

Review Article

Protective Effects of Dietary Antioxidants against Vanadium-Induced Toxicity: A Review

Iwona Zwolak 

Laboratory of Oxidative Stress, Centre for Interdisciplinary Research, The John Paul II Catholic University of Lublin, Konstantynów 1 J, 20-708 Lublin, Poland

Correspondence should be addressed to Iwona Zwolak; iglina@kul.lublin.pl

Received 12 July 2019; Accepted 23 November 2019; Published 9 January 2020

Academic Editor: Fabiana Morroni

Copyright © 2020 Iwona Zwolak. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Vanadium (V) in its inorganic forms is a toxic metal and a potent environmental and occupational pollutant and has been reported to induce toxic effects in animals and people. *In vivo* and *in vitro* data show that high levels of reactive oxygen species are often implicated in vanadium deleterious effects. Since many dietary (exogenous) antioxidants are known to upregulate the intrinsic antioxidant system and ameliorate oxidative stress-related disorders, this review evaluates their effectiveness in the treatment of vanadium-induced toxicity. Collected data, mostly from animal studies, suggest that dietary antioxidants including ascorbic acid, vitamin E, polyphenols, phytosterols, and extracts from medicinal plants can bring a beneficial effect in vanadium toxicity. These findings show potential preventive effects of dietary antioxidants on vanadium-induced oxidative stress, DNA damage, neurotoxicity, testicular toxicity, and kidney damage. The relevant mechanistic insights of these events are discussed. In summary, the results of studies on the role of dietary antioxidants in vanadium toxicology appear encouraging enough to merit further investigations.

1. Introduction

It is well known that one of the mechanisms associated with the toxicity of heavy metals is oxidative stress [1] defined as an imbalance between the production of reactive oxygen species and antioxidant defenses, which may lead to tissue injury [2]. Currently, the preferred medical treatment of metal poisoning includes chelation therapy, which facilitates removal of the excess of the metal from soft tissues and excretion in urine. However, the serious side effects that may occur during the chelation therapy such as depletion of essential minerals, prooxidant effects, hepatic and renal toxicity, and no removal of heavy metals from intracellular compartments are the major drawbacks of this treatment [3]. Since oxidative stress plays a pivotal role in the pathogenesis of metal toxicity, the use of antioxidants as a supplementary therapy to conventional chelation treatment has been proposed [3, 4].

In literature, the beneficial action of dietary and plant-derived antioxidants on toxicity of some of the heavy metals has been reported. For example, flavonoids such as epigallocatechin gallate and curcumin have been shown to possess

protective activity against cadmium-induced nephrotoxicity in rats [5, 6]. Catechin hydrate has been found to reduce cytotoxic and genotoxic effects of cadmium in human peripheral blood lymphocytes [7]. Sulforaphane (organosulfur compound) protected human mesenchymal stem cells against cadmium-induced changes in nuclear morphology, depletion of mitochondrial membrane potential, and alteration of gene expression [8]. The administration of epigallocatechin gallate attenuated arsenic-induced oxidative damage in the liver of rats [9]. An inhibitory effect of dietary supplement containing a mixture of grape seed extract, tea polyphenols, and probiotics on toxicity of lead to mice has also been reported [10]. A human study with car battery workers found that garlic supplementation significantly reduced signs of occupational chronic lead poisoning such as irritability, headache, and mean systolic blood pressure [11]. In the same study, garlic has also significantly decreased blood lead concentrations.

In this review, the focus is placed on studies evaluating the use of natural antioxidant compounds against vanadium toxicity. In industry, vanadium is a widely used transition metal, and the global demand and production of vanadium

is on the increase [12]. This certainly raises concerns over the detrimental effects of vanadium excess on human and animal health. People who were reported to encounter vanadium toxicity are those occupationally exposed to vanadium as well as those living in areas with high vanadium content in the air or water (described in the next chapter). In view of the fact that oxidative stress is an underlying mechanism of vanadium-induced toxicity, natural compounds with antioxidant properties are gaining increasing attention as cheap and safe antidotes against vanadium. To this end, this review explores the past and current data on the effectiveness of antioxidants of diet (vitamins C and E, polyphenols, phytochemicals, and plant extracts) in treatment of vanadium toxicity examined in animal and cell culture models. Additionally, possible direct and indirect mechanisms that could be involved in the beneficial activity of these antioxidants against vanadium have also been suggested. Studies identified in this review may help in the development of dietary strategies to improve protection of humans at high risk of vanadium toxicity.

2. Overview of Vanadium

Vanadium occurs as a natural component of the earth crust (in various minerals, coal, and crude oil) and is released to the environment mainly due to human activities. The unique chemical and physical features of vanadium compounds make it an indispensable material in many industries. Its compounds are frequently used in the production of steel and titanium-aluminum alloys, as catalysts in the sulfuric acid manufacture, and in the production of pigments, inks, and varnishes [13, 14]. The latest use of vanadium involves green technologies and the production of vanadium-based redox flow batteries, which can store electricity produced from renewable sources such as wind or sun. These very efficient and increasingly popular energy storage systems have already been installed, e.g., in China, the USA, Germany, and Japan [15]. The industrial use of vanadium is on the increase and so is the release of vanadium to the environment [12]. Vanadium is one of the elements listed on the second drinking water contaminant candidate list (CCL-2) that was announced by the United States Environmental Protection Agency in 2005. This is a list of contaminants that are known or anticipated to occur in public water systems and may require future regulations [16]. Vanadium was reported to contribute to soil pollution. For example, soils from the stone coal smelting district in Hunan province of China had vanadium concentrations in the range from 168 to 1538 mg/kg, which substantially exceeded Canadian soil quality standards (130 mg/kg) [17]. Yang et al. [12] conducted research of the national soil pollution in China, and the results showed that 26.49% of soils were contaminated with vanadium in the southwest of China and 8.6% of the national soil pollution survey samples were contaminated with vanadium presenting a threat to the public and environment. Heavy oil combustion contributes to the release of vanadium as a component adhering to fine particulate matter (PM_{2.5}) observed in large urban and industrial agglomerations such as New York City, the USA [18], and Jeddah, Saudi Arabia [19].

High groundwater concentrations of vanadium of natural geological sources have been noted in volcanic areas. For example, in some areas of Mt. Etna, groundwater vanadium content exceeded the Italian legal limit of 140 µg/l; the consequently estimated daily intake of vanadium in children calculated in this study was in the range of 0.4-11 µg/kg/bw, which was much higher than the estimated daily intakes of 0.09-0.34 µg/kg/day reported in the literature [20]. Vanadium excess can be toxic and detrimental to human health like any other metal. For instance, occupational inhalation exposure to vanadium was found to induce, e.g., acute respiratory symptoms in boiler makers [21], DNA damage in blood cells of workers from a vanadium pentoxide factory [22], and altered neurobehavioral functions in Chinese workers [23]. In turn, environmental overexposures to vanadium oxides attached to fine particulate matter (PM_{2.5}) were associated with, e.g., increased risk of respiratory symptoms in children of New York City [24] and a higher risk of cardiovascular and respiratory hospitalizations of older people in US counties [25]. Recently, urinary vanadium concentrations during pregnancy were positively associated with impaired fetal growth [26] and preterm or early-term delivery [27] in China. Association between the high level of trace elements (including vanadium) in the drinking water in the Mt. Etna volcanic area and the increased thyroid cancer incidence was suggested by Malandrino et al. [28]. A suicidal death after ingestion of an undetermined amount of ammonium vanadate has also been reported [29]. In addition, laboratory-based studies conducted in animal models or cell cultures found that vanadium exposure can induce a variety of toxic effects such as cardiovascular effects (e.g., vascular endothelial dysfunction [30] and arterial hypertension [31]), immune toxicity (e.g., damage to the spleen [32] and thymus [33]), neurotoxicity (e.g., hippocampal alterations [34] and memory loss [35]), developmental disturbances (e.g., increased embryoletality and skeletal defects [36]), and pulmonary toxicity [37]. It should be added that, besides the dose of vanadium and the route of vanadium exposure, many other factors such as the form of vanadium (inorganic versus organic forms) and interactions with other elements such as selenium [38, 39] or magnesium [40] can also influence vanadium toxicity (depicted in Figure 1).

Along with the studies of the toxic effects of vanadium, many investigators have been focused on the examination of potential medical applications of this mineral. These include antidiabetic (insulin-mimetic) actions, antiviral effects, and anticarcinogenic activity. Among these effects, the antidiabetic action of vanadium complexes with organic ligands has been very intensively studied since 1990s [41]. One of such compounds is bis(ethylmaltolato)oxidovanadium(IV) (BEOV), which entered into stage II clinical trials. However, due to kidney problems in some patients, BEOV as an antidiabetic agent could not progress to the next phase of research. Indeed, the risks associated with vanadium intoxication such as vanadium-induced reactive oxygen species generation, adverse effects on the immune system, and a risk of mutagenesis are listed among the arguments against the antidiabetic application of vanadium [42]. Domingo and Gomez [43] reviewed the results of past and

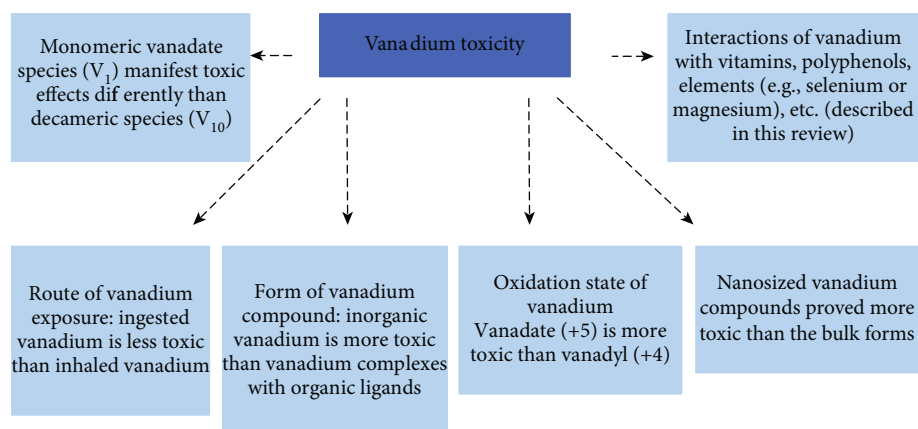


FIGURE 1: Factors affecting the toxicity of vanadium, according to References [13, 130, 131].

recent human studies on vanadium in diabetes and concluded that the use of vanadium compounds in oral diabetes therapy is misplaced. Vanadium compounds have attracted interest of researchers as potential antitumor agents. For example, promising *in vitro* and *in vivo* anticancer activity was demonstrated for oxidovanadium(IV) complex with flavonoid chrysin [44, 45]. Vanadium as vanadyl sulfate has been used by weight training athletes as a nutritional supplement that can increase muscle mass [46]. The role of vanadium in muscle development has been emphasized to be associated with its insulin-mimetic properties and anabolic effects [47]. So far, however, human studies have failed to demonstrate significant effects of vanadium on the body composition and performance enhancement, and the use of vanadium as a sport nutrition supplement is not recommended [46]. Vanadium is also a well-known constituent of the most commercialized titanium alloy named Ti-6Al-4V, which has been widely used in the manufacture of biomedical implants such as artificial hip joints, knee joints, and dental implants due to its excellent physical and mechanical properties [48]. Again, however, the potential cytotoxicity of vanadium limits the medical value of the Ti-6Al-4V alloy [49]. Recently, for example, a case of systemic allergic dermatitis to vanadium has been reported in a patient following placement of a titanium alloy (Ti-6Al-4V) plate in the left foot [50]. Summing up, due to the intensive use of vanadium in industry and the vanadium environmental pollution often related with it as well as the popularity of vanadium-based dietary supplements and medicinal applications of vanadium compounds, increasing numbers of humans are likely to experience the exposure to vanadium compounds in the near future.

3. Metabolism and Vanadium Detoxification Modes

Vanadium enters the human body via the gastrointestinal tract or respiratory system. In the bloodstream, transferrin is the major serum protein of vanadium transport from blood into tissues [51]. Other serum proteins, i.e., albumin, hemoglobin, and immunoglobulin, and low-molecular ligands, e.g., lactate and citrate, can be involved in the blood transport

of vanadium as well. From the blood, vanadium is transferred to different tissues such as the liver, kidney, heart, spleen, brain, and bones [52]. Final excretion of absorbed vanadium occurs through urine [53]. In the human body, vanadium can exist in oxidation state +5 (vanadate ions) or +4 (vanadyl cations). Cellular uptake of vanadium species proceeds via receptor-mediated endocytosis of vanadium-laden proteins (transferrin, albumin), phosphate or sulfate ion channels, or membrane citrate transporters [52]. Reductants, e.g., glutathione, ascorbic acid, or NADH, convert pentavalent vanadium to a tetravalent state (vanadyl), the latter being regarded as a predominant oxidation state of vanadium within the cell. Simultaneously, oxidants such as NAD⁺, O₂, and O₂²⁻ can oxidize vanadyl back to vanadate [54, 55].

Metabolic detoxification of vanadium possibly involves (1) reduction of vanadate to vanadyl by cellular reductants (mentioned above) and (2) complexation reactions during which vanadyl interacts with cellular agents such as reduced glutathione (GSH), an oxidized form of glutathione (GSSG), L-cysteine, and cystine forming stable, nonharmful complexes [56]. In addition, vanadium accumulates in bones by replacing bone phosphate in apatite Ca₅(PO₄)OH with vanadate [41]. The storage of vanadium in bones is also recognized as a potent detoxification mechanism of vanadium in animals [56].

Pharmacological treatment of vanadium poisoning is based on chelation therapy, which is the basic medical strategy for the treatment of acute and chronic intoxication with metals. Chelating agents are organic or inorganic compounds that can bind metal ions to form a stable, water-soluble complex with low toxicity, which are subsequently excreted from the body [3, 56]. With regard to the treatment of vanadium toxicity, calcium disodium ethylenediaminetetraacetate (CaNa₂EDTA) has been found to enhance excretion of vanadium in calves. Simultaneously, however, the chelator did not protect against pathological damage caused by vanadium [57]. Another chelator, Tiron (4,5-dihydroxybenzene-1,3-disulfonate), was partly effective in reducing vanadium-induced behavioral toxicity in rats [58]. Other potential chelating antidotes for vanadium intoxication include desferrioxamine B, *meso*-2,3-disulfanylsuccinic acid (DMSA), and 2,3-disulfanylpropane-1-sulfonic acid (DMPS) [56].

However, besides benefits, the therapy with the drugs mentioned above may also induce side effects. For example, CaNa_2EDTA was reported to cause nephrotoxicity, fever, headache, hepatotoxicity, and gastrointestinal symptoms. In addition, prolonged administration of CaNa_2EDTA decreases the levels of essential metals. Treatment with DMSA and DMPS was reported to be associated with skin reactions, gastrointestinal discomfort, and elevated liver enzymes [3].

In contrast to the aforementioned chelating compounds, ascorbic acid was suggested to be a very effective and safe pharmacologic agent for the treatment of vanadium toxicity in humans [56]. Detoxification of vanadium by ascorbic acid mainly relies on ascorbic acid-mediated reduction of vanadate to vanadyl and its high capacity to scavenge reactive oxygen species. Furthermore, vanadyl was found to interact with oxidation products of ascorbic acid forming stable complexes, which may allow excretion of vanadium from the organism [59]. In addition, the results of studies from our group have shown that pyruvic acid could be another potential antidote for the treatment of vanadium toxicity [60]. The studies showed that this alpha-keto acid protected against vanadium-induced oxidative stress and cytotoxicity in a cell culture model. The mechanism of protection probably involves antioxidative effects of pyruvate, especially its ability to neutralize hydrogen peroxide, but still more research is required to elucidate this issue [60].

4. Role of Oxidative Stress in Vanadium-Induced Toxicity

Increased generation of reactive oxygen species (ROS) and oxidative stress play a predominant role in vanadium-induced cytotoxicity. For example, vanadate-induced cytotoxicity towards mouse epidermal JB6 cells [61] and monkey epithelial Ma104 cells [62] was demonstrated to be related with increased hydrogen peroxide (H_2O_2) formation. Vanadium-mediated formation of the hydroxyl radical ($\text{OH}\cdot$) by activated human neutrophils was shown by ESR spectroscopy after *in vitro* exposure of these cells to vanadium in the +2, +3, and +4 valence states [63]. In an *in vivo* experimental study, significantly increased levels of hydroxyl radical and superoxide anion ($\text{O}_2^{\cdot-}$) were detected in the cerebellum of sodium metavanadate-treated rats [64]. The vanadium-induced production of ROS occurs as a result of interconversion between V^{4+} species and V^{5+} species by the action of cellular oxidants (e.g., O_2 and H_2O_2) and antioxidants within the cytoplasmic compartment, as described below. For example, the bioreduction of vanadate with NADPH in the presence of NADPH oxidase leads to the formation of vanadyl and superoxide radical. The superoxide is next decomposed by superoxide dismutase (SOD) to less toxic hydrogen peroxide and oxygen. In a Fenton-like reaction, vanadyl can be oxidized with hydrogen peroxide to vanadate with generation of highly reactive hydroxyl radicals [62]. The reaction of vanadate with superoxide anions leads to the formation of peroxovanadyl [$\text{V}(+4)\text{-OO}\cdot$]. This radical can use hydrogen from NADPH and transform to vanadyl hydroperoxide, which in turn can decompose to vanadate and hydrogen peroxide via reaction with hydrogen [65].

Moreover, vanadium can directly affect the mitochondrial inner membrane, which subsequently may impair electron transfer between respiratory complexes causing generation of ROS in mitochondria [66]. ROS are implicated in mediating the deleterious actions of vanadium in cells and tissues through their reactions with cellular lipids, proteins, and nucleic acids. A reaction of lipids with ROS (lipid peroxidation) has a chain character and leads to oxidative degeneration of phospholipids in cell membranes. The two major consequences of lipid peroxidation include changes in membrane biophysical properties (e.g., increased permeability and altered fluidity) and generation of lipid peroxidation end products, many of which are toxic to cells [67]. Various studies indicate that vanadium-induced lipid peroxidation is implicated in toxic effects of vanadium compounds on the liver [68, 69], kidney [70], and brain [71]. In addition, vanadium-mediated oxidative stress manifested by protein oxidation or DNA oxidation has also been observed in vanadium-exposed animals [71] and humans [22], respectively. ROS and oxidative stress have also been reported to contribute to vanadium-induced pulmonary inflammation [72], neurotoxicity [73], and carcinogenic-related effects [74].

5. Dietary Antioxidants in the Prevention of Vanadium Toxicity

It is well known that many edible plants are the main source of natural compounds acting as exogenous antioxidants. Exogenous antioxidants cannot be produced in the body and therefore must be provided through daily nutrition. They reinforce our intrinsic antioxidant system in the protection of the organism against reactive oxygen species-mediated injuries [75]. As shown below in this review, research studies indicate that vanadium toxicity, which is strongly associated with prooxidant mechanisms, can be efficiently reduced or alleviated by dietary and plant-derived antioxidants. The details of the studies on the protective effects of exogenous antioxidants against vanadium adverse actions are presented in Table 1.

5.1. Vitamins C and E. Very early studies already explored the efficiency of vitamin C (ascorbic acid, ascorbate) in the prevention and treatment of vanadium toxicity. For example, a study by Jones and Basinger [76] found that vitamin C was effective against acute vanadate and vanadyl intoxication in mice. Vitamin C, similar to a chelating agent Tiron, was reported to increase urinary excretion of vanadium in mice following acute exposure to vanadyl sulfate [77]. In contrast, other early studies in rats failed to show that vitamin C could influence urinary elimination or tissue concentration of vanadium [78, 79]. More recently, very few studies investigated the interactions of vitamin C and vanadium. In one published study, vitamin C partly enhanced the serum antioxidant status and egg quality in ammonium metavanadate-intoxicated hens [80].

Some studies focused on the role of vitamin E (α -tocopherol) in the treatment of vanadium toxicity. For example, a study by Chandra et al. [81] provided *in vivo* evidence that vitamin E acetate decreased sodium metavanadate-

TABLE 1: Summary of the effects of dietary antioxidants on vanadium toxicity in animal and cell culture models.

Vanadium compounds	Dietary antioxidants	Animal/cell culture model	Effects compared to vanadium-treated animals or cells	Ref.
Vanadium-vitamins				
NH ₄ VO ₃ 5 and 10 mg/kg diet	Vitamin C 50 and 100 mg/kg diet	L hens	↑ Egg quality; ↑ serum SOD activity; ↓ serum LPO	[80]
V ₂ O ₅ 40 mg/kg, ip	Vitamin C (100 mg/kg, ip) or vitamin E (20 mg/kg by gavage)	Hsd:ICR mice (♂)	↓ Micronucleated polychromatic erythrocytes	[83]
NaVO ₃ 0.4 mg V/kg bw, ip	Vitamin E acetate 50 and 100 mg/kg bw, orally	SD rats (♂)	↑ Reproductive organ weight; ↑ sperm number and morphology; ↑ testicular steroidogenic enzyme activities; ↑ serum testosterone, LH and FSH levels; ↑ testicular SOD, CAT activities; ↓ testicular LPO; ↓ testicular histopathological changes	[81]
NaVO ₃ 3 mg/kg bw, ip	Vitamin E 500 mg/kg bw, orally	Nursing albino rats	In pups coexposed to vitamin E and vanadium through lactation: ↑ body weight gain, ↑ brain weight, ↓ reactive astrogliosis, ↑ locomotor and exploratory activity, ↑ hanging latency	[82]
NaVO ₃ 3 mg/kg bw, ip (pubs)	Vitamin E 500 mg, orally (dams)	W rats (dams and pubs)	In vanadium-treated pups exposed to vitamin E through lactation: ameliorated histopathological changes in the testes, lungs, and liver	[126]
Vanadium-polyphenols (flavanols)				
NH ₄ VO ₃ 5, 10, and 15 mg V/kg diet	Tea polyphenols 600 and 1000 mg/kg diet	L hens	↑ Hepatic GST and GPx activities; ↑ production and egg quality	[92]
NH ₄ VO ₃ 10 mg V/kg diet	Tea polyphenols 600 and 1000 mg/kg diet	L hens	↓ Intestinal microflora diversity; ↓ duodenal cell apoptosis; ↑ cecum butyrate acid content	[93]
NH ₄ VO ₃ 5 mg/kg bw, ip	Epigallocatechin gallate 5 mg/kg bw, ip	W rats (♂)	↓ Renal LPO; ↑ renal CAT, SOD, and GPx activities; ↑ serum vitamin E and A levels; ↓ histopathological changes in the kidneys	[95]
NH ₄ VO ₃ 10 mg/kg diet	Epigallocatechin gallate 130 mg/kg diet	L hens	↑ Eggshell color; ↑ protoporphyrin IX content (in eggshell); ↓ uterine LPO; ↑ uterine GST activity; ↑ Nrf2 and HO-1 gene and protein level (in uterus); ↑ phospho-P38 MAPK protein level (in uterus)	[96]
Vanadium-polyphenols (flavonones)				
NaVO ₃ 1 mg/kg bw, ip	Glucosyl hesperidin 25 and 50 mg/kg bw, orally	SD rats (♂)	↑ Reproductive organ weight; ↑ sperm count, motility, and morphology; ↓ sperm DNA fragmentation; ↑ serum testosterone levels; ↓ testicular LPO; ↑ testicular SOD and CAT activities; ↓ testicular histopathological changes	[127]
Vanadium-polyphenols (stilbenes)				
NH ₄ VO ₃ 5 mg/kg bw, ip	Resveratrol 50 mg/kg, orally	SD rats (♂)	↑ Body weight gain; ↓ blood urea nitrogen and creatinine level; ↑ renal SOD activity; no effects of resveratrol on vanadium-induced histopathological changes in the kidneys	[128]

TABLE 1: Continued.

Vanadium compounds	Dietary antioxidants	Animal/cell culture model	Effects compared to vanadium-treated animals or cells	Ref.
Vanadium-phytosterols				
NaVO ₃ 3 mg/kg, ip	Stigmasterol 100 µg, orally	BALB/c mice (♂)	↓ Hippocampal LPO and H ₂ O ₂ levels; ↑ hippocampal SOD and CAT activities; ↑ learning and memory; ↑ locomotor and exploratory activity; ↑ hanging latency; ↓ damage to myelin sheaths, ↑ MBP expression ↑ Learning and memory; ↑ locomotor and exploratory activity; ↑ hanging latency; ↓ brain LPO and H ₂ O ₂ levels; ↓ damage to myelin sheaths; ↑ MBP expression; ↑ SOD, CAT activities and GSH level in the brain	[106]
NaVO ₃ 3 mg/kg, ip	β-Sitosterol 100 µg, orally	BALB/c mice (♂)	↑ SOD, CAT activities and GSH level in the brain	[107]
Vanadium-organosulfur compounds (isothiocyanates)				
VOSO ₄ 34.4 and 68.8 µM	R-sulforaphane 5 µM	HepG2, Caco-2, and Vero cells	↓ ROS; ↓ mitochondrial depolarization; ↑ lysosomal integrity; ↓ DNA damage (comet assay); ↓ cell death	[129]
Vanadium-plant extracts				
NaVO ₃ 100 and 200 µM	<i>Moringa oleifera</i> leaf extract 0.063 mg/well, 0.01 and 0.02 mg/ml	Mouse hippocampal H22 cells	↓ Superoxide levels; ↓ DNA damage (comet assay)	[115]
NaVO ₃ 3 mg/kg, ip	<i>Grewia carpinifolia</i> leaf extract 200 mg/kg, orally	Swiss mice (♂)	↑ Locomotor and exploratory activity; ↑ hanging latency; ↑ motor coordination	[116]
NH ₄ VO ₃ 5 mg/kg bw, ip	Green tea <i>Camellia sinensis</i> decoction 66 g of leaves/l, as a drink	W rats (♂)	↓ LPO in the kidney, liver, and testes; ↑ vitamins E and A in serum; ↓ CAT and SOD activities in erythrocytes	[117]
NH ₄ VO ₃ 60 mg/kg bw (drinking water)	<i>Malva sylvestris</i> decoction 0.2 g of dry plant/kg bw	W rats (♂)	↓ Renal LPO, CAT, SOD, and GPx activities; ↓ histopathological changes in the kidneys	[70]
NH ₄ VO ₃ 5 mg/kg bw, ip	Essential oil of <i>Salvia officinalis</i> 15 mg/kg bw, orally	W rats (♂)	↓ Plasma renal markers (creatinine, urea, uric acid, and LDH); ↓ renal LPO and protein carbonyls; normalized CAT, SOD, and GPx activities in the kidney; ↓ renal histopathological changes	[113]

Abbreviations: bw: body weigh; CAT: catalase; FSH: follicle-stimulating hormone; GPx: glutathione peroxidase; GSH: reduced glutathione; GST: glutathione S-transferase; H₂O₂: hydrogen peroxide; HO-1: heme oxygenase; ip: intraperitoneal; LDH: lactate dehydrogenase; LH: luteinizing hormone; L hen: Lohmann hen; LPO: lipid peroxidation; MAPK: mitogen-activated protein kinases; MBP: myelin basic protein; NaVO₃: sodium metavanadate; NH₄VO₃: ammonium metavanadate; Nrf2: nuclear factor erythroid 2-related factor 2; ROS: reactive oxygen species; SD rat: Sprague Dawley rat; SOD: superoxide dismutase; V: vanadium; V₂O₅: vanadium pentoxide; W rats: Wistar rats; ↑: increased; ↓: decreased.

induced oxidative stress and histopathological changes in the testes of rats. Furthermore, vitamin E was demonstrated to exhibit protective activity against sodium metavanadate-mediated neurotoxicity in rat pups [82]. In this study, vitamin E increased performance in neurobehavioral tests (though not statistically significantly) and decreased reactive astrogliosis in brain tissue of vanadium-treated animals. Both vitamins C and E exhibited protective activity against vanadium pentoxide-induced genotoxicity measured using a micronucleus assay in mouse polychromatic erythrocytes [83].

The antioxidant effect of vitamins E and C is related to their high reactivity as hydrogen (vitamin E) or electron donors (vitamin C) to free radical oxidants, which prevent oxidative damage to cells and tissues, as described below. Vitamin E is a fat-soluble vitamin; in the form of α-tocopherol, it is a major antioxidant located within biological membranes playing a role in protecting from lipid peroxidation. α-Tocopherol breaks the chain reactions of lipid peroxidation through the mechanism of donation of a hydrogen atom from its phenolic hydroxyl group to lipid peroxy radical resulting in the formation of stable lipid hydroperoxide and

unreactive tocopheroxyl radicals [84]. Vitamin C readily donates electrons to oxygen-related radicals (e.g., hydroxyl and peroxy radicals), sulfur radicals, and nitrogen-oxygen radicals [85]. In addition, vitamin C is able to regenerate α -tocopherol by reducing tocopheroxyl radical to its original form (reduced α -tocopherol) [86].

5.2. Polyphenols. Polyphenols (also known as phenolics) are the most numerous and highly diverse group of phytochemicals. They are divided into four subclasses (flavonoids, stilbenes, lignans, and phenolic acids) based on the number of phenolic rings and structural elements that bind these rings to one another. In plants, polyphenols serve a protective role, defending the plant against ultraviolet radiation, cold temperatures, droughts, and incoming pathogens [87, 88]. In humans, polyphenol-rich diet has been associated with the prevention of diseases like certain cancers, cardiovascular diseases, type 2 diabetes, and neurodegenerative disorders [89]. Studies suggest that they can also provide protection against cadmium and lead toxicity [90]. These health-promoting effects of polyphenols are explained by their antioxidant, anti-inflammatory, and metal-chelating actions [88, 91].

As described in Table 1, some very recent studies have shown that polyphenols can be protective against vanadium toxicity in animals. Accordingly, tea polyphenols (a mixture of catechin, epigallocatechin gallate, and caffeine) alleviated vanadium-induced toxic effects on liver antioxidant enzymes [92] and vanadium-mediated epithelial cell apoptosis of the duodenum [93] in hens. Other investigations have focused on a specific polyphenolic compound, namely, epigallocatechin gallate (EGCG). This polyphenol, classified to the group of flavonoids, is the most abundant catechin from green tea infusion with high antioxidant activity [94]. Studies have shown that administration of this phenolic compound to animals protected against oxidative stress and histopathological changes induced by ammonium metavanadate in rat kidneys [95]. Epigallocatechin gallate had also a protective effect on vanadium-induced oxidative stress in the uterus of hens [96]. There are different mechanisms by which polyphenols can exert their antioxidant actions. These include (1) direct radical-scavenging activity by H-atom transfer or by electron transfer from polyphenol to an unstable free radical [97], (2) chelation of metal ions such as iron or copper thereby preventing them from the production of free radicals [98], (3) inhibition of enzymes that can generate radicals, e.g., cytochrome P₄₅₀ isoforms, lipoxygenases, and cyclooxygenases, and (4) synergistic interactive effects of polyphenols with other antioxidants such as phytosterols [99, 100]. However, which of these mechanisms is responsible for the antioxidant effect of phenolics against vanadium-induced toxicity remains to be elucidated.

In addition, polyphenolic compounds (and other phytochemicals) may prove beneficial for the treatment of vanadium toxicity through their ability to activate Nrf2 signaling. Nuclear factor erythroid 2-related factor 2 (Nrf2) is a transcription factor, which upon activation translocates to the nucleus and binds to antioxidant response element (ARE) sequences inducing expression of different cytopro-

protective enzymes. Nrf2-mediated enzymes include antioxidant enzymes such as catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx), and heme oxygenase-1 (HO-1) as well as enzymes responsible for the synthesis and regeneration of glutathione (GSH) such as glutamate cysteine ligase (GCL), glutathione synthetase (GSS), and glutathione reductase (GR) [101]. Thus, activated Nrf2 can significantly increase the antioxidant response in cells to fight oxidative stress associated with vanadium toxicity. Furthermore, vanadium was shown *in vitro* to exert a negative effect on the Nrf2 pathway by inhibiting both the translocation of the Nrf2 factor to the nucleus and the expression of the Nrf2 inducible enzyme NAD(P)H:quinone oxidoreductase 1 (NQO-1) in Hepa 1c1c7 cells [102]. Very recent *in vivo* data provided evidence that dietary vanadium down-regulated Nrf2 and heme oxygenase-1 expression in the uterus of hens and coexposure to epigallocatechin gallate prevented this effect and additionally markedly reduced the vanadium-induced uterine oxidative stress [96]. The mechanisms of Nrf2 activation by epigallocatechin gallate were suggested to involve phosphorylation of Nrf2 serine/threonine residues by protein kinase P38-MAPK, which could upregulate Nrf2 nuclear translocation and subsequent ARE binding [96].

Moreover, some cytoprotective actions of polyphenols may be related to their positive effects on gut microflora. It is suggested that polyphenols through not yet known mechanisms promote beneficial intestinal flora (e.g., bifidobacteria) and inhibit invasive species [89]. Intestinal microbes produce short-chain fatty acids, including butyrate, acetate, and propionate, during fermentation of dietary fiber. These short-chain fatty acids, and butyrate in particular, have been linked with beneficial effects on the metabolism of epithelial cells and preventive effects against colonic cancer [103]. A recent study showed that dietary supplementation of vanadium reduced cecum butyrate acid content and tea polyphenols prevented this reduction in hens [93]. The authors suggested that, by increasing the butyrate content, polyphenols protected duodenal cells from vanadium-induced apoptosis.

5.3. Phytosterols. Phytosterols are a group of steroid compounds present in plant food with the highest amount found in vegetable oils [104]. In plants, their function is to stabilize the phospholipid bilayer of cell membranes. In functional (and structural) terms, they are analogous with cholesterol in humans. Dietary intake of plant sterols by humans has been shown to block both biliary and dietary absorption of cholesterol in the intestine, which helps to lower blood cholesterol levels [105]. In addition, phytosterols have been associated with other beneficial health effects in humans and animals such as reduced risk of heart diseases, anti-inflammatory activities, and prevention of certain cancers [104].

Although limited in number, there are studies evaluating the effects of phytosterols on vanadium-induced toxicity, as described below. The oral administration of stigmasterol and β -sitosterol significantly attenuated neurobehavioral impairments such as deficits in exploration, learning and

memory disabilities, and decreased motor coordination, which were induced by sodium metavanadate in mice [106, 107]. Additionally, in these studies, the two sterols mentioned above exerted an inhibitory effect on hydrogen peroxide generation and lipid peroxidation and improved the activities of superoxide dismutase and catalase in brain tissue of vanadium-exposed mice. Therefore, the authors proposed that the neuroprotective effects of stigmasterol and β -sitosterol could be associated with reduced levels of oxidative stress [106, 107]. The exact mechanism of the antioxidant activity of phytosterols is not well understood. Nevertheless, some *in vitro* studies were undertaken to clarify this issue. For example, it has been found that the extract of the Asian shrub *Aglaia oligophylla*, which contained β -sitosterol and stigmasterol in addition to oligophyllolic acid, possessed high antioxidant activity measured by cupric reducing antioxidant capacity (CUPRAC) and ferric reducing antioxidant power (FRAP) assays [108]. The antioxidant effect of β -sitosterol and stigmasterol observed in this study has been attributed to their ability to donate electrons from their hydroxyl or carboxyl groups directly to the free radical thus neutralizing it [108]. Additionally, another *in vitro* study suggested that β -sitosterol prevented superoxide anion and hydrogen peroxide production by RAW 264.7 macrophages stimulated by phorbol myristate acetate (PMA) due to enhancement of endogenous intracellular antioxidant defenses [109]. In line with this, the study with the same cell culture model as above found that β -sitosterol recovered glutathione (GSH) levels and the GSH/total glutathione ratio and enhanced the activities of antioxidant enzymes, probably through the estrogen receptor/phosphatidylinositol 3-kinase (PI3-kinase) pathway [110]. More recently, *in vivo* research implicated the Nrf2 transcription factor in beneficial effects of β -sitosterol against N-diethylnitrosamine- and ferric nitrilotriacetate-induced nephrotoxicity in rats [111]. So far, however, there are no data available that could indicate which exact mechanism could be involved in the beneficial effects of phytosterols against vanadium intoxication.

5.4. Plant Extracts. Some studies were designed to explore the protective effects of formulations obtained from parts of plants, e.g., leaves or flowers, rather than effects of single isolated components against vanadium-induced toxicity. For example, the results reported that *Malva sylvestris* (prepared as decoction of leaves and flowers) alleviated ammonium metavanadate-induced nephrotoxic effects in rats measured by lipid peroxidation, antioxidant enzyme activities, and histopathological changes [70]. The authors suggested that the antioxidant activities of flavonoids were mostly involved in these beneficial effects. In addition, *M. sylvestris*, particularly its leaves, is also rich in carotenoids and tocopherols and the flowers of this plant exhibited high content of ascorbic acid. All these components are well known for their radical-scavenging activity [112]. The essential oil of the sage *Salvia officinalis* exerted a similar antioxidant effect against ammonium metavanadate-induced renal toxicity [113] as that shown by *M. sylvestris* extract. The sage essential oil contains a diverse group of bioactive phytochemicals with antioxidant,

anti-inflammatory, and nephroprotective properties like camphor, α -pinene, α -thujene, carveol, or α -terpineol, which were assumed to contribute to the protective activity of *S. officinalis* [113, 114]. The leaf extracts of plants such as *Moringa oleifera* [115] and *Grewia carpinifolia* [116] were also reported to have beneficial effects against vanadium-induced neurotoxicity, and green tea (*Camellia sinensis*) [117] was active against vanadium-induced renal, hepatic, and testicular lipid peroxidation in rodents. The protective effect of green tea could be probably attributed to green tea polyphenols, among which catechins (epicatechin, epigallocatechin, epicatechin gallate, and epigallocatechin gallate) are the most abundant tea polyphenols showing many health-promoting benefits including antioxidant (discussed in the earlier section), anti-inflammatory, and antimutagenic effects [118]. In turn, *M. oleifera* is a well-known herbal plant in Africa and Asia, whose leaves contain a diverse mixture of bioactive phytochemicals such as phenolic acids (caffeic acid, gallic acid), flavonoids (e.g., quercetin, kaempferol, and catechins), saponins, tannins, vitamins, and carotenoids. They have been reported to underlie the health protective potential of *Moringa* [119] and may contribute to the alleviation of vanadium-mediated toxicity.

5.5. Other Possible Dietary Supplements against Vanadium. Some mineral components have also been reported to reduce toxicity associated with vanadium exposure. For example, the essential metal zinc was found to reduce the vanadium-induced DNA damage (comet assay) in melanocytes *in vitro* [120]. Oral supplementation of zinc sulfate prevented lipid peroxidation and normalized the activities of antioxidant enzymes in the testis of sodium metavanadate-exposed rats [121]. As shown by the *in vitro* study of Bay et al. [122], zinc has the ability to reduce the levels of hydroxyl free radicals generated by vanadium in a Fenton-like reaction and prevent a decline in glutathione levels, which may constitute mechanisms through which zinc exhibits its protective effects against vanadium injury. Another element, i.e., magnesium, has shown a protective potential against vanadium as well. As reviewed by Ścibior [40], when given orally during vanadium intoxication in rats, magnesium caused the following effects: it reduced the level of vanadium in erythrocytes and whole blood, limited accumulation of vanadium in kidney and the cerebral hemisphere, decreased lipid peroxidation in the liver, and prevented the decrease in glutathione transferase activity in erythrocytes. Lastly, selenium either alone or in conjunction with a chelating agent Tiron exhibited protective effects against vanadium-induced injury in rats [38, 123]. However, converse results were obtained in mammalian cells *in vitro* (CHO-K1 cells) where vanadyl-induced cytotoxicity was increased in the presence of low doses of selenite (0.5 and 1 μ M) [39]. The contradictory results on vanadium and selenium interactions appear to confirm the well-known “two-faced” character of selenium, most probably caused by a very narrow range in selenium concentrations between necessity and toxicity. This certainly raises concerns over the use of selenium (especially as selenite) as a potential antidote against vanadium.

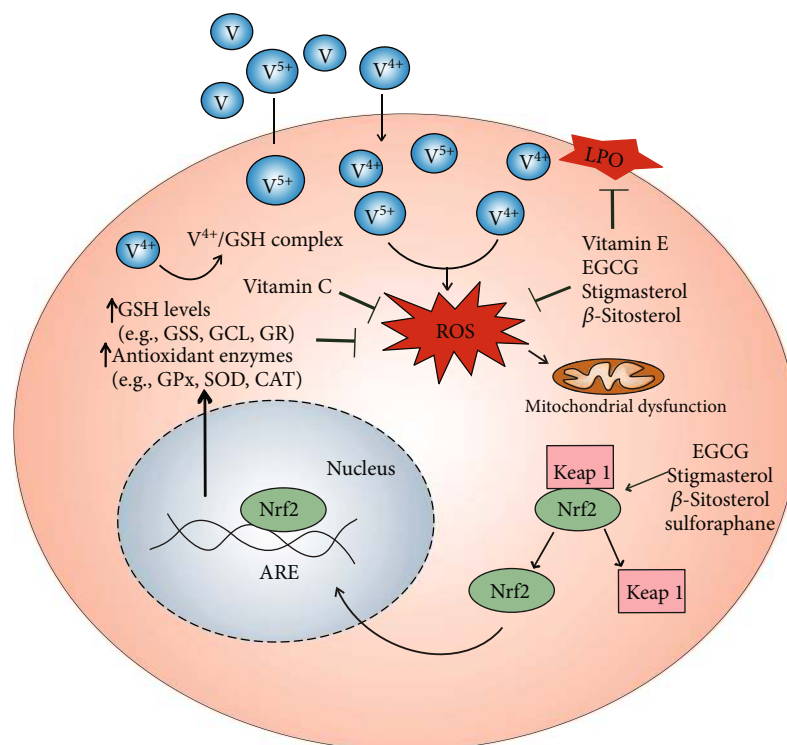


FIGURE 2: Scheme showing the putative cellular sites of action targeted by some dietary antioxidants during vanadium intoxication. Whether polyphenols (e.g., EGCG) and phytosterols (e.g., stigmasterol and β -sitosterol) act as direct antioxidants (through their scavenging activity) or/and indirect antioxidants (e.g., by inducing Nrf2 binding to ARE) remains to be confirmed. Abbreviations: ARE: antioxidant response element; CAT: catalase; EGCG: epigallocatechin gallate; GCL: glutamate cysteine ligase; GPx: glutathione peroxidase; GR: glutathione reductase; GSS: glutathione synthetase; Keap1: Kelch-like ECH-associated protein 1; LPO: lipid peroxidation; Nrf2: nuclear factor (erythroid-derived 2)-like 2; ROS: reactive oxygen species; SOD: superoxide dismutase; V⁵⁺: pentavalent vanadium; V⁴⁺: tetravalent vanadium; —|: inhibition; —>: stimulation.

6. Conclusions

This review provides an updated overview of the role of different dietary-derived antioxidants in vanadium-induced toxicity. In general, the studies show the therapeutic protective effects of vitamins C and E, tea polyphenols, phytosterols, and some plant extracts against vanadium. As expected, the beneficial action of these natural compounds is based on their ability to reduce vanadium-mediated oxidative stress (some possible cellular sites of protection are summarized in Figure 2). The mechanism of action of vitamins C and E probably includes direct removal of reactive oxygen species from the intracellular compartment (by hydrophilic vitamin C) and within the cell membrane (by lipid-soluble vitamin E). As mentioned in this review, polyphenols and phytosterols have also the ability to neutralize reactive oxygen species directly via donating electrons or hydrogen atoms from their hydroxyl (or carboxyl) groups. However, some authors suggest that polyphenols perform their antioxidant activity through other mechanisms, e.g., induction of antioxidant and phase II enzymes, which are indirect modes of antioxidant activity rather than acting through direct reactive oxygen species scavenging mechanisms. The reason for that would be the lower reduction potential and bioavailability of polyphenols compared to endogenous antioxidants

[124]. So far, only one study has assessed the indirect antioxidant mechanism of polyphenols against vanadium showing increased Nrf2 and HO-1 expression as a protective mode of epigallocatechin gallate action in vivo [96]. Therefore, the mechanisms of action of exogenous antioxidants in prevention of vanadium toxicity remain to be further clarified. In addition, beneficial effects of extracts from medicinal plants such as *Moringa oleifera* or *Malva sylvestris* against vanadium have also received attention. The major advantage of plant extracts is their content of a mixture of different phytochemicals and nutrients which, via synergistic/additive interactions, are suggested to be more health effective than isolated phytochemicals [75, 125].

In conclusion, although the investigations cited in this review show that supplementation with dietary antioxidants has beneficial effects on vanadium poisoning, further studies have to be conducted to draw more definitive statements. The following points have been identified as topics for future research:

- (1) Still, more studies are needed on the role of vitamins C and E in the toxicology of vanadium. The relatively low cost and wide therapeutic window (especially for vitamin C) of these nutrients make them attractive antidotes against vanadium poisoning

- (2) The precise mechanism of the activity of polyphenols and phytoosterols against vanadium should be explored
- (3) Therapy with plant extracts containing a mixture of different phytochemicals could be an interesting alternative to single compound treatment

Conflicts of Interest

The author declares that she have no conflicts of interest.

Acknowledgments

This work was supported by statutory funds of the John Paul II Catholic University of Lublin, Poland.

References

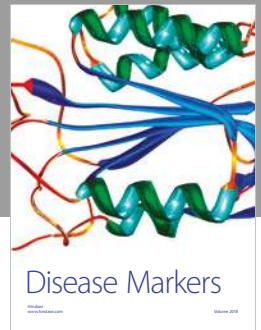
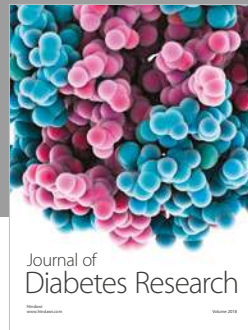
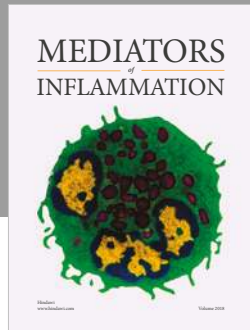
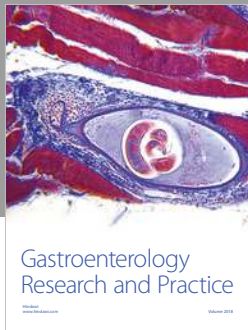
- [1] B. Sharma, S. Singh, and N. J. Siddiqi, "Biomedical implications of heavy metals induced imbalances in redox systems," *BioMed Research International*, vol. 2014, Article ID 640754, 26 pages, 2014.
- [2] D. J. Betteridge, "What is oxidative stress," *Metabolism*, vol. 49, 2 Supplement 1, pp. 3–8, 2000.
- [3] S. J. S. Flora and V. Pachauri, "Chelation in metal intoxication," *International Journal of Environmental Research and Public Health*, vol. 7, no. 7, pp. 2745–2788, 2010.
- [4] M. E. Ferrero, "Rationale for the successful management of EDTA chelation therapy in human burden by toxic metals," *BioMed Research International*, vol. 2016, Article ID 8274504, 13 pages, 2016.
- [5] J. Chen, L. Du, J. Li, and H. Song, "Epigallocatechin-3-gallate attenuates cadmium-induced chronic renal injury and fibrosis," *Food and Chemical Toxicology*, vol. 96, pp. 70–78, 2016.
- [6] A. J. Akinyemi, N. Onyebueke, O. A. Faboya, S. A. Onikanni, A. Fadaka, and I. Olayide, "Curcumin inhibits adenosine deaminase and arginase activities in cadmium-induced renal toxicity in rat kidney," *Journal of Food and Drug Analysis*, vol. 25, no. 2, pp. 438–446, 2017.
- [7] A. A. Alshatwi, T. N. Hasan, A. M. Alqahtani et al., "Delineating the anti-cytotoxic and anti-genotoxic potentials of catechin hydrate against cadmium toxicity in human peripheral blood lymphocytes," *Environmental Toxicology and Pharmacology*, vol. 38, no. 2, pp. 653–662, 2014.
- [8] N. A. O. Alkharashi, V. S. Periasamy, J. Athinarayanan, and A. A. Alshatwi, "Sulforaphane alleviates cadmium-induced toxicity in human mesenchymal stem cells through *POR* and *TNFSF10* genes expression," *Biomedicine & Pharmacotherapy*, vol. 115, article 108896, 2019.
- [9] X. D. Han, Y. Y. Zhang, K. L. Wang, Y. P. Huang, Z. B. Yang, and Z. Liu, "The involvement of Nrf2 in the protective effects of (-)-epigallocatechin-3-gallate (EGCG) on NaAsO₂-induced hepatotoxicity," *Oncotarget*, vol. 8, no. 39, pp. 65302–65312, 2017.
- [10] Q. Zhai, L. Yang, J. Zhao, H. Zhang, F. Tian, and W. Chen, "Protective effects of dietary supplements containing probiotics, micronutrients, and plant extracts against lead toxicity in mice," *Frontiers in Microbiology*, vol. 9, article 2134, 2018.
- [11] S. Kianoush, M. Balali-Mood, S. R. Mousavi et al., "Comparison of therapeutic effects of garlic and d-penicillamine in patients with chronic occupational lead poisoning," *Basic & Clinical Pharmacology & Toxicology*, vol. 110, no. 5, pp. 476–481, 2012.
- [12] J. Yang, Y. Teng, J. Wu et al., "Current status and associated human health risk of vanadium in soil in China," *Chemosphere*, vol. 171, pp. 635–643, 2017.
- [13] WHO, "Vanadium," in *WHO air quality guidelines for Europe*, vol. 91 of European Series, World Health Organisation, WHO Regional Publications, 2nd edition, 2000.
- [14] IARC, International Agency for Research on Cancer, "IARC Monographs on the evaluation of carcinogenic risks to humans," in *Cobalt in hard metals and cobalt sulfate, gallium arsenide, indium phosphide and vanadium pentoxide*, vol. 86, International Agency for Research on Cancer, Lyon, France, 2006.
- [15] C. Doetsch and J. Burfeind, "Vanadium redox flow batteries," in *Storing energy. With special reference to renewable energy surces*, T. M. Letcher, Ed., Elsevier, Oxford, 2016, Chapter 12.
- [16] EPA, *Fact Sheet: the drinking water contaminant candidate list – the source of priority contaminants for the drinking water program*, United States Environmental Protection Agency. EPA 815-F-05-001. Office of Water, US Environmental Protection Agency, Washington, DC, 2005.
- [17] X. Y. Xiao, M. Yang, Z. H. Guo, Z. C. Jiang, Y. N. Liu, and X. Cao, "Soil vanadium pollution and microbial response characteristics from stone coal smelting district," *Transactions of the Nonferrous Metals Society of China*, vol. 25, no. 4, pp. 1271–1278, 2015.
- [18] M. Imtiaz, M. S. Rizwan, S. Xiong et al., "Vanadium, recent advancements and research prospects: a review," *Environment International*, vol. 80, pp. 79–88, 2015.
- [19] M. Khodeir, M. Shamy, M. Alghamdi et al., "Source apportionment and elemental composition of PM_{2.5} and PM₁₀ in Jeddah City, Saudi Arabia," *Atmospheric Pollution Research*, vol. 3, no. 3, pp. 331–340, 2012.
- [20] G. Arena, C. Copat, A. Dimartino et al., "Determination of total vanadium and vanadium(V) in groundwater from Mt. Etna and estimate of daily intake of vanadium(V) through drinking water," *Journal of Water and Health*, vol. 13, no. 2, pp. 522–530, 2015.
- [21] M. A. Woodin, Y. Liu, D. Neuberg, R. Hauser, T. J. Smith, and D. C. Christiani, "Acute respiratory symptoms in workers exposed to vanadium-rich fuel-oil ash," *American Journal of Industrial Medicine*, vol. 37, no. 4, pp. 353–363, 2000.
- [22] V. A. Ehrlich, A. K. Nersesyan, K. Atefie et al., "Inhalative exposure to vanadium pentoxide causes DNA damage in workers: results of a multiple end point study," *Environmental Health Perspectives*, vol. 116, no. 12, pp. 1689–1693, 2008.
- [23] H. Li, D. Zhou, Q. Zhang et al., "Vanadium exposure-induced neurobehavioral alterations among Chinese workers," *Neurotoxicology*, vol. 36, pp. 49–54, 2013.
- [24] M. Patel, L. Hoepner, R. Garfinkel et al., "Ambient metals, elemental carbon and wheeze and cough in New York City children through 24 months of age," *American Journal of Respiratory and Critical Care Medicine*, vol. 180, no. 11, pp. 1107–1113, 2009.
- [25] M. L. Bell, K. Ebisu, R. D. Peng, J. M. Samet, and F. Dominici, "Hospital admissions and chemical composition of fine particle air pollution," *American Journal of Respiratory and Critical Care Medicine*, vol. 179, no. 12, pp. 1115–1120, 2009.
- [26] J. Hu, Y. Peng, T. Zheng et al., "Effects of trimester-specific exposure to vanadium on ultrasound measures of fetal

- growth and birth size: a longitudinal prospective prenatal cohort study,” *Lancet Planet Health*, vol. 2, no. 10, pp. e427–e437, 2018.
- [27] J. Hu, W. Xia, X. Pan et al., “Association of adverse birth outcomes with prenatal exposure to vanadium: a population-based cohort study,” *Lancet Planet Health*, vol. 1, no. 6, pp. e230–e241, 2017.
- [28] P. Malandrino, M. Russo, A. Ronchi et al., “Increased thyroid cancer incidence in a basaltic volcanic area is associated with non-anthropogenic pollution and biocontamination,” *Endocrine*, vol. 53, no. 2, pp. 471–479, 2016.
- [29] B. Boulassel, N. Sadeg, O. Roussel, M. Perrin, and H. Belhadj-Tahar, “Fatal poisoning by vanadium,” *Forensic Science International*, vol. 206, no. 1–3, pp. e79–e81, 2011.
- [30] A. Montiel-Dávalos, A. Gonzalez-Villava, V. Rodriguez-Lara, L. F. Montaño, T. I. Fortoul, and R. López-Marure, “Vanadium pentoxide induces activation and death of endothelial cells,” *Journal of Applied Toxicology*, vol. 32, no. 1, pp. 26–33, 2012.
- [31] P. Boscolo, M. Carmignani, A. R. Volpe et al., “Renal toxicity and arterial hypertension in rats chronically exposed to vanadate,” *Occupational and Environmental Medicine*, vol. 51, no. 7, pp. 500–503, 1994.
- [32] G. Pinon-Zarate, V. Rodriguez-Lara, M. Rojas-Lemus et al., “Vanadium pentoxide inhalation provokes germinal center hyperplasia and suppressed humoral immune responses,” *Journal of Immunotoxicology*, vol. 5, no. 2, pp. 115–122, 2008.
- [33] M. Ustarroz-Cano, I. Garcia-Pelaez, S. Cervantes-Yepez, N. Lopez-Valdez, and T. I. Fortoul, “Thymic cytoarchitecture changes in mice exposed to vanadium,” *Journal of Immunotoxicology*, vol. 14, no. 1, pp. 9–14, 2017.
- [34] O. R. Folarin, A. M. Snyder, D. G. Peters, F. Olopade, J. R. Connor, and J. O. Olopade, “Brain metal distribution and neuro-inflammatory profiles after chronic vanadium administration and withdrawal in mice,” *Front. Neuroanat*, vol. 11, p. 58, 2017.
- [35] O. Folarin, F. Olopade, S. Onwuka, and J. Olopade, “Memory Deficit Recovery after Chronic Vanadium Exposure in Mice,” *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 4860582, 7 pages, 2016.
- [36] J. L. Domingo, “Vanadium: a review of the reproductive and developmental toxicity,” *Reproductive Toxicology*, vol. 10, no. 3, pp. 175–182, 1996.
- [37] E. A. Rondini, D. M. Walters, and A. K. Bauer, “Vanadium pentoxide induces pulmonary inflammation and tumor promotion in a strain-dependent manner,” *Particle and Fibre Toxicology*, vol. 7, no. 1, p. 9, 2010.
- [38] S. S. Haider, A. A. Abdel-Gayoum, M. El-Fakhri, and K. M. Ghwarsha, “Effect of selenium on vanadium toxicity in different regions of rat brain,” *Human & Experimental Toxicology*, vol. 17, no. 1, pp. 23–28, 1998.
- [39] I. Zwolak, “Increased Cytotoxicity of vanadium to CHO-K1 cells in the presence of inorganic selenium,” *Bulletin of Environmental Contamination and Toxicology*, vol. 95, no. 5, pp. 593–598, 2015.
- [40] A. Ścibior, “Vanadium (V) and magnesium (Mg) – in vivo interactions: a review,” *Chemico-Biological Interactions*, vol. 258, pp. 214–233, 2016.
- [41] D. Rehder, “The future of/for vanadium,” *Dalton Transactions*, vol. 42, no. 33, pp. 11749–11761, 2013.
- [42] T. Scior, J. Guevara-Garcia, Q. T. Do, P. Bernard, and S. Laufer, “Why Antidiabetic Vanadium Complexes are Not in the Pipeline of “Big Pharma” Drug Research? A Critical Review,” *Current Medicinal Chemistry*, vol. 23, no. 25, pp. 2874–2891, 2016.
- [43] J. L. Domingo and M. Gomez, “Vanadium compounds for the treatment of human diabetes mellitus: a scientific curiosity? A review of thirty years of research,” *Food and Chemical Toxicology*, vol. 95, pp. 137–141, 2016.
- [44] I. E. León, J. F. Cadavid-Vargas, I. Tiscornia et al., “Oxidovanadium(IV) complexes with chrysin and silibinin: anticancer activity and mechanisms of action in a human colon adenocarcinoma model,” *JBIC Journal of Biological Inorganic Chemistry*, vol. 20, no. 7, article 1298, pp. 1175–1191, 2015.
- [45] I. E. Leon, J. F. Cadavid-Vargas, A. Resasco et al., “In vitro and in vivo antitumor effects of the VO-chrysin complex on a new three-dimensional osteosarcoma spheroids model and a xenograft tumor in mice,” *Journal of Biological Inorganic Chemistry*, vol. 21, no. 8, pp. 1009–1020, 2016.
- [46] A. B. Hodgson, R. Baskerville, L. M. Burke, S. J. Stear, and L. M. Castell, “A–Z of nutritional supplements: dietary supplements, sports nutrition foods and ergogenic aids for health and performance: part 42,” *British Journal of Sports Medicine*, vol. 47, no. 4, pp. 247–248, 2013.
- [47] U. Repinc and L. Benedik, “Determination of vanadium in dietary supplements,” *Acta Chimica Slovenica*, vol. 51, pp. 59–65, 2004.
- [48] A. Polishetty, G. Littlefair, and K. Praveen Kumar, “Machinability assessment of titanium alloy Ti-6Al-4V for biomedical applications,” *Advanced Materials Research*, vol. 941–944, pp. 1985–1990, 2014.
- [49] C. Oldani and A. Dominguez, *Titanium as a biomaterial for implants, recent advances in arthroplasty*, S. Fokter, Ed., InTech, 2012, <http://www.intechopen.com/books/recent-advances-in-arthroplasty/titanium-as-a-biomaterial-for-implants>.
- [50] S. Engelhart and R. J. Segal, “Allergic reaction to vanadium causes a diffuse eczematous eruption and titanium alloy orthopedic implant failure,” *Cutis*, vol. 99, no. 4, pp. 245–249, 2017.
- [51] D. Sanna, G. Micera, and E. Garribba, “On the transport of vanadium in blood serum,” *Inorganic Chemistry*, vol. 48, no. 13, pp. 5747–5757, 2009.
- [52] S. Trevino, A. Díaz, E. Sánchez-Lara, B. L. Sanchez-Gaytan, J. M. Perez-Aguilar, and E. González-Vergara, “Vanadium in biological action: chemical, pharmacological aspects, and metabolic implications in diabetes mellitus,” *Biological Trace Element Research*, vol. 188, no. 1, pp. 68–98, 2019.
- [53] D. Rehder, “The role of vanadium in biology,” *Metallomics*, vol. 7, no. 5, pp. 730–742, 2015.
- [54] J. Korbecki, I. Baranowska-Bosiacka, I. Gutowska, and D. Chlubek, “Biochemical and medical importance of vanadium compounds,” *Acta Biochimica Polonica*, vol. 59, no. 2, pp. 195–200, 2012.
- [55] D. Rehder, “The potentiality of vanadium in medicinal applications,” *Future Medicinal Chemistry*, vol. 4, no. 14, pp. 1823–1837, 2012.
- [56] E. J. Baran, “Vanadium detoxification: chemical and biochemical aspects,” *Chemistry & Biodiversity*, vol. 5, no. 8, pp. 1475–1484, 2008.
- [57] B. Gummow, C. J. Botha, and M. C. Williams, “Chronic vanadium poisoning in calves and its treatment with calcium

- disodium ethylenediaminetetraacetate," *Veterinary Research Communications*, vol. 30, no. 7, pp. 807–822, 2006.
- [58] D. J. Sanchez, M. T. Colomina, J. L. Domingo, and J. Corbella, "Prevention by sodium 4,5-dihydroxybenzene-1,3-disulfonate (Tiron) of vanadium-induced behavioral toxicity in rats," *Biological Trace Element Research*, vol. 69, no. 3, pp. 249–259, 1999.
- [59] E. G. Ferrer and E. J. Baran, "Reduction of vanadium(V) with ascorbic acid and isolation of the generated oxovanadium(IV) species," *Biological Trace Element Research*, vol. 83, no. 2, pp. 111–119, 2001.
- [60] I. Zwolak and D. Gołębiowska, "Protective activity of pyruvate against vanadium-dependent cytotoxicity in Chinese hamster ovary (CHO-K1) cells," *Toxicology and Industrial Health*, vol. 34, no. 5, pp. 283–292, 2018.
- [61] C. Huang, Z. Zhang, M. Ding et al., "Vanadate induces p53 transactivation through hydrogen peroxide and causes apoptosis," *The Journal of Biological Chemistry*, vol. 275, no. 42, pp. 32516–32522, 2000.
- [62] L. S. Capella, M. R. Gefé, E. F. Silva et al., "Mechanisms of vanadate-induced cellular toxicity: role of cellular glutathione and NADPH," *Archives of Biochemistry and Biophysics*, vol. 406, no. 1, pp. 65–72, 2002.
- [63] H. Fickl, A. J. Theron, H. Grimmer et al., "Vanadium promotes hydroxyl radical formation by activated human neutrophils," *Free Radical Biology & Medicine*, vol. 40, no. 1, pp. 146–155, 2006.
- [64] S. Cuesta, D. Francés, and G. B. García, "ROS formation and antioxidant status in brain areas of rats exposed to sodium metavanadate," *Neurotoxicology and Teratology*, vol. 33, no. 2, pp. 297–302, 2011.
- [65] A. M. Evangelou, "Vanadium in cancer treatment," *Critical Reviews in Oncology/Hematology*, vol. 42, no. 3, pp. 249–265, 2002.
- [66] Y. Zhao, L. Ye, H. Liu et al., "Vanadium compounds induced mitochondria permeability transition pore (PTP) opening related to oxidative stress," *Journal of Inorganic Biochemistry*, vol. 104, no. 4, pp. 371–378, 2010.
- [67] A. Catalá, "Lipid peroxidation of membrane phospholipids generates hydroxy-alkenals and oxidized phospholipids active in physiological and/or pathological conditions," *Chemistry and Physics of Lipids*, vol. 157, no. 1, pp. 1–11, 2009.
- [68] M. Samira, T. Mounira, K. Kamel et al., "Hepatotoxicity of vanadyl sulfate in nondiabetic and streptozotocin-induced diabetic rats," *Canadian Journal of Physiology and Pharmacology*, vol. 96, no. 11, pp. 1076–1083, 2018.
- [69] A. Ścibior, D. Gołębiowska, and I. Niedźwiecka, "Magnesium can protect against vanadium-induced lipid peroxidation in the hepatic tissue," *Oxidative Medicine and Cellular Longevity*, vol. 2013, Article ID 802734, 11 pages, 2013.
- [70] W. Marouane, A. Soussi, J. C. Murat, S. Bezzine, and A. el Feki, "The protective effect of *Malva sylvestris* on rat kidney damaged by vanadium," *Lipids in Health and Disease*, vol. 10, no. 1, p. 65, 2011.
- [71] I. L. Usende, J. O. Olopade, B. O. Emikpe, A. A. Oyagbemi, and A. A. Adedapo, "Oxidative stress changes observed in selected organs of African giant rats (*Cricetomys gambianus*) exposed to sodium metavanadate," *International Journal of Veterinary Science and Medicine*, vol. 6, no. 1, pp. 80–89, 2019.
- [72] L. Wang, D. Medan, R. Mercer et al., "Vanadium-induced apoptosis and pulmonary inflammation in mice: role of reactive oxygen species," *Journal of Cellular Physiology*, vol. 195, no. 1, pp. 99–107, 2003.
- [73] O. I. Fatola, F. A. Olaolorun, F. E. Olopade, and J. O. Olopade, "Trends in vanadium neurotoxicity," *Brain Research Bulletin*, vol. 145, pp. 75–80, 2019.
- [74] I. Zwolak, "Vanadium carcinogenic, immunotoxic and neurotoxic effects: a review of in vitro studies," *Toxicology Mechanisms and Methods*, vol. 24, no. 1, pp. 1–12, 2014.
- [75] J. Bouayed and T. Bohn, "Exogenous Antioxidants—Double-Edged Swords in Cellular Redox State: Health Beneficial Effects at Physiologic Doses versus Deleterious Effects at High Doses," *Oxidative Medicine and Cellular Longevity*, vol. 3, no. 4, 237 pages, 2010.
- [76] M. M. Jones and M. A. Basinger, "Chelate antidotes for sodium vanadate and vanadyl sulfate intoxication in mice," *Journal of Toxicology and Environmental Health*, vol. 12, no. 4–6, pp. 749–756, 1983.
- [77] J. L. Domingo, M. Gomez, J. M. Llobet, and J. Corbella, "Chelating agents in the treatment of acute vanadyl sulphate intoxication in mice," *Toxicology*, vol. 62, no. 2, pp. 203–211, 1990.
- [78] M. Gómez, J. L. Domingo, J. M. Llobet, and J. Corbella, "Effectiveness of some chelating agents on distribution and excretion of vanadium in rats after prolonged oral administration," *Journal of Applied Toxicology*, vol. 11, no. 3, pp. 195–198, 1991.
- [79] M. Gómez, J. L. Domingo, J. M. Llobet, and J. Corbella, "Evaluation of the efficacy of various chelating agents on urinary excretion and tissue distribution of vanadium in rats," *Toxicology Letters*, vol. 57, no. 2, pp. 227–234, 1991.
- [80] J. P. Wang, K. R. He, X. M. Ding et al., "Effect of dietary vanadium and vitamin C on egg quality and antioxidant status in laying hens," *Journal of Animal Physiology and Animal Nutrition*, vol. 100, no. 3, pp. 440–447, 2016.
- [81] A. K. Chandra, R. Ghosh, A. Chatterjee, and M. Sarkar, "Amelioration of vanadium-induced testicular toxicity and adrenocortical hyperactivity by vitamin E acetate in rats," *Molecular and Cellular Biochemistry*, vol. 306, no. 1–2, pp. 189–200, 2007.
- [82] J. O. Olopade, I. O. Fatola, and F. E. Olopade, "Vertical administration of vanadium through lactation induces behavioural and neuromorphological changes: protective role of vitamin E," *Nigerian Journal of Physiological Sciences*, vol. 26, no. 1, pp. 055–060, 2011.
- [83] M. del Carmen García-Rodríguez, L. M. Hernández-Cortés, and M. A. Altamirano-Lozano, "In Vivo Effects of Vanadium Pentoxide and Antioxidants (Ascorbic Acid and Alpha-Tocopherol) on Apoptotic, Cytotoxic, and Genotoxic Damage in Peripheral Blood of Mice," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 6797851, 11 pages, 2016.
- [84] K. Hajibabaei, "Antioxidant properties of vitamin E," *Annals of Research in Antioxidants*, vol. 1, article e22, 2016.
- [85] S. J. Padayatty, A. Katz, Y. Wang et al., "Vitamin C as an antioxidant: evaluation of its role in disease prevention," *Journal of the American College of Nutrition*, vol. 22, no. 1, pp. 18–35, 2003.
- [86] M. Valko, M. Izakovic, M. Mazur, C. J. Rhodes, and J. Telser, "Role of oxygen radicals in DNA damage and cancer

- incidence," *Molecular and Cellular Biochemistry*, vol. 266, no. 1/2, pp. 37–56, 2004.
- [87] R. Tsao, "Chemistry and biochemistry of dietary polyphenols," *Nutrients*, vol. 2, no. 12, pp. 1231–1246, 2010.
- [88] A. Tresserra-Rimbau, R. M. Lamuela-Raventos, and J. J. Moreno, "Polyphenols, food and pharma. Current knowledge and directions for future research," *Biochemical Pharmacology*, vol. 156, pp. 186–195, 2018.
- [89] H. Cory, S. Passarelli, J. Szeto, M. Tamez, and J. Mattei, "The role of polyphenols in human health and food systems: a mini-review," *Frontiers in Nutrition*, vol. 5, p. 87, 2018.
- [90] Q. Zhai, A. Narbad, and W. Chen, "Dietary strategies for the treatment of cadmium and lead toxicity," *Nutrients*, vol. 7, no. 1, pp. 552–571, 2015.
- [91] M. T. Mitjavila and J. J. Moreno, "The effects of polyphenols on oxidative stress and the arachidonic acid cascade. Implications for the prevention/treatment of high prevalence diseases," *Biochemical Pharmacology*, vol. 84, no. 9, pp. 1113–1122, 2012.
- [92] Z. H. Yuan, K. Y. Zhang, X. M. Ding et al., "Effect of tea polyphenols on production performance, egg quality, and hepatic antioxidant status of laying hens in vanadium-containing diets," *Poultry Science*, vol. 95, no. 7, pp. 1709–1717, 2016.
- [93] Z. H. Yuan, J. P. Wang, K. Y. Zhang et al., "Effect of vanadium and tea polyphenols on intestinal morphology, microflora and short-chain fatty acid profile of laying hens," *Biological Trace Element Research*, vol. 174, no. 2, pp. 419–427, 2016.
- [94] S. Legeay, M. Rodier, L. Fillon, S. Faure, and N. Clere, "Epigallocatechin gallate: a review of its beneficial properties to prevent metabolic syndrome," *Nutrients*, vol. 7, no. 7, pp. 5443–5468, 2015.
- [95] A. Soussi, R. Abdennabi, F. Ghorbel, J. C. Murat, and A. F. El Feki, "Ameliorated effects of (-)-epigallocatechin gallate against toxicity induced by vanadium in the kidneys of Wistar rats," *Biological Trace Element Research*, vol. 180, no. 2, pp. 239–245, 2017.
- [96] J. Wang, Z. Yuan, K. Zhang et al., "Epigallocatechin-3-gallate protected vanadium-induced eggshell depigmentation via P38MAPK-Nrf2/HO-1 signaling pathway in laying hens," *Poultry Science*, vol. 97, no. 9, pp. 3109–3118, 2018.
- [97] M. Leopoldini, T. Marino, N. Russo, and M. Toscano, "Antioxidant properties of phenolic compounds: H-atom versus electron transfer mechanism," *The Journal of Physical Chemistry A*, vol. 108, no. 22, pp. 4916–4922, 2004.
- [98] M. Szymonowicz and M. Kolanek, "Flavonoids and their properties to form chelate complexes," *Biotechnology and Food Science*, vol. 76, pp. 35–41, 2012.
- [99] M. Vivancos and J. J. Moreno, "Effect of resveratrol, tyrosol and beta-sitosterol on oxidised low-density lipoprotein-stimulated oxidative stress, arachidonic acid release and prostaglandin E2 synthesis by RAW 264.7 macrophages," *The British Journal of Nutrition*, vol. 99, no. 6, pp. 1199–1207, 2008.
- [100] C. E. Storniolo, N. Martínez-Hovelman, M. Martínez-Huélamo, R. M. Lamuela-Raventos, and J. J. Moreno, "Extra virgin olive oil minor compounds modulate mitogenic action of oleic acid on colon cancer cell line," *Journal of Agricultural and Food Chemistry*, vol. 67, no. 41, pp. 11420–11427, 2019.
- [101] A. L. Stefanson and M. Bakovic, "Dietary regulation of Keap1/Nrf2/ARE pathway: focus on plant-derived compounds and trace minerals," *Nutrients*, vol. 6, no. 9, pp. 3777–3801, 2014.
- [102] A. Anwar-Mohamed and A. O. El-Kadi, "Down-regulation of the detoxifying enzyme NAD(P)H:quinone oxidoreductase 1 by vanadium in Hepa 1c1c7 cells," *Toxicology and Applied Pharmacology*, vol. 236, no. 3, pp. 261–269, 2009.
- [103] S. M. McNabney and T. M. Henagan, "Short chain fatty acids in the colon and peripheral tissues: a focus on butyrate, colon cancer, obesity and insulin resistance," *Nutrients*, vol. 9, no. 12, article 1348, 2017.
- [104] R. J. Ogbe, D. O. Ochalefu, S. G. Mafulul, and O. B. Olaniru, "A review on dietary phytosterols: their occurrence, metabolism and health benefits," *Asian Journal of Plant Science and Research*, vol. 5, pp. 10–21, 2015.
- [105] Ghafoorunissa, "Impact of quality of dietary fat on serum cholesterol and coronary heart disease: focus on plant sterols and other non-glyceride components," *National Medical Journal of India*, vol. 22, no. 3, pp. 126–132, 2009.
- [106] O. E. Adebisi, J. O. Olopade, and F. O. Olayemi, "Sodium metavanadate induced cognitive decline, behavioral impairments, oxidative stress and down regulation of myelin basic protein in mice hippocampus: ameliorative roles of β -sitosterol, and stigmasterol," *Brain and Behavior*, vol. 8, no. 7, article e01014, 2018.
- [107] O. E. Adebisi, F. O. Olayemi, J. O. Olopade, and N. H. Tan, "Beta-sitosterol enhances motor coordination, attenuates memory loss and demyelination in a vanadium-induced model of experimental neurotoxicity," *Pathophysiology*, vol. 26, no. 1, pp. 21–29, 2019.
- [108] Y. Y. S. Yu, N. K. Kassim, K. H. Musa, and A. Abdullah, "Measurement of antioxidant activity and structural elucidation of chemical constituents from *Aglaia oligophylla* Miq," *International Proceedings of Chemical, Biological and Environmental Engineering*, vol. 95, pp. 1–7, 2016.
- [109] J. J. Moreno, "Effect of olive oil minor components on oxidative stress and arachidonic acid mobilization and metabolism by macrophages RAW 264.7," *Free Radical Biology and Medicine*, vol. 35, no. 9, pp. 1073–1081, 2003.
- [110] M. Vivancos and J. J. Moreno, " β -Sitosterol modulates antioxidant enzyme response in RAW 264.7 macrophages," *Free Radical Biology and Medicine*, vol. 39, no. 1, pp. 91–97, 2005.
- [111] R. Sharmila, G. Sindhu, and P. M. Arockianathan, "Nephroprotective effect of β -sitosterol on N-diethylnitrosamine initiated and ferric nitrilotriacetate promoted acute nephrotoxicity in Wistar rats," *Journal of Basic and Clinical Physiology and Pharmacology*, vol. 27, no. 5, pp. 473–482, 2016.
- [112] L. Barros, A. M. Carvalho, and I. C. Ferreira, "Leaves, flowers, immature fruits and leafy flowered stems of *Malva sylvestris*: a comparative study of the nutraceutical potential and composition," *Food and Chemical Toxicology*, vol. 48, no. 6, pp. 1466–1472, 2010.
- [113] F. G. Koubaa, R. Abdennabi, A. S. B. Salah, and A. El Feki, "Microwave extraction of *Salvia officinalis* essential oil and assessment of its GC-MS identification and protective effects versus vanadium-induced nephrotoxicity in Wistar rats models," *Archives of Physiology and Biochemistry*, vol. 125, no. 5, pp. 404–413, 2018.
- [114] A. Ghorbani and M. Esmaeilzadeh, "Pharmacological properties of *Salvia officinalis* and its components," *Journal of Traditional and Complementary Medicine*, vol. 7, no. 4, pp. 433–440, 2017.

- [115] O. O. Igado, J. Glaser, M. Ramos-Tirado et al., "Isolation of a novel compound (MIMO2) from the methanolic extract of *Moringa oleifera* leaves: protective effects against vanadium-induced cytotoxicity," *Drug and Chemical Toxicology*, vol. 41, no. 3, pp. 249–258, 2018.
- [116] O. E. Adebisi, J. O. Olopade, and F. O. Olayemi, "Neuroprotective effect of *Grewia carpinifolia* extract against vanadium induced behavioural impairment," *Folia Veterinaria*, vol. 60, no. 4, pp. 5–13, 2016.
- [117] A. Soussi, F. Croute, J. P. Soleilhavoup, A. Kammoun, and A. El Feki, "Impact of green tea on oxidative stress induced by ammonium metavanadate exposure in male rats," *Comptes Rendus Biologies*, vol. 329, no. 10, pp. 775–784, 2006.
- [118] S. M. Chacko, P. T. Thambi, R. Kuttan, and I. Nishigaki, "Beneficial effects of green tea: a literature review," *Chinese Medicine*, vol. 5, no. 1, p. 13, 2010.
- [119] I. Matic, A. Guidi, M. Kenzo, A. Galgani, and M. Mattei, "Investigation of medicinal plants traditionally used as dietary supplements: a review on *Moringa oleifera*," *Journal of Public Health in Africa*, vol. 9, no. 3, 2018.
- [120] A. R. Volpe, P. Cesare, P. Aimola, M. Boscolo, G. Valle, and M. Carmignani, "Zinc opposes genotoxicity of cadmium and vanadium but not of lead," *Journal of Biological Regulators and Homeostatic Agents*, vol. 25, no. 4, pp. 589–601, 2011.
- [121] A. K. Chandra, R. Ghosh, A. Chatterjee, and M. Sarkar, "Vanadium-induced testicular toxicity and its prevention by oral supplementation of zinc sulphate," *Toxicology Mechanisms and Methods*, vol. 17, no. 4, pp. 175–187, 2007.
- [122] B. H. Bay, K. H. Sit, R. Paramanatham, and Y. G. Chan, "Hydroxyl free radicals generated by vanadyl[IV] induce cell blebbing in mitotic human Chang liver cells," *Biometals*, vol. 10, no. 2, pp. 119–122, 1997.
- [123] S. Shrivastava, A. Jadon, and S. Shukla, "Effect of tiron and its combination with nutritional supplements against vanadium intoxication in female albino rats," *The Journal of Toxicological Sciences*, vol. 32, no. 2, pp. 185–192, 2007.
- [124] C. Ly, J. Yockell-Lelievre, Z. M. Ferraro, J. T. Arnason, J. Ferrier, and A. Gruslin, "The effects of dietary polyphenols on reproductive health and early development," *Human Reproduction Update*, vol. 21, no. 2, pp. 228–248, 2015.
- [125] P. Rasoanaivo, C. W. Wright, M. L. Willcox, and B. Gilbert, "Whole plant extracts versus single compounds for the treatment of malaria: synergy and positive interactions," *Malaria Journal*, vol. 10, no. S1, 2011.
- [126] F. A. Olaolorun, A. A. Obasa, H. A. Balogun, O. O. Aina, and J. O. Olopade, "Lactational vitamin E protects against the histotoxic effects of systemically administered vanadium in neonatal rats," *Nigerian Journal of Physiological Sciences*, vol. 29, no. 2, pp. 125–129, 2014.
- [127] B. Vijaya Bharathi, G. Jaya Prakash, K. M. Krishna et al., "Protective effect of alpha glucosyl hesperidin (G-hesperidin) on chronic vanadium induced testicular toxicity and sperm nuclear DNA damage in male Sprague Dawley rats," *Andrologia*, vol. 47, no. 5, pp. 568–578, 2015.
- [128] S. Zendeboodi, A. Esmaili, A. Movahed et al., "The attenuative effects of oral resveratrol on renal changes induced by vanadium injection in rats," *Journal of Renal Injury Prevention*, vol. 8, no. 2, pp. 127–132, 2019.
- [129] G. Visalli, A. Facciola, M. P. Bertuccio, I. Picerno, and A. Di Pietro, "In vitro assessment of the indirect antioxidant activity of sulforaphane in redox imbalance vanadium-induced," *Natural Product Research*, vol. 31, no. 22, pp. 2612–2620, 2017.
- [130] M. Aureliano, "Decavanadate Toxicology and Pharmacological Activities: V₁₀ or V₁, Both or None?," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 6103457, 8 pages, 2016.
- [131] S. K. Ghosh, R. Saha, and B. Saha, "Toxicity of inorganic vanadium compounds," *Research on Chemical Intermediates*, vol. 41, no. 7, pp. 4873–4897, 2015.



Hindawi

Submit your manuscripts at
www.hindawi.com

