

Quantitative evidence for conserved longevity pathways between divergent eukaryotic species

Erica D. Smith^{1,2}, Mitsuhiro Tsuchiya¹, Lindsay A. Fox¹, Nick Dang¹, Di Hu¹, Emily O. Kerr¹,
Elijah D. Johnston¹, Bie N. Tchao¹, Diana N. Pak¹, K. Linnea Welton¹, Daniel E.L. Promislow³,
James H. Thomas⁴, Matt Kaeberlein^{2,5} and Brian K. Kennedy^{1,5}

¹Department of Biochemistry, University of Washington, Seattle, WA 98195 USA, ²Department
of Pathology, University of Washington, Seattle, WA 98195 USA, ³Department of Genetics,
University of Georgia, Athens, GA 30602 USA, ⁴Department of Genome Sciences, University of
Washington, Seattle, WA 98195 USA, ⁵To whom correspondence should be addressed:

Brian K. Kennedy
Department of Biochemistry
University of Washington
Box 357350
Seattle WA 98195
(206)685-0111
bkenn@u.washington.edu

Matt Kaeberlein
Department of Pathology
University of Washington
Box 357470
Seattle WA 98195
(206)543-4849
kaeber@u.washington.edu

Running Title: Conserved modulators of life span in worms and yeast

Keywords: aging, longevity pathways, yeast, worms, TOR, dietary restriction

ABSTRACT

Studies in invertebrate model organisms have been a driving force in aging research, leading to the identification of many genes that influence life span. Few of these genes have been examined in the context of mammalian aging, however, and it remains an open question whether and to what extent the pathways that modulate longevity are conserved across different eukaryotic species. Using a comparative functional genomics approach, we have performed the first quantitative analysis of the degree to which longevity genes are conserved between two highly divergent eukaryotic species, the yeast *Saccharomyces cerevisiae* and the nematode *Caenorhabditis elegans*. Here, we report the replicative life span phenotypes for single-gene deletions of the yeast orthologs of worm aging genes. We find that 15% of these yeast deletions are long-lived. By contrast, only 2.3% of a random set of deletion mutants are long-lived—a statistically significant difference. Among the longevity determining ortholog pairs, we note a substantial enrichment for genes involved in an evolutionarily conserved pathway linking nutrient sensing and protein translation. In addition, we have identified several conserved aging genes that may represent novel longevity pathways. Together, these findings indicate that the genetic component of life span determination is significantly conserved between divergent eukaryotic species, and suggest pathways that are likely to play a similar role in mammalian aging.

[Supplemental research data is included.]

INTRODUCTION

It has been argued that aging is not a genetically programmed process because there is minimal selective advantage associated with long-term post-reproductive survival of the individual (Partridge and Gems 2006). In the natural environment, survival of many organisms is heavily dependent on extrinsic factors such as predation rather than age-associated mortality. Thus, there is little evidence supporting the idea that alleles that influence the rate of individual aging have been selected for that function. Nevertheless, empirical data show that aging is genetically determined. Single gene mutations that significantly decrease mortality have been identified in each of the major model organisms used for gerontological research. Indeed, several hundred genes have been reported that influence longevity in yeast, worms, or flies (Guarente and Kenyon 2000; Kaeberlein 2006; Warner 2003).

Given that aging is not programmed, it can be argued that life span in divergent eukaryotic species, living in different environments, may be influenced by different factors. Therefore, the genetic pathways that modulate aging in one organism may differ substantially from those in another. It has been demonstrated that a handful of orthologous genes (e.g. Sir2-family proteins, insulin/IGF-like receptor proteins, and the target of rapamycin (TOR) kinase family) modulate longevity in different organisms (Kaeberlein 2006). However, no studies have yet quantified the degree to which genetic modifiers of aging are evolutionarily conserved. Using two divergent eukaryotic species, the unicellular budding yeast *Saccharomyces cerevisiae* and the multicellular nematode *Caenorhabditis elegans*, evolutionarily separated by approximately 1.5 billion years (Wang et al. 1999), we have quantitatively addressed this question by asking whether yeast

orthologs of known worm aging genes are more likely to modulate yeast aging than randomly selected yeast genes.

RESULTS

Based on a survey of the aging literature, we have compiled a set of 276 *C. elegans* genes for which increased life span results from decreased gene expression or altered protein function (Table S1) (see SAGE KE database at <http://uwaging.org/genesdb/index.php>) (Boehm and Slack 2005; Curran and Ruvkun 2007; Dillin et al. 2002; Hamilton et al. 2005; Hansen et al. 2005; Hansen et al. 2007; Lee et al. 2003; Pan et al. 2007; Schafer et al. 2006; Ventura et al. 2005; Winkelbauer et al. 2005). Using the basic local alignment search tool for proteins, or BLASTp, we searched the yeast proteome for proteins with high sequence similarity to each of the proteins encoded by the 276 known worm aging genes. A set of putative yeast orthologs was defined based on a stringent reciprocal best hit (RBH) criterion (described in Methods). Due to an ancient duplication event in the yeast genome (Wolfe and Shields 1997), we allowed for 1:2 worm:yeast ortholog sets by including both the best hit and the next best hit, when BLASTing worm against yeast, if the BLASTp scores were within 10% of each other. From this analysis, we identified 103 yeast orthologs that were associated with 78 out of the 276 worm aging genes (Table 1, Table S2). The 103 yeast genes that code for these proteins are defined as the RBH set.

To account for possible redundancy within protein families, a second set of yeast homologs to the worm aging genes was identified based on less stringent homology criteria (described in Methods). This set of related proteins (RPs) is likely to contain nearly all of the true ortholog

pairs, but also includes several proteins related by sequence similarity but with different biological functions. A total of 393 yeast genes (including the 103 RBH genes) are contained in the RP set (Table 1, Table S2).

We determined replicative life span (RLS) for single-gene deletion mutants corresponding to genes contained in the RP set (Table S3). Of the 393 genes in the RP set, 264 are viable as deletion alleles and are present in the yeast ORF deletion collection (Winzeler et al. 1999). Replicative life span was measured for each of these 264 single-gene deletion mutants from the *MAT α* (isogenic to BY4742) deletion collection. Deletion strains that tested as significantly long-lived in the initial analysis were verified by independently measuring the replicative life span of the corresponding deletion mutant from the *MAT α* (isogenic to BY4741) deletion collection. This approach of initial testing in one mating type followed by verification in the opposite mating type is the same method used in the genome-wide screen for aging-related genes in yeast (Kaeberlein et al. 2005).

Based on the previously reported partial genome-wide analysis of replicative life span in yeast (Kaeberlein et al. 2005), we estimate that the random chance of a yeast gene deletion mutant being long-lived is 2.3% (13 long-lived strains from a pool of 564 randomly chosen deletion mutants). To account for the possibility that evolutionarily ancient and conserved genes are more likely to be associated with aging, we determined the frequency of aging-related genes in a subset of conserved genes. Based on RBH criteria (see Methods), we determined that 146 yeast genes in the random pool of 564 have putative worm orthologs. In this subset of conserved

genes, 3.4 % (5/146) of the deletion mutants were long-lived. Thus, the expected frequency of aging-related genes in a random set of yeast genes with worm orthologs is 3.4%.

Of the 76 genes examined in the RBH set, 11 deletion mutants were found to be significantly long-lived (Table 2, Figure 1, Fig. S1). After controlling for the potential effects of evolutionary conservation, we see a four-fold enrichment for aging-related genes in yeast genes orthologous to worm genes associated with aging (G test: $\chi_1^2 = 8.57$, $P = 0.0034$). The 76 yeast genes in the RBH set correspond to 56 worm genes. Twenty of the 56 worm aging genes had two yeast orthologs. By chance alone, these 20 genes are approximately twice as likely to have at least one yeast ortholog associated with aging. Even after controlling for the potential confounding effects of duplicate orthologs (see Methods), we still find that our set of yeast orthologs was enriched for genes associated with aging (G test: $\chi_1^2 = 5.37$, $P = 0.021$). Therefore, we propose that yeast orthologs of worm aging genes are more likely regulate yeast aging than yeast orthologs of randomly chosen worm genes, and we infer from these findings that the pathways modulating longevity are partially conserved between these two divergent species.

Analysis of the single gene deletion mutants corresponding to the larger set of yeast RP's was carried out as described above for the RBH set. Of the 264 deletion strains examined, 25 were found to have a significant increase in replicative life span (9.5%), a 3.7-fold enrichment over the random set (G-test: $\chi_1^2 = 12.03$, $P = 0.00052$) (Table 2, Fig. S1). The reduced frequency of long-lived strains among the RP set relative to the RBH set likely reflects the increased number of yeast genes that are not true orthologs of worm aging genes.

In this study, we have identified 25 genes that modulate yeast replicative life span, of which 22 have not been previously reported (Table 2, Figure 1, Fig. S1). Using the Princeton Protein Orthology Database (P-POD, <http://ortholog.princeton.edu/findorthofamily.html>), we were able to identify clear human orthologs for at least 15 of the 25 worm-yeast ortholog pairs, including all genes in the RBH set (Table 2) (Heinicke et al. 2007). The novel genes in the RBH set found to influence yeast aging include a transcription elongation factor (*SPT4*), an ATP-dependent metalloprotease (*AFG3*), an inositol polyphosphate 5-phosphatase (*INP53*), two ribosomal proteins (*RPL9A* and *RPL19A*), and three translation initiation factors (*TIF4631*, *TIF1*, and *TIF2*) (Figure 1). Strikingly, 7 of the 11 genes in the RBH set with a conserved role in longevity are associated with protein translation.

To determine whether the ortholog pairs identified as conserved aging genes in this study are overrepresented in specific functional classes, we performed gene ontology analysis on the worm aging genes, their yeast orthologs in the RBH set, and the 11 conserved aging genes within the RBH set (see Methods, Table S4). Focusing on the cellular component category, we noted that ribosome-associated genes are significantly enriched in each case, with 6/11 of the conserved aging genes in the RBH set annotated with this gene ontology term (Figure 2A). In the biological process category, these ribosome-associated genes are annotated as being involved in translation initiation, ribosome biogenesis, and protein biosynthesis or translation (Table S4). Although mitochondrial-associated genes are highly enriched in the set of worm aging genes and the corresponding yeast orthologs, the conserved aging genes in the RBH set are not statistically enriched for mitochondrial-associated genes (Figure 2A).

Genetic and functional evidence suggests that decreased TOR or S6K activity is one mechanism by which dietary restriction extends life span (Kennedy et al. 2007). There are many downstream targets of TOR signaling, including translation, autophagy, and the stress response (Wullschleger et al. 2006). In this study, we have identified translation initiation factors as highly conserved longevity modifiers, consistent with the hypothesis that these factors modulate longevity in a linear pathway with TOR and DR (Figure 2B). Because our analysis was limited to loss-of-function mutations, however, it is unlikely that we would identify genes involved in autophagy or the stress response using this approach. Thus, we do not exclude the possibility that TOR signaling modulates longevity in divergent eukaryotes through regulation of both translation and other downstream targets.

DISCUSSION

Invertebrate models have been a driving force in aging research, but the degree to which the genetic modifiers of longevity are conserved between different organisms has remained largely unexplored. We have compared two divergent eukaryotes (worms and yeast) and found that the ortholog of a gene with a known longevity role in worms is statistically more likely to play a similar role in yeast. Worms and yeast are separated evolutionarily by a greater distance (~1.5 billion years) than worms and mammals (~1 billion years) (Wang et al. 1999); therefore, quantitative evidence for conservation of longevity pathways between these species would suggest similar overlap between invertebrates and mammals. Indeed, we find that at least 15 of the aging-related ortholog pairs between yeast and worms have clear human orthologs. We believe that many of these genes, and the pathways within which they function, are likely to

modulate aging in mammals, and that mammalian orthologs of these genes pairs are reasonable candidates as potential therapeutic targets for age-associated diseases.

For several reasons, we believe that the observed overlap between worm and yeast aging genes is likely to be an underestimate of the true conservation among longevity genes. First, our rigorous criteria for verification of long-lived yeast deletion mutants (see Methods), and known issues of aneuploidy in the yeast ORF deletion collection, both create a low false positive rate, but may fail to identify some deletion mutants with increased replicative life span. As evidence for this idea, we note several cases (15 in the RBH set) in which the deletion mutant is significantly long-lived in *MAT α* but not *MAT α* (Table S3). While it is possible that these deletions regulate aging in a mating type-specific manner, more likely scenarios are that (1) the deletion would become significant in both mating types with analysis of a greater number of mother cells or (2) the deletion strain was incorrectly annotated or has acquired a suppressor mutation in one mating type or the other. None of these deletions were included in the reported set of long-lived yeast strains. Second, there is an unknown false positive rate among genes reported to be long-lived in *C. elegans*. Third, the longevity modulating role of nearly half of the worm aging genes in our set of 276 was defined in large-scale RNAi screens, and the efficiency of RNAi-mediated knock-down is known to vary. Similarly, many long-lived *C. elegans* mutants correspond to partial loss of function alleles, rather than null alleles. In contrast, all of the yeast data from this study were derived from single-gene deletion mutants that (with the notable exception of the ribosomal protein pairs) result in complete loss of protein activity. Finally, other methods of ortholog identification, such as the use of protein-protein interaction networks, may increase the number of ortholog pairs that have a conserved role in life span determination. For example, previous

studies have argued that *SCH9* is the functional ortholog of *akt-1* (Longo and Finch 2003), and more recently, *SCH9* was shown to have ribosomal S6 kinase activity (Urban et al. 2007).

Because of our stringent reciprocal best hit requirement, however, *SCH9* falls into the RP group in this study, although functional evidence indicates it is orthologous to a known *C. elegans* aging gene.

If aging is not the result of a genetic program, why should orthologs of the genes modulating aging in one organism regulate aging in another? Through careful examination of the pathways that have a conserved role in modulating longevity, we propose a possible explanation. Nutrient-responsive signaling pathways and translational control have been closely linked to life span extension by dietary restriction in a number of studies (Kaeberlein 2007). There is likely to be selective pressure on eukaryotic organisms to assess food availability and alter behavior accordingly in order to increase fitness. Life span modulation by dietary restriction, and genetic mimetics thereof, may be under selective pressure, indirectly, by virtue of their close apposition to organismal responses to food availability. Other pathways modulating aging may not be so tightly partnered to phenotypes under high selection, and therefore may be more variable across eukaryotic species. If true, these findings may help resolve the apparent paradox inherent in the argument that aging is non-programmed but genetically conserved.

METHODS

Ortholog identification

For the purposes of this study, we have defined a worm aging gene as any worm gene whose decreased function has been reported to increase life span (Table S1). Computational algorithms

can identify putative ortholog pairs between yeast and worms; however, this determination is complicated by the fact that there are no definitive parameters for determining true orthology. Here, we used a custom program to analyze BLASTp matches between WS156 (longest splice form for each gene) and SGD1 protein sequences downloaded from Wormbase (www.wormbase.org) and Saccharomyces Genome Database (SGD, <http://www.yeastgenome.org>), respectively. All-by-all BLASTp searches of worm to yeast and yeast to worm were done using the -m 8 output option without filtering and with an E-value cutoff of 0.01. BLASTp hits with an E-value greater than 0.001 or a BLAST bit score below 40 were excluded from analysis in this study.

In order to identify yeast orthologs of the 276 reported worm aging genes, we have adopted a multiple stringency level strategy. The reciprocal best hit (RBH) group contains protein pairs that meet a reciprocal best BLASTp match requirement with at least 20% sequence identity and 20% amino acid alignment. Reciprocal best hits were identified using blast bit scores as the measure of similarity. To allow for 1:2 orthologs, we have included in this group any yeast proteins with a BLASTp score within 10% of the best hit. The RBH group represents putative ortholog pairs and many, if not all, of these ortholog pairs are true orthologs. However, it is likely that other true ortholog pairs are not represented here due to the stringent reciprocal best hit requirement. The ‘related proteins’ (RP) group contains the top six yeast homologs to each worm aging gene, with at least 20% sequence identity and 10% amino acid alignment. This group allows us to address the possibility of redundancy of function within protein families in the context of aging. It is important to note that the RBH group is completely overlapping (included) in the ‘related proteins’ group. Approximately 1/3 of the orthologs and homologs in

the ‘related proteins’ group cannot be tested in the replicative life span assay because these genes are essential or the deletion strains are very sick (and thus not represented in the *MAT α* haploid deletion collection) (Table S2). Ortholog identification is complete for currently known worm aging genes; however, we will continue to periodically refine our putative ortholog sets as new worm aging genes are reported and as gene predictions are updated.

Yeast replicative life span analysis

Single-gene deletion mutants corresponding to all non-essential genes were analyzed for replicative aging. The life span of a minimum of 20 cells from each strain in both sets was determined in the *MAT α* haploid collection. Sign Wilcoxon Rank Sum tests were performed to determine whether the strain exhibited an enhanced life span relative to experiment-matched control cells (BY4742) (Table S3). One of three decisions was made at this point. If the deletion strain was statistically long-lived ($P < 0.05$), 20 cells from the haploid *MAT α* collection were examined to confirm that long life span was linked to the deletion in question. While the ORF deletion set has been immensely valuable to the yeast research community, there are examples of aneuploidy in a small fraction of deletions and growth suppressors have arisen in certain cases. In deletion strains where data from the 20 cells analyzed was ambiguous, we chose to examine the life span of more cells. In general, less than 100 cells were examined in total for these *MAT α* strains, although in a few cases more cells were analyzed. If the life span of one of these deletions was long-lived after collection of more data, verification was performed in the *MAT α* set. Finally, in cases where *MAT α* deletion strains were unlikely to be long-lived, based on analysis of 20 cells, no further analysis was performed.

Statistical analysis of worm aging genes with duplicate yeast orthologs

Our statistical analysis of conserved aging genes included worm genes that had two yeast orthologs. If the probability that a randomly chosen yeast gene is associated with aging is given by ϕ , then worm genes with two yeast orthologs are approximately twice as likely to have one or more yeast ortholog associated with aging ($1-(1-\phi)^2 \approx 2\phi$, assuming ϕ is small). Thus, we may overestimate the degree to which having a worm ortholog associated with aging increases the likelihood that a yeast gene is associated with aging. To control for our allowance of 1:2 orthologs, we performed a more conservative analysis, calculating the probability that deletion of one of two putative yeast orthologs (corresponding to a single worm aging gene, and chosen at random) would result in a long-lived phenotype. Of the 56 worm aging genes in the RBH group, 36 have a single yeast ortholog (3 of which are aging-related) and 20 worm genes have duplicate yeast orthologs (7 of which have at least one yeast ortholog that is aging related). For one worm aging gene (F57B9.6), both yeast orthologs are associated with aging in yeast, so the probability that a randomly chosen ortholog is associated with aging is 1. For the other six worm genes that have duplicate orthologs (one of which is aging related), the probability is 1/2. So, based on the life span data collected, we would expect 7 of the 56 worm aging genes to have a yeast ortholog that is associated with aging. Even taking duplicate orthologs into account, we find that yeast orthologs of worm aging genes are significantly enriched for genes that impact longevity (Fisher's G test, $G=5.37$, $p=0.021$).

Gene ontology (GO) analysis

All GO terms associated with each of the 276 worm aging genes were downloaded from WormMart (www.wormbase.org) and analyzed by a custom program similar to SGD GO Term

Finder, <http://db.yeastgenome.org/cgi-bin/GO/goTermFinder.pl>). GO terms that are statistically enriched (using a cutoff of $P < 0.05$ after Bonferroni correction) relative to the GO term frequency in entire worm genome. Individual statistical tests were computed using Fisher's exact test, which is based on the hypergeometric distribution as described at SGD (<http://www.yeastgenome.org/help/goTermFinder.html>).

GO term analysis of yeast genes was done using SGD GO Term Finder, including all ORFs in the background set and the default settings for annotation source, annotation type, and evidence codes. The P-value cut-off used in this analysis was $P < 0.05$ (after Bonferroni correction).

ACKNOWLEDGMENTS

This work was funded by an award from the Ellison Medical Foundation to B.K.K. and M.K. and by National Institutes of Health Grant R01 AG024287 to B.K.K. E.D.S. was supported by NIH training grant P30 AG013280. This work was performed while M.K. was an AFAR research grant recipient. D.E.L.P. is funded by an award from the Ellison Medical Foundation.

FIGURE LEGENDS

Figure 1. Novel modulators of yeast replicative life span. Replicative life span is significantly extended by deletion of 11 out of 76 yeast ORFs in the RBH set, including the transcription elongation factor *SPT4* (A), ATP-dependent metalloprotease *AFG3* (B), inositol polyphosphate 5-phosphatase *INP53* (C), ribosomal proteins *RPL9A* (D) and *RPL19A* (E), and translation initiation factors *TIF4631* (F), *TIF1* (G), and *TIF2* (H). Deletion alleles of *TOR1*, *RPL6B*, and

IDH2 have been previously shown to increase RLS (Kaeberlein et al. 2005). Life span data from both mating types was combined to generate these mortality curves.

Figure 2. Conserved aging-related genes are enriched for ribosome-associated genes. (A) Gene ontology analysis of the 276 worm aging genes, their yeast putative yeast orthologs in the reciprocal best hit set (RBH, gene total =103), and the 11 conserved aging genes (Table 2) revealed a significant enrichment in the ribosome cellular component when compared to the entire worm or yeast genome ($*P < 0.05$, after Bonferroni Correction for multiple comparisons). Mitochondrial genes are highly enriched in worm aging genes and their corresponding yeast homologs and orthologs, but there is no significant enrichment for mitochondrial genes in the group of conserved longevity genes identified in this study. Essential genes were included in the gene ontology analysis but replicative life span was not measured for these (RBH/ribosome = 8; RBH/mitochondrion = 8). (B) Conserved aging genes highlight the role of the TOR signaling pathway in modulating longevity. Orthologous gene pairs are shown in parentheses (worm genes are in blue and yeast genes are in red). The nutrient sensing kinases are enclosed in the gray box. Many downstream targets of TOR signaling, including ribosomal proteins and translation initiation factors, play a conserved role in life span determination. (**SCH9* was recently shown to have S6K activity, but also has homology to worm aging genes in the Akt family of kinases, such as *akt-1* and *sgk-1*).

Table 1. Genes that modulate longevity are conserved between worms and yeast. Replicative life span (RLS) was measured in viable deletion mutants of putative yeast orthologs (as defined by reciprocal best hit (RBH) analysis) of worm aging genes. This group of putative orthologs was found to be statistically enriched for genes that determine life span compared to a random set of yeast genes (expected frequency). RLS was also measured in yeast homologs (related proteins) of worm aging genes to account for redundancy of function within protein families and identify novel modulators of longevity. (* ‘Essential Yeast Genes’ includes genes essential for viability and other genes not represented in the ORF deletion collection.)

	RBH	Related Proteins
Gene Pairs	103	507
Worm Genes	78	161
Yeast Genes	103	393
Essential Yeast Genes*	27	129
Yeast Genes Analyzed	76	264
Corresponding Worm Genes	56	137
Long-lived Yeast Deletion Strains	11	25
Expected frequency	3.4%	2.6%
Observed Frequency (y/y)	14.5%	9.5%
<i>p</i> -value	0.0034	0.00052

Table 2. Conserved longevity genes. Replicative life span (RLS) was initially measured in *MAT α* ORF deletion strains and confirmed in *MAT α* deletion strains. Statistical significance was determined using Sign Wilcoxon Rank Sum tests. The corresponding worm aging gene for each yeast gene is given; (*) denotes those cases in which more than one worm aging gene is homologous to the yeast gene listed (the closest worm homolog is listed). Human orthologs of yeast genes were identified using P-POD (Heinicke et al. 2007). Human orthologs are not listed (–) in cases where the yeast and worm aging genes are related but not orthologous. (^ Previously described deletion strain (Kaeberlein et al. 2005); ND = not done).

Group	Worm Gene	Yeast Gene	<i>MATα</i>		<i>MATα</i>		Human Ortholog(s)
			RLS extension (%)	p-value	RLS extension (%)	p-value	
RBH	<i>spg-7</i>	<i>AFG3</i>	23.3	1.1E-02	21.4	2.1E-02	<i>AFG3L2</i>
	<i>F43G9.1*</i>	<i>IDH2</i>	17.3	2.2E-03	16.9	5.2E-03	<i>IDH3A</i>
	<i>unc-26</i>	<i>INP53</i>	30.7	1.1E-03	10.2	5.1E-02	<i>SYNJI, SYNJ2</i>
	<i>rpl-19</i>	<i>RPL19A</i>	28.4	6.1E-03	48.2	7.6E-03	<i>RPL19</i>
	<i>rpl-6</i>	<i>RPL6B</i>	13.9	2.9E-03	32.7	1.1E-04	<i>RPL6</i>
	<i>rpl-9</i>	<i>RPL9A</i>	29.4	1.6E-06	19.7	5.1E-03	<i>RPL9</i>
	<i>spt-4</i>	<i>SPT4</i>	40.9	3.7E-11	38.7	1.3E-11	<i>SUPT4H1</i>
	<i>inf-1*</i>	<i>TIF1</i>	29.3	4.8E-04	14.6	6.6E-03	<i>EIF4A2, EIF4A1</i>
	<i>inf-1*</i>	<i>TIF2</i>	33.8	1.4E-02	14.9	4.8E-02	<i>EIF4A2, EIF4A1</i>
	<i>ifg-1</i>	<i>TIF4631</i>	21.2	7.1E-03	22.8	3.3E-02	<i>EIF4G1, EIF4G3</i>
	<i>let-363*</i>	<i>TOR1</i>	20.3	7.6E-04	21.6	3.7E-03	<i>FRAP1</i>
	RP	<i>W09H1.5</i>	<i>ADH1</i>	23.2	4.0E-04	14.9	6.2E-03
<i>T27F7.3</i>		<i>ALG12</i>	24.7	7.1E-03	23.8	3.0E-02	–
<i>B0511.6*</i>		<i>DBP3</i>	33.6	4.3E-03	21.5	4.0E-02	–
<i>sem-5</i>		<i>HSE1</i>	25.5	1.7E-03	13.7	3.7E-02	–
<i>F43G9.1</i>		<i>IDH1</i>	16.2	1.2E-02	9.0	2.9E-02	–
<i>JC8.10</i>		<i>INP51</i>	13.2	4.1E-03	10.6	2.6E-02	<i>SYNJI, SYNJ2</i>
<i>pdk-1</i>		<i>PKH2</i>	22.2	1.8E-03	14.4	2.0E-02	<i>PDPK1</i>
<i>eat-6</i>		<i>PMR1</i>	37.0	4.7E-03	14.7	3.2E-03	–
<i>C06E7.1*</i>		<i>SAM1</i>	18.9	4.0E-02	12.9	2.4E-02	<i>MATIA, MAT1B</i>
<i>rsks-1*</i>		<i>SCH9[^]</i>	40.0	1.4E-24	ND	ND	<i>RPS6KBI, SGK2</i>
<i>Y46H3C.6</i>		<i>SIS2</i>	46.6	4.1E-08	27.7	4.1E-03	–
<i>pos-1</i>		<i>TIS11</i>	32.2	1.1E-03	25.7	5.1E-03	–
<i>erm-1</i>		<i>YGR130C</i>	31.9	1.4E-02	19.2	1.5E-02	–
<i>rab-10</i>		<i>YPT6</i>	30.3	3.9E-04	10.7	8.1E-03	–

REFERENCES

- Boehm, M. and F. Slack. 2005. A developmental timing microRNA and its target regulate life span in *C. elegans*. *Science* **310**: 1954-1957.
- Curran, S.P. and G. Ruvkun. 2007. Lifespan regulation by evolutionarily conserved genes essential for viability. *PLoS Genet* **3**: e56.
- Dillin, A., A.L. Hsu, N. Arantes-Oliveira, J. Lehrer-Graiwer, H. Hsin, A.G. Fraser, R.S. Kamath, J. Ahringer, and C. Kenyon. 2002. Rates of behavior and aging specified by mitochondrial function during development. *Science* **298**: 2398-2401.
- Guarente, L. and C. Kenyon. 2000. Genetic pathways that regulate ageing in model organisms. *Nature* **408**: 255-262.
- Hamilton, B., Y. Dong, M. Shindo, W. Liu, I. Odell, G. Ruvkun, and S.S. Lee. 2005. A systematic RNAi screen for longevity genes in *C. elegans*. *Genes Dev* **19**: 1544-1555.
- Hansen, M., A.L. Hsu, A. Dillin, and C. Kenyon. 2005. New genes tied to endocrine, metabolic, and dietary regulation of lifespan from a *Caenorhabditis elegans* genomic RNAi screen. *PLoS Genet* **1**: 119-128.
- Hansen, M., S. Taubert, D. Crawford, N. Libina, S.J. Lee, and C. Kenyon. 2007. Lifespan extension by conditions that inhibit translation in *Caenorhabditis elegans*. *Aging Cell* **6**: 95-110.
- Heinicke, S., M.S. Livstone, C. Lu, R. Oughtred, F. Kang, S.V. Angiuoli, O. White, D. Botstein, and K. Dolinski. 2007. The Princeton Protein Orthology Database (P-POD): a comparative genomics analysis tool for biologists. *PLoS ONE* **2**: e766.
- Kaeberlein, M. 2006. Genome-wide approaches to understanding human ageing. *Hum Genomics* **2**: 422-428.
- Kaeberlein, M., Kennedy, B.K. 2007. Hot Topics in Aging: Protein Translation, 2007. *Aging Cell* **In press**.
- Kaeberlein, M., R.W. Powers, 3rd, K.K. Steffen, E.A. Westman, D. Hu, N. Dang, E.O. Kerr, K.T. Kirkland, S. Fields, and B.K. Kennedy. 2005. Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. *Science* **310**: 1193-1196.
- Kennedy, B.K., K.K. Steffen, and M. Kaeberlein. 2007. Ruminations on dietary restriction and aging. *Cell Mol Life Sci* **64**: 1323-1328.
- Lee, S.S., R.Y. Lee, A.G. Fraser, R.S. Kamath, J. Ahringer, and G. Ruvkun. 2003. A systematic RNAi screen identifies a critical role for mitochondria in *C. elegans* longevity. *Nat Genet* **33**: 40-48.
- Longo, V.D. and C.E. Finch. 2003. Evolutionary medicine: from dwarf model systems to healthy centenarians? *Science* **299**: 1342-1346.
- Pan, K.Z., J.E. Palter, A.N. Rogers, A. Olsen, D. Chen, G.J. Lithgow, and P. Kapahi. 2007. Inhibition of mRNA translation extends lifespan in *Caenorhabditis elegans*. *Aging Cell* **6**: 111-119.
- Partridge, L. and D. Gems. 2006. Beyond the evolutionary theory of ageing, from functional genomics to evo-gero. *Trends Ecol Evol* **21**: 334-340.
- Schafer, J.C., M.E. Winkelbauer, C.L. Williams, C.J. Haycraft, R.A. Desmond, and B.K. Yoder. 2006. IFTA-2 is a conserved cilia protein involved in pathways regulating longevity and dauer formation in *Caenorhabditis elegans*. *J Cell Sci* **119**: 4088-4100.

- Urban, J., A. Souillard, A. Huber, S. Lippman, D. Mukhopadhyay, O. Deloche, V. Wanke, D. Anrather, G. Ammerer, H. Riezman, J.R. Broach, C. De Virgilio, M.N. Hall, and R. Loewith. 2007. Sch9 is a major target of TORC1 in *Saccharomyces cerevisiae*. *Mol Cell* **26**: 663-674.
- Ventura, N., S. Rea, S.T. Henderson, I. Condo, T.E. Johnson, and R. Testi. 2005. Reduced expression of frataxin extends the lifespan of *Caenorhabditis elegans*. *Aging Cell* **4**: 109-112.
- Wang, D.Y., S. Kumar, and S.B. Hedges. 1999. Divergence time estimates for the early history of animal phyla and the origin of plants, animals and fungi. *Proc Biol Sci* **266**: 163-171.
- Warner, H.R. 2003. Subfield history: use of model organisms in the search for human aging genes. *Sci Aging Knowledge Environ* **2003**: RE1.
- Winkelbauer, M.E., J.C. Schafer, C.J. Haycraft, P. Swoboda, and B.K. Yoder. 2005. The *C. elegans* homologs of nephrocystin-1 and nephrocystin-4 are cilia transition zone proteins involved in chemosensory perception. *J Cell Sci* **118**: 5575-5587.
- Winzeler, E.A., D.D. Shoemaker, A. Astromoff, H. Liang, K. Anderson, B. Andre, R. Bangham, R. Benito, J.D. Boeke, H. Bussey, A.M. Chu, C. Connelly, K. Davis, F. Dietrich, S.W. Dow, M. El Bakkoury, F. Foury, S.H. Friend, E. Gentalen, G. Giaever, J.H. Hegemann, T. Jones, M. Laub, H. Liao, N. Liebundguth, D.J. Lockhart, A. Lucau-Danila, M. Lussier, N. M'Rabet, P. Menard, M. Mittmann, C. Pai, C. Rebischung, J.L. Revuelta, L. Riles, C.J. Roberts, P. Ross-MacDonald, B. Scherens, M. Snyder, S. Sookhai-Mahadeo, R.K. Storms, S. Veronneau, M. Voet, G. Volckaert, T.R. Ward, R. Wysocki, G.S. Yen, K. Yu, K. Zimmermann, P. Philippsen, M. Johnston, and R.W. Davis. 1999. Functional characterization of the *S. cerevisiae* genome by gene deletion and parallel analysis. *Science* **285**: 901-906.
- Wolfe, K.H. and D.C. Shields. 1997. Molecular evidence for an ancient duplication of the entire yeast genome. *Nature* **387**: 708-713.
- Wullschleger, S., R. Loewith, and M.N. Hall. 2006. TOR signaling in growth and metabolism. *Cell* **124**: 471-484.

Figure 1

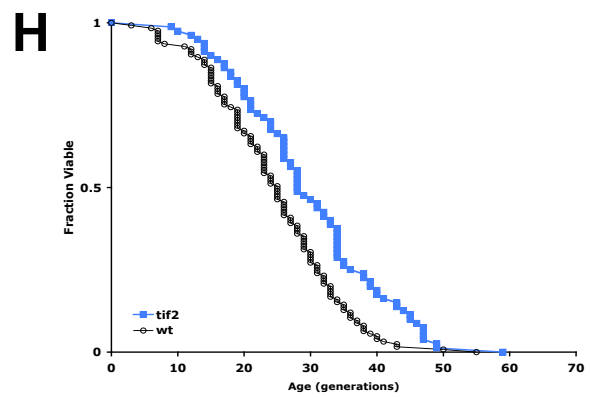
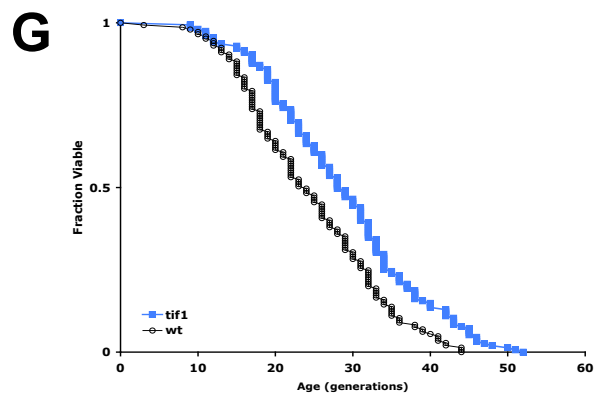
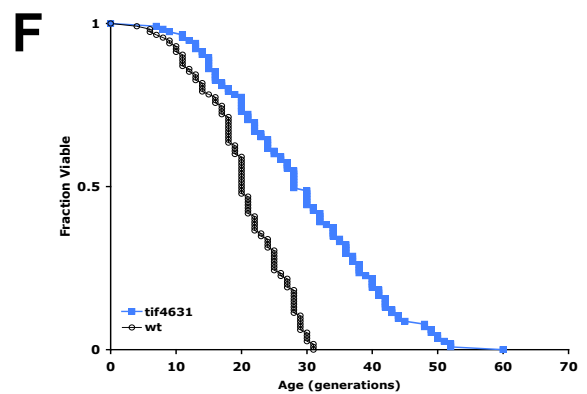
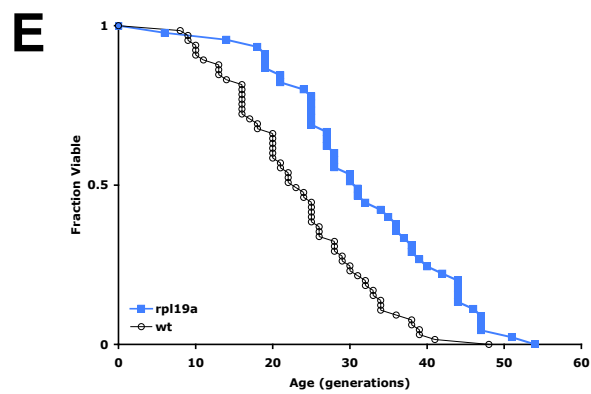
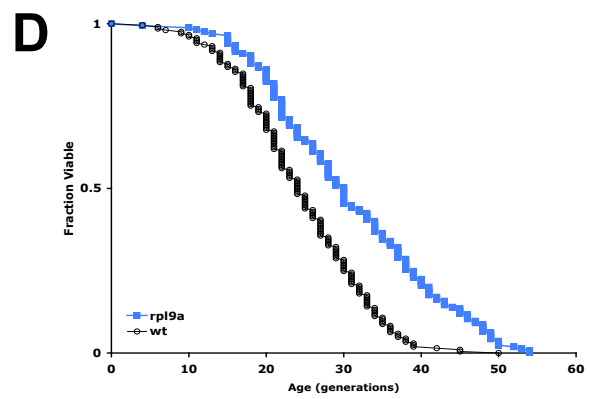
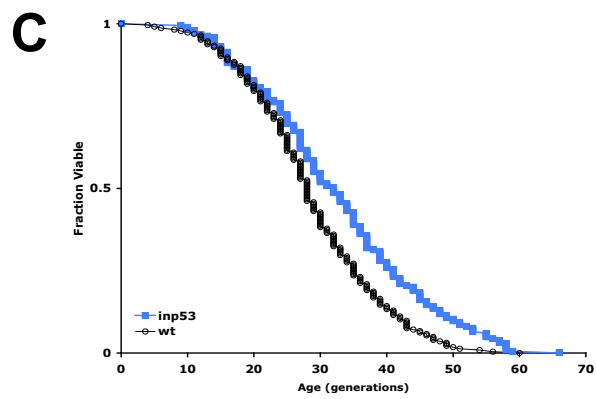
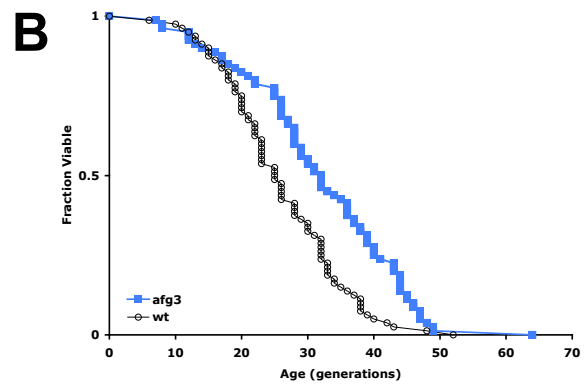
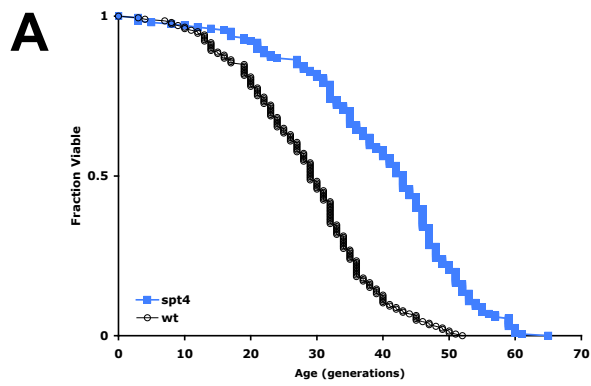
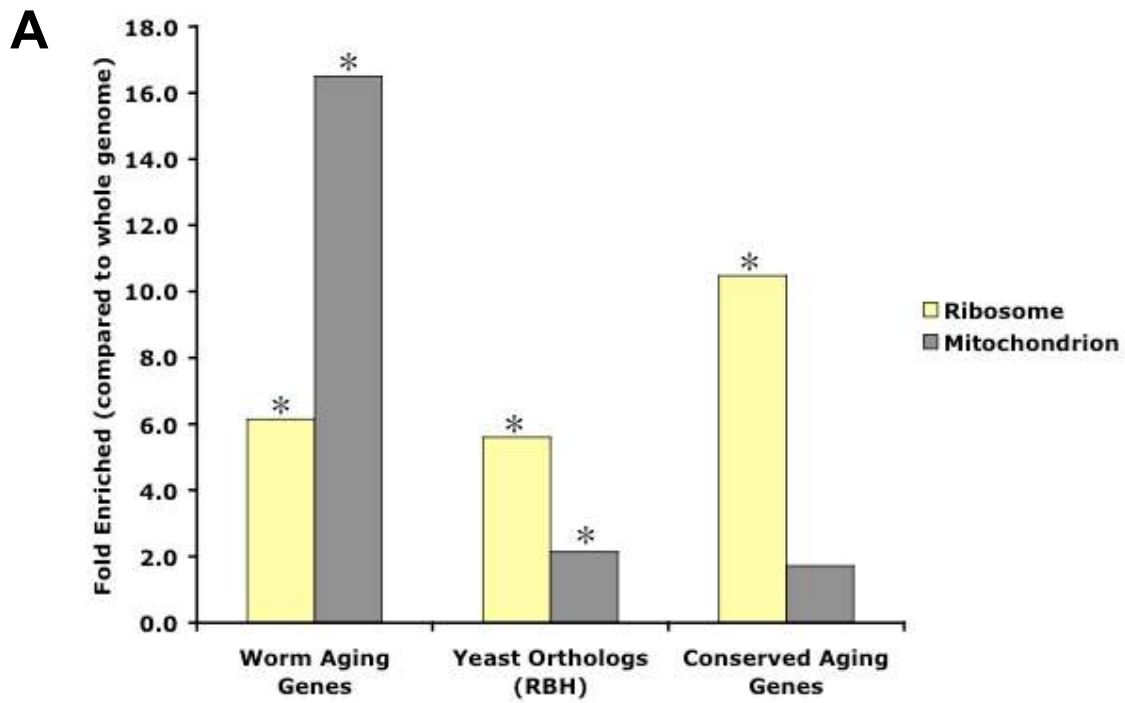
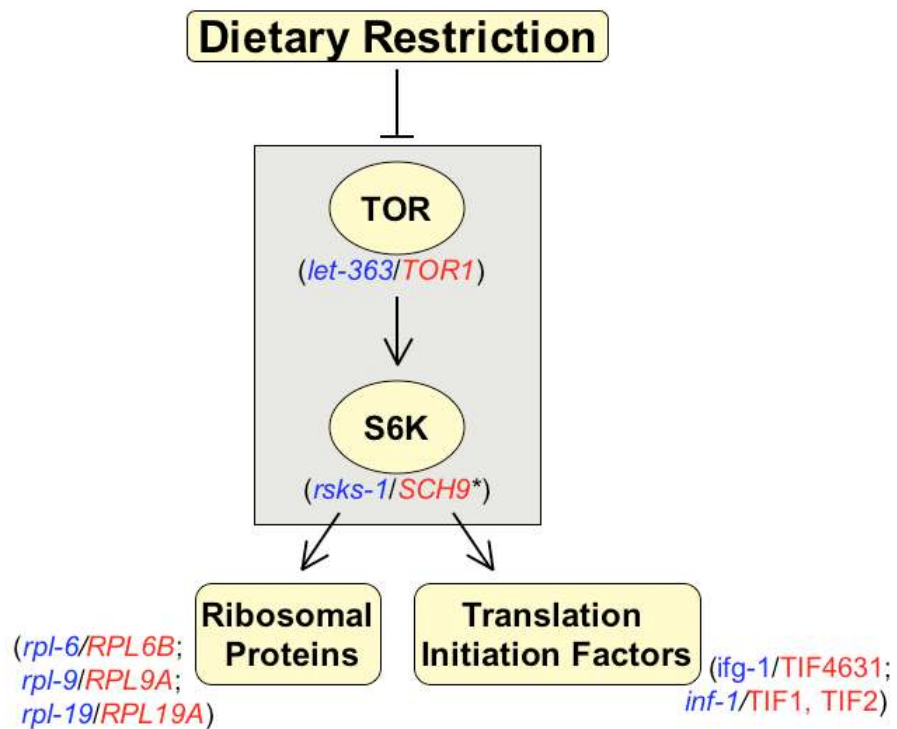
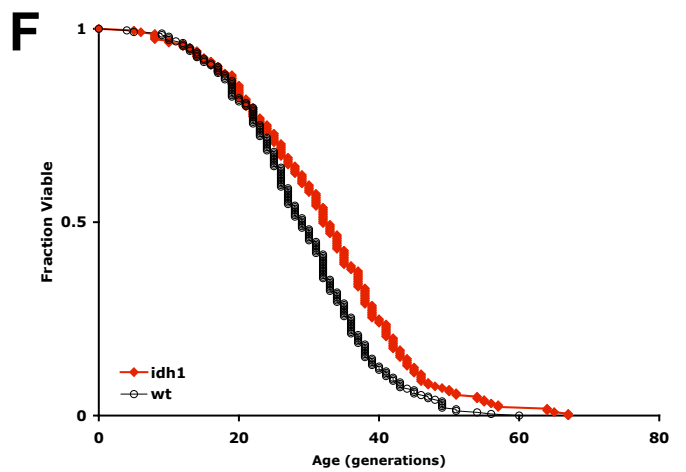
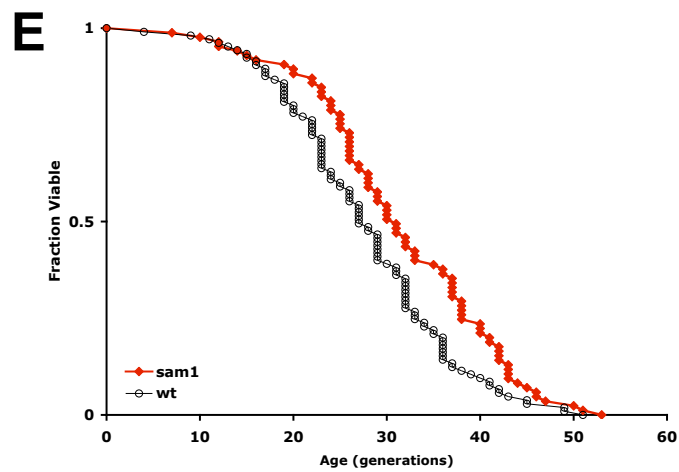
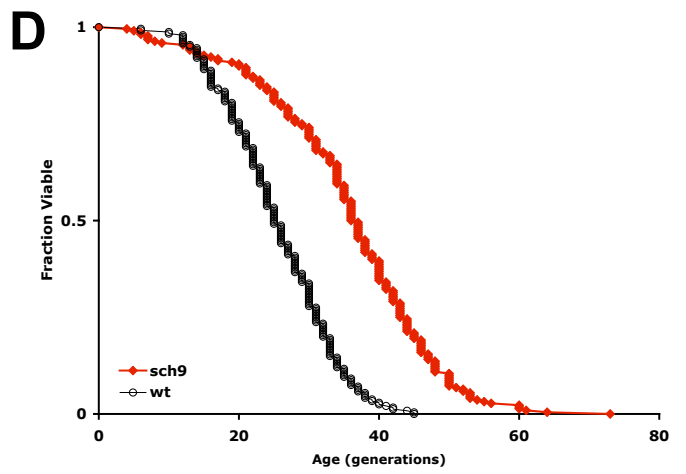
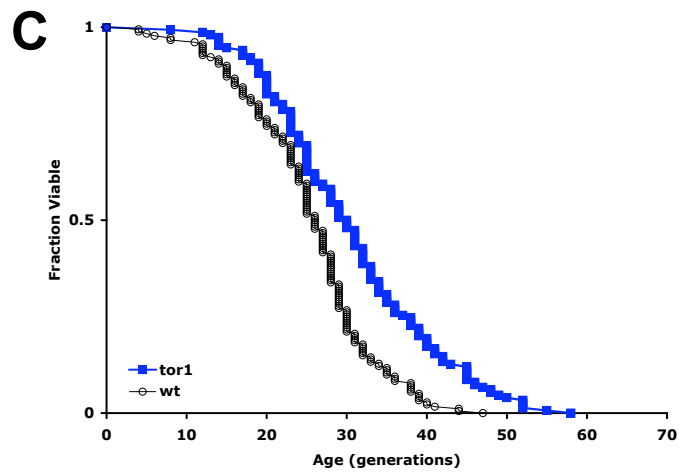
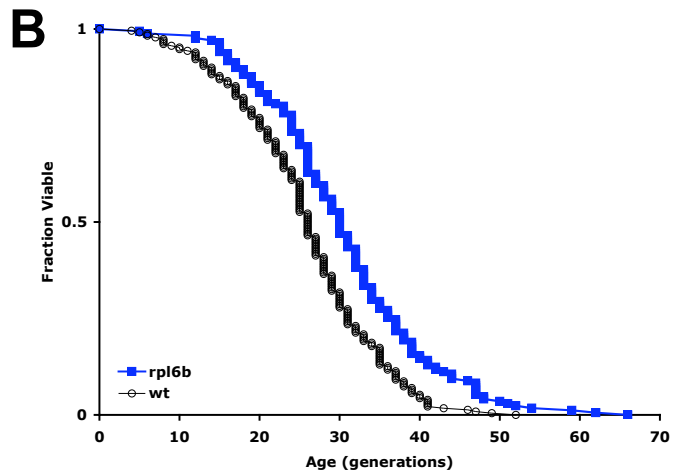
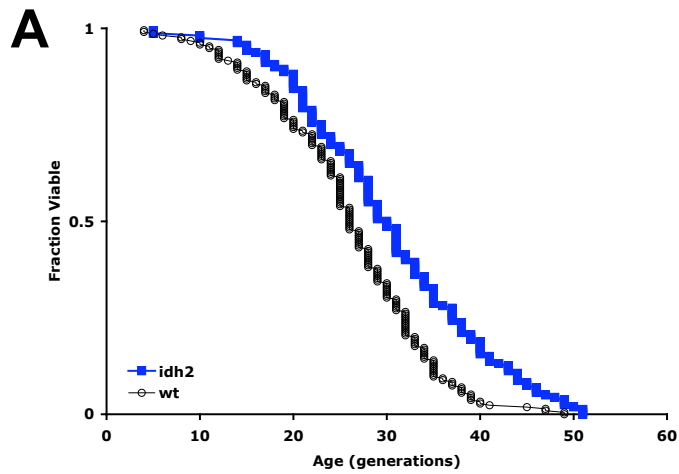


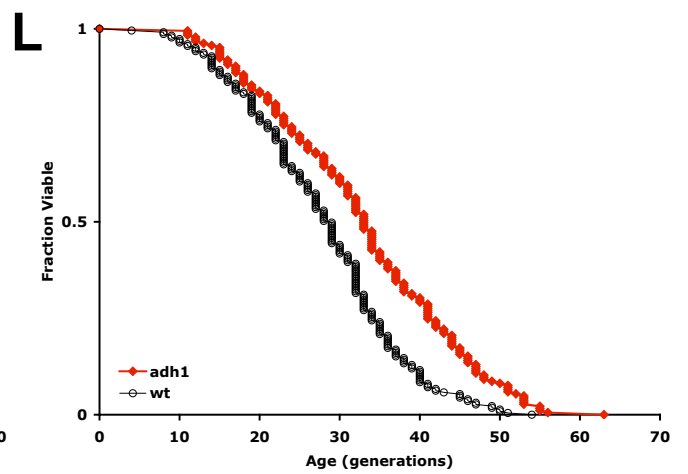
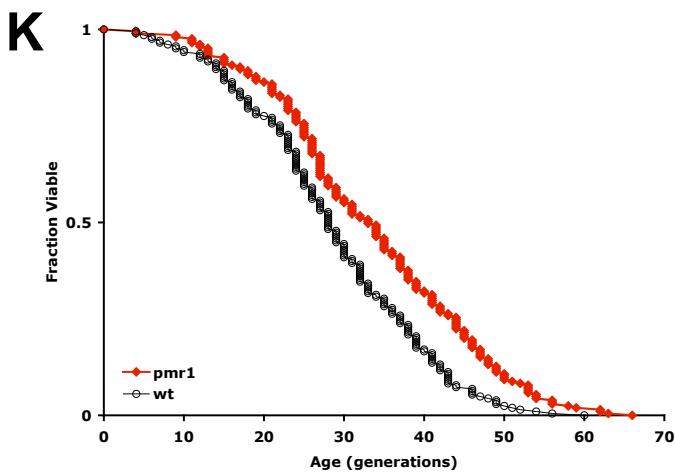
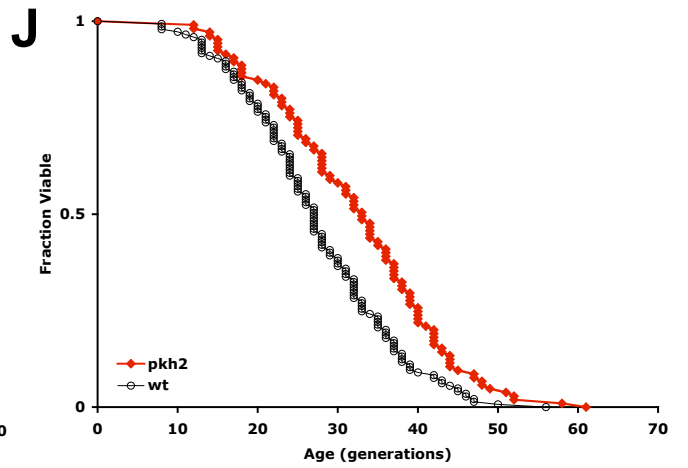
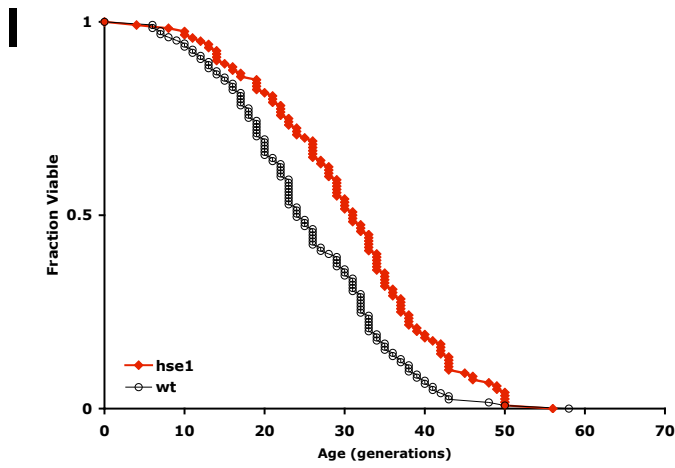
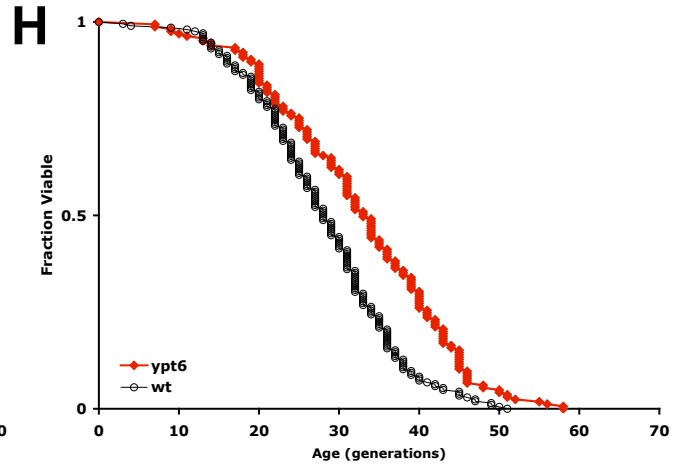
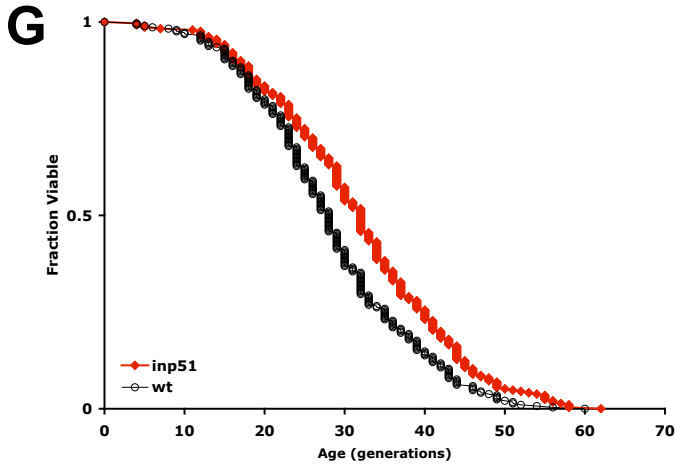
Figure 2



B







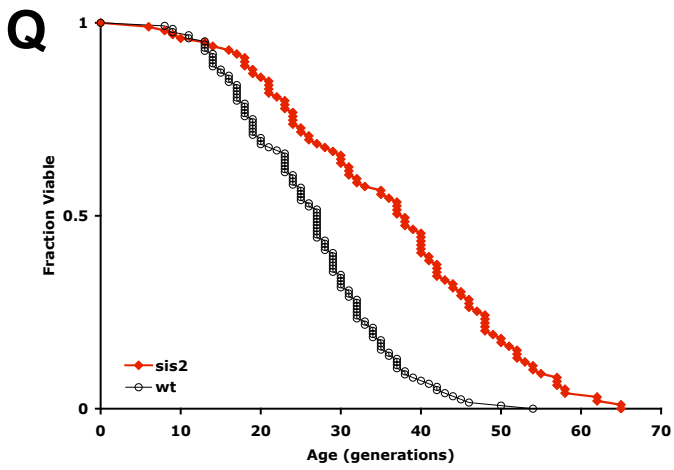
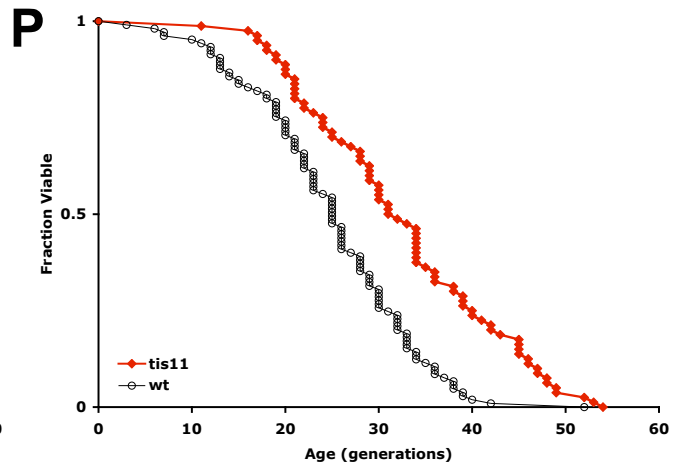
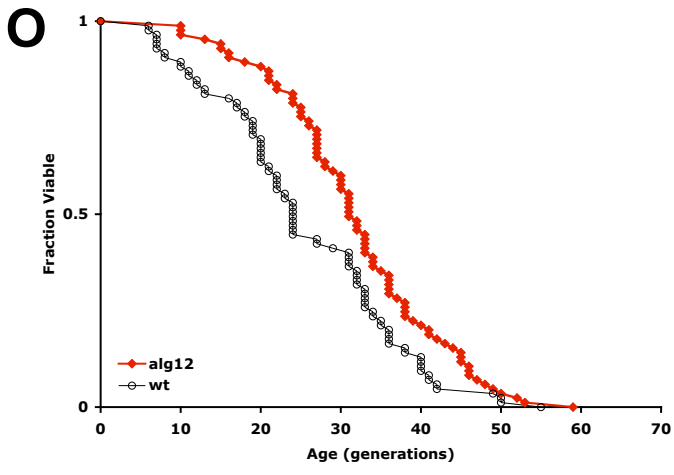
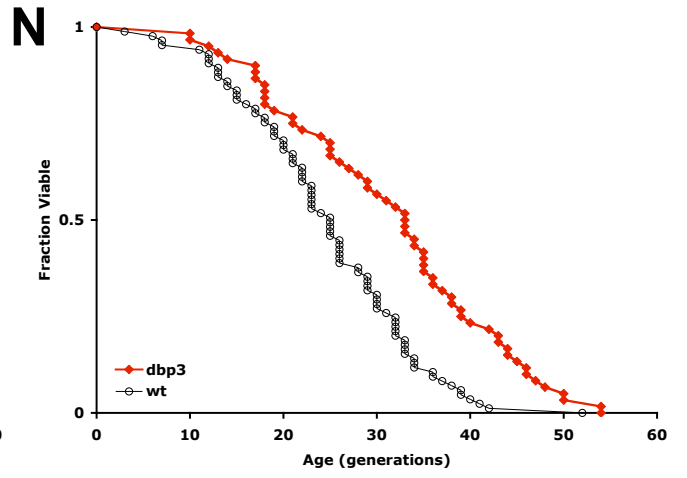
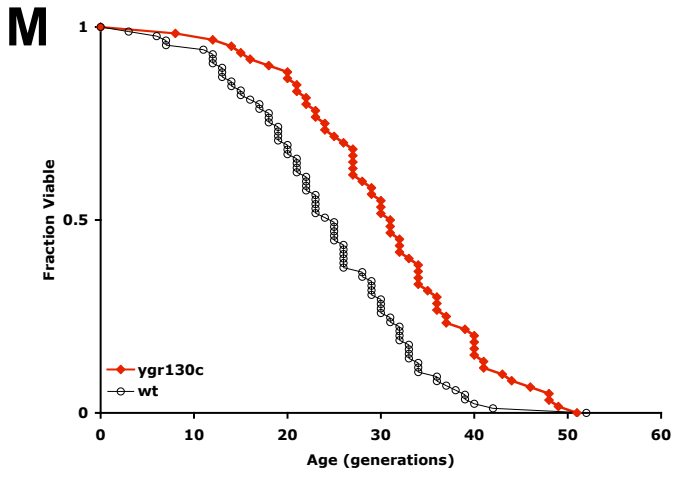


Figure S1. Mortality curves for conserved aging genes. Replicative life span is significantly extended by deletion of *IDH2* (A), *RPL6B* (B), *TOR1* (C), *SCH9* (D), *SAMI* (E), *IDH1* (F), *INP51* (G), *YPT6* (H), *HSE1* (I), *PKH2* (J), *PMR1* (K), *ADH1* (L), *YGR130C* (M), *DBP3* (N), *ALG12* (O), *TIS11* (P), and *SIS2* (Q). Mortality curves for the other conserved aging genes are shown in Figure 1. Curves shown in blue indicate deletion of a gene in the RBH set, while those shown in red indicate deletion of a gene in the RP set. Life span data from both mating types was combined to generate these mortality curves.