

## Research Article

# Purification, Characterization, and Effect of Thiol Compounds on Activity of the *Erwinia carotovora* L-Asparaginase

Suchita C. Warangkar and Chandrahas N. Khobragade

Biotechnology Research Laboratory, School of Life Sciences, Swami Ramanand Teerth Marathwada University, Nanded 431606, India

Correspondence should be addressed to Chandrahas N. Khobragade, cnkhobragade@rediffmail.com

Received 22 May 2009; Accepted 31 July 2009

Academic Editor: Roberto Fernandez-Lafuente

Copyright © 2010 S. C. Warangkar and C. N. Khobragade. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

L-asparaginase was extracted from *Erwinia carotovora* and purified by ammonium sulfate fractionation (60–70%), Sephadex G-100, CM cellulose, and DEAE sephadex chromatography. The apparent Mr of enzyme under non-denaturing and denaturing conditions was 150 kDa and  $37 \pm 0.5$  kDa, respectively. L-asparaginase activity was studied in presence of thiols, namely, L-cystine (Cys), L-methionine (Met), N-acetyl cysteine (NAC), and reduced glutathione (GSH). Kinetic parameters in presence of thiols (10–400  $\mu$ M) showed an increase in  $V_{max}$  values (2000, 2223, 2380, 2500, and control 1666.7  $\mu$ moles  $mg^{-1}min^{-1}$ ) and a decrease in  $K_m$  values (0.086, 0.076, 0.062, 0.055 and control 0.098 mM) indicating nonessential mode of activation.  $K_A$  values displayed propensity to bind thiols. A decrease in  $V_{max}/K_m$  ratio in concentration plots showed inverse relationship between free thiol groups (NAC and GSH) and bound thiol group (Cys and Met). Enzyme activity was enhanced in presence of thiol protecting reagents like dithiothreitol (DTT), 2-mercaptoethanol (2-ME), and GSH, but inhibited by p-chloromercuribenzoate (PCMB) and iodoacetamide (IA).

## 1. Introduction

L-asparaginase (L-asparagine amino hydrolase, E.C.3.5.1.1) catalyzes the hydrolysis of L-asparagine into L-aspartic acid and ammonia [1–5]. L-asparaginase from *E. coli* and *E. chrysanthemii* has been used as chemotherapeutics in Acute Lymphoblastic Leukemia (ALL) for the past three decades [6]. The L-asparaginases from *E. coli* and *Erwinia* possess different immunological specificities and offer an important alternative therapy if a patient becomes hypersensitive to one of the enzymes [7]. The comparison of the two enzymes leads to the conclusion that *E. coli*-asparaginase can be recommended for first-line therapy, reserving *Erwinia*-asparaginase for allergic patients, because most patients allergic to the former are not immediately allergic to the latter [8]. It has been reported that thiol compounds such as N-acetyl cysteine and glutathione, in a fixed concentration potentiates the activity of asparaginase from *Cylindrocarpum obtusiporum* MB-10 [9] and *Pseudomonas stutzeri* MB-405 [10]. Enzyme activation by thiol group especially from glutathione N-acetyl cysteine is not an uncommon event in enzymology.

For instance, catalytic activities of cysteine protease from *Schistosoma mansoni* [11],  $N^{\omega}$ -phosphoarginine hydrolase from rat liver [12], reticulocytic protein kinase [13], glyoxylase [14], the family C G-protein coupled receptors, that is, the metotropic glutamate receptors were profoundly accelerated by the addition of GSH [15]. The Chloropicrin activation and reductive dechlorination by Glutathione lead to a decrease in the toxicity index as assayed by *Salmonella* mutagenicity test [16]. The molecular mechanism behind the activation effects of enzymes by glutathione have been the scope of the investigations for researchers. In catalysis by N-terminal hydrolase, Glutathione acts as an additional efficient molecule associated with redox changes [17]. Asparaginase activation is considered to be due to the involvement of free thiol group in the enzyme catalysis [18]. The Glutathione molecules can also binds to the G-protein coupled receptor which acts as a potentiator and agonist. Glutathione binds to family C-receptors in part via bonding interactions between the free  $\alpha$ -amino and  $\alpha$ -carboxyl groups of the ligands and a set of highly conserved residues in the binding pocket [19, 20].

In this paper, an attempt has been made to isolate asparaginase producing bacteria from soil, to extract, purify, characterize and examine the effect of four thiol compounds on the kinetic properties of asparaginase obtained from *E. carotovora*. We have investigated the effects of various concentrations of L-cystine (Cys), L-methionine (Met), N-acetyl cysteine (NAC), and reduced Glutathione (GSH) on the maximum velocity ( $V_{max}$ ) of asparagine hydrolysis and the apparent Michaelis-Menten constant ( $K_m$ ) of the enzyme in order to explain the mechanism of activation of asparaginase activity. The effect of some metal ions is also studied on the activity of L-asparaginase. The activation phenomenon caused due to thiol compounds could be exploited to treat ALL in future.

## 2. Materials and Methods

**2.1. Screening of Microbial Source.** The soil bacteria producing L-asparaginase was primarily screened by using the standard method [21, 22] and then selectively enriched into the growth medium containing yeast extract (1.0%, wt/vol) and L-asparagine (1.5%, wt/vol). A bacterium was grown at 28°C with aeration in Erlenmeyer flasks on a rotary shaker. The cells were harvested by centrifugation near the end of exponential phase of growth and stored at -10°C until needed.

**2.2. Asparaginase Production.** Culture medium was prepared with tryptone (5.0 g/L), yeast extract (5.0 g/L), fructose (1.0 g/L), and  $K_2HPO_4$  (5.0 g/L). The initial pH of the medium was adjusted to 7.0. Erlenmeyer flask (250 mL) containing 50 mL of culture medium was inoculated with 1000  $\mu$ L (2%, v/v) of bacterial suspension with a 0.8 absorbance at 620 nm. For inoculum preparation, two days old slant culture was scraped from agar surface, added to 0.85% sterile saline solution, and mixed until a homogeneous suspension was obtained. The flasks were incubated at 30°C on a rotary shaker at 120 rpm for 2-3 days.

**2.3. Asparaginase Activity Assay.** L-asparaginase activity was assayed by a modified method of Mashburn and Wriston [23]. A 0.1 mL cell suspension or purified enzyme solution, 0.9 mL sodium borate buffer (0.1 M, pH 8.5) and 1 mL L-asparagine (0.04 M) solution were combined and incubated for 10 minutes at 37°C. The reaction was stopped by the addition of 0.5 mL of 15% wt/vol trichloro acetic acid. The reaction contents were centrifuged at 8000 rpm. The supernatant was collected and 0.2 mL supernatant was diluted to 8 mL with distilled water. The resulting mixture was treated with 1.0 mL of Nessler's reagent and 1.0 mL of 2.0 M NaOH. The color reaction was allowed to proceed for 15 minutes before the absorbance at 500 nm was determined. The absorbance was then compared with a standard curve prepared from solutions of ammonium sulfate as the ammonia source. One international unit of L-asparaginase is the amount of enzyme which liberates 1  $\mu$ mole of ammonia in 1 minute at 37°C [24]. Determinations of protein concentrations were made on whole cell suspension or crude enzyme preparations [25].

**2.4. Purification of Bacterial L-Asparaginase.** Cells were harvested after 24 hours by centrifugation and washed with phosphate buffer ( $Na_2HPO_4$ , 4.76 g;  $KH_2PO_4$ , 4.54 g; Triton X-100, 0.125 mL; distilled water to 1 liter; pH adjusted to 7.0). Lysozyme spheroplast formation with subsequent release of L-asparaginase was carried out [26]. Washed cells (300 g) were suspended in 2 liters of 20% sucrose-0.033 M tris hydroxymethyl aminomethane (tris buffer, pH 8.0). Successive treatments of the cells with 0.01 volume of 0.1 M EDTA and lysozyme (5 mg/mL) of suspension were added at 5°C. The suspension was stirred gently, and osmotic fragility was checked at 15-minute intervals by determining the absorbance at 490 nm. After the decrease in absorbance had ceased, the suspension was centrifuged at 10 000 $\times$ g for 2 hours. To analyze for efficiency of extraction, the spheroplast was resuspended in 2 L of the original sucrose-tris solution. Analyses for enzyme content were made on the resuspended spheroplast and on the supernatant fluid from the final centrifugation [27]. All purification steps were carried out at 4°C unless stated otherwise.

Finely powdered ammonium sulfate was added to the protein to the final concentration of 70% saturation and stirred for 1 hour at 10°C. The mixture was left for 12 hours at 4°C. This was followed by centrifugation at 9000 rpm for 30 minutes at 4°C. The precipitate was dissolved in the phosphate buffer pH 8.5 and dialyzed overnight at 4°C against the same buffer [28].

The 10 mL of diluted supernatant was applied onto the Sephadex G-100 gel filtration column (2.3 cm  $\times$  120 cm), pre-equilibrated with 50 mM tris-HCl buffer pH 8.5 containing 100 mM KCl. The fractions were collected with 1 L of 40 mM sodium phosphate buffer (pH 6.0) at 10 mL/min flow rate (each fraction containing 10 mL). The L-asparaginase activity was assayed by the direct Nesslerization. Fractions containing L-asparaginase activity were pooled and dialyzed against 20 mM sodium phosphate buffer for 24 hours at 4°C.

The dialyzed sample was applied to the column of CM cellulose (2.0  $\times$  12.5 cm) that was pre-equilibrated with 0.01 M sodium phosphate buffer (pH 8.5). It was eluted with the NaCl gradient (0.1–0.5 M) and 0.01 M sodium phosphate buffer (pH 6.0). The fractions were collected and assayed for L-asparaginase activity. Active fractions were collected and used in the subsequent steps of purification.

A chromatography column (2.5 $\times$ 25 cm) was packed with DEAE sephadex and equilibrated with 0.1 M tris HCl buffer containing 2 mM  $MgCl_2$  (pH 8.6). The desalted enzyme thus obtained from dialysis was loaded on to the top of the column and eluted with 1 L of linear gradient tris HCl buffer composed of sodium chloride (0.1 M). The flow rate of the column was adjusted to 10 mL/min constantly to determine the protein concentration in the eluant (280 nm uv/vis Shimadzu). The 10 mL fractions were pooled and assayed for the protein and enzyme activity.

**2.5. Electrophoresis.** The mass of the native protein was determined by native gel electrophoresis on 10% acryl amide gels in tris-glycine (pH 8.5) using Genei Mini-V 8  $\times$  10 vertical gel electrophoresis system (Banglore Genei, India). Gels were stained with coomassie brilliant

blue. Myosin rabbit muscle (205 kDa), phosphorylase b (97.4 kDa), BSA (66 kDa), ovalbumin (43 kDa), carbonic anhydrase (29 kDa), soybean trypsin inhibitor (14.3 kDa), and aprotinin (6.5 kDa) were used as protein standards in native gel, while Phosphorylase b (97.3 kDa), bovine serum albumin (66 kDa), ovalbumin (43 kDa), soybean trypsin inhibitor (20 kDa), and lysozyme (14.3 kDa) were used as molecular weight markers for Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE). SDS-PAGE was carried out in 3 mm slab gel of 6% acrylamide in a tris borate buffer pH 7.2 containing 0.1% SDS. The gels were stained with 0.025% coomassie brilliant blue R-250 and destained by 1.0% acetic acid solution [29].

**2.6. Molecular Weight Determination by Gel Filtration.** The molecular weight of the enzyme subunit was estimated by gel filtration chromatography through a column (2.3 cm  $\times$  120 cm) of sephadex G-100 [30]. The column was equilibrated with 50 mM Tris HCl buffer, pH 8.5, containing 100 mM KCl. The column was calibrated with blue dextran and standard molecular weight marker proteins: Bovine Serum Albumin (66 kDa), Egg albumin (45 kDa), Carbonic anhydrase (29 kDa), and Cytochrome c (12.4 kDa). The elution volume ( $V_e$ ) of each marker protein and void volume ( $V_o$ ) of the column were estimated. A plot of  $V_e/V_o$  against log molecular weight was used to determine the molecular weight of L-asparaginase.

**2.7. Effect of Reagents on L-Asparaginase Activity.** The effect of different compounds was tested by preincubation (37°C for 20 minutes) of properly diluted enzyme with the respective reagents. The working concentrations of metal ions and thiol compounds were set at 0.5–10 mM and other thiol compounds were set at 10–400  $\mu$ M, respectively. After pre incubation, enzymatic assay was performed under optimal conditions, and enzyme activity was expressed as the percentage of the activity observed without additions of metal ions, thiol group blocking reagents, and thiols.

**2.8. Kinetic Measurements.** In all cases, enzymatic activity was assayed under temperature and pH optima. Alternatively data were fitted to the Michaelis-Menten equation to obtain the kinetic constants by applying linear regression. Activation of asparagine hydrolysis was studied in terms of change in values of kinetic parameters ( $K_m$  and  $V_{max}$ ) in presence and absence of thiol compounds. The values of parameters were determined from Lineweaver-Burk plots. The binding constants ( $K_A$ ) were obtained by plotting  $K_m/V_{max}$  versus thiol concentration. Kinetic analyses were also performed in the presence of fixed NAC and GSH concentrations (10–400  $\mu$ M) using different asparagine concentrations in the range of 0.02 to 0.2 mM. Enzymatic activity was determined as mentioned above taking into account that aspartate formation was linear for 40 minutes, in the presence of GSH and NAC under all conditions studied. On the other hand, Lineweaver-Burk analyses were performed to determine the binding constant ( $K_A$ ) for both GSH and NAC with the asparaginase interactions. These constants were calculated

by using the secondary replots of  $1/\Delta$  slope versus  $1/[\text{thiol compound}]$  according to Segel [31]. The  $\Delta$  slope values were obtained from individual Lineweaver-Burk plots. The constants  $\alpha$  and  $\beta$  refer to the fold change in the  $K_m$  and  $V_{max}$ , respectively, obtained in the presence of nonessential activator. In  $1/\Delta$  slope versus  $1/[\text{thiol}]$  replots, the  $x$  and  $y$  intercepts correspond to  $-\beta/\alpha K_A$  and  $\beta V_{max}/K_m$  ( $\beta-\alpha$ ), respectively. By assigning the values for  $\alpha$  and  $\beta$ ,  $K_A$  values are estimated accurately.

### 3. Results and Discussions

**3.1. Screening of Microbial Source.** The organism has been characterized as the genus *Erwinia*. It is a gram negative motile rod with peritrichous flagella (0.5  $\times$  1.0–3.0  $\mu$ m), sometimes occurs singly or in pairs and also forms short chains. On minimal agar slants colonies are shiny, smooth and circular off-white colored. Blood agar was not hemolysed. Growth as well as liquefaction of gelatin was seen in gelatin stabs. The bacillus was negative for cytochrome oxidase activity but positive for catalase activity. It converts nitrate to nitrite. Acid was produced with slight amount of gas from sugars like fructose, galactose, dextrose, and sucrose, and  $\beta$ -methyl glucoside. Various organic compounds are utilized as a source of carbon and energy viz citrate, lactate, formate, galacturonate, but tartarate, malate, benzoate, oxalate, and propionate do not serve as source of carbon and energy. There was no pigmentation on minimal agar. Casein and cotton seed oil was hydrolyzed at 37°C in medium which confirms the differentiation in between subspecies of *E. carotovora*.

**3.2. Purification of Bacterial L-Asparaginase.** L-asparaginase was purified from *E. carotovora*, using spheroplast formation, ammonium sulfate fractionation, sephadex G-100 gel filtration, CM cellulose, and DEAE Sephadex column chromatography. The recovery of enzyme is shown in Table 1. The specific activity of the enzyme increased with every step of purification with a minimum loss in quantity, giving a final recovery of 36.5% (Table 1). After every purification procedure, the peak fractions with the enzyme activity were analyzed using SDS-PAGE. The enzyme precipitated out between 60 and 70% of ammonium sulfate saturation. Removal of salt from the enzyme by Sephadex G-100 was found to be suitable after that step enzyme loss was negligible. Peak fractions of Sephadex G-100 chromatography were pooled together and loaded onto equilibrated CM cellulose column for purification and in the eluant two protein peaks and one enzyme activity peak are obtained (Figure 1(a)). Enzyme activity peak was matched with the second protein peak. Further peak fractions were collected from CM cellulose chromatography and loaded onto equilibrated DEAE Sephadex column for final purification. The elution profile has shown two peaks for proteins and one for enzyme activity. Enzyme activity was detected only in trace amount in the peak I fractions. While peak II has showed lower protein concentration but higher enzyme activity (Figure 1(b)).

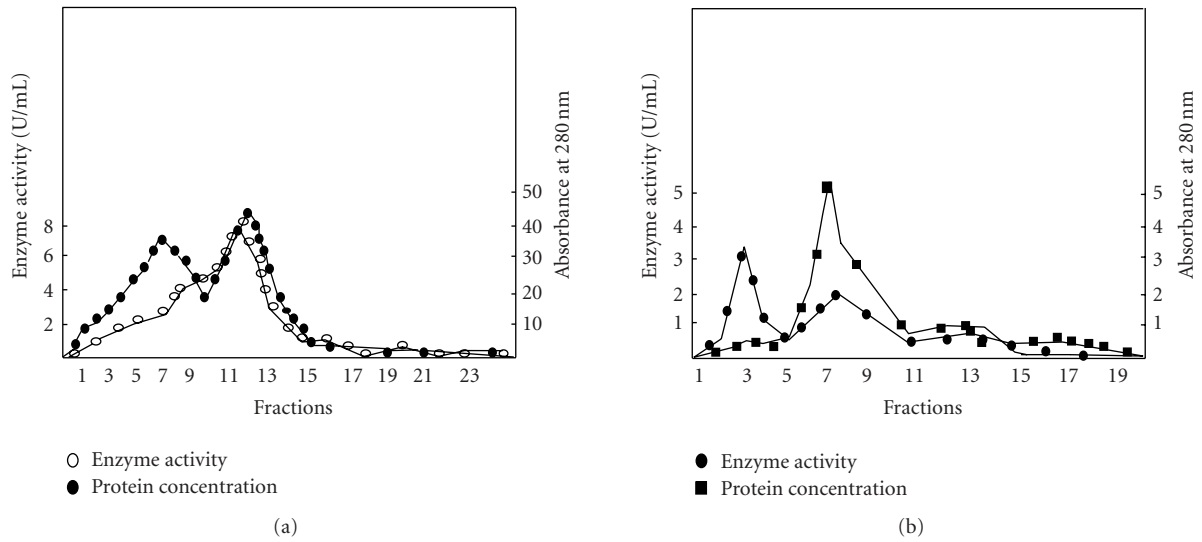


FIGURE 1: Purification of L-asparaginase from the *E. carotovora*. (a) CM cellulose chromatography of the active fractions collected from the Sephadex G-100 gel filtration column. (b) DEAE cellulose Anion Exchange Chromatography of the active fractions collected from the CM cellulose chromatography column.

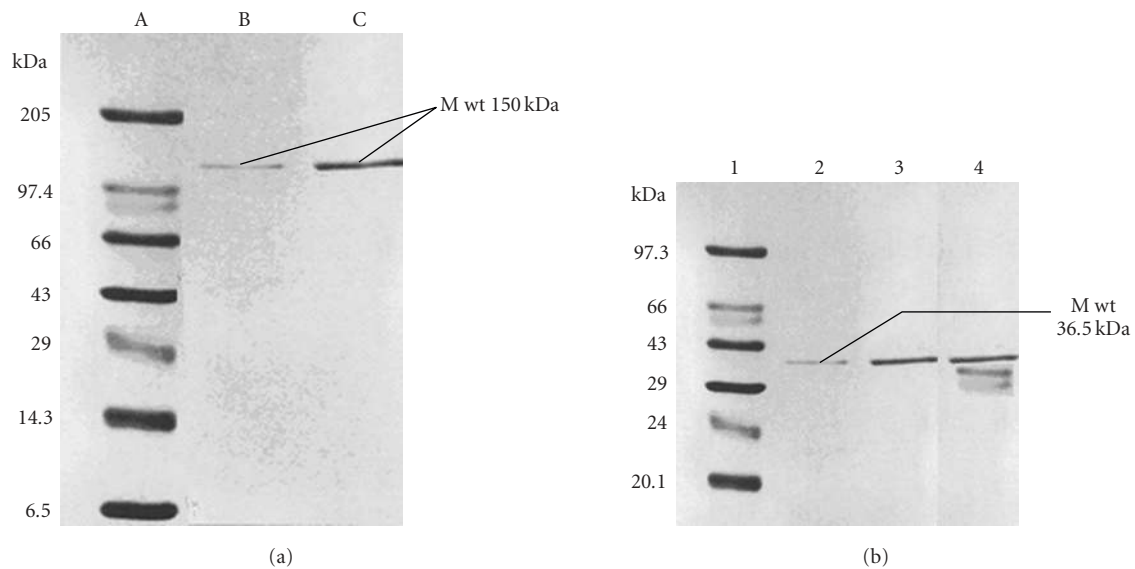


FIGURE 2: (a) Native PAGE of the purified L-asparaginase from the *E. carotovora*, (i) molecular weight marker proteins, (ii) purified asparaginase 10  $\mu\text{g}/\text{mL}$ , (iii) purified asparaginase 25  $\mu\text{g}/\text{mL}$ . (b) SDS-PAGE of the purified L-asparaginase from the *E. carotovora*, lane 1 molecular weight marker proteins, lane 2 purified asparaginase after DEAE sephadex chromatography, lane 3 purified asparaginase after CM Cellulose chromatography, lane 4 crude preparation.

TABLE 1: L-asparaginase purification summary.

Purification steps	Total activity (IU)	Total Protein (mg)	Specific activity (IU/mg)	Purification fold	Recovery (%)
Spheroplast suspension	6800	5000	1.36	1.0	100
Ammonium sulfate precipitation	5000	1400	3.57	2.62	73
Sephadex G-100 gel filtration	4100	248.9	16.4	12.11	60
CM Cellulose	3172	10.2	310.9	228.6	46
DEAE sephadex	2482	2.4	1034	752.9	36.5



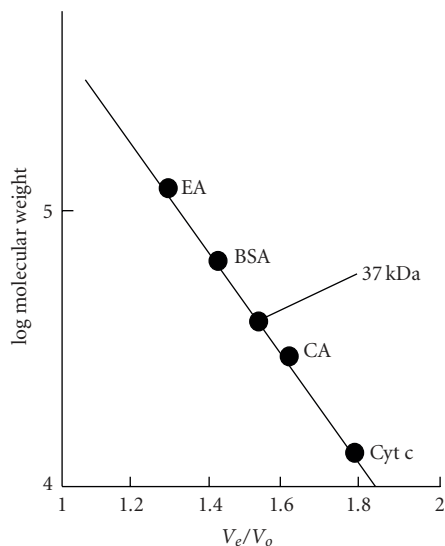


FIGURE 3: Calibration curve for the determination of molecular weight of L-asparaginase by gel filtration chromatography.

### 3.3. Electrophoresis and Molecular Weight Determination.

The L-asparaginase from *E. carotovora* is active as homotetramer and its crystal structure (PDB ID 1ZCF) and function have been thoroughly characterized with the approximate molecular mass of 150 kDa as determined by native PAGE (Figure 2(a)) [32, 33]. The molecular mass of enzyme subunit is  $36.5 \pm 0.5$  kDa as observed from SDS-PAGE separation and gel filtration. The SDS-PAGE of the enzyme preparation from different purification steps showed that the resolved electrophoretic bands were progressively improved from the crude extract to the final step of purification. It revealed only one distinctive band that was indicated by the pure preparation of L-asparaginase (Figure 2(b)). Native PAGE has exhibited an approximate molecular weight 150 kDa, which indicates that L-asparaginase purified from *E. carotovora* was homogeneous. SDS-PAGE and gel filtration chromatography indicated that enzyme subunit is one band with approximate molecular weight of 36.5 kDa and 37 kDa, respectively (Figure 3). This value was equal to tumor inhibitory L-asparaginase [32].

### 3.4. Effect of Reagents on L-Asparaginase Activity.

L-asparaginase activity was assayed in the presence of different reagents (Table 2). Among the salts tested, considerable loss of activity was observed only with  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Mg}^{2+}$  and  $\text{Zn}^{2+}$ , whereas  $\text{Na}^+$  and  $\text{K}^+$  acting somewhat as an enhancer. This was also true for EDTA. The data indicates that asparaginase may not be the metallo-enzyme. Among the amino acids tested, only L-cysteine and histidine stimulate the relative activity, while others had not observable effect. Inhibition of enzyme activity in presence of  $\text{Hg}^{2+}$ ,  $\text{Cd}^{2+}$ , and  $\text{Zn}^{2+}$  might be indicative of essential vicinal sulfhydryl groups of the enzyme for productive catalysis. Furthermore, stimulation of the activation with reducing agents like 2-mercaptoethanol (2-ME), dithiothreitol (DTT), and reduced glutathione (GSH) and inhibition in the

TABLE 2: Influence of different metal ions and reagents on L-asparaginase activity.

Addition	Concentration (mM)	Relative activity (%)
No addition	none	100
$\text{Na}^+$ (NaCl)	1.0	117
$\text{K}^+$ (KCl)	1.0	133
$\text{Mg}^{2+}$ ( $\text{MgCl}_2$ )	1.0	24
$\text{Ca}^{2+}$ ( $\text{CaCl}_2$ )	1.0	78
$\text{Mn}^{2+}$ ( $\text{MnCl}_2$ )	1.0	71
$\text{Zn}^{2+}$ ( $\text{ZnCl}_2$ )	0.5	06
$\text{Fe}^{3+}$ ( $\text{FeCl}_3$ )	0.5	47
$\text{Hg}^{2+}$ ( $\text{HgCl}_2$ )	1.0	0
$\text{Ni}^{2+}$ ( $\text{NiCl}_2$ )	1.0	51
$\text{Cu}^{2+}$ ( $\text{CuCl}_2$ )	1.0	0
$\text{Cd}^{2+}$ ( $\text{CdCl}_2$ )	1.0	0
EDTA	0.5	117
L-Cysteine	25	148
L-Histidine	25	114
Glutathione (reduced)	0.5	136
Thiourea	0.5	114
Thioacetic acid	0.5	106
Thioacetamide	0.5	110
2-mercaptoethanol	0.5	112
Dithiothreitol	0.5	86
p-chloro mercury benzoic acid	0.5	0
Iodoacetamide	0.5	0
Sodium dodecyl sulfate	2.0	119
	3.0	22
Urea	2.0	108
$\text{H}_2\text{O}_2$	1.0	24

presence of thiol group blocking reagents, namely, PCMB (P-chloro mercury benzoate) and IA (iodoacetate) provided additional proof for the role of sulfhydryl groups in the catalytic activity of the enzyme. The enzyme completely lost its activity at 4.0 M urea and only 22% of activity was retained at 3.0 M sodium dodecyl sulfate (SDS).

**3.5. Kinetic Measurements.** The enzyme showed typical Michaelis-Menten kinetics at lower substrate concentrations and the apparent  $K_m$  value for L-asparagine is 0.098 mM. The apparent  $K_m$  and  $V_{max}$  values were tentatively determined from both linear and nonlinear regressions using asparagine and glutamine as substrates. The lower apparent  $K_m$  value for L-asparagine indicates that the purified L-asparaginase has higher affinity for the substrate asparagine. The efficiency of substrate utilization was estimated by  $V_{max}/K_m$  ratios (Table 3) and the hydrolysis efficiency of asparagine was at least 11 925 fold higher than that of L-glutamine.

**3.6. Effect of Thiol Compounds on L-Asparaginase.** The enzyme activity was determined in the presence of different

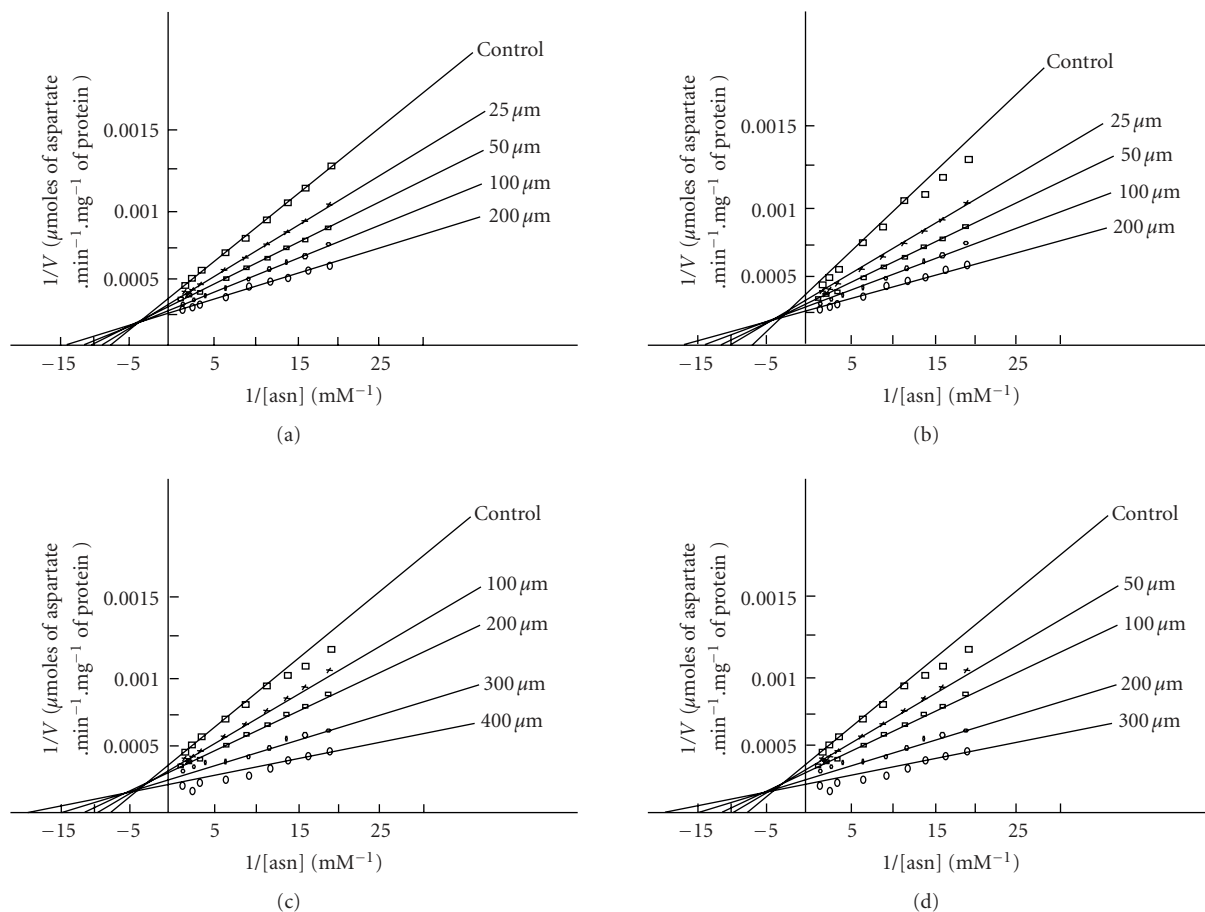


FIGURE 4: Lineweaver-Burk analyses of activity shown by asparaginase from *E. carotovora* in the presence of different concentrations of Cys (a), Met (b), NAC (c), and GSH (d).

TABLE 3: Kinetic constants of partially purified asparaginase.

Substrate	Linear regression (Lineweaver-Burkplot)			Nonlinear regression (Michaelis-Menten equation)		
	$K_m$ (mM)	$V_{max}$ ( $\mu\text{moles mg}^{-1}\text{min}^{-1}$ )	$V_{max}/K_m$	$K_m$ (mM)	$V_{max}$ ( $\mu\text{moles mg}^{-1}\text{min}^{-1}$ )	$V_{max}/K_m$
L-asparagine	0.096	1632.6	17,006	0.098	1666.7	17,007
L-glutamine	2.86	4.08	1.426	3.04	4.74	1.559

thiol compounds. A lower concentration of GSH and NAC in the medium has exhibited stimulatory effects on L-asparaginase catalytic activity. The activation was fully characterized by performing Lineweaver-Burk analyses using different concentration of Cys, Met, NAC, and GSH (10–400  $\mu\text{M}$ ) (Figure 4). An increase in amount of GSH and NAC in the reaction medium revealed an increase in  $V_{max}$  and a decrease in  $K_m$  values, which corresponds to a nonessential activation [34, 35]. The secondary replot of  $1/\Delta$  slope versus  $1/[\text{thiols}]$  (Figure 5) enabled us to know  $\alpha$ ,  $\beta$ , and binding constant  $K_A$  values for each thiol compounds (Table 4). The constants  $\alpha$  and  $\beta$  refer to the fold change in  $K_m$  and  $V_{max}$ , respectively, in the presence of each thiol compound. From Table 4, it is evident that the  $K_A$  value for the enzyme decreases from Cys, Met, NAC, and GSH. As a consequence, the concentration of each thiol compound

at which maximum stimulation was achieved was lowest for Cys and Met. This data suggests that relatively specific interaction may take place between the free thiol group and enzyme, as a critical determinant of this interaction as well as activation. Thiol compound binding may lead to decrease in the  $K_m$  which is more evident with GSH ( $\alpha = 0.062$ ). In this sense,  $\alpha$  value gradually decreases when the thiol group of compound is not free for interaction with enzyme. However, the magnitude of nonessential activation is almost same for every free thiol group containing compounds, taking into account the  $\beta$  values (Table 4). The catalytic activity of asparaginase from *E. carotovora* was not further increased, when the concentrations of each thiol compound were increased in the assay medium more than the concentration (400  $\mu\text{M}$ ) reported for the nonessential activation. All the data described previously were expressed in terms of relative

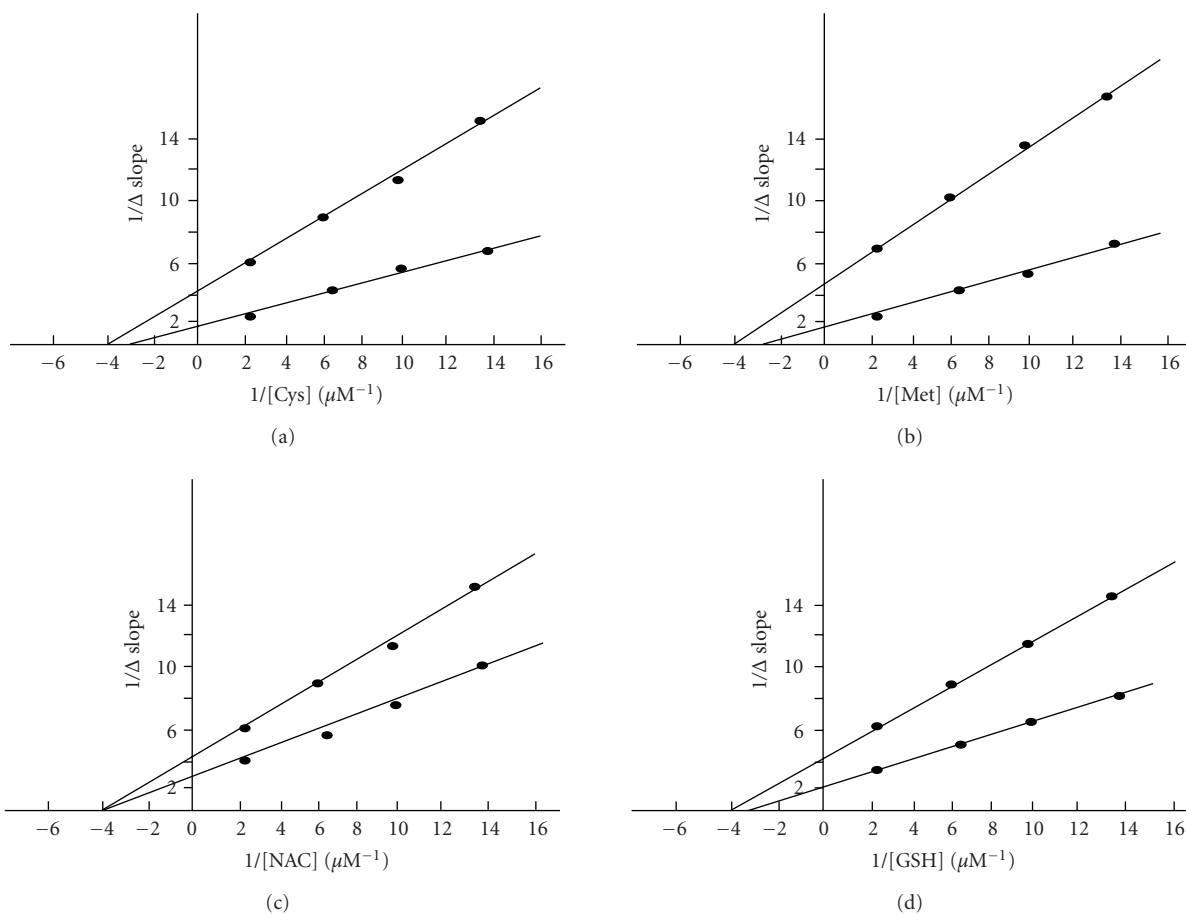


FIGURE 5: The secondary replots of  $1/\Delta$  slope versus  $1/[\text{Cys}]$  (a),  $1/\Delta$  slope versus  $1/[\text{Met}]$  (b),  $1/\Delta$  slope versus  $1/[\text{NAC}]$  (c), and  $1/\Delta$  slope versus  $1/[\text{GSH}]$  (d).

$V_{\max}/K_m$  ratio and plotted versus thiol compound concentrations (Figure 6). For each of the four thiol compounds at the lower concentrations, these replots exhibited positive slope, indicating that GSH and NAC act as activators at these concentrations, while Cys and Met at lower concentrations result in a negative slope indicating inhibition. It is obvious to observe an inverse relationship between the free and bound thiol group of the compounds and the breakpoint in the  $V_{\max}/K_m$  ratio versus thiol compound concentration plots.

Bacterial asparaginases have been the subject of considerable medical interest and are being employed in the therapy of acute lymphoblastic leukemia. Its therapeutic potential is now well established, as it has remarkably induced remission in most of the patients suffering with ALL. A comparative examination of preparations of *Erwinia* L-asparaginase made in USSR and Germany was recommended for clinical use. The catalytic geometry and the secondary structure around the active sites of asparaginase from *E. coli*, *E. carotovora*, and *E. chrysanthemii* are closely similar but their substrate specificity pockets are quite different. To explain the mechanism of asparaginase activation by GSH and NAC, it is to conceive that asparaginase possesses the thiol group binding domain with high affinity towards free-SH group

containing effectors. Using this model, the most hydrophilic compounds such as GSH and NAC would bind more effectively to the activator site and convert the asparaginase from one conformation to another conformation, that is, State I to State II (Scheme 1). The later form is catalytically more active than the free form of the enzyme, as is evident from the increase in  $V_{\max}/K_m$  ratio. The asparaginase activation induced by GSH and NAC can be considered as a case of nonessential activation. In the extensive kinetic analyses of  $1/\Delta$  slope versus  $1/[\text{thiol compound}]$ , values of  $\beta V_{\max}/K_m$  ( $\beta\text{-}\alpha$ ) are determined, which are approximately same for NAC and GSH, but different for Cys and Met. This supports the hypothesis that all thiol group containing amino acids and compounds may interact with the same activator site on asparaginase. This kind of stimulatory action has been also studied with the enzyme glyoxalase, but it is concentration dependant. The mechanism of NAC and GSH mediated activation in mitogen activated protein kinase was redox dependant and requires reactive oxygen species (ROS) sensitive mechanism for regulation of TNF- $\alpha$  biosynthesis [13]. Molecular docking experiments with GSH in G-protein coupled receptor have shown that the peptide binds in a "bent" conformation by protruding the terminal glycine residue upward in the pocket. At the same

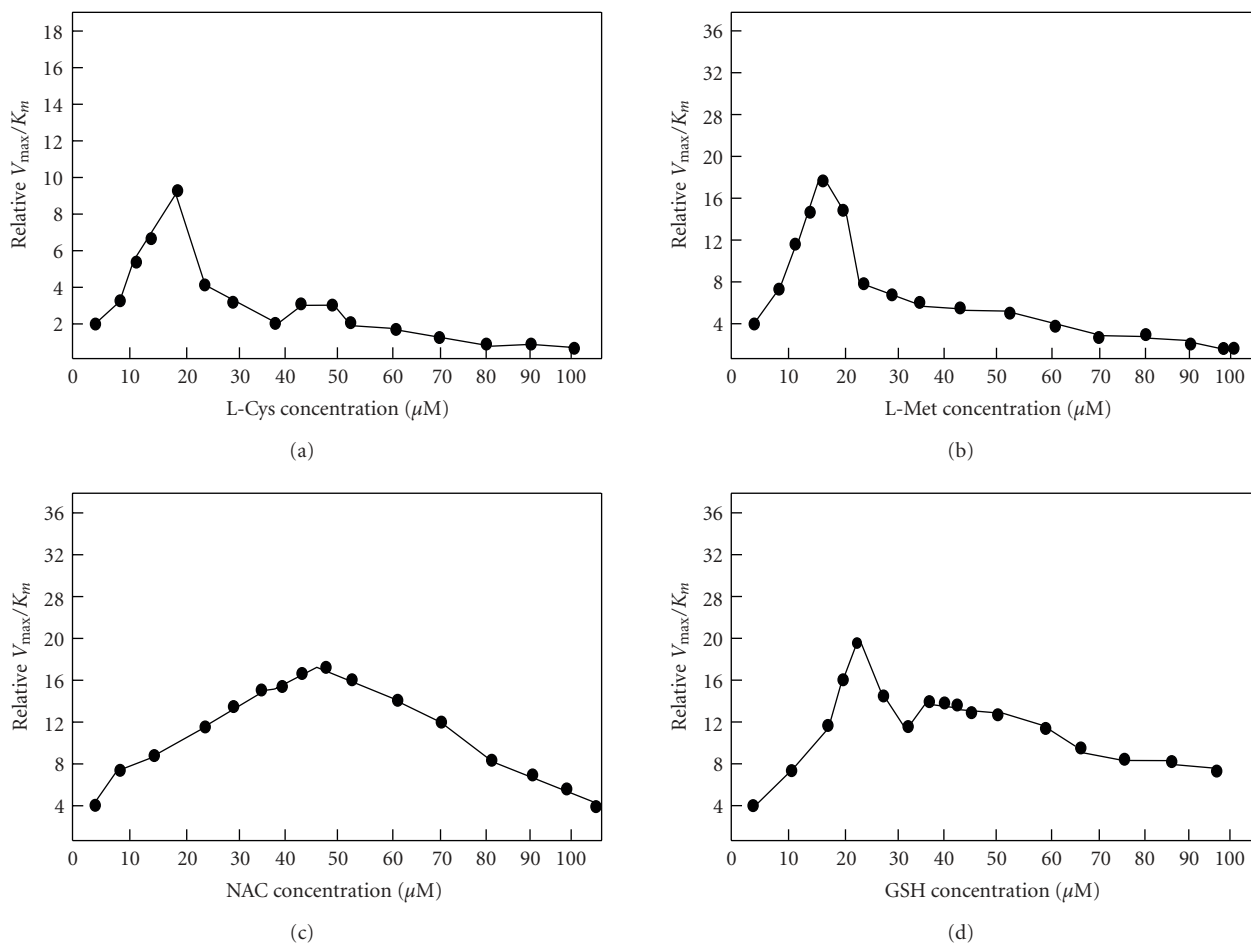


FIGURE 6: Effect of thiol containing compounds on the relative  $V_{\max}/K_m$  ratio of L-asparaginase from *E. carotovora*.

TABLE 4: Summary of activation parameters and affinity constants of thiol containing compounds to asparaginase.

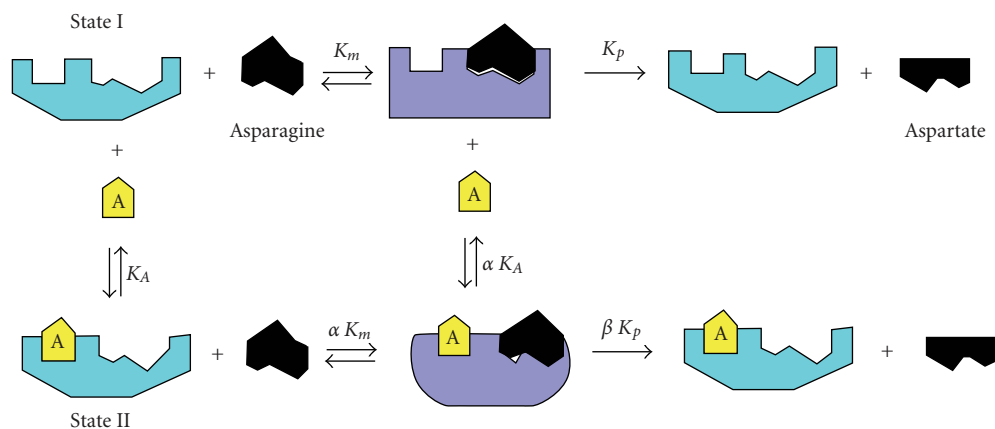
Thiol compounds	$\alpha$	$\beta$	$K_A$ (mM)	$K_m$ (mM)	$V_{\max}$ ( $\mu\text{moles mg}^{-1}\text{min}^{-1}$ )	$V_{\max}/K_m$
control	—	—	—	0.098	1666.7	17,007
Cys	0.589	1.87	293	0.086	2000	23,255
Met	0.138	1.96	229	0.076	2223	29,250
NAC	0.081	2.27	128	0.062	2380	38,387
GSH	0.062	1.89	14.6	0.055	2500	45,454

time, it was also proposed that like free amino acids, GSH acts in conjunction with divalent cation ligands promoted closure of the Venus fly trap domain and initiate receptor activation [15]. This effect was probably the cause of a more flexible enzyme conformation, which has a higher catalytic activity. The authors ascribed this effect to an enzyme conformational change provoked by free thiol group containing amino acids with the enzyme at a locus other than that for substrate binding (Scheme 2). In asparaginase, the lower concentrations of free thiol group containing non polar amino acids should change the enzyme structure to a more active conformation due to a more effective interaction with the hydrophilic activator site. Higher concentrations of

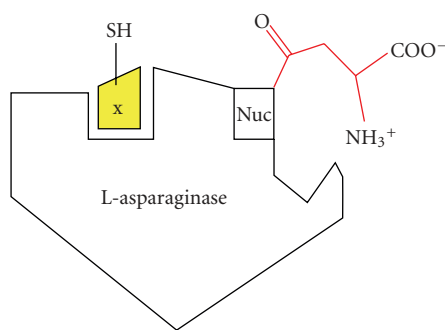
NAC and GSH were resulted in an uncompetitive mode of inhibition of enzyme in its activated state. It was proposed that once the activator site of asparaginase is filled and all enzyme molecules are activated, the excess of molecules may compete with enzyme substrate complex (ES) during the deamination step of the hydrolysis, accounting for the uncompetitive inhibition and lowering the turnover.

The whole data of the present work indicate that NAC and GSH may binds to the activator site, which is very hydrophobic pocket nearer to the hydrophilic active site. In the present paper, we have described the extraction, purification, characterization, and activation of asparaginase from *E. carotovora* by GSH and NAC and kinetic events





SCHEME 1: Model for the nonessential activation of asparaginase by thiol containing compounds.



SCHEME 2: Acyl enzyme intermediate attack by nucleophile Tyr/Thr. The probable acyl enzyme intermediate of asparaginase from *E. coli* where Thr is a nucleophile in the N-terminal position.

accompanying with the increase of thiol concentration in a reaction medium. This biocatalyst in the presence of thiol compounds will be more useful to improve a potential biotechnological purpose and to increase the catalytic efficiency of asparaginase as the therapeutic agent in the treatment of ALL and other blood system tumors. The comparison of crystal structures, active sites, combined crystallographic, thermal stability, and cytotoxic experiments has shown that *E. carotovora* asparaginase is 30 times less toxic against the human leukemia cell lines. But, denaturing experiments showed that *E. carotovora* asparaginase has decreased thermodynamic stability as compared to the *E. coli* asparaginase and get rapidly inactivated in the presence of urea [33]. Our studies may find some alternative to stabilise the structure and function of *E. carotovora* asparaginase by adding thiols.

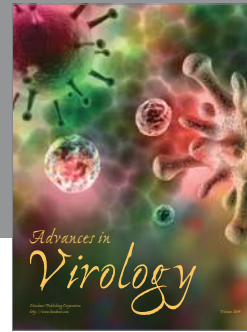
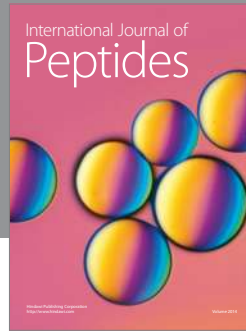
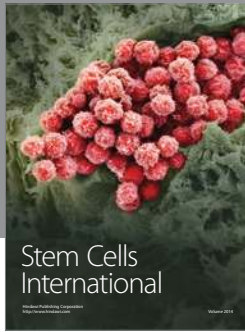
## Acknowledgments

The author Suchita C. Warangkar acknowledges UGC, India, for awarding the non-SAP-UGC meritorious fellowship. The authors are also thankful to the Director of the School of Life Sciences, S. R. T. M. University, Nanded, India for providing all the necessary facilities.

## References

- [1] A. A. Borisova, M. A. Eldarov, A. A. Zhgun, et al., "Purification and some properties of recombinant *Erwinia carotovora* L-asparaginase, expressed in *E. coli* cells," *Biomeditsinskaya Khimiya*, vol. 49, no. 5, pp. 502–507, 2003.
- [2] S. H. Fisher and L. V. Wray Jr., "Bacillus subtilis 168 contains two differentially regulated genes encoding L-asparaginase," *Journal of Bacteriology*, vol. 184, no. 8, pp. 2148–2154, 2002.
- [3] A. C. T. North, H. E. Wade, and K. A. Cammack, "Physicochemical studies of L-Asparaginase from *Erwinia carotovora*," *Nature*, vol. 224, no. 5219, pp. 594–595, 1969.
- [4] H. L. Ammon, K. C. Murphy, K. Chandrasekhar, and A. Wlodawer, "Preliminary crystallographic study of an l-asparaginase from *Vibrio succinogenes*," *Journal of Molecular Biology*, vol. 184, no. 1, pp. 179–181, 1985.
- [5] K. Sieciechowicz, R. J. Ireland, and K. W. Joy, *Plant Physiology*, vol. 77, pp. 506–508, 1985.
- [6] K. Aghaiypour, A. Wlodawer, and J. Lubkowski, "Structural basis for the activity and substrate specificity of *Erwinia chrysanthemi* L-asparaginase," *Biochemistry*, vol. 40, no. 19, pp. 5655–5664, 2001.
- [7] S.-M. Lee, M. H. Wroble, and J. T. Ross, "L-asparaginase from *Erwinia carotovora*: an improved recovery and purification process using affinity chromatography," *Applied Biochemistry and Biotechnology*, vol. 22, no. 1, pp. 1–11, 1989.
- [8] M. Duval, S. Suci, A. Ferster, et al., "Comparison of *Escherichia coli*-asparaginase with *Erwinia*-asparaginase in the treatment of childhood lymphoid malignancies: results of a randomized European Organisation for Research and Treatment of Cancer—Children's Leukemia Group phase 3 trial," *Blood*, vol. 99, no. 8, pp. 2734–2739, 2002.
- [9] S. K. Raha, S. K. Roy, S. K. Dey, and S. L. Chakrabarty, "Purification and properties of an L-asparaginase from *Cylindrocarpus obtusisporus* MB-10," *Biochemistry International*, vol. 21, no. 6, pp. 987–1000, 1990.
- [10] S. Manna, A. Sinha, R. Sadhukhan, and S. L. Chakrabarty, "Purification, characterization and antitumor activity of L-asparaginase isolated from *Pseudomonas stutzeri* MB-405," *Current Microbiology*, vol. 30, no. 5, pp. 291–298, 1995.
- [11] C. L. Chappell, M. H. Dresden, and D. W. Walters, "Glutathione activation of a cysteine proteinase from *Schistosoma mansoni*," *Biochimica et Biophysica Acta*, vol. 913, no. 3, pp. 335–341, 1987.

- [12] M. Kuba, H. Ohmori, and A. Kumon, "Characterization of N( $\omega$ )-phosphoarginine hydrolase from rat liver," *European Journal of Biochemistry*, vol. 208, no. 3, pp. 747–752, 2005.
- [13] V. Ernst, D. H. Levin, and I. M. London, "Inhibition of protein synthesis initiation by oxidized glutathione: activation of a protein kinase that phosphorylates the  $\alpha$  subunit of eukaryotic initiation factor 2," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 75, no. 9, pp. 4110–4114, 1979.
- [14] J. Giršavičius and P. A. Heyfetz, "Mechanism of glyoxalase activation by glutathione," *Nature*, vol. 136, no. 3442, pp. 645–646, 1935.
- [15] M. Wang, Y. Yao, D. Kuang, and D. R. Hampson, "Activation of Family C G-protein-coupled receptors by the tripeptide glutathione," *The Journal of Biological Chemistry*, vol. 281, no. 13, pp. 8864–8870, 2006.
- [16] M. Schneider, G. B. Quistad, and J. E. Casida, "Glutathione activation of chloropicrin in the Salmonella mutagenicity test," *Mutation Research*, vol. 439, no. 2, pp. 233–238, 1999.
- [17] O. I. Aruoma, B. Halliwell, B. M. Hoey, and J. Butler, "The antioxidant action of N-acetylcysteine: its reaction with hydrogen peroxide, hydroxyl radical, superoxide, and hypochlorous acid," *Free Radical Biology and Medicine*, vol. 6, no. 6, pp. 593–597, 1989.
- [18] B. Halliwell, "Antioxidant defence mechanisms: from the beginning to the end (of the beginning)," *Free Radical Research*, vol. 31, no. 4, pp. 261–272, 1999.
- [19] D. Kuang, Y. Yao, M. Wang, N. Pattabiraman, L. P. Kotra, and D. R. Hampson, "Molecular similarities in the ligand binding pockets of an odorant receptor and the metabotropic glutamate receptors," *The Journal of Biological Chemistry*, vol. 278, no. 43, pp. 42551–42559, 2003.
- [20] P. Luu, F. Acher, H.-O. Bertrand, J. Fan, and J. Ngai, "Molecular determinants of ligand selectivity in a vertebrate odorant receptor," *Journal of Neuroscience*, vol. 24, no. 45, pp. 10128–10137, 2004.
- [21] W. Chen, L. Wu, W. T. Frankenberger Jr., and A. C. Chang, "Soil enzyme activities of long-term reclaimed wastewater-irrigated soils," *Journal of Environmental Quality*, vol. 37, supplement 5, pp. S36–S42, 2008.
- [22] R. Gulati, R. K. Saxena, and R. Gupta, "A rapid plate assay for screening L-asparaginase producing micro-organisms," *Letters in Applied Microbiology*, vol. 24, no. 1, pp. 23–26, 1997.
- [23] L. T. Mashburn and J. C. Wriston Jr., "Tumor inhibitory effect of L-asparaginase," *Biochemical and Biophysical Research Communications*, vol. 12, no. 1, pp. 50–55, 1963.
- [24] R. E. Peterson and A. Ciegler, "L-asparaginase production by *Erwinia* aroideae," *Applied microbiology*, vol. 18, no. 1, pp. 64–67, 1969.
- [25] O. H. Lowry, N. J. Rosenbrough, A. L. Farr, and R. J. Randall, *The Journal of Biological Chemistry*, vol. 193, pp. 265–275, 1951.
- [26] M. H. Malamy and B. L. Horecker, "Release of alkaline phosphatase from cells of *Escherichia coli* upon lysozyme spheroplast formation," *Biochemistry*, vol. 3, no. 12, pp. 1889–1893, 1965.
- [27] P. W. Buck, R. Elsworth, G. A. Miller, K. Sargeant, J. L. Stanley, and H. E. Wade, "The batch production of L-asparaginase from *Erwinia carotovora*," *Journal of General Microbiology*, vol. 65, no. 3, p. i, 1971.
- [28] R. K. Scopes, *Protein Purification, Principles and Practice*, Springer, New York, NY, USA, 3rd edition, 1994.
- [29] U. K. Laemmli, "Cleavage of structural proteins during the assembly of the head of bacteriophage T4," *Nature*, vol. 227, no. 5259, pp. 680–685, 1970.
- [30] P. Andrews, "Estimation of the molecular weights of proteins by Sephadex gel-filtration," *Biochemical Journal*, vol. 91, no. 2, pp. 222–233, 1964.
- [31] International Union of Pure and Applied Chemistry, "Symbolism and terminology in chemical kinetics (Provisional)," *Pure and Applied Chemistry*, vol. 53, no. 3, pp. 753–771, 1981.
- [32] J. C. Wriston Jr. and T. O. Yellin, "L-asparaginase: a review," *Advances in Enzymology and Related Areas of Molecular Biology*, vol. 39, pp. 185–248, 1973.
- [33] A. C. Papageorgiou, G. A. Posypanova, C. S. Andersson, N. N. Sokolov, and J. Krasotkina, "Structural and functional insights into *Erwinia carotovora* L-asparaginase," *FEBS Journal*, vol. 275, no. 17, pp. 4306–4316, 2008.
- [34] I. H. Segel, "Enzyme activation," in *Enzyme Kinetics*, pp. 227–272, Wiley Interscience, New York, NY, USA, 1975.
- [35] J. Krasotkina, A. A. Borisova, Y. V. Gervaziev, and N. N. Sokolov, "One-step purification and kinetic properties of the recombinant L-asparaginase from *Erwinia carotovora*," *Biotechnology and Applied Biochemistry*, vol. 39, no. 2, pp. 215–221, 2004.



**Hindawi**

Submit your manuscripts at  
<http://www.hindawi.com>

