

**Workshop Report: Putative Adverse Outcome Pathways (AOP) Relevant to Neurotoxicity**

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**Workshop Report: Putative Adverse Outcome Pathways (AOP) Relevant to Neurotoxicity**

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## Abstract

The Adverse Outcome Pathway (AOP) framework provides a template that facilitates understanding of complex biological systems and the pathways of toxicity that result in adverse outcomes (AOs). The AOP starts with an molecular initiating event (MIE) in which a chemical interacts with a biological target(s), followed by a sequential series of KEs, which are cellular, anatomical and/or functional changes in biological processes, that ultimately result in an AO manifest in individual organisms and populations. It has been developed as a tool for a knowledge-based safety assessment that relies on understanding mechanisms of toxicity, rather than simply observing its adverse outcome.

A large number of cellular and molecular processes are known to be crucial to proper development and function of the central (CNS) and peripheral nervous systems (PNS). However, there are relatively few examples of well-documented pathways that include causally linked MIEs and KEs that result in adverse outcomes in the CNS or PNS. As a first step in applying the AOP framework to adverse health outcomes associated with exposure to exogenous neurotoxic substances, the EU Reference Laboratory for Alternatives to Animal Testing (EURL ECVAM) organized a workshop (March 2013, Ispra, Italy) to identify potential AOPs relevant to neurotoxic and developmental neurotoxic outcomes. Although the AOPs outlined during the workshop are not fully described, they could serve as a basis for further, more detailed AOP development by further identification of data gaps and application of modified Bradford-Hill criteria (Meek et al., 2014) to be useful for human health risk assessment.

**Key words:** *pathways of neurotoxicity, molecular initiating event, key events, adverse outcome pathway, in vitro testing, predictive toxicology*

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## Introduction to AOP concept

Regulatory toxicology is undergoing a transformation from decision-making relying solely on apical animal testing to biological pathway-based approaches that include cellular, tissue and organismal dynamics. A key driver for this change is a collective desire of all stakeholders (academia, industry, and regulators) to better utilize the latest scientific thinking and tools within the safety assessment process (Vinken 2013). Success will improve protection of human health and the environment, considerably reduce animal testing, significantly increase the rate of data collection and provide opportunities for industrial innovation and competitiveness. Central to this transformation is a shift towards knowledge-based weigh-of-evidence paradigm for hazard assessment. This transformative process must include development of cost efficient and less time consuming toxicity testing methods that predict the impact of chemicals on human and ecological health (Collins et al., 2008). The ultimate goal would involve routine toxicity testing conducted in human-cell based test systems (NRC, 2007) to more accurately predict the potential adverse effects of chemicals rather than having to rely on *in vivo* animal models. Such predictive toxicology is still emerging and will require considerably more research and development before its principles and processes are mature enough to translate into mainstream regulatory practice (Thomas et al., 2013; Patlewicz and Lander, 2013; Adeleve et al., 2014). One area that needs particular attention is how to actually harvest, curate and manage relevant knowledge so that it informs AOP development that serves regulatory needs.

Extant literature contains an extraordinary amount of information on the mechanisms by which chemicals alter cellular signaling pathways mostly generated in the context of toxicological science or in relation to basic biomedical research. However, generation of this data is only one step in the process of use in risk decisions. The distillation of knowledge from such data requires

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3 functional understanding of complex biological systems and underlying system dynamics, as  
4 well as xenobiotic-induced perturbations that lead to dysfunction and failure.  
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8 The Adverse Outcome Pathway (AOP) framework has been developed as a tool to organize data  
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10 across multiple levels of biological organization to identify correlative and causal linkages  
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12 between the events, that when sufficiently perturbed by chemical exposures, result in adverse  
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14 outcomes. (Ankley et al, 2010). The AOP framework not only provides a means to adapt  
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16 mechanistic understanding for regulatory decision making, but it also provides a useful tool for  
17  
18 consolidating, managing and exchanging knowledge amongst the research community. The  
19  
20 concept of the AOP structure encompasses functional systems biology/toxicology with the  
21  
22 ultimate goal of predicting systems behaviour. It is important to appreciate, however, that an  
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24 AOP is not a description of a biological system per se, but is a higher-level depiction of the  
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26 sequence of toxicological events that lead to dysfunction or failure of the system, given a certain  
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28 set of circumstances or boundary conditions.  
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34 AOPs can vary in resolution and expanse and can include both qualitative and quantitative  
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36 descriptions of key events (KE) and their interlinking causal relationships (Fig. 1). Initial  
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38 development and elucidation of an AOP begins with a proposed pathway (Ankely et al., 2010;  
39  
40 Watanabe et al., 2011). Recent published guidance and template documents for development  
41  
42 and assessment of AOPs (OECD, 2013) suggest whenever possible anchoring the AOP at the  
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44 molecular initiating event (MIE) and the adverse outcome (AO) at the individual or population  
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46 level (i.e. the regulatory effect/endpoint of concern). However, AOP development may originate  
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48 at any step in the pathway, whether nearer to the initial chemical-biological interaction, the MIE,  
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50 or somewhere in-between initiation and AO. The AOP is then further developed by identifying  
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52 and describing the intermediate KEs and the causal or correlative relationships between them. An  
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3 additional aspect of this process is transitioning from a correlative-based AOP to one based on  
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5 causative links between the MIE, KE, and AO within the pathway (Meek et al., 2014). This  
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7 evidence can be drawn from various sources, but typically comprises relevant studies described  
8  
9 in the literature, or results from experiments specifically designed and undertaken for the purpose  
10  
11 of AOP development. Finally, an AOP can be enhanced with quantitative linkages that allow for  
12  
13 more accuracy and surety in predicting outcomes from biomarkers of upstream key events or  
14  
15 MIEs. This progress from correlative to causative and then quantitative is paralleled by  
16  
17 concomitant decreases in uncertainty in the model predictions and increases confidence for use  
18  
19 by regulatory decision makers. In 2012 the Organisation for Economic Cooperation and  
20  
21 Development (OECD) launched the AOP Development Program, which is coordinated by the  
22  
23 OECD Extended Advisory Group on Molecular Screening and Toxicogenomics (EAG-MST).  
24  
25 The OECD guidance document and template for developing and assessing AOPs aims to ensure  
26  
27 consistency in approach and compliance with AOP standards related to content, structure and  
28  
29 presentation (OECD, 2013). An AOP Knowledge Base (AOP-KB) has also been recently  
30  
31 developed which includes an AOP-Wiki, slated for release later in 2014  
32  
33 (<http://www.aopwiki.org>). The AOP-Wiki is a web-accessible collaboration-space for AOP  
34  
35 development teams to work together in an efficient and convenient manner for the capture,  
36  
37 classification and evaluation of AOPs. It will also serve to crowd-source knowledge on a global  
38  
39 scale to refine existing AOPs and trigger the development of new ones where gaps in the AOP  
40  
41 landscape are identified. The AOP-KB is envisioned to be the primary hub for the regulatory  
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43 science community to rapidly and efficiently access AOP knowledge to serve their needs.  
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53 As a first step in applying the AOP framework to adverse neurological outcomes caused by  
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55 exposure to xenobiotics, the EU Reference Laboratory for Alternatives to Animal Testing  
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3 (EURL ECVAM) joined forces with the Safety Evaluation Ultimately Replacing Animal Testing  
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5 (SEURAT-1) consortium to organize a workshop (March 2013, Ispra). The goal of the workshop  
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7 was to identify potential AOPs relevant to neurotoxicity and developmental neurotoxicity. The  
8  
9 outlined AOPs identified during this workshop are by no means comprehensive for  
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11 neurotoxicity, however, they do provide a starting point to stimulate discussion and to which  
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13 information can and should be added in the future.  
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### 20 **Challenges for neurotoxicity AOP development**

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24 Development and use of the AOP framework for neurological outcomes following  
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26 developmental or adult exposure has been hampered by a number of serious challenges. A major  
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28 concern for neurotoxicity is a general lack of understanding of the MIEs that are causally  
29  
30 responsible for altered KEs triggering AOs. For example, the relationship between  
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32 developmental lead ( $Pb^{2+}$ ) exposure and adverse cognitive outcomes in children is well described  
33  
34 (Sanders et al., 2009); however, the initial molecular interactions between  $Pb^{2+}$  molecules and  
35  
36 cellular targets that are causatively linked to adverse cognitive outcomes (e.g. IQ) are still not  
37  
38 well understood. The lack of a known pathobiology of many neurodevelopmental disorders  
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40 hampers AOPs development. For example, where is the locus of IQ loss from developmental  
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42 lead exposure? The same can be said for a wide number of other well-known developmental and  
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44 adult neurotoxicants (e.g., methylmercury, alcohol, polychlorinated biphenyls). Additionally,  
45  
46 many human neurological disorders may have diverse pathophysiology that underlies similar  
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48 clinical phenotypes, or conversely, diverse clinical outcomes that result from similar  
49  
50 pathophysiology. For example, autism spectrum disorder (ASD) is a neurodevelopmental malady  
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52 with an increasing incidence and is more prevalent in males (McDonald and Paul, 2010).  
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3 However, it is now clear that ASD is an umbrella term for multiple disorders with overlapping  
4 clinical symptoms, suggesting that there are shared and unique pathophysiological mechanisms  
5 which have yet to be identified.  
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10 There are, however, a limited number of neurotoxic outcomes that do have well defined  
11 pathophysiological outcomes and MIEs. One example, developed as an AOP below, is acute  
12 neuronal sodium channel disruption and consequent behavioural effects, exemplified by p,p'-  
13 DDT and pyrethroids (Shafer et al., 2005) (see AOP on *Acute neurotoxic effects of pyrethroids*  
14 *mediated by disruption of voltage-gated sodium channels*). Another example are the well-known  
15 peripheral neuropathies induced by a number of chemicals, including organophosphates, carbon  
16 disulfide, pyridoxine (Vitamin B6), 2,5-hexandione and acrylamide (LoPachin and DeCaprio,  
17 2005; Rao et al., 2014) (see AOP on *Binding of certain organophosphates to NTE results in*  
18 *delayed neuropathy*).  
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31 In these cases the AOs are well-described AOs in the peripheral systems of multiple species  
32 correlatively and/or causatively linked to MIEs. For example, 2,5-hexandione forms irreversible  
33 covalent bonds (adducts) with proteins (LoPachin and DeCaprio, 2005; Graham et al., 2005).  
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38 The lack of known pathophysiology for a specific AO does not make it difficult to propose an  
39 AOP or to hypothesize MIEs. However, it does pose challenges in developing the empirical data  
40 needed to move from a proposed AOP to a causal or quantitative AOP. This has important  
41 consequences for the development and acceptance of more efficient and predictive testing  
42 methods for detecting chemicals that may lead to AOs of concern. An AOP that contains good  
43 correlative and/or causative links between the MIE or early KEs provides risk managers an  
44 increasing level of confidence to make regulatory decisions (Ankley et al., 2010). For example,  
45 the known causative relationships between estrogen receptor binding, activation of downstream  
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3 cellular ER-based signalling pathways, and adverse impacts on reproductive function facilitates  
4 the use of quantitative structure-activity relationships (QSAR) and chemical structure-based  
5 read-across models that can be used to make regulatory decisions (Schmeider et al., 2004; 2003).  
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10 Indeed, this model is already in use as part of the OECD ToolBox (Mombeli, 2012).  
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### 13 14 15 **Common key events for neurotoxic outcomes** 16

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19 Application of the AOP concept for hazard identification aims for faster, cheaper and more  
20 predictive neurotoxicity evaluation by including in vitro human-based systems in the testing  
21 strategy (NRC, 2007). Such a proposal was made for assessing the potency of compounds to  
22 induce skin sensitization, a complex procedure involving a variety of KE carried out by skin and  
23 immune cells. However, in the case of skin sensitization, the MIE triggering the AOP is the same  
24 for many skin sensitizers: covalent interaction of electrophilic substances with cellular proteins  
25 (MacKay et al., 2013). The functional and structural heterogeneity of the nervous system,  
26 coupled with the dynamics of brain development, suggests that a broad array of MIEs may be  
27 involved in adverse neurological outcomes. This complexity teamed with the previously  
28 mentioned dearth of well-accepted MIEs for developmental or adult neurotoxicity makes  
29 development of alternative test methods a serious challenge. One interim solution to this problem  
30 is the identification of downstream KEs that are common to multiple MIEs and pathways. On-  
31 going efforts to develop DNT screening methods based on common cellular phenotypes is an  
32 example of this approach (Coecke et al., 2007; Lein et al., 2007; Bal-Price et al., 2010; Crofton  
33 et al., 2011; Bal-Price et al. 2012). Alternatively, cellular signalling molecules common to  
34 multiple pathways could be utilized for assay development, which enables chemical testing in a  
35 medium- to high-throughput manner, enhancing the effectiveness and robustness of testing. This  
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3 approach might seem too reductionist for reflecting the complex issue of neurotoxicity with its  
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5 diverse MIEs and AOs. However, scientific data is needed to understand the degree to which this  
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7 type of testing strategy based on common KEs linked to the one or more AOs is able to be  
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9 predictive.

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11 For example, common KEs were identified that might possibly serve as endpoints for *in vitro*  
12  
13 neurotoxicity testing with a high predictivity for hazard potential. These include cytoskeleton  
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15 alterations (AOP V, VI, VII, VIII), impaired mitochondrial function (AOP II, V), increased  
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17 oxidative stress (AOP V, VI, VIII, X) and altered neuronal firing rates (AOP I, III, IV, V, VI,  
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19 VII, IX, X). It is important to note that some of these KEs are common to many cell and tissue  
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21 types, not just nervous system tissues, (e.g, markers of oxidative stress, mitochondrial function).  
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25 Data is needed to demonstrate how these non-neuronal specific KEs are linked to nervous system  
26  
27 specific adverse outcomes. A classic example is the generation of the toxic cation MPP<sup>+</sup> from  
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29 MPTP by glial cells, the subsequent destruction of neural dopaminergic neurons in a specific  
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31 brain region, the substantia nigra, that results in Parkinson's like symptoms. While the exact  
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33 reasons for the specificity of MPTP for these neurons remains controversial, with one  
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35 explanation that selective uptake of MPP<sup>+</sup> by membrane transporters in dopaminergic cells is  
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37 responsible for the targeting of dopamine neurons (Jenner and Marsden, 1986; Tipton and  
38  
39 Singer, 1993).  
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46 Predictivity of assay endpoints based on common KEs, as well as variable combinations of them  
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48 will need to be tested using a set of neurotoxic compounds (Crofton et al., 2011). This approach  
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50 may lead to a defined test battery for neurotoxicity evaluation.  
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52  
53 Within the proposed AOPs developed at the Workshop (see Appendices) two sub-groups can be  
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55 identified. The AOPs I, II, III, IV, V and VI are strongly related to neurotransmitter receptor  
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3 MIEs located in the cell membrane. These targets all have critical functions for  
4 neurotransmission. The AOPs VIII, VII, IX and X are more related to events associated with  
5 general molecular and cellular support or defence mechanisms. Accordingly, there will be  
6 interlinks within these two groups as well, and likely across these two groups. The process of  
7 neurotransmission is a fine-tuned, multi-event process of various ion channels and receptors that  
8 finally depolarizes the membrane. Neurotoxicant-induced alterations in voltage-gated ion  
9 channels (see AOP IV) directly affects the functionality of neuronal N-methyl-D-aspartate  
10 (NMDA) receptors (see AOPs I, II). During depolarisation the influx of  $Ca^{2+}$  ions is a crucial  
11 cellular process for multiple physiological functions, including synaptic plasticity (e.g. AOP II  
12 and IX). It is also a common intracellular process that may adversely affect the integrity of  
13 neural cells (see AOP V, VI, VII, VIII, IX, X). Therefore, within the description of the AOPs  
14 there are commonalities between pathways caused by the complexity of the neurobiological  
15 processes related to normal functioning of the human brain.  
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34 Further development of the ten AOPs outlined here, as well as development of new AOPs for  
35 neurotoxicity require generation of new data as well as mining existing data. In this regard it is of  
36 utmost importance that the test systems mimic human physiology as closely as possible (NRC,  
37 2007), i.e. co-culture of neurons and glia cells, expression and sensitivity of receptors,  
38 presence/absence of signalling molecules and pathways especially in a spacio-temporal context.  
39 Development and use of these models must also account for inter-species differences in brain  
40 physiology as responses to the same compounds may differ between human and rodent *in vitro*  
41 models (Gassmann et al., 2010; Harrill et al., 2011).  
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### Considering life stage-specific susceptibility in neurotoxicity AOPs

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8 Ideally, AOPs for neurotoxicity should consider specific life stages such as development or aging  
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10 as significant age-related susceptibilities in response to chemical exposure are well documented  
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12 (Rice and Barone, 2000; Landrigan et al., 2010). The development of AOPs that are based on life  
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14 stage-specific KEs in nervous system development and aging, AOP-dependent hazard and risk  
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16 assessment, should include not just the fetal and infants, but also juveniles and the elderly as  
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18 major vulnerable subpopulations.  
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22 During brain development several processes occur primarily, or exclusively during this time.  
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24 Key processes such as the commitment and differentiation of neural progenitor cells (NPCs)  
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26 followed by glial and neuronal cell proliferation, migration, differentiation into specific neuronal  
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28 and glial subtypes, axonal and dendritic outgrowth, formation and pruning of synapses,  
29  
30 myelination, programmed cell death, ontogeny of neurotransmitters and receptors, and  
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32 development of the blood brain barrier (BBB) are critical for functional brain development (Lein  
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34 et al., 2005; Bal-Price et al., 2012; Stiles and Jernigan, 2010). Disruption of any of these  
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36 processes by neurotoxic compounds may modify neuronal/glial cell function leading to adverse  
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38 alterations in neuroanatomy, neurophysiology, neurochemistry and neurobehavior. Complex and  
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40 dynamic glial/neuronal processes occur during discrete developmental windows that differ across  
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42 brain regions, and this spatiotemporal variation influences the variable sensitivity of the  
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44 developing brain to the same chemical exposure at different developmental stages (Barone et al.,  
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46 2000; Rice et al., 2000; Lein et al., 2005). Insults that occur early during CNS development have  
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48 the potential to cause more widespread impacts throughout the brain, while those occurring later  
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50 in development may only affect a specific structure or structures. For example, methylmercury  
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3 has more widespread effects if exposure occurs early in CNS development, relative to exposures  
4 that occur later in development, which result in more focused insults in the cortex and  
5 cerebellum (Burbacher et al., 1990). Another critical instance with regard to life stage-  
6 specificities is given by the developmental switch of neuronal GABAergic responses from  
7 excitation to inhibition. This switch is dependent on GABA-induced GABA<sub>A</sub> receptor activation  
8 (Ganguly et al. 2001). Therefore, interference with GABA receptors during development and  
9 after brain maturation (see AOP III) is likely to cause distinct AOs (Westerholz et al., 2010)

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20 Aging of the human brain is also characterised by processes that are unique or predominant at  
21 this life stage such as decreased neuronal cell volume, loss in cell numbers, reduced synaptic  
22 density/connectivity and declines in cognitive function (Walhovd et al., 2014). In addition,  
23 neurons show evidence of DNA damage, elevated reactive oxygen species (ROS) production,  
24 Ca<sup>2+</sup>-signalling disturbances, mitochondrial dysfunction and increased neuroinflammation with  
25 increasing age (Bishop et al., 2010). Moreover, declining hippocampal neurogenesis associated  
26 with aging of hippocampal neural stem/progenitor cells has been proposed to contribute to aging-  
27 related cognitive decline (van Wijngaarden and Franklin, 2013). Identification of NPC  
28 dysfunction in the aging hippocampal regenerative niche suggests a parallel between aging and  
29 basic processes of neurodevelopment as NPC proliferation and neuronal differentiation, that are  
30 necessary for formation and maintenance of brain, function throughout life (see AOP X).  
31 Importantly, these molecular biomarkers and biological pathways associated with aging are also  
32 implicated in neurodegenerative diseases, suggesting an overlap between biological pathways  
33 associated with ageing and neurodegenerative brain disorders.

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A representative example of such life stage-dependent neurotoxicity is anaesthetic exposure.  
There is growing concern that anaesthetics exposure causes learning impairment, memory



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3 deficits and behavioural abnormalities in young subjects, and accelerated cognitive decline in the  
4 elderly (Rohan et al. 2005; Johnson et al. 2002; Moller et al. 1998). Although the MIE(s)  
5 responsible for the AO is (are) not clear, it is suggested that the extent of neuroapoptosis,  
6 neuronal network assembly, neuro- and synaptogenesis determine the qualitative and quantitative  
7 aspects of toxicity in the developing brain (reviewed in Jevtovic-Todorovic et al., 2013). In the  
8 elderly, clinical evidence for postoperative cognitive decline is present while underlying  
9 molecular mechanisms still need to be elucidated (Jevtovic-Todorovic et al., 2013).

10  
11 An additional example of vulnerable life stages for neurotoxicity is adolescence. While  
12 developmental changes in the adolescent brain are more subtle than those in the first 4 years of  
13 life (Paus et al., 2001), magnetic resonance imaging (MRI) analyses of humans during juvenile  
14 and adolescence periods have clearly demonstrated structural and functional changes in  
15 synaptogenesis and connectivity in the human brain throughout adolescence and into adulthood  
16 (Giedd, 2004; 2008). Ethanol acts on multiple processes occurring during adolescent  
17 development (reviewed in Guerri and Pascual, 2010). It attenuates N-methyl-D-aspartate  
18 (NMDA)-mediated synaptic activity to a larger extent in the immature than in the mature  
19 hippocampus and thus, more potently inhibits the induction of long-term potentiation (LTP) in  
20 immature versus mature animals (Swartzwelder et al., 1995a; b). MIEs, cellular processes and  
21 pathways critical for brain development during adolescence may overlap with those important  
22 for development and aging of the nervous system.

23  
24 What is the relevance of life stage-susceptibility for neurotoxicity AOPs development? First,  
25 AOP developers need to think in a broader context for incorporating specific aspects of brain  
26 development and brain aging into the neurotoxicity mode of action (MoA) portfolio. This means  
27 that besides 'classical' AOP for neurotoxicity like interference with neurotransmitter receptors or  
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3 inhibition of acetylcholinesterase (AOP I-V), other pathways specific for brain development and  
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5 aging should be considered when developing AOPs for neurotoxicity. As a result, life stage-  
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7 specific key events will increase the number of AOPs for neurotoxicity. One example for such  
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9 life stage-specific issues in AOP development is the notable increase in brain oxidative stress and  
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11 neuroinflammation with age (Perluigi et al., 2013; Hsieh and Yang, 2013). These suggest  
12  
13 common key events in mitochondrial (AOP VII), inflammatory (AOP VIII) and epigenetic (AOP  
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15 X) pathways within AOPs when addressing neurotoxicity in the elderly. Interestingly, two of the  
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17 draft AOPs (AOP VII and AOP VIII) are interlinked and influence each other, rendering it  
18  
19 difficult to separate them in a clear manner (Gemma et al., 2007). Importantly, in vitro models  
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21 for key events should mimic the relevant stage of neural/glia differentiation and maturation for  
22  
23 developing AOPs specific for the developing, adolescent, adult and aging nervous systems. As  
24  
25 molecular targets and thus AOPs are numerous at all life stages and likely vary in importance  
26  
27 across life stages, the AOPs presented here are just the initial step in a process that will continue  
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29 to expand and be refined over the upcoming years as the science progresses.  
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### 39 **Future directions for development of Neurotoxicity AOPs**

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43 With few exceptions, development of AOPs for developmental and adult neurotoxicity is a  
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45 relatively recent concept. There are many different directions that could be taken as work in this  
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47 area proceeds. A goal of the Workshop was to outline the directions that might prove most  
48  
49 fruitful for the AOP concept to be employed effectively in environmental decision-making by  
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51 risk assessors and others. The following directions should be considered as high priority:  
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- Cataloguing the current state of knowledge regarding known or putative AOPs for neurotoxicity
- Identifying AOPs specific for Developmental Neurotoxicology
- Prioritizing AOP development
- Identifying KEs that are amenable to High and Medium throughput screening
- Demonstrating the utility of the AOP approach to risk assessors using case studies

Each of these priorities is discussed briefly below.

#### *Cataloguing the current state of knowledge regarding known or putative AOPs*

This workshop report presents examples of 10 draft AOPs related to neurotoxicity and/or developmental neurotoxicity, but it was outside of the scope of the workshop to attempt to identify and catalog all of the known or putative AOPs that are related to neurotoxicity. Indeed, even the AOPs presented here are to be considered first drafts that require additional data, description and detail prior to use. These AOPs range from those that are mostly complete, such as those outlining pyrethroid effects on VGSCs or GABA<sub>A</sub> receptor mediated excitotoxicity and convulsions, to those that need substantial work to better establish linkages between proposed MIEs and KEs, such as the AOP for neuroinflammation leading to neurodegeneration. As mentioned previously, the AOP-Wiki will facilitate international collaboration in developing a knowledge base related to AOPs. As an important first step in utilizing AOPs for neurotoxicity and developmental neurotoxicity in risk decisions, it will be important for researchers to populate the AOP-Wiki with examples such as those developed at this Workshop. This will help to identify the most complete AOPs, and allow identification and prioritization of data gaps that require additional data collection. In a time when resources are limited and it is not possible to

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2  
3 address every data gap to have a “complete” AOP, prioritization of which data gaps are the most  
4  
5 crucial for risk decision-making is necessary for effective and efficient prioritizing of research  
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7 needs. Further, AOPs do not necessarily need to be complete to be useful and informative for  
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9 decision-making, particularly if the data gaps and uncertainties are identified and understood.  
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### 12 13 14 15 *Identifying AOPs for Developmental Neurotoxicology*

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17 As discussed above, it is challenging to develop AOPs for developmental neurotoxicity due to  
18  
19 the complex symphony of events that occur during nervous system development. Still,  
20  
21 development of AOPs following chemical exposure during development is crucial and should be  
22  
23 a high priority given the increasing incidence of childhood neurological syndromes such as  
24  
25 autism spectrum disorders, attention deficit hyperactivity disorder (ADHD) and others that have  
26  
27 significant consequences for society (Bloom et al 2009; McDonald and Paul., 2010; Landrigan  
28  
29 et al., 2012). However, as the etiology of these disorders, and in particular the role of  
30  
31 environmental chemicals is not well understood, a more critical and useful initial goal would be  
32  
33 to develop AOPs that are linked to adverse outcomes that traditionally have been used to make  
34  
35 risk decisions for environmental chemicals. Doing so would allow researchers to take advantage  
36  
37 of the existing databases in the public literature as well as publically available databases (e.g.  
38  
39 ToxRefDB, <http://www.epa.gov/ncct/toxrefdb/>) to hypothesize and test putative AOPs for  
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41 developmental neurotoxicity. Furthermore, it will facilitate adoption of the AOP concept by risk  
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43 assessors, as they will be familiar with the described AOs.  
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### *Prioritizing AOP development*

There are thousands of macromolecules (e.g. receptors, transporters, nucleic acids, lipids) in the brain that form a vast number of potential targets for xenobiotic chemicals. If even a small percentage of these function as MIEs, the task of developing AOPs for all of these poses an almost insurmountable hurdle. One way to address this challenge is to move beyond the linear approach to modelling AOPs that has dominated past research efforts. The initial models developed for the International Programme on Chemical Safety (IPCS) mode-of-action (MOA) framework (IPCS, 2007), as well as many AOPs (Crofton and Zoeller, 2005; Watanabe et al., 2010; Bushnell et al., 2010), describe a series of one-to-one relationships, in which the MIE initiates sequential KEs eventually leading to the AO. This linear sequence likely does not capture the whole complexity of molecular and cellular biology implicated in a neurotoxic response. Thus, systems approaches should be used to develop computational models of the networks of MIEs and KEs that collectively influence the AO. For example, Kleinstreuer et al. (2013) developed a novel multicellular agent-based model of vasculogenesis using the CompuCell3D (<http://www.compuCell3d.org/>), which incorporates vascular endothelial growth factor signals, pro- and anti-angiogenic inflammatory chemokine signals, and the plasminogen activating system of enzymes and proteases to recapitulate disruption of vascular formation by environmental chemicals. Computational models of neurotoxicity that link networks of cellular and organ level systems provide one approach to reduce the complexity of the challenge of developing predictive models of neurotoxicity for use in regulatory decisions.

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3 *Identifying Key Events that lend themselves to High and Medium throughput screening*  
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5 One of the advantages of understanding an AOP is that the knowledge can be used in a  
6 predictive manner. Whether it be a simple “read across” approach using structural information  
7 about a chemical and its interactions with a molecular target (MIE) or a more quantitative  
8 approach, AOPs foster the ability to predict potential AOs for chemicals that have not been  
9 evaluated for toxicity in a test system.  
10

11 A high priority for neurotoxicity AOPs is to identify KEs that represent points of convergence  
12 across multiple AOPs and that are biological responses, which can be easily incorporated into  
13 high or medium throughput screening assays. There are clear advantages and disadvantages to  
14 this approach. KEs occurring at more apical points in AOPs (i.e., closer to the AO) will by  
15 definition give rise to screening assays that detect broad classes of chemicals, but may not have  
16 the capability of distinguishing which “upstream” AOP has been activated by a chemical or class  
17 of chemicals (Woodruff et al., 2008). By contrast, assays based on early MIEs in an AOP will be  
18 test for direct interaction between chemicals and the biological target (e.g., receptor, enzyme)  
19 and thus be more specific to individual chemicals or chemical classes. This will facilitate SAR  
20 and QSAR model development. In any event, approaches that yield rapid, reliable and high(er)  
21 throughput screening methods for detecting chemicals with the potential for neurotoxicity and  
22 developmental neurotoxicity are of high priority and data from screens that are based on AOPs  
23 will provide scientifically sound rationale to those making risk decisions about chemicals based  
24 on in vitro data.  
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3 *Demonstrate the utility of the AOP approach to risk assessors using case studies*

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6 This workshop presents 10 proposed AOPs related to neurotoxicity, but it was beyond the scope  
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8 of the workshop to finalize comprehensive AOPs or demonstrate how use for risk decision-  
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10 making. Demonstration of the real-world applicability of neurotoxicity AOPs is paramount to  
11  
12 having this concept more readily accepted and utilized by risk assessors. As such, an important  
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14 goal for the neurotoxicology research community should be to identify a small number of case  
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16 studies that demonstrate how applying an AOP approach to a risk-decision problem can improve  
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18 the speed and/or confidence in a risk decision, or allows information from one chemical to be  
19  
20 applied more broadly to an entire class of chemicals. Candidate AOPs for case studies may  
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22 include the pyrethroids/sodium channels, thyroid hormone AOP, or the GABA<sub>A</sub> or NMDA-  
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24 related AOPs; however, this has to be proven by empirical data.  
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32 **Further development of the outlined AOPs to neurotoxicity**

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36 In this report, using the OECD template and information from literature searches, initial work  
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38 has been conducted to identify and develop AOPs relevant to neurological outcomes. In all cases,  
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40 the authors were told to identify an MIE or a putative MIE, followed by responses at the cellular,  
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42 tissue, organ, organism and population level, with each AOP summarized in a flow diagram. The  
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44 presented AOPs are often based on a few well-studied model neurotoxicants and a summary of  
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46 the qualitative understanding of each AOP has been briefly described. The KEs have been  
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48 further evaluated in a correlative manner based on the available published data and the subjective  
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50 interpretation of the strength of the scientific evidence.  
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55 The AOP descriptions are based on the OECD guidelines following the rule that two anchors  
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57 should be identified: one MIE linked in a causative manner to one AO. Indeed, in most of the  
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3 described AOPs this principle has been followed. However, available scientific knowledge for  
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5 some complex AOPs (e.g AOP VIII: *Neuroinflammation*) suggests more than one putative MIE  
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7 and AO. These draft AOPs will need further development following the OECD AOP  
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9 Framework, and at thus a decision will be needed to identify the most salient MIE to link with  
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11 the AO.  
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14  
15 The main aim of the workshop was to identify a set of putative AOPs related to neurotoxicity  
16  
17 and developmental neurotoxicity that could be further elaborated. The next step for development  
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19 of these AOPs will be application of the modified Bradford Hill considerations following the full  
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21 OECD Template and Guidance on Developing and Assessing the Completeness of Adverse  
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23 Outcome Pathway (OECD, 2013). This implies evaluation of biological plausibility,  
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25 concordance of dose-response, temporal concordance, consistency, and specificity of association  
26  
27 between MIE and AO in a quantitative manner (Meek et al, 2014). These issues were outside the  
28  
29 scope of this Workshop and are thus not included in the proposed AOPs. In the near future, the  
30  
31 ultimate goal for the listed AOPs is their submission to the OECD AOP Development Program.  
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42  
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44  
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46  
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48  
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50  
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8  
9

### 10 11 12 **Declaration of Interest (DOI)** 13

14  
15 The employment affiliation of the authors is shown on the cover page. The authors have sole  
16  
17 responsibility for the writing and content of this paper. The contributing authors were  
18  
19 participants of the workshop organized by the EURL ECVAM. The external workshop  
20  
21 participants, were invited on the basis of a survey by EURL ECVAM neurotoxicity experts of  
22  
23 the latest literature to identify those with specific expertise in the relevant research fields. The  
24  
25 workshop organization was financially supported by the European Commission, including the  
26  
27 cost of travelling and per diem of all invited external experts. The strategy for preparing the  
28  
29 report, the literature selected for review, the conclusions drawn and the recommendations made  
30  
31 are exclusively the collective scientific output of the workshop participants and do not  
32  
33 necessarily represent the views of the participants' employers.  
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## Appendix: Examples of the putative AOPs for neurotoxicity

### I. Adverse Outcome Pathway on: *Binding of antagonist to an NMDAR during synaptogenesis contributes to impairment of learning and memory abilities*

*Cristina Suñol*

#### 1. Introduction

Learning and memory are processes that rely on functioning of the glutamate receptor N-methyl-D-aspartate (NMDAR), which is a postsynaptic channel protein permeable to  $\text{Na}^+$  and  $\text{Ca}^{2+}$ . Pre-synaptically released glutamate binds to  $\alpha$ -Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA)/glutamate receptor, which when repetitively activated depolarizes the postsynaptic cell, eventually leading to the relief of the  $\text{Mg}^{2+}$  block of the NMDAR. Activation of the NMDAR regulates neurodevelopment, results in long-term potentiation (LTP) and long-term depression (LTD) and affects neuronal synaptic plasticity, thought to be dependent on different receptor subunits. The crucial role of the NMDAR in synaptic plasticity is supported by the general scientific consensus on the effect of NMDAR blockade/deletion on LTP (Hassel, 2006).

All functional NMDARs are tetrameric complexes, containing the essential subunit NR1 and one or more different NR2 types (NR2A, B, C and D). The necessary subunit NR1 needs to associate with other NR2 subunits that regulate channel gating and  $\text{Mg}^{2+}$  dependency. During synaptogenesis there is a switch from NR2B to NR2A expression in the cortex and hippocampus that will be relevant for the duration of channel opening. Switching from the NR2B to the NR2A subunit is thought to underlie functional alteration of the NMDAR during synaptic maturation, and it is generally believed that activation of the NR2A and NR2B subunits results in LTP and LTD, respectively (Miwa et al., 2008). Consequently, alterations in the expression of the NR2A



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3 and NR2B subunits during synaptogenesis could affect the learning and memory processes  
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5 (Morris et al., 1986).  
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## 8 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

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10  $Pb^{2+}$  is an environmental neurotoxicant that interferes with neurotransmission during central  
11 nervous system (CNS) development. It passes the human placenta and accumulates in fetal tissue  
12 during gestation (David et al., 1972). Even low levels of exposure to  $Pb^{2+}$  seem to cause  
13 surprisingly significant functional damage to children's CNS (Lanphear et al., 2005). A RfD  
14 (reference dose) of 10 microgram/dL in blood has been considered to be protective for years  
15 however, subtle neurological effects in children have been observed at lower doses (Chiodo et  
16 al., 2004; Min et al., 2007, 2009). This suggests that a definitive RfD still needs to be established  
17 for  $Pb^{2+}$ . Therefore, preventing  $Pb^{2+}$  exposure is crucial. Exposure to  $Pb^{2+}$  has been reduced  
18 during the past decades (Braun et al., 2012) however, blood levels of lead still pose a warning  
19 mark as the values reported in several cohorts exceed 5 microgram/dL (Kim et al., 2013;  
20 Palaniappan et al., 2011). Recent studies define the critical period window for  $Pb^{2+}$  exposure to  
21 induce developmental deficits as being at prenatal and early childhood period (before two-year  
22 old) (Braun et al., 2012; Ethier et al., 2012; Wasserman et al., 2000).  
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## 40 **3. Identification of the molecular initiating event (MIE)**

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42 The MIE in this AOP is the binding of  $Pb^{2+}$  to NMDAR during synaptogenesis.  $Pb^{2+}$  acts as non-  
43 competitive, voltage-independent, NMDAR antagonist inhibiting NMDA-induced  $Ca^{2+}$  currents  
44 ( $IC_{50}$  around 1- 10  $\mu M$ ) with similar potency for NR2A- and NR2B-subunits of the receptors  
45 (Alkondon et al., 1990). It has been shown that  $Pb^{2+}$  inhibits [ $^3H$ ] MK801 binding to the  
46 NMDAR in brain homogenates ( $IC_{50}$ : 0.55  $\mu M$ ) (Lasley and Gilbert, 1999).  
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3 NMDAR sensitivity to  $Pb^{2+}$  binding is higher at the earlier developmental stages when compared  
4  
5 to mature neurons (Guilarte and Miceli, 1992; Rajanna et al., 1997), indicating the relevance of  
6  
7 this MIE to developmental neurotoxicity.  
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#### 10 **4. Identification of the responses on the cellular/tissue level that may be an adverse** 11 **outcome or linked to the final adverse outcome** 12 13

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15 Key cellular effects of the sustained exposure of primary cultures of hippocampal neurons to  
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17  $Pb^{2+}$  during synaptogenesis is manifested by the decreased expression of NR2A-subunit of  
18  
19 NMDARs at synapses and in an increased targeting of NR2B-NMDARs to dendritic spines  
20  
21 (without increased NR2B-NMDARs expression) (Neal et al., 2011; Zhang et al., 2002). It has  
22  
23 been reported that NR2A-containing NMDAR are critical for protein synthesis in dendrites,  
24  
25 which may have relevance in the control of synaptic plasticity (Tran et al., 2007). Decreased  
26  
27 expression of NR2A-containing NMDAR leads to the reduced calcium currents and decreased  
28  
29 release of glutamate in the hippocampus of  $Pb^{2+}$ -exposed rats as observed using microdialysis  
30  
31 (Lasley and Gilbert, 2002) resulting in lowered NMDAR activation. Postsynaptic NMDAR  
32  
33 activation regulates the generation and release of brain-derived neurotrophic factor (BDNF) in a  
34  
35 retrograde trans-synaptic way. Due to the reduced NMDAR receptor activation observed after  
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37 exposure of hippocampal cultured neurons to  $Pb^{2+}$  resulted in a reduction of both cellular  
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39 proBDNF protein synthesis and BDNF release (Neal et al., 2010) and reduced BDNF levels in  
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41 the rat cortex and hippocampus (Baranowska-Bosiacka et al., 2013). The decreased release of  
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43 BDNF is a critical key event as it affect neuronal processes such as survival, growth,  
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45 differentiation and synaptogenesis (Acheson et al., 1995) affecting the learning and memory  
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47 processes (Yamada et al., 2003).  
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3 Exposure to  $Pb^{2+}$  also reduced the levels of the presynaptic proteins synaptophysin and  
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5 synaptobrevin, which are proteins involved in vesicular neurotransmitter release (Neal et al.,  
6  
7 2010). The key cellular events triggered by binding of antagonist to NMDA receptor during  
8  
9 synaptogenesis are summarized in Fig. 2.  
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12 Moreover, chronic  $Pb^{2+}$  exposure of rats increases the threshold for LTP induction in the  
13  
14 hippocampus at non-maximal train stimulation (Gilbert et al., 1996). This could be linked to the  
15  
16 decreased expression of the NR2A that is involved in LTP response. The alterations in the  
17  
18 expression ratio of these two subunits (NR2A and NR2B) during synaptogenesis could affect the  
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20 learning and memory processes as it is suggested based on in vivo studies (Morris et al., 1986).  
21  
22 Indeed,  $Pb^{2+}$ -exposed rats exhibit deficits in acquisition of a water maze spatial learning task that  
23  
24 correlates with the reduction in the maintenance of in vivo hippocampal LTP (Nihei et al., 2000).  
25  
26 Some of these effects are mimicked by other NMDAR antagonists, like the decrease in NR2A  
27  
28 subunit expression by exposure to memantine during synaptogenesis in vitro (Maler et al., 2005)  
29  
30 and the reduced levels of presynaptic proteins by DL-2-amino-5-phosphonovalerate (APV) (Neal  
31  
32 et al., 2010). The modulation of hippocampus-prefrontal cortex synaptic transmission and  
33  
34 disruption of executive cognitive functions was also observed in animal model induced by  
35  
36 noncompetitive NMDAR antagonist MK-801 (Blot et al., 2013).  
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#### 43 **5. Identification of the responses on the organ level that may be an adverse outcome or** 44 45 **linked to the final adverse outcome** 46 47

48 Rats exposed to  $Pb^{2+}$  during development and resulting in blood lead concentrations close to  
49  
50 those found in exposed children showed reduced protein and mRNA expression levels of NR2A  
51  
52 subunit in the hippocampus, whereas those of NR2B were unchanged (Nihei and Guilarte, 1999;  
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54 Zhang et al., 2002). Rats chronically exposed to  $Pb^{2+}$  during the postweaning period showed  
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3 decreased MK-801 binding to the NMDAR, particularly in hippocampal regions (Cory-Slechta et  
4 al., 1997a; b). A decrease in the number of pyramidal and granule cells in the hippocampus and  
5  
6 reduced adult neurogenesis of hippocampal granule cells displaying aberrant dendritic  
7  
8 morphology was also found in vivo (Baranowska-Bosiacka et al., 2013; Verina et al., 2007;  
9  
10 Gilbert et al., 2005; Jaako-Movits et al., 2005). Prenatal exposure to the NMDAR antagonist  
11  
12 phencyclidine resulted in reduced proliferation of neuronal progenitors and decreased density of  
13  
14 glutamatergic neurons in the hippocampus decreasing glutamatergic neurotransmission showing  
15  
16 behavioral deficits in cognitive memory and sensorimotor gating until adulthood (Toriumi et al.,  
17  
18 2012).

#### 24 25 **6. Identification of the responses on the organism level that may be an adverse outcome or** 26 27 **linked to the final adverse outcome**

28  
29 Neonatal treatment with NMDAR antagonists has been reported to affect learning and memory  
30  
31 and to produce hyperlocomotion (Harris LW, et al., 2003; Kawabe et al., 2007; Kawabe and  
32  
33 Miyamoto, 2008). Likewise, Pb<sup>2+</sup> exposure during development has been reported to produce  
34  
35 deficits in acquisition and retention tasks, learning and memory functions (Bijoor et al., 2012;  
36  
37 Massaro et al., 1986), delayed synaptogenesis (McCauley et al., 1982), impaired capacity for  
38  
39 LTP (Gilbert et al., 1999) and for short-term and long-term depression (Ruan et al., 2000) .  
40  
41 Schematic representation of MIEs, cellular key events and organ/organism effects is described in  
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43 Fig. 2.  
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#### 48 49 **7. Identification of the overall effects on the population**

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51 Epidemiological studies show association of Pb<sup>2+</sup> exposure with cognitive and motor deficits  
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53 (Braun et al., 2012; Hornung et al., 2009), visual brain development (Ethier et al., 2012), ADHD  
54  
55 (Min et al., 2007), behavioral signature (Chiodo et al., 2004), antisocial behavior (Dietrich et al.,  
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3 2001) and children intelligence measured by various techniques at age 3 and 4, 5 and 7  
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5 respectively (McCarthy GCI; Wechsler Preschool and Primary Scale of Intelligence-Revised,  
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7 WPPSI-R IQ and Wechsler Intelligence Scale for Children-version III, WISC-III IQ)  
8  
9 (Wasserman et al., 2000). A magnetic resonance spectroscopy case report study reported  
10  
11 alterations in brain metabolism compatible with neuronal loss and decline in intellectual  
12  
13 functioning in a child with blood lead levels about 40 microgram/dl, which showed inappropriate  
14  
15 school learning (Trope et al., 1998) while magnetic resonance imaging studies found reduced  
16  
17 adult brain volume (Brubaker et al., 2010). Follow-up analysis of the Cincinnati Lead Study  
18  
19 Cohort (253 children) suggested that averaged lifetime blood lead concentrations in excess of 20  
20  
21 micrograms/dL were associated with deficits in performance IQ (PIQ) on the order of  
22  
23 approximately 7 points when compared to children with mean concentrations less or equal to 10  
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25 micrograms/dL (Dietrich et al., 1993) showing that link between developmental consequences of  
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27 low to moderate prenatal and postnatal lead exposure and decreased intellectual capacity of  
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29 children.  
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### 36 **8. Is the AOP specific to certain life stages (DNT or aging)?**

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38 This AOP is specific to neurodevelopmental life stage, especially when there is a shift in the  
39  
40 expression of the NR2 subunits of the NMDAR increasing the expression of the NR2A subunit  
41  
42 with respect to the NR2B subunit.  
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### 45 **9. How much are initiating and key events conserved across species?**

46  
47 This is not known. The conservation of the MIE and key events across species would depend on  
48  
49 the phylogenetic conservation of the NMDAR where  $Pb^{2+}$  interacts and the NR2B/A switch  
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51 produced during neuronal development.  
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3 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be**  
4 **considered**  
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8 As mentioned above solid epidemiological data in humans demonstrate unequivocally that  
9 learning and memory are impaired in children exposed to  $Pb^{2+}$  during development. Learning  
10 and memory are processes that rely on the right expression and functioning of NMDAR in the  
11 hippocampus and cortex during development. Several laboratories provided evidence that  
12 hippocampal LTP is affected after treatment of laboratory animals undergoing development with  
13  $Pb^{2+}$  (reviewed in Toscano and Guilarte, 2005). Strong in vitro data suggest that  $Pb^{2+}$  could act as  
14 an antagonist of the NMDAR. However, the exact molecular mechanism by which  $Pb^{2+}$  inhibits  
15 NMDAR is not completely clear. A proposed mechanism involves the  $Pb^{2+}$  interaction at a  
16 divalent cation binding site that is responsible for glycine binding (Toscano and Guilarte, 2005).  
17 In rodents,  $Pb^{2+}$  binds to NMDAR of developing brain with higher affinity than does in adult  
18 brain. Whether in humans this is the case remains to be elucidated.  
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34 Furthermore, exposure to  $Pb^{2+}$  during development results in reduced expression of NMDAR  
35 subunits in the hippocampus based on animal models (Toscano and Guilarte, 2005). It is not  
36 clear whether the decreased expression of NR2A subunit by  $Pb^{2+}$  is linked to the entire  
37 inhibition of the NMDAR activity or to the selective reduced expression of the NR2A subunit.  
38 Indeed, it has been shown that the reduction in NR2A subunit presence in dendritic spines  
39 induced by  $Pb^{2+}$  exposure is possibly due to entire inhibition of NMDAR function (not selective  
40 decrease in NR2A subunit expression) as treatment with a specific NMDAR inhibitor (APV)  
41 caused similar effects (Toscano and Guilarte, 2005). However, the mechanism behind this  
42 observation has not been fully elucidated yet. It has been found that under physiological  
43 conditions the developmental increase in NR2A subunits is mediated by calcium influx derived  
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3 by both NMDAR and L-type  $\text{Ca}^{2+}$  channels, pointing out the complexity of NMDAR subunit  
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5 expression, which is not only genetically-mediated but also activity-dependent (Toscano and  
6  
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8  
9 Guilarte, 2005).

10  
11 Further studies are also needed to determine to what extent decreased expression of NR2A  
12  
13 subunit is causality linked to impairment of learning and memory in human. Specifically needed  
14  
15 are studies that characterize both the anatomical and physiological changes that caused by altered  
16  
17 NR2A expression, and importantly, exactly how these changes result in decreased cognitive  
18  
19 function. While, knockout studies in mice showing similar effects to  $\text{Pb}^{2+}$ -exposed rats, it is  
20  
21 unlikely that  $\text{Pb}^{+}$  exposure in children is equivalent to knocking out or knocking down a gene.  
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24 Thus, critical dose dynamic studies are needed to better mimic the level of alternations in NR2A  
25  
26 likely to be found in  $\text{Pb}^{+}$  exposed children. Future elaboration of this AOP requires quantitative  
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28 description of the cellular KEs that should be shown to be linked with AO in a causative manner.  
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34 **II. Adverse Outcome Pathway on: *Binding of agonist to NMDA receptor causes***  
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36 ***excitotoxicity that mediates neuronal cell death, contributing to reduction (or loss) of***  
37  
38 ***cognitive, sensory and motor function***

39  
40 *André Schrattenholz*

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43  
44 **1. Introduction**

45  
46 NMDAR-mediated excitotoxicity is upstream of neuroinflammation and apoptosis (see AOP  
47  
48 VIII on *Multiple molecular initiating events trigger neuroinflammation leading to*  
49  
50 *neurodegeneration*) and thus has been implicated in many important human pathologies, ranging  
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52 from amyotrophic lateral sclerosis, Alzheimer's and Parkinson's diseases, depression, epilepsy,  
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54 trauma and stroke to schizophrenia.  
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3 The major excitatory neurotransmitter in the mammalian CNS is L-glutamate (Glu), activating a  
4 variety of ionotropic and metabotropic receptors. The ionotropic NMDARs are ligand-gated  
5 calcium channels tightly regulated by a complex set of endogenous ligands and ions like  $Mg^{2+}$ .  
6  
7 They are activated by Glu, glycine (Gly) and depolarization (relief of voltage-dependent  
8 magnesium block). They are redox sensors (dithiol-bridges) and allosteric proteins regulated by a  
9 variety of factors like polyamines, zinc, lipid environment and phosphorylation. The four  
10 subunits composing the receptor are coded by seven genes (NR1, NR2A-D and NR3A-B), and  
11 the eventual architecture is shaped by alternative splicing, RNA-editing and extensive  
12 posttranslational modifications, like phosphorylation and glycosylation.  
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16 The underlying functional regulation of NMDARs depends on interacting proteins and cofactors,  
17 like membrane-bound receptor tyrosine kinases, cholesterol-rich membrane domains (lipid rafts),  
18  $Ca^{2+}$ -related mitochondrial feedback-loops and sub-synaptic structural elements like post-  
19 synaptic density protein of 95 kD (PSD-95). Thus, NMDARs are at the core of highly dynamic  
20 molecular systems relevant for cognition and memory (see AOP I on: *Binding of antagonist to*  
21 *an NMDAR during synaptogenesis contributes to impairment of learning and memory abilities*)  
22 and in particular responsive to alterations of access of compounds to the CNS by dysfunction of  
23 the BBB (Schrattenholz and Soskic, 2006).  
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26  
27 For toxicology, it is important that the NMDAR-dependent molecular machinery initiates and  
28 stabilizes neuronal plasticity and thus is tightly connected to brain energy metabolism  
29 (ATP/NADH) (see AOP VII *Binding of inhibitors to the mitochondrial respiration chain*  
30 *complex I, II, III or IV or interaction of uncouplers with oxidative phosphorylation decreases or*  
31 *blocks ATP production resulting in neurodegeneration*).  
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3 Under a variety of stressful conditions it can derail towards increasingly irreversible  
4 pathophysiological conditions, all converging on  $\text{Ca}^{2+}$  homoeostasis, but with different MIE's,  
5 ranging from epigenetic events (Chandrasekar, 2013), secondary neuronal damage after e.g.  
6 organophosphate poisoning (Chen, 2012) to direct interaction of neurotoxic compounds, e.g.  
7 domoic acid (DA), methyl mercury, with glutamate transport (Khandare et al., 2013; Liu et al.,  
8 2013) or certain biphenyls with related ion channels (Westerink, 2013).

## 17 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

19 The routes of exposure leading to excitotoxicity can be via inhalation, passage of the  
20 gastrointestinal system or by skin penetration, but the toxicant has to pass the BBB, usually by  
21 passive transport by appropriate hydrophobic and moderate polar properties (Grumetto et al.,  
22 2013) or by complex (co)transport mechanisms involving e.g. certain viral proteins (Silverstein  
23 et al., 2012). The permeability of the BBB also changes under oxidative stress, ischemia and  
24 during ageing (Enciu et al., 2013). Domoic acid (DA) a natural toxin that accumulates in mussels  
25 and shellfish is an analogue of L-glutamate. Prenatal exposure to DA has been associated with  
26 damage to neurons in different brain regions, decreased levels of brain gamma-aminobutyric acid  
27 (GABA) and increased glutamate levels (Hogberg et al., 2011). It has also been shown to cross  
28 the placenta reaching the brain tissue of the fetus and accumulate in the amniotic fluid.  
29 Moreover, a higher quantity of DA remains in the milk as compared to the plasma and therefore  
30 a new born baby can be more exposed than the mother (Maucher and Ramsdell, 2005).

## 47 **3. Identification of the Molecular Initiating Event (MIE)**

49 The MIE is the prolonged binding of Glu or an analogue (plus co-agonist glycine and plus  
50 depolarization via non-NMDARs) to NMDAR leading to long lasting opening of the channel  
51 resulting in the excessive intracellular  $\text{Ca}^{2+}$  concentrations. Any chemicals that can directly  
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3 activate NMDAR or indirectly via triggering increased level of endogenous glutamate could  
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5 initiate this AOP. Over activation of NMDAR leads to  $\text{Ca}^{2+}$ -dependent kinases activation  
6  
7 triggering cascades of events resulting in neuronal cell death (Fig. 3A). If certain thresholds of  
8  
9 intracellular  $\text{Ca}^{2+}$  are breached, intrinsic apoptosis and mitochondrial transition follow  
10  
11 downstream leading to neurodegeneration as an adverse outcome (Schrattenholz and Soskic,  
12  
13 2006).  
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18 **4. Identification of the responses on the cellular/tissue level that may be an adverse**  
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20 **outcome or linked to the final adverse outcome**  
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22 NMDAR over activation results in excitotoxicity, by which neurons are damaged and finally  
23  
24 killed. It is caused by excessive stimulation of NMDARs by Glu or its analogues allowing high  
25  
26 levels of  $\text{Ca}^{2+}$  to enter the cell.  $\text{Ca}^{2+}$  influx into cells activates a number of enzymes, including  
27  
28 phospholipases, endonucleases, and proteases such as calpain. These enzymes go on to damage  
29  
30 cell structures such as components of the cytoskeleton, membrane, and DNA. At the same time  
31  
32  $\text{Ca}^{2+}$  overload leads to mitochondrial damage, overproduction of free radicals, opening of the  
33  
34 mitochondrial permeability transition pore, energy depletion, poly-ADP ribosylation, activation  
35  
36 of apoptotic signaling pathways (Fig. 3A and B) (see AOP on *Binding of inhibitors to the*  
37  
38 *mitochondrial respiration chain complex I, II, III or IV or interaction of uncouplers with*  
39  
40 *oxidative phosphorylation decreases or blocks ATP production resulting in neurodegeneration*)  
41  
42 leading to apoptosis, autophagy and up regulation of inflammatory mediators (see AOP on  
43  
44 *Multiple molecular initiating events trigger neuroinflammation leading to neurodegeneration*).  
45  
46 Additionally, the mitochondrial production of reactive oxygen species (ROS) inhibits glial  
47  
48 EAAT2 function leading to further increases in the glutamate concentration at the synaptic cleft  
49  
50 and further rises in postsynaptic  $\text{Ca}^{2+}$  levels.  
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3 In the mammalian brain NMDA receptors are also responsible for activity-dependent Hebbian  
4 behavior of synapses and the formation of synaptic plasticity via LTP of postsynaptic excitatory  
5 potentials. Thus, they represent the key protein necessary for the formation of memory and  
6 cognition.  
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### 12 **5. Identification of the responses on the organ level that may be an adverse outcome or** 13 **linked to the final adverse outcome** 14

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16 The consequence of excitotoxicity and activation of the intrinsic pathway of apoptosis is  
17 neurodegeneration. Depending on the overall individual genetic, epigenetic and ontological  
18 predisposition, the brain regions where neurodegeneration occurs first can differ dramatically,  
19 producing different adverse outcomes. For example, organophosphate poisoning will primarily  
20 affect the cholinergic system and predominantly brain regions like the hippocampus. In other  
21 cases other brain regions like substantial nigra or spinal cord neurons may be affected more  
22 severely (Rahn et al., 2012; Gonda, 2012).  
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### 34 **6. Identification of the responses on the organism level that may be the final adverse** 35 **outcome or linked to the final adverse outcome** 36 37

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39 Depending on the CNS region primarily affected, there can be cognitive, behavioral or motor  
40 deficits due to excitotoxicity induced neurodegeneration. Excitotoxicity been implicated in  
41 many important human neurodegenerative diseases such as amyotrophic lateral sclerosis  
42 (Spalloni et al., 2013), Alzheimer's (Lakhan et al., 2013) and Parkinson's diseases (Mehta et al.,  
43 2013), depression, epilepsy, trauma, stroke and schizophrenia (Beal, 1992; Deutsch et al., 2001,  
44 Duman, 2009; Cho, 2013). Another important site of neurodegeneration e.g. due to exposure to  
45 antibiotics or cytostatics are the sensory neurons of the inner ear with subsequent hearing loss  
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3 (Deavall et al., 2012; Langer et al., 2013). Schematic representation of MIE, cellular key events  
4 and organ/organism effects is described in Fig. 3B.  
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### 7. Identification of the overall effects on the population

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9  
10 There is broad agreement that oxidative stress/excitotoxicity is one of the major contributing  
11 factors underlying neurodegenerative disorders like Alzheimer's (Lakhan et al., 2013) and  
12 Parkinson's diseases (Mehta et al., 2013), Amyotrophic lateral sclerosis (ALS) (Spalloni et al.,  
13 2013), but also it has been involved in conditions such as autism (Essa et al., 2013), neuropathic  
14 pain and others (Lipton, 2005).  
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### 8. Is the AOP specific to certain life stages (DNT or aging)?

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23  
24 Indeed, the physiological roles of glutamate and other neurotransmitter receptors are changing  
25 dramatically throughout development, brain maturation and aging (Paradies et al., 2013; Groebe  
26 et al., 2010; Oh et al., 2013; Crompton, 2004). At different life stages they undergo structural  
27 changes and define in a very dynamic way the relatively narrow threshold window of  
28 intracellular  $\text{Ca}^{2+}$  concentration changes which are essential for cognitive processes, but also  
29 integrate epigenetic, nutritional, individual life time histories of toxic exposures and infections  
30 and their role in excitotoxicity and neurodegeneration. NMDAR  $\text{Ca}^{2+}$ -related threshold triggers  
31 neuronal apoptosis induced by excessive  $\text{Ca}^{2+}$  leading to neurodegeneration (adverse outcome).  
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### 9. How much are initiating and key events conserved across species?

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45 They are relatively well conserved in mammals with similar physiological roles. In lower species  
46 there are still the same conserved pathways, but have different physiological roles: glutamate  
47 receptors are e.g. peripheral in muscle of insects and not expressed in the CNS as in the case of  
48 mammals (Di Antonio, 2006).  
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## 10. Challenges for further AOP development: strength, data gaps and uncertainties to be considered

Based on the cited references in the text it can be concluded that there is a general agreement that the excitotoxicity triggered by over activation of NMDAR is capable of inducing neurodegeneration. Indeed, NMDA agonists caused excitotoxic neurodegeneration in a dose-dependent manner that was also proportional to the increase in intracellular  $\text{Ca}^{2+}$  assessed in cultured cortical neurons (Sattler et al., 1998).

However, the individual genetic risk profiles for neurodegeneration, the complexity of contributing factors over the life time of mammalian brains and in particular the dynamic role of the blood brain barrier (damaged by chronic inflammation) make it difficult to extrapolate quantitative results from in vitro and in vivo models to human neuropathologies. The corresponding systems biology and modeling of the complex interactions and feedback signaling, typical for neurotransmission, is still at an early stage. However, the computational and data acquisition tools are developing fast in a context of personalized medicine (OMICS technologies, Next Generation Sequencing).

Also, it is important to note that glutamate, like each and every other neurotransmitter not only activates ionotropic receptors like the NMDA receptor, but at the same time, in the neighboring synaptic structures, activates metabotropic G-protein-coupled receptors. These receptors trigger a variety of downstream effects with completely different time scales and contribute to the complex regulation of cellular responses. Furthermore, recent findings suggest that increased voltage-gated calcium channels (VGCCs) density or activity in specific brain regions can also augment intracellular calcium levels leading to the increased glutamate release, causing over activation of NMDA receptor, promoting neurodegeneration (Cataldi, 2013).

Another complexity of this AOP is the presence of three AOs linked to the same MIE instead of one. However, this has been done intentionally so future developers could choose one of the three AOs and elaborate further connecting a specific brain structure with a defined neurodegenerative disorder. The largest limitation of this AOP is linked to the lack of temporal concordance (i.e.  $KE_{up}$  precedes  $KE_{down}$ ). Mitochondrial dysfunction and ER stress are ubiquitous cellular process that influence and perpetuate a wide array of cellular dysfunction, including facilitating the development of neurodegenerative diseases (Mei et al., 2013). Similarly, just as collapse of  $Ca^{2+}$  homeostasis can lead to ER stress, ER stress also contribute to increased intracellular  $Ca^{2+}$  (Mei et al., 2013). These complex cellular KEs have to be further described in a causative and quantitative manner to provide confidence that they are responsible for specific neurodegenerative pathologies (AO).

### III. Adverse Outcome Pathway on: *Binding of antagonist to GABA<sub>A</sub> receptor results in hyperexcitability and convulsions*

*Cristina Suñol*

#### 1. Introduction

The amino acid  $\gamma$ -aminobutyric acid (GABA) is ubiquitously present in the mammalian CNS where it is in charge of the inhibitory transmission signals between neurons. The GABAergic system mediates a series of physiological functions. Altered GABAergic function, mainly related to hyperexcitability, has a role in neurological and psychiatric disorders in humans. The most prominent GABA receptor is the GABA<sub>A</sub> receptor (GABA<sub>A</sub>R), which is a ligand-gated Cl<sup>-</sup> channel belonging to the family of ionotropic receptors. The GABA<sub>A</sub>R is the site of action of GABA released from presynaptic neurons as well as of many neuroactive drugs, among them

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2  
3 benzodiazepines, barbiturates, ethanol, neurosteroids and anesthetics. The activation of the  
4 GABA<sub>A</sub>R by GABA or other agonists leads to an increased membrane conductance to Cl<sup>-</sup> that  
5  
6 generally induces a membrane hyperpolarization and the consequent reduction in the probability  
7  
8 of action potential firing, eventually causing neuronal inhibition (Olsen and Betz, 2006). The Cl<sup>-</sup>  
9  
10 flux is inhibited by convulsant agents like bicuculline and picrotoxin, which act as competitive  
11  
12 and non-competitive GABA<sub>A</sub>R antagonists, respectively (Krishek et al., 1996). While bicuculline  
13  
14 reduces Cl<sup>-</sup> flux by decreasing the opening frequency and mean open time of the channel by  
15  
16 binding at the GABA recognition binding, picrotoxin decreases the channel opening probability  
17  
18 by binding at separate sites that block the chloride channel.  
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## 24 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

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26  
27 Hyperexcitability symptoms like anxiety and convulsions have been reported to occur after acute  
28  
29 exposure to chlorinated pesticides belonging to the family of cyclodienes and  
30  
31 hexachlorocyclohexane, produced by both intentional and non-intentional ingestion (Moses and  
32  
33 Peter, 2010; Parbhu et al., 2009; Durukan et al., 2009). Although some of these pesticides are not  
34  
35 in use nowadays in developed countries, they are still found in human fluids and tissues (Cassidy  
36  
37 et al., 2005; Shen et al., 2007; Vizcaino et al., 2011) due to their high lipophilicity and body  
38  
39 accumulation. These lipophilic compounds easily cross the BBB and target the GABA<sub>A</sub>R in the  
40  
41 CNS.  
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45  
46 Besides picrotoxin, the chemicals that block Cl<sup>-</sup> conductance through the ion channel of GABA<sub>A</sub>  
47  
48 receptor include: pentylenetetrazol, plant toxins like cicutoxin and oenanthotoxin, and pesticides  
49  
50 such as lindane and cyclodienes (Wyrembek et al., 2010; Allan and Harris 1986; Vale et al.,  
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52 2003).  
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### 3. Identification of the molecular initiating event (MIE)

MIE is triggered by binding of antagonist to GABA<sub>A</sub>R leading to the inhibition of GABA transmission (Fig. 4A). Organochlorine cyclodienes and  $\gamma$ -hexachlorocyclohexane potently interact with the GABA<sub>A</sub>R at the picrotoxin recognition site. Competitive inhibition of [<sup>35</sup>S]TBPS binding by these compounds is induced at concentrations in the nanomolar-micromolar range (Lawrence and Casida, 1984; Pomés et al., 1993; Huang and Casida, 1996; Ratra et al., 2001). Finally, they do not directly interact with the recognition sites for GABA and benzodiazepine at the GABA<sub>A</sub>R (Vale et al., 1997). Altogether these data suggest that organochlorine cyclodienes and  $\gamma$ -hexachlorocyclohexane bind to the picrotoxin recognition site at the GABA<sub>A</sub>R. It is also noticeable that pentylenetetrazole inhibits [<sup>35</sup>S]TBPS binding (Maksay and Ticku, 1985). This drug is used as a chemoconvulsant in animal seizure models for the screening of drugs effective against myoclonic seizures in humans (Mcnamara, 2006). Finally, cicutoxin and related compounds inhibited [<sup>3</sup>H]EBOB binding at the GABA<sub>A</sub> receptor (Uwai et al., 2000).

### 4. Identification of the responses on the cellular/tissue level that may be an adverse outcome or linked to the final adverse outcome

The binding of the organochlorine pesticides to the picrotoxin recognition site results in the blocking of the chloride channel operated by GABA. Inhibition of GABA-induced currents and of GABA-induced Cl<sup>-</sup> flux has been demonstrated by means of electrophysiological and Cl<sup>-</sup> uptake (<sup>36</sup>Cl) methods in cultured neuronal cells and neural preparations (Gant et al., 1987a; Obata et al., 1988; Pomés et al., 1994; Nagata et al., 1996; Vale et al., 2003; Ikeda et al., 1998; Galofré et al., 2010). A significant correlation was found for inhibition of [<sup>35</sup>S]TBPS binding and GABA-induced Cl<sup>-</sup> flux for these compounds (Obata et al., 1988; Pomés et al., 1994). Reduced or blocked Cl<sup>-</sup> influx is leading to inhibition or decreased GABA transmission resulting in

1  
2  
3 increased excitability. Induced alterations in neuronal network function are contributing to the  
4 anxiety and can cause seizures and convulsions (Fig. 4A).  
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8 **5. Identification of the responses on the organ level that may be an adverse outcome or**  
9 **linked to the final adverse outcome**  
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11  
12 The inhibition of the major inhibitory neurotransmission system alters the balance between  
13 excitation and inhibition in the brain, as a result of changes in network activity (Zhang and Sun,  
14 2011). In this regard, an acute dose of  $\gamma$ -hexachlorocyclohexane induced an increase in the innate  
15 excitability of the granule cells in the hippocampus (Joy and Albertson 1988) and of glucose  
16 uptake in several regions of the brain (Sanfeliu et al., 1989) that is compatible with its excitatory  
17 activity. On the other hand, no major histologic alterations in the CNS have been reported after  
18 acute exposure to cyclodiene and  $\gamma$ -hexachlorocyclohexane organochlorine pesticides in animal  
19 models (Omer, 1970; Castro et al., 1992). This is not unexpected as in neurotoxicity many potent  
20 neurotoxic agents produce no morphological but functional changes (Ray, 1997).  
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34 **6. Identification of the responses on the organism level that may be an adverse outcome or**  
35 **linked to the final adverse outcome**  
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38 Inhibition or decrease of GABA neurotransmission induces alterations in neuronal network  
39 function (Fig. 4A) as the balance between excitory and inhibitory neuronal activity is disrupted  
40 resulting in seizure or convulsions observed in animal experimental models and in humans  
41 (Omer, 1970; Carvalho et al., 1991; Suñol et al., 1989; Moses and Peter, 2010; Parbhu et al.,  
42 2009; Durukan et al., 2009). Furthermore, subconvulsant doses of these pesticides reduced the  
43 intensity of electrical stimulation required to evoke seizures in amygdala kindled animals  
44 (Gilbert 1995; Gilbert and Mack 1995). Schematic representation of MIE, cellular key events  
45 and organ/organism effects is described in Fig. 4A.  
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## 7. Identification of the overall effects on the population

Several case reports have been published for the convulsant activity of organochlorine cyclodiene and  $\gamma$ -hexachlorocyclohexane pesticides. Although these effects are mainly induced after ingestion of relatively high doses of the toxicant, which is a rare condition nowadays, recent reports still attest to the existence of this exposure in humans and its toxicological consequences (Moses and Peter, 2010; Parbhu et al., 2009; Durukan et al., 2009).

## 8. Is the AOP specific to certain life stages (DNT or aging)?

There is no evidence suggesting that this AOP might be specific to certain life stages. However, there is a lack of studies regarding the effect during neurodevelopment especially at low concentrations.

## 9. How much are initiating and key events conserved across species?

The GABA<sub>A</sub>R is phylogenetically conserved across species (Garcia-Reyero et al., 2011) and the ligand binding properties at the GABA<sub>A</sub>R are similar among human, mammal, avian and fish brain (Cole et al., 1984).

## 10. Challenges for further AOP development: strength, data gaps and uncertainties to be considered

A reasonable causative correlation can be depicted for the inhibition of GABA<sub>A</sub>R (measured as inhibition of [<sup>35</sup>S]TBPS binding and of GABA-induced Cl<sup>-</sup> flux and the toxic doses (LD<sub>50</sub>) in experimental animals exposed to cyclodienes and  $\gamma$ -hexachlorocyclohexane (Fig. 4B). Also, a causative correlation has been reported for the inhibition of [<sup>35</sup>S]TBPS binding and the convulsant dose of several pentylenetetrazole analogues (Squires et al., 1984). Data on absorption distribution metabolism and excretion (ADME) of GABA antagonists are not available. In addition, it is not well-established that different expression (increase/decrease) and

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localization of specific GABA<sub>A</sub>Rs subunits that form the receptors render these same receptors more sensitive (or not) to GABA<sub>A</sub>R antagonists, something that has been demonstrated to be important in animal models of epilepsy (Sperk et al., 2004). It is also not entirely clear whether the altered neuronal network function (increased excitability) is the only GABA-related mechanism leading to anxiety. Indeed, the GABA<sub>A</sub>R-mediated currents are also known to be controlled by ion-regulatory molecules such as the neuronal Cl<sup>-</sup> and/or HCO<sub>3</sub><sup>-</sup> transporters (e.g. K-Cl co-transporter isoform 2, KCC2 and Na<sup>+</sup>-independent and Na<sup>+</sup>-dependent Cl-HCO<sub>3</sub> exchangers AE3 and NDCBE, respectively) and cytosolic carbonic anhydrases (e.g. CA2 and CA7). Furthermore, TrkB and calpain have emerged as important factors in GABAergic signaling during epileptogenesis and epilepsy. The above regulators of ionic plasticity that seem to be implicated in seizures and convulsions induction due to disturbance of GABAergic transmission should be also included in this AOP. Future elaboration of this AOP requires quantitative description of the cellular KEs that should be shown to be linked with AO in a causative manner.

#### IV. Adverse Outcome Pathway on: *Binding of Pyrethroids to Voltage-gated Sodium*

##### *Channels induces acute neurotoxicity*

*Timothy Shafer*

##### **1. Introduction**

Pyrethroid insecticides have been used for pest control for over 50 years. Consequently, much is known regarding their acute neurotoxicity. In insects as well as mammals, two distinct poisoning syndromes (Adverse Outcomes) have been identified. Type I or T type syndrome, is characterized by hyperreactivity, aggressive sparring and tremor, while type II or CS syndrome is

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3 characterized by pawing and burrowing, choreoathetosis, salivation (Verschoyle and Barnes,  
4  
5 1972; Verschoyle and Aldridge, 1980). These signs and symptoms, as well as the effects of  
6  
7 pyrethroids on behavior have been demonstrated in many different laboratories and have been  
8  
9 extensively reviewed (Gammon et al., 1981; Lawrence and Casida, 1982; Soderlund et al., 2002;  
10  
11 Wolansky and Harrill, 2008). This class of compounds has been well studied at several different  
12  
13 levels of biological organization and there is a solid database of literature to support development  
14  
15 of an adverse outcome pathway for neurotoxicity following acute exposure. By contrast, the  
16  
17 developmental neurotoxicity of pyrethroids as a class is not as well understood, and there is not a  
18  
19 sufficient database for the development of an AOP (Shafer et al., 2005). Thus, the AOP  
20  
21 described below applies only to acute neurotoxicity.  
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## 27 **2. Short characterization of the exposure to the chemicals relevant to the selected AOP**

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29 Exposure to pyrethroids can occur via a variety of routes, due to the fact that these compounds  
30  
31 are used for pest control on a variety of food crops, as well as for indoor pest control. Primary  
32  
33 routes of exposure are therefore oral and dermal. Because of discontinued uses for  
34  
35 organophosphorous (OP) pesticides in the last decade, uses of pyrethroids have increased, thus  
36  
37 exposure potential for this class of compounds has increased.  
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## 41 **3. Identification of the Molecular Initiating Event (MIE)**

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43 The molecular initiating event for pyrethroid neurotoxicity is binding of these compounds to  
44  
45 voltage-gated sodium channels (VGSC) in neurons of the central and peripheral nervous system.  
46  
47 This has been well documented in numerous laboratories and different types of preparations  
48  
49 from insects, arthropods, mammals and other species. This topic has been extensively reviewed  
50  
51 (Soderlund and Bloomquist, 1989; Vijverberg and van den Bercken, 1990; Narahashi, 1982;  
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53 1992; 1996; Bloomquist, 1993; 1996; Ray, 2001; Bradberry et al., 2005), and thus will only be  
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3 briefly summarized here. Mammalian VGSCs are a multimeric protein comprised of an alpha  
4 and two beta subunits. The alpha subunit is sufficient for channel function, and multiple isoforms  
5  
6 have been identified in mammalian neurons. VGSCs in neurons are critical for the generation and  
7  
8 propagation of action potentials, electrical signals that transmit information from one end of the  
9  
10 nerve cell to the other. These channels open in response to slight changes in voltage across the  
11  
12 neuronal cell membrane, and allow sodium ions to enter the nerve cell and depolarize the  
13  
14 membrane further. The channels then close (or "inactivate") even in the presence of ongoing  
15  
16 depolarization. The alpha subunit of VGSCs has distinct binding sites for a variety of  
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18 neurotoxins, including saxitoxin, batrachotoxin, scorpion toxin and others (Ogata and Ohishi,  
19  
20 2002). Pyrethroids bind to a site on the alpha subunit of VGSC (Trainer et al., 1997) that is  
21  
22 distinct from these other binding sites (O'Reilly et al., 2006), and this binding interferes with the  
23  
24 open and closing of VGSC by delaying the kinetics, or transitions, between different open,  
25  
26 closed and inactivated states, of the channel. When measured at the level of the entire population  
27  
28 of VGSC in an individual cell, this results in short to long-lasting "tail currents" through VGSC  
29  
30 when a depolarizing stimulus is ended. If membrane voltage is examined, depolarization under  
31  
32 normal circumstances generates a single action potential. VGSCs modified by type I compounds  
33  
34 depolarize the cell membrane above the threshold for action potential generation, resulting in a  
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36 series of action potentials (repetitive firing). Type II compounds cause greater membrane  
37  
38 depolarization, diminishing the sodium electrochemical gradient and subsequent action potential  
39  
40 amplitude. Eventually, membrane potential becomes depolarized above the threshold for action  
41  
42 potential generation (depolarization-dependent block). This is the result of sodium continuing to  
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44 enter the cell through those channels that have been modified by pyrethroids. Several lines of  
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46 evidence link this MIE to the adverse outcomes described above, including chemical structure  
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3 (Type I and II pyrethroids have different structures and alter VGSC differently), stereospecificity  
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5 (pyrethroids exist as stereospecific isomers and the more toxic isomers have greater effects on  
6  
7 VGSC), and the presence of mutations of sodium channels that are related to pyrethroid  
8  
9 resistance.  
10

#### 11 **4. Identification of the responses on the cellular/tissue level that may be an adverse** 12 **outcome or linked to the final adverse outcome** 13

14  
15 The net result of the alterations in VGSC kinetics described above is an alteration of neuronal  
16  
17 excitability. If the membrane voltage of a neuronal cell is examined, a depolarization event that  
18  
19 generates a single action potential under normal circumstances, results in a series of action  
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21 potentials (repetitive firing) when VGSCs are modified by type I compounds. This is the result of  
22  
23 the pyrethroid modified channels in the membrane causing sufficient depolarization to trigger  
24  
25 additional action potentials. By contrast, type II compounds cause greater membrane  
26  
27 depolarization, which results in a diminution of the sodium electrochemical gradient and  
28  
29 subsequent action potential amplitude. Eventually, membrane potential becomes depolarized  
30  
31 above the threshold for action potential generation (depolarization-dependent block). These  
32  
33 effects of pyrethroids have been well characterized in a number of different types of preparations  
34  
35 from insects, mammals and other species. In many of these cases, it is possible to record both the  
36  
37 VGSC currents as well as the action potential firing in the same preparation thus, the changes in  
38  
39 neuronal excitability are well-linked to the MIE. The relationship between pyrethroid-induced  
40  
41 alterations in VGSC function and disrupted membrane excitability (membrane depolarization  
42  
43 and/or changes in firing of action potentials) is well established and has been demonstrated in a  
44  
45 number of species in vitro. Using crayfish giant axon, Salgado and co-workers (Salgado et al.,  
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47 1989) demonstrated temperature-dependent actions of fenvalerate on VGSC function as well as  
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3 membrane excitability. In addition, voltage-dependent potassium currents were not altered by  
4 fenvalerate. Lund and Narahashi (1981) demonstrated that cyphenothrin disrupts sodium channel  
5 function, depolarized the membrane, and upon stimulation of the nerve, induces repetitive firing  
6 that is blocked by TTX. These same authors demonstrated that tetramethrin altered VGSC  
7 function and induced repetitive firing in squid axon membranes (Lund and Narahashi, 1982).  
8 Similarly pyrethroids alter sodium current at the node of Ranvier and depolarization and/or  
9 induce repetitive in peripheral nerves from frogs (Vijverberg and Van den Bercken, 1979,  
10 reviewed in Vijverberg and Van den Bercken, 1982). Similar findings have also been reported in  
11 mammalian neurons in vitro, wherein pyrethroids modify VGSC function and produce changes  
12 in membrane excitability in dorsal root ganglion neurons (Tabarean and Narahashi, 1998) and  
13 cerebellar Purkinje neurons (Song and Narahashi, 1996).

#### 24 25 26 27 28 29 **5. Identification of the responses on the organ level that may be an adverse outcome or** 30 **linked to the final adverse outcome** 31

32  
33 The alterations in neuronal excitability are consistent with reports from in vivo measurements of  
34 modified neurophysiological endpoints. Following deltamethrin administration the typical period  
35 of supranormal nerve excitability of compound action potentials (CAP) in rat tail nerve was  
36 increased from ~30 msec to ~400 msec (Parkin and Le Quesne, 1982). Similar changes in CAP  
37 were reported following administration of fenvalerate or allethrin (Nozaki et al., 1995). When  
38 dorsal root potentials were recorded from urethane anesthetized rats, cismethrin increased the  
39 amplitude of the potential by up to 142% (Smith, 1980). Recordings in the rat hippocampus  
40 demonstrate that a variety of pyrethroids disrupt hippocampal neurophysiology, including  
41 paired-pulse inhibition (Gilbert et al., 1989; Joy et al., 1990). Finally, EEG measurements  
42 demonstrated that administration of cypermethrin produced bursts of epileptic activity following  
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3 the first administration, and on subsequent days of administration these were accompanied by  
4 shaking, myoclones, and tonic-clonic seizures (Condés-Lara et al., 1999). Thus, data from in  
5 vitro studies indicate that pyrethroid modification of VGSC leads to changes in membrane  
6 excitability. In vivo, changes in neurophysiological measures occur that are consistent with  
7 alterations in action potentials, and correlate with behavioral changes.  
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#### 10 11 12 13 14 15 **6. Identification of the responses on the organism level that may be the final adverse** 16 **outcome or linked to the final adverse outcome** 17

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19 As described above, there are two distinct syndromes of acute pyrethroid neurotoxicity that are  
20 observed after high-dose acute exposure to pyrethroids. Data from studies on auditory startle and  
21 functional observational battery protocols confirm this profile of toxicity based on structure--  
22 pyrethroids historically categorized as Type I and Type II cause different toxicity profiles in  
23 rodents. Auditory startle studies (Crofton and Reiter, 1984; Crofton and Reiter, 1988) are also  
24 consistent with two types of pyrethroid effects; with Type I compounds producing an increase in  
25 startle amplitude (permethrin, bifenthrin, cismethrin) and Type II compounds producing a  
26 decrease in startle amplitude (deltamethrin, cypermethrin, cyfluthrin, and flucythrinate).  
27 However, some compounds that produce alterations in the auditory startle reflex parameters are  
28 not predicted by the absence or presence of the  $\alpha$ -cyano group (fenvalerate) (Wolansky and  
29 Harrill, 2008). Although this relationship between effects at the sodium channel level and  
30 behavioral changes is highly correlated, true causality is difficult to determine because of the  
31 lack of suitable methods to simultaneously assess sodium channel function at the cellular level in  
32 behaving animals. Schematic representation of MIE, cellular key events and organ/organism  
33 effects is described in Fig. 5.  
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## 7. Identification of the overall effects on the population

For humans, acute neurotoxicity following pyrethroid poisoning has little impact on the population level. Individuals who are affected typically recover if properly cared for and the most significant population effect is an association between poisoning severity and lost work time (Walters et al., 2009). There is no clear indication in the literature that there are permanent adverse effects following acute pyrethroid poisoning (Ray and Forshaw, 2000; Bradberry et al., 2005). From an ecological perspective, the most significant population effect is selection for resistant strains of insect pests. Several different point mutations have been found in the voltage-gated sodium channels of resistant insects, which supports the proposed AOP (Casida and Durkin, 2013).

## 8. Is the AOP specific to certain life stages (DNT or aging)?

Age-related differences to pyrethroid neurotoxicity have been documented in rodents (Sheets et al., 1994; Sheets, 2000; reviewed in Shafer et al., 2005). In the context of AOPs, these age related differences appear to be largely due to pharmacokinetics (Cantalamesa, 1993). With respect to the key initiating event, the functional and pharmacological diversity of sodium channel expression varies by tissue and stage of development (reviewed in Meacham et al., 2008; Mandel, 1992). What is currently unclear is whether or not these developmentally expressed subunits contribute to differential sensitivity of individuals to pyrethroids based on age. A study by Meacham et al using rodent  $Na_v1.2$  and  $Na_v1.3$  channels expressed in oocytes indicated that the developmentally expressed  $Na_v1.3$  channel is more sensitive than the adult  $Na_v1.2$  channel to modification by type II pyrethroids. However, a study by Tan and Soderlund (2009) found that the human  $Na_v1.2$  and  $Na_v1.3$  channels were not different in their sensitivity to pyrethroids.

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3 Thus, additional work is needed in this area to understand whether pharmacodynamic differences  
4 contribute to age-related sensitivity to pyrethroids.  
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### 7 8 **9. How much are initiating and key events conserved across species?** 9

10 Voltage-gated sodium channels are highly conserved across species and play a key role in  
11 control of electrical excitability of the nervous system. As evidenced by the fact that both type I  
12 and type II syndromes of pyrethroid neurotoxicity can be described in insects, mammals and  
13 other species, the overall AOP is well conserved. However, there are known differences in the  
14 susceptibility to pyrethroid neurotoxicity between mammals and insects, which can be related to  
15 the AOP. First, there are differences in the pharmacokinetics of pyrethroid metabolism that can  
16 account for some of the differential toxicity between mammals and insects (Glickman and  
17 Casida, 1982; Godin et al., 2007; reviewed in Narahashi, 1996; Soderlund et al., 2002). Second,  
18 it is well known that actions of pyrethroids on VGSC are inversely temperature-dependent  
19 (Salgado et al., 1989); thus, insects, which are cold-blooded, are more susceptible than mammals  
20 due to greater modification of VGSC by pyrethroids in the former species. Finally, molecular  
21 biology studies have demonstrated that there are species differences in critical VGSC residues  
22 that are important for pyrethroid action. For example, mutation of mammalian VGSC channels to  
23 resemble more closely insect channels can increase the sensitivity of the channel to deltamethrin  
24 by 100 fold (Vais et al., 2000). In addition, more recent evidence indicates that there are  
25 differences in the sensitivity of VGSC from humans and rodents to pyrethroids (Meacham et al.,  
26 2008; Tan et al., 2009). Recently, a much larger cross-species comparison has been completed  
27 that demonstrated good cross-species predictivity of pyrethroid toxicity based on similarities in  
28 the VGSC alpha subunit (LaLone et al., 2013).  
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## 10. Challenges for further AOP development: strength, data gaps and uncertainties to be considered

In general this AOP is well documented. It is clear that interactions with VGSC is an MIE responsible for the acute neurotoxicity of pyrethroids. However, to better link experimental data to human, research is needed on which subtypes of VGSCs are the most susceptible. The compounds which trigger the MIE have been well studied and there is a solid database of literature to support development of an AOP for neurotoxicity following acute exposure.

While this AOP is well established, there are some important knowledge gaps, uncertainties and limitations. One of the greatest uncertainties is related to the heterogeneity of VGSC in mammals; mammalian VGSC are complex, and this creates uncertainty in correlating effects on specific VGSC isoforms in vitro to in vivo symptomology caused by pyrethroid toxicity in mammals. There are at least 10 distinct genes that encode sodium channel  $\alpha$  subunit proteins (Nav1.1-Nav1.9; Nav1.X) in humans and mice (Plummer and Meisler, 1999; Goldin, 2001). In addition, four  $\beta$  subunits ( $\beta$ 1- $\beta$ 4) (Yu et al., 2003) have been identified to date. The heterogeneity of the sodium channel can also be enhanced by the alternative mRNA splicing of some isoforms (Sarao et al., 1991; Schaller et al., 1992; Gustafson et al., 1993). Differential sensitivity of these channels has been demonstrated in mammalian neurons, in vitro (Ginsburg and Narahashi, 1993; Tatebayashi and Narahashi, 1994; Choi and Soderlund, 2006). While it is clear that interactions with VGSC are pivotal in the acute neurotoxicity of pyrethroids, exactly which subtypes are the most susceptible and their relative contributions are currently not completely understood. One of the limitations of the current data set is that there is no good method to measure directly VGSC function in vivo, thus the data describing pyrethroid interactions with VGSC are based on in vitro studies. Further, as there is no readily available

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3 biomarker of pyrethroid effect on VGSC, it is difficult to compare directly between in vivo and  
4  
5 in vitro concentrations. Another major data gap is the causative link between the known  
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7 alterations in neuronal channel kinetics and the types of neurological outcomes. Type I and Type  
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9 II pyrethroids produce vastly difference behavioral signs (Wolancky and Harrill, 2012). These  
10  
11 difference may, or may not, be linked to differences in isoforms mentioned above. This gap  
12  
13 includes a lack of studies examining pyrethroid effects on neurophysiological function in vivo.  
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15 Thus it is this aspect of the AOP that has the largest uncertainty. Nevertheless, available data  
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17 support a well-established AOP.  
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24 **V. Adverse Outcome Pathway on: *Binding of certain organophosphates to NTE results in***  
25  
26 ***delayed neuropathy***

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29 *Alan Hargreaves, Anna Forsby, Mamta Behl and Magdalini Sachana*  
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32  
33 **1. Introduction**  
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35 OPs comprise a diverse group of compounds that are used extensively in pesticide formulations,  
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37 aviation fluids, lubricants and flame retardants. Certain but not all OPs used in aviation fluids or  
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39 as oil additives (e.g. tri-ortho-cresyl phosphate [TOCP]) and as insecticides (e.g. chlorpyrifos,  
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41 dichlorvos, isofenphos, methamidophos, mipafox, trichlorfon, trichlorat,  
42  
43 phosphamidon/mevinphos) have been shown to induce a central/peripheral sensory-motor distal  
44  
45 axonopathy known as OP-induced delayed neuropathy (OPIDN), the clinical symptoms of which  
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47 appear up to several weeks after exposure (Weiner and Jortner, 1999; Abou Donia and Lapadula,  
48  
49 1990; Lotti and Moretto, 2005). Pathological symptoms of OPIDN include numbness, cognitive  
50  
51 dysfunction, ataxia and muscle weakness and the extent and severity of these symptoms greatly  
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53 depend on the exposure level and intervention measures.  
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3 The primary target of OPs that is capable of inducing OPIDN is considered to be neuropathy  
4 target esterase (NTE) (Johnson, 1990). OPIDN inducers irreversibly inhibit NTE far more potent  
5 than they do for acetylcholinesterase (AChE), the primer target of OPs associated with acute  
6 neurotoxicity. This type of delayed neuropathy includes inhibition of axonal transport, impaired  
7 nerve regeneration and cytoskeletal disruption (reviewed in Hargreaves, 2012).

## 14 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

15  
16 Exposure to OPIDN inducers can occur by multiple routes, including oral or dermal uptake and  
17 inhalation (e.g. contaminated dust and spray drift) (Lotti and Moretto, 2005). This can occur via a  
18 range of sources including occupational exposure, environmental pollution, intentional or  
19 unintentional contaminated food sources, dietary intake, self-poisoning (Jokanović and  
20 Kosanović, 2010). It is worth mentioning that some OPs may need to be metabolized into a  
21 neuropathic form, as it is in the case of TOCP.

## 22 **3. Identification of the Molecular Initiating Event (MIE)**

23  
24 In the case of OPIDN, MIE has been identified as the covalent binding of inducers to the active  
25 site of the esterase NTE (Johnson, 1975; Johnson MK, 1990). However, not all OPs are capable  
26 of inducing OPIDN. It has been found that only those OPs that can cause not only inhibition but  
27 also aging of NTE can ultimately induce OPIDN (Johnson, 1975; Johnson MK, 1990). The aging  
28 process involves loss of an R-group from the phosphoryl moiety resulting in formation of a  
29 negatively charged phosphoryl group that is still covalently bonded to the active site serine of  
30 NTE (Richardson et al., 2013).

31  
32 Several other OPs can inhibit the enzyme activity of NTE without causing its aging and  
33 consequently do not produce OPIDN (Richardson et al., 2013). Both in vitro and ex vivo studies  
34 using brain tissue from treated hens supported that OP inducers can age NTE whereas other OPs

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3 do not (reviewed in Hargreaves, 2012). These non-ageable NTE inhibitors have been used in the  
4  
5 past towards the protection against OPIDN from subsequently administered neuropathic OPs as  
6  
7 they have been proved potent blockers of NTE (Richardson et al., 2013).  
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10 NTE is known to have lipid hydrolase activity and belongs to the family called patatin-like  
11  
12 phospholipase domain-containing proteins and seems to play an important role in axonal and  
13  
14 synaptic integrity (Glynn, 2013). Interestingly, mutations in the catalytic domain of NTE in  
15  
16 humans lead to the development of a condition that is comparable to OPIDN named NTE-related  
17  
18 motor neuron disorder (NTE-MND) (Richardson et al., 2013).  
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#### 22 **4. Identification of the responses on the cellular/tissue level that may be an adverse** 23 24 **outcome or linked to the final adverse outcome** 25 26

27 The binding by OPs to NTE is resulting in inhibition of its enzyme activity and disrupted  $\text{Ca}^{2+}$   
28  
29 homeostasis, activation of  $\text{Ca}^{2+}$ /calmodulin kinase, altered phosphorylation, distribution and  
30  
31 reduced levels of cytoskeletal proteins (Fig. 6). However, the cellular events that occur between  
32  
33 NTE inhibition and the clinical manifestation of AO are not completely understood. Although  
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35 there are experimental studies demonstrating that calcium channel blockers can ameliorate the  
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37 signs and symptoms of OPIDN by restoring calcium balance, the same has not been noted for  
38  
39 NTE enzymatic activity (Guilherme et al., 2012). Another KE is mitochondrial dysfunction  
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41 resulting in oxidative stress and ATP depletion, which can potentiate the disruption of  $\text{Ca}^{2+}$   
42  
43 homeostasis leading to the inhibition of axonal transport both in vitro and in vivo that severely  
44  
45 affects nerve regeneration processes (Gultekin et al., 2000; Zhang et al., 2007). OP-induced  
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47 increase in the intracellular free  $\text{Ca}^{2+}$  concentration activates calpain, which degrades structural  
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49 proteins, whose cleavage disrupts axonal transport, cell signaling and axonal integrity  
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60 (Hargreaves, 2012).

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## **5. Identification of the responses on the organ level that may be an adverse outcome or linked to the final adverse outcome**

8 Energy depletion, elevated ROS formation, disruption of Ca<sup>2+</sup> homeostasis, impaired synaptic  
9 signal transmission, cytoskeletal disruption and the ensuing impairment of axonal transport,  
10 ultimately result in distal to proximal degeneration of the nerve terminal and neuropathy. Large  
11 and thin nerve fibers in the PNS are typically damaged first but the CNS neurons are also  
12 affected in some instances (Dobbs, 2009). In the CNS, degeneration of myelin sheets and  
13 “swelling” of axons has been observed (Dobbs, 2009).  
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22 Histopathological analysis performed postmortem in humans and animals exposed to OPIDN  
23 inducers revealed axonal degeneration that initially involves focal but nonterminal areas of the  
24 axon and then spreads to damage the entire distal axon. Prior the onset of OPIDN aggregation  
25 and accumulation of neurofilaments in peripheral nerves have been described, whereas, after the  
26 appearance of OPIDN symptoms changes in mitochondria and their accumulation have been also  
27 detected (Jokanović and Kosanović, 2010), pointing out the importance of these two KEs in the  
28 pathology of the adverse outcome.  
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## **6. Identification of the responses on the organism level that may be the final adverse outcome or linked to the final adverse outcome**

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OPIDN is considered a rare neurodegenerative disorder in humans and is characterized mainly by the following symptoms due to damage in peripheral nerves (reviewed in Jokanović and Kosanović, 2010):

- 51 • Mild effects in sensory organs, e.g. “restless legs”, tingling or lost sensation in hands  
52 and feet  
53
- 54 • Pain in hands and feet, headache  
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- Muscle weakness / abnormal posture
- Ataxia and gait problems
- Cognitive dysfunction
- Behavioral effects

Physical examination of humans or animals exposed to OPIDN inducers revealed motor polyneuropathy with weakness of distal limb muscles. These findings have been further supported by electrophysiological evaluation of patients diagnosed with OPIDN showing acute denervation of affected muscles with abnormal spontaneous activity (Jokanović and Kosanović, 2010; Lotti and Moretto, 2005). Schematic representation of MIE, cellular key events and organ/organism effects is described in Fig. 6.

### **7. Identification of the overall effects on the population**

Most documented cases of exposure are through food consumption, occupational exposure and environmental pollution. For example, the first major outbreak of OPIDN was found to be caused by the substitution of castor oil with TOCP in 'Ginger Jake' health remedy, resulting in a flaccid paralysis of the extremities in thousands of individuals in the USA during the 1930s (Weiner and Jortner, 1999). Since then a substantial number of clinical cases of OPIDN have been reported and reviewed in Jokanović and Kosanović, 2010 and Lotti and Moretto, 2005.

### **8. Is the AOP specific to certain life stages (DNT or aging)?**

This AOP on OPIDN is specific to the adult life stage. Indeed, adult hens (18 weeks old) are used as animal model of choice to measure NTE and identify OPs that cause OPIDN (Doherty, 2006). In contrast, adult mice treated with known OPIDN inducers do not show clinical signs or axonal degeneration but only few swollen axons in the brain stem (Veronesi et al., 1991). Only

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3 long time of exposure (9 months) to these compounds can cause axonal degeneration and  
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6 paralysis in mice (Lapadula et al., 1985).

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8 In humans, clinical cases of OPIDN have mainly been reported in adults (Jokanovic et al., 2011).

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10 In rare cases of young individuals diagnosed with OPIDN the clinical signs were considerably  
11  
12 milder, whereas their recovery was faster and complete compared to adults (Lotti, 1992;  
13  
14 Jokanović et al., 2011). It is believed that the reason behind this difference in sensitivity to  
15  
16 develop OPIDN depending on the age is related to the repair mechanisms that seem to be more  
17  
18 active in young rather than old individuals (Glynn, 2000).

### 21 22 **9. How much are initiating and key events conserved across species?**

23  
24 NTE was identified initially in adult vertebrate neuronal tissue. However, NTE is also present in  
25  
26 a variety of non-neuronal tissues such as intestine, placenta and lymphocytes. NTE is highly  
27  
28 conserved across species including mammals, insects, nematodes, and yeast (Moser et al., 2000).

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30 The remaining KEs outlined in this AOP, are also conserved across species and observed in a  
31  
32 variety of cellular models.

### 33 34 35 36 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be** 37 38 **considered**

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40 Perhaps the clearest evidence for causal links between MIE and the final AO is that pretreatment  
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42 of experimental animals with reversible inhibitors of NTE prevents the induction of OPIDN. For  
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44 example, prior administration of certain carbamates, sulphonyl fluorides and phosphinates to  
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46 hens, cats and/or rats has been found to prevent or significantly ameliorate the symptoms of  
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48 OPIDN as determined by clinical, histopathological and/or electrophysiological measurements  
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50 (Carrington, 1989). Although capable of binding to the active site of NTE and inhibiting its  
51  
52 activity, these agents do not induce aging.

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3 Studies in genetically modified mice have shown that NTE is important for normal blood vessel  
4 and placental development and that the death of embryos is not associated with the deletion of  
5 NTE (Moser et al., 2004). On the contrary, conditional mutant strains where NTE was deleted  
6 only in neuronal tissue appeared no embryo-lethality. These animals were sacrificed at the age of  
7  
8 3-4 months and histopathological examination revealed vacuolization and a dramatic  
9 redistribution of the rough endoplasmic reticulum in the hippocampus and the thalamus, whereas  
10 loss of Purkinje cells was also detected in the cerebellum (Akossoglou et al., 2004). Studies with  
11 heterozygous ( $Nte^{+/-}$ ) showed that the presence of 40% less brain NTE than in normal mice  
12 results in clinical signs and lesions that are not detectable in cases of OPIDN (Winrow et al.,  
13 2003). For example, neurodegeneration and loss of endoplasmic reticulum are not encountered in  
14 the typical pathology of OPIDN. Similarly, hyperactivity presented in  $Nte^{+/-}$  animals is not a  
15 clinical symptom related to OPIDN. However, these controversial findings may be attributed to  
16 known resistance of mice to OPIDN (Veronesi et al., 1991) and this is probably the reason why  
17 the studies with transgenic mice cannot confirm the direct association of NTE with the AO.  
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20 Protection to clinical symptoms of OPIDN can also be given by  $Ca^{2+}$  channel blockers. These  
21 blockers given prior to OP exposure relieve symptoms of OPIDN, most importantly these drugs  
22 given after the OP are still able to alleviate clinical symptoms (reviewed in Emerick et al., 2012).  
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25 Although these experimental data shows that there is strong causal relationship between calcium  
26 levels and AO, the same is not true for the relationship between NTE inhibition and calcium.  
27 Indeed, no published data is available showing reversibility in NTE levels after treatment with  
28  $Ca^{2+}$  channel blockers, rendering this KE relationship moderate.  
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31  $Ca^{2+}$  channel blockers also prevent the increase of OP-induced calpain activity in the nerves of  
32 hens (reviewed in Song and Xie, 2012). Calpain activation has been suggested to be involved in  
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3 the onset and development of OPIDN and its reduced activation by  $\text{Ca}^{2+}$  channel blockers has  
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5 been followed by attenuation of clinical symptoms and histopathological findings related to  
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7 OPIDN (Song and Xie, 2012). However, the same review concluded that further experimental  
8  
9 work is required on proteolytic pathways to shed light on the direct or indirect role of calpain in  
10  
11 the pathogenesis of OPIDN (Song and Xie, 2012).  
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14  
15 Oxidative stress has not been extensively studied in the development of OPIDN and although  
16  
17 antioxidants are known to protect from OP acute neurotoxicity, no studies are available about  
18  
19 their protective role from OPs that induce delayed neuropathy. The same is truth for some more  
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21 KEs of this AOP, like mitochondrial and cytoskeletal dysfunction, where the causative  
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23 relationship between these changes and the NTE inhibition remains to be established.  
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30 **VI. Adverse Outcome Pathway on: *Impairment of learning and memory induced by binding***  
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32 ***of electrophilic chemicals to SH(thiol)-group of proteins and non-protein molecules in***  
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34 ***neuronal and glial cells during development***

35  
36 *Christoph van Thriel and Magdalini Sachana*  
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41 **1. Introduction**  
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43 Several, chemically unrelated neurotoxins (acrylamide, methylmercury, acetaldehyde, acrolein,  
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45 etc.) share the physicochemical property of being electrophilic (LoPachin and Barber, 2006).  
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47 Thus, they share the ability to form adducts or in other words to modify nucleophilic sulfhydryl  
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49 groups (SH- or thiol-groups) from either low- or high-molecular weight biomolecules (LoPachin  
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51 and Barber, 2006). This interaction of chemicals with SH-groups has been found to be  
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53 responsible, partially, for the disturbance of redox cell balance due to increase in reactive oxygen  
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3 species (ROS) and decrease in antioxidant capacity (Farina et al., 2011). The induced oxidative  
4 stress leads to damage of proteins, lipids and nucleic acids eventually resulting in cell death and  
5 neurodegeneration (Andersen, 2004). Furthermore, the binding to SH-groups of synaptic proteins  
6 by electrophilic neurotoxicants has also been shown to modulate many pre- and postsynaptic  
7 events of neurotransmission, disrupting  $Ca^{2+}$  signaling and finally, causing severe adverse  
8 neurobehavioral effects (LoPachin and Barber, 2006). During the development of this AOP  
9 particular emphasis was given to methylmercury (MeHg) exposure during brain development, as  
10 the key cellular events described below are involved in MeHg induced developmental  
11 neurotoxicity and its binding to and modification of SH-containing proteins.  
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## 24 **2. Short characterization of the exposure to the chemicals relevant to the selected AOP**

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26 Mercury (Hg) is available in the environment in three distinct chemical forms (elemental  
27 mercury vapor, inorganic mercury salts and organic mercury) and thus, all routes of exposure  
28 (e.g. inhalation, dermal, oral) might contribute to its uptake. Mercury compounds are mainly  
29 released in the aquatic environment from anthropogenic activities, where they undergo  
30 biomethylation causing contamination of fish. MeHg-containing fish meat represents a major  
31 source of human exposure. MeHg has high affinity to -SH groups leading to the formation of a  
32 MeHg cysteine complex (Cys-SHgMe or MeHg-S-Cys) in fish meat that influence not only the  
33 bioavailability but also the neurochemical and neurobehavioral toxicity of the parent compound  
34 (Harris et al., 2003; Berntssen et al., 2004). By mimicking the essential proteinogenic amino acid  
35 methionine, cysteinyl-bound MeHg can initially be absorbed by an organism and then be taken  
36 up by cells and redistributed into subcellular compartments. This likeness to methionine can  
37 facilitate the crossing of MeHg cysteine complex through specific barriers (e.g. BBB) via  
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3 specialized amino acid transporters (Kerper et al., 1992). Thus, this MIE can influence not only  
4  
5 the toxicodynamics but also the toxicokinetics of the electrophilic neurotoxicants.  
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### 8 **3. Identification of the Molecular Initiating Event (MIE)**

9  
10 MeHg reacts with specific cysteine residues (SH-groups) on astrocytic proteins like Glu  
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12 transporters and small molecules such as glutathione (GSH) (LoPachin and Barber, 2006; Farina  
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14 et al., 2011). MeHg interacts with SH-groups of GSH not only in astrocytes but also in neuronal  
15  
16 cells as it is the most abundant intracellular low molecular weight molecule in all organs,  
17  
18 including CNS and this interaction has been demonstrated both in vitro and in vivo (Farina et al.,  
19  
20 2011). However, this is not the only target of MeHg that leads to increased levels of reactive  
21  
22 oxygen/nitrogen species (ROS/RNS). The direct binding by MeHg to specific SH-containing  
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24 proteins in mitochondria of neuronal cells, including respiratory chain complexes and  
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26 mitochondrial creatine kinase found in vitro and in vivo further contributes to imbalances in  
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28 oxidative metabolism and increased levels of ROS/RNS (Farina et al., 2011).  
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34 Furthermore, electrophilic compounds have been proposed to interact with SH-groups of proteins  
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36 involved in pre- and post-synaptic processes related to neurotransmitter storage, release, uptake  
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38 and binding in neurons (LoPachin and Barber, 2006), thus affecting both intra- and extra-cellular  
39  
40 neurotransmitter homeostasis. In case of MeHg, Glu dyshomeostasis occurs not only by  
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42 inhibiting Glu uptake into astrocytes but also by increasing the spontaneous release of Glu  
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44 (Farina et al., 2011). Moreover, disturbance in neurotransmission by MeHg is not only limited to  
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46 Glu but extends also to GABA neurotransmission (Sadiq et al., 2012).  
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50 Finally, MeHg and other electrophilic neurotoxicants form adducts with SH-groups that also  
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52 function as acceptors for redox modulators such as nitric oxide (NO) (Taqatqeh et al., 2009) that  
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54 is known to affect various protein complexes at synapse level (e.g. NMDARs) (LoPachin and  
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3 Barber, 2006). While some of the modulatory processes (e.g. NO redox modulation) are  
4 reversible and relevant for normal physiology (Lei et al. 1992), electrophilic neurotoxins (e.g.  
5 acrylamide, acrolein) might cause long-lasting SH-group binding that subsequently causes  
6 molecular and cellular events that are not reversible. Such an effect is also described for cyanide  
7 (Sun et al., 1999). Clearly, it can be stated that targeting of SH-containing proteins and non-  
8 protein molecules of neuronal and glial cells during development can be the MIE that can  
9 potential trigger subsequent key cellular events that are described below, ultimate resulting in  
10 impairment of learning and memory.  
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#### 13 **4. Identification of the responses on the cellular/tissue level that may be an adverse** 14 **outcome or linked to the final adverse outcome** 15 16

17 The described MIE can initiate a cascade of parallel events by targeting glutamatergic synapses,  
18 permanently blocking or modifying normal neurotransmission. MeHg has been found to interact  
19 with SH-groups from proteins involved in the modulation of intracellular  $Ca^{2+}$  levels such as  
20 ligand-, voltage-gated channels and transporters that can promote or block the release, storage  
21 and uptake of neurotransmitters (Farina et al., 2011). Importantly, MeHg exposure leads to  
22 postsynaptic intracellular increase of  $Ca^{2+}$  and activation of important pathways involved in cell  
23 death, which follows the increased levels of extracellular Glu (reviewed in Ni et al., 2012). In  
24 addition, the elevated Glu levels may result also in overactivation of NMDARs leading to  
25 neuronal cell death induced by excitotoxicity (Berliocchi et al., 2005) (see AOP II on *Binding of*  
26 *agonist to NMDA receptor causes excitotoxicity that mediates neuronal cell death, contributing*  
27 *to reduction (or loss) of cognitive, sensory and motor function*).  
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30 Moreover, it is well documented that MeHg can disrupt mitochondrial structure and function by  
31 binding to the specific SH-containing proteins of the respiratory chain complexes and  
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mitochondrial creatine kinase (reviewed in Farina et al., 2011), resulting in mitochondrial dysfunction and ROS overproduction. Mitochondrial damage contributes to many pathways of toxicity, potentially leading to neurodegeneration (see AOP VII on *Binding of inhibitors to the mitochondrial respiration chain complex I, II, III or IV or interaction of uncouplers with oxidative phosphorylation decreases or blocks ATP production resulting in neurodegeneration*). This mitochondrial dysfunction further causes  $\text{Ca}^{2+}$  overload and stimulation of neuronal nitric oxide synthase (nNOS) increasing production of NO that nitrosylates a specific cysteine thiolate on both the NR1 and NR2A subunits and final blockage of NMDA receptor (LoPachin and Barber, 2006) (see AOP I on *Binding of antagonist to an NMDAR during synaptogenesis contributes to impairment of learning and memory abilities*). It has been suggested that this nitrosylation of brain proteins may be involved in neurotoxicity and neurodegenerative disorders (Brown, 2010).

Although it is well documented that disruption of  $\text{Ca}^{2+}$  homeostasis and mitochondrial function play a central role in MeHg-induced neurotoxicity, it has not been yet elucidated whether these changes are primary or secondary events after interaction of MeHg with other cellular components. Furthermore, it is not clear if more mechanisms (e.g. ROS production) rather than direct binding of MeHg to astrocyte Glu transporters interferes with the oxidative and inhibitory effect on the same transporters.

Besides the induced oxidative stress, MeHg exposure can also directly bind to SH-group of low molecular weight compounds involved in the antioxidant defense mechanisms. MeHg has been found to bind to GSH reducing the functionality of this important antioxidant in ROS scavenging, contributing further to the imbalance in oxidative metabolism and the increased

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3 levels of ROS. It is worth mentioning that CNS is particularly susceptible to oxidative insults and  
4  
5 is, therefore, very dependent on its GSH content, especially during development.  
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8 Such multiple and interactive molecular and cellular events might cause disturbance in neuronal  
9  
10 cell physiology and morphology, especially during development (e.g. reduce activity dependent  
11  
12 neuronal plasticity), resulting in the neurobehavioral phenotype of MeHg exposure.  
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15 **5. Identification of the responses on the organ level that may be an adverse outcome or**  
16  
17 **linked to the final adverse outcome**  
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20 Even without the presence of neurodegeneration, perturbation of synaptic neurotransmission by  
21  
22 redox modulation of transmitter-gated ion channels can cause adverse effects as seen in the  
23  
24 pathophysiology of seizures (Sanchez et al., 2000). Impaired GSH functioning is relevant in  
25  
26 schizophrenia models and is an early biomarker of nigral neuron degeneration (Do et al., 2009).  
27  
28 However, chronic exposure to SH-binding neurotoxins can lead to neurodegeneration caused by  
29  
30 the downstream pathways triggered by altered intracellular  $Ca^{2+}$  concentrations as well as by the  
31  
32 increased oxidation of cellular proteins, lipids and nucleic acids.  
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36 **6. Identification of the responses on the organism level that may be the final adverse**  
37  
38 **outcome or linked to the final adverse outcome**  
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41 Some electrophilic compounds such as MeHg and acrylamide that are capable of interacting with  
42  
43 functional cysteine residues of proteins are known or suspected human developmental  
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45 neurotoxicants, respectively (Ko et al., 1999; Sorgel et al., 2002; Farina et al., 2011).  
46  
47 Experimental evidence has shown that the developing CNS is more susceptible to neurotoxic  
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49 effects of MeHg than the adult brain (Grandjean and Landrigan, 2006). These findings were  
50  
51 further supported by epidemiological studies showing that prenatal or early postnatal exposure to  
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53 this electrophile can cause decrease in IQ, mental retardation, dose-related impairments in  
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3 memory, attention, language and visuospatial perception (Bellinger, 2013) or impaired test  
4 scores in standardize neuropsychological tests (White et al., 2011). Schematic representation of  
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6 MIEs, cellular KEs and organ/organism effects is described in Fig. 7.  
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## 9 10 **7. Identification of the overall effects on the population**

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12 The population effects of MeHg exposure have recently been described in a review of several  
13 suspected DNT compounds (Bellinger, 2013). Apart from IQ loss, MeHg has also been found to  
14 contribute to neurodevelopmental disorders, including autism (Kern et al., 2012), defective  
15 learning and memory processes and Attention deficit hyperactivity disorder (ADHD) (Bellinger,  
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17 2013).  
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## 24 **8. Is the AOP specific to certain life stages (DNT or aging)?**

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26 Since some functions of the various receptors change across certain life stages (e.g. GABAergic  
27 signaling in the developing brain) the AOP might be more relevant for particular stages of  
28 development. Aging is associated with a lot of neurobiological changes including the expression  
29 of glutamatergic receptors (Hof et al., 2002) and thus, this AOP might also be relevant during  
30 aging.  
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38 In astrocytes, GSH production is dependent not only on the L- $\gamma$ -glutamyl-cysteine synthase but  
39 also on available cellular Glu levels (Wu et al., 2001; Mates et al., 2002). Interestingly,  
40 astrocytes are tightly related to glutamatergic transmission and antioxidant defense (Araque and  
41 Perea, 2004; Takuma et al., 2004) and play important role during different processes of brain  
42 development, including migration and synaptogenesis, pointing out their importance in MeHg-  
43 induced developmental neurotoxicity.  
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52 The decreased GSH levels in the CNS have been reported to be more pronounced in young rather  
53 than adult rodents exposed to MeHg (Farina et al., 2011). Indeed, it is well known that the  
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3 developing brain is even more vulnerable to oxidative stress than mature brain because it  
4 contains limited amounts of protective enzymes and antioxidants and is formed by higher  
5 number of neurons than glia, while at the same time it is characterized by increased metabolic  
6 demand related to growth.  
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### 12 **9. How much are initiating and key events conserved across species?**

13 As described in the AOP I *Binding of antagonist to an NMDAR during synaptogenesis*  
14 *contributes to impairment of learning and memory abilities* and AOP II. *Binding of agonist to*  
15 *NMDA receptor causes excitotoxicity that mediates neuronal cell death, contributing to*  
16 *reduction (or loss) of cognitive, sensory and motor function*, the cellular and molecular targets  
17 (e.g. NMDARs) are well conserved across species. The same is true for the antioxidant  
18 machinery that is affected by toxin-binding to GSH (Nava et al., 2009).  
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### 29 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be** 30 **considered**

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32 For some aspects of this AOP (e.g. NMDA receptor mediated excitotoxicity) there is evidence  
33 for cause-effect relationship, however, the direct effects of electrophilic neurotoxins on  
34 regulatory sites within the synapses are not fully elucidated. Only for acrylamide solid evidence  
35 for neurotoxicity that is partly caused by redox signaling in the synaptic compartment exists  
36 (LoPachin and Gavin, 2012). For the same electrophile there also experimental studies showing  
37 that presynaptic buildup of cysteine adducts is progressive and closely correlated to the  
38 development of acrylamide neurological symptoms (LoPachin and Gavin, 2012). In the case of  
39 MeHg, although there is a good understanding of the KEs that mediate its neurotoxicity, our  
40 understanding of the primary critical targets of MeHg remains limited, meaning that it is not  
41 clear if mitochondrial dysfunction and  $\text{Ca}^{2+}$  homeostasis disruption occur prior or after the MeHg  
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3 interaction with key protein or no-protein molecules (Farina et al., 2011). As mentioned before,  
4  
5 the initial idea of this AOP is based on organic chemistry and the general principle that  
6  
7 electrophiles form covalent bonds with nucleophiles. Such an approach is relevant for  
8  
9 computational toxicity but large batteries of test compounds have not been tested with respect to  
10  
11 their affinity for regulatory cysteine residues yet. Moreover, the way in which such a covalent  
12  
13 binding would affect the functionality of the synapse and subsequently the whole brain has not  
14  
15 been studied in sufficient detail to identify causal links between the MIE and KEs.  
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20 It is important to emphasize that there are no available comparative studies among developing  
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22 and adult brain on the differential expression of specific SH-containing targets for MeHg or other  
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24 compounds that would be covered by this AOP. Additionally, studies are lacking that identify  
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26 cellular events prone to MeHg electrophilicity that are specific to critical periods of brain  
27  
28 development.  
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32 Besides the interaction of electrophilic neurotoxicants to SH-containing proteins and non-protein  
33  
34 molecules, these compounds are known to bind to another group of proteins named  
35  
36 selenoproteins much earlier and at a higher level (Farina et al., 2011), pointing out the possibility  
37  
38 for multiple MIEs in the presented AOP that could potentially be explored in the future.  
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40 Different organic selenocompounds have been proposed as potential therapeutic agents to  
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42 MeHg-induced neurotoxicity as seleno-containing intermediates are produced and interact with  
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44 MeHg in a higher affinity, protecting by this way the SH-containing biomolecules (Farina et al.,  
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48 2013).  
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3 **VII. Adverse Outcome Pathway on: *Binding of inhibitors to the mitochondrial respiration***  
4 ***chain complex I, II, III or IV or interaction of uncouplers with oxidative phosphorylation***  
5 ***decreases or blocks ATP production resulting in neurodegeneration.***  
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10 *Alicia Paini, Brigitte Landesmann, Jochem Lousse and Anna Bal-Price*  
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15 **1. Introduction**  
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17 In the CNS mitochondria play a pivotal role in neuronal and glial cell survival and cell death  
18 because they are regulators of both energy metabolism and apoptotic/necrotic pathways (Fiskum,  
19 2000; Wieloch, 2001; Friberg and Wieloch, 2002). The production of the ATP via oxidative  
20 phosphorylation (OXPHOS) is critical for maintaining ionic gradients across the cell membranes  
21 necessary for neuronal excitability and for execution of complex processes such as  
22 neurotransmission and plasticity (Kann and Kovács, 2007; Nunnari and Suomalainen, 2012). In  
23 addition, mitochondria are also involved in numerous other cellular functions including  $\text{Ca}^{2+}$   
24 signalling, steroid synthesis (Kang and Pervaiz, 2012), lipid and phospholipid metabolism, and  
25 the biosynthesis of essential intermediates, including heme and iron-sulfur clusters (Green, 1998;  
26 Hajnóczky et al., 2006; McBride et al., 2006). They also contribute to various cellular stress  
27 responses, such as deregulation of cellular  $\text{Ca}^{2+}$  homeostasis (Graier et al., 2007), ROS  
28 production and release of pro-apoptotic factors (Nunnari and Suomalainen, 2012). In general  
29 mitochondrial dysfunction is considered to be an early event in neurotoxicity. Exposure to  
30 xenobiotics can cause mitochondrial damage leading to decreased (or entirely blocked) ATP  
31 production triggering a cascade of events culminating in apoptotic and/or necrotic neuronal cell  
32 death.  
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The mitochondrion is formed by an outer and inner membrane, which create two separate compartments, the internal matrix and an intermembrane space. The electron transport chain (ETC) (Fig. 8A), embedded in the inner membrane, transfers electrons from the matrix into the complexes (I-IV). The electrons lose free energy at each step (complex I, III, and IV), generating a proton gradient used for the production of ATP in the matrix by complex V (Alberts et al., 2002). Due to their structural and functional complexity, mitochondria present multiple targets for compounds and numerous AOPs can be developed related to mitochondrial dysfunction. Mitochondrial dysfunction can be caused by direct interference with one of the ETC complexes (I-IV) (MIE) or as further downstream KE involved in different toxicity mechanisms induced by various mitochondria-independent MIEs (see AOP VIII on *Multiple molecular initiating events trigger neuroinflammation leading to neurodegeneration*). The described framework can serve as an initial base for further development of multiple AOPs related to mitochondrial dysfunction resulting in different adverse outcome depending on the affected brain structure.

## 34 **2. Short characterization of the exposure to the chemical relevant to the selected AOP**

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Several classes of chemicals, entering the body via oral, dermal, or inhalation exposure, are known to induce neurotoxicity via mitochondrial dysfunction by different mechanisms. Well-studied examples of chemicals that induce neurotoxic adverse effects via mitochondrial dysfunction are pesticides such as rotenone, paraquat and 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), which inhibit complex I (Desplats et al., 2012; Lin et al., 2012; Sava et al., 2007). Also under pathological conditions (e.g. inflammation) endogenously formed increased amounts of nitric oxide can compromise mitochondrial respiration through inhibition of Complex IV (Bal-Price and Brown, 2001) causing excitotoxicity resulting in neurodegeneration (Brown and Bal-Price, 2003) (see AOP on *Binding of agonist to NMDA*



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3 *receptor causes excitotoxicity that mediates neuronal cell death, contributing to reduction (or*  
4 *loss) of cognitive, sensory and motor function). Certain drugs, such as gramicidins, act as*  
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6 uncouplers of OXPHOS (Katsu et al., 1987; Luvisetto and Azzone, 1989). These examples  
7  
8 indicate that sources of exposure can be diverse, ranging for example from occupational  
9  
10 exposure (pesticides) to food and drug intake.  
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### 13 **3. Identification of the Molecular Initiating Event (MIE)**

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17 There are two main mechanisms underlying the MIEs that cause the common KE triggered by  
18  
19 inhibition of mitochondrial respiration. The MIE can be induced by:  
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22 **1) Chemicals that directly bind to the complexes of the respiratory chain leading to the inhibition**  
23  
24 **of ATP production. They may inhibit each of the four complexes of the ETC or ATP synthase**  
25  
26 **directly (Wallace and Starkov, 2000).**  
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29 **2) Chemicals that act through uncoupling of OXPHOS, by acting as alternative electron**  
30  
31 **acceptors since they (a) accept electrons from the ETC and feed them back at the site of higher**  
32  
33 **redox potential, or (b) become reduced by an electron carrier of the respiratory chain, producing**  
34  
35 **heat (Rousset et al., 2004).**  
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### 38 **4. Identification of the response on the cellular/tissue level that may be an adverse outcome**

39  
40 **or linked to the final adverse outcome.**

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43 Inhibition of the complexes of the respiratory chain or uncoupling of OXPHOS primarily results  
44  
45 in mitochondrial dysfunction eventually resulting in neuronal and/or glial cell death. Thereby,  
46  
47 mitochondria dysfunction triggered by inhibition of mitochondrial respiration or uncoupling of  
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49 oxidative phosphorylation results in the decreased ATP level that is linked in a causative manner  
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51 to the following events observed at the cellular level: (a) the loss of the mitochondrial membrane  
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53 potential, (b) the loss of mitochondrial protein import and protein biosynthesis, (c) reduced  
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3 activities of enzymes of the mitochondrial respiratory chain and the Krebs cycle, (d) elevated  
4 levels of reactive oxygen species (ROS) production, (e) the loss of mitochondrial motility,  
5  
6 causing a fail to re-localize to the sites with increased energy demands, such as synapses (f) the  
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8 destruction of the mitochondrial network, and (g) increased mitochondrial  $\text{Ca}^{2+}$  uptake, causing  
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10  $\text{Ca}^{2+}$  overload (Graier et al., 2007), (h) the rupture of the mitochondrial inner and outer  
11  
12 membranes, leading (i) to the release of mitochondrial pro-death factors, including cytochrome *c*  
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14 (Cyt. *c*), apoptosis-inducing factor, or endonuclease G (Braun, 2012; Martin, 2011; Correia et al.,  
15  
16 2012; Cozzolino et al., 2013).  
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21 This leads to overproduction of free radicals, activation of cell death signalling pathways and up  
22  
23 regulation of inflammatory mediators (see AOP VIII on *Multiple molecular initiating events*  
24  
25 *trigger neuroinflammation leading to neurodegeneration*). Indeed, excessive  $\text{Ca}^{2+}$  uptake into  
26  
27 mitochondria has been shown to inhibit ATP synthesis, breakdown and depletion of  
28  
29 mitochondrial phospholipids in cellular membranes (Pieczenik and Neustadt, 2007) inducing  
30  
31 mitochondrial permeability transitions pore (mPTP) opening. This results in mitochondrial  
32  
33 swelling, which can lead to the release of proapoptotic proteins such as cytochrome *c*,  
34  
35 Smac/Diablo, HtrA2/Omi, etc., triggering caspase-dependent apoptosis. Severe mitochondrial  
36  
37 cytochrome *c* release is also a precursor of necrotic cell death (Lewen et al., 2000). Whether a  
38  
39 cell dies by apoptosis or necrosis following a specific compound treatment, depends on cell  
40  
41 type and ATP levels (Bal-Price and Brown, 2000). Low levels of ATP (< 30% of control) inhibit  
42  
43 apoptotic cell death in favour of necrosis.  
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## 50 51 **5. Identification of the response on the organ level**

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53 Chemicals, that act as inhibitors of mitochondrial ETC complexes or as uncouplers of OXPHOS,  
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55 can induce necrotic and/or apoptotic cell death. Induced cell death triggered by mitochondrial  
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dysfunction could contribute to the pathology of various neurodegenerative disorders depending on the extent and the affected brain structure where cell death takes place. Dopaminergic neurons in the substantia nigra (SN, pars compacta) have been found to be particularly sensitive to oxidative stress and inhibition of mitochondrial respiration is the key events that contribute to the pathological features of Parkinson's disease (PD) (Perfeito et al., 2013). Environmental chemicals such as rotenone or MPTP have been reported to induce selective degeneration of the nigrostriatal pathway leading to formation of alpha-synuclein-positive inclusions in dopaminergic neurons of the SN, possibly by inhibiting mitochondrial complex I and increasing oxidative stress (Alam and Schmidt, 2002; Fleming et al., 2004; Sherer et al., 2003). As a result, the dopamine release in the striatum is reduced affecting processes related to motor control and various cognitive functions. Recently, increased risk for PD was found in amphetamine users, possibly due to amphetamine induced mitochondrial dysfunction, oxidative stress and  $\alpha$ -synuclein aggregation (Perfeito et al., 2012). Pesticides, such as paraquat and rotenone caused selective cell death of dopaminergic neurons, resulting in specific lesions in the substantia nigra and striatum (Costello et al. 2009, Wu et al. 2013), suggesting that these toxicants may be associated with Parkinson's disease.

Oxidative stress and diminished energy metabolism also occur in the brains of Alzheimer's disease (AD) patients, mainly in such structures as locus coeruleus, frontal, occipital and mid-temporal cortex as well as caudate. In particular, permeability transition pore formation is strongly linked with neurodegenerative disorders including AD (Rao et al., 2013).

## **6. Identification of the response on the organism**

A neurotoxicant-induced decrease of dopaminergic neurons and the resulting decrease in motor control can lead to severe adverse responses in the organism. A decrease in locomotor activity

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3 and an increase in catalepsy (muscular rigidity), resulting from the degeneration of dopaminergic  
4 neurons, have been observed in animals following rotenone treatment (Alam and Schmidt, 2002;  
5 Fleming et al., 2004; Sherer et al., 2003). Since many of the responses induced by rotenone (and  
6 other complex I inhibitors, such as MTPT) in rodents are similar to the symptoms of Parkinson's  
7 disease (Betarbet et al., 2000; 2002), chemicals that act as inhibitors or uncouplers of  
8 mitochondrial respiration could strongly contribute to the pathology of Parkinson disease.  
9 However, the specific AO caused by inhibition of mitochondrial respiration or uncoupling of  
10 oxidative phosphorylation will depend on the cell type of the brain structure where these MIEs  
11 will be triggered. Here the description of the AO is linked to PD as the dopaminergic neurons are  
12 one of the most sensitive to mitochondria damage and oxidative stress suggesting that the key  
13 events described at the cellular level (Fig. 8B) could contribute to the overall pathology of the  
14 Parkinson disease, however they are other neuropathologies lined to the same MIE and KEs  
15 described in this outlined AOP. Schematic representation of MIE, cellular key events and  
16 organ/organism effects is described in Fig. 8B.

### 36 **7. Identification of the overall effects on the population**

37  
38 Mitochondrial damage is a hallmark for Alzheimer's, Parkinson's, Huntington's diseases, and  
39 amyotrophic lateral sclerosis (Martin, 2011; Correia et al., 2012; Cozzolino et al., 2013). These  
40 neurodegenerative diseases represent a real concern for public health and for health care costs.  
41 Particularly, the prevalence of patients with Alzheimer's and Parkinson's disease increases  
42 dramatically with aging (Kawas et al. 2000, de Lau and Breteler 2006). For both diseases, less  
43 than 5 percent of the cases are genetically transmitted, suggesting that environmental factors are  
44 involved in the most common idiopathic form (Tsang and Soong 2003). In humans, there is  
45 epidemiological evidence linking Parkinson's disease to exposure to pesticides such as rotenone  
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3 (Tanner et al., 2011) and paraquat (alone or in combination with other environmental  
4 contaminants, particularly maneb) (Costello et al. 2009, Berry et al. 2010, Wang et al. 2011a).  
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### 8 **8. Is the AOP specific to certain life stages (DNT or aging)?**

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10 Mitochondrial dysfunction can lead to neuropathology at any life stage (development, maturation  
11 or aging). However, the kind of adverse outcome is life-stage dependent. Therefore the  
12 development of a specific AOP related to mitochondrial dysfunction must be life stage specific  
13 and for *in vitro* testing the applied cell model and endpoints must be selected accordingly. It is  
14 well documented that mitochondria play a central role in the process of brain development and  
15 aging across different species (Bratic and Larsson, 2013; Lee and Wei, 2012). During brain  
16 development high energy demanding processes are taking place (e.g. cell proliferation, migration  
17 differentiation etc.) and decreased level of ATP will affect these critical developmental  
18 processes. A wide spectrum of alterations in mitochondria are described in the course of human  
19 aging including (a) increased disorganization of mitochondrial structure, (b) decline in  
20 mitochondrial oxidative phosphorylation (OXPHOS) function, (c) accumulation of mtDNA  
21 mutations, (d) increased mitochondrial production of ROS and (e) increased extent of oxidative  
22 damage to DNA, proteins, and lipids. Therefore adverse outcomes from chemical-induced  
23 mitochondrial damage can be further potentiated with increasing age. This age-dependence is  
24 even more pronounced in women, as estrogens and estrogen receptors play a pivotal role in  
25 regulating energy expenditures and protecting against oxidative stress in the mitochondria.  
26 Estrogen, androgen and progesterone receptors are also found in human brain (Henderson and  
27 Diaz Brinton, 2010). Peroxide production by mitochondria (in particular brain synaptic  
28 mitochondria) from males is higher than that from females of the same age and the mitochondrial  
29 glutathione content is higher in females than in males (Borras et al., 2003). Hormonal deficit in  
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3 post-menopausal women has been proposed to be one risk factor in AD since two thirds of AD  
4 patients are women (Grimm et al 2012). There is a greater incidence of PD in men than in  
5 women (Baldereschi et al., 2000), persisting across age groups (Baldereschi et al., 2000; Bower  
6 et al., 1999). Further, age at onset tends to be later in women compared to men, though more  
7 data are needed in this area (Pavon et al., 2010).

### 15 **9. How much are initiating and key events conserved across species?**

17 The key events described in Fig. 8B are general mitochondria-related events, which can be  
18 observed in animal models used for toxicity testing, as well as in humans. However, the  
19 sensitivity of different species to these chemicals in relation to critical changes in key events and  
20 resulting adverse outcomes differ and have to be studied on a case by case basis, taking into  
21 consideration which brain structure is affected.

### 29 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be 30 considered**

34 The development of neurodegenerative diseases gives some evidence for the link between  
35 mitochondrial injury and human neuropathology. The information collected on mitochondrial  
36 respiration inhibitors and uncouplers of oxidative phosphorylation (MIEs) provide correlative  
37 evidence that the described cellular key events are linked to neuronal cell death (Fig. 8B). In  
38 animals, such mitochondrial damage-related neuronal cell death is linked to different types of  
39 adverse outcomes, dependent on the brain structure that is affected. However, the causative  
40 relationship between the same key events induced by described MIEs leading in a causative  
41 manner to a specific neurodegenerative disorders in humans is not that clear. Indeed, a major  
42 unknown is how alternations in ubiquitous no-specific cellular processes such as mitochondrial  
43 and oxidative phosphorylation lead to specific adverse neurological outcomes. Additional  
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3 complexity of the issue is added by the various possible windows of exposure from  
4  
5 neurodevelopment through the aging, which might drive the neurodegenerative outcome,  
6  
7 sometimes taking place many years later on. Further development of this AOP should be focused  
8  
9 on providing dose- and time- and dynamic studies that demonstrate how these common cellular  
10  
11 events are linked to cellular changes in the specific brain structure, potentially contributing to a  
12  
13 defined neurodegenerative disorder relevant to a brain structure where neurodegeneration takes  
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15 place.  
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22 **VIII. Adverse Outcome Pathway on: *Multiple molecular initiating events trigger***  
23 ***neuroinflammation leading to neurodegeneration.***  
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26  
27 *Florianne Monnet-Tschudi and Anna Bal-Price*  
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32 **1. Introduction**  
33

34 Neuroinflammation is can be triggered by several cellular key events (Wyss-Coray and Mucke,  
35  
36 2002), such as neuronal stress, injury, or death (Kreutzberg, 1995; Kreutzberg, 1996; Monnet-  
37  
38 Tschudi et al., 2007), demyelination (Defaux et al., 2010), or direct activation of microglia and  
39  
40 astrocytes by neurotoxicants (Eskes et al., 2002; Eskes et al., 2003). When neuroinflammation  
41  
42 becomes chronic (Kraft and Harry, 2011) and/or acquires a neurodegenerative phenotype (Kigerl  
43  
44 et al., 2009), it can lead to neurodegeneration, which is the adverse outcome. Indeed  
45  
46 neuroinflammation is a component of neurodegenerative diseases such as Alzheimer's disease,  
47  
48 Parkinson's disease, Multiple Sclerosis (Neumann, 2001), playing a secondary or an active  
49  
50 primary role in the disease process (Hirsch and Hunot, 2009). As several classes of toxicants are  
51  
52 able to induce neuroinflammation, the MIEs can be multiple.  
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3 Neuroinflammation is characterized by the activation of both microglial cells and astrocytes  
4 (Graeber and Streit, 1990; Aschner, 1998, Streit et al., 1999; Monnet-Tschudi et al., 2007). When  
5  
6 activated, both glial cell types undergo changes in cell morphology and physiology, accompanied  
7  
8 by increased expression and/or release of pro-inflammatory cytokines, chemokines, eicosanoids,  
9  
10 metalloproteins, and stress proteins (Dong and Benveniste, 2001), as well as by the production of  
11  
12 reactive oxygen (ROS) and nitrogen species (RNS) (Brown and Bal-Price, 2003).

13  
14  
15 Neuroinflammation can have both neuro-protective/-reparative and neurodegenerative  
16  
17 consequences (Monnet-Tschudi et al., 2007). Under normal physiological conditions, microglial  
18  
19 cells participate in surveillance of immune status (Kreutzberg, 1995; Kreutzberg, 1996; Aloisi,  
20  
21 2001; Rivest, 2009) and of neuronal integrity (Nimmerjahn et al., 2005). In the event of brain  
22  
23 cell stress or injury, microglial and astrocytic cells are activated. Within the spectrum of  
24  
25 microglial activation, two distinct activation states are described (Perego et al., 2011; Gordon  
26  
27 2003; Ponomarev et al., 2005; Maresz et al., 2008; Mosser and Edwards, 2008; Kigerl et al.,  
28  
29 2009): The M1 activation state is induced by interferon-gamma and/or other pro-inflammatory  
30  
31 cytokines and is characterized by increased expression of integrin alpha M (Itgam) and cluster of  
32  
33 differentiation 86 (CD86) and the release of pro-inflammatory cytokines (TNF-alpha, IL-1beta,  
34  
35 IL-6) and ED1 staining, and generally causes neurodegeneration. The M2 activation state  
36  
37 depends on the presence of IL-4 and IL-13 (Ponomarev et al., 2007; Maresz et al., 2008; Perego  
38  
39 et al., 2011) and induces the expression of mannose receptor 1 (MRC1), arginase1 (Arg 1) and  
40  
41 Ym1/2; it is involved in repair processes.

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44 Chronic neuroinflammation can induce secondary injury (Kraft and Harry, 2011), and appears to  
45  
46 play a role in the onset and pathological outcome of Alzheimer's and Parkinson's diseases  
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48 (McNaull et al., 2010; Tansey and Goldberg, 2009), motor neuron diseases, multiple sclerosis,  
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3 meningitis and AIDS dementia (Brown and Bal-Price, 2003). McNaull and coworkers (McNaull  
4 et al., 2010) suggested that early developmental onset of brain inflammation could be linked to  
5  
6 late onset of Alzheimer's disease. Several reports suggest that exposure to heavy metals during  
7  
8 development showed delayed symptoms, or cause silent damage, that are revealed only under  
9  
10 conditions that challenge the functional capacities (Stern et al., 2001; Fortune and Lurie, 2009).  
11  
12 Heavy metal exposure triggers a neuroinflammatory response (Tiffany-Castiglioni et al., 1989;  
13  
14 Charleston et al., 1994; Pompili et al., 2004; Monnet-Tschudi et al., 2007; White et al., 2007),  
15  
16 and is considered to be a risk factor in the etiology of Alzheimer's disease (Mutter et al., 2004;  
17  
18 Wu et al., 2008). Similarly, neuroinflammation was also observed following pesticide exposure  
19  
20 (Zurich et al., 2004; Binukumar et al., 2011; Mitra et al., 2011) and is associated with  
21  
22 Parkinson's disease (Wang et al., 2011a).  
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## 29 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

30  
31 The neurotoxic potential of environmental chemicals depends on their ability to cross the BBB.  
32  
33 Entry into the brain depends on exchange between three compartments, the plasma, the cerebro-  
34  
35 spinal fluid and the brain parenchyma, taking into account also possible efflux mechanisms, drug  
36  
37 binding and drug metabolism (Liu et al., 2008). Toxicants can use amino acid or ion transporters  
38  
39 to cross the BBB (Shimizu et al., 2001; Ose et al., 2010; Wang et al., 2011b; Corvino et al.,  
40  
41 2013; Farina et al., 2013).  
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46 Regarding the characterization of the exposure itself, long-term exposure to low doses of  
47  
48 toxicants is more relevant to human environmental exposure possibly linked to  
49  
50 neurodegenerative disorders. Widespread neurodegeneration can take years to develop. It was  
51  
52 shown in humans and monkeys that microglial activation and chronic neuron damage continues  
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3 for years after the initial exposure, suggesting that active pathology continues for a long time  
4  
5 after the toxin has been metabolized and eliminated (Taetzsch and Block, 2013).  
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### 8 **3. Identification of the Molecular Initiating Events (MIEs)**

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10 As several classes of toxicants are able to trigger a neuroinflammatory response, the molecular  
11  
12 initiating events can be very diverse. Binding to thiol and/or selenol containing proteins (See  
13  
14 AOP VI on *Impairment of learning and memory induced by binding of electrophilic chemicals to*  
15  
16 *SH(thiol)-group of proteins and non-protein molecules in neuronal and glial cells during*  
17  
18 *development*) leads to modification of the oxidation state of proteins and/or depletion of  
19  
20 glutathione, or interferences with the respiratory chain in mitochondria (see AOP VII on *Binding*  
21  
22 *of inhibitors to the mitochondrial respiration chain complex I, II, III or IV or interaction of*  
23  
24 *uncouplers with oxidative phosphorylation decreases or blocks ATP production resulting in*  
25  
26 *neurodegeneration*) are MIEs described for mercury and the herbicide paraquat, respectively  
27  
28 (Farina et al., 2013). They lead to oxidative stress as an intermediate cellular key event. Binding  
29  
30 of glutamate to NMDA receptors can induce either excitotoxicity (see AOP II on: *Binding of*  
31  
32 *agonist to NMDA receptor causes excitotoxicity that mediates neuronal cell death, contributing*  
33  
34 *to reduction (or loss) of cognitive, sensory and motor function*) such as described for the heavy  
35  
36 metal trimethyl tin (TMT) and kainate (Jeong et al., 2011; Little et al., 2012; Corvino et al.,  
37  
38 2013), or synapse impairment, as after lead (Pb<sup>2+</sup>) exposure, which is a potent non-competitive  
39  
40 NMDA receptor antagonist (see AOP I on *Binding of antagonist to an NMDAR during*  
41  
42 *synaptogenesis contributes to impairment of learning and memory abilities*) (Neal and Guilarte,  
43  
44 2010). Inhibition of acetylcholinesterase (AChE), as triggered by the organophosphates soman  
45  
46 and parathion (Zurich et al., 2004; Collombet et al., 2007), or other unknown molecular events  
47  
48 leading to interferences with Sonic Hedgehog, or 15-deoxy- $\Delta$  prostaglandin J<sub>2</sub> (15d-PGJ<sub>2</sub>)  
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3 signaling pathways following low dose exposure to the mycotoxin ochratoxin A (Hong et al.,  
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5 2002; Zurich et al., 2005; Sandström et al., submitted), are also possible MIEs.  
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#### 8 **4. Responses on the cellular/tissue level**

##### 9 *Effects on neurons*

10  
11 The intracellular calcium ( $\text{Ca}^{2+}$ ) overload due to glutamate dyshomeostasis and excitotoxicity  
12  
13 (see AOP II on *Binding of agonist to NMDA receptor causes excitotoxicity that mediates*  
14  
15 *neuronal cell death, contributing to reduction (or loss) of cognitive, sensory and motor function*)  
16  
17 can lead to cytoskeletal disruption or apoptosis. Such a cascade of key events has been described  
18  
19 following mercury (Sanfeliu et al., 2003) or TMT exposure (Corvino et al., 2013). Cytoskeleton  
20  
21 instability has also been observed following ochratoxin A treatment, as evidenced by a decrease  
22  
23 in the expression of the non-phosphorylated and the phosphorylated form of neurofilaments  
24  
25 heavy chain (Sandström et al., submitted). Oxidative stress caused by paraquat exposure, can  
26  
27 lead to neuronal death by apoptosis and secondary induction of a neuroinflammatory response  
28  
29 (Choi et al., 2010; Klintworth et al., 2009). Thus it is generally well accepted that neuronal stress  
30  
31 or mild neuronal injury (Nimmerjahn et al., 2005), as well as changes in excitability (Janigro and  
32  
33 Costa, 1987) are sufficient to trigger a neuroinflammatory response with a possible rescue  
34  
35 process. However, when neuronal cell death is occurring, the neuroinflammatory response has  
36  
37 the potential to exacerbate this state and lead to a sustained neurodegenerative process (Eskes et  
38  
39 al., 2003).  
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##### 48 *Effects on oligodendrocytes*

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50 Although oligodendrocytes are very sensitive to oxidative stress and to excitotoxicity (Gonsette,  
51  
52 2008), demyelination is not often described as a primary target after a neurotoxic insult, but it  
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54 may occur rather as secondary effect, due to neuronal/oligodendroglial interactions (Zoupi et al.,  
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2011). Ochratoxin A was shown to induce a demyelination by decreasing myelin basic protein and myelin oligodendrocyte glycoprotein content. It may also interfere with oligodendrocyte maturation by decreasing the expression of markers of maturation (Sandström et al., submitted).

### ***Effects on microglial cells***

The heavy metals mercury and TMT are able to activate directly microglial cells, modulating thus cytokine release (Eskes et al., 2002). Presence of microglial cells appeared to be necessary for paraquat-induced dopaminergic neurotoxicity (Wu et al., 2005; Taetzsch and Block, 2013), suggesting a direct effect of paraquat on microglia. In a histotypic environment, microglial cells may become reactive secondary to neuronal or oligodendroglial stress. Such observations were made *in vivo* following sub-clinical long term methyl mercury (Charleston et al., 1994; Charleston et al., 1996), TMT (Corvino et al., 2013) or paraquat exposure (Mangano et al., 2012) and *in vitro* after mercury and TMT treatments (Figiel and Dzwonek, 2007; Monnet-Tschudi et al., 1995a;b). Recently, ochratoxin A exposure was shown to cause microglial activation expressing the M1 degenerative phenotype with an upregulation of the pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$  and IL-6) and a downregulation of the anti-inflammatory cytokine (IL-4). This response was downstream of ochratoxin-induced effects on neurons, astrocytes and oligodendrocytes (Sandström et al., submitted).

### ***Effects on astrocytes***

It is generally accepted that cytokines (IL-1 $\beta$ , IL-6) released by reactive microglial cells induce astrocyte reactivity, called astrogliosis (Banati et al., 1993; Van Wagoner and Benveniste, 1999). Astrogliosis is characterized by an upregulation of glial fibrillary acidic protein (GFAP), a cytoskeleton protein (Eng et al., 2000). Astrogliosis was observed *in vivo* and *in vitro* after TMT (Monnet-Tschudi et al., 1995a; Monnet-Tschudi et al., 1995b; Pompili et al., 2004), mercury

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3 (Charleston et al., 1996; Monnet-Tschudi et al., 1996; Roda et al., 2008) and paraquat exposure  
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5 (McCormack et al., 2002). In addition, 27 compounds out of 86 with different chemical  
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7 properties and undefined MIEs, tested in the EU project AcuteTox, were able to induce an  
8  
9 upregulation of GFAP mRNA expression attesting for astrocyte reactivity (Zurich et al., 2013).  
10  
11 Ochratoxin A caused an atypical astrocyte reaction with a decrease of GFAP expression (Zurich  
12  
13 et al., 2005). This unusual astrocyte reactivity is accompanied by a decrease of metallothioneins  
14  
15 (Sandström et al., submitted), localized in astrocytes (Miyazaki et al., 2011). A treatment with  
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17 metallothioneins during ochratoxin exposure caused a decrease of microglial reactivity,  
18  
19 suggesting that in the case of ochratoxin the astrocyte perturbations are upstream of the  
20  
21 microglial reactivity (Sandström et al., submitted).  
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27 Following specific toxicant-induced cellular perturbations, cross-talk between the different  
28  
29 neural cells play a crucial role in the triggering, control, evolution, and consequences of  
30  
31 neuroinflammation. As example, microglial reactivity following neuronal stress can be due to a  
32  
33 loss of chemokine control (Blank and Prinz, 2013; Chapman et al., 2000; Streit et al., 2001).  
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35 Whereas demyelination- or neural cell death-induced neuroinflammation can be triggered by  
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37 early cytokines/chemokines production by the different neural cells themselves (Peferoen et al.,  
38  
39 2013) or the release of intracellular content acting on specific receptors such as DAMPS  
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41 (Damage Associated Molecular Pathways) (Marin-Teva et al., 2011). The mechanisms by which  
42  
43 activated microglia and astrocytes can kill neurons and induce/exacerbate the neurodegenerative  
44  
45 process has been suggested to include the release of nitric oxide that causes inhibition of  
46  
47 neuronal respiration, ROS and RNS production, and rapid glutamate release resulting in  
48  
49 excitotoxic death of neurons (Brown and Bal-Price, 2003; Kraft and Harry, 2011; Taetsch and  
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3 Block, 2013). These feedback loops amplify the inflammatory response and lead to a self-  
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5 sustained neuroinflammation that exacerbates the neurodegenerative process.  
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8 **5. Identification of the responses on the organ level that may be an adverse outcome or**  
9  
10 **linked to the final adverse outcome**

11  
12 Cortical regions and hippocampus show a higher sensitivity to heavy metals (Fiedorowicz et al.,  
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14 2001; Falluel-Morel et al., 2012; Schneider et al., 2012). This may be due to differential  
15  
16 accumulation of mercury, as observed following exposure to high concentrations (Hamilton et  
17  
18 al., 2011), or to differential vulnerability. Changes in genes involved in the amyloid cascade  
19  
20 related to Alzheimer's disease were observed in the cortex of monkeys following Pb<sup>2+</sup> exposure  
21  
22 early in life (Zawia and Basha, 2005; Wu et al., 2008). In addition, aggregation of the β-amyloid  
23  
24 peptide was particularly enhanced in these monkeys after re-exposure to Pb<sup>2+</sup> (Basha et al.,  
25  
26 2005). The particular sensitivity of cortical areas to heavy metal exposure together with the  
27  
28 increased amyloid peptide deposition suggest a link between heavy metal exposure and  
29  
30 Alzheimer's pathology (Mutter et al., 2004; Castoldi et al., 2008). Pesticides, such as paraquat  
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32 and rotenone caused specific lesions in the substantia nigra and striatum (Costello et al., 2009;  
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34 Wu et al., 2013), suggesting that these toxicants may be associated with Parkinson's disease.  
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41 **6. Identification of the responses on the organism level that may be an adverse outcome or**  
42  
43 **linked to the final adverse outcome**

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45 As consequence of neurotoxicant-induced alterations of brain specific regions, behavioral  
46  
47 perturbations can occur. Heavy metal exposure is associated with memory deficits (Chen et al.,  
48  
49 2012; Wang et al., 2012; Kaur and Nehru, 2013; Lam et al., 2013), which may be related to tau  
50  
51 hyperphosphorylation (Olivieri et al., 2000; Rahman et al., 2012). Such alterations give weight to  
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3 the hypothesis that heavy metals increase the risk of developing a neurodegenerative disease of  
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5 Alzheimer's type (Mutter et al., 2004).

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8 Decrease in locomotor activity and lesion in the striatum were observed following paraquat and  
9  
10 ochratoxin treatment (Sava et al., 2006; Prakash et al., 2013). In addition, paraquat exposure  
11  
12 resulted in a robust accumulation of  $\alpha$ -synuclein, reinforcing the link between pesticide exposure  
13  
14 and the development of Parkinson's disease (Costello et al., 2009; Berry et al., 2010; Wang et al.,  
15  
16 2011a). Schematic representation of MIEs, cellular key events and organ/organism effects is  
17  
18 described in Fig. 9.  
19

## 20 21 22 **7. Identification of the overall effects on the population**

23  
24 Neuroinflammation is a hallmark of Alzheimer's and Parkinson's diseases and the prevalence of  
25  
26 patients with these disease is increasing dramatically with aging (Kawas et al., 2000; de Lau and  
27  
28 Breteler, 2006). Therefore the neurodegenerative disorders represent a real concern for public  
29  
30 health. For both diseases, less than 5 percent of the cases have a genetic background, suggesting  
31  
32 that environmental factors are involved in the most abundant, idiopathic form (Tsang and Soong,  
33  
34 2003). In humans, there is epidemiological evidence linking Parkinson's disease with exposure  
35  
36 to paraquat alone or in combination with other environmental contaminants, particularly maneb  
37  
38 (Costello et al., 2009; Berry et al., 2010; Wang et al., 2011a). In Parkinson's disease,  
39  
40 inflammation is mainly associated with microglia activation that can underlie the  
41  
42 neurodegeneration of neurons in the substantia nigra and anti-inflammatory treatment in PD  
43  
44 patients exerts a neuroprotective effect.  
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## 50 51 **8. Is the AOP specific to certain life stages (DNT or aging)?**

52  
53 Neuroinflammation is not specific to brain development or aging, as it happens at both life stages  
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55 but it may be regulated differently during these two life time. There was some controversy about  
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3 the developmental regulation of neuroinflammation, with the final consensus that during  
4 development the glial reactivities depend on the extent of the insult (Morioka and Streit, 1991).  
5  
6 The triggering of an inflammatory response during early development may be related to the  
7  
8 hypothesis of Landrigan and coworkers (Landrigan et al., 2005) of early environmental origins  
9  
10 of neurodegenerative disease in later life. Aging is associated with a low-grade chronic  
11  
12 neuroinflammation (Giunta et al., 2008), which may be exacerbated in the presence of  
13  
14 environmental toxicants.  
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### 19 20 **9. How much are initiating and key events conserved across species?**

21  
22 Toxicant-induced neuroinflammation, tau hyperphosphorylation, and formation of insoluble  
23  
24 amyloid peptides have been described *in vivo* in different mammalian species, mice, rats and  
25  
26 monkeys (Charleston et al., 1994; Zawia and Basha, 2005; Wu et al., 2008; Blesa et al., 2012,  
27  
28 Corvino et al., 2013), and *in vitro* in mice and rats and in neuroblastoma cells of human origin  
29  
30 (Olivieri et al., 2000; Aschner et al., 2007; Monnet-Tschudi et al., 2007; Fiedorowicz et al.,  
31  
32 2008).  
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### 36 37 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be** 38 39 **considered**

40  
41 The fact that environmental toxicants trigger inflammatory responses is a robust observation. It is  
42  
43 also accepted that neuroinflammation contributes to the pathogenesis of neurodegenerative  
44  
45 diseases. However, The link between environmental exposures , neuroinflammation and human  
46  
47 neurodegenerative diseases is weak. Because microglial/astrocyte activation and chronic neuron  
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49 damage continues for years after initial exposure (Taetzsch and Block, 2013), and because in  
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51 Parkinson's and Alzheimer's diseases related cognitive deficits appeared only when  
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3 neurodegeneration was widespread (Marsden, 1990; Lichtenstein et al., 2010), the link between  
4  
5 toxicant exposure and neurodegenerative diseases is difficult to study.  
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8 A major main data gap is the elucidation of a causative link between toxicant-induced  
9  
10 neuroinflammation and neurodegenerative diseases. There are some correlative links, such as the  
11  
12 association of occupational exposure to paraquat and Parkinson's disease (Costello et al., 2009;  
13  
14 Berry et al., 2010; Wang et al., 2011a), or early exposure to Pb<sup>2+</sup> and the increased formation of  
15  
16 amyloid plaques (Zawia and Basha, 2005; Wu et al., 2008), or the finding of increased level of  
17  
18 mercury in brain and blood of Alzheimer's patients (Hock et al., 1998), however, it is still  
19  
20 matter of controversy. The difficulty of studying such links is due to the fact that  
21  
22 neurodegenerative diseases are complex, multifactorial, depends on gene-environment  
23  
24 interactions and have a slow temporal evolution (Sherer et al., 2002; Steece-Collier et al., 2002;  
25  
26 Tsang and Soong, 2003; Mutter et al., 2004). This AOP deserves further development which  
27  
28 should focus on providing better correlative and causative links between exposure and activation  
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30 of key-events, identification of MIEs, and linking changes to specific adverse outcomes through  
31  
32 an more robust understanding of the cellular processes by which neuroinflammation leads to  
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34 neurodegeneration.  
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43 **IX. Adverse Outcome Pathway on: *The interaction of non-dioxin-like PCBs with ryanodine***  
44  
45 ***receptors (RyRs) causes their sensitization affecting neuronal connectivity that results in***  
46  
47 ***behavioral deficits (developmental neurotoxicity)***  
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50 *Pamela J. Lein*  
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## 1. Introduction

Polychlorinated biphenyls (PCBs) are synthetic chlorinated aromatic hydrocarbons that are non-flammable, chemically stable and have high boiling points. PCBs are often classified according to their molecular structure as dioxin-like or non-dioxin-like (NDL). PCB congeners with < 2 *ortho*-chlorine substituents are typically coplanar and exhibit dioxin-like toxicity while congeners with >1 *ortho*-substituted chlorine are often non-coplanar and classified as NDL (Safe, 1993). Commercial PCB mixtures were widely used in several industries for their insulation and heat dissipating properties and these compounds were broadly incorporated into a variety of common products such as pesticide extenders, plastics, varnishes, adhesives, carbonless copy paper, newsprint, fluorescent light ballasts and caulking compounds (Ross, 2004). In the late 1970s, commercial production of PCBs was banned in many countries because of their persistence in the environment coupled with growing concerns regarding their carcinogenic potential. While environmental and human PCB levels dropped significantly between 1970 and 1995 as a result of the production ban, recent studies of temporal trends show no or only a slight decrease in PCB levels since the mid-1990s (Hornbuckle et al., 2006), Chronic low level PCB exposures remain a significant public health concern because human studies indicate an association between PCB body burdens and immune system dysfunction, cardiovascular disease, and impairment of the developing nervous system. Of these various adverse health effects, developmental neurotoxicity is considered a particularly vulnerable endpoint in PCB toxicity. Critical reviews and meta-analyses of epidemiological studies have concluded that the weight of evidence indicates a negative association between developmental exposure to environmental PCBs and measures of neuropsychological function in infancy or childhood. (Schantz et al., 2003; Carpenter, 2006; Korrick and Sagiv, 2008; Winneke, 2011).

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3 Combined *in utero* and lactational PCB exposure correlates with decreased scores on IQ tests,  
4 impaired learning and memory, psychomotor difficulties, and attentional deficits. Because of  
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6 discrepancies between studies with respect to the spectrum and persistence of adverse  
7  
8 neurobehavioral outcomes, confounding co-exposures and differences in congener profiles that  
9  
10 comprise the exposure, questions have been raised concerning the causative role of PCBs in  
11  
12 human developmental neurotoxicity (Winneke, 2011). However, experimental findings in  
13  
14 animal models confirm that developmental PCB exposure causes deficits in learning and  
15  
16 memory (Schantz et al., 1989; Hany et al., 1999; Widholm et al., 2001; Sable et al., 2006; Yang  
17  
18 et al., 2009) and sensorimotor functions (Roegge et al., 2004; Nguon et al., 2005; Powers et al.,  
19  
20 2006). More recently, it has been posited that developmental exposure to PCBs contributes to  
21  
22 increased risk of neurodevelopmental disorders (Grandjean and Landrigan, 2006; Landrigan,  
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24 2010; Landrigan et al., 2012; Stamou et al., 2013); however, there are as yet no epidemiological  
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26 data to support this hypothesis.  
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## 33 34 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

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36 Humans are exposed to PCBs via oral, inhalation and dermal routes of exposure, and since PCBs  
37  
38 readily cross the placenta and are concentrated in breast milk, fetuses and infants are at risk from  
39  
40 *in utero* and lactational exposures, respectively. Environmental PCB contamination is not static:  
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42 mass flux studies demonstrate that large quantities of PCBs are deposited and volatilized every  
43  
44 year from Lake Michigan (Hornbuckle et al., 2006). PCBs in the environment bioaccumulate  
45  
46 and biomagnify up the food chain, thus, consumption of contaminated meat, fish, poultry and  
47  
48 dairy products is a predominant route of human exposure. Exposure also occurs via secondary  
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50 PCB sources, such as release from building materials including caulking and paint (Jamshidi et  
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52 al., 2007; Thomas et al., 2012), as well as from contemporary unintentional sources, most  
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3 notably commercial paint pigments (Hu and Hornbuckle, 2010). There is still considerable risk  
4  
5 of human exposure to PCBs as corroborated by recent reports from the United States of  
6  
7 widespread exposure to PCBs among women of childbearing age (Thompson and Boekelheide,  
8  
9 2013) and levels of PCBs in the indoor air of elementary schools that exceed the EPA's 2009  
10  
11 public health guidelines (Thomas et al., 2012).  
12

13  
14 NDL PCBs with multiple *ortho* chlorine substitutions are particularly stable and predominate  
15  
16 over dioxin-like congeners in environmental samples (Kostyniak et al., 2005; Hwang et al.,  
17  
18 2006; Martinez and Hornbuckle, 2011) and in human tissues (DeCaprio et al., 2005; Schantz et  
19  
20 al., 2010; Marek et al., 2013; Megson et al., 2013). *Ortho*-substituted congeners with the highest  
21  
22 activity towards ryanodine receptors (RyRs) collectively represent 40-50% of total PCBs  
23  
24 currently found in environmental and biotic samples and their net effects are likely to be additive  
25  
26 (Pessah et al., 2006). Consistent with these reports, analyses of PCB levels in human brains  
27  
28 obtained from the general adult population detected predominantly *ortho*-substituted congeners  
29  
30 at concentrations ranging from 0.07 to 12-ng/g wet weight (Dewailly et al., 1999, Covaci et al.,  
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32 2002, Chu et al., 2003).  
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### 38 39 **3. Identification of the Molecular Initiating Event (MIE)**

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41 A molecular initiating event for PCB developmental neurotoxicity is the interaction of NDL  
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43 PCBs with Ryanodine Receptors (RyRs) in neurons of the central nervous system. RyRs are  
44  
45 microsomal  $\text{Ca}^{2+}$  channels that are broadly expressed throughout the mammalian brain and  
46  
47 associate with cytosolic, endoplasmic reticulum (ER)-anchored and ER luminal proteins to form  
48  
49 local  $\text{Ca}^{2+}$  release units (CRUs). These CRUs regulate  $\text{Ca}^{2+}$  release from the ER and modify  
50  
51 gating responses and signal gain of plasma membrane ion channels, notably NMDA receptors  
52  
53 and voltage-gated  $\text{Ca}^{2+}$  channels. Thus, RyRs function to modulate the amplitude and  
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3 spatiotemporal fluctuation of intracellular  $\text{Ca}^{2+}$  during cell activation (Pessah et al., 2010).  
4  
5 Significant to the neurotoxic potential of PCBs, RyR channel activity regulates a variety of  
6  
7 physiological and pathophysiological processes in the developing and mature central nervous  
8  
9 system (Berridge, 2006; Pessah et al., 2010).  
10

11  
12 Nanomolar concentrations of NDL PCB congeners interact with RyRs to dramatically increase  
13  
14 their sensitivity to activation by nanomolar  $\text{Ca}^{2+}$  and to attenuate their sensitivity to inhibitory  
15  
16 feedback by millimolar  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  (Pessah and Wong, 2001), thereby stabilizing the RyR in  
17  
18 its full open conformation (Samso et al., 2009). NDL PCBs potently and selectively sensitize  
19  
20 both RyR1 and RyR2 channel activities, and PCB-triggered  $\text{Ca}^{2+}$  release from ER membrane  
21  
22 vesicles can be selectively blocked by pretreatment with either FK506 or rapamycin without  
23  
24 inhibiting responses to other RyR channel activators such as caffeine (Pessah et al., 2010).  
25  
26 Rapamycin and FK506 interfere with NDL PCB actions in the same concentration range that  
27  
28 promotes the dissociation of the FKBP12/RyR1 complex, suggesting that NDL PCBs interact  
29  
30 with a binding site formed at the FKBP12/RyR1 complex interface to enhance channel open  
31  
32 probability. However, an allosteric mechanism has not been ruled out.  
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39 A stringent structure-activity relationship has been identified for RyR sensitization by NDL  
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41 PCBs with PCB 95 (2,2',3,5'6-pentachlorobiphenyl) being the most potent and efficacious  
42  
43 congener identified to date (Pessah et al., 2010). Non-coplanar PCBs possessing 2-3 chlorine  
44  
45 *ortho* substitutions are the most potent RyR activators (Pessah et al., 2006), which is consistent  
46  
47 with findings from multiple laboratories that non-coplanar, but not coplanar, PCBs increase  
48  
49 intracellular  $\text{Ca}^{2+}$  in neurons [reviewed in (Kodavanti, 2005)]. Two important aspects of the  
50  
51 PCB structure-activity relationship towards RyR1 include: (1) chlorine substitutions at the *ortho*-  
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53 positions which restrict the biphenyl rings to non-coplanarity; and (2) the contribution of *para*-  
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3 substituents which can reduce or eliminate activity (Pessah et al., 2010). The 2,3,6-Cl  
4 configuration on one ring with *ortho*- and *meta*-chloro substitutions on the other is optimal for  
5 recognizing a binding site within the RyR1 complex and for sensitizing channel activation. In  
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substituents which can reduce or eliminate activity (Pessah et al., 2010). The 2,3,6-Cl configuration on one ring with *ortho*- and *meta*-chloro substitutions on the other is optimal for recognizing a binding site within the RyR1 complex and for sensitizing channel activation. In general, PCBs lacking at least one *ortho*-substitution are inactive toward RyR1 and RyR2, regardless of the degree of chlorination, whereas *para*-chloro substitution lowers the efficacy towards RyR1 regardless of the presence of one or more *ortho*-substitutions. Since hydroxylated PCBs are appearing in human tissues, there is currently great interest in the biological activity of these metabolites relative to the corresponding parent structures. The 4-OH derivative of PCB 30 (4'-OH-PCB 30) was found to be significantly more active toward RyR1 than the parent PCB 30 (2,4,6-Cl). Thus, a *para*-OH group on the phenyl ring that carries no other deactivating substitution confers potency and efficacy towards activating RyR1.

#### **4. Identification of the response on the cellular/tissue level that may be an adverse outcome or linked to the final adverse outcome**

Critical determinants of neuronal connectivity include neuronal apoptosis (Barone et al., 2000; Sastry and Rao, 2000, Martin, 2001) and dendritic morphogenesis (Kennedy, 2000; Matus, 2000), and both apoptosis and dendritic morphogenesis are regulated by Ca<sup>2+</sup>-dependent signaling [reviewed in (Pessah et al., 2010)]. As determined by Ca<sup>2+</sup> imaging of dissociated cultures of primary rat hippocampal neurons, PCB 95, a potent RyR sensitizer, enhanced synchronized Ca<sup>2+</sup> oscillations in somata and dendrites. This effect was blocked by ryanodine, indicating that PCB 95 increases spontaneous Ca<sup>2+</sup> oscillations in neurons by stabilizing RyR channels in the open configuration (Wayman et al., 2012a). Consistent with this observation, picomolar to nanomolar concentrations of PCB 95 activate two Ca<sup>2+</sup>-dependent signaling pathways in cultured hippocampal neurons: (1) sequential activation of CaMKK, CaMKI $\alpha$ / $\gamma$ , and

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3 MEK/ERK and CREB, which increases transcription of Wnt2 (Wayman et al., 2012a); and (2)  
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5 CREB-mediated miR132 upregulation, which suppresses the translation of p250GAP (Lesiak et  
6  
7 al., 2013). In cultured hippocampal neurons, the former signaling pathway mediates PCB 95-  
8  
9 induced dendritic growth (Wayman et al., 2012a), whereas the latter mediates PCB 95-induced  
10  
11 synaptogenesis, which is evident as increased spine density and increased frequency of miniature  
12  
13 excitatory post-synaptic currents (Lesiak et al., 2013).  
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16  
17 The dendrite- and synapse-promoting activity of PCB 95 was blocked by pharmacologic  
18  
19 antagonism of RyRs by FLA 365 or by siRNA knockdown of RyR1 or RyR2, and exposure of  
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21 sister cultures to PCB 66 (2,3,4',4-tetrachloro-biphenyl), a congener that has negligible RyR  
22  
23 activity, did not alter dendritic arborization relative to vehicle controls (Wayman et al., 2012b,  
24  
25 Lesiak et al., 2013). Dendritic growth in hippocampal slice cultures was similarly enhanced by  
26  
27 PCB 95 and inhibited by pharmacologic blockade or siRNA knockdown of RyRs (Wayman et  
28  
29 al., 2012b). PCB 95, but not PCB 66, also promotes dendritic growth in primary cultures of  
30  
31 cortical neurons via RyR-dependent mechanisms (Yang et al., 2009).  
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35  
36 Aroclor 1254 (10  $\mu$ M), a commercial PCB mixture comprised predominantly of NDL PCBs, as  
37  
38 well as the NDL congener PCB 47 (1  $\mu$ M) significantly increase caspase-dependent apoptosis in  
39  
40 primary cultures of hippocampal but not cortical neurons (Howard et al., 2003). In contrast,  
41  
42 PCB 77, a congener with little to no RyR activity, has no effect on apoptosis in either neuronal  
43  
44 cell type. The pro-apoptotic activity of PCB 47 was inhibited by the RyR antagonist FLA 365  
45  
46 and by the antioxidant  $\alpha$ -tocopherol but not by antagonists of the IP3 receptor (xestospongin C),  
47  
48 L-type calcium channel (verapamil), or NMDA receptor (APV) (Howard et al., 2003).  
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52 There is also functional evidence that NDL PCBs interfere with neuronal connectivity at the  
53  
54 cellular/tissue level. First, PCB 95 but not PCB 66 altered excitability in hippocampal slice  
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3 cultures (Wong et al., 1997). Second, two different NDL congeners, PCB 95 and PCB 170  
4 altered the ratio of excitatory to inhibitory neurotransmission in hippocampal slice cultures, and  
5  
6 this effect was blocked by the RyR antagonist dantrolene (Kim et al., 2009). Third, the NDL  
7  
8 congener PCB 136, which enantiospecifically sensitizes RyRs (Pessah et al., 2009), exhibits the  
9  
10 same enantiomeric specificity on dendritic arborization and synchronized  $Ca^{2+}$  oscillations in  
11  
12 hippocampal neurons cultured on microelectrode arrays (Yang et al., 2013).  
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17 **5. Identification of the responses on the organ level that may be an adverse outcome or**  
18  
19 **linked to the final adverse outcome**  
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22 Gestational and lactational exposure to Aroclor 1254 (A1254) in the maternal diet significantly  
23  
24 increased dendritic arborization of pyramidal neurons in the CA1 region of the hippocampus of  
25  
26 weanling rats (Wayman et al., 2012b). In a separate study of experience-dependent dendritic  
27  
28 growth, gestational and lactational exposure to Aroclor 1254 promoted dendritic growth in  
29  
30 cerebellar Purkinje cells and neocortical pyramidal neurons among untrained animals but  
31  
32 attenuated or reversed experience-dependent dendritic growth among Morris water maze-trained  
33  
34 littermates (Yang et al., 2009). A1254 is comprised predominantly of NDL PCB congeners  
35  
36 (Kostyniak et al. 2005), and consistent with the hypothesis that these congeners are primarily  
37  
38 responsible for the effects of A1254 on neuronal connectivity, developmental exposure to PCB  
39  
40 95 in the maternal diet significantly increased dendritic growth of CA1 pyramidal neurons in the  
41  
42 hippocampus of untrained weanling rats at the lower doses tested (0.1 – 1.0 mg/kg/d in the  
43  
44 hippocampus of untrained weanling rats at the lower doses tested (0.1 – 1.0 mg/kg/d in the  
45  
46 maternal diet) but not at the highest dose tested (6.0 mg/kg/d in the maternal diet) (Wayman et  
47  
48 al., 2012b).  
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51  
52 PCBs have also been shown to increase apoptosis in the developing brain. Caspase-3 activity  
53  
54 was significantly increased in the cortex, hippocampus and cerebellum of newborn but not  
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3 weanling rats exposed to Aroclor 1254 at 1.0 mg/kg/d in the maternal diet (Yang and Lein,  
4  
5 2010). The most prominent effect was observed in the cerebellum, and PCB-induced apoptosis  
6  
7 in this brain region was confirmed by TUNEL. Further evidence that PCBs modulate  
8  
9 development of neuronal networks at the organ level is the demonstration that developmental  
10  
11 exposure to PCB 95 significantly enhanced the ratio of excitatory to inhibitory currents within  
12  
13 the primary auditory cortex (A1) of weanling rats (Kenet et al., 2007). This effect was  
14  
15 associated with irregularly shaped topographic organization of A1 and disruption of the critical  
16  
17 period plasticity that underlies normal postnatal auditory system development.  
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22 **6. Identification of the response on the organism level that may be the final adverse**  
23  
24 **outcome or linked to the final adverse outcome**  
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27 Structural plasticity of dendrites is considered the cellular substrate of learning and memory  
28  
29 (Leuner and Shors, 2004), and developmental A1254 exposure causes subtle but statistically  
30  
31 significant delays in learning and memory that exhibit an inverted dose-related response similar  
32  
33 to that observed for experience-dependent dendritic plasticity in A1254-treated animals (Yang et  
34  
35 al., 2009). Similarly, perinatal exposure to PCB 95 has been shown to persistently alter activity  
36  
37 levels and behavior in the radial arm maze in adult rats (Schantz et al., 1997). Perinatal exposure  
38  
39 to a mixture of the NDL PCB 47 and the dioxin-like PCB 77 has recently been reported to alter  
40  
41 social behaviors in rats (Jolous-Jamshidi et al., 2010). Schematic representation of MIE, cellular  
42  
43 key events and organ/organism effects is described in Fig. 10.  
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48 **7. Identification of the overall effects on the population**  
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51 Population-based studies have consistently demonstrated that PCBs negatively impact  
52  
53 neuropsychological function in exposed children (Schantz et al., 2003; Carpenter, 2006; Korrnick  
54  
55 and Sagiv, 2008). Similar to the behavioral deficits observed in experimental models of PCB  
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3 developmental neurotoxicity, the subtle effects of PCBs on cognitive function in children may be  
4  
5 overcome by training or increasing age (Schantz et al., 2003; Carpenter, 2006; Korrick and  
6  
7 Sagiv, 2008). Such subtle effects may have significant biological and social costs when  
8  
9 considered at the population level (Weiss, 2000; Grandjean et al., 2007). More recently, NDL  
10  
11 PCBs have been implicated as environmental risk factors for complex neurodevelopmental  
12  
13 disorders such as autism and ADHD (Grandjean and Landrigan, 2006; Eubig et al., 2010;  
14  
15 Landrigan, 2010, Landrigan et al., 2012; Stamou et al., 2013). Abnormalities in dendritic shape  
16  
17 and impaired experience-dependent dendritic plasticity are the most consistent pathologic  
18  
19 correlate of behavioral deficits in heritable and environmentally triggered neurodevelopmental  
20  
21 disorders (Fukuda et al., 2005; Bourgeron, 2009; Garey, 2010; Svitkina et al., 2010; Penzes et  
22  
23 al., 2011). PCB 95 effects on dendritic arborization were blocked by pharmacological  
24  
25 antagonism or siRNA knockdown of the  $\text{Ca}^{2+}$ /calmodulin kinase-I (CaMKI)-CREB-Wnt  
26  
27 signaling pathway (Wayman et al., 2012a). Genes encoding these same  $\text{Ca}^{2+}$ -dependent signaling  
28  
29 molecules are implicated as ASD susceptibility genes (Krey and Dolmetsch, 2007; Pessah and  
30  
31 Lein, 2008), and the proteins encoded by these genes are upregulated in iPSC-derived neurons  
32  
33 from Timothy syndrome patients (Pasca et al., 2011).  $\text{Ca}^{2+}$  imaging studies of cultured rat  
34  
35 hippocampal neurons (Wayman et al., 2012a) revealed that acute exposure to the NDL congener  
36  
37 PCB 95 promotes the same bursting type of  $\text{Ca}^{2+}$  activity as was reported in iPSC-derived  
38  
39 neurons expressing gene mutations that confer ASD susceptibility, specifically the gain-of-  
40  
41 function missense mutation in the L-type  $\text{Ca}^{2+}$  channel  $\text{CaV1.2}$  that causes Timothy syndrome  
42  
43 (Pasca et al., 2011) and the *FMR1* premutation (Cao et al., 2012, Liu et al., 2012) (see section 2).  
44  
45 Collectively, these observations suggest that this  $\text{Ca}^{2+}$ -dependent signaling pathway represents a  
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3 potential convergent molecular target for both NDL PCBs and ASD risk genes that interfere with  
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5 a final common path of activity-dependent dendritic arborization and plasticity.  
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### 8. Is the AOP specific to certain life stages (DNT or aging)?

9  
10 The answer to this question is not known. Certainly there are age-related differences in  
11  
12 vulnerability to the neurotoxicity of NDL PCBs (Winneke, 2011) but it is not clear if this is  
13  
14 predominantly a reflection of age-related differences in toxicokinetic or toxicodynamic factors.  
15  
16 RyRs are expressed in the mammalian brain throughout life, but their spatiotemporal expression  
17  
18 patterns change as a function of developmental stage [reviewed in (Pessah et al., 2010)].  
19  
20 Functionally, RyRs not only regulate neurodevelopment and physiological processes in the  
21  
22 central nervous system, but they are also implicated in Ca<sup>2+</sup> dysregulation associated with aging  
23  
24 and neurodegeneration [reviewed in (Thibault et al., 2007)]. With respect to the key cellular  
25  
26 events downstream of RyR sensitization, altered patterns of neuronal apoptosis not only impact  
27  
28 neuronal connectivity in the developing brain, but also influence the susceptibility of the adult  
29  
30 brain to subsequent environmental insults or aging (Langston et al., 1999; Barlow et al. 2007),  
31  
32 and altered patterns of dendritic growth and plasticity are thought to contribute to  
33  
34 neurodegenerative diseases (de Ruiter and Uylings, 1987; Jagadha and Becker, 1989; Flood and  
35  
36 Coleman, 1990). Thus, it is plausible that this AOP may be relevant to not only PCB toxicity in  
37  
38 the developing brain, but also PCB toxicity in the adult and aging brain; however, additional  
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40 work is needed in this area to determine whether this is the case.  
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### 9. How much are initiating and key events conserved across species?

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49 Ryanodine receptors were first identified because of the pronounced actions of the plant alkaloid  
50  
51 ryanodine on insects and vertebrate striated muscles; but subsequently they have been detected in  
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53 a wide range of species including nematodes, mollusks, arthropods, fish, reptiles, amphibians  
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3 and birds (Rossi and Sorrentino, 2004). However, RyR expression profiles vary across species:  
4  
5 in vertebrate, three isoforms of RyRs have been identified (RyR1, RyR2 and RyR3); whereas in  
6  
7 in vertebrates, only one RyR isoform has been cloned. By contrast, in most avian, amphibian and  
8  
9 fish skeletal muscles, two isoforms of RyRs, named  $\alpha$  and  $\beta$ , that correspond to mammalian  
10  
11 RyR1 and RyR3 are expressed. A third isoform, which is better recognized by antibodies against  
12  
13 the mammalian RyR2 than against avian  $\alpha$  and  $\beta$  isoforms and is likely to represent the  
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15 homologous of mammalian RyR2, has been detected in chicken heart. Whether the action of  
16  
17 NDL PCBs on RyR activity is conserved across species remains to be determined. However, it  
18  
19 is known that the key events of  $\text{Ca}^{2+}$ -dependent dendritic arborization, synapse formation and  
20  
21 neuronal apoptosis are conserved across species (Lein et al., 2005).  
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## 26 27 **10. Challenges for further AOP development: strength, data gaps and uncertainties to be** 28 29 **considered** 30

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32 In vitro studies establish a causal link between the MIE, RyR sensitization, and PCB effects on  
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34 dendritic arborization and synaptogenesis. Dendritic growth in primary dissociated cortical or  
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36 hippocampal cells or in hippocampal slice cultures is promoted by PCB 95, a congener with  
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38 potent RyR activity, but not by PCB 66, a congener with negligible RyR activity (Yang et al.,  
39  
40 2009; Wayman et al., 2012b). Blocking RyR activity using either pharmacological approaches  
41  
42 or siRNA knockdown of RyR prevented PCB 95 enhancement of both synchronized  $\text{Ca}^{2+}$   
43  
44 oscillations (Wayman et al., 2012a) and dendritic growth (Yang et al., 2009; Wayman et al.,  
45  
46 2012b), and blocked activation of the CaMK-CREB-Wnt2 signaling pathway (Wayman et al.,  
47  
48 2012a). Similarly, PCB 95-induced synapse formation, as defined by increased spine density  
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50 and increased mEPSCs in primary cultures of dissociated hippocampal cells or hippocampal slice  
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52 cultures, was blocked by pharmacologic antagonism or siRNA knockdown of RyR (Lesiak et al.,  
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3 2013). Structure-activity relationship studies similarly support a causal link between RyR  
4 sensitization and PCB-induced neuronal apoptosis: the RyR active congener PCB 47, but not the  
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6 RyR inactive congener PCB 77, induced apoptosis in primary cultures of dissociated  
7  
8 hippocampal cells (Howard et al., 2003). Furthermore, the pro-apoptotic activity of PCB 47 was  
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10 inhibited pharmacologic block of RyR but not by pharmacologic block of IP3 or voltage-gated  
11  
12 Ca<sup>2+</sup> channels (Howard et al., 2003).

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17 In vivo, developmental exposure to A1254 enhanced basal levels of dendritic growth but blocked  
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19 experience-dependent dendritic growth in weanling rats (Lein et al., 2007; Yang et al., 2009;  
20  
21 Wayman et al., 2012b), while developmental exposure to PCB 95 increased basal levels of  
22  
23 dendritic growth in the weanling rat hippocampus (Wayman et al., 2012b) and disrupted the  
24  
25 balance of neuronal inhibition to excitation in the developing rat auditory cortex (Kenet et al.,  
26  
27 2007). Several lines of evidence suggest that RyR sensitization contributed to these PCB effects.  
28  
29 First, these changes in neuronal connectivity were associated with exposure to NDL PCBs with  
30  
31 high affinity for the RyR. Second, PCB-induced changes in dendritic growth and plasticity were  
32  
33 coincident with increased [3H]ryanodine binding sites and RyR expression in the brains of  
34  
35 untrained animals and inhibition of training-induced RyR upregulation (Yang et al., 2009).  
36  
37 Increased RyR expression in brain tissues has also been associated with PCB-induced changes in  
38  
39 gene expression (Royland and Kodavanti, 2008; Royland et al., 2008) and locomotor activity  
40  
41 (Roegge et al., 2006). Third, the dose relationship for PCB effects on dendritic growth and  
42  
43 plasticity was similar to that of PCB effects on RyR expression but not to that of PCB effects on  
44  
45 thyroid hormone levels or sex-steroid-dependent developmental endpoints (Yang et al., 2009).  
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47 Fourth, the dose relationship for PCB effects on dendritic growth and plasticity was also similar  
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49 to that of PCB effects on spatial learning and memory in the Morris water maze (Yang et al.,  
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3 2009). This is consistent with the extensive literature documenting the robust effect of activity  
4 or experience on the development and refinement of synaptic connections, which not only  
5 patterns neural circuitry during development but also underlies associative learning (Pittenger  
6 and Kandel, 2003). Furthermore, altered patterns of dendritic growth and plasticity are  
7 associated with impaired neuropsychological function in experimental models (Berger-Sweeney  
8 and Hohmann, 1997), and abnormalities in dendritic shape and experience-dependent plasticity  
9 are the most consistent pathologic correlate of behavioral deficits in heritable and  
10 environmentally triggered neurodevelopmental disorders (Fukuda et al., 2005; Bourgeron, 2009;  
11 Garey, 2010, Svitkina et al., 2010; Penzes et al., 2011).

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25 While a causal link between the MIE and key events at the cellular/tissue level is well-  
26 established in vitro, and key events identified in vitro are recapitulated in vivo, there remain  
27 uncertainties as to whether the MIE is causally linked to altered in vivo neuronal connectivity  
28 and behavioral deficits. This uncertainty arises in part because other biological activities have  
29 been ascribed to non-coplanar PCBs, including increased intracellular levels of ROS (Fonnum et  
30 al., 2006; Mariussen and Fonnum, 2006), disruption of thyroid hormone signaling (Zoeller,  
31 2007; Crofton, 2008) and decreased levels of dopamine (Mariussen and Fonnum, 2006). RyR  
32 activity has been implicated in these other biological activities [reviewed in (Pessah et al.,  
33 2010)], which raises the interesting question of whether these other biological activities are  
34 causally related to PCB developmental neurotoxicity, and if so, do they represent divergent or  
35 convergent mechanisms of PCB developmental neurotoxicity? The observation that RyRs play  
36 critical roles in diverse tissue types and in numerous cellular processes (Berridge, 2006) raises  
37 another interesting challenge associated with the identification of RyR sensitization as a MIE in  
38 PCB developmental neurotoxicity. What factor(s) determine the specificity of PCB toxicity?  
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3 Why do PCBs seem to preferentially target the developing nervous system? Certainly the timing  
4 of exposure will influence the biological outcomes of PCB exposures, as will pharmacokinetic  
5 parameters such as dosage, the metabolites produced, and distribution of PCBs within the body.  
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10 But other factors that could be equally important include expression patterns of RyRs and the  
11 complement of accessory proteins that comprise the calcium release unit as well as the  
12 antioxidant capacity of the cell. An important data gap in this context is whether NDL PCBs  
13 exert comparable effects on all 3 RyR isoforms that are expressed in the brain. Another  
14 interesting data gap that emerges from consideration of the structure-activity relationship of PCB  
15 interactions with the RyR is whether the RyR functions as a target for other toxicants that  
16 possess non-coplanar structures. Obvious candidates include the polybrominated diphenyl ethers  
17 (Kim et al., 2011) and triclosan (Cherednichenko et al., 2012).  
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31 **X. Adverse Outcome Pathway on: *The interaction of redox cycling chemicals with NADH***  
32 ***cytochrome b5 reductase and NADH-quinone oxidoreductase results in NAD<sup>+</sup> formation***  
33 ***causing reduced adult neurogenesis.***  
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38 *Ellen Fritsche*  
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## 42 **1. Introduction**

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44 Adult neurogenesis of hippocampal neural progenitor cells (NPCs) takes place in human brains  
45 up to old age (Eriksson et al., 1998) and contributes to brain function in the adult mammal  
46 (Dupret et al., 2008). During the physiological process of aging, a decline in hippocampal NPC  
47 function is observed (Altman and Das, 1965; Kempermann et al., 1998; Kuhn et al., 1996; Seki  
48 and Arai, 1995; van Praag et al., 2005) correlating with a decline in learning and memory tasks  
49 (Aizawa et al., 2011; Chang et al., 2008; Drapeau et al., 2003; Driscoll et al., 2006; Kempermann  
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3 et al., 1998; Kempermann et al., 2002; Klempin et al., 2013; Kronenberg et al., 2006; Montaron  
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5 et al., 2006; Shors et al., 2001; van Praag et al., 2005). To study adult neurogenesis the rodent  
6  
7 has proven to be a valuable *in vivo* model. However, with regard to the mechanisms responsible  
8  
9 for decreasing neurogenesis with age, it has to be kept in mind that there seem to be species  
10  
11 differences between primates (possibly humans) (Aizawa et al., 2011) and rodents (Eisch and  
12  
13 Petrik, 2012) regarding the extent of adult neurogenesis (Eisch and Petrik, 2012) and processes  
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15 contributing to an aging hippocampus.  
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20 Oxidative stress contributes to loss-of-function during NPC aging (reviewed in van Wijngaarden  
21  
22 and Franklin, 2013). Thereby, the intracellular redox state seems to govern cell fate during  
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24 differentiation as an oxidized intracellular environment favours glial over neuronal  
25  
26 differentiation of SGZ neural stem cells (Prozorovski et al., 2008). Hence, increased ROS  
27  
28 generation might influence NPC proliferation, differentiation and fate determination (van  
29  
30 Wijngaarden and Franklin, 2013).  
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## 34 **2. Characterization of the exposure to the chemicals relevant to the selected AOP**

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36 ROS-producing chemicals are able to enter adult brains and cause oxidative stress. Routes of  
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38 exposure can be via skin, lung or the gastro-intestinal tract. After entering the circulation, the  
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40 compound has to pass the BBB either by passive diffusion or active uptake. ROS-producing  
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42 compounds include paraoxon (Jafari et al., 2012b), diazinon (Jafari et al., 2012a), ammonium  
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44 acetate (Satpute et al., 2012), acrylamide (Lakshmi et al., 2012), adriamycin (doxorubicin) (Joshi  
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46 et al., 2005) and paraquat (Chen et al., 2010). In addition to chemical compounds, gamma-  
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48 irradiation also produces oxidative stress in brain tissue (Riley, 1994; Zhao and Robbins, 2009).  
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### 3. Identification of the Molecular Initiating Event (MIE)

The MIE prompting this AOP is the interaction of redox cycling chemicals (R) with enzymes (NADH-cytochrome *b*<sub>5</sub> reductase, NADH-quinone oxidoreductase) catalysing electron transport from NADH to the chemical, thereby forming a radical R<sup>•</sup> and the NADH oxidation product NAD<sup>+</sup> (Dinis-Oliveira et al., 2006; Shimada et al., 1998). These processes happen at the cytosolic side of the outer mitochondrial membrane and thus do not directly involve or alter the mitochondrial electron transport chain (Shimada et al., 1998). The radical R<sup>•</sup> causes non-enzymatic generation of the reactive oxygen species (ROS)  $\cdot\text{O}_2^-$ . Enzymatic detoxification of  $\cdot\text{O}_2^-$  involves superoxide dismutase and glutathione peroxidase, thereby producing the oxidized form of glutathione (GSH), GSSG (Andreyev et al., 2005; Zhu and Shan, 2009). Glutathione reductase then reconstitutes GSSG to reduced glutathione by forming NADP<sup>+</sup> (reviewed in Belenky et al., 2007; Bolton et al., 2000). The mitochondrial enzyme nicotinamide nucleotide transhydrogenase (NNT) is able to reconstitute NADP<sup>+</sup> to NADPH by generating NAD<sup>+</sup> (Olgun, 2009). Thus, NAD<sup>+</sup> is formed by two different mechanisms: as the MIE by direct chemical interference with electron-donating enzymes and secondly by the first series of key events, i.e. ROS formation, detoxification and reconstitution of GSH. One example of such a redox-cycling chemical is paraquat (PQ). PQ directly interacts with NADH-cytochrome *b*<sub>5</sub> reductase and NADH-quinone oxidoreductase, thereby accepting one electron from NADH reducing PQ<sup>2+</sup> to PQ<sup>•+</sup> and forming NAD<sup>+</sup> (Belenky et al., 2007; Dinis-Oliveira et al., 2006; Shimada et al., 1998). ROS effects on cells, tissues and organs are multifarious and depend on the amount of intracellular redox active species. While a certain low level of ROS produced by mitochondrial respiration is nowadays considered 'physiological signalling molecules', large excess of ROS causes macromolecular and thus cellular damage with deathly consequences. ROS concentrations triggering this AOP

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3 are meant to be below causing cell death, but function as signalling molecules by shifting the  
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5 cellular redox state towards the oxidative side. Such ROS production occurs during aging in  
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7 humans and experimental animals (reviewed in van Wijngaarden and Franklin, 2013) as well as  
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9 in response to toxicant exposure (Cheng et al., 2009; Merzoug et al., 2011; Milatovic et al., 2009;  
10  
11 Ojha et al., 2013) or radiation damage (Robbins and Zhao, 2004; Zhao et al., 2007). Thus, aging  
12  
13 poses a higher susceptibility to adverse effects of exogenous ROS due to raised basal  
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15 intracellular levels.  
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#### 20 **4. Identification of the responses on the cellular/tissue level that may be an adverse** 21 **outcome or linked to the final adverse outcome** 22 23

24 NAD<sup>+</sup> is a necessary co-factor and activator for the histone deacetylase (HDAC) Sirt-1 (Araki et  
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26 al., 2004; Lin et al., 2004), the mammalian homologue of the yeast, drosophila and C. elegans  
27  
28 Sir2 protein (Imai et al., 2000). Sirt-1 lacks a DNA binding domain and has to be recruited to  
29  
30 target promoters by sequence-specific transcription factors to induce chromatin remodelling and  
31  
32 regulate gene expression (Rosenberg and Parkhurst, 2002). One of the transcription factors  
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34 associating with Sirt-1 is the transcriptional co-repressor Hes1 (Takata and Ishikawa, 2003),  
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36 which is expressed in neural stem/progenitor cells (NS/PCs) and prevents premature neuronal  
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38 differentiation into DCX (double cortin)<sup>+</sup> cells by repressing activation of the pro-neuronal basic  
39  
40 helix-loop-helix (bHLH) transcription factor Mash1 (Ishibashi et al., 1995); reviewed by (Libert  
41  
42 et al., 2008). Redox state contributes to NS/PC proliferation and neuronal differentiation:  
43  
44 oxidation-mediated *Mash-1* repression in NS/PCs is blocked in the absence of Sirt-1 or Hes-1 *in*  
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46 *vitro* (Prozorovski et al., 2008). It is highly likely that NAD<sup>+</sup> is the signalling molecule linking  
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48 redox state and Sirt-1-dependent repression of neuronal differentiation.  
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3 **5. Identification of the responses on the organ level that may be an adverse outcome or**  
4 **linked to the final adverse outcome**  
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8 *In vivo*, Sirt-1 regulates the neurogenic potential of NS/PCs in the early postnatal as well as in  
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10 the adult subventricular zone (SVZ) and adult hippocampus (Prozorovski et al., 2008; Saharan et  
11  
12 al., 2013). In adult mouse SVZ and hippocampi, Sirt-1 knock down results in a significant  
13  
14 increase in neuronal production, whereas Sirt-1 overexpression or activation by resveratrol, a  
15  
16 potent chemical Sirt-1 activator, prevent adult neural precursors from differentiating into neurons  
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18 (Saharan et al., 2013). A pro-oxidative state by systemic glutathione depletion decreases NS/PC  
19  
20 proliferation and generation of young neurons and increases the number of GFAP<sup>+</sup> cells at the  
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22 same time in the SVZ in a Sirt-1 dependent manner, demonstrating contribution of Sirt-1 to  
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24 neural fate decision in the oxidative milieu (Prozorovski et al., 2008); reviewed by (Libert et al.,  
25  
26 2008). One repression target of Sirt-1 is the pro-neural gene Mash-1 (Ishibashi et al., 1995) and  
27  
28 Mash-1 regulates neural versus glial fate of embryonic and adult NS/PC (Nieto et al., 2001;  
29  
30 Parras et al., 2004). During the aging process, oxidative damage occurs in the hippocampus of  
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32 mice (Nicolle et al., 2001). Feeding middle aged mice with already impaired NS/PC proliferation  
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34 and generation of DCX<sup>+</sup> young neurons with the NO-donor flurbiprofen, which amongst others  
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36 has antioxidative properties, restores these NS/PC functions almost to levels of young controls  
37  
38 (L'Episcopo et al., 2013) suggesting causal involvement of oxidative stress in age-related decline  
39  
40 of NS/PC functions. These are related to altered wnt-signaling (L'Episcopo et al., 2013) and a  
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42 crosstalk between Sirt-1- and wnt-signaling was reported in a different context earlier (Holloway  
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44 et al., 2010). Whether this aging-related, oxidation state-dependent, wnt-mediated decline in  
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46 NS/PC functions is determined by NAD<sup>+</sup>-dependent Sirt-1 activation needs further clarification.  
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## **6. Identification of the responses on the organism level that may be the final adverse outcome or linked to the final adverse outcome**

Decreased adult neurogenesis by waning NS/PC proliferation and/or generation of new, young neurons result in the adverse outcome of impaired cognitive function that mirrors changes characteristic of aging (Aizawa et al., 2011; Drapeau et al., 2003; Driscoll et al., 2006; Kempermann et al., 1998; Montaron et al., 2006; Shors et al., 2001). Two notions support this statement: (i) Interventions that enhance or reduce hippocampal neurogenesis cause improved (Chang et al., 2008; Kempermann et al., 1998; Kempermann et al., 2002; Klempin et al., 2013; Kronenberg et al., 2006; Montaron et al., 2006; van Praag et al., 2005) or impaired, aging-characteristic (Montaron et al., 2006; Shors et al., 2001) cognitive functions, respectively. (ii) Creating an inducible transgenic strategy allowing specific ablation of adult-born hippocampal neurons in mice causes impairment of spatial relational memory in these animals (Dupret et al., 2008).

Spatial learning and memory impairment in Sirt-1 knockout mice imply that Sirt-1 is involved in hippocampal-dependent cognitive functions (Michan et al., 2010). Feeding aged mice an antioxidative saffron diet that improves learning and memory measured by a passive avoidance task supports the notion that oxidative stress indeed triggers the impaired adverse outcome (Papandreou et al., 2011). Schematic representation of MIE, cellular key events and organ/organism effects is described in Fig. 11.

## **7. Identification of the overall effects on the population**

In humans, cognitive deficits are attributed to reduced adult neurogenesis (reviewed in Knoth et al., 2010; van Wijngaarden and Franklin, 2013). By current knowledge, such stem cell exhaustion belongs to the nine identified hallmarks of aging (reviewed in Lopez-Otin et al.,

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3 2013). Thereby, oxidative stress is one of the main contributors to NS/PC aging (reviewed in  
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5 (van Wijngaarden and Franklin, 2013). Such age-related decline in NS/PC function can be  
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7 accelerated by exogenous noxae, which increase the oxidative burden in the brain. In the human  
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9 population, chemotherapy or gamma-irradiation for cancer treatment cause cognitive impairment  
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11 probably due to compromised NC/PC function (reviewed in Gibson and Monje, 2012; Greene-  
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13 Schloesser et al., 2012; Monje and Dietrich, 2012). One discussed mechanism of chemotherapy-  
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15 or gamma-irradiation-induced cognitive changes is the generation of ROS (Ahles and Saykin,  
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17 2007; Greene-Schloesser et al., 2012). Such accelerated cognitive decline poses a large financial  
18  
19 and social burden on society. Whether environmentally relevant compounds trigger the same  
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21 AOP in humans is so far not known. However, accumulating data from animals and humans  
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23 suggest that antioxidative strategies have the ability to improve cognition in the elderly  
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25 (reviewed in Joseph et al., 2009).  
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### 31 **8. Is the AOP specific to certain life stages (DNT or aging)?**

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33 The current body of literature supports the hypothesis that age-associated increases in the  
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35 generation of ROS might influence CNS stem cell proliferation, differentiation and fate  
36  
37 determination (reviewed in Takata and Ishikawa, 2003). While mitochondrial dysfunction  
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39 increases as a function of age raising endogenous ROS formation (reviewed in Bishop et al.,  
40  
41 2010), antioxidative defence capacities decrease during the aging process (Li et al., 2012;  
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43 Saharan et al., 2013). Therefore, susceptibility to disturbance of the delicate cellular redox  
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45 balance is likely to increase with age. Moreover, BBB permeation changes within the function of  
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47 age, adding to the higher susceptibility for this AOP during aging (Simpson et al., 2010).  
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## 9. How much are initiating and key events conserved across species?

The general concept of adult neurogenesis is conserved between species. However, there are some uncertainties and some already known species differences. (I) MIE: Direct and indirect mechanisms contribute to the generation of NAD<sup>+</sup>. It is not known if these are quantitatively conserved across species. (II) Cellular ROS detoxifying capacities determine NAD(P)<sup>+</sup>/NAD(P)H ratios. Whether detoxification capacities of adult NS/PCs are comparable across species has never been studied. (III) Concerning gamma-irradiation of brains, rats tolerate a 5-times higher dose than humans (Monje and Palmer, 2003). The underlying molecular rationale is not known. (IV) In rodents, SVZ and hippocampal neurogenesis contribute to cognition. In humans, current evidence suggests that SVZ is negligible beyond late infancy (Sanai et al., 2011; Wang et al., 2011c). (V) Whether human Sirt-1 also regulates the human Mash-1 homologue Hash-1 and thus regulates adult NS/PC fate decision is not known. However, this pathway seems to be homologous in human embryonic stem cells (Zhang et al., 2011). (VI) Aging studies in non-human primates suggest that signalling pathways and NS/PC phenotypes during the physiological brain aging process might be different between primates (maybe including humans) and rodents (Aizawa et al., 2011).

## 10. Challenges for further AOP development: strength, data gaps and uncertainties to be considered

Causalities between the MIE, KEs 1-5 and the AO are very stringent. However, data linking the MIE directly and causally with the AO are missing. Moreover, human data on chemical exposure and affected hippocampal function neurogenesis is not available.

Although causality within this AOP is quite stringent, it has not yet been determined if the above-mentioned indeed increase NAD<sup>+</sup> in regenerative zones of the brain and cause decreased

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3 adult neurogenesis by this mechanism chemicals (Chen et al., 2010; Jafari et al., 2012a,b; Joshi  
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5 et al., 2005; Lakshmi et al., 2012; Satpute et al., 2012). Moreover, the proposed higher  
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7 susceptibility towards ROS-induced impairment in NS/PC function has not specifically been  
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9 addressed experimentally.  
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Review Only

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## Legends of figures

**Fig. 1.** Relationships between the uncertainty in AOP models, the data and resources needed to develop the models and the regulatory domains of applicability.

**Fig. 2.** Schematic representation of the putative AOP on *Binding of antagonist to an NMDAR during synaptogenesis contributes to impairment of learning and memory abilities*. The binding of antagonist to NMDAR in hippocampus during synaptogenesis leads to inhibition of the receptor activity and to the delay in the ontogeny of the NR2A subunit, contributing to decreased  $\text{Ca}^{2+}$  influx into neurons and decreased glutamate release, causing a concomitant reduction of the BDNF release. The resulting cellular key events eventually lead to impaired capacity for processes underlying learning and memory. Brain-derived neurotrophic factor (BDNF); N-methyl-D-aspartate receptor (NMDAR); long-term potentiation (LTP); long-term depression (LTD).

**Fig. 3A.** Cascade of events taking place after NMDAR over activation leading to neuronal cell death. Over activation of the ligand-gated calcium channel NMDAR leads to various stress responses like endoplasmic reticulum (ER) stress, which is also termed as “unfolded protein” response and is related to protein aggregation ( $\beta$ -amyloid,  $\alpha$ -synuclein in Alzheimer and Parkinson disease), mitochondrial apoptosis, oxidative stress, with subsequent activation of poly-ADP-ribose polymerase-1 (PARP-1), resulting in rapid energy depletion of apoptotic neurons. Calcium signaling via NMDAR and calcium-dependent kinases is in a normal “healthy” setting absolutely key to memory and cognition. NMDARs are allosteric proteins tightly regulated by activity and numerous endogenous factors maintaining cytoskeletal and nuclear integrity of the neuron and its dendritic contacts. One key aspect is maintenance of energy homeostasis which

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3 explains the crucial role of mitochondria and NADH in neurodegenerative processes. PARG is  
4 the antagonistic enzyme to PARP-1 degrading poly-ADP-ribosylated modifications.  
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8 **Fig. 3B.** Schematic representation of the putative AOP on *Binding of agonist to NMDA receptor*  
9 *causes excitotoxicity that mediates neuronal cell death, contributing to reduction (or loss) of*  
10 *cognitive, sensory and motor function.* The binding of exogenous agonist to NMDAR causes  
11 over activation of the receptor and collapse of calcium homeostasis, a key regulator of synaptic  
12 plasticity underlying cognition. Intracellular calcium overload mediates chemical-induced  
13 neurodegeneration induced by key cellular events such as mitochondrial dysfunction, oxidative  
14 and ER stress. Depending on the degree, time of exposure and structure of the brain where  
15 neurodegeneration takes place, it can result in different adverse outcomes such as reduction (or  
16 loss) of cognitive, sensory or motor function. N-methyl-D-aspartate receptor (NMDAR);  
17 Adenosine triphosphate (ATP); endoplasmic reticulum (ER); reactive oxygen species (ROS).  
18 (\*see AOP on *Binding of inhibitors to the mitochondrial respiration chain complex I, II, III or IV*  
19 *or interaction of uncouplers with oxidative phosphorylation decreases or blocks ATP production*  
20 *resulting in neurodegeneration*).  
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38 **Fig. 4A.** Schematic representation of the putative AOP on *Binding of antagonist to GABA<sub>A</sub>*  
39 *receptor results in hyperexcitability and convulsions.* By blocking the activity of the GABA<sub>A</sub>R,  
40 the inhibitory GABAergic neurotransmission is reduced, due to the reduced Cl<sup>-</sup> influx, resulting  
41 in increased excitatory activity in neuronal network. A severe deregulation of the balance  
42 between excitation and inhibition beyond physiological levels is involved in several pathologies  
43 of the central nervous system such as seizures and/or /convulsions (Adverse Outcome).  $\gamma$ -  
44 aminobutyric acid A receptor (GABA<sub>A</sub>R).  
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3 **Fig. 4B.** Correlation between acute toxicity in vivo (LD50) and inhibition of the GABA<sub>A</sub>R  
4 induced by convulsant drugs and pesticides acting at the GABA<sub>A</sub>R. Symbols (correlations):  
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6 squares for the inhibition of GABA-induced Cl<sup>-</sup> flux (R<sup>2</sup> = 0.613; p = 0.004) and triangles for the  
7  
8 inhibition of [<sup>35</sup>S] TBPS binding (r<sup>2</sup> = 0.371; p = 0.02). Values are extracted from references  
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10 cited along the text (Obata et al., 1988; Pomés et al., 1994)  
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13 **Fig. 5.** Schematic representation of the putative AOP on *Binding of pyrethroids to voltage-gated*  
14 *sodium channels induces acute neurotoxicity*. This AOP is initiated by binding of pyrethroids to  
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16 a distinct site on voltage-gated sodium channels (VGSCs). This binding interaction between  
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18 pyrethroids and VGSCs results in alterations in the kinetics of VGSC activation and inactivation  
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20 (opening and closing of the channel). The degree of perturbation of VGSC kinetics is structure-  
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22 dependent, and a continuum of lengths of modification of kinetics have been observed in a  
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24 variety of species, including mammals. At the cellular level, altered VGSC kinetics give rise to  
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26 changes in membrane excitability that also depend on the length of modification, with short-  
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28 lasting modifications resulting in repeated firing of action potentials and long lasting  
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30 modifications resulting in membrane depolarization and ultimately, depolarization-dependent  
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32 block of action potential propagation. In turn, these cellular changes result in changes in the  
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34 activity at the organ level, resulting in changes in activity in different neuronal pathways that  
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36 ultimately lead to the two different Adverse Outcomes that are observed as Type I (T) or II (CS)  
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38 syndromes described in the main text.  
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48 **Fig. 6.** Schematic representation of the putative AOP on *Binding of certain organophosphates to*  
49 *NTE results in delayed neuropathy*. Binding of some specific organophosphates (OPs) to  
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51 neuropathy target esterase (NTE) causes inhibition and aging of the enzyme followed by cellular  
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53 key events such as disruption of Ca<sup>2+</sup> homeostasis, mitochondrial dysfunction with subsequent  
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3 energy depletion and buildup of oxidative stress. One or more of the above cellular effects could  
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6 disrupt the regulation of cell signaling, causing altered phosphorylation of cytoskeletal proteins  
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8 that lead to cytoskeletal dysfunction. Energy depletion due to mitochondrial dysfunction would  
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10 also impact on cytoskeletal motor protein activity and axonal transport with subsequent  
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12 impairment of nerve regeneration or neurite development. Prolongation of these cellular key  
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14 events causes degeneration of axons in the CNS and PNS, leading to peripheral neuropathy  
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16 (adverse outcome).  
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20 **Fig. 7.** Schematic representation of the putative AOP on *Impairment of learning and memory*  
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22 *induced by binding of electrophilic chemicals to SH(thiol)-group of proteins and non-protein*  
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24 *molecules in neuronal and glial cells during development.* By binding of a compound to  
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26 SH(thiol)-groups of proteins and non-protein molecules (Molecular Initiation Event) during brain  
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28 development and the subsequent functional modification of their function leads to several  
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30 distinct cellular key events that depend on the function and location of these proteins in the  
31  
32 specific brain cell type and the brain structure. Mainly proteins and non-protein molecules  
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34 associated with mitochondria, antioxidant defense, and glutamate storage, release and uptake are  
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36 targeted. This binding leads to the depletion of reduced glutathione, increased of extracellular  
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38 Glu levels inducing over activation of NMDAR, possible neuronal/glia dysfunction of  
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40 respiratory chain complexes, triggering oxidative and nitrosative stress causing neuronal cell  
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42 death. The induced neurodegeneration contributes to the decreased neuronal network formation  
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44 and function responsible for the deficits in developmental learning and memory processes (AO).  
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Glutathione (GSH); glutamate (Glu) N-methyl-D-aspartate receptor (NMDAR); neuronal nitric  
oxide synthase (nNOS); nitric oxide (NO); thiol- (SH-). (\*see AOP on *Binding of inhibitors to  
the mitochondrial respiration chain complex I, II, III or IV or interaction of uncouplers with*

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3 *oxidative phosphorylation decreases or blocks ATP production resulting in neurodegeneration ;*  
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6 *\*\*see AOP on Binding of agonist to NMDA receptor causes excitotoxicity that mediates*  
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8 *neuronal cell death, contributing to reduction (or loss) of cognitive, sensory and motor function;*  
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10 *\*\*\*see AOP on Binding of antagonist to an NMDAR during synaptogenesis contributes to*  
11 *impairment of learning and memory abilities).*  
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15 **Fig. 8A.** Schematic representation of the mitochondrial electron transport chain (ETC). Electrons  
16 are transferred from the matrix via NADH (nicotinamide adenine dinucleotide) reduction into the  
17 complex I and via FADH<sub>2</sub> (FAD: flavin adenine dinucleotide) reduction into complex II. The  
18 Coenzyme Q transfers electrons from complex I and II to complex III. Cytochrome C further  
19 transfers these electrons from complex III to the final electron acceptor complex IV, where  
20 oxygen is reduced to water. A proton gradient is generated that is used for production of ATP by  
21 complex V (ATP synthase).  
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31 **Fig. 8B.** Schematic representation of the putative AOP on *Binding of inhibitors to the*  
32 *mitochondrial respiration chain complex I, II, III or IV or interaction of uncouplers with*  
33 *oxidative phosphorylation decreases or blocks ATP production resulting in neurodegeneration.*  
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39 Binding of inhibitors to the ETC complexes or uncoupling of OXPHOS induces inhibition of  
40 mitochondrial respiration leading to reduction of the ATP production. The decreased level of  
41 ATP leads to the cellular effects such as disturbed calcium homeostasis, causing ROS  
42 production, disruption of mitochondrial membrane potential, cytochrome c release resulting in  
43 apoptotic or necrotic cell death. If the significant cell death of dopaminergic neurons takes place  
44 through these mechanisms in substantia nigra, the symptoms of Parkinson's disease might be  
45 triggered.  
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3 Adenosine triphosphate (ATP); cytosolic (c); Calcium ( $\text{Ca}^{2+}$ ); cytochrome C (Cyt C); electron  
4 transport chain, (ETC); mitochondria (mt); oxidative phosphorylation (OXPHOS); reactive  
5 oxygen species (ROS); Substantia Nigra pars compacta (SNpc) \*see AOP on *Binding of agonist*  
6 *to NMDA receptor causes excitotoxicity that mediates neuronal cell death, contributing to*  
7 *reduction (or loss) of cognitive, sensory and motor function; \*\*see AOP on Multiple molecular*  
8 *initiating events trigger neuroinflammation leading to neurodegeneration.*  
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17 **Fig. 9.** Schematic representation of the putative AOP on *Multiple molecular initiating events*  
18 *trigger neuroinflammation leading to neurodegeneration.* Several MIEs can lead to intermediate  
19 cellular key events that cause cell type specific effects, followed by the neuroinflammatory  
20 response. The triggered cellular effects such as mitochondrial dysfunction, oxidative stress,  
21 depletion of reduced GSH and excitotoxicity lead to neuronal damage (axonal demyelination,  
22 synapse impairment and cytoskeleton disruption) and glia activation (microglia and astrocytes)  
23 causing neuroinflammation leading to neurodegeneration. Neurodegeneration mediated through  
24 these pathways are well documented in the brain structures which are linked to PD and AD.  
25 Molecular Initiating Event (MIE); Acetylcholinesterase (AChE); glial fibrillary acidic protein  
26 (GFAP); glutathione (GSH); N-methyl-D-aspartate receptor (NMDAR), Parkinson disease (PD);  
27 Alzheimer disease; (\*see AOP on: *Binding of antagonist to an NMDAR during synaptogenesis*  
28 *contributes to impairment of learning and memory abilities; \*\* AOP on: Impairment of learning*  
29 *and memory induced by binding of electrophilic chemicals to SH(thiol)-group of proteins and*  
30 *non-protein molecules in neuronal and glial cells during development; \*\*\* AOP on: Binding of*  
31 *agonist to NMDA receptor causes excitotoxicity that mediates neuronal cell death, contributing*  
32 *to reduction (or loss) of cognitive, sensory and motor function; \*\*\*\* AOP on: Binding of*  
33 *inhibitors to the mitochondrial respiration chain complex I, II, III or IV or interaction of*  
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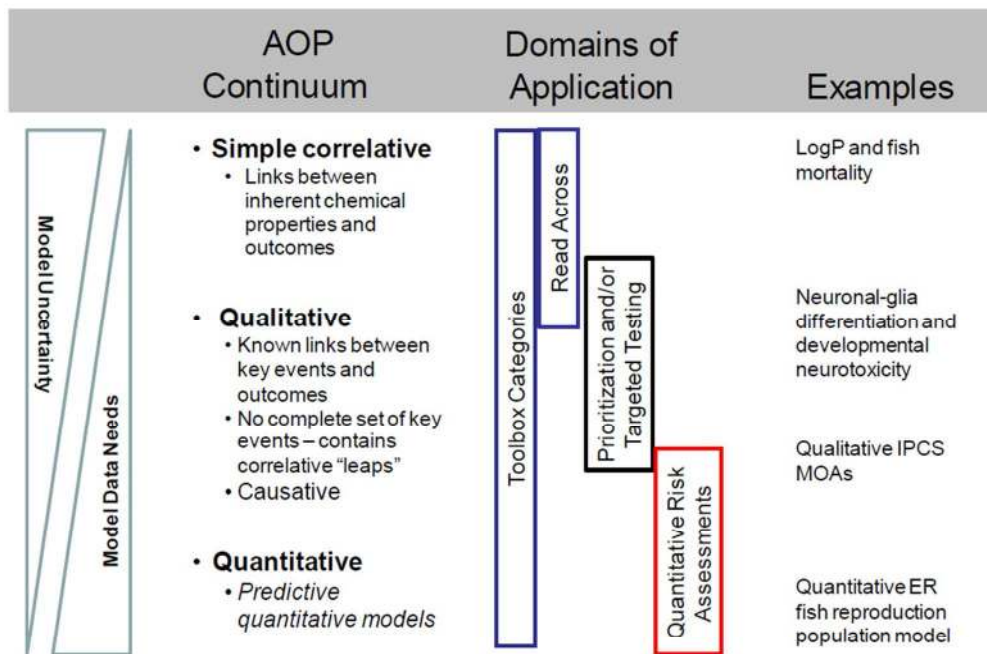


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3 *uncouplers with oxidative phosphorylation decreases or blocks ATP production resulting in*  
4 *neurodegeneration).*  
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8 **Fig. 10:** Schematic representation of the putative AOP on *The interaction of non-dioxin-like*  
9 *PCBs with ryanodine receptors (RyRs) causes their sensitization affecting neuronal connectivity*  
10 *that results in behavioral deficits (developmental neurotoxicity).* NDL PCBs sensitize RyR  
11 activity and alter Ca<sup>2+</sup>-dependent signaling mechanisms that link neuronal activity to dendritic  
12 growth and plasticity and to neuronal apoptosis. These cellular effects alter normal patterns of  
13 neuronal connectivity in the brain and contribute to behavioral and psychomotor deficits  
14 observed at the organismal level (adverse outcome). Ryanodine receptor (RyR).  
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25 **Fig. 11.** Schematic representation of the putative AOP on *The interaction of redox cycling*  
26 *chemicals with NADH cytochrome b5 reductase and NADH-quinone oxidoreductase results in*  
27 *NAD<sup>+</sup> formation causing reduced adult neurogenesis.* The MIE of this AOP is the interaction of  
28 a redox-active chemical with NADH-cytochrome c reductase or NADH-quinone oxidoreductase  
29 on the cytoplasmic site of the outer mitochondrial membrane associated to mitochondrial  
30 complex I (**Fig. 8A**). NAD<sup>+</sup> is formed either directly by this MIE or secondary through  
31 formation of  $\cdot$  O<sub>2</sub><sup>-</sup> by the redox cycler and subsequent oxidation of GSH to GSSG, which is  
32 reconstituted under generation of NADH<sup>+</sup> and finally NAD<sup>+</sup>. NAD<sup>+</sup> is a necessary co-factor  
33 activating the HDAC Sirt-1. By recruiting Hes-1, Sirt-1 represses expression of the pro-neuronal  
34 gene Mash-1 thus shifting neural progenitor cell fate to the glial side. On the organ level, this  
35 causes diminished neuronal regeneration in the hippocampus with decreased cognitive  
36 performance as the AO. Glutathione (GSH); oxidized glutathione (GSSG); histone deacetylase  
37 (HDAC); nicotinamide adenine dinucleotide (NAD<sup>+</sup>); reduced form of nicotinamide adenine  
38 dinucleotide (NADH); neural stem/progenitor cells (NS/PC).  
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JRC/EPA AOP Workgroup Meeting RTP 03/01/12

Fig.1.  
85x62mm (300 x 300 DPI)

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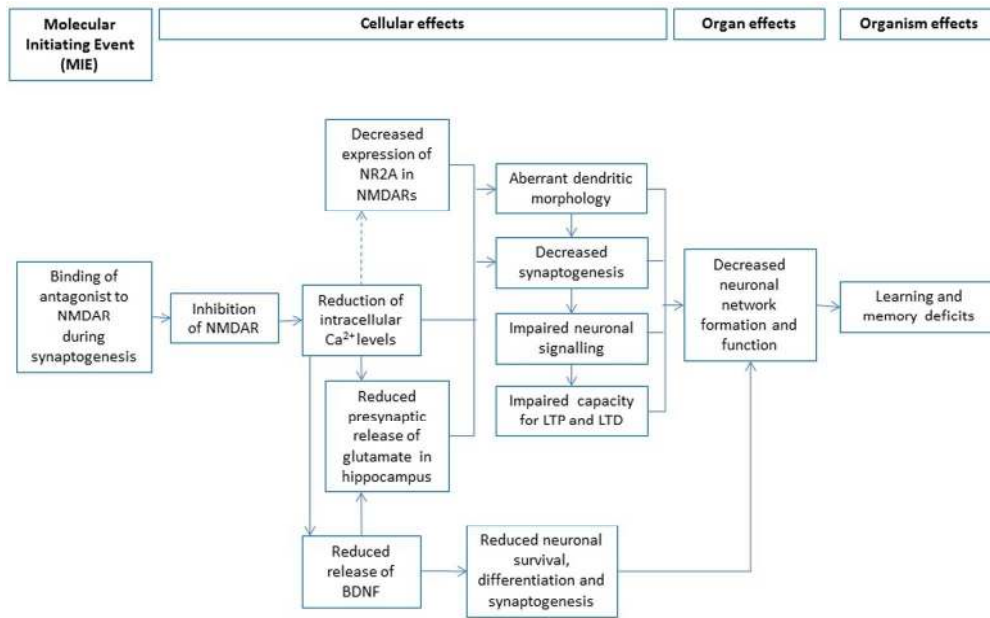


Fig. 2.  
253x159mm (300 x 300 DPI)

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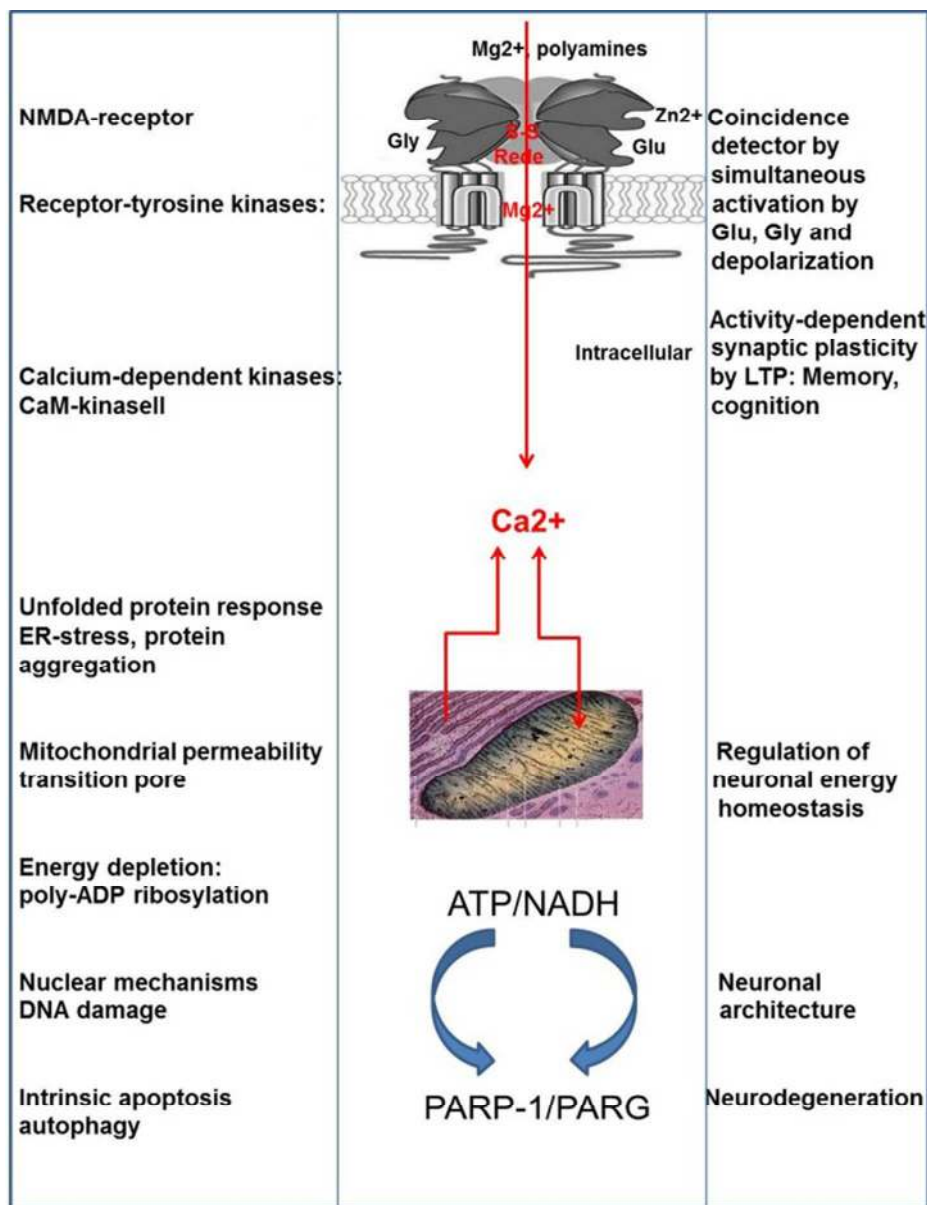


Fig. 3A.  
83x103mm (300 x 300 DPI)

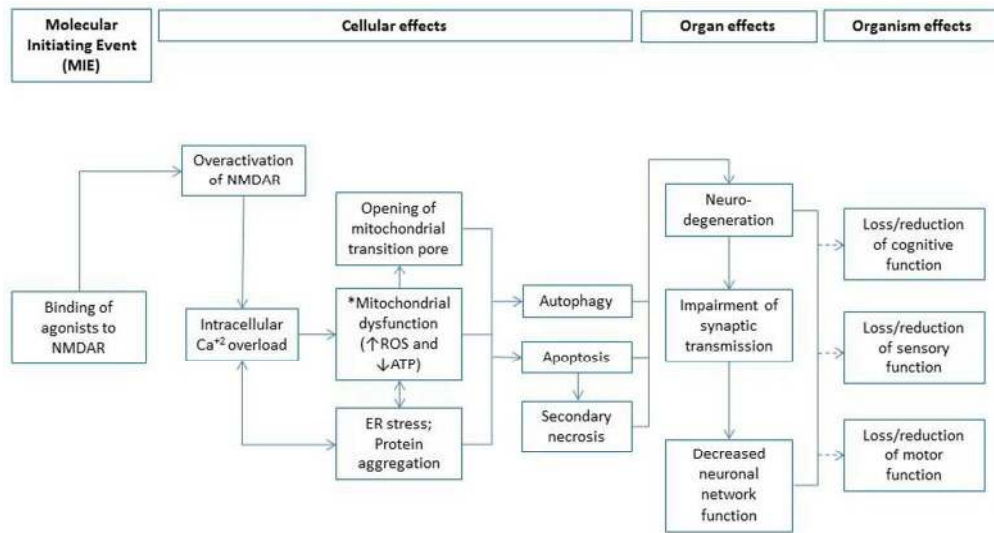


Fig. 3B.  
251x135mm (300 x 300 DPI)

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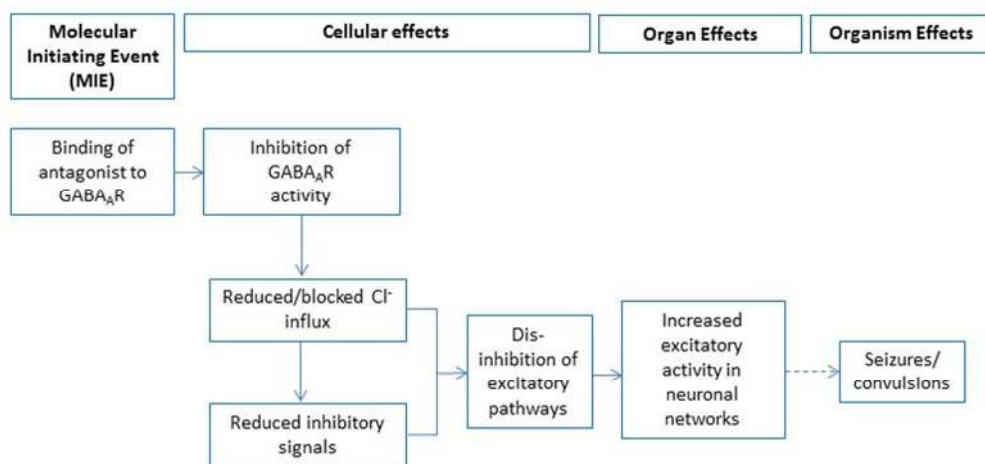


Fig. 4A.  
208x97mm (300 x 300 DPI)

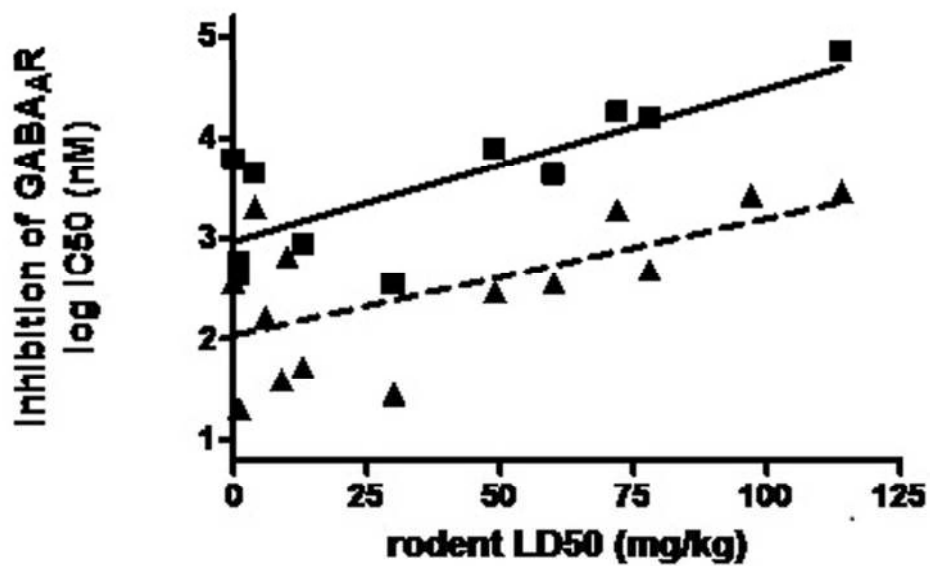


Fig. 4B.  
95x60mm (300 x 300 DPI)



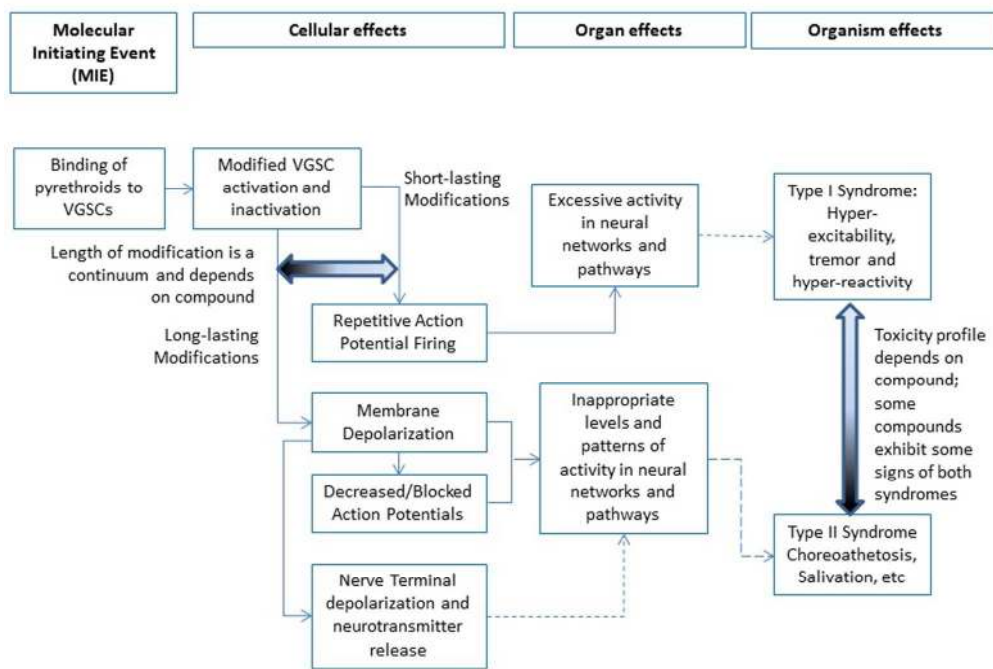


Fig. 5.  
223x151mm (300 x 300 DPI)

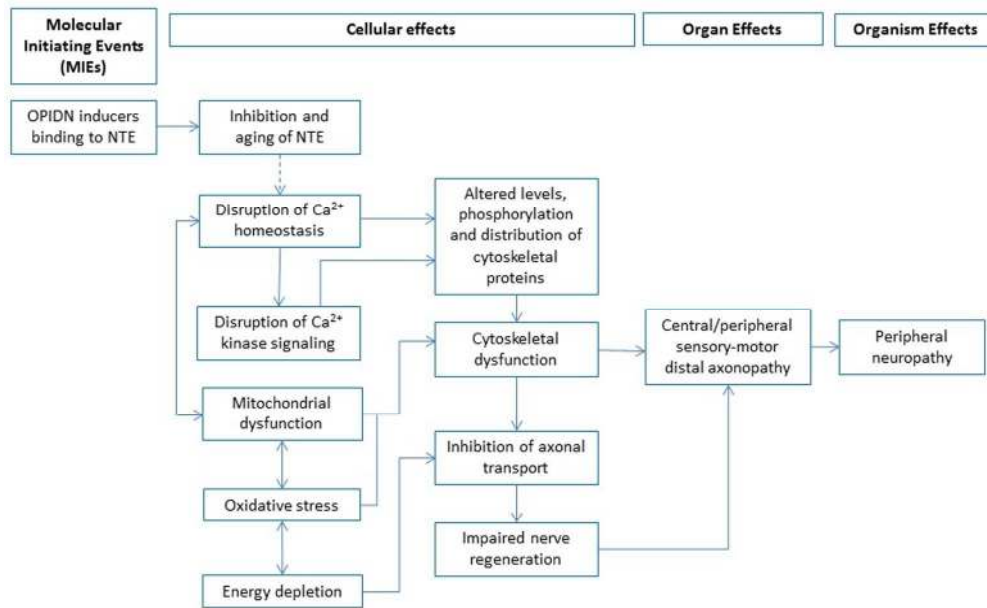


Fig. 6.  
240x149mm (300 x 300 DPI)

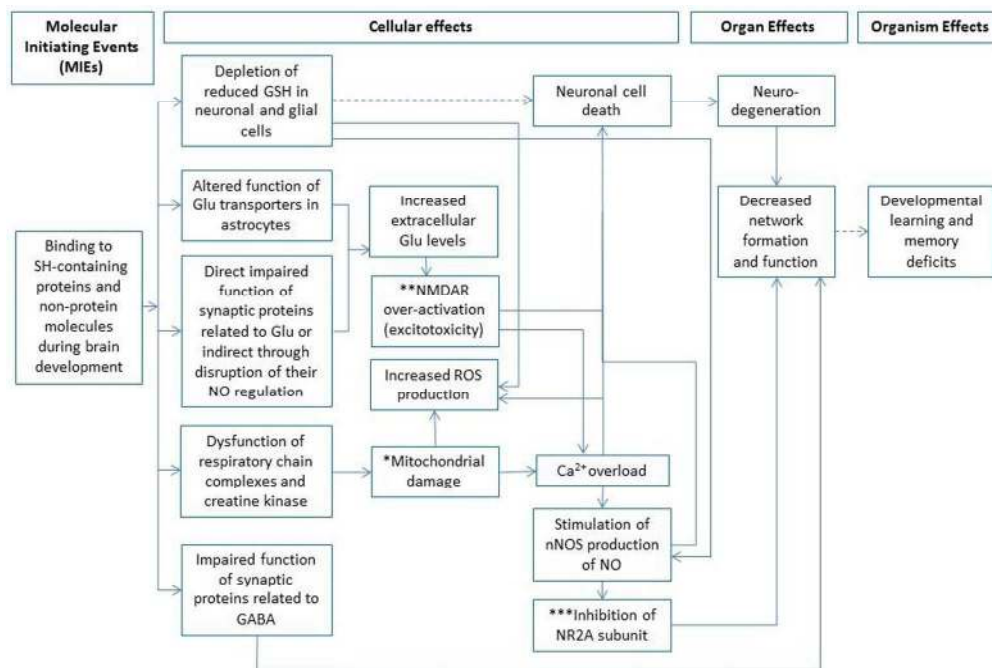


Fig. 7.  
253x170mm (300 x 300 DPI)

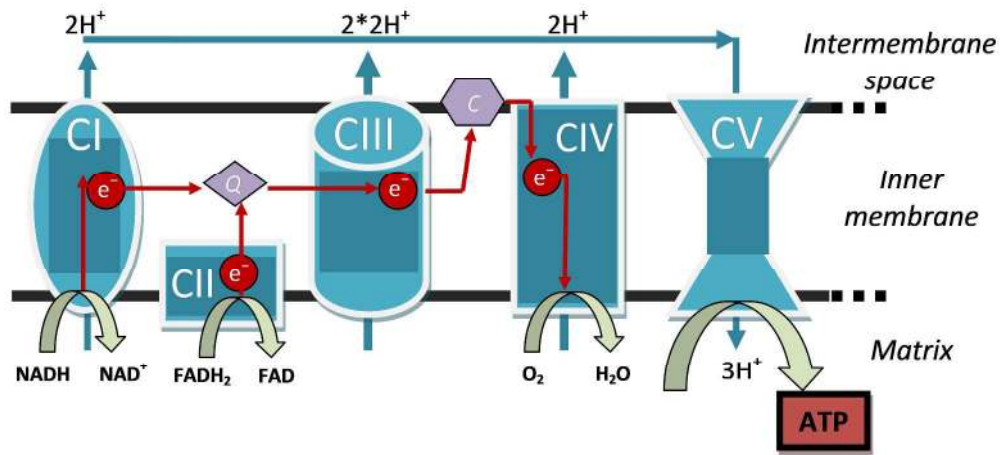


Fig. 8A.  
762x364mm (300 x 300 DPI)

Pre-Review Only

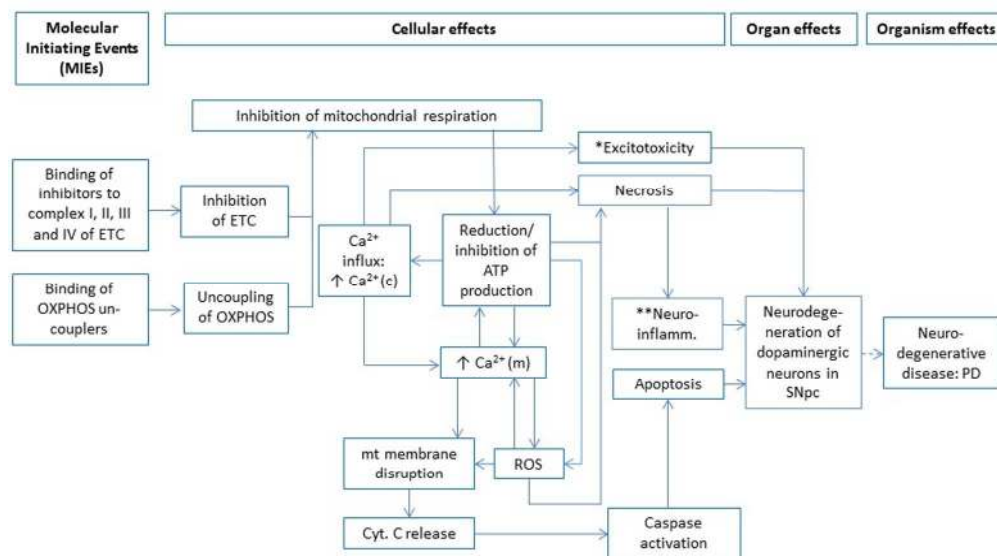


Fig. 8B.  
252x143mm (300 x 300 DPI)

Review Only

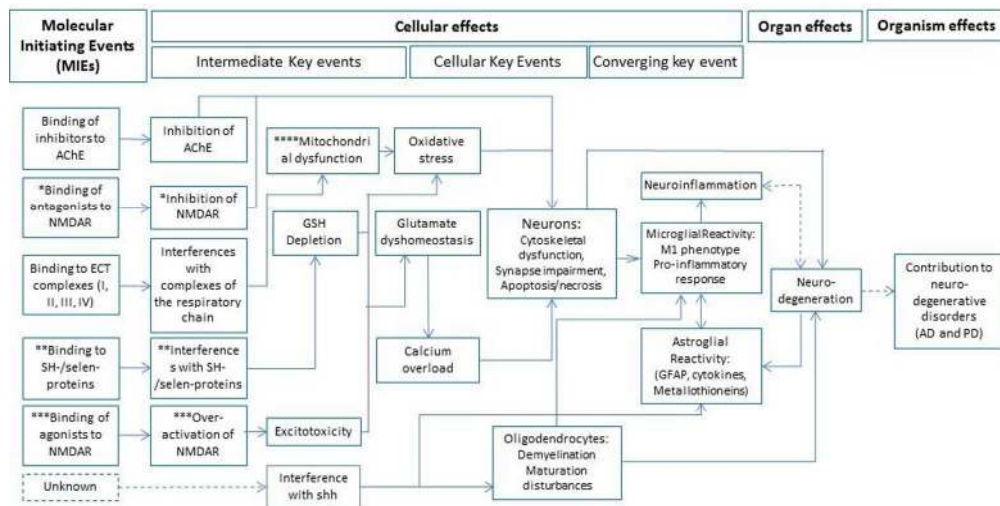


Fig. 9.  
253x129mm (300 x 300 DPI)

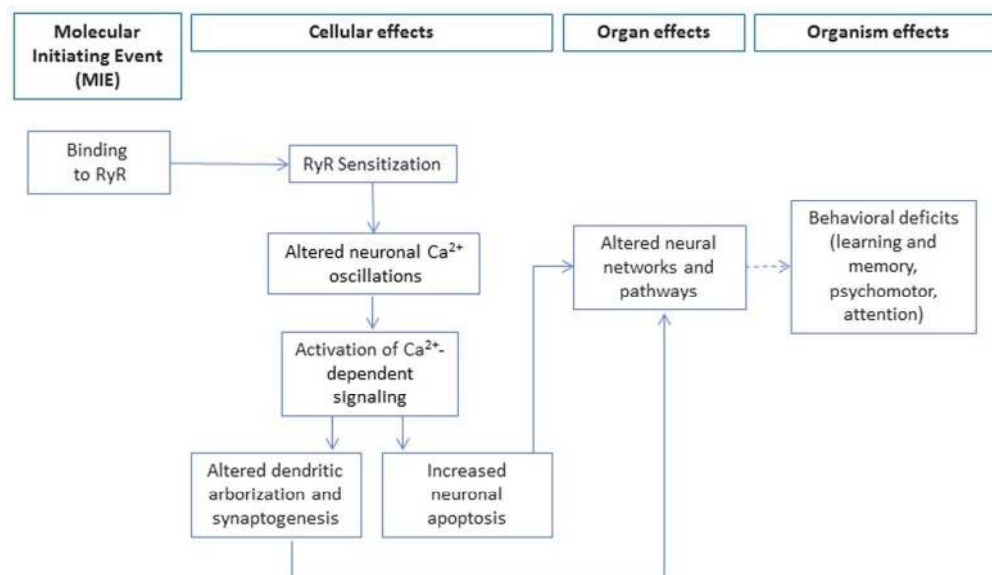


Fig. 10.  
210x122mm (300 x 300 DPI)

Review Only



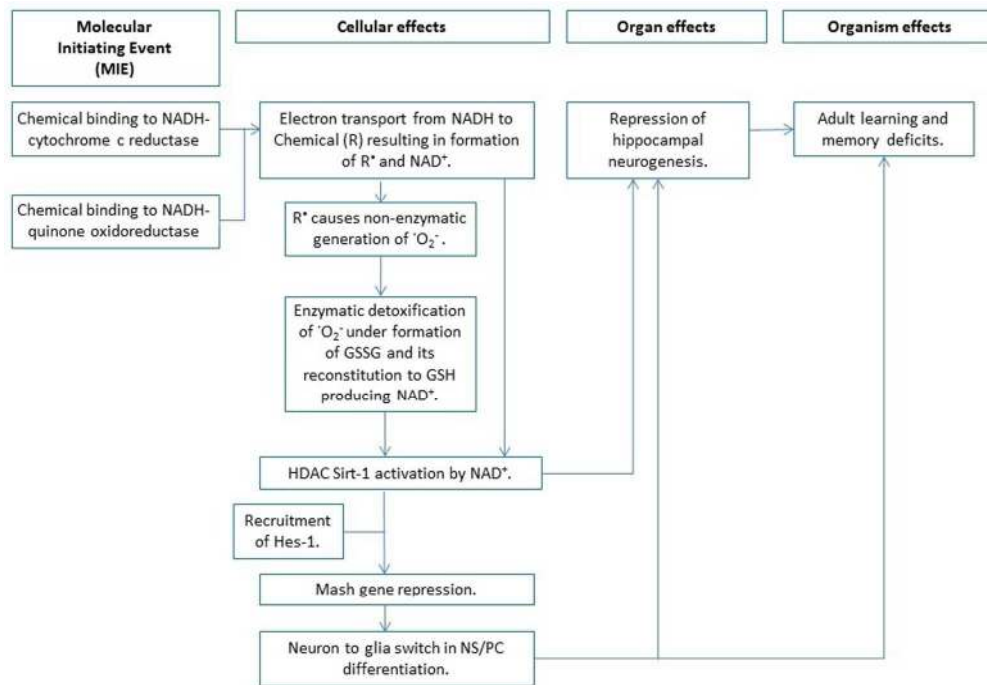


Fig. 11.  
241x167mm (300 x 300 DPI)