

Radiation Damage and Nanofabrication in TEM and STEM

Ray Egerton

Physics Department, University of Alberta, Edmonton, Canada T6G 2E1

regerton@ualberta.ca

Abstract: Different aspects of electron-beam damage are summarized, together with some quantitative evaluation. TEM and STEM are compared in terms of information-to-damage ratio. Electron-beam fabrication is briefly considered in terms of resolution and writing speed.

Keywords: transmission electron microscope, electron-beam damage, knock-on damage, electron charge damage, electron heat damage

Introduction

Radiation damage in the transmission electron microscope (TEM) has been a problem ever since researchers in the late 1940s put organic specimens into the electron beam, hoping to see details beyond the limit of a light microscope, or even processes occurring within living cells. But as in the initial studies of radioactive materials or the first atom-bomb tests, there was little recognition of the damaging effect of ionizing radiation. This process starts with the breaking of chemical bonds, causing structural damage on an atomic scale (observable from TEM diffraction patterns), and leads to longer-range disruption of structure (seen in TEM images) and the removal of some chemical elements (known as mass loss, measurable by energy-loss or x-ray spectroscopy).

With the development of brighter electron sources, improved aberration-corrected optics, and better electron detectors, damage has emerged as an important limiting factor in many areas of electron microscopy.

Electron-beam damage has many aspects, as described in a recent review paper [1], and what follows is, in part, an update to that article, collecting together various mathematical formulas and references to recent publications. From time to time, people find new ways of using electron beams to fabricate small structures, so that creative aspect of radiation damage will also be discussed briefly.

Radiation Effects in an Electron Microscope

An electron beam can cause permanent change to a TEM specimen (thickness t) through several physical mechanisms as discussed below.

Heating. Within the irradiated volume (radius R), inelastic scattering generates heat that is removed by conduction (for example, to an annular heat sink at distance R_0 away). For a specimen of density ρ and specific heat C_p , the temperature rise is exponential with a time constant $\tau_h = R^2 \ln(R_0/R) [\rho C_p / (2\kappa)]$, typically 1 ps for a small probe of diameter $2R=1$ nm. The final temperature rise is $\Delta T \sim I_b E_m (\text{eV}) \ln(R_0/R) / (2\pi\kappa\lambda_i)$, where E_m ($\sim 7Z$ for atomic number $Z < 30$) is the mean energy loss per inelastic collision, λ_i is the inelastic mean free path, and κ is the thermal conductivity of the specimen. For most materials, $\kappa > 0.1$ W/m/K at room temperature and ΔT does

not exceed 10 K for beam currents typical of a modern TEM (Figure 1) so thermal decomposition cannot account for most damage observations. However, polymers such as polystyrene (with $\kappa \sim 0.01$ W/m/K at $T=100$ K) can soften and fall apart in the beam, assisted by forces due to electrostatic charging. With the condenser aperture removed, thermionic-source TEMs have allegedly generated beam currents high enough to melt metallic specimens.

Charging. Inelastic scattering causes emission of secondary and Auger electrons, creating a local charge Q , a radial conduction current I_c , and an increasing surface potential V_s that reduces the total yield $Y(V_s)$ from each surface (Figure 2). This charge accumulation has an RC time constant $\tau_q = (V_s/I_c) (Q/V_s) = \epsilon_0 \epsilon_r / \sigma$ that depends on the electrical conductivity σ and permittivity ϵ_r of the sample, but not on the beam current or radius. For a poor-quality insulator, the result may be a steady-state condition: $I_c = I_b Y(V_s)$, with $V_s = I_b Y(V_s) \ln(R_0/R) / (\pi\sigma t)$ and a charge density within the irradiated volume given by $\rho_e = Q / (\pi R^2 t) = 2\epsilon_0 \epsilon_r I_b Y(V_s) / (\pi\sigma t R^2)$. The voltage gradient is highest at the edge of the probe: $dV/dr = V_s / [R \ln(R_0/R)]$.

Table 1 shows estimates based on these macroscopic formulas, taking $t=100$ nm, $R_0=30$ μm , and $Y(V_s) = 10^{-2}$, for a STEM probe ($2R=1$ nm, $I_b=0.4$ nA) and for TEM illumination ($2R=5$ μm , $I_b=100$ nA, values given in parentheses). The charge density ρ is given in terms of electron units per atom.

Radiolysis. Inelastic scattering causes ionization damage (radiolysis): the breakage of chemical bonds and loss of atomic structure followed by ejection of atoms from the sample (mass loss). For an incident beam with uniform current density, the

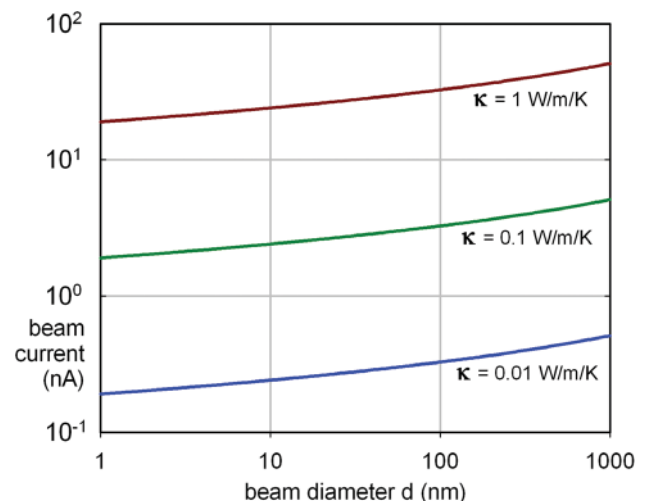


Figure 1: Beam current (in nA) giving $\Delta T=10$ K for specimens of thermal conductivity κ , calculated assuming $E_m=40$ eV, $\lambda_i=100$ nm, $R_0=30$ μm .

TEM signal (for example, diffracted-peak intensity) is often proportional to $\exp(-D/D_c)$, where D is the electron dose (product of current density and exposure time) and D_c is a critical or characteristic dose, dependent on the accelerating voltage and specimen temperature. For a non-uniform beam, the decay is no longer exponential (Figure 3).

Radiolysis in an organic specimen is reduced (by a factor of typically 2 to 20) by cooling to around 100°K, supposedly by suppressing the diffusion of radiation-induced reaction products (single atoms, radicals, ionized species). Chemical scavengers such as antioxidants may also curtail structural damage at room temperature [2], but not at 100°K where diffusion is already largely eliminated. Encapsulation in ice or between layers of carbon is found to reduce damage and is employed in cryo-EM (together with cooling) to permit structural measurements on proteins and other important biological molecules [3].

Any substantial heating effect of a beam will increase the radiation sensitivity, giving a *direct* dose-rate effect (D_c falling with increasing dose rate). Conversely, mass transport and associated structural damage may be diffusion-limited, giving an *inverse* response (Figure 4).

Knock-on damage. High-angle *elastic* scattering causes knock-on displacement of individual atoms of the specimen. This process is inefficient, so it is most commonly observed in metals and semiconductors where radiolysis is absent. The interaction is highly localized and has been used to produce changes on an atomic scale [4]. There is evidence that inelastic scattering can supply some of the energy needed for displacement of a particular atomic species, allowing modification at primary-beam energies below the threshold value calculated from relativistic kinematics [5].

STEM versus TEM for Radiation-Sensitive Specimens

A question of longstanding concern is how much damage scanning transmission electron microscopy (STEM) produces, relative to TEM and for the same amount of retrieved information. Beam heating is less, but only because the current achievable in a small electron probe is much below that used for TEM imaging.

Charging effects are time-dependent and sensitive to dose rate, which is typically a factor of 10^3 – 10^5 higher in STEM. As seen in Table 1, the charging time for resistive specimens may exceed the pixel dwell time (0.16 μ s for a 250×250 raster recorded in 10 s). The specimen would then discharge between adjacent frames if there were no charge trapping. Random or sparse sampling (as opposed to conventional raster scanning) might allow less charge accumulation and damage [6]. A similar situation may apply to short-pulse beams [7,8], except that

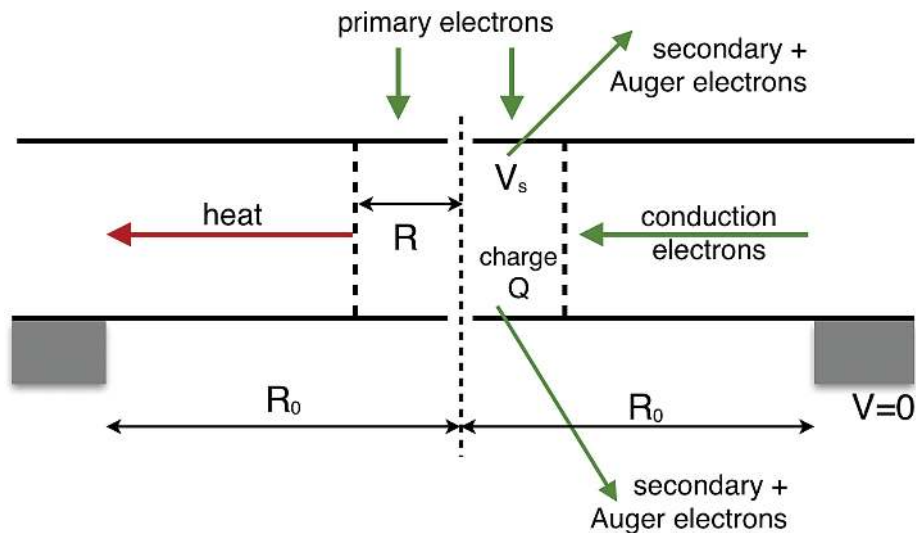


Figure 2: Left, radial heat flow from the irradiated volume of specimen (radius R) to a heat sink at distance R_0 . Right, electron current arising from emission of secondary and Auger electrons, creating a surface potential V_s and accumulated charge Q .

Table 1: Charging calculated assuming classical electrostatics and ohmic conduction.

Specimen	σ (S/m)	ϵ_r	τ	V_s (volt)	ρ_e (e/atom)	dV/dr (MV/m)
Al_2O_3	10^{-8}	2.5	2 ms	70 (6000)	$0.14 (10^{-6})$	$10^4 (10^3)$
Ice (-5°C)	10^{-7}	90	8 ms	7 (600)	$0.5 (10^{-5})$	$10^3 (10^2)$
Pure Si	10^{-3}	12	0.1 μ s	$10^{-3} (0.1)$	$10^{-5} (10^{-10})$	0.3 (0.02)
am-C	10^3	10	0.01 ps	$10^{-9} (10^{-7})$	$10^{-11} (10^{-16})$	$10^{-7} (10^{-13})$

Coulomb repulsion between beam electrons precludes sub- μ m probes with very high current density [9,10].

For good insulators, including most organic specimens, the electric field just outside a STEM probe well exceeds the breakdown strength (100–1000 MV/m), as seen in the last column of Table 1. Calculated assuming ohmic conduction, V_s and ρ will then be overestimates, but local dielectric breakdown might itself be damaging.

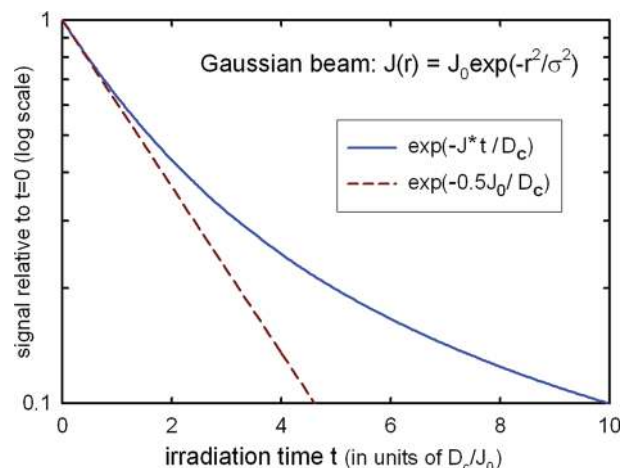


Figure 3: Damage response for a Gaussian or a diffraction-limited probe (blue curve) compared with the exponential decay calculated for a current density equal to half the peak value.

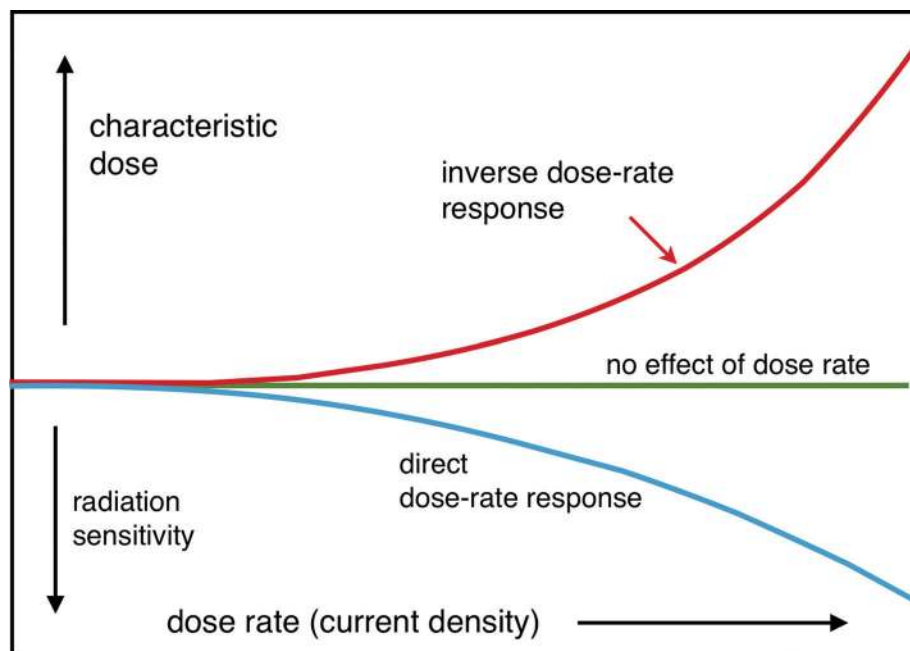


Figure 4: Possible dose-rate effects: *direct* (due to heating or charging) and *inverse* (arising from diffusion limits).

The charge density ρ_e is much higher in STEM mode and might even increase the amount of bond breaking; time-dependent density-functional calculations [11] can hopefully answer this question. Such an effect would lead to a *direct* dose-rate dependence of damage, the amount increasing above some current-density threshold that should increase with increasing primary energy due to the reduced secondary-electron and Auger yield.

In an inorganic oxide, the electrostatic field can give rise to ionic drift, resulting in phase segregation or phase transformation [12] that could be worse in STEM mode. But STEM elemental mapping might benefit from diffusion limits (which reduce radiolytic mass loss) if fast-recording EELS and EDX detectors are available [13].

Charging effects are reduced or eliminated (for example, in cryo-EM imaging of biomolecules) by coating the sample with a thin layer of amorphous carbon [14,15]. Graphene may work even better, as a conducting layer and diffusion barrier [16].

After irradiation by x-rays, *dark progression* of damage is sometimes observed [17]. If it turns out to be significant for electron irradiation of thin specimens, single-frame STEM acquisition would be advantageous [13].

Table 2: Approximate values of the dose D , hole diameter d , time to write 10^6 holes, and the damage mechanism for material removal by a beam (1 nm diameter, current 0.4 nA) of 200 keV electrons.

Material	$D(\text{C}/\text{cm}^2)$	Diameter d	10^6 -time	Mechanism	Ref.
PMMA	2×10^{-3}	5 nm	40 ms	radiolysis	[24]
NaCl	100	4 nm	30 min	radiolysis	[27]
Al_2O_3	5000	1 nm	28 hr	charging	[29]
MoS_2	1600	0.3 nm	9 hr	e-sputter	[5]
Graphene	8000	0.3 nm	44 hr	e-sputter	[35]
Diamond	2×10^5	0.3 nm	46 day	knock-on	[35]

Increasing signal-collection efficiency is just as important as reducing damage. Phase-contrast provides the largest signal from light-element samples and has traditionally been carried out using fixed-beam TEM mode. But using recently developed algorithms and perhaps combined with ptychography, STEM might be capable of comparable or greater efficiency [18–22].

Consequences for Small-Probe Fabrication

Ultraviolet lithography is the basis of microelectronics fabrication and relies on the radiolysis of organic “resists” for patterning. Electron-beam lithography can utilize the same resists but is carried out serially using a focused electron beam. A $50 \mu\text{C}/\text{cm}^2$ dose of 20 keV electrons is said to reduce the molecular weight of 70% of PMMA molecules, crosslinking about 2% of them [23]. After chemical development of the latent image, linewidths down to 20 nm are achievable [23], or below 10 nm with 200 kV aberration-corrected optics [24]. For a sensitive

resist, there is always a statistical limit to spatial resolution due to the small number of beam electrons producing the damage [25]. The resolution of organic polymers may also be limited by their large molecular size [26], which has provided an incentive for using smaller molecules.

A focused field-emission probe has been used to create holes of 1–5 nm diameter in metal halides [27] and oxides [28,29]. If used as a *self-developing* resist that volatilizes during exposure, these inorganic materials avoid the need for wet-chemical processing. Each hole could represent one bit of stored information, the small dimensions enabling high information density, but the sensitivity is well below that of organic resists (Table 2). It may be true that the *Encyclopedia Britannica* could have been written on a pinhead [30], but that procedure would have taken at least a year.

Organics and halides may damage by radiolysis, but hole drilling in amorphous alumina appears to involve electrostatic charging, as suggested by the existence of a threshold current density. If the charge density ρ_e exceeds about 0.1 e/atom, interatomic repulsion causes a Coulomb explosion, and ions are emitted from the specimen surfaces [31].

Any damage process that depends on inelastic scattering is limited in resolution by the long-range nature of the electrostatic interaction between beam and atomic electrons (Coulomb delocalization). Another fundamental limit arises from lateral motion of the secondary electrons, which cause most of the damage in organic materials [32]. As a result of these two effects, it appears that about 50% of the energy is deposited *outside* a 1 nm diameter probe [33,34].

These limits are avoided by basing the fabrication on knock-on displacement of individual atoms, the only damage process

in electrically conducting materials. For thicker samples and high incident energies, knock-on means displacement of lattice atoms to interstitial sites. But for 2D materials (for example, MoS₂, graphene) and lower energies, the displacement is predominantly electron-beam sputtering of surface atoms. Both processes are included in Table 2, which illustrates the limitation on writing speed that results from the low probability of high-angle elastic scattering.

Conclusions

Radiation damage is a curse for electron microscopy of beam-sensitive specimens but a blessing for electron-beam fabrication, which could result in devices far smaller than those possible with UV light. Progress in our understanding has been slow, but time-dependent quantum calculations and further advances in experimental technique should lead to improved control over the damage process.

References

- [1] RF Egerton, *Micron* 119 (2019) doi: 10.1016/j.micron.2019.01.005.
- [2] B Kuei and ED Gomez, *Nat Comm* (2021) doi:10.1038/s41467-020-20363-1.
- [3] R Henderson and CJ Russo, *Microsc Microanal* 25 (Suppl. 2) (2019) doi:10.1017/S1431927619000758.
- [4] AV Krasheninnikov and F Banhart, *Nat Mater* 6 (2007) doi: 10.1038/nmat1996.
- [5] S Kretschmer et al., *Nano Lett* 20 (2020) doi: 10.1021/acs.nanolett.0c00670.
- [6] A Stevens et al., *Appl Phys Lett* 112 (2018) doi.org/10.1063/1.5016192.
- [7] EJ VandenBussche and DJ Flannigan, *Nano Lett* 19 (2019) doi.org/10.1021/acs.nanolett.9b03074.
- [8] C Jing et al., *Microsc Microanal* 26 (Suppl. 2) (2020) doi:10.1017/S1431927620018589.
- [9] RF Egerton, *Adv Struct Chem Imaging* 1 (2015) doi.org/10.1186/s40679-014-0001-3.
- [10] JCHSpence, *Struct Dyn* 4 (2017) doi.org/10.1063/1.4984606.
- [11] Z Cai et al., *Chem Sci* 10 (2019) doi.org/10.1039/C9SC04100A.
- [12] N Jiang, *Rep Prog Phys* 79 (2016) doi.org/10.1088/0034-4885/79/1/016501.
- [13] RF Egerton and H Qian, *Microsc Microanal* 25 (Suppl. 2) (2019) doi:10.1017/S1431927619005695.
- [14] RM Glaeser and KH Downing, *Microsc Microanal* 10 (2004) doi:10.1017/S1431927604040668.
- [15] RM Glaeser, *Meth Enzymol* 579 (2016) doi.org/10.1016/bs.mie.2016.04.010.
- [16] G Algara-Siller et al., *Appl Phys Lett* 103 (2013) doi.org/10.1063/1.4830036.
- [17] M Warkentin et al., *J Synchrotron Rad* 20(2013) doi.org/10.1107/S0909049512048303.
- [18] F Krumeich et al., *Micron* 49 (2013) doi.org/10.1016/j.micron.2013.03.006.
- [19] R Close et al., *Ultramicroscopy* 159 (2015) doi.org/10.1016/j.ultramicro.2015.09.002.
- [20] I Lazić et al., *Ultramicroscopy* 160 (2016) doi.org/10.1016/j.ultramicro.2015.10.011.
- [21] DN Johnstone et al., *Microsc Microanal* 25 (Suppl. 2) (2019) doi:10.1017/S1431927619009462.
- [22] D Ren et al., *Ultramicroscopy* 208 (2020) doi: 10.1016/j.ultramicro.2019.112860.
- [23] CDW Wilkinson and SP Beaumont, “Electron Beam Nanolithography” in *The Physics and Fabrication of Microstructures and Microdevices*, MJ Kelly and C Weisbuch, eds. (1986) Springer. doi.org/10.1007/978-3-642-71446-7_3.
- [24] VR Manfrinato et al., *Nano Lett* 17 (2017) doi.org/10.1021/acs.nanolett.7b00514.
- [25] PA Crozier, *J Vac Sci Technol B* 26 (2008) https://doi.org/10.1116/1.2834560.
- [26] AN Broers et al., *Microelectronic Eng* 32 (1996) doi.org/10.1016/0167-9317(95)00368-1.
- [27] A Muray et al., *J Vac Sci Technol B* 3 (1985) https://doi.org/10.1116/1.583265.
- [28] IG Salisbury et al., *Appl Phys Lett* 45 (1984) https://doi.org/10.1063/1.95115.
- [29] JL Hollenbeck and RC Buchanan, *J Mater Res* 5 (1990) https://doi.org/10.1557/JMR.1990.1058.
- [30] CJ Humphreys et al., *Scanning Microsc Suppl* 4 (1990) 185–92.
- [31] J Cazaux, *Ultramicroscopy* 60 (1995) doi.org/10.1016/0304-3991(95)00077-1.
- [32] B Wu and AR Neureuther, *J Vac Sci Technol B* 19 (2001) doi.org/10.1116/1.1421548.
- [33] RF Egerton, *Microscopy* 67 (2018) doi.org/10.1093/jmicro/dfx089.
- [34] RF Egerton and M Malac, *Microsc Microanal* 10 (Suppl. 2) (2004) https://doi.org/10.1017/S1431927604880541.
- [35] F Banhart, *Rep Prog Phys* 62 (1999) https://doi.org/10.1088/0034-4885/62/8/201.

MT

Expand your Knowledge of Microscopy with

MSA Membership!

Whether your primary focus is in the biological or the physical sciences, MSA takes your knowledge to the next level!

Members Receive:

- A personal subscription to MSA's official journal, *Microscopy and Microanalysis*, and MSA's popular bi-monthly magazine, *Microscopy Today*.
- Peer Networking through the Society's Focused Interest Groups and Local Affiliated Societies.
- MSA Awards Programs, Scholarships, Speaker Opportunities, and much more!

Join MSA Today!

For more information:
visit www.microscopy.org

