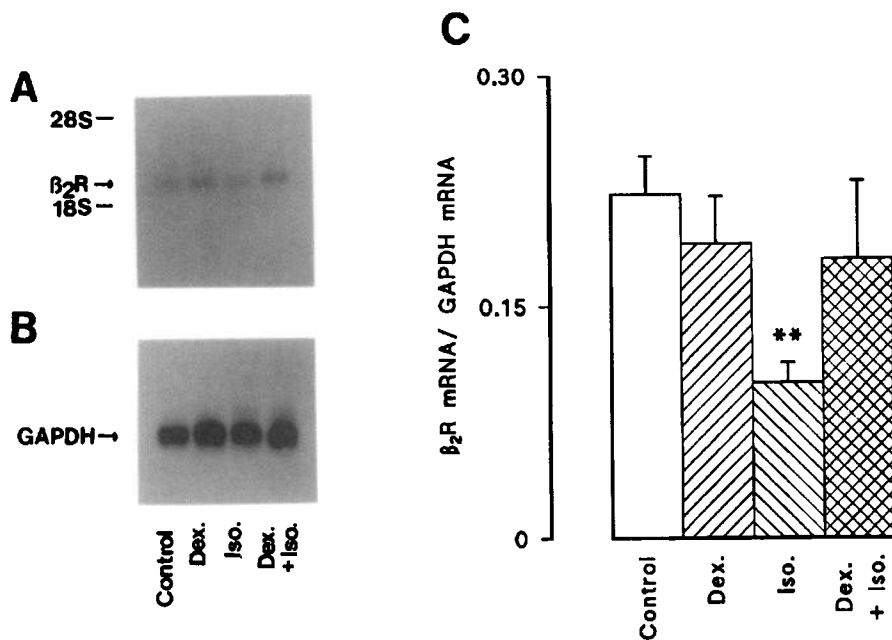


Figure 4. Effects of treatment with Dex and/or Iso on  $\beta_2$ -adrenergic receptor mRNA in rat lung. (A) Representative autoradiogram from Northern blot of rat  $\beta_2$ -receptor mRNA using rat full-length  $\beta_2$ -receptor cDNA. The same membrane as used in Fig. 3. was hybridized with a  $^{32}\text{P}$ -labeled full-length rat  $\beta_2$ -receptor cDNA probe. The size of the mRNA (in kilobases) was estimated from rRNA markers as 2.2 kb. (B) Representative autoradiogram from Northern blot of rat GAPDH mRNA. (C) Densitometric measurement of  $\beta_2$ -receptor mRNA from control, Dex-treated, Iso-treated, and Dex and Iso-treated rat lungs ( $n = 7$  in each group).  $\beta_2$ -receptor mRNA was normalized to that for GAPDH mRNA. Significance of difference from the control value, \*\*  $P < 0.01$ .

combined treatment of Dex and Iso may be complex. Glucocorticoids alone have been shown to selectively upregulate  $\beta_2$ -receptors in brown fat, 3T3-L1 preadipocytes, and 3T3-F442

adipocytes, with no effect or an actual decrease in the expression of  $\beta_1$ -receptors (29, 36, 37).

The molecular explanation for regulation of receptor

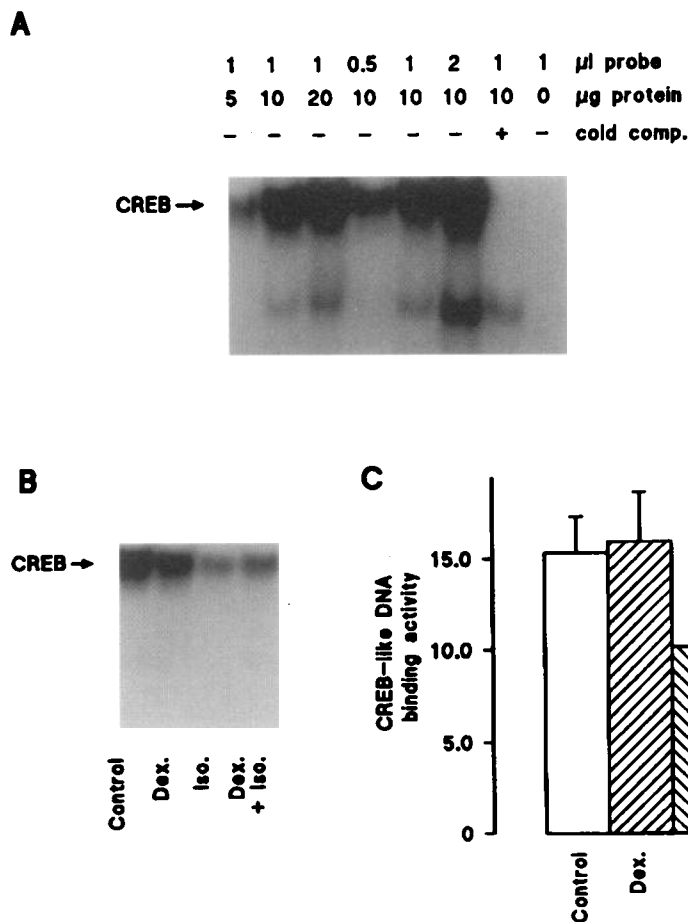
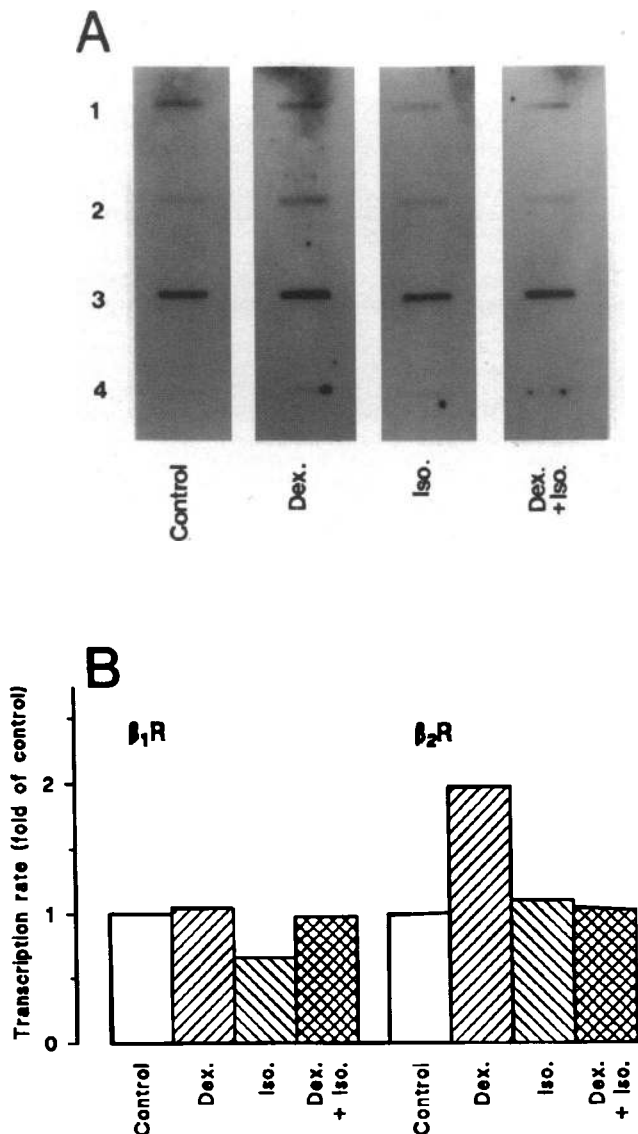


Figure 5. Effects of treatment with Dex and/or Iso on CREB-like DNA-binding activity in rat lung. (A) Specificity of the nuclear protein-CREB consensus oligonucleotide complex. The indicated amounts of nuclear protein from one control animal were incubated with the indicated amounts of  $^{32}\text{P}$ -labeled CREB consensus oligonucleotide in the presence or absence of unlabeled competitor oligonucleotide. Arrow indicates the specific complex. (B) Representative autoradiogram from gel shift assay of CREB in rat lung. Nuclear protein from one control, one Dex-treated, one Iso-treated, and one Dex and Iso-treated rat lung was hybridized with a  $^{32}\text{P}$ -labeled CREB consensus oligonucleotide. Arrow indicates the specific complex. (C) Densitometric measurement of CREB-like DNA-binding activity from control, Dex-treated, Iso-treated, and Dex and Iso-treated rat lungs ( $n = 5$  in each group). Significant different from the control value. \*  $P < 0.05$ .



**Figure 6.** Effects of treatment with Dex and/or Iso on the transcription rates of  $\beta_1$ - and  $\beta_2$ -adrenergic receptor gene in rat lung. (A) Representative autoradiograms from nuclear run-on transcription assays on  $\beta_1$ - and  $\beta_2$ -receptor gene in control, Dex-treated, Iso-treated, and Dex and Iso-treated rat lung. The tissues were frozen and nuclei were prepared as outlined in Methods. Transcription was performed using [ $\alpha^{32}$ P]UTP and unlabeled nucleotides. Labeled RNA was then isolated and hybridized either to plasmid (10  $\mu$ g/slot) containing the full-length human  $\beta_1$ -receptor cDNA, rat  $\beta_2$ -receptor cDNA, GAPDH cDNA, or to the plasmid lacking the cDNA insert (pGEM-3Z) as control. 1,  $\beta_1$ -receptor; 2,  $\beta_2$ -receptor; 3, GAPDH; 4, pGEM-3Z. (B) Densitometric measurement of  $\beta_1$ - and  $\beta_2$ -receptor transcription rate from control, Dex-treated, Iso-treated, and Dex and Iso-treated rat lungs. The transcription rate was calculated as the ratio of  $\beta_1$ - or  $\beta_2$ -receptor cDNA signal relative to the GAPDH cDNA signal. Average values from two separate experiments are shown.

mRNA, the balance and interplay between glucocorticoid induced upregulation and agonist-promoted downregulation, has been demonstrated in cultured cell lines (13). Glucocorticoids enhance the rate of transcription at a GRE in the 5'-noncoding portion of the gene which is responsible for the steroid-induced upregulation of receptor mRNA, and the stability of mRNA

**Table II. Summary of Results**

	Treatment		
	Dex	Iso	Dex + Iso
<b><math>\beta_1</math>-Receptors</b>			
Number	↑	↓	↓
mRNA	No change	↓	↓
Rate of transcription	No change	↓	No change
<b><math>\beta_2</math>-Receptors</b>			
Number	↑	↓	No change
mRNA	No change	↓	No change
Rate of transcription	↑	No change	No change
<b>CREB-like DNA-binding activity</b>			
	No change	↓	No change

appears to play no major role in the glucocorticoid effect. On the other hand,  $\beta$ -agonists reduce the half-life of mRNA and this is responsible for the short-term agonist-induced downregulation of receptor mRNA, without any effect on the rate of transcription. An intriguing finding in the present study was the observation that there was no change on the steady state level of  $\beta_1$ - and  $\beta_2$ -receptor mRNA after chronic in vivo glucocorticoid treatment, indicating that the observed increase in  $\beta_1$ - and  $\beta_2$ -receptor density might reflect the involvement of different molecular mechanisms. For  $\beta_1$ -receptor, the synthesis of receptor protein may be unchanged but the stability of receptor protein may be increased. On the other hand, both an increase in the rate of transcription of the  $\beta_2$ -receptor gene as well as an consequent increase in the synthesis of the receptor protein may occur. After chronic in vivo  $\beta$ -agonist, the observed decrease in  $\beta_1$ -receptor mRNA and rate of transcription, followed by a decrease in  $\beta_1$ -receptor density, suggests that it is primarily the reduced rate of transcription which is responsible for the  $\beta$ -agonist-induced downregulation of  $\beta_1$ -receptor mRNA. On the other hand, the decrease in  $\beta_2$ -receptor mRNA and density without any detectable decline in the rate of transcription indicates that  $\beta$ -agonist might destabilize  $\beta_2$ -receptor mRNA. In these studies with chronic in vivo  $\beta$ -agonist and/or glucocorticoid treatment, it is not possible to measure whether mRNA stability is changed, as it is not possible to perform the transcriptional blocking studies that can be carried out in vitro. Recently, a regulatory factor, known as  $\beta$ -adrenergic receptor-binding protein, has been identified that selectively modulates the stability of  $\beta_2$ -receptor mRNA in DDT<sub>1</sub>-MF2 cells (38). The abundance of this factor varies inversely with the level of receptor mRNA, being induced by  $\beta$ -agonists that downregulate receptor mRNA, and being reduced by glucocorticoids that upregulate receptor mRNA.

It is now increasingly recognized that chronic treatment with  $\beta$ -agonist bronchodilators may result in desensitization to the protective effects of  $\beta$ -agonists on the airways (3-6). Our results suggest that concomitant treatment with glucocorticoids should prevent changes in  $\beta_2$ -receptors which mediate most of the antiasthma effects of  $\beta$ -agonists, whereas the desensitization of  $\beta_1$ -receptors would remain. Taken together, our findings suggest that different molecular mechanisms may be involved in the regulation of  $\beta_1$ - and  $\beta_2$ -receptor expression after chronic in vivo  $\beta$ -agonist and/or glucocorticoid treatment. This may be

of clinical benefit, since some of the cardiovascular side effects of  $\beta$ -agonists may be mediated via stimulation of cardiac  $\beta_1$ -receptors.

## Acknowledgments

The authors are grateful to Dr. R. J. Lefkowitz (Duke University, Durham, NC) for providing the cDNA probe for human  $\beta_1$ -adrenergic receptors and to Dr. T. R. Bai (University of British Columbia, St. Paul's Hospital, Vancouver, BC, Canada) for providing the cDNA probe for rat GAPDH. The authors also wish to thank Mr. Paul Seldon for printing the photographs.

This study was supported by National Institutes of Health grant HL-45947, the British Lung Foundation, and the National Asthma Campaign (United Kingdom).

## References

1. Tattersfield, A. E., and P. J. Barnes. 1992.  $\beta_2$ -Agonists and corticosteroids: new developments and controversies. *Am. Rev. Respir. Dis.* 146:1637-1641.
2. Barnes, P. J. 1993.  $\beta$ -Adrenoceptors on smooth muscle, nerves and inflammatory cells. *Life Sci.* 52:2101-2109.
3. O'Connor, B. J., S. L. Aikman, and P. J. Barnes. 1992. Tolerance to the non-bronchodilator effects of inhaled  $\beta_2$ -agonists in asthma. *N. Engl. J. Med.* 327:1204-1208.
4. Cockcroft, D., C. P. McParrand, S. A. Britto, V. A. Swystun, and C. Rutherford. 1993. Regular inhaled salbutamol and airway responsiveness to allergen. *Lancet.* 342:833-837.
5. Sears, M. R., D. R. Taylor, C. G. Print, D. G. Lake, Q. Li, E. M. Flannery, D. M. Yates, M. K. Lucas, and G. P. Herbison. 1990. Regular inhaled  $\beta$ -agonist treatment in bronchial asthma. *Lancet.* 336:1391-1396.
6. Taylor, D. R., M. R. Sears, G. P. Herbison, E. M. Flannery, C. G. Print, D. C. Lake, D. M. Yates, M. K. Lucas, and Q. Li. 1993. Regular inhaled  $\beta$ -agonist in asthma: effects on exacerbation and lung function. *Thorax.* 48:134-138.
7. Nishikawa, M., J. C. W. Mak, H. Shirasaki, and P. J. Barnes. 1993. Differential down-regulation of pulmonary  $\beta_1$ - and  $\beta_2$ -adrenoceptor messenger RNA with prolonged *in vivo* infusion of isoprenaline. *Eur. J. Pharmacol. (Mol. Pharmacol.)* 247:131-138.
8. Nishikawa, M., J. C. W. Mak, H. Shirasaki, S. E. Harding, and P. J. Barnes. 1994. Long term exposure to norepinephrine results in down-regulation and reduced mRNA expression of pulmonary  $\beta$ -adrenergic receptors in guinea pig. *Am. J. Respir. Cell Mol. Biol.* 10:91-99.
9. Davies, A. O., and R. J. Lefkowitz. 1984. Regulation of  $\beta$ -adrenergic receptors by steroid hormones. *Annu. Rev. Physiol.* 46:119-130.
10. Mano, K., A. Akbarzadel, and R. G. Townley. 1979. Effect of hydrocortisone on beta-adrenergic receptors in lung membrane. *Life Sci.* 25:1925-1930.
11. Davies, A. O., and R. J. Lefkowitz. 1981. Agonist promoted high affinity state of the beta-adrenergic receptor in human neutrophils: modulation by corticosteroids. *J. Clin. Endocrinol. & Metab.* 53:703-708.
12. Hui, K. K., M. E. Conolly, and D. P. Tashkin. 1982. Reversal of human lymphocyte beta-adrenoceptor desensitization by glucocorticoids. *Clin. Pharmacol. & Ther.* 32:556-565.
13. Hadcock, J. R., H. Wang, and C. C. Malbon. 1989. Agonist-induced destabilization of  $\beta$ -adrenergic receptor mRNA. Attenuation of glucocorticoid-induced up-regulation of  $\beta$ -adrenergic receptors. *J. Biol. Chem.* 264:19928-19933.
14. Collins, S., J. Altschmied, O. Herbsman, M. G. Caron, P. L. Mellon, and R. J. Lefkowitz. 1990. A cAMP response element in the  $\beta_2$ -adrenergic receptor gene confers transcriptional autoregulation by cAMP. *J. Biol. Chem.* 265:19330-19335.
15. Malbon, C. C., and J. R. Hadcock. 1988. Evidence that glucocorticoid response elements in the 5' non-coding region of the hamster  $\beta_2$ -adrenergic receptor gene are obligate for glucocorticoid regulation of hamster mRNA levels. *Biochem. Biophys. Res. Commun.* 154:676-681.
16. Collins, S., M. G. Caron, and R. J. Lefkowitz. 1992. From ligand binding to gene expression: new insights into the regulation of G-protein-coupled receptors. *Trends Biochem. Sci.* 17:37-40.
17. Collins, S., M. G. Caron, and R. J. Lefkowitz. 1988.  $\beta_2$ -Adrenergic receptors in hamster smooth muscle cells are transcriptionally regulated by glucocorticoids. *J. Biol. Chem.* 263:9067-9070.
18. Hadcock, J. R., and C. C. Malbon. 1988. Regulation of  $\beta$ -adrenergic receptors by permissive hormones; glucocorticoids increase steady-state levels of receptor mRNA. *Proc. Natl. Acad. Sci. USA.* 85:8415-8419.
19. Galant, S. P., L. Duriseti, S. Underwood, and P. A. Insel. 1978. Decreased beta-adrenergic receptors on polymorphonuclear leukocytes after adrenergic therapy. *N. Engl. J. Med.* 229:933-936.
20. Hauck, R. W., M. Böhm, S. Gengenback, L. Sunder-Plassmann, G. Fruhmman, and E. Endmann. 1990.  $\beta_2$ -Adrenoceptors in human lung and peripheral mononuclear leukocytes of untreated and terbutaline-treated patients. *Chest.* 98:376-381.
21. Carstairs, J. R., A. J. Nimmo, and P. J. Barnes. 1985. Autoradiographic visualization of beta-adrenoceptor subtypes in human lung. *Am. Rev. Respir. Dis.* 132:541-547.
22. Lowry, O. H., N. J. Rosebrough, A. L. Farr, and R. J. Randall. 1951. Protein measurement with the Folin phenol reagent. *J. Biol. Chem.* 193:265-275.
23. Gocayne, J., D. A. Robinson, M. G. Fitzgerald, F.-Z. Chung, A. R. Kerlavage, K.-U. Lentes, J. Lai, C.-D. Wang, C. M. Fraser, and J. C. Venter. 1987. Primary structure of rat cardiac  $\beta$ -adrenergic and muscarinic cholinergic receptors obtained by automated DNA sequence analysis: further evidence for a multigene family. *Proc. Natl. Acad. Sci. USA.* 84:8296-8300.
24. Subramaniam, M., D. Colvard, P. E. Keeting, K. Rasmussen, B. L. Riggs, and T. C. Spelsberg. 1992. Glucocorticoid regulation of alkaline phosphatase, osteocalcin, and proto-oncogenes in normal human osteoblast-like cells. *J. Cell. Biochem.* 50:411-424.
25. Chromczynski, P., and N. Sacchi. 1987. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* 162:156-159.
26. Peters, M. J., I. M. Adcock, C. R. Brown, and P. J. Barnes. 1995.  $\beta$ -Adrenoceptor agonists interfere with glucocorticoid receptor DNA binding in rat lung. *Eur. J. Pharmacol. (Mol. Pharmacol.)* 289:275-281.
27. Greenberg, M. E., and T. P. Bender. 1992. Identification of newly transcribed RNA. In *Current Protocols in Molecular Biology*. F. M. Ausubel, R. Brent, R. E. Kingston, D. D. Moore, J. G. Seidman, J. A. Smith, and K. Struhl, editors. John Wiley & Sons, New York. 4.10.1-4.10.9.
28. Hamid, Q. A., J. C. W. Mak, M. N. Sheppard, B. Corrin, J. C. Venter, and P. J. Barnes. 1991. Localization of  $\beta_2$ -adrenoceptor messenger RNA in human and rat lung using *in situ* hybridization: correlation with receptor autoradiography. *Eur. J. Pharmacol. (Mol. Pharmacol.)* 206:133-138.
29. Frielle, T., S. Collins, K. W. Daniel, M. G. Caron, R. J. Lefkowitz, and B. K. Kobilka. 1987. Cloning of the cDNA for the human  $\beta_1$ -adrenergic receptor. *Proc. Natl. Acad. Sci. USA.* 84:7920-7924.
30. Feve, B., L. J. Emorine, M. M. Briand-Sutren, F. Lasnier, A. D. Strosberg, and J. Pariault. 1990. Differential regulation of  $\beta_1$ - and  $\beta_2$ -adrenergic receptor protein and mRNA levels by glucocorticoids during 3T3-F442A adipose differentiation. *J. Biol. Chem.* 265:16343-16349.
31. Hadcock, J. R., and C. C. Malbon. 1988. Down-regulation of  $\beta$ -adrenergic receptors: agonist-induced reduction in receptor mRNA levels. *Proc. Natl. Acad. Sci. USA.* 85:5021-5025.
32. Collins, S., J. Ostrowski, and R. J. Lefkowitz. 1993. Cloning and sequence analysis of the human  $\beta_1$ -adrenergic receptor 5'-flanking promoter region. *Biochim. Biophys. Acta.* 1172:171-174.
33. McGraw, D. W., S. E. Chai, F. C. Hiller, and L. E. Cornett. 1993.  $\beta_2$ -adrenergic receptor and mRNA levels in rat lungs following *in vitro* administration of glucocorticoids. *Am. Rev. Respir. Dis.* 147:A275. (Abstr.)
34. Mak, J. C. W., M. Nishikawa, and P. J. Barnes. 1995. Glucocorticosteroids increase  $\beta_2$ -adrenergic receptor transcription in human lung. *Am. J. Physiol.* 268:L41-L46.
35. Rosewicz, S., A. R. McDonald, B. A. Maddux, I. D. Goldfine, R. J. Miesfeld, and C. D. Logsdon. 1988. Mechanism of glucocorticoid receptor down-regulation by glucocorticoids. *J. Biol. Chem.* 263:2581-2584.
36. Scarpace, P. J., L. A. Baresi, and J. E. Morley. 1988. Glucocorticoids modulate  $\beta$ -adrenoceptor subtypes and adenylate cyclase in brown fat. *Am. J. Physiol.* 255:E153-E158.
37. Nakada, M. T., K. M. Haskell, D. J. Ecker, J. M. Stadel, and S. T. Crooke. 1989. Genetic regulation of  $\beta_2$ -adrenergic receptors in 3T3-L1 fibroblasts. *Biochem. J.* 260:53-59.
38. Port, J. D., L.-Y. Huang, and C. C. Malbon. 1992.  $\beta$ -Adrenergic agonists that down-regulate receptor mRNA upregulate a M<sub>r</sub> 35,000 protein(s) that selectively binds to  $\beta$ -adrenergic receptor mRNAs. *J. Biol. Chem.* 267:24103-24108.