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Pumping a Ring-Sliding Molecular Motion by a Light-Powered Molecular Motor

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Supporting Information

ABSTRACT: Designing artificial molecular machines to execute complex mechanical tasks, like coupling rotation and translation to accomplish transmission of motion, continues to provide important challenges. Herein, we demonstrated a novel molecular machine comprising a second-generation light-driven molecular motor and a bistable [1]rotaxane unit. The molecular motor can rotate successfully even in an interlocked [1]rotaxane system through a photoinduced *cis*-to-*trans* isomerization and a thermal helix inversion, resulting in concomitant transitional motion of the [1]rotaxane. The transmission process was elucidated via ¹H NMR, ¹H–¹H COSY, HMQC, HMBC, and 2D ROESY NMR spectroscopies, UV–visible absorption spectrum, and density functional theory calculations. This is the first demonstration of a molecular motor to rotate against the appreciably noncovalent interactions between dibenzo-**24**-crown-**8** and *N*-methyltriazolium moieties comprising the rotaxane unit, showing operational capabilities of molecular motors to perform more complex tasks.



■ INTRODUCTION

Inspired by biological molecular machines (such as ATPases, kinesin, bacterial flagellar motors, etc.) widely present in nature,¹⁻³ artificial molecular machines have been successfully constructed by synthetic chemists to mimic the diverse functions of biological systems.⁴ Extensive efforts have been focused on the construction, manipulation, and application of artificial molecular machines.^{5,6} Diversified molecular machines working at a single-molecule scale or amplifying their motion in a collective manner have been well realized, exhibiting the great significance and possibilities of molecular machines." Biological molecular machines and macroscopic machines have reminded us of the significance to couple two or more machinery operations in a transmission mechanism. Hence, it would be intriguing if such mechanical transmission mechanism could be achieved in the artificial molecular system. Recent efforts have included the successful coupling of the photo-isomerization of azobenzene with mechanical rotation motion,⁸ and the coupled intramolecular mechanical transmission from the light-driven unidirectional rotary motion to the synchronous sliding-and-rotation movement of a connected biaryl rotor.9 These achievements supported strategies and guided inspirations for the design and construction of mechanically coupled artificial molecular machines. However, designing multifunctional mechanical molecular systems remains challenging, especially in addressing the fundamental question of whether we can combine molecular rotary motion with linear sliding motion in a mechanically interlocked system.¹⁰

Molecular motors are representative classes of molecular machines, which can perform directional rotation under light and heat, have been proven to operate successfully in a variety of environments.^{11–14} While rotaxanes are representative classes of mechanically interlocked molecules,¹⁵ in particular, bistable [2]rotaxane based on the noncovalent interactions between the crown ether with ammonium and triazolium stations, have been extensively studied because of their clear shuttling process in response to pH.¹⁶ Herein, our motivation is to integrate these two types of molecular units into a single-molecular system, which would be a great model to explore if molecular motor can operate against the appreciably non-covalent interactions in an interlocked [1]rotaxane, and if the linear motion of the rotaxane can be driven via a transmission

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mechanism. Compared to the previously reported [1]rotaxane transmission system,¹⁰ we mainly focus on the operation of molecular motors in interlocking structures, whether the molecular motor can achieve the four-step rotational cycle under the constraint of certain noncovalent interactions, which would provide deeper insight into the operational capabilities of the second-generation molecular motors in more complex mechanical molecular systems. We envision that this research would push the general exploration of multicomponent molecular machines working by coupled motion at the molecular scale, and further give unprecedented insights to the nanoworld.

The envisioned design of a molecular machine with distinct mechanical units allowing transmission of motion, that is, the [1]rotaxane *cis*-18 H^+ molecular motor system, is shown in Figure 1a. A light-driven second-generation molecular motor is



Figure 1. (a) Chemical structures of *cis*-18H⁺ and (b) schematic representation of photo-thermal driven process of [1]rotaxane stable *cis*-18. The mechanical transmission cycle is initiated with the deprotonated stable *cis*-18 by adding base DBU, wherein the DB24C8 ring locates at the MTA sites because of their host-guest combination. UV irradiation drives the first half-cycle rotation of the motor part. This rotation motion further transmits into the DB24C8 ring, leading to its dissociation with MTA sites and subsequent sliding along the long carbon chain, thus yielding unstable *trans*-18. Then the THI of the motor unit produces stable *trans*-18, and irradiation further drives the second half-cycle rotation, transmitting the sliding motion of the DB24C8 ring back to the MTA site, yielding unstable *cis*-18. A second THI from unstable *cis*-18 to stable *cis*-18 completes the final step of the one cycle.

covalently immobilized in a [1]rotaxane. The upper rotor part of the motor is modified with a crown ether macrocycle, dibenzo-24-crown-8 (DB24C8); and the lower stator part of the motor is connected with a linear thread, which contains two distinguishable recognition sites relative to DB24C8, the benzylalkylammonium hexafluorophosphate (BAA) station and a *N*-methyltriazolium hexafluorophosphate (MTA) station, as well as an auxiliary recognition site, the secondary amide (SA) group.¹⁷ The linear component is mechanically threaded into the macrocycle with a bulky 3,5-di-*tert*butylphenyl group applied as stopper.¹⁸ First, the macrocycle DB24C8 is stationary at the BAA recognition site because of the stronger binding constant^{16b,d,e} (Figure 1a). After adding base DBU to the system, the BAA station is deprotonated, leading to the movement of the DB24C8 macrocycle from the BAA site to the weaker MTA recognition site^{16b,d,e} (Figure 1b). Then the stable cis-18 can be transformed into unstable *trans*-18 via photo-isomerization under UV irradiation (λ = 365 nm). At this moment, the molecular motor would rotate against the host-guest combination of the macrocycle DB24C8 and guest MTA, resulting in the "pulling" process of the DB24C8 ring away from the MTA recognition site. Then the unbound macrocycle is expected to slide along the alkyl chain (AC) region near the SA group because of the overall molecular configuration (trans motor). After heating $(60 \ ^{\circ}C)$ the system, unstable *trans*-18 will undergo a thermal helix inversion (THI) and transform into stable trans-18, at this stage, the geometry of the entire molecular system basically remains. Continued UV irradiation and heating can trigger the molecular motor to perform a four-step cycle of directional rotation, and the rotor part of the molecular motor would pump the macrocycle slide along the AC region to the MTA station back and forth, thus realizing a transmission behavior in the bifunctional mechanical system from molecular rotational motion to linear sliding motion. This manner of molecular machinery motion would prove that the molecular motor resists appreciably noncovalent interactions, which has been proven to lock the rotation of a molecular motor in a specific molecular system.¹⁹

RESULTS AND DISCUSSION

Design. The design of *cis*-18H⁺ is based on the following principles: (i) the S-S molecular motor is selected because of its long half-life at room temperature,^{14b,20} which is convenient to track the entire driving process by NMR spectroscopy; (ii) crown ether DB24C8 is selected because of its flexibility, which might facilitate the mechanical transmission process and the successful full-cycle rotation of the motor unit; (iii) the length of carbon chain between MTA station and BAA station should be sufficient to provide spatial pathway for the ringsliding motion, meanwhile enabling the successful THI process to complete the full rotation cycle of the confined molecular motor; (iv) considering that only MTA station exists in the system after deprotonation of BAA station, in this case, the DB24C8 ring is hard to leave the current station, it is necessary to introduce another weaker SA group to assist the subsequent photothermal-driven process; 17 (v) the MTA station is chosen considering the weaker interactions between the MTA and DB24C8 instead of BAA station with a high-affinity interaction with the crown ether which would lock the rotation of the molecular motor;¹⁹ (vi) the BAA station is still employed to template the rotaxane formation to enable the efficient synthesis of the interlocked structure, and meanwhile the acid/base-responsive capability of BAA sites would enable the facile removal of the high-affinity synthesis template by deprotonation.^{5h,16d,e}

Synthesis. The synthesis of [1]rotaxane *cis*-18H⁺ is illustrated in Scheme 1. The target *cis*-18H⁺ was divided into three components—molecular motor, DB24C8 macrocycle, and linear thread. One of the key challenges is to synthesize a bifunctional molecular motor *cis*-8 with groups at precisely defined positions in stator and rotor to allow rotaxane assembly in such a way the rotary motor function is not compromised. Toward this goal, first a bromine-substituted rotor and a triflate-modified stator were synthesized in multiple steps (detailed are given in Scheme S1). Subsequently, a

Scheme 1. Partial Synthesis of [1]Rotaxane *cis*-18H⁺, NonInterlocked Reference Compound *cis*-21H⁺ and Corresponding [2]rotaxane 25H⁺



mixture of cis-8 and trans-8 isomers was obtained through Barton-Kellogg diazo-thioketone coupling²¹ of thioketone 4 and hydrazone 5 followed by an alkaline hydrolysis reaction. The isomers were separated by flash column chromatography and the structure of cis-8 was confirmed by X-ray single crystal diffraction (Figure S1). Azide-modified linear thread 12 and borate-modified DB24C8 macrocycle 13 were synthesized by following literature procedures (Schemes S1 and S2). Next, the DB24C8 macrocycle was mounted to the molecular motor by Suzuki coupling reaction. Alkynyl-substituted cis-16 was obtained by subsequent etherification. Finally, [1]rotaxane cis-17 was obtained through a typical self-threading in apolar solvent, in which a copper(I)-catalyzed Huisgen [5] alkyneazide 1,3-dipolar cycloaddition was employed to covalently bridge between cis-16 and thread 12,²² at this moment, two different configuration isomers of cis-17 generated because the single bond of the naphthalene unit is asymmetrically attached to catechol ring of DB24C8 (Scheme S2). After a mild methylation reaction by CH₂I, the final [1]rotaxane cis-18H⁺ was successfully obtained. Furthermore, the isomer rotaxane trans-18H⁺ was synthesized following a similar strategy (Scheme S2). In addition, we also synthesized an noninterlocked reference compound *cis*-21H⁺ and a corresponding [2] rotaxane **25H**⁺ through a similar synthesis method (Scheme 1). All compounds were fully characterized by ¹H NMR, ¹³C NMR spectroscopy and high-resolution electrospray ionization (HR-ESI).

Structure Verification of [1]Rotaxane *cis*-18H⁺. To further confirm the structure of [1]rotaxane *cis*-18H⁺, a comparison of the ¹H NMR spectra among the interlocked *cis*-18H⁺, noninterlocked *cis*-21H⁺ and the corresponding [2]rotaxane 25H⁺ was performed (Figure 2). The characteristic proton signals were assigned by ¹H-¹H COSY, HMQC, and HMBC (Figures S2-S4, S6, and S12). Compared to the noninterlocked *cis*-21H⁺ (Figure 2a), certain absorptions of *cis*-18H⁺ have split into two groups (Figure 2b), such as proton H₁₄ on the MTA, aromatic protons H₃, H₂₀, and H₂₁, methyl proton H₁₅ and tertiary butyl proton H₁, this is caused by the two different configuration isomers, which are produced during



Figure 2. ¹H NMR (500 MHz, 298 K, CD₃CN) spectra of (a) noninterlocked reference compound *cis*-21H⁺, (b) [1]rotaxane *cis*-18H⁺ (inset: magnification of proton H_{20}), (c) corresponding [2]rotaxane 25H⁺.

the self-threading and "Click Chemistry" reaction (Scheme S2). Distinctive changes of other ¹H NMR signals were observed as well, for instance the methylene protons H₄ and H₅ near the **BAA** station shifted downfield ($\Delta \delta = 0.33$ ppm, $\Delta \delta$ = 0.13 ppm, respectively), which is attributed to the hydrogen-bonding interactions with the oxygen atoms of the crown ether moiety. The amide proton H₉ and the tert-butyl protons H₁ shifted upfield ($\Delta \delta = -0.17$ ppm, $\Delta \delta = -0.14$ ppm, respectively) because of the shielding effect of the crown ether ring. In addition, a comparison with the corresponding [2]rotaxane 25H⁺, which is also an interlocked structure, shows that the ¹H NMR signals of protons near the BAA station are highly consistent (Figure 2c). The structure of cis-18H⁺ was further confirmed by 2D ROESY spectroscopy (Figure S5, S7, and S13) and both *cis*-18H⁺ and [2]rotaxane 25H⁺ showed a strong ROEs effect between DB24C8 and the BAA station, while cis-21H⁺ did not show any obvious ROEs

effect. The combined data clearly indicate that $cis-18H^+$ features an interlocked [1]rotaxane structure.

Operation of Reference [2]Rotaxane. In order to fully understand the movement of the macrocycle after deprotonation of the **BAA** recognition site. Two distinct reference [2]rotaxane $24H^+$ and $25H^+$ were synthesized (Schemes 1c and S3). The localization of the **DB24C8** was investigated via ¹H NMR spectroscopy (Figure 3b-e). In the initial state, the



Figure 3. (a) Scheme representation of the DBU-triggered motion of the [2]rotaxane 24H⁺ and rotaxane 25H⁺; ¹H NMR (400 MHz, 298 K, CD₃CN) spectra of (b) [2]rotaxane 24H⁺; (c) [2]rotaxane 24 obtained after adding 1.50 equiv of DBU to [2]rotaxane 24H⁺; ¹H NMR (400 MHz, 298 K, CD₂Cl₂) spectra of (d) [2]rotaxane 25H⁺; (e) [2]rotaxane 25 obtained after adding 1.50 equiv of DBU to rotaxane 25H⁺.

DB24C8 was located on the BAA station (Figures S8, S9, S12, and S13), after addition of DBU to the 24H⁺ and 25H⁺, respectively, the deprotonated 24 and 25 can be obtained. The obvious changes of ¹H NMR spectra between protonated rotaxane 24H⁺ and deprotonated rotaxane 24 showed that the DB24C8 would move off the BAA station and was confined in the AC region after addition of DBU (Figure 3b,c). Furthermore, the protons H_4 and H_5 near the BAA station shifted upfield ($\Delta \delta = -0.51$ ppm, $\Delta \delta = -0.37$ ppm, respectively) because they lost their hydrogen bonding interactions with the oxygen atoms of DB24C8, while the protons H₁ shifted downfield ($\Delta \delta$ = 0.06 ppm) because of the disappearance of the DB24C8 shielding effect. Meanwhile, the protons on the AC region shifted upfield ($\Delta \delta = -0.08$ ppm) because of the shielding effect of the DB24C8. The protons H₉, H₁₀, H₁₁, H₁₂, H₁₃, and H₁₄ expectedly shifted downfield slightly ($\Delta \delta$ = 0.25 ppm, $\Delta \delta$ = 0.07 ppm, $\Delta \delta$ = 0.05 ppm, $\Delta \delta$ = 0.06 ppm, $\Delta \delta = 0.08$ ppm, $\Delta \delta = 0.09$ ppm, respectively) ascribed to the deshielding effect of DB24C8. In this state, the DB24C8 ring kept a procumbent configuration, in which the benzene groups of DB24C8 would be closed to the triazole and the SA group. In addition, the protons on the AC region showed a strong correlation with the protons on the DB24C8

ring according to the 2D ROESY spectrum (Figure S11). All the above results confirmed that the DB24C8 ring moved from the BAA station to the AC region after adding DBU to the rotaxane 24H⁺. Interestingly, this result seems inconsistent with previous work, which indicated that the amide moieties could serve as molecular stations for DB24C8¹⁷ The key factor is the weaker binding force between the SA group and DB24C8, leading to the DB24C8 ring staying in the AC region under thermodynamic control. Similarly, the comparison between the ¹H NMR spectra of protonated rotaxane 25H⁺ and deprotonated rotaxane 25 shows that the DB24C8 would move away from BAA and combine with MTA station after addition of DBU (Figure 3d,e), which is consistent with a typical bistable [2]rotaxane.^{16b,d,e} Remarkably, the proton H_{14} on the MTA station dramatically shifted downfield ($\Delta \delta = 0.61$ ppm) because of the deshielding effect from the aromatic rings of the DB24C8. Meanwhile, the protons H₁₃, H₁₅, and H₁₆ obviously shifted upfield ($\Delta \delta = -0.51$ ppm, $\Delta \delta = -0.45$ ppm, $\Delta \delta = -0.43$ ppm, respectively) because of the shielding effect of the DB24C8. The localization of the DB24C8 was confirmed though 2D ROESY spectroscopy (Figure S15).

The abovementioned results illustrate the movement of the macrocycle in the presence and absence of the MTA station. In the presence of a stronger secondary MTA recognition site, DB24C8 would stay at the MTA station, whereas in the presence of only a weak SA group, the DB24C8 ring prefers to move to a thermodynamically stable AC region. The detailed understanding of the states of rotaxanes, which potentially exist in *cis*-18H⁺, will provide a good experimental basis for the subsequent characterization and analysis of [1]rotaxane *cis*-18H⁺.

Operation of [1]Rotaxane *cis***-18H**⁺. Because the **DB24C8** ring of [1]rotaxane *cis***-18H**⁺ is located at the **BAA** station in the initial state, the base DBU needs to be added to the system triggering it to enter stable *cis***-18**. It should be noted that the ¹H NMR spectra of current *cis***-18H**⁺ in Figure 4a and the same *cis***-18H**⁺ in Figure 2b are not exactly consistent, especially the ratio of doublet H_{14} because they were prepared in different batches, considering that the





synthesis conditions are difficult to achieve completely consistent, resulting in small differences in the ratio of the two configuration isomers. The comparison of the ¹H NMR spectra between protonated cis-18H⁺ and deprotonated stable cis-18 confirmed that DB24C8 could move from the BAA recognition site to the MTA recognition site after addition of DBU (Figure 4a,b). A large number of protons underwent significant chemical shift changes, which is consistent with the presence of [2]rotaxane 25H⁺ (Figure 3d,e). The proton H_{14} shifted downfield ($\Delta \delta = 0.11$ ppm) because of the hydrogen bonding interaction with the oxygen atoms of the DB24C8 ring. On the other hand, the protons H₁₂, H₁₃, H₁₅, H₁₆, and H₁₇ shifted upfield ($\Delta \delta$ = -0.17 ppm, $\Delta \delta$ = -0.20 ppm, $\Delta \delta$ = -0.24 ppm, $\Delta\delta = -0.21$ ppm, $\Delta\delta = -0.22$ ppm, respectively), as a result of the shielding effect of the DB24C8 ring. The protons H₄ and H₅ near the BAA station shifted upfield and downfield ($\Delta \delta = -0.59$ ppm, $\Delta \delta = 0.11$ ppm, respectively) because of the departure of the DB24C8 ring. In particular, the methyl proton H₂₇ on the rotor moiety dramatically shifted downfield ($\Delta \delta$ = 0.19 ppm) because the **DB24C8** macrocycle recognized with the MTA site, affecting the spatial configuration of the molecular motor (Figure 8). Compared with noninterlocked reference compound cis-21H⁺, the ¹H NMR signals did not change significantly after addition of DBU (Figure 4c,d). In addition, the MTA proton H_{14} of stable cis-18 exhibited a strong ROE effect with DB24C8 protons (a-f) (Figures S19 and S20). These results unequivocally confirmed that the DB24C8 ring could move from the BAA recognition site to the MTA recognition site after addition of DBU.

We hypothesized that the rotational motion of the molecular motor would drive the dissociation of the noncovalent host– guest combination between the **DB24C8** ring and **MTA** recognition site. This process was investigated via ¹H NMR spectroscopy (Figure 5) (see partial enlargement in Figures S21–S24). Upon irradiation of UV light at 365 nm for 2 h, stable *cis*-18 converts into unstable *trans*-18 because of the photoisomerization (Figure 6a,b) with concomitant change in



Figure 5. ¹H NMR (500 MHz, 298 K, CD₃CN) spectra of (a) [1]rotaxane stable *cis*-18; (b) [1]rotaxane unstable *trans*-18 obtained after stable *cis*-18 was irradiated with UV (365 nm) for 2 h; (c) [1]rotaxane stable *trans*-18 obtained after unstable *trans*-18 was heated at 60 °C for 24 h in the dark; (d) [1]rotaxane stable *trans*-18 obtained after adding 1.50 equiv of DBU to *trans*-18H⁺.



Figure 6. Schematic representation of the photo-thermal driven process of (a) [1]rotaxane stable *cis*-18 and (b) [1]rotaxane stable *trans*-18.

the ${}^1\!\mathrm{H}$ NMR signals of the protons on the molecular motor. The protons H_{20} and H_{21} obviously shifted downfield ($\delta = 0.94$ ppm, $\Delta \delta$ = 1.28 ppm, respectively), and the proton H₁₉ shifted upfield ($\delta = -0.36$ ppm) (Figure S21), which indicated that the shielding effect and the deshielding effect of the naphthalene have been eliminated in the trans isomer. Proton H_{27} shifted downfield ($\delta = 0.25$ ppm) (Figure S24); this chemical shift can be attributed to the increased deshielding effect of the pseudo-equatorial methyl group. Furthermore, the aliphatic protons H_{28} and $H_{28'}$ in the upper-half of the motor shifted upfield ($\delta = -0.12$ ppm, $\delta = -0.16$ ppm, respectively) (Figure S22) because of the difference in the conformational structure of the six-membered ring in stable cis-18 and unstable *trans*-18. These results suggest that the [1] rotaxane stable *cis*-18 has transformed from the stable cis state to unstable trans state, was consistent with previous reports.^{12b,14b} It is important to note that extended irradiation for 2 h resulted in a photostationary state (PSS) with a ratio of 58:42 (unstable trans-18/stable cis-18), calculated by the relative integration of the signals for the methyl group of each isomers. Unsurprisingly, the ratio of two isomers was lower than that in previous reports^{12b,14b} because the interlocked [1]rotaxane limited the movement of the molecular motor to some extent. It should be emphasized that the chemical shifts of protons on the thread part also changed significantly. The protons H₁₂, H₁₃, H₁₅, H₁₆, H₁₇, and H₁₈ near the MTA station were all shifted downfield $(\Delta \delta = 0.13 \text{ ppm}, \Delta \delta = 0.21 \text{ ppm}, \Delta \delta = 0.46 \text{ ppm}, \Delta \delta = 0.26$ ppm, $\Delta \delta = 0.35$ ppm, $\Delta \delta = 0.34$ ppm, respectively) because of the disappearance of the shielding effect from the DB24C8 ring (Figure S22). Proton H_{14} on the MTA station shifted upfield ($\Delta \delta$ = 0.30 ppm) because of the absence of hydrogen bonding interactions with the oxygen atoms of DB24C8. These results indicated that DB24C8 of stable cis-18 is located away from the MTA recognition site after illumination. Though the ¹H NMR signals of protons H₉ on the SA group and H₆, H₇, H₈, and H₁₀ near the SA group did not change significantly, it is worth noting that the protons on the AC region obviously shifted upfield ($\Delta \delta = -0.04$ ppm) (Figure S23), which could be attributed to the shielding effect of DB24C8. Meanwhile, the strong ROE effects between the crown ether hydrogen (a–f) and AC region were observed by the 2D ROESY spectrum (Figure S33). The results confirmed that DB24C8 of stable cis-18 moved from the MTA station to AC region after UV irradiation, which was consistent with the abovementioned reference [2]rotaxane 24 formed by addition DBU to [2]rotaxane 24H⁺ (Figure 3b,c). After heating the unstable trans-18 at 60 °C overnight in the dark, the molecular



Figure 7. (a) UV–vis absorption spectra (CH₂Cl₂, 283 K) of [1]rotaxane stable *cis*-**18H**⁺ before and after adding the DBU, UV irradiation for 60 min, heating for 24 h; (b) [1]rotaxane stable *trans*-**18H**⁺ before and after adding the DBU, UV irradiation for 60 min, heating for 24 h.

motor underwent a THI process from an unstable trans state to a stable trans state. Considering no major structural change between these two states, most signals showed no obvious shift (Figure 5c). However, the H₂₈ and H_{28'} on the upper-half of molecular motor-shifted downfield (Figure S22), and the protons H₂₇ on the methyl significantly shifted upfield ($\Delta \delta =$ -0.42 ppm) (Figure S24), both of which indicated that the unstable *trans*-18 had transformed into stable *trans*-18. Therefore, the motor of stable *cis*-18 had been operated successfully for half cycle.

Operation of [1]Rotaxane trans-18H⁺. To further verify the authenticity of the abovementioned half cycle, we synthesized a reference compound trans-18H⁺ through a comparable strategy used for *cis*-18H⁺ (Scheme S2). Similarly, the DB24C8 ring of [1]rotaxane trans-18H⁺ located at the BAA station in the initial state, after adding the DBU to the system, BAA station was deprotonated. As expected, only the methylene protons H₄ near the BAA station shifted upfield because of deprotonation ($\Delta \delta = 0.24$ ppm), the ¹H NMR signals of protons in other regions did not change obviously (Figure S34) because the DB24C8 ring cannot move to the MTA recognition site after deprotonation because of the limitation of the spatial structure of stable trans-18. By comparing the ¹H NMR spectra of stable trans-18 obtained from deprotonation of trans-18H⁺ and the same stable trans-18 obtained from stable cis-18 after UV irradiation and heating, though some unexpected byproduct accumulated because of sequential isomeric reactions without purification, partial characteristic signal peaks were highly consistent (Figure 5c,d). The results further confirmed that stable cis-18 can be operated according to the set program after illumination and heating. Based on the cumulated data, it can be concluded that the second-generation light-driven molecular motor can resist the considerable noncovalent interactions to allow directional rotation and accomplish a shuttling movement of the rotaxane via a transmission mechanism.

To further prove the complete four-step cycle of stable *cis*-**18**, the solution of stable *trans*-**18** was irradiated under UV (365 nm) for 2 h and subsequently heated at 60 °C for 24 h in the dark. However, the signals of ¹H NMR spectra were too complicated to unequivocally assign because of the incomplete photoisomerization and THI processes (Figures S25 and S26). To further explore these two processes, the latter half cycle was accomplished by driving stable *trans*-**18** directly, which was obtained from synthetic *trans*-**18H**⁺ after adding DBU (Figures 6b and S34). Upon the UV irradiation, the molecular motor stable *trans*-**18** would transform into an unstable *cis* state from a stable trans state. At the meantime, the **DB24C8** ring would move to the **MTA** recognition site because of the decrease of tension in the molecule. Notably, the long and flexible thread component enabled this movement. In this process, the corresponding ¹H NMR signals changed significantly (Figure S41a,b), notably, the protons H_{20} and H_{21} dramatically shifted upfield ($\delta = -0.91$ ppm, $\Delta \delta = -1.10$ ppm, respectively), and the proton H₁₉ shifted downfield ($\delta = 0.35$ ppm), and the proton H_{27} obviously shifted downfield ($\delta = 0.25$ ppm), which indicated that the stable trans-18 transformed into unstable cis-18. Importantly, the proton H_{14} on the MTA station dramatically shifted downfield ($\Delta \delta = 0.40$ ppm) because of the hydrogen bonding interactions with the oxygen atoms of **DB24C8**. Meanwhile the methyl proton H_{15} on the MTA station shifted upfield significantly ($\Delta \delta = -0.46$ ppm), attributed to the shielding effect from the DB24C8 ring. The protons H₁₂, H₁₃, H₁₆, H₁₇, and H₁₈ near the MTA station all shifted upfield ($\Delta \delta$ = -0.07 ppm, $\Delta \delta$ = -0.13 ppm, $\Delta \delta$ = -0.23 ppm, $\Delta\delta = -0.34$ ppm, $\Delta\delta = -0.37$ ppm, respectively). A clear ROE effect was observed between the protons H_a-H_f of DB24C8 and H_{14} at the MTA station (Figure S43). These results confirm that the DB24C8 unit of unstable cis-18 have moved from the AC region to the MTA station after illumination. After heating the solution of unstable cis-18 at 60 °C 24 h in the dark, the molecular motor underwent a THI process from an unstable cis state to a stable cis state. The observed ¹H NMR signals were consistent with that of an independent sample of stable cis-18 (Figure S41c,d). The above results proved that in principle, the entire four-step cycle of [1]rotaxane stable *cis*-18 can be successfully executed.

UV-Vis Absorption Spectroscopy. The photochemical and thermal isomerization processes of the [1]rotaxane cis-18H⁺ were furthermore studied by UV-vis absorption spectroscopy in CH₂Cl₂. After adding DBU to the system, a slight red shift in the maximum absorption was observed (Figure 7a), indicating a geometrical change of the molecular motor because of the movement of rotaxane from BAA station to MTA station. This result provides an explanation for the chemical shift of proton H₂₇ after the deprotonation of cis-18H⁺ (Figure 4a,b). Upon UV irradiation at 10 °C, the maximum absorption gradually shifted from 382 to 365 nm, which was consistent with increased strain at the central double bond, and hence the generation of a higher-energy unstable isomer. Allowing the solutions to heat to 60 °C for 24 h resulted in a red-shift, which was consistent with thermal isomerization to the corresponding stable isomers. Similar photochemical and thermal isomerization behavior of stable trans-18 was detected using UV-vis analysis (Figure 7b). The dynamic process was consistent with that observed for stable cis-18 (vide supra), and in agreement with the photochemical properties of related molecular motors reported earlier by our

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Figure 8. Energy-minimized structures of [1]rotaxane stable cis-18 at four-step cycle optimized using DFT at the RB3LYP/6-311+G (d) level.

group.^{11a,12b,14b} The results showed that the rotational motion of the molecular motor unit was not restrained in the mechanically interlocked [1]rotaxane, even in the presence of the rotaxane host-guest interaction.

DFT Calculations. To get a more detailed insight into the entire transmission mechanism of [1]rotaxane stable cis-18 and support the proposed structural changes, the energy optimization states of stable cis-18 in the four-step cycle were calculated by density functional theory (DFT), and all optimizations were performed on the RB3LYP/6-311+G (d) level of theory. The optimized geometries are shown in the Figure 8. The results indicate that the DB24C8 ring of stable cis-18 is located on the near MTA station instead of just above it, which confirm the peculiar changes of proton H₂₇ after the protonation of cis-18H⁺ (Figure 5b) because movement of the DB24C8 ring to the MTA site would distort the motor molecules. Upon UV irradiation, the stable cis-18 transformed into unstable trans-18, the optimized geometry showed that the DB24C8 ring is located on the AC region, which is consistent with the aforementioned ¹H NMR analysis (Figure 5). Meanwhile the DB24C8 ring has moved about 14.4 Å compared with the initial position. In this photoisomerization process, the electronic energies increase dramatically (the relative energy ΔE_1 = +22.8 kcal/mol) because of the crowded equatorial orientation of the methyl group in a trans configuration. After heating, the unstable trans-18 transforms into stable trans-18, the molecular motor undergoes a THI, leading to a slightly changed position of the DB24C8 ring. Simultaneously, the electronic energies decreased because of the recovery of the axial orientation (the relative energy ΔE_2 = -7.6 kcal/mol). In the next stage of the process, stable *trans*-18 isomerizes into unstable cis-18 upon the UV irradiation, and the DB24C8 ring returns back to the MTA site. Meanwhile, the axial orientation of methyl group transforms into an equatorial orientation, which should be accompanied by an increase in energy, but the energy dramatically decreased (ΔE_3 = -6.7 kcal/mol). This is a reasonable result in view of the formation of the cis isomer with more relaxed molecular geometry. After the final THI process, unstable cis-18 converted into stable cis-18, and the energy further decreased $(\Delta E_4 = -8.5 \text{ kcal/mol})$ because of the more stable axial orientation of the methyl group. The detailed data of the fourstep state were also summarized in Tables S1-S8 and Figures S44-S47. It can be concluded that the variation in energy and stability of the isomers in the light-driven process of stable cis-18 was not only dependent on the change in orientation of methyl,20a but is also governed by the entire molecular geometry. Compared to the cis isomers, trans isomers are more distorted resulting in higher energy, which may be another reason for the decrease in ratio of unstable trans isomers and stable cis isomers in the PSS state during the photoisomerization.

CONCLUSIONS

In summary, we have demonstrated the design, characterization and operation of a composite motor-[1]rotaxane contained a light-driven molecular motor and a bistable rotaxane. Detailed NMR experiments and molecular dynamics simulation confirmed that the molecular motor can perform directional rotation, even being integrated with an interlocked [1]rotaxane with considerable noncovalent interactions between macrocycle DB24C8 and MTA recognition sites being present. Demonstrating the complete four-step cycle using interrupted light and heat stimuli, it is evident that unidirectional rotary motion can be transmitted successfully into translational motion. These studies provide deeper insight into the operational capabilities of the second-generation molecular motors in more complex mechanical molecular systems and an important step on our route to explore molecular machines that can perform more complex mechanical tasks.

EXPERIMENTAL SECTION

General Methods and Details. Chemicals were purchased from Acros, Aldrich, TCI, Adamas, or Merck and used as received unless otherwise stated. Solvents were reagent grade, which were dried and distilled prior to use according to standard procedures. All reactions were carried out under an atmosphere of dry argon unless otherwise stated. NMR experiments (1H NMR, 13C NMR, 1H-1H COSY, HMQC, HMBC, and 2D Roesy) were measured on a Brüker AVANCE III 400, AVANCE III 500 or Ascend 600 spectrometer. High-resolution mass spectra (HR-MS) were tested on a LCT Premier XE mass spectrometer by using ESI analyzed by time-of-flight (TOF). The UV-vis absorption spectra and fluorescence spectra were also recorded on a Varian Cary 100 spectrometer and a Varian Cary Eclipse (1 cm quartz cell used), respectively. Photo-driven experiments were carried out by a PerfectLight PL-LED 100 with a constant wavelength of 365 nm. The X-ray diffraction intensity data were collected at 273 or 200 K on a Rigaku RAXIS RAPID IP imaging plate system with Mo K α radiation ($\lambda = 0.71073$ Å). The CIF files for the crystallographic data have been deposited in the Cambridge Crystallographic Data Centre, and the CCDC number is 1882649.

Syntheses of Molecular Motor 8 and Compound 12. 3-Hydroxy-9H-thioxanthen-9-one (2). To a solution of compound 1^{23} (20 g, 82.6 mmol) in toluene was added anhydrous aluminum trichloride (22 g, 165.2 mmol) slowly. Then the mixture was stirred

under reflux with argon atmosphere for 8 h. After the reaction was over, concentrated hydrochloric acid (40 mL) and ice water (60 mL) was added carefully and slowly to quench the reaction. The more ice water (250 mL) was added to the reaction solution, the more mass of white solid was precipitated. The precipitate was filtered under reduced pressure, washing the filter cake with toluene and water, then the white solid was dried under vacuum for 16 h to afford compound 2 (18.8 g, 90%) as a white powder. ¹H NMR (400 MHz, DMSO, 298 K): δ (ppm) 10.93 (s, 1H), 8.42 (d, *J* = 8 Hz, 1H), 8.34 (d, *J* = 8.8 Hz, 1H), 7.72 (m, 2H), 7.54 (t, *J* = 6.4 Hz, 1H), 7.06 (d, *J* = 2 Hz, 1H), 7.01 (dd, *J* = 8.8, 2 Hz, 1H). ¹³C{¹H} NMR (100 MHz, DMSO, 298 K): δ (ppm) 178.1, 162.0, 139.2, 136.6, 132.9, 132.2, 129.3, 129.0, 127.01, 126.7, 121.4, 116.8, 110.8. HR-MS (ESI-TOF) (*m*/*z*): [M - H]⁻ calcd for C₁₃H₇O₂S, 227.0172; found, 227.0177.

9-Oxo-9H-thioxanthen-3-yl Trifluoromethanesulfonate (3). To a solution of compound 2 (3 g, 13.1 mmol) in anhydrous dichloromethane was added anhydrous pyridine (10.5 mL, 131.0 mmol), then trifluoromethanesulfonic anhydride (5.6 g, 19.8 mmol) was added dropwise under ice bath. After addition, the mixture was stirred at room temperature with argon atmosphere for 16 h. Then the solvent was removed, and the residue was dissolved in dichloromethane (150 mL). The solution was washed with 1 N HCl aqueous solution (100 mL) and brine (100 mL), and then the organic layer was dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, PE/EA = 100:1) to afford compound 2 (4.7 g, 95%) as a white solid. ¹H NMR (400 MHz, $CDCl_3$, 298 K): δ (ppm) 8.71 (d, J = 9.2 Hz, 1H), 8.61 (dd, J = 8.0Hz, 1.2, 1H), 7.68 (m, 1H), 7.60 (dd, J = 8.0, 0.8 Hz, 1H), 7.54 (m, 1H), 7.52 (d, J = 2.4 Hz, 1H), 7.37 (dd, J = 8.8, 2.4 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 178.8, 151.5, 139.7, 136.5, 129.9, 126.8, 126.1, 119.4, 118.7 (q, J = 319 Hz), 118.3. HR-MS (ESI-TOF) (m/z): $[M + H]^+$ calcd for $C_{14}H_8F_3O_4S_2$, 360.9811; found, 360.9801.

9-Thioxo-9H-thioxanthen-3-yl Trifluoromethanesulfonate (4). To a solution of compound 2 (3 g, 13.1 mmol) in anhydrous dichloromethane was added anhydrous pyridine (10.5 mL, 131.0 mmol), then trifluoromethanesulfonic anhydride (5.6 g, 19.8 mmol) was added dropwise under ice bath. After addition, the mixture was stirred at room temperature with argon atmosphere for 16 h. Then the solvent was removed, and the residue was dissolved in dichloromethane (150 mL). The solution was washed with 1 N HCl aqueous solution (100 mL) and brine (100 mL), and then the organic layer was dried over Na2SO4 and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, PE/EA = 100:1) to afford compound 2 (4.7 g, 95%) as a white solid. ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 8.71 (d, J = 9.2 Hz, 1H), 8.61 (dd, J = 8.0 Hz, 1.2, 1H), 7.68 (m, 1H), 7.60 (dd, J = 8.0, 0.8 Hz, 1H), 7.54 (m, 1H), 7.52 (d, J = 2.4 Hz, 1H), 7.37 (dd, J = 8.8, 2.4 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 178.8, 151.5, 139.7, 136.5, 129.9, 126.8, 126.1, 119.4, 118.7 (q, J = 319 Hz), 118.3. HR-MS (ESI-TOF) (m/z): $[M + H]^+$ calcd for $C_{14}H_8F_3O_4S_2$ 360.9811; found, 360.9801.

(Z,E)-9-(8-Bromo-2-methyl-2,3-dihydro-1H-benzo[f]thiochromen-1-ylidene)-9H-thioxanthen-3-yl Trifluoromethanesulfonate (7). Compound 5^{24} (500 mg, 1.6 mmol) was dissolved in DMF and cooled under an ice bath, then iodosobenzene diacetate (509 mg, 1.6 mmol) was added slowly, the mixture was stirred for 5 min, compound 4 (588 mg, 1.6 mmol) was added. After that, the reaction solution was allowed to warm to room temperature and stirred for another 2 h. Then the solvent was removed under reduced pressure. The crude episulfide 6 was obtained as sticky liquid and used directly for the next step without further purification. The above crude episulfide 6 was dissolved in toluene, triphenylphosphine (839 mg, 3.2 mmol) was added, and then the mixture was heated under reflux for overnight. After that, the solvent was removed under reduced pressure, the residue was purified by column chromatography $(SiO_2, PE/DCM = 20:1)$ to afford compound 7 (650 mg, 64%) as yellow powder, which was identified by ¹H NMR as a 44:56 mixture of cis/trans isomers. The isomers were used in the next step without further separation. ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm)

7.72 (dd, J = 8, 2 Hz, 1H), 7.63 (d, J = 8.4 Hz, 1H), 7.58 (m, 1H), 7.52 (dd, J = 8.4, 2.4 Hz, 1H), 7.35–7.43 (m, 2H), 7.32 (d, J = 8.8 Hz, 1H), 7.27 (m, 1H), 7.08 (m, 1H), 6.83 (td, J = 7.6, 1.2 Hz, 1H), 6.50 (td, J = 7.6, 1.2 Hz, 1H), 6.37 (m, 1H), 4.04 (m, 1H), 3.69 (m, 1H), 3.10 (dd, J = 11.6, 2.8 Hz, 1H), 0.79 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 147.5, 139.1, 137.5, 136.3, 136.0, 133.9, 133.7, 133.2, 132.6, 131.4, 130.8, 130.3, 129.7, 129.2, 129.0, 127.7, 126.8, 126.2, 125.8, 125.5, 120.6, 119.2, 118.8 (q, J = 305 Hz), 118.5, 36.9, 32.3, 19.1. HR-MS (ESI-TOF) (m/z): [M + Na]⁺ calcd for C₂₈H₁₈BrF₃O₃S₃Na, 656.9446; found, 656.9448.

(Z,E)-9-(8-Bromo-2-methyl-2,3-dihydro-1H-benzo[f]thiochromen-1-ylidene)-9H-thioxanthen-3-ol (cis-8, trans-8). To a solution of compound 7 (500 mg, 0.78 mmol) in dioxane was added tetraethylammonium hydroxide (229 mg, 1.56 mmol) aqueous solution (25%). The reaction mixture was stirred at room temperature for 2 h. Then the solvent was removed under reduced pressure, the residue was acidified with hydrochloric acid solution (1 N), and extracted with DCM (3×20 mL). The combined organic layer was washed with water $(2 \times 20 \text{ mL})$, brine (15 mL), and then the organic layer was dried over Na2SO4 and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, PE/EA = 50:1) to afford cis-8 (194 mg, yield 49%) and trans-7 (150 mg, yield 38%) as a white solid. cis-8: ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.71 (d, J = 2 Hz, 1H), 7.60 (dd, J = 8.8 Hz, 1H), 7.56 (dd, J = 7.6, 1.2 Hz, 1H), 7.47 (t, J = 8.8 Hz, 1H), 7.37 (d, J = 8.8 Hz, 1H), 7.35 (td, J = 7.6, 1.2 Hz, 1H), 7.29 (dd, J = 7.6, 1.6 Hz, 1H), 6.25 (d, J = 8.4 Hz, 1H), 5.92 (dd, J = 8.4, 2.4 Hz, 1H), 4.09 (m, 1H), 3.72 (m, 1H), 3.10 (dd, J = 11.2, 2.8 Hz, 1H), 0.73 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 154.8, 135.8, 135.7, 135.3, 134.9, 134.7, 132.2, 131.0, 0.9, 130.8, 129.6, 129.5, 129.2, 128.9, 128.2, 127.2, 127.2, 126.6, 126.3, 126.2, 126.1, 126.1, 112.8, 112.5, 36.4, 30.9, 22.0, 17.6, 13.0. HR-MS (ESI-TOF) (*m*/*z*): [M – H]⁻ calcd for C₂₇H₁₈BrOS₂, 500.9988; found, 500.9997. trans-8: ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.69 (d, J = 2 Hz, 1H), 7.48 (d, J = 8.4 Hz, 1H), 7.44 (d, J = 8.4 Hz, 1H), 7.40 (d, J = 6.8 Hz, 1H), 7.37 (d, J = 6.8 Hz, 1H), 7.29 (dd, J = 8, 0.8 Hz, 1H), 7.10 (d, J = 2.4 Hz, 1H), 7.04 (dd, J = 9.2, 2.4 Hz, 1H), 6.83 (dd, J = 8.4, 2.4 Hz, 1H), 6.78 (td, J = 7.6, 1.2 Hz, 1H), 6.44 (td, J = 7.6, 1.2 Hz, 1H), 6.37 (dd, J = 7.6, 1.2 Hz, 1H), 5.02 (s, 1H), 4.09 (m, 1H), 3.70 (m, 1H), 3.10 (dd, J = 11.2, 2.8 Hz, 1H), 0.76 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 178.9, 154.4, 138.4, 137.5, 135.5, 135.2, 134.8, 133.9, 132.1, 130.9, 132.4, 131.5, 129.5, 129.0, 128.5, 127.5, 126.6, 126.4, 126.2, 126.0, 125.5, 124.5, 118.3, 114.4, 113.7, 37.2, 29.7, 19.2. HR-MS (ESI-TOF) (m/ z): $[M - H]^-$ calcd for C₂₇H₁₈BrOS₂, 500.9988; found; 500.9998.

tert-Butyl(5-((12-azidododecyl)amino)-5-oxopentyl)(3,5-di-tert*butylbenzyl)carbamate (11).* To a solution of compound 9^{25} (2.0 g, 4.7 mmol) in anhydrous dichloromethane was added compound 10^{26} (1.2 g, 4.7 mmol), a catalytic amount of DMAP, then EDCI (1.8 g, 9.5 mmol) was added dropwise under ice bath. After addition, the mixture was stirred at room temperature with argon atmosphere for 16 h. Then the solution was extracted with DCM $(3 \times 100 \text{ mL})$ and brine (100 mL), the organic layer was dried over Na2SO4 and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, PE/EA = 100:1) to afford compound 11 (2.4 g, 80%) as a transparent oil. ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.31 (s, 1H), 7.05 (s, 2H), 4.37 (m, 2H), 3.22 (m, 4H), 2.17 (m, 2H), 1.75 (m, 2H), 1.53–1.66 (m, 8H), 1.39–1.53 (m, 9H), 1.29–1.33 (m, 18H), 1.20–1.29 (m, 16H). $^{13}C{^{1}H}$ NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 172.9, 155.9, 150.8, 137.5, 121.6, 121.3, 121.0, 79.5, 51.4, 45.3, 39.4, 35.9, 34.7, 31.4, 29.6, 29.4, 29.4, 29.4, 29.2, 29.0, 28.7, 28.4, 27.9, 26.9, 26.8, 26.6, 22.9. HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{37}H_{65}N_5O_3Na$, 650.4980; found, 650.4986.

5-((12-Azidododecyl)amino)-N-(3,5-di-tert-butylbenzyl)-5-oxopentan-1-aminium Hexafluorophosphate(V) (12). To a solution ofcompound 11 (2.0 g, 3.2 mmol) in dichloromethane was added TFA(5.3 g, 47 mmol). After addition, the mixture was stirred at roomtemperature under argon atmosphere for 3 h, the solvent was

removed under reduced pressure. Then to a solution of the residue in methanol was added saturated aqueous solution of NH_4PF_6 (100 mL), a white precipitate appeared quickly, the solution turned into milky white, the mixture was stirred vigorously for 30 min. The solvent was removed, the residue was extracted with DCM and washed with distilled water. The organic layer was collected and dried over Na₂SO₄, after concentrating in vacuo, compound 12 was obtained (2.0 g, 95%) as a transparent oil without further purification. ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 7.46 (s, 1H), 7.35 (s, 1H), 7.34 (s, 1H), 4.13 (s, 2H), 3.24 (t, J = 6.8 Hz, 2H), 3.14 (m, 2H), 3.08 (t, J = 6.4 Hz, 2H), 2.24 (t, J = 6.0 Hz, 2H), 1.62–1.88 (m, 4H), 1.54-1.62 (m, 2H), 1.41-1.53 (m, 4H), 1.31 (s, 18H), 1.20-1.28 (m, 16H). $^{13}\text{C}\{^{1}\text{H}\}$ NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 174.2, 151.7, 130.4, 124.2, 123.4, 52.0, 51.0, 46.5, 39.3, 34.6, 32.9, 29.2, 29.2, 29.1, 28.9, 28.9, 28.7, 28.4, 26.5, 26.3, 24.9, 19.8. HR-MS (ESI-TOF) (m/z): $[M - PF_6^-]^+$ calcd for $C_{32}H_{58}N_5O$, 528.4636; found, 528,4651

Syntheses of Target Compound cis-18H⁺, trans-18H⁺ and Reference Compound cis-21H+. cis-14, trans-14. To a solution of cis-8 (450 mg, 0.79 mmol) in DMF was added a compound 13²⁷ (689 mg, 1.2 mmol), potassium carbonate (331 mg, 2.4 mmol), and tetrakis (triphenylphosphine) palladium (91 mg, 0.079 mmol). After addition, the mixture was stirred at reflux under argon atmosphere for 8 h. The solvent was removed under reduced pressure, and then the residue was acidified with hydrochloric acid solution (1 N) and extracted with DCM (3 \times 20 mL). The combined organic layer was washed with water $(2 \times 20 \text{ mL})$ and brine (15 mL), and then the organic layer was dried over Na2SO4 and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, DCM/ methanol = 100:1) to afford cis-14 as a white solid (480 mg, yield 70%). ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.66 (d, J = 1.2 Hz, 1H), 7.61 (d, J = 7.6 Hz, 2H), 7.56 (d, J = 7.6 Hz, 2H), 7.37 (d, J = 8.4 Hz, 1H), 7.32 (td, J = 7.6, 0.8 Hz, 1H), 7.22 (m, 2H), 7.10 (m, 2H), 6.80–6.93 (m, 6H), 6.22 (d, J = 8.4 Hz, 1H), 5.93 (dd, J = 8.4, 2.0 Hz, 1H), 4.18 (m, 2H), 4.04-4.15 (m, 7H), 3.60-3.91 (m, 17H), 3.10 (dd, J = 11.2, 2.4 Hz, 1H), 0.78 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 154.9, 152.0, 148.9, 148.8, 148.6, 136.6, 136.4, 136.0, 135.3, 135.2, 134.6, 132.3, 131.7, 131.4, 130.1, 129.8, 127.8, 127.5, 127.4, 126.0, 126.0, 125.2, 124.9, 121.7, 120.3, 114.3, 113.6, 113.1, 70.5, 70.4, 69.5, 69.4, 69.3, 37.3, 29.7, 19.2. HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{51}H_{50}O_9S_2Na$, 893.2788; found, 893.2818. trans-14 was prepared according to the same procedure as above. trans-14: ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm): 7.59 (d, J = 1.6 Hz, 1H), 7.53 (d, J = 8.8 Hz, 1H), 7.48 (d, *J* = 8.4 Hz, 1H), 7.33 (d, *J* = 8.4 Hz, 1H), 7.30 (d, *J* = 8.4 Hz, 1H), 7.12 (dd, J = 8.8, 2.0 Hz, 2H), 7.07 (d, J = 2.4 Hz, 1H), 7.00 (s, 2H), 6.75-6.90 (m, 7H), 6.57 (s, 1H), 6.23-6.37 (m, 2H), 4.04-4.14 (m, 8H), 4.02 (m, 1H), 3.66–3.92 (m, 16H), 3.61 (m, 1H), 3.01 (dd, J = 11.2, 2.8 Hz, 2H), 0.69 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 162.8, 153.3, 148.9, 138.7, 137.2, 135.3, 134.0, 131.7, 129.0, 128.5, 127.8, 127.6, 126.4, 125.4, 125.0, 124.7, 121.5, 114.4, 114.0, 113.1, 71.3, 69.9, 69.6, 69.36, 37.26, 36.7, 19.0. HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{51}H_{50}O_9S_2Na_7$ 893.2788; found, 893.2794.

cis-16, trans-16. To a solution of cis-14 (200 mg, 0.23 mmol) in CH₃CN was added compound 15²⁸ (82 mg, 0.34 mmol) and potassium carbonate (95 mg, 0.69 mmol). After addition, the mixture was stirred at reflux under argon atmosphere for 8 h. After the solvent was removed under reduced pressure, the residue was extracted with DCM $(3 \times 50 \text{ mL})$ and washed with brine (50 mL), water (50 mL), and then the organic layer was dried over Na2SO4 and concentrated in vacuo. The crude product was purified by column chromatography $(SiO_2, DCM/methanol = 100:1)$ to afford *cis*-16 as a white powder (183 mg, yield 85%). trans-15 was prepared according to the same procedure as above. cis-16: ¹H NMR (400 MHz, CD₃Cl, 298 K): δ (ppm) 7.69 (d, J = 1.6 Hz, 1H), 7.54–7.65 (m, 4H), 7.38 (d, J = 8.4 Hz, 1H), 7.35 (td, J = 7.6, 1.2 Hz, 1H), 7.29 (dd, J = 7.6, 1.2 Hz, 1H), 7.24 (dd, J = 9.2, 1.2 Hz, 1H), 7.10 (m, 2H), 6.86 (d, J = 2.8 Hz, 1H), 6.86-6.91 (m, 4H), 6.92 (d, J = 8.8 Hz, 1H), 6.34 (d, J = 8.8 Hz, 1H), 6.00 (dd, J = 8.8, 2.8 Hz, 1H), 4.24 (t, J = 4.4 Hz, 2H), 4.134.21 (m, 6H), 4.11 (m, 1H), 3.89-3.98 (m, 8H), 3.80-3.88 (m, 8H), 3.69–3.79 (m, 3H), 3.11 (dd, J = 11.2, 2.8 Hz, 1H), 2.14 (td, J = 7.2, 2.8 Hz, 2H), 1.85 (t, J = 2.8 Hz, 1H), 1.72 (t, J = 2.4 Hz, 2H), 0.78 (d, J = 6.8 Hz, 3H). HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{56}H_{56}O_{9}S_{2}Na$, 959.3258; found, 959.3263. ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 157.4, 149.0, 148.8, 148.4, 138.5, 137.4, 136.2, 135.6, 134.5, 134.2, 133.8, 132.0, 131.6, 131.2, 129.8, 128.9, 128.6, 128.4, 127.6, 127.5, 127.4, 126.3, 126.6, 126.0, 125.8, 125.4, 125.0, 124.8, 124.3, 124.6, 121.4, 119.9, 114.1, 114.0, 113.1, 112.9, 83.2, 71.2, 69.8, 69.5, 69.4, 69.3, 69.0, 66.3, 37.0, 31.9, 29.6, 27.9, 19.1, 15.0. trans-16: ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.67 (d, J = 1.6 Hz, 1H), 7.62 (d, J = 8.4 Hz, 1H), 7.56 (d, J = 8.8 Hz, 1H), 7.50 (d, J = 8.4 Hz, 1H), 7.38 (d, J = 8.4 Hz, 1H), 7.27 (d, J = 7.6 Hz, 1H), 7.21 (dd, J = 8.8, 1.6 Hz, 1H), 7.18 (d, J = 6.8 Hz, 1H), 7.11 (m, 2H), 6.83-6.95 (m, 6H), 6.71 (m, 1H), 6.38 (m, 2H), 4.23 (m, 2H), 3.80-3.88 (m, 6H), 3.73 (m, 1H), 3.11 (dd, J = 11.2, 2.8 Hz, 1H), 2.44 (td, J = 7.2, 2.8 Hz, 2H), 2.04 (m, 2H), 2.00 (t, J = 2.8 Hz, 1H), 1.72 (m, 2H), 0.80 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 157.4, 149.0, 148.8, 148.4, 138.5, 137.4, 136.2, 135.6, 134.5, 133.8, 132.0, 131.6, 131.2, 129.8, 128.9, 128.6, 128.3, 127.5, 127.4, 126.3, 126.0, 125.8, 125.6, 125.0, 124.8, 124.7, 119.9, 114.1, 114.0, 113.1, 112.9, 83.3, 71.2, 69.5, 69.4, 69.3, 69.0, 66.3, 37.1, 31.9, 29.6, 28.0, 18.9, 15.1. HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{56}H_{56}O_9S_2Na$, 959.3258; found, 959.3261.

cis-17H⁺, trans-17H⁺. To a solution of cis-16 (100 mg, 0.106 mmol) in DCM (10 mL) was added compound 12 (72 mg, 0.106 mmol), the mixture was stirred at room temperature for 30 min. Then the solution was diluted to 250 mL with DCM, and Cu(CH₃CN)₄PF₆ (40 mg, 0.106 mmol) was added. After that, the reaction solution was stirred at room temperature under argon atmosphere for 24 h. The solution was extracted with DCM $(3 \times 50 \text{ mL})$ and distilled water (50 mL), then the organic layer was dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography $(SiO_2, DCM/MeOH = 100:1)$ to afford compound *cis*-17H⁺ (94 mg, 30%) as a white solid. *trans*-17H⁺ was prepared according to the same procedure as mentioned above. cis-17H+: 1H NMR (400 MHz, CD_3CN , 298 K): δ (ppm) 7.80 (s, 1H), 7.69 (d, J = 8.4 Hz, 1H), 7.65 (d, J = 7.6 Hz, 1H), 7.64 (dd, J = 8.4, 1.2 Hz, 1H), 7.62 (d, J = 8.4 Hz, 1H), 7.60 (d, J = 8.4 Hz, 1H), 7.37–7.50 (m, 4H), 7.31–7.37 (m, 2H), 7.23-7.27 (m, 4H), 7.04-7.13 (m, 1H), 6.99-7.13 (m, 1H), 6.81–6.97 (m, 6H), 6.28 (dd, J = 8.4, 1.6 Hz, 1H), 5.96 (dd, J = 8.4, 2.0 Hz, 1H), 4.66 (m, 2H), 4.24 (m, 2H), 4.02-4.21 (m, 9H), 3.92-4.01 (m, 1H), 3.76-3.91 (m, 4H), 3.69-3.76 (m, 4H), 3.67 (m, 2H), 3.55-3.67 (m, 4H), 3.54-3.40 (m, 4H), 3.34-3.20 (m, 4H), 3.11 (dd, J = 11.2, 2.8 Hz, 1H), 3.05 (m, 2H), 2.56 (m, 2H), 1.88 (m, 2H), 1.78 (m, 2H), 1.50 (m, 2H), 1.17-1.23 (m, 16H), 1.09-1.17 (m, 18H), 0.69 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 171.7, 157.0, 151.1, 147.8, 147.7, 147.4, 147.0, 136.1, 135.8, 135.7, 135.7, 135.2, 133.8, 131.8, 131.6, 130.8, 129.7, 129.6, 127.6, 127.6, 127.5, 126.9, 126.6, 125.8, 124.9, 124.7, 124.6, 124.1, 123.9, 123.3, 121.3, 119.6, 119.6, 118.7, 112.9, 112.5, 112.0, 111.9, 111.2, 111.1, 70.4, 70.2, 70.0, 69.2, 68.2, 68.0, 67.1, 52.5, 49.5, 49.5, 48.5, 38.6, 36.6, 34.7, 34.3, 33.1, 31.6, 30.8, 29.6, 29.5, 29.4, 29.0, 28.5, 28.4, 28.3, 27.9, 26.7, 26.1, 25.7, 25.5, 24.6, 22.3, 21.5, 18.0. HR-MS (ESI-TOF) (m/z): $[M - PF_6]^+$ calcd for C₈₈H₁₁₄N₅O₁₀S₂, 1464.8002; found, 1464.8013. *cis*-17H⁺: ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 7.87 (s, 1H), 7.70 (d, J = 8.8 Hz, 1H), 7.62 (m, 2H), 7.53 (d, J = 8.4 Hz, 1H), 7.45 (s, 1H), 7.44 (m, 1H), 7.40 (m, 1H), 7.39 (d, J = 1.2 Hz, 1H), 7.30 (d, J = 8.0 Hz, 1H), 7.27 (m, 1H), 7.25 (d, J = 1.2 Hz, 1H), 7.22 (dd, J = 8.4, 2.4 Hz, 1H), 7.18 (s, 1H), 7.13 (dd, J = 8.4, 1.6 Hz, 1H), 7.06 (s, 1H), 7.01 (d, J = 9.6 Hz, 1H), 6.85–6.99 (m, 5H), 6.75 (m, 1H), 6.45 (m, 2H), 4.67 (t, J = 13.2 Hz, 1H), 4.42 (m, 2H), 4.29 (m, 2H), 4.13–4.22 (m, 4H), 4.10 (m, 1H), 3.94-4.09 (m, 4H), 3.90 (m, 1H), 3.67-3.87 (m, 10H), 3.40–3.67 (m, 4H), 3.32 (m, 2H), 3.20 (m, 1H), 3.14 (d, J = 11.2 Hz, 2H), 2.82 (m, 2H), 2.00 (m, 2H), 1.77 (m, 2H), 1.64 (m, 2H), 1.51 (m, 2H), 1.37 (m, 2H), 1.25–1.29 (m, 16H), 1.23 (d, J = 2.0 Hz, 18H), 0.67 (d, J = 6.4 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 171.9, 164.0, 158.4, 151.9, 148.3, 147.6,

found, 1464.7997. [1]Rotaxane cis-18H⁺, [1]Rotaxane trans-18H⁺. To a solution of cis-17H⁺ (90 mg, 0.056 mmol) in DCM (2 mL) was added excessive iodomethane (2 mL), the solution was kept in a pressure tube and sealed with a screw cap. Then the mixture was stirred at 40 °C for 24 h. After that, the solvent was removed under reduced pressure. Then to a solution of the residue in methanol was added saturated aqueous solution of NH₄PF₆ (100 mL), a white precipitate appeared quickly, the solution turned into milky white, the mixture was stirred vigorously for 30 min. Then the solvent was removed, the residue was extracted with DCM and washed with distilled water. The organic layer was collected and dried over Na2SO4, after concentrating in vacuo. The crude product was purified by column chromatography $(SiO_2, DCM/MeOH = 100:1)$ to afford compound *cis*-18H⁺ (80 mg, 80%) as a white solid. trans-18H⁺ was prepared according to the same procedure as above. cis-18H⁺: ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 8.03 (d, J = 8.8 Hz, 1H), 7.86 (s, 1H), 7.72 (d, J = 8.8 Hz, 1H), 7.62-7.70 (m, 4H), 7.38-7.49 (m, 4H), 7.30-7.38 (m, 3H), 7.20-7.30 (m, 4H), 7.07-7.10 (m, 2H), 6.84-7.02 (m, 5H), 6.38 (d, I = 8.8 Hz, 1H), 6.18 (m, 1H), 6.02 (dd, I = 8.8, 2.8 Hz, 1H), 4.67 (m, 2H), 4.41 (m, 2H), 3.99-4.31 (m, 9H), 3.91 (s, 3H), 3.58-3.88 (m, 14H), 3.37–3.58 (m, 4H), 3.13 (dd, J = 11.2, 2.4 Hz, 1H), 3.01 (m, 2H), 2.73 (m, 2H), 2.04 (m, 2H), 1.87 (m, 2H), 1.26 (m, 2H), 1.14 (m, 2H), 0.68 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CD₂Cl₂, 298 K): δ (ppm) 172.0, 156.4, 151.3, 147.9, 147.6, 147.1, 145.7, 144.2, 138.8, 136.2, 135.7, 135.6, 135.2, 134.4, 132.7, 131.9, 131.8, 131.6, 130.9, 130.9, 130.0, 129.7, 128.4, 127.7, 127.5, 127.4, 126.7, 126.3, 125.9, 125.0, 124.6, 124.4, 124.1, 123.8, 123.3, 121.6, 120.1, 118.8, 118.6, 113.5, 113.2, 112.8, 112.6, 111.8, 111.6, 111.3, 70.5, 70.1, 68.7, 68.3, 68.1, 66.4, 51.4, 48.8, 45.3, 39.2, 37.1, 35.7, 35.1, 34.6, 33.6, 31.8, 31.0, 29.8, 29.6, 29.4, 29.3, 28.9, 28.7, 28.3, 28.0, 27.1, 26.5, 26.1, 25.9, 25.4, 24.8, 22.6, 20.2, 18.6. HR-MS (ESI-TOF) (m/z): $[M - PF_6^-]^+$ calcd for $C_{89}H_{117}F_6N_5O_{10}PS_2$, 1624.7878; found, 1624.7881, $[M - 2PF_6^{-}]^{2+}$ calcd for $C_{89}H_{117}N_5O_{10}S_{27}$ 739.9115; found, 739.9116. trans-18H+: 1H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 8.14 (d, J = 8.8 Hz, 1H), 7.87 (s, 1H), 7.72 (d, J = 8.8 Hz, 1H), 7.57 (t, J = 8.4 Hz, 2H), 7.48 (m, 1H), 7.44 (d, J = 4.0 Hz, 1H), 7.42 (d, J = 2.4 Hz, 1H), 7.40 (m, 1H), 7.35 (m, 1H), 7.31 (d, J = 7.6 Hz, 1H), 7.28 (d, J = 2.0 Hz, 1H), 7.26 (m, 2H), 7.24 (d, J = 2.4 Hz, 1H), 7.21 (m, 1H), 7.15 (d, J = 8.8, 2.4 Hz, 1H), 7.03 (d, J = 2.8 Hz, 1H), 7.02 (d, J = 2.8 Hz, 1H), 6.97 (m, 2H), 6.88-6.94 (m, 3H), 6.78 (m, 1H), 6.46 (m, 1H), 4.67 (m, 2H), 4.31-4.51 (m, 4H), 4.21 (m, 1H), 4.17 (d, J = 3.6 Hz, 2H), 4.15 (d, J = 3.6 Hz, 1H), 4.03-4.13 (m, 4H), 3.80-3.93 (m, 4H), 3.68-3.80 (m, 9H), 3.59-3.67 (m, 4H), 3.46 (m, 2H), 3.36 (m, 2H), 3.17 (m, 2H), 3.14 (m, 2H), 3.07 (m, 2H), 2.86 (m, 2H), 2.11 (m, 2H), 2.02 (m, 2H), 1.85 (m, 2H), 1.66 (m, 2H), 1.54 (m, 2H), 1.32-1.38 (m, 16H), 1.24 (s, 18H), 0.69 (d, J = 6.8 Hz, 3H). trans-18H⁺: ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 8.14 (d, J = 7.2 Hz, 1H), 7.87 (s, 1H), 7.72 (d, J = 8.8 Hz, 1H), 7.57 (t, J = 8.4 Hz, 2H), 7.47 (m, 1H), 7.44 (d, J = 4.4 Hz, 1H), 7.42 (d, J = 2.4 Hz, 1H), 7.40 (s, 1H), 7.35 (m, 1H), 7.31 (d, J = 7.6 Hz, 1H), 7.28 (m, 1H), 7.26 (m, 2H), 7.24 (d, J = 2.4 Hz, 1H), 7.21 (m, 1H), 7.15 (dd, J = 8.4, 2.4 Hz, 1H), 7.03 (d, J = 2.8 Hz, 1H), 7.02 (d, J = 2.8 Hz, 1H), 6.97 (m, 2H), 6.88-6.95 (m, 4H), 6.77 (m, 1H), 6.46 (m, 1H), 4.67 (m, 2H), 4.43 (m, 2H), 4.20–4.38 (m, 4H), 4.17 (d, J = 3.6 Hz, 2H), 4.15 (d, J = 3.6 Hz, 3H), 3.98-4.13 (m, 5H), 3.68-3.89 (m, 9H), 3.43-4.67 (m, 8H), 3.36 (m, 2H), 3.12-3.25 (m, 3H), 3.07 (m, 2H), 2.86 (m, 2H), 2.11 (m, 2H), 2.02 (m, 2H), 1.77-1.90 (m, 4H), 1.66 (m, 2H), 1.33-1.38 (m, 16H), 1.24 (s, 16H), 0.69 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CD₂Cl₂, 298 K): δ (ppm) 172.0, 156.4, 151.3, 147.9, 147.6, 147.3, 145.7, 144.2, 138.8, 136.2, 135.7, 135.6, 135.2, 134.4, 132.7, 131.9, 131.9, 131.8, 131.6, 130.9, 129.7, 128.4,

127.7, 127.5, 127.4, 126.7, 126.3, 125.9, 125.0, 124.6, 124.2, 123.8, 123.4, 121.6, 120.1, 118.7, 113.2, 112.7, 111.6, 111.3, 70.8, 70.5, 70.1, 68.7, 68.5, 68.3, 68.1, 66.4, 48.8, 39.2, 37.1, 35.7, 35.1, 34.6, 33.6, 31.8, 31.1, 31.0, 29.8, 29.4, 29.3, 28.9, 28.7, 28.3, 28.0, 27.1, 26.1, 25.9, 25.4, 22.6, 20.2, 18.6. HR-MS (ESI-TOF) (m/z): $[M - PF_6^{-}]^+$ calcd for $C_{89}H_{117}F_6N_5O_{10}PS_2$, 1624.7878; found, 1624.7866.

cis-19. To a solution of cis-15 (100 mg, 0.068 mmol) in DCM (10 mL) was added compound 11 (46 mg, 0.068 mmol), the mixture was stirred at room temperature for 30 min. Then the solution was diluted to 250 mL with DCM, and Cu(CH₃CN)₄PF₆ (25 mg, 0.068 mmol) was added. After that, the reaction solution was stirred at room temperature under argon atmosphere for 24 h. Then the solution was extracted with DCM $(3 \times 50 \text{ mL})$ and distilled water (50 mL), the organic layer was dried over Na₂SO₄ and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, DCM/ MeOH = 100:1) to afford compound cis-19 (102 mg, 60%) as a transparent oil. ¹H NMR (400 MHz, CDCl₃, 298 K): δ (ppm) 7.92 (s, 1H), 7.77 (d, J = 5.2 Hz, 1H), 7.64–7.75 (m, 3H), 7.45–7.54 (m, 2H), 7.37-7.45 (m, 3H), 7.35 (s, 1H), 7.27 (s, 1H), 7.21 (d, J = 6.4 Hz, 1H), 7.17 (m, 2H), 6.90–7.06 (m, 6H), 6.40 (s, 1H), 6.36 (d, J = 6.8 Hz, 1H), 6.02 (d, J = 6.8 Hz, 1H), 4.43 (m, 2H), 4.05-4.33 (m, 10H), 3.82-3.98 (m, 9H), 3.68-3.70 (m, 9H), 3.49 (m, 2H), 3.16 (m, 3H), 2.62 (t, J = 6.0 Hz, 2H), 2.13 (m, 2H), 1.85 (m, 2H), 1.47-1.57 (m, 10H), 1.36-1.38 (m, 18H), 1.33-1.36 (m, 9H), 1.20-1.32 (m, 16H), 0.77 (d, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 172.2, 157.0, 150.7, 148.9, 148.7, 148.4, 146.3, 138.2, 136.1, 135.9, 135.6, 135.3, 135.0, 133.3, 131.8, 131.6, 130.8, 130.7, 129.7, 129.5, 127.6, 127.5, 126.8, 126.5, 125.7, 124.7, 124.6, 124.5, 123.7, 121.4, 121.3, 121.1, 120.8, 119.5, 118.7, 116.6, 114.1, 114.0, 112.7, 112.0, 78.8, 70.5, 69.4, 69.3, 68.9, 68.8, 68.7, 67.0, 54.2, 49.6, 49.5, 38.7, 36.6, 35.5, 34.4, 31.6, 31.5, 30.4, 29.9, 29.3, 29.1, 28.9, 28.6, 28.3, 27.6, 26.5, 26.0, 22.8, 22.3, 21.4, 18.1, 13.3. HR-MS (ESI-TOF) (m/z): $[M + Na]^+$ calcd for $C_{93}H_{121}N_5O_{12}S_2Na$, 1586.8345; found, 1586.8336.

cis-20. To a solution of cis-19 (50 mg, 0.032 mmol) in DCM (2 mL) was added excessive iodomethane (2 mL), the solution was stay in a pressure tube and sealed with a screw cap. Then the mixture was stirred at 40 °C for 24 h. After that, the solvent was removed under reduced pressure. Then to a solution of the residue in methanol was added saturated aqueous solution of NH₄PF₆ (100 mL), a white precipitate appeared quickly, the solution turned into milky white, the mixture was stirred vigorously for 30 min. The solvent was removed, and the residue was extracted with DCM and washed with distilled water. The organic layer was collected and dried over Na2SO4, after concentrating in vacuo. The crude product was purified by column chromatography (SiO₂, DCM/MeOH = 100:1) to afford compound cis-20 (27 mg, 50%) as a white solid. ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 7.92 (s, 1H), 7.86 (d, J = 1.6 Hz, 1H), 7.70 (d, J =8.8 Hz, 1H), 7.65 (m, 2H), 7.60 (d, J = 8.8 Hz, 1H), 7.44 (d, J = 7.2 Hz, 1H), 7.40 (d, J = 8.4 Hz, 1H), 7.28–7.37 (m, 3H), 7.23 (td, J = 8.4, 1.6 Hz, 1H), 7.19 (s, 1H), 7.15 (dd, J = 8.4, 2.8 Hz, 1H), 7.09 (d, *J* = 1.2 Hz, 1H), 6.96 (d, *J* = 8.4 Hz, 1H), 6.87–6.93 (m, 4H), 6.31 (s, 1H), 6.29 (d, J = 8.8 Hz, 1H), 5.95 (dd, J = 8.4, 2.4 Hz, 1H), 4.35 (s, 2H), 4.32 (d, J = 7.2 Hz, 1H), 4.21 (m, 2H), 4.13 (m, 2H), 4.10 (m, 4H), 4.05 (m, 1H), 3.89 (s, 3H), 3.76-3.87 (m, 9H), 3.75 (m, 2H), 3.64–3.72 (m, 8H), 3.14 (m, 2H), 3.11 (dd, J = 11.2, 2.4 Hz, 1H), 3.06 (m, 2H), 2.03 (m, 4H), 1.86 (m, 2H), 1.79 (m, 2H), 1.53 (m, 2H), 1.38–1.50 (m, 10H), 1.28–1.33 (m, 18H), 1.27 (s, 9H), 1.21–1.25 (m, 16H), 0.72 (d, J = 6.8 Hz, 3H). $^{13}C{^{1}H}$ NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 159.8, 156.2, 150.5, 148.5, 148.4, 148.0, 143.6, 135.7, 135.6, 135.4, 135.0, 134.9, 133.1, 131.4, 131.0, 130.4, 130.4, 129.5, 129.5, 129.3, 127.4, 127.2, 127.2, 126.7, 126.4, 125.5, 124.5, 124.4, 124.3, 124.2, 123.5, 122.7, 121.1, 120.6, 119.2, 118.4, 113.8, 112.4, 112.3, 111.8, 78.6, 78.0, 77.6, 70.0, 70.0, 69.0, 69.0, 68.6, 68.4, 68.4, 66.2, 54.0, 53.2, 38.4, 36.8, 36.3, 35.2, 34.2, 31.3, 30.4, 30.3, 29.1, 29.0, 28.9, 28.7, 28.1, 27.4, 26.5, 26.3, 25.7, 25.2, 22.5, 22.1, 19.4, 17.7, 13.0. HR-MS (ESI-TOF) (m/z): $[M - PF_6^-]^+$ calcd for C₉₄H₁₂₄N₅O₁₂S₂, 1578.8682; found, 1578.8682.

 $cis-21H^+$. To a solution of compound cis-20 (100 mg, 0.064 mmol) in dichloromethane was added TFA (73 mg, 0.64 mmol). After

addition, the mixture was stirred at room temperature under argon atmosphere for 3 h, the solvent was removed under reduced pressure. Then to a solution of the residue in methanol was added saturated aqueous solution of NH₄PF₆ (5 mL), a white precipitate appeared quickly, the solution turned into milky white, the mixture was stirred vigorously for 30 min. The solvent was removed, and the residue was extracted with DCM and washed with distilled water. The organic layer was collected and dried over Na2SO4, after concentrating in vacuo, compound 21H⁺ was obtained (90 mg, 88%) as a white solid without further purification. ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm) 7.91 (s, 1H), 7.82 (d, J = 1.6 Hz, 1H), 7.66 (d, J = 8.8 Hz, 1H), 7.59 (m, 3H), 7.37 (m, 2H), 7.34 (m, 1H), 7.23-7.32 (m, 4H), 7.21 (dd, J = 8.4 Hz, 2.0 Hz, 1H), 7.16 (td, J = 8.8, 2.0 Hz, 1H), 7.07 (dd, J = 8.4, 2.4 Hz, 1H), 7.01 (s, 2H), 6.97 (m, 2H), 6.92 (m, 2H),6.82 (d, J = 2.4 Hz, 1H), 6.29 (d, J = 8.4 Hz, 1H), 6.24 (m, 1H), 5.93 (dd, J = 8.4, 2.4 Hz, 1H), 4.20-4.34 (m, 6H), 4.09-4.20 (m, 6H),4.03 (m, 1H), 3.84 (s, 3H), 3.64-3.78 (m, 11H), 3.52-3.62 (m, 8H), 3.08 (m, 3H), 2.99 (m, 2H), 2.68 (d, J = 7.2 Hz, 2H), 1.96 (m, 4H), 1.71 (m, 4H), 1.47 (m, 4H), 1.29–1.42 (m, 16H), 1.20 (s, 18H), 0.61 (d, J = 6.8 Hz, 1H). ¹³C{¹H} NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 173.7, 156.7, 151.4, 148.0, 147.9, 147.6, 135.8, 135.4, 135.2, 135.1, 135.0, 134.6, 131.6, 131.4, 130.6, 130.2, 129.4, 127.4, 127.2, 126.7, 126.3, 125.6, 124.6, 124.4, 123.9, 123.0, 122.5, 122.4, 120.7, 115.8, 115.5, 114.2, 112.5, 111.7, 68.5, 68.3, 68.2, 68.0, 67.9, 67.8, 67.8, 67.4, 67.3, 67.3, 66.7, 54.0, 51.7, 49.3, 46.3, 39.0, 36.4, 34.3, 31.3, 30.6, 30.3, 29.6, 29.0, 28.9, 28.9, 28.8, 28.8, 28.7, 28.7, 28.5, 28.3, 28.0, 26.3, 25.7, 24.7, 22.1, 21.2, 19.9, 17.8, 13.1. HR-MS (ESI-TOF) (m/z): $[M - 2PF_6^-]^+$ calcd for $C_{89}H_{117}N_5O_{10}S_2$, 740.4132; found, 740.4136.

Syntheses of Reference [2]Rotaxane 24H⁺ and 25H⁺. 1.3-Dimethoxy-5-(pent-4-yn-1-yloxy)benzene (22). To a solution of 3,5dimethoxyphenol (500 mg, 3.25 mmol) in CH₃CN was added compound 15 (774 mg, 3.25 mmol) and potassium carbonate (449 mg, 3.25 mmol). After addition, the mixture was stirred at reflux under argon atmosphere for 8 h. Then the solvent was removed under reduced pressure, the residue was extracted with DCM $(3 \times 100 \text{ mL})$ and washed with brine (100 mL), water (100 mL), then the organic layer was dried over Na2SO4, and concentrated in vacuo. The crude product was purified by column chromatography (SiO₂, PE) to afford 22 as a white powder (608 mg, yield 85%). ¹H NMR (400 MHz, $CDCl_{3}$, 298 K): δ (ppm) 6.08 (m, 3H), 4.03 (t, J = 6.0 Hz, 2H), 3.77 (s, 6H), 2.40 (td, J = 7.2, 2.8 Hz, 2H), 1.98 (m, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃, 298 K): δ (ppm) 161.4, 160.7, 93.3, 92.9, 83.4, 68.8, 66.1, 55.2, 28.0, 15.1. HR-MS (ESI-TOF) (m/z): $[M + H]^+$ calcd for C₁₃H₁₇O₃, 221.1172; found, 221.1155.

[2]Rotaxane 24H⁺. To a solution of 12 (100 mg, 0.148 mmol) in DCM (10 mL) was added compound 22 (33 mg, 0.148 mmol), compound 23^{29} (66 mg, 0.148 mmol) and Cu(CH₃CN)₄PF₆ (56 mg, 0.148 mmol). The mixture was stirred at room temperature under argon atmosphere for 24 h. Then the solution was extracted with DCM $(3 \times 50 \text{ mL})$ and distilled water (50 mL), the organic layer was dried over Na2SO4, and concentrated in vacuo. The crude product was purified by column chromatography (SiO2, DCM/MeOH = 100:1) to afford compound $24H^+$ (158 mg, 80%) as a white solid. ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ (ppm) 7.21 (s, 1H), 7.17 (s, 1H), 7.15 (s, 2H), 6.70-6.80 (m, 8H), 5.84-5.94 (m, 3H), 5.52 (t, J = 5.6 Hz, 1H), 4.49 (m, 2H), 4.12 (m, 2H), 4.08 (m, 4H), 3.93 (m, 4H), 3.77 (t, J = 6.0 Hz, 1H), 3.65 (m, 4H), 3.58 (m, 4H), 3.56 (s, 6H), 3.44 (m, 4H), 3.23 (m, 4H), 3.01 (m, 2H), 2.91 (m, 2H), 2.68 (t, J = 7.6 Hz, 2H), 1.67 (m, 4H), 1.14–1.34 (m, 8H), 1.06–1.10 (m, 16H), 1.05 (s, 18H). ${}^{13}C{}^{1}H$ NMR (100 MHz, CD₂Cl₂, 298 K): δ (ppm) 171.8, 161.6, 160.9, 151.3, 147.6, 146.9, 131.7, 124.2, 123.4, 121.7, 120.9, 112.8, 93.3, 92.8, 70.6, 70.1, 68.3, 67.0, 55.3, 50.1, 48.8, 39.4, 35.1, 34.7, 31.1, 30.3, 29.7, 29.6, 29.5, 29.5, 29.4, 29.3, 29.3, 28.9, 26.9, 26.4, 26.0, 22.4, 22.0. HR-MS (ESI-TOF) (m/z): [M - $PF_6^{-}]^+$ calcd for $C_{69}H_{106}N_5O_{12}$, 1196.7833; found, 1196.7847.

[2]Rotaxane **25H**⁺. To a solution of **24H**⁺ (50 mg, 0.037 mmol) in DCM (2 mL) was added excessive iodomethane (2 mL), the solution was stayed in a pressure tube and sealed with a screw cap. Then the mixture was stirred at 40 $^{\circ}$ C for 24 h. After that, the solvent was

removed under reduced pressure. Then to a solution of the residue in methanol (2 mL) was added saturated aqueous solution of NH₄PF₆ (2 mL), a white precipitate appeared quickly, the solution turned into milky white, the mixture was stirred vigorously for 30 min. The solvent was removed, and the residue was extracted with DCM (3 \times 50 mL) and washed with distilled water (50 mL). The organic layer was collected and dried over Na₂SO₄, after concentrating in vacuo. The crude product was purified by column chromatography (SiO₂, DCM/MeOH = 80:1) to afford compound $25H^+$ (41 mg, 50%) as a white solid. ¹H NMR (400 MHz, CD₃CN, 298 K): δ (ppm): 8.15 (s, 1H), 7.39 (s, 1H), 7.29 (d, J = 1.6 Hz, 2H), 7.25 (m, 2H), 7.03 (s, 2H), 6.85–6.98 (m, 8H), 6.11 (s, 1H), 6.10 (s, 1H), 6.06 (d, J = 2.0 Hz, 2H), 4.67 (m, 2H), 4.45 (t, J = 7.2 Hz, 2H), 4.13-4.23 (m, 4H), 4.10 (s, 3H), 4.08-4.10 (m, 2H), 3.99-4.08 (m, 4H), 3.78-3.87 (m, 4H), 3.73 (s, 6H), 3.67-3.72 (m, 4H), 3.59-3.66 (m, 4H), 3.51-3.45 (m, 4H), 3.24 (m, 2H), 3.03 (m, 2H), 2.98 (t, J = 7.2 Hz, 2H), 2.13 (m, 2H), 1.89 (m, 2H), 1.82 (t, J = 7.2 Hz, 2H), 1.46 (m, 2H), 1.34-1.46 (m, 4H), 1.21-1.30 (m, 16H), 1.18 (s, 18H). ¹³C{¹H} NMR (100 MHz, CD₃CN, 298 K): δ (ppm) 171.2, 161.4, 160.2, 160.2, 150.8, 147.2, 143.9, 131.6, 127.5, 123.8, 123.1, 121.1, 117.0, 112.3, 92.9, 92.6, 70.2, 69.7, 67.8, 65.9, 54.7, 54.0, 53.3, 52.3, 48.3, 38.5, 37.0, 34.4, 34.1, 30.3, 29.0, 29.0, 28.9, 28.7, 28.6, 28.2, 26.3, 25.9, 25.4, 25.3, 22.1, 21.9, 19.49. HR-MS (ESI-TOF) (m/z): [M -PF₆⁻]⁺ calcd for C₇₀H₁₀₉F₁₂N₅O₁₂P₂, 1356.7709; found, 1356.7701, $[M - 2PF_6^-]^{2+}$ calcd for $C_{70}H_{109}N_5O_{12}$, 605.9031; found, 605.9025.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.9b00783.

Details of synthetic route, ¹H, ¹³C NMR, ¹H–¹H COSY, HMQC, HMBC, 2D ROESY and HR-MS spectra, and DFT calculation methods (PDF)

Crystallographic data for compound cis-8 (CIF)

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Notes

The authors declare no competing financial interest.

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