# Pseudomonas syringae subsp. savastanoi (ex Smith) subsp. nov., nom. rev., the Bacterium Causing Excrescences on Oleaceae and Nerium oleander L.

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From a study of the so-called bacterial canker of ash, caused by a variant of "*Pseudomonas savastanoi*" (Smith) Stevens, it became evident that this variant and the variants of "*P. savastanoi*" which cause olive knot and oleander knot can be distinguished from one another on the basis of their pathogenicity and host range. All isolates of "*P. savastanoi*" were recently classified by Dye et al. (Plant Pathol. **59:**153–168, 1980) as members of a single pathovar of *P. syringae* van Hall. It appears, however, that these isolates differ sufficiently from the other members of *P. syringae* to justify subspecies rank for them. The following classification and nomenclature are therefore proposed: *Pseudomonas syringae* subsp. *savastanoi* (ex Smith) subsp. nov., nom. rev., to include the olive pathogen (pathovar *oleae*), the ash pathogen (pathovar *fraxini*), and the oleander pathogen (pathovar *nerii*). The type strain of *P. syringae* subsp. *savastanoi* is ATCC 13522 (= NCPPB 639).

"Pseudomonas savastanoi" (Smith 1908) Stevens 1913 was previously used as the name of the bacterium which causes pernicious excrescences on several species of the Oleaceae. (Names in quotation marks are not on the Approved Lists of Bacterial Names [12], have not been validly published since 1 January 1980, and therefore do not have standing in bacterial nomenclature.) This bacterium was first adequately described from Olea europea L. by Smith (15), who named it "Bacterium savastanoi." A variant from the common ash (Fraxinus excelsior L.) was then described by Brown in 1932 (2) under the name "B. savastanoi var. fraxini"; it was later named "P. savastanoi subsp. fraxini" by Dowson in 1943 (5). The variant from Nerium oleander L. (Apocynaceae), which was first described by Ferraris (7) in 1926 as "P. tonelliana," was described more adequately by Smith (14), who renamed it "P. savastanoi subsp. nerii'' in 1928.

My recent studies of the so-called bacterial canker of common ash (7a, 7b) have yielded biochemical, serological, and pathological data that cast additional light on the nature of the isolates obtained from ash, olive, oleander, privet, and jasmin.

In their proposed nomenclature of the plantpathogenic fluorescent pseudomonads, Young et al. (23) did not distinguish among the different variants, and they classified all isolates of "P. savastanoi" as members of P. syringae pathovar savastanoi. Their interpretation of the term pathovar, however, is in this case not in agreement with the definition of the term as recommended in the International Code of Nomenclature of Bacteria (8).

According to the available data, the present classification and nomenclature of the organisms under discussion are inadequate. The purpose of this paper is to rectify this situation.

## MATERIALS AND METHODS

Bacterial strains. The strains studied are listed in Table 1.

Methods. Descriptions of the methods employed are reported elsewhere (7b).

## RESULTS

Over 40 biochemical and physiological features were determined for the strains. All "P. savastanoi" strains had nearly the same characteristics. Significant variances were found only in the production of levan and in the hydrolysis of pectate (not correlated to host plant or pathogenicity) and in the production of indolyl-acetic acid and cytokinin-like compounds. The latter two substances are not produced (or are produced only in very small amounts) by isolates from ash, whereas they are produced by isolates from other host plants. The tests which appear to be useful in differentiating between "P. savastanoi" and P. syringae are listed in Table 2. For differential characters, also see Sands et al. (11)

All of the "P. savastanoi" strains produced similar titers with an antiserum prepared against strain NCPPB 639 (from Olea europea). The antiserum was absorbed with a cross-reacting P. syringae strain (NCPPB 191). After cross-ab-

Strain <sup>a</sup>	Origin		
Stram	Host	Country	
"P. savastanoi"			
PD 109	F. excelsior L.	Netherlands	
PD 116	F. excelsior L.	Netherlands	
PD 119	F. excelsior L.	Netherlands	
PD 120	F. excelsior L.	Netherlands	
PD 159	F. excelsior L.	Netherlands	
PD 160	F. excelsior L.	Netherlands	
PD 161	F. excelsior L.	Netherlands	
PD 179	F. excelsior L.	Netherlands	
PD 206	F. excelsior L.	Netherlands	
NCPPB 1464	F. excelsior L.	U.K.	
NCPPB 1006	F. excelsior L.	<b>U.K</b> .	
CNBP 1838	F. excelsior L.	France	
NCPPB 639	O. europea L.	Yugoslavia	
NCPPB 2327	O. europea L.	Italy	
NCPPB 640	N. oleander L.	Yugoslavia	
PD 181	N. oleander L.	Spain	
NCPPB 2328	Ligustrum japonicum Thbg.	Italy	
CNBP 1751	Jasminum sp.	Greece	
P. syringae			
NCPPB 281	Syringa vulgaris L.	U.K.	
NCPPB 191	Persea americana Mill.	Israel	
NCPPB 981	Populus canadensis Mnch. 'Eugenei'	U.K.	
"P. mors-prunorum"	• -		
NCPPB 560	Host unknown	U.K.	
"P. maculicola"			
IPO 154	Brassica oleracea L.	Netherlands	
Saprophytic fluorescent pseudomonads			
PD 117	F. excelsior L.	Netherlands	
NCPPB 1465 <sup>b</sup>	F. excelsior L.	U.K.	

TABLE 1. Strains used in this study

<sup>a</sup> The strains are maintained under these names in the collections referred to as follows: PD, Culture Collection of the Plant Protection Service, Wageningen, The Netherlands; NCPPB, National Collection of Plant-Pathogenic Bacteria, Harpenden, U.K.; CNBP, Collection Nationale de Bactéries Phytopathogènes, Angers, France; IPO, Culture Collection of the Research Institute for Plant Protection, Wageningen, The Netherlands.

<sup>b</sup> NCPPB 1465 was described as a deviating pathogenic strain of "*P. savastanoi* subsp. *fraxini*" by Šutić and Dowson (19). In my hands it deviated morphologically, biochemically, and serologically from *P. syringae* subsp. *savastanoi* and was found to be nonpathogenic on four different hosts. It should therefore be regarded as a saprophytic pseudomonad.

sorption, only the "P. maculicola" strain showed close antigenic relationship to "P. savastanoi" strains.

No significant morphological differences were found between the strains of "P. savastanoi," and only small differences were observed between these strains and the other phytopathogenic pseudomonads tested.

The results of the pathogenicity tests are presented in Table 3. The host plants used were *Fraxinus excelsior* L., *Olea europea* L., *Nerium oleander* L., and *Forsythia intermedia* Zab.

# DISCUSSION

From my previous studies (7b) and those of others (2, 11, 18, 19, 21, 22), it has become apparent that "P. savastanoi" isolates from different hosts are almost indistinguishable morphologically, biochemically, and serologically.

Only the production of indolyl-acetic acid and cytokinin-like compounds differentiated between the isolates from ash and those from other host plants. These substances are most likely related to pathogenicity, as will be discussed below.

However, the isolates from these different host plants show different pathogenicities and host ranges (4, 7b, 14, 18, 20). The isolates from ash can be clearly distinguished from those of other hosts by a deviating pathogenicity (18; Janse, in press). They evoke wartlike necrotic bark swellings with abundant periderm instead of large, parenchymatous galls. This can possibly be explained by the restricted production of growth substances by isolates from ash. The other isolates of "P. savastanoi" were found to produce these substances in rather large amounts in vitro, and elevated levels were also

 TABLE 2. Biochemical tests useful in differentiating

 P. syringae subsp. savastanoi from P. syringae

 subsp. syringae<sup>a</sup>

Tests	P. syringae subsp. savastanoi	P. syringae subsp syringae
Hydrolysis of:		
Gelatin	_	+
Esculin	-	+
Arbutin	<ul> <li>or weak</li> </ul>	+
Casein	<ul> <li>or weak</li> </ul>	+
Acid from:		
D-(+)-Raffinose	-	+
Erythritol	-	+
Alkali from:		
L-(+)-Tartrate	+	-

<sup>a</sup> Also see Sands et al. (11).

found in their galls (1, 17, 22). The host range of the isolates from ash is limited to the *Oleaceae*.

The isolates from olive differ from the isolates from ash by producing large galls instead of necrotic bark swellings; they differ from isolates from oleander in host range (Table 3). Isolates from olive usually do not infect *N. oleander*, and it was for this reason that the oleander organism was originally described as a separate species, "*P. tonelliana*" (7). Although on two occasions (10, 22) strains from olive have been reported to infect *N. oleander*, this is generally not the case.

The isolates from oleander form galls and can therefore be distinguished from isolates from ash; they differ from isolates from olive in host range (Table 3).

On the basis of these facts, it is concluded that the bacterial isolates causing excressences on ash, olive, and oleander must be ranked separately at the level of pathovar as defined in the *Bacteriological Code* (8) and not as interpreted by Dye et al. (6). Determination of the status of the jasmin and privet isolates requires a more comprehensive host-range study.

Isolates of "P. savastanoi" were recently named P. syringae pathovar savastanoi (6, 23) as a result of investigations (e.g., 3, 9, 11) which have shown that "P. savastanoi" is closely related to P. syringae van Hall. However, isolates belonging to "P. savastanoi" can be readily distinguished biochemically, serologically, and pathologically from P. syringae (Table 2) and its subgroups (11); they should therefore be considered at the subspecies level. As the epithet "savastanoi" has had no standing in bacterial nomenclature since 1 January 1980 (12), it is here revived for bacterial pathogens causing excressences on Oleaceae and N. oleander L.

Description of *Pseudomonas syringae* subsp. savastanoi (ex Smith 1908) subsp. nov., nom. rev. (sa·vas·ta'no·i· M.L. gen. noun savastanoi of Savastano, named for L. Savastano, the first to study olive knot).

Gram-negative, nonsporeforming rods with rounded ends, 0.3 to 0.7 by 1.0 to 1.8  $\mu$ m, occurring singly or in pairs; motile by means of one to five polar flagella. Rather slow-growing, gray-white, smooth, glistening, raised and circular or slightly irregular to undulate colonies are produced on nutrient agar; levan negative or levan positive on nutrient-sucrose (5%) agar; produces a weak, blue-green fluorescent, diffusible pigment on King's B medium; some strains produce a brown diffusible pigment. Metabolism is respiratory. Oxidase negative, catalase positive. Acid is produced from D-(+)-galactose, glucose, D-(+)-ribose, sucrose (slow), D-(+)xylose, and mannitol; no acid is produced from maltose, D-(+)-raffinose, erythritol, or salicin; alkali is produced from L-(+)-tartrate; esculin, arginine, gelatin, and starch are not hydrolyzed; generally, arbutin and casein are not hydrolyzed. Nitrates are not reduced. H<sub>2</sub>S is not produced from cysteine. No growth occurs in nutrient broth at 37°C or with 5% NaCl. Hypersensitivity is produced in tobacco leaves. The deoxyribonucleic acid contains 60 mol% guanine plus cytosine (3). Causes galls and wartlike excrescences on various species of Oleaceae and N. oleander L. Gall-forming isolates produce indolyl-acetic acid and cytokinin-like substances in vitro.

The type strain of this subspecies is ATCC 13522 (= NCPPB 639). This strain was isolated by D. Šutič from *Olea europea* in Yugoslavia. Its description is identical to that of the subspecies, but with the following modifications: levan negative on nutrient-sucrose (5%) agar; casein hydrolysis is weak; produces galls on *O. europea* L. and *F. excelsior* L.

The following pathovars of *P. syringae* subsp. savastanoi are recognized: pathovar oleae, causing parenchymatic galls on various species of the Oleaceae (15, 16); pathovar nerii, causing parenchymatic galls or wartlike excrescences on *N. oleander* L. and various species of the Olea-

TABLE 3. Results of pathogenicity tests with isolates of *P. syringae* subsp. *savastanoi* on different host plants

Isolate(s) from:				
	Ash	Olive	Oleander	Forsythia
Ash	+	+	_	_
Olive	$\oplus$	Ð	_	_
Oleander	Ť	ĕ	Ð	_
Jasmin	-	ĕ	-	Ŧ
Privet	$\oplus$	Ť	-	-

"Symbols: +, Necrotic swellings;  $\oplus$ , parenchymatous galls; -, no pathogenic reaction.

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ceae (14); and pathovar *fraxini*, causing wartlike excressences on *F. excelsior* L. and *O. europea* L. (2, 5, 13).

### ACKNOWLEDGMENT

I am indebted to H. J. Miller for his encouragement and help during this investigation.

#### **REPRINT REQUESTS**

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